Check for updates

OPEN ACCESS

EDITED BY Rita Raisman-Vozari, Paris Brain Institute – CNRS UMR 7225 – Inserm U 1127 – Sorbonne Université UM75, France

REVIEWED BY Ruixiang Wang, The University of Iowa, United States Mauricio Dos Santos Pereira, University of São Paulo, Brazil

*CORRESPONDENCE Behroo Mirza Agha b.mirzaagha@uleth.ca Robert J. McDonald r.mcdonald@uleth.ca

RECEIVED 27 January 2025 ACCEPTED 10 April 2025 PUBLISHED 09 May 2025

CITATION

Mirza Agha B, Monteith M, Earl J, Ganske K, Kaloa T, McDonald KJ, Nixon AG, Panjwani M, Robinson D, Rusnak V, Mohajerani MH, Kovalchuk I, Sutherland RJ, Hong NS and McDonald RJ (2025) Effects of cannabidiol (CBD) treatment on age-related cognitive decline in C57 mice. *Front. Aging Neurosci.* 17:1567650. doi: 10.3389/fnagi.2025.1567650

COPYRIGHT

© 2025 Mirza Agha, Monteith, Earl, Ganske, Kaloa, McDonald, Nixon, Panjwani, Robinson, Rusnak, Mohajerani, Kovalchuk, Sutherland, Hong and McDonald. This is an open-access article distributed under the terms of the **Creative Commons Attribution License (CC** BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Effects of cannabidiol (CBD) treatment on age-related cognitive decline in C57 mice

Behroo Mirza Agha^{1*}, Merrin Monteith¹, Jarem Earl¹, Keona Ganske¹, Tina Kaloa¹, Kelan J. McDonald¹, Abigail G. Nixon¹, Maleeha Panjwani¹, Danika Robinson¹, Valeria Rusnak¹, Majid H. Mohajerani², Igor Kovalchuk³, Robert J. Sutherland¹, Nancy S. Hong¹ and Robert J. McDonald^{1*}

¹Canadian Centre for Behavioural Neuroscience, Department of Neuroscience, University of Lethbridge, Lethbridge, AB, Canada, ²Douglas Research Centre, Department of Psychiatry, McGill University, Montréal, QC, Canada, ³Department of Biological Sciences, University of Lethbridge, Lethbridge, AB, Canada

Aging is associated with cognitive decline, and currently, there are no approved medications that can prevent these impairments. Recently, cannabinoids derived from *Cannabis sativa* have emerged as promising therapeutic compounds with neuroprotective, anti-inflammatory, and cognitive-enhancing properties. Despite their benefits, further research is needed to fully understand their efficacy across various conditions. This study investigates the effects of cannabidiol (CBD) on memory impairment and brain inflammation in aging mice. Fourteen-monthold C57 mice were administered CBD orally for 7 months and subsequently evaluated between 19 and 21 months of age using behavioral tasks that are sensitive to dysfunction of the perirhinal cortex, hippocampus, amygdala, and various brain regions that are crucial for motor control and coordination. The findings of this study indicate that CBD reduces inflammatory response in the brain and improves cognitive decline associated with aging.

KEYWORDS

aging, age-related cognitive decline, cannabidiol (CBD), learning and memory, hippocampus, prefrontal cortex, inflammation, acetylcholine

Introduction

Aging is inevitable, and it is a universal process that brings about biological, psychological, and physiological changes in individuals (Burke and Barnes, 2006). The brain also undergoes physical and neurobiological changes that can lead to impaired cognition. These changes are considered a normal part of the aging process (Murman, 2015; Peters, 2006). In contrast, Alzheimer's disease (AD) is characterized behaviorally by progressive memory impairment, confusion, loss of cognitive abilities, and various brain changes, including abnormal protein accumulation [amyloid-beta and tau (Braak et al., 1994; Braak and Braak, 1991)], inflammation (Akiyama, 2000), and neuron and synapse loss (Tzioras et al., 2022).

Normal cognitive aging is different from AD, and the anatomical and morphological changes in the brain due to aging differ from those observed in AD in both humans and animals. In aging humans and rodents, a decrease in hippocampus (HPC) volume is observed, likely due in part to a decrease in HPC neurogenesis or shrinkage of neurons, which in turn can contribute to deficits in HPC-dependent

memory tasks. Notably, significant cell death, reduction in dendritic length or branching, and decrease in spine density are not typical features of the normal aging process (Burke and Barnes, 2006; Driscoll et al., 2006; Driscoll and Sutherland, 2005; Rapp and Gallagher, 1996; Rasmussen et al., 1996; Raz, 2000; West et al., 1994). Similarly, cognitive changes that occur during normal aging differ from those observed in AD in both humans and animals. For example, both young and old rats can learn a cognitive task in the same number of trials. However, older rats tend to shift away from using HPC-dependent strategies and instead opt for alternative approaches. Consequently, they show deficits in HPCdependent spatial learning and memory in tasks such as the Morris water task (MWT) and Barnes maze (Barnes, 1979; Foster et al., 2012; Gage et al., 1984). Similar to rats, changes in spatial strategy are also observed in older rhesus macaques and humans (Driscoll and Sutherland, 2005; Rapp et al., 1997).

To date, several approaches that can prevent dementia in both humans and animal models have been investigated. These approaches include pharmacological and non-pharmacological interventions, such as lifestyle modifications, cognitive training and mental stimulation, and sleep improvement (Homolak et al., 2018; Li et al., 2023; Stefaniak et al., 2022; Yu et al., 2021). In recent years, cannabinoids, active compounds found in the plant Cannabis sativa, have emerged as novel therapeutic compounds to treat a variety of neurodegenerative diseases (Pagano et al., 2022; Ramírez et al., 2005). Given their recognized neuroprotective properties (Ben-Shabat et al., 1998; Marsicano et al., 2002) and anti-inflammation effects (Burstein, 2015; Nagarkatti et al., 2009), cannabinoids have also been proposed to enhance cognitive function (Bachmeier et al., 2013). Nevertheless, numerous review studies on the effect of cannabinoids on dementia have concluded that the available evidence is inconclusive and insufficient (Bilbao and Spanagel, 2022; Bosnjak Kuharic et al., 2021; Peprah and Mccormack, 2019). Although cannabinoids may have a minimal impact on familial AD, in which more aggressive brain changes and pathology are observed than in the sporadic form of AD (McDonald, 2002; McDonald et al., 2010) or normal aging, they may potentially offer symptomatic relief to individuals experiencing normal cognitive decline. There are three major components in Cannabis sativa: phytocannabinoids, terpenes, and flavonoids. Phytocannabinoids are cannabinoids that are naturally found in the cannabis plant. Delta-9-tetrahydrocannabinol (THC) and cannabidiol (CBD) are the major constituents of cannabis that are of medical interest. THC is the primary psychoactive component, and CBD is the primary non-psychotomimetic component (Wang et al., 2022). CBD modulates the endocannabinoid system chiefly through indirect mechanisms. It acts as a negative allosteric modulator at the CB1 receptor (Laprairie et al., 2015), reducing the receptor's response despite increased anandamide. The action of CBD on the CB2 receptor is more complex and context-dependent, with some studies suggesting weak inverse agonism or allosteric modulation (McPartland et al., 2015; Pertwee, 2008). In addition, CBD also interacts with various neurotransmitter systems (Abame et al., 2021; Chrestia et al., 2022; McPartland et al., 2015).

The present study investigated the effect of CBD on memory decline and brain pathology in aging mice. In this study, 14-monthold C57 mice were given either CBD or a vehicle orally for 7 months and were subjected to behavioral tasks at the age of 19–21 months, such as novel object recognition (NOR), MWT, discriminative fear conditioning to context (DFCTC), and balance beam (BB), that are sensitive to the dysfunction of networks centered on the perirhinal cortex (PRh), HPC, amygdala, and motor coordination, respectively. We hypothesized that aged mice that received CBD would perform better in all these tasks compared to those that received vehicle. We also quantified the volume of HPC and inflammation markers (astrocytes and microglia) in the HPC and medial prefrontal cortex (mPFC). Finally, we assessed the status of cholinergic neurons in the medial septum (MS)/diagonal band (DB) that project to the HPC and neocortex.

Methods

Ethics approval

All experiments were carried out in accordance with Canadian Council of Animal Care and approved by the University of Lethbridge Animal Welfare Committee.

Animals

In this study, we used 19 (15 male and 4 female) C57BL/6 mice (14 months of age at the beginning of the study), weighing 27–44 g [28–44 g (CBD group) and 27–37 g (vehicle group)], bred and raised at the Canadian Centre for Behavioural Neuroscience Vivarium, University of Lethbridge. The animals were housed in pairs under a 12:12 h light/dark cycle, with the lights on at 7:30 a.m. and temperature set at 22°C. All mice had *ad libitum* access to food and water. All treatments, training, and testing were carried out during the light phase of the cycle at the same time every day.

Experimental design

Mice were randomly assigned to two groups: CBD (n = 10, 8 male and 2 female mice) and vehicle (n = 9, 7 male and 2 female mice), and they were subjected to identical handling, testing, and treatment procedures. Starting at 14 months of age, for the following 7 months, they were daily treated with either CBD or vehicle in Nutella. Behavioral testing in NOR, BB, MWT, and DFCTC tasks was started when the mice were 19 months old and lasted for 2 months. All behavioral tasks were carried out during the light cycle at the same time every day. The day after the last day of testing, mice were perfused at 21 months of age, and their brains were extracted for further histological analysis (Figure 1).

Drugs, preparation, and administration

The CBD powder (I.K. lab) was stored in a -20° C freezer before dilution. All dilutions were carried out in a biosafety cabinet fume hood. To prevent product contamination, instruments and materials were sprayed with 100% ethanol and placed under UV light for 15 min except for CBD and oil (PC 100% pure grapeseed oil, No Frills). The oil was filtered using a sterile 0.22-micron



syringe filter (FroggaBio, Canada). A stock solution of oil and CBD powder (20 mg/ml) was heated on a rotating plate to 40° C to allow the powder to dissolve. The solution was aliquoted in Eppendorf tubes and stored at -20° C. While the CBD oil was being used, it was stored at 4° C for 3–4 days.

The treatment was carried out daily by mixing CBD or vehicle in 0.25 g of Nutella. During the treatment, mice were transferred from their housing cages to individual holding cages, which contained a water bottle and a weigh boat with the Nutella/CBD grapeseed oil mix for the CBD group and Nutella/grapeseed oil only for the vehicle group. Mice were weighed every other day to determine the amount of CBD/vehicle (20 ml/kg) they would receive. Holding cages were cleaned daily with Virkon, and water bottles were replaced weekly. Mice were placed in the holding cages for a maximum of 1 h or until every mouse had finished eating its Nutella mix. The dose followed in this study was based on pilot studies assessing dose–response curves that were conducted in our laboratory (Nixon et al., 2022) and previous research (Esposito et al., 2007, 2011; Hayakawa et al., 2008; Martín-Moreno et al., 2011).

Behavioral tasks

Novel object recognition

This task is based on the demonstration that mice have an innate preference to explore novel items without requiring external motivation, reward, or punishment. Research has shown that this task is dependent on the PRh (Kealy and Commins, 2011). When mice that become familiar with two similar objects during the training day are provided with a novel object along with a familiar object on a subsequent test day in a familiar environment, they will spend more time exploring the novel object. This pattern of behavior indicates that mice formed memories of the objects during training and noticed the presence of a novel object during testing (Antunes and Biala, 2011). The NOR test was carried out as described in previous studies (Mehla et al., 2017, Mehla et al., 2019; Vogel-Ciernia and Wood, 2014). Briefly, the mice were transferred from their housing room to the experimental room and familiarized with a white square box (50 cm width \times 50 cm length \times 30 cm height) with a bedding-covered floor by placing them in this box for 10 min daily for 3 days. Training was conducted 24 h after the last habituation day. The mice were placed in the box and allowed to explore two similar objects for 10 min. A test session (test 1) was conducted 24 h after training, in which one of the familiar objects was replaced with a novel object with a different geometry and texture. Mice were given 5 min to explore the familiar and novel objects. A second test session (test 2) was conducted 24 h later, in which the new object from the previous day became the familiar object and a novel object was placed in the box. The mice were allowed 5 min to explore the objects. A retention test was conducted 1 month later, in which mice were placed in the box with the familiar object from test one and a completely new object that they had never interacted with previously. To account for side preference among mice, on all test days, half of the mice in each group had the new object placed on the right and other half had it placed on the left. During habituation, training, testing, and retention, mice were individually placed in the box, and after removing each mouse from the testing box, feces were removed and the corncob bedding was stirred to equally distribute any odor cues. In addition, the objects were wiped with 70% isopropyl alcohol to mask odor cues after each mouse and allowed to completely dry before placing the next mouse in the box. An overhead video camera (Panasonic HDC-SDT750, China) recorded the mice's behavior for further analysis. The investigation ratio (IR) for the novel object was calculated by dividing the time spent by mice exploring the novel object by the total exploration time for both objects for each group.

Balance beam

This test serves as an assessment tool to detect age-related changes in balance and motor coordination, given that physical decline and reduced mobility are common characteristics of aging (Orenduff et al., 2021). In the BB task, mice were required to walk on an elevated narrow aluminum beam (1 cm in diameter, 100 cm long, and 50 cm above a foam pad) to reach an enclosed escape box. Initially, mice were trained by placing them at distances of 10 cm, 50 cm, and 90 cm from the escape box and were required to complete one successful crossing or a maximum of six trials at each distance. On the second training day, mice were placed 90 cm from the escape box and were given a maximum of six trials or until

they completed three successful crossings. On the third day (test day), mice were placed 90 cm from the escape box and given three trials. During training and testing, the beam and the escape box were cleaned with 70% isopropyl alcohol. Furthermore, the mean latency (in seconds) was scored manually.

Morris water task

This task is widely recognized and used to study hippocampaldependent spatial learning and memory (Morris, 1981; Sutherland et al., 1982). Rodents learn to orient themselves using distal visual cues to locate the hidden platform. The MWT was conducted as reported in previous studies (Mehla et al., 2017, Mehla et al., 2019). In brief, the pool (154 cm in diameter, 50 cm deep) was filled with water to a depth of 40 cm and made opaque by adding non-toxic white paint, and water temperature was maintained at 22 \pm 1°C. The pool was divided into four quadrants. A circular platform (11 cm in diameter) was kept 0.5-1 cm below the water surface in one of the quadrants. Three distinct visual cues were placed around the pool to help mice navigate to the platform position. For 8 days of the acquisition phase, each mouse received four distributed training trials from each quadrant randomly. The trial was completed once the mouse found the platform or 60s had elapsed. If the mouse failed to find the platform on a given trial, it was guided to find the platform. After finding the platform or aided placement, the mouse remained on the platform for 10 s. Data were recorded using an automated tracking system (HVS Image Hampton, UK) and analyzed using wtr2100 software (Actimetrics). The latency to find the platform was used as an indicator of the spatial learning ability of mice. The swimming speed of mice was analyzed to rule out the involvement of motor function as a confounding factor. The thigmotaxis behavior of mice, as an indicator of anxiety levels, during the acquisition phase was also analyzed as described in previous studies (Mehla et al., 2023, 2022). A single probe trial was conducted on the 9th day to evaluate spatial memory performance. The data collected during the probe trial were analyzed, and the time spent by mice in the target quadrant and the average time spent in the other three quadrants were measured. Furthermore, average proximity and annulus crossing were analyzed to assess the spatial accuracy of the search for the platform during the probe trial.

Discriminative fear conditioning to context

In this task, mice learn to associate a specific environment with a fear-inducing stimulus, which is useful in the investigation of memory and fear responses (Antoniadis and McDonald, 1999; Antoniadis, 2000). In the DFCTC test, both groups underwent a pre-exposure day during which they explored two different contexts, each with distinct shapes, colors, and scent cues (a white square with amyl acetate scent and a stripped black-and-white triangle with a Vicks scent). These contexts were connected by an alleyway, allowing mice to explore both contexts for 10 min. The amount of time each mouse spent in each context was used to determine the paired context, where they would receive a foot shock, and the unpaired context, where nothing would happen. During the training days (days 1–8), mice were alternatively placed in the paired and unpaired contexts (counterbalanced) for 5 min every day, spending a total of 4 days in each context. In the paired context, mice received three 2-s foot shocks (0.5 mA) at the second, third, and fourth minutes of the session. On days 9 and 10, mice were placed in their assigned context and filmed for 5 min to assess freezing behavior. On day 11 (preference test), the two contexts were connected by the alleyway, and mice were filmed for 10 min to determine the time spent in each context. The pre-exposure, unpaired context training test, and preference test were conducted in room A, whereas only the paired context training was carried out in room B.

Histology

After the completion of behavioral tests, mice were anesthetized and perfused transcardially with 1× phosphate buffered saline (PBS) followed by 4% paraformaldehyde (PFA) in 1× PBS. The brain was removed, post-fixed in 4% PFA overnight at 4°C, and then cryoprotected in a 30% sucrose solution in $1 \times PBS$ and 0.02% sodium azide. The brain was sectioned coronally at 40 µm using a freezing microtome. One brain in the aged CBD group was poorly perfused, and therefore, only Nissl staining could be performed for this brain, resulting in 18 brains being used for immunohistochemistry (IHC) analysis. Nissl staining and IHC procedures for identifying astrocytes [glial fibrillary acidic protein (GFAP)] and microglia [ionized calcium-binding adaptor molecule 1 (Iba1)] were carried out on a subset of sections, with every ninth section being mounted on a charged Superfrost Plus Microscope Slide (VWR). In IHC procedures, choline acetyltransferase (ChAT) was used to stain cholinergic neurons, and every fifth section was mounted. For Nissl staining, sections were stained with cresyl violet and cover-slipped using Permount (Fisher Scientific). For GFAP, Iba1, and ChAT, slides were washed in Tris-buffered saline (TBS) and blocked in a solution containing 3% goat serum and 0.3% Triton-X in TBS for 2 h. The slides were then incubated with primary antibodies in TBS with 0.3% Triton-X at room temperature in a dark humid chamber for 24 h. The following primary antibodies were used: rabbit anti-GFAP (polyclonal, Ab7260, Abcam, 1:2,000); rabbit anti-Iba1 (SAF4318, 019-19741, Wako, 1:1,000); and rabbit anti-ChAT (monoclonal, Ab178850, Abcam, 1:500). After the first incubation, the slides were washed three times for 10 min and then incubated with the secondary antibody, goat anti-rabbit-alexa-594 (IgG [H+L], Invitrogen, 1:1,000), in TBS with 0.3% Triton-X for 24 h for GFAP, Iba1, and ChAT. The slides were again washed three times for 10 min. Then, Iba1 sections were incubated with DAPI (1:2,000 of the 20 ug/ml stock in TBS) for 1 h. After that, the slides were washed once for 10 min. Then, the sections on the slides were covered with Vectashield H-1000 (Vector Laboratories), coverslipped, and later sealed with nail polish. Finally, whole slides were imaged using Nanozoomer (2.0-RS, Hamamatsu, Japan) with $40 \times$ objective magnification.

Histological analysis

The HPC volume in each mouse was estimated following the Cavalieri method utilizing a set of 12 cross-sections of the hippocampal area, starting from -0.94 mm and terminating at

-3.08 mm relative to Bregma (Paxinos and Franklin, 2019) using a light microscope with $10 \times$ objective magnification. GFAP and Iba1 images were analyzed using ImageJ (NIH) and Ilastik (WWW download) (Berg et al., 2019) software.

Statistical analysis

Statistical analyses were conducted using SPSS (v.22.0) statistical software package. Results were presented as mean \pm SE. A repeated-measures ANOVA was used to determine statistically significant differences across days during the acquisition phase of MWT, between the target and the average of other quadrants during the probe trial of MWT, and for the DFCTC task. An independent-sample *t*-test was used to measure significant differences in the NOR, BB, proximity and annulus crossing in the MWT, HPC

volume, GFAP and Iba1 percent coverage, and ChAT counts. Bonferroni correction was used for *post-hoc* comparison. A *p*-value of < 0.05 was considered statistically significant.

Results

CBD improved long-term retention on the NOR

During the training session, no significant difference was observed in exploration time for identical objects A1 and A2 in vehicle and CBD groups. The IR of the novel object in both groups is shown in Figure 2. During tests 1 and 2, a two-tailed one-sample *t*-test revealed that all mice explored the novel object significantly more than the familiar object, as determined by the IR value higher than 0.5 for the aged CBD group for both test 1 and test 2 [$t_{(9)}$ =



FIGURE 2

Effect of CBD on learning and memory function of 19-month-old C57BL/6 mice in the novel object recognition (NOR) task. (a) Investigation ratio for vehicle and CBD groups in test 1, in which a familiar and a new object are used. (b) Investigation ratio for vehicle and CBD groups in test 2, in which the new object in test 1 has become familiar and another new object is used. (c) Investigation ratio for vehicle and CBD groups 1 month later with the familiar object in test 1 and a completely novel object. *P < 0.05 and ***P < 0.001 are considered statistically significant. a—as compared to 0.5 investigation ratio (chance level). Vehicle group (n = 9; 7 male and 2 female), CBD group (n = 10; 8 male and 2 female).



to cross the beam in three trials for vehicle and CBD groups. (b) Average (mean \pm SEM) number of foot slips in three trials for vehicle and CBD groups. (c) Average (mean \pm SEM) number of falls in three trials for vehicle and CBD groups. Vehicle group (n = 9; 7 male and 2 female), CBD group (n = 10; 8 male and 2 female).



Effect of CBD on spatial learning and memory function of 19-month-old C57BL/6 mice in the Morris water task (MWT). (a) Mean latency (mean \pm SEM) to find the hidden platform during the acquisition phase for vehicle and CBD groups. (b) Mean swimming speed (mean \pm SEM) of mice during the acquisition phase for vehicle and CBD groups. (c) Percent time spent in the target quadrant and average of other three quadrants during the probe trial for vehicle and CBD groups. (d) Percent time of annulus crossing during the probe trial for vehicle and CBD groups. (e) Mean (mean \pm SEM) number of annulus crossing during the probe trial for vehicle and CBD groups. (f) Mean (mean \pm SEM) proximity during the probe trial for vehicle and CBD groups. **P* < 0.05 and ***P* < 0.01, are considered statistically significant. a–as compared to day 1 in CBD group. b–as compared to day 1 in vehicle group. Vehicle group (*n* = 9; 7 male and 2 female), CBD group (*n* = 10; 8 male and 2 female).

14.630, P < 0.001; and $t_{(9)} = 4.976$, P < 0.001, respectively] and for the aged vehicle group for test 1 and test 2 [$t_{(8)} = 2.646$, P = 0.029; and t ₍₈₎ = 2.389, P = 0.044, respectively]. During the one-month retention test, mice that received CBD explored the novel object significantly more than mice in the vehicle group, as determined by the IR value higher than 0.5 [$t_{(9)} = 2.876$, P = 0.018 for the aged CBD group; and $t_{(8)} = 1.669$, P = 0.134 for the aged vehicle group].

CBD had no effect on motor balance and coordination

The mean latency, number of foot slips, and number of falls in the BB test in aged vehicle and CBD groups are shown in Figure 3. The latency to cross the beam, number of foot slips, and number of falls were not significantly different between the vehicle and CBD groups, as confirmed by independent sample *t*-tests $[t_{(17)} = 0.104, P = 0.919; t_{(16.745)} = 0.556, P = 0.585; and t_{(17)} = 0.653, P = 0.522$, respectively].

CBD improved spatial memory consolidation

The latency of aged mice to reach the hidden platform over eight training days is shown in Figure 4a. Both vehicle and CBD groups showed learning, which was confirmed by a repeatedmeasures ANOVA indicating a significant effect of days [$F_{(7,119)}$ = 4.163, P < 0.01, $\eta_p^2 = 0.197$, observed power = 0.985]. No significant effects of group [$F_{(1,17)} = 0.017$, P = 0.899, $\eta_p^2 =$



0.001] and interaction $[F_{(7,119)} = 1.360, P = 0.229, \eta_p^2 = 0.074]$ were observed. Although some learning occurred, the performance of both groups of aging subjects was impaired compared with younger mice [day $F_{(7,196)} = 13.951$, P < 0.001, $\eta_p^2 = 0.333$; group $F_{(2,28)} = 9.618$, P < 0.001, $\eta_p^2 = 0.407$; and their interaction $F_{(14,196)} = 2.772$, P < 0.001, $\eta_p^2 = 0.165$]. Young mice were not included in this study, but Supplementary Figure 1 shows the current MWT acquisition data compared with young mice that were run on the same task and training regimen in our laboratory. The swimming speed of mice over the eight training days is represented in Figure 4b, which shows that there is no difference in the swimming speed between both groups across training days. A repeated-measures ANOVA confirmed no significant effects of day $[F_{(7,119)} = 1.204, P = 0.306, \eta_p^2 = 0.066]$, group $[F_{(1,17)} =$ 0.042, P = 0.840, $\eta_p^2 = 0.002$], and their interaction $[F_{(7,119)} =$ 0.410, P = 0.894, $\eta_p^2 = 0.024$]. The thigmotaxis behavior of mice during the eight training days is presented in Figure 4c. A repeatedmeasures ANOVA showed that both groups had significantly reduced thigmotaxis behavior as training progressed $[F_{(7,119)} =$ 23.996, P < 0.001, $\eta_p^2 = 0.585$, observed power = 1.000]; however, no significant effects of group [$F_{(1,17)} = 1.540$, P = 0.232, $\eta_p^2 =$ 0.083] and interaction $[F_{(7,119)} = 0.625, P = 0.632, \eta_p^2 = 0.035]$ were observed.

The quadrant preference of both vehicle and CBD groups on the probe trial is represented in Figure 4d. A significant main effect of quadrant [$F_{(1,17)} = 7.356$, P = 0.015, $\eta_p^2 = 0.302$, observed power = 0.725] was observed, in which the CBD group spent significantly more time in the target quadrant rather than in the remaining quadrants compared with the vehicle group (P =0.027). Annulus crossing on the probe day is shown in Figure 4e. Although the average annulus crossing on the probe trial was higher in the CBD group than in the vehicle group, there was no significant difference between them [$t_{(17)} = -1.482$, P = 0.157]. The average proximity of mice to the platform in the probe trial is represented in Figure 4f. No significant difference was observed in the proximity measure between vehicle and CBD groups [$t_{(17)} =$ 1.260, P = 0.225].



CBD had no effect on discriminative fear conditioning to context

The results of the pre-exposure day are illustrated in Figure 5a, which shows that aged mice in vehicle and CBD groups spent approximately the same amount of time in each of the contexts. A repeated-measures ANOVA comparing time spent in the two contexts revealed no effects of context [$F_{(1,17)} = 1.725$, P = 0.207, $\eta_p^2 = 0.092$, observed power = 0.236], group [$F_{(1,17)} = 0.041$, P = 0.843, $\eta_p^2 = 0.002$, observed power = 0.054], and their interaction [$F_{(1,17)} = 0.201$, P = 0.660, $\eta_p^2 = 0.012$, observed power = 0.071]. During the freezing tests (Figure 5b), mice in the vehicle and CBD groups showed significantly more freezing behavior in the



group (n = 9; 7 male and 2 female).

paired context than in the unpaired context, with no significant difference. This was confirmed by a repeated-measures ANOVA, in which there was a main effect of context $[F_{(1,17)} = 11.860, P]$

= 0.003, η_p^2 = 0.411, observed power = 0.900], but there was no effects of group [$F_{(1,17)}$ = 0.275, P = 0.607, η_p^2 = 0.016] and their interaction [$F_{(1,17)}$ = 0.006, P = 0.937, η_p^2 = 0.000]. As shown in



Figure 5c, on the preference test day, mice in both groups preferred the unpaired context and spent the majority of the time there as compared to the paired context. This was confirmed by a repeated-measures ANOVA, in which there was a main effect of context $[F_{(1,17)} = 401.740, P < 0.001, \eta_p^2 = 0.959$, observed power = 1.000], but there were no effects of group $[F_{(1,17)} = 0.578, P = 0.458, \eta_p^2 = 0.033]$ and their interaction $[F_{(1,17)} = 1.132, P = 0.302, \eta_p^2 = 0.062]$.

No effect of CBD on HPC volume

The HPC volume in the vehicle and CBD groups is shown in Figure 6. There was no significant difference in the HPC volume of aged mice in vehicle and CBD groups, which was confirmed by an independent *t*-test [$t_{(16)} = 0.394$, P = 0.699, CI (-0.743, 1.109)].

CBD decreased astrocytes in HPC

Astrocytes labeled by GFAP in the mPFC, HPC, and PRh in vehicle and CBD groups are shown in Figure 7. Aged mice that received CBD had decreased astrocytosis in their HPC, which was confirmed by an independent *t*-test [$t_{(16)} = 2.303$, P = 0.035, CI (0.075, 2.067)]. No significant difference was found in the mPFC and PRh.

CBD decreased microglia in mPFC

Microglia labeled by Iba1 in the mPFC and HPC in vehicle and CBD groups are shown in Figure 8. A marginally significant difference was found in the mPFC of aged mice as the mPFC of the CBD group had less percent coverage of microglia than that of the



vehicle-treated mice [$t_{(16)} = 2.098$, P = 0.052, CI (-0.009, 1.960)]. No significant difference in microgliosis was found in the HPC between aged mice that received vehicle or CBD [$t_{(16)} = -0.025$, P = 0.981, CI (-0.935, 0.913)].

CBD had no effect on ChAT

The number of ChAT-positive neurons per slice labeled by ChAT IHC in the MS and DB in vehicle and CBD group is shown in Figure 9. No significant difference in the number of ChAT-positive neurons was observed in the MS and DB of aged mice in the vehicle and CBD groups, which was confirmed by an independent *t*-test [MS t $_{(16)} = -1.584$, P = 0.133, CI (-1.695, 0.223); DB t $_{(16)} = 0.725$, P = 0.479, CI (-0.595, 1.268)].

Discussion

This study assessed the effect of a cannabinoid treatment on various brain regions associated with learning and memory and the related brain pathology in aged C57 mice. Fourteen-month-old mice were administered vehicle or CBD orally daily for 7 months. At 19 months of age, behavioral testing was started using NOR, BB, MWT, and DFCTC. After behavioral testing was completed, mice were perfused at 21 months of age for histological assessment. Nissl staining was used to measure HPC cell volume, and IHC analysis was carried out using GFAP, Iba1, and ChAT. The results indicated that aged mice treated with CBD showed improved performance on the NOR and MWT, suggesting that CBD treatment has a positive impact on object memory processes mediated by the PRh and spatial memory functions centered on the HPC. Brain pathology results showed that the aged mice treated with CBD showed reduced inflammation in the HPC and mPFC, but not in the PRh. There was no indication that CBD differentially affected the pathology of the mice depending on sex; however, the number of female subjects in this study was limited. Furthermore, longterm administration of CBD did not negatively affect learning and memory functions, or the brain of the aged subjects, based on our suite of measures. Taken together, these results suggest that oral CBD treatments can improve memory processes that are impaired due to aging. Brain regions and associated functions are mediated by structures in the medial temporal lobe that are typically compromised in age-related cognitive decline, and inflammatory responses in some of these brain regions were also reduced. Importantly, evidence was also observed suggesting that long-term CBD treatments do not have negative impacts on these same functions and brain regions, nor do they cause negative side effects in aged mice subjects.

Perirhinal cortex and object memory processes

One of the key studies conducted on rats and monkeys has suggested that the PRh is important for recognition memory (Kealy and Commins, 2011). Anatomical and electrophysiological evidence is consistent with this finding as the PRh receives and processes extensive visual and olfactory input from the neocortex (Burwell, 2000) and seems crucial for visual and olfactory recognition memory functions (Brown et al., 2010; Buckley et al., 2001; Gaffan, 1994; Hannesson et al., 2005; Kaut and Bunsey, 2001; Kaut et al., 2003; Kealy and Commins, 2011; Mumby et al., 2002; Mumby and Pinel, 1994; Mumby et al., 2007; Suzuki, 1996). Another study has suggested that the specific role of the PRh lies in one aspect of recognition memory: familiarity detection, not recollection (Brown and Aggleton, 2001). In aged rats, deficits in PRh-dependent behavioral tasks are evident, with a marked reduction in the firing rate activity of different cell types within this region while rats learn a novel object task. This reduction may lead to a reduced focus on details and an overemphasis on processing general or gist-like information (Maurer et al., 2017). This impairment is also observed in aged primates (Burke et al., 2011) and humans (Ryan et al., 2012).

Hippocampus and spatial memory processes

Half a century of evidence converging from different fields is consistent with the notion that the HPC is a central structure of a spatial/relational learning and memory network (Eichenbaum et al., 1988; Hirsh, 1974; O'Keefe and Nadel, 1978). The HPC seems essential when an organism must learn about relationships between cues to solve a particular task. For example, rats with HPC damage have been reported to be impaired at the acquisition and retention of various spatial tasks such as the MWT (Morris et al., 1982; Sutherland et al., 1982), a finding that is reliable and valid to date (McDonald and Hong, 2000; McDonald et al., 2004).

In the present study, mice in the CBD and vehicle groups did not show normal learning curves or low asymptotic levels of performance like young adult mice. To further explain this finding, we included a young adult mouse group from another study to show the normal pattern of learning on this task (Supplementary Figure 1). Clearly, the aged subjects in both groups were impaired in the acquisition of the task although they did show some limited improvements. However, during the probe trial, only the CBD-treated mice spent more time swimming in the target quadrant. These results suggest that the CBD treatment has some impact on the ability of the aged mice to remember the spatial location 24 h later.

The fact that neither the acquisition of the task nor measures of spatial specificity (annulus crossings or proximity to escape location) were affected suggests that some general spatial navigation process was improved in the CBD-treated mice. One possibility is that the treatment targeted the ventral HPC. This sub-region of the HPC is presumed to be involved in getting the subject to the general goal area, and these experiences with the escape platform would result in the formation of a precise spatial representation of the exact goal location in the dorsal HPC, leading to spatial specificity (Gruber and McDonald, 2012). Consistent with this idea, both dorsal and ventral HPC contribute to the acquisition of spatial information though the former plays a more prominent role in spatial learning and navigation (Ferbinteanu et al., 2003) and has been shown to be required for spatial specificity (McDonald et al., 2018; Ruediger et al., 2012). These analyses suggest that the CBD treatment acts on the ventral HPC specifically. However, further work is necessary to confirm this observation.

Finally, it is interesting to note that the aged model used in present experiments did not show depletions of cholinergic neurons in the MS/DB; projections from these neurons provide the HPC with extensive acetylcholine tone. These depletions are observed in human AD patients and other preclinical rodent models of AD (Craig et al., 2011; Mehla et al., 2023, 2022) and can be attributable to certain treatment interventions. This pattern suggests a relationship between protein malformations associated with AD and cholinergic depletions independent of aging.

Modes of action of cannabinoids on the brain

Cannabinoids exert their effects on the brain primarily by interacting with the endocannabinoid system, a complex cellsignaling network that regulates various physiological processes. This system includes two primary receptors, CB1 and CB2, though cannabinoids can also influence other receptors and pathways indirectly. THC, the main psychoactive component of cannabis, directly activates both CB1 and CB2 receptors in the brain, while CBD, which is non-psychotomimetic, acts on these receptors indirectly. CB1 receptor activation primarily modulates neuronal function, whereas CB2 activation influences immune cells such as microglia (Benito et al., 2008; Burstein, 2015; Nagarkatti et al., 2009; Pertwee, 2008).

In addition to cannabinoids, terpenoids play a key role in the therapeutic effects of cannabis and contribute to its unique aroma and the associated entourage effect. The entourage effect indicates that cannabinoids and terpenes act synergistically to produce effects that are more potent or distinct than when the compounds are used in isolation (Ben-Shabat et al., 1998; Ferber et al., 2020). Research has shown that cannabis extracts that contain a mixture of cannabinoids and terpenes are more effective at reducing inflammation than CBD or THC alone (Russo, 2011). Therefore, we hypothesize that an extract containing CBD, THC, and terpenoids would produce more effective therapeutic effects. Future research needs to examine the effects of such extracts on the brain.

Cannabinoid treatment efficacy might depend on the severity of neurodegenerative disease

The efficacy of cannabinoid treatment remains a topic of ongoing research, largely due to the complex interactions between cannabinoids and the endocannabinoid system. In vitro studies have suggested that repeated activation of CB1 and CB2 receptors can reduce brain pathology and cognitive deficits associated with AD by decreasing AB levels, inflammation, oxidative stress, excitotoxicity, and ischemia (Benito et al., 2008; Paloczi et al., 2018; Ramírez et al., 2005). Similarly, in vivo studies in the APP \times PS1 mouse model of AD have shown improvements in social and object recognition, but no effect on Aβ levels (Cheng et al., 2014). However, several studies conducted in our laboratory have found little to no impact on AD progression. Specifically, investigations using different dosages of THC and CBD in the APP-NL-G-F mouse model of AD have revealed no significant effect on memoryrelated behavioral tasks or inflammation (Nixon et al., 2022), and a combination of cannabis extract used to improve pathology or memory deficits in the same mouse model has also shown no effect (Robinson et al., 2023). Based on the results of the present experiments, it seems that the severity and aggressiveness of the type of age-related cognitive decline determine whether cannabinoids, or any treatment, might be effective or not.

Inflammation and age-related cognitive decline

There is a well-established association between inflammation and age-related dementia. As people grow older, the immune system undergoes changes that lead to a state of chronic inflammation, often referred to as "inflammaging" (Franceschi and Campisi, 2014). This state is characterized by increased levels of pro-inflammatory markers in the bloodstream and tissues. Neuroinflammation is a hallmark of many forms of dementia and involves the activation of microglia and astrocytes, which can become overactive in response to injury or disease (Franceschi and Campisi, 2014). The findings of the present study suggest that chronic CBD treatment in aging mitigates astrocyte-mediated inflammation, probably by modulating astrocytic signaling pathways. One proposed mechanism is that CBD inhibits equilibrative nucleoside transporter 1 (ENT1), which is responsible for the uptake of adenosine from the extracellular space into cells (Carrier et al., 2006; Ibeas Bih et al., 2015). By inhibiting ENT1, CBD increases the extracellular concentration of adenosine, which results in enhanced interaction with adenosine receptors on astrocytes. This interaction in turn activates adenylate cyclase and subsequently protein kinase A. These signaling events result in the downregulation of pro-inflammatory cytokines and other inflammatory mediators, thereby reducing inflammation. However, further studies are required to comprehensively understand molecular signaling pathways and to identify potential common or distinct mechanisms by which CBD regulates inflammation and synaptic functions.

Other potential targets of CBD

Emerging evidence from animal studies suggests that CBD promotes adult neurogenesis, particularly in the HPC, by enhancing the survival of newborn neurons in the dentate gyrus (Wolf et al., 2010) and by interacting with the nuclear transcription factor PPAR γ , which increases neurogenic activity (Esposito et al., 2011). Moreover, CBD has been shown to modulate neurotransmitter systems such as upregulating norepinephrine (Abame et al., 2021). These effects highlight the potential of CBD as a promising therapeutic target for future research, particularly in addressing age-related dementia.

Caveats

One caveat associated with our study is that it primarily used male mice, with only a small number of female subjects, which limits our conclusions on potential sex-dependent effects of CBD. This disparity in the number of female and male subjects occurred because we used mice that were available at the time of the study; due to unpredictable litter size and sex distribution, the inclusion of a small number of female mice was unavoidable. To address concerns regarding potential sex-dependent effects, we carefully investigated the raw data from female subjects, and their values were well within the range of the male subjects. Thus, the inclusion of only two female mice per group did not affect our primary conclusions and allowed us to modestly increase our overall sample size.

Another caveat is the mode of administration of CBD. It has been reported that oral administration may not be the most effective method for optimal bioavailability and brain penetration. Although oral CBD delivery may have lower bioavailability than other routes such as intravenous administration, it remains a practical and widely used method, particularly in translational research. Marusich and Wiley (2023) found that the THC administration route has minimal impact on potency and duration of effect in C57 mice (except for aerosolized delivery) although THC and CBD have distinct pharmacokinetic properties. In addition, daily intraperitoneal or subcutaneous injections for 7 months would have been impractical due to stress-related effects on the animals and the increased risk of injection-related variability. Thus, oral administration was chosen for its feasibility, translational relevance, ability to minimize procedural stress, and consistency in treatment administration across experimenters.

Finally, the length of the treatment used in our study might be considered unrealistic. The daily CBD treatment lasted 7 months, which is equivalent to 10–20 years of human life. Our aim was to evaluate the long-term effects of CBD supplementation in an attempt to understand the preventative potential of CBD over a considerable part of the animal's lifespan rather than a shortterm therapeutic intervention. Given that preventative strategies such as dietary and vitamin supplements in humans are frequently taken for many years or even lifelong, we believe our approach realistically reflects how CBD might be employed in a human preventative context.

Conclusion

The findings of this study show that CBD targets inflammatory responses in the brain and can improve cognitive decline associated with aging. It is possible that the effects of CBD treatment can be enhanced if an extract with THC and terpenoids is used.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary material, further inquiries can be directed to the corresponding authors.

Ethics statement

The animal study was approved by the University of Lethbridge Animal Welfare Committee. The study was conducted in accordance with the local legislation and institutional requirements.

Author contributions

BM: Formal analysis, Writing - original draft, Writing - review & editing, Data curation, Investigation, Validation, Visualization. MM: Methodology, Writing - review & editing, Formal analysis, Investigation. JE: Methodology, Writing review & editing, Investigation. KG: Methodology, Writing review & editing, Investigation. TK: Methodology, Writing review & editing, Investigation. KM: Methodology, Writing review & editing, Investigation. AN: Methodology, Writing review & editing, Investigation. MP: Methodology, Writing review & editing, Investigation. DR: Investigation, Methodology, Writing - review & editing. VR: Methodology, Writing review & editing, Investigation. MHM: Resources, Writing review & editing. IK: Resources, Writing - review & editing, Methodology. RS: Resources, Writing - review & editing. NH: Writing - review & editing, Methodology, Conceptualization, Data curation, Formal analysis, Investigation, Validation, Visualization. RM: Conceptualization, Writing - review & editing, Funding acquisition, Resources, Supervision, Writing - original draft.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This work was supported

by the Canadian Institutes of Health Research (CIHR) grant PJT-175091 to RM. AN received a CIHR scholarship. MM, TK, and VR received an Alberta Innovates summer research scholarship.

Acknowledgments

The authors thank Di Shao for animal breeding, Cecilia Badenhorst for pilot study, Chayston Williams for histological analysis, and the University of Lethbridge Animal Care Services.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Generative AI statement

The author(s) declare that no Gen AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fnagi.2025. 1567650/full#supplementary-material

SUPPLEMENTARY FIGURE 1

Effect of CBD on spatial learning and memory function of 19-month-old C57BL/6 mice in MWT task compared with 3-months old C57BL/6 mice from another study conducted in our lab. **P < 0.01 is considered statistically significant. a—aged CBD group Day 8 compared to Day 8 of 3-months old mice. b—aged Vehicle group Day 8 compared to Day 8 of 3-months old mice.

References

Abame, M. A., He, Y., Wu, S., Xie, Z., Zhang, J., Gong, X., et al. (2021). Chronic administration of synthetic cannabidiol induces antidepressant effects involving modulation of serotonin and noradrenaline levels in the hippocampus. *Neurosci. Lett.* 744:135594. doi: 10.1016/j.neulet.2020.1 35594

Akiyama, H. (2000). Inflammation and Alzheimer's disease. Neurobiol. Aging 21, 383–421. doi: 10.1016/S0197-4580(00)00124-X

Antoniadis, E. (2000). Amygdala, hippocampus and discriminative fear conditioning to context. *Behav. Brain Res.* 108, 1–19. doi: 10.1016/S0166-4328(99)00121-7

Antoniadis, E. A., and McDonald, R. J. (1999). Discriminative fear conditioning to context expressed by multiple measures of fear in the rat. *Behav. Brain Res.* 101, 1–13. doi: 10.1016/S0166-4328(98)00056-4

Antunes, M., and Biala, G. (2011). The novel object recognition memory: neurobiology, test procedure, and its modifications. *Cogn. Process.* 13, 93–110. doi: 10.1007/s10339-011-0430-z

Bachmeier, C., Beaulieu-Abdelahad, D., Mullan, M., and Paris, D. (2013). Role of the cannabinoid system in the transit of beta-amyloid across the blood-brain barrier. *Molec. Cell. Neurosci.* 56, 255–262. doi: 10.1016/j.mcn.2013.06.004

Barnes, C. A. (1979). Memory deficits associated with senescence: a neurophysiological and behavioral study in the rat. *J. Comp. Physiol. Psychol.* 93, 74–104. doi: 10.1037/h0077579

Benito, C., Tolón, R. M., Pazos, M. R., Núñez, E., Castillo, A. I., and Romero, J. (2008). Cannabinoid CB2 receptors in human brain inflammation. *Br. J. Pharmacol.* 153, 277–285. doi: 10.1038/sj.bjp.0707505

Ben-Shabat, S., Fride, E., Sheskin, T., Tamiri, T., Rhee, M.-H., Vogel, Z., et al. (1998). An entourage effect: inactive endogenous fatty acid glycerol esters enhance 2-arachidonoyl-glycerol cannabinoid activity. *Eur. J. Pharmacol.* 353, 23–31. doi:10.1016/S0014-2999(98)00392-6

Berg, S., Kutra, D., Kroeger, T., Straehle, C. N., Kausler, B. X., Haubold, C., et al. (2019). Ilastik: interactive machine learning for (bio)image analysis. *Nat. Methods* 16, 1226–1232. doi: 10.1038/s41592-019-0582-9

Bilbao, A., and Spanagel, R. (2022). Medical cannabinoids: a pharmacology-based systematic review and meta-analysis for all relevant medical indications. *BMC Med.* 20:259. doi: 10.1186/s12916-022-02459-1

Bosnjak Kuharic, D., Markovic, D., Brkovic, T., Jeric Kegalj, M., Rubic, Z., Vuica Vukasovic, A., et al. (2021). Cannabinoids for the treatment of dementia. *Cochr. Datab. System. Rev.* 9:CD012820. doi: 10.1002/14651858.CD0128 20.pub2

Braak, F., Braak, H., and Mandelkow, E.-M. (1994). A sequence of cytoskeleton changes related to the formation of neurofibrillary tangles and neuropil threads. *Acta Neuropathol.* 87, 554–567. doi: 10.1007/BF00293315

Braak, H., and Braak, E. (1991). Neuropathological stageing of Alzheimer-related changes. Acta Neuropathol. 82, 239–259. doi: 10.1007/BF00308809

Brown, M. W., and Aggleton, J. P. (2001). Recognition memory: What are the roles of the perirhinal cortex and hippocampus? *Nat. Rev. Neurosci.* 2, 51–61. doi: 10.1038/35049064

Brown, M. W., Warburton, E. C., and Aggleton, J. P. (2010). Recognition memory: Material, processes, and substrates. *Hippocampus* 20, 1228–1244. doi: 10.1002/hipo.20858

Buckley, M. J., Booth, M. C. A., Rolls, E. T., and Gaffan, D. (2001). Selective perceptual impairments after perirhinal cortex ablation. *J. Neurosci.* 21, 9824–9836. doi: 10.1523/JNEUROSCI.21-24-09824.2001

Burke, S. N., and Barnes, C. A. (2006). Neural plasticity in the ageing brain. *Nat. Rev. Neurosci.* 7, 30–40. doi: 10.1038/nrn1809

Burke, S. N., Wallace, J. L., Hartzell, A. L., Nematollahi, S., Plange, K., and Barnes, C. A. (2011). Age-associated deficits in pattern separation functions of the perirhinal cortex: a cross-species consensus. *Behav. Neurosci.* 125, 836–847. doi: 10.1037/a0026238

Burstein, S. (2015). Cannabidiol (CBD) and its analogs: a review of their effects on inflammation. *Bioorg. Med. Chem.* 23, 1377–1385. doi: 10.1016/j.bmc.2015.01.059

Burwell, R. D. (2000). The parahippocampal region: corticocortical connectivity. Ann. N. Y. Acad. Sci. 911, 25–42. doi: 10.1111/j.1749-6632.2000.tb06717.x

Carrier, E. J., Auchampach, J. A., and Hillard, C. J. (2006). Inhibition of an equilibrative nucleoside transporter by cannabidiol: a mechanism of cannabinoid immunosuppression. *Proc. Nat. Acad. Sci.* 103, 7895–7900. doi: 10.1073/pnas.0511232103

Cheng, D., Low, J. K., Logge, W., Garner, B., and Karl, T. (2014). Chronic cannabidiol treatment improves social and object recognition in double transgenic APPswe/PS1∆E9 mice. *Psychopharmacology* 231, 3009–3017. doi: 10.1007/s00213-014-3478-5

Chrestia, J. F., Esandi, M., del, C., and Bouzat, C. (2022). Cannabidiol as a modulator of α7 nicotinic receptors. *Cell. Molec. Life Sci.* 79:564. doi: 10.1007/s00018-022-04600-y

Craig, L. A., Hong, N. S., and McDonald, R. J. (2011). Revisiting the cholinergic hypothesis in the development of Alzheimer's disease. *Neurosci. Biobehav. Rev.* 35, 1397–1409. doi: 10.1016/j.neubiorev.2011.03.001

Driscoll, I., Howard, S. R., Stone, J. C., Monfils, M. H., Tomanek, B., Brooks, W. M., et al. (2006). The aging hippocampus: a multi-level analysis in the rat. *Neuroscience* 139, 1173–1185. doi: 10.1016/j.neuroscience.2006.01.040

Driscoll, I., and Sutherland, R. J. (2005). The aging hippocampus: navigating between rat and human experiments. *Rev. Neurosci.* 16, 87–121. doi: 10.1515/REVNEURO.2005.16.2.87

Eichenbaum, H., Fagan, A., Mathews, P., and Cohen, N. J. (1988). Hippocampal system dysfunction and odor discrimination learning in rats: Impairment of

facilitation depending on representational demands. *Behav. Neurosci.* 102, 331–339. doi: 10.1037/0735-7044.102.3.331

Esposito, G., Scuderi, C., Savani, C., Steardo, L., De Filippis, D., Cottone, P., et al. (2007). Cannabidiol in vivo blunts β -amyloid induced neuroinflammation by suppressing IL-1 β and iNOS expression. Br. J. Pharmacol. 151, 1272–1279. doi: 10.1038/sj.bjp.0707337

Esposito, G., Scuderi, C., Valenza, M., Togna, G. I., Latina, V., De Filippis, D., et al. (2011). Cannabidiol reduces $A\beta$ -induced neuroinflammation and promotes hippocampal neurogenesis through PPAR γ involvement. *PLoS ONE* 6:e28668. doi: 10.1371/journal.pone.0028668

Ferber, S. G., Namdar, D., Hen-Shoval, D., Eger, G., Koltai, H., Shoval, G., et al. (2020). The "entourage effect": terpenes coupled with cannabinoids for the treatment of mood disorders and anxiety disorders. *Curr. Neuropharmacol.* 18, 87–96. doi: 10.2174/1570159X17666190903103923

Ferbinteanu, J., Ray, C., and McDonald, R. J. (2003). Both dorsal and ventral hippocampus contribute to spatial learning in Long-Evans rats. *Neurosci. Lett.* 345, 131–135. doi: 10.1016/S0304-3940(03)00473-7

Foster, T. C., DeFazio, R. A., and Bizon, J. L. (2012). Characterizing cognitive aging of spatial and contextual memory in animal models. *Front. Aging Neurosci.* 4:12. doi: 10.3389/fnagi.2012.00012

Franceschi, C., and Campisi, J. (2014). Chronic inflammation (inflammaging) and its potential contribution to age-associated diseases. *J. Gerontol. Series A* 69, S4–S9. doi: 10.1093/gerona/glu057

Gaffan, D. (1994). Dissociated effects of perirhinal cortex ablation, fornix transection and amygdalectomy: evidence for multiple memory systems in the primate temporal lobe. *Exper. Brain Res.* 99, 411–22. doi: 10.1007/BF00228977

Gage, F. H., Dunnett, S. B., and Björklund, A. (1984). Spatial learning and motor deficits in aged rats. *Neurobiol. Aging* 5, 43–48. doi: 10.1016/0197-4580(84)90084-8

Gruber, A. J., and McDonald, R. J. (2012). Context, emotion, and the strategic pursuit of goals: interactions among multiple brain systems controlling motivated behavior. *Front. Behav. Neurosci.* 6. doi: 10.3389/fnbeh.2012.00050

Hannesson, D. K., Howland, J. G., Pollock, M., Mohapel, P., Wallace, A. E., and Corcoran, M. E. (2005). Anterior perirhinal cortex kindling produces long-lasting effects on anxiety and object recognition memory. *Eur. J. Neurosci.* 21, 1081–1090. doi: 10.1111/j.1460-9568.2005.03938.x

Hayakawa, K., Mishima, K., Hazekawa, M., Sano, K., Irie, K., Orito, K., et al. (2008). Cannabidiol potentiates pharmacological effects of Δ 9-tetrahydrocannabinol via CB1 receptor-dependent mechanism. *Brain Res.* 1188, 157–164. doi: 10.1016/j.brainres.2007.09.090

Hirsh, R. (1974). The hippocampus and contextual retrieval of information from memory: a theory. *Behav. Biol.* 12, 421-444. doi: 10.1016/S0091-6773(74)92231-7

Homolak, J., Mudrovčić, M., Vukić, B., and Toljan, K. (2018). Circadian rhythm and Alzheimer's disease. *Med. Sci.* 6:52. doi: 10.3390/medsci6030052

Ibeas Bih, C., Chen, T., Nunn, A. V. W., Bazelot, M., Dallas, M., and Whalley, B. J. (2015). Molecular targets of cannabidiol in neurological disorders. *Neurotherapeutics* 12, 699–730. doi: 10.1007/s13311-015-0377-3

Kaut, K. P., and Bunsey, M. D. (2001). The effects of lesions to the rat hippocampus or rhinal cortex on olfactory and spatial memory: retrograde and anterograde findings. *Cogn. Affect. Behav. Neurosci.* 1, 270–286. doi: 10.3758/CABN.1.3.270

Kaut, K. P., Bunsey, M. D., and Riccio, D. C. (2003). Olfactory learning and memory impairments following lesions to the hippocampus and perirhinal-entorhinal cortex. *Behav. Neurosci.* 117, 304–319. doi: 10.1037/0735-7044.117.2.304

Kealy, J., and Commins, S. (2011). The rat perirhinal cortex: a review of anatomy, physiology, plasticity, and function. *Prog. Neurobiol.* 93, 522–548. doi: 10.1016/j.pneurobio.2011.03.002

Laprairie, R. B., Bagher, A. M., Kelly, M. E. M., and Denovan-Wright, E. M. (2015). Cannabidiol is a negative allosteric modulator of the cannabinoid CB1 receptor. *Br. J. Pharmacol.* 172, 4790–4805. doi: 10.1111/bph.13250

Li, D., Jia, J., Zeng, H., Zhong, X., Chen, H., and Yi, C. (2023). Efficacy of exercise rehabilitation for managing patients with Alzheimer's disease. *Neural Regener. Res.* 19, 2175–2188. doi: 10.4103/1673-5374.391308

Marsicano, G., Moosmann, B., Hermann, H., Lutz, B., and Behl, C. (2002). Neuroprotective properties of cannabinoids against oxidative stress: role of the cannabinoid receptor CB1. *J. Neurochem.* 80, 448–456. doi: 10.1046/j.0022-3042.2001.00716.x

Martín-Moreno, A. M., Reigada, D., Ramírez, B. G., Mechoulam, R., Innamorato, N., Cuadrado, A., et al. (2011). Cannabidiol and other cannabinoids reduce microglial activation in vitro and in vivo: relevance to Alzheimer's disease. *Mol. Pharmacol.* 79, 964–973. doi: 10.1124/mol.111.071290

Marusich, J. A., and Wiley, J. L. (2023). Δ 9-tetrahydrocannabinol discrimination: effects of route of administration in mice. *Drug Alcohol Depend. Rep.* 9:100205. doi: 10.1016/j.dadr.2023.100205

Maurer, A. P., Burke, S. N., Diba, K., and Barnes, C. A. (2017). Attenuated activity across multiple cell types and reduced monosynaptic connectivity in the aged perirhinal cortex. *J. Neurosci.* 37, 8965–8974. doi: 10.1523/JNEUROSCI.0531-17.2017

McDonald, R. J. (2002). Multiple combinations of co-factors produce variants of age-related cognitive decline: a theory. *Canadian J. Exper. Psychol.* 56, 221–239. doi: 10.1037/h0087399

McDonald, R. J., Balog, R. J., Lee, J. Q., Stuart, E. E., Carrels, B. B., and Hong, N. S. (2018). Rats with ventral hippocampal damage are impaired at various forms of learning including conditioned inhibition, spatial navigation, and discriminative fear conditioning to similar contexts. *Behav. Brain Res.* 351, 138–151. doi: 10.1016/j.bbr.2018.06.003

McDonald, R. J., Craig, L. A., and Hong, N. S. (2010). The etiology of agerelated dementia is more complicated than we think. *Behav. Brain Res.* 214, 3–11. doi: 10.1016/j.bbr.2010.05.005

McDonald, R. J., and Hong, N. S. (2000). Rats with hippocampal damage are impaired on place learning in the water task when overtrained under constrained conditions. *Hippocampus* 10, 153–161. doi: 10.1002/(SICI)1098-1063(2000)10:2 < 153::AID-HIPO4 > 3.0.CO;2-A

McDonald, R. J., Hong, N. S., and Devan, B. D. (2004). The challenges of understanding mammalian cognition and memory-based behaviours: an interactive learning and memory systems approach. *Neurosci. Biobehav. Rev.* 28, 719–745. doi: 10.1016/j.neubiorev.2004.09.014

McPartland, J. M., Duncan, M., Di Marzo, V., and Pertwee, R. G. (2015). Are cannabidiol and Δ 9-tetrahydrocannabivarin negative modulators of the endocannabinoid system? *A systematic review. Br. J. Pharmacol.* 172, 737–753. doi: 10.1111/bph.12944

Mehla, J., Deibel, S. H., Karem, H., Hong, N. S., Hossain, S. R., Lacoursiere, S. G., et al. (2023). Repeated multi-domain cognitive training prevents cognitive decline, anxiety and amyloid pathology found in a mouse model of Alzheimer disease. *Commun. Biol.* 6:1145. doi: 10.1038/s42003-023-05506-6

Mehla, J., Deibel, S. H., Karem, H., Hossain, S., Lacoursiere, S. G., Sutherland, R. J., et al. (2022). Dramatic impacts on brain pathology, anxiety, and cognitive function in the knock-in APPNL-G-F mouse model of Alzheimer disease following long-term voluntary exercise. *Alzheimer's Res. Ther.* 14:143. doi: 10.1186/s13195-022-01085-6

Mehla, J., Lacoursiere, S., Stuart, E., McDonald, R. J., and Mohajerani, M. H. (2017). Gradual cerebral hypoperfusion impairs fear conditioning and object recognition learning and memory in mice: potential roles of neurodegeneration and cholinergic dysfunction. J. Alzheimer's Dis. 61, 283–293. doi: 10.3233/JAD-170635

Mehla, J., Lacoursiere, S. G., Lapointe, V., McNaughton, B. L., Sutherland, R. J., McDonald, R. J., et al. (2019). Age-dependent behavioral and biochemical characterization of single APP knock-in mouse (APPNL-G-F/NL-G-F) model of Alzheimer's disease. *Neurobiol. Aging* 75, 25–37. doi: 10.1016/j.neurobiolaging.2018.10.026

Morris, R. G. M. (1981). Spatial localization does not require the presence of local cues. *Learn. Motiv.* 12, 239–260. doi: 10.1016/0023-9690(81)90020-5

Morris, R. G. M., Garrud, P., Rawlins, J. N. P., and O'Keefe, J. (1982). Place navigation impaired in rats with hippocampal lesions. *Nature* 297, 681–683. doi: 10.1038/297681a0

Mumby, D. G., Glenn, M. J., Nesbitt, C., and Kyriazis, D. A. (2002). Dissociation in retrograde memory for object discriminations and object recognition in rats with perirhinal cortex damage. *Behav. Brain Res.* 132, 215–226. doi: 10.1016/S0166-4328(01)00444-2

Mumby, D. G., and Pinel, J. P. J. (1994). Rhinal cortex lesions and object recognition in rats. *Behav. Neurosci.* 108, 11–18, doi: 10.1037/0735-7044.108.1.11

Mumby, D. G., Piterkin, P., Lecluse, V., and Lehmann, H. (2007). Perirhinal cortex damage and anterograde object-recognition in rats after long retention intervals. *Behav. Brain Res.* 185, 82–87. doi: 10.1016/j.bbr.2007.07.026

Murman, D. (2015). The impact of age on cognition. Semin. Hear. 36, 111-121. doi: 10.1055/s-0035-1555115

Nagarkatti, P., Pandey, R., Rieder, S. A., Hegde, V. L., and Nagarkatti, M. (2009). Cannabinoids as novel anti-inflammatory drugs. *Future Med. Chem.* 1, 1333–1349. doi: 10.4155/fmc.09.93

Nixon, A. G., University of Lethbridge. Faculty of Arts and Science. (2022). The effects of cannabinoid compounds on the brain and behaviour of mouse models of Alzheimer's disease. Thesis Department of Neuroscience.

O'Keefe, J., and Nadel, L. (1978). The Hippocampus as a Cognitive Map. Oxford: Clarendon Press.

Orenduff, M. C., Rezeli, E. T., Hursting, S. D., and Pieper, C. F. (2021). Psychometrics of the balance beam functional test in C57BL/6 mice. *Comp. Med.* 71, 302–308. doi: 10.30802/AALAS-CM-21-000033

Pagano, C., Navarra, G., Coppola, L., Avilia, G., Bifulco, M., and Laezza, C. (2022). Cannabinoids: therapeutic use in clinical practice. *Int. J. Mol. Sci.* 23, 3344. doi: 10.3390/ijms23063344

Paloczi, J., Varga, Z. V., Hasko, G., and Pacher, P. (2018). Neuroprotection in oxidative stress-related neurodegenerative diseases: role of endocannabinoid system modulation. *Antiox. Redox Signal.* 29, 75–108. doi: 10.1089/ars.201 7.7144

Paxinos, G., and Franklin, K. B. J. (2019). *Paxinos and Franklin's the Mouse Brain in Stereotaxic Coordinates*. Amsterdam: Academic Press.

Peprah, K., and Mccormack, S. (2019). Medical Cannabis for the Treatment of Dementia: A Review of Clinical Effectiveness and Guidelines. Ottawa (ON).

Pertwee, R. G. (2008). The diverse CB1 and CB2receptor pharmacology of three plant cannabinoids: $\Delta9$ -tetrahydrocannabinol, cannabidiol and $\Delta9$ -tetrahydrocannabivarin. Br. J. Pharmacol. 153, 199–215. doi: 10.1038/sj.bjp.0707442

Peters, R. (2006). Ageing and the brain. Postgrad. Med. J. 82, 84–88. doi: 10.1136/pgmj.2005.036665

Ramírez, B. G., Blázquez, C., del Pulgar, T. G., Guzmán, M., and de Ceballos, M. L. (2005). Prevention of Alzheimer's disease pathology by cannabinoids: neuroprotection mediated by blockade of microglial activation. *J. Neurosci.* 25, 1904–1913. doi: 10.1523/JNEUROSCI.4540-04.2005

Rapp, P. R., and Gallagher, M. (1996). Preserved neuron number in the hippocampus of aged rats with spatial learning deficits. *Proc. Nat. Acad. Sci.* 93, 9926–9930. doi: 10.1073/pnas.93.18.9926

Rapp, P. R., Kansky, M. T., and Roberts, J. A. (1997). Impaired spatial information processing in aged monkeys with preserved recognition memory. *Neuroreport* 8, 1923–1928. doi: 10.1097/00001756-199705260-00026

Rasmussen, T., Schliemann, T., Sørensen, J. C., Zimmer, J., and West, M. J. (1996). Memory impaired aged rats: no loss of principal hippocampal and subicular neurons. *Neurobiol. Aging* 17, 143–147. doi: 10.1016/0197-4580(95)02032-2

Raz, N. (2000). "Aging of the brain and its impact on cognitive performance: integration of structural and functional findings," in *The Handbook of Aging and Cognition, 2nd ed.* (Mahwah, NJ, US: Lawrence Erlbaum Associates Publishers).

Robinson, D. A., University of Lethbridge, Faculty of Arts and Science. (2023). The impacts of a cannabis extract on brain pathology and memory impairments associated with Alzheimer's disease in an APP-NL-G-F transgenic mouse model. Department of Neuroscience.

Ruediger, S., Spirig, D., Donato, F., and Caroni, P. (2012). Goal-oriented searching mediated by ventral hippocampus early in trial-and-error learning. *Nat. Neurosci.* 15, 1563–1571. doi: 10.1038/nn.3224

Russo, E. B. (2011). Taming THC: potential cannabis synergy and phytocannabinoid-terpenoid entourage effects. *Br. J. Pharmacol.* 163, 1344–1364. doi: 10.1111/j.1476-5381.2011.01238.x

Ryan, L., Cardoza, J. A., Barense, M. D., Kawa, K. H., Wallentin-Flores, J., Arnold, W. T., et al. (2012). Age-related impairment in a complex object discrimination task that engages perirhinal cortex. *Hippocampus* 22, 1978–1989. doi: 10.1002/hipo.22069

Stefaniak, O., Dobrzyńska, M., Drzymała-Czyż, S., and Przysławski, J. (2022). Diet in the prevention of Alzheimer's disease: current knowledge and future research requirements. *Nutrients* 14:4564. doi: 10.3390/nu14214564

Sutherland, R. J., Kolb, B., and Whishaw, I. Q. (1982). Spatial mapping: definitive disruption by hippocampal or medial frontal cortical damage in the rat. *Neurosci. Lett.* 31, 271–276. doi: 10.1016/0304-3940(82)90032-5

Suzuki, W. A. (1996). The anatomy, physiology and functions of the perirhinal cortex. *Curr. Opin. Neurobiol.* 6, 179–186. doi: 10.1016/S0959-4388(96)80071-7

Tzioras, M., McGeachan, R. I., Durrant, C. S., and Spires-Jones, T. L. (2022). Synaptic degeneration in Alzheimer disease. *Nat. Rev. Neurol.* 19, 19–38. doi: 10.1038/s41582-022-00749-z

Vogel-Ciernia, A., and Wood, M. A. (2014). Examining object location and object recognition memory in mice. *Curr. Prot. Neurosci.* 69, 1–17. doi: 10.1002/0471142301.ns0831s69

Wang, Y., Hong, Y., Yan, J., Brown, B., Lin, X., Zhang, X., et al. (2022). Low-dose delta-9-tetrahydrocannabinol as beneficial treatment for aged APP/PS1 mice. *Int. J. Mol. Sci.* 23:2757. doi: 10.3390/ijms23052757

West, M. J., Coleman, P. D., Flood, D. G., and Troncoso, J. C. (1994). Differences in the pattern of hippocampal neuronal loss in normal ageing and Alzheimer's disease. *Lancet* 344, 769–772. doi: 10.1016/S0140-6736(94)92338-8

Wolf, S. A., Bick-Sander, A., Fabel, K., Leal-Galicia, P., Tauber, S., Ramirez-Rodriguez, G., et al. (2010). Cannabinoid receptor CB1 mediates baseline and activityinduced survival of new neurons in adult hippocampal neurogenesis. *Cell Commun. Signal.* 8:12. doi: 10.1186/1478-811X-8-12

Yu, T.-W., Lane, H.-Y., and Lin, C.-H. (2021). Novel therapeutic approaches for Alzheimer's disease: an updated review. *Int. J. Mol. Sci.* 22:8208. doi: 10.3390/ijms22158208