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Exploring efficient and effective mammalian models for Alzheimer's disease

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The aim of this study was to explore and discuss efficient and effective mammalian models for Alzheimer's disease (AD). In this study, efficient AD models are characterized by a small body size, a short lifespan, and rapid development of the main pathology including amyloid plaque formation. Effective AD models are expected to exhibit not only the main pathology, but also co-pathology associated with other neurodegenerative diseases (e.g., Lewy body dementia), systemic disturbances such as disrupted central–peripheral homeostasis, and sleep–circadian failures. This reflects recent findings indicating that AD is far more multifactorial than previously assumed. Although further investigation is required, non-human primates, particularly common marmosets (*Callithrix jacchus*), and dogs (*Canis lupus familiaris*) are candidates of promising and effective AD models. Tree shrews (*Tupaia belangeri*), guinea pigs (*Cavia porcellus*), and evolutionary related species including degus (*Octodon degus*) constitute an alternative group of AD models that remain underexplored but potentially efficient and effective. These mammalian models, together with hypothesis-driven mouse models and advances in data science technologies including omics and imaging analyses, may lead to breakthroughs in AD research, resulting in the development of effective prevention and treatment for AD.

KEYWORDS

amyloid- β , α -synuclein, blood brain barrier, cerebral amyloid angiopathy, marmoset, dog, tree shrew, rodent

1 Introduction

Alzheimer's disease (AD), the most common form of dementia, is characterized by the presence of β -amyloid ($A\beta$)-containing extracellular plaques and tau-containing intracellular neurofibrillary tangles in the brain (Hardy and Selkoe, 2002; Bloom, 2014; Scheltens et al., 2021). In amyloid hypotheses, form(s) of $A\beta$, such as plaques and soluble oligomers, in the brain initiates a pathophysiological cascade leading the tau pathology, neuro-inflammation related to activation of microglia and astrocyte, neuronal death and cognitive decline (Selkoe and Hardy, 2016; Cline et al., 2018). However, prevention and treatment targeting the brain $A\beta$ have not been as successful as the amyloid hypotheses had expected.

AD may be far more multifactorial than previously assumed, regarding co-pathology and systemic abnormalities. For example, emerging evidences have supported that AD brains frequently share the pathology (co-pathology) associated with other dementias such as Lewy body dementia (LBD) and frontotemporal lobe dementia (FTLD) through the

interplay among A β , tau, α -synuclein and TAR DNA-binding protein of 43 kDa (TDP-43) (Robinson et al., 2018; Sengupta and Kayed, 2022). Also, AD may extend beyond the brain, involving systemic alterations (Wang et al., 2017; Cheng et al., 2020; Xu et al., 2025). At least 10 to 20 multilevel factors at molecular, cellular, tissue-organ and individual levels are then associated with AD: (1) molecular level: A β , tau, α -synuclein, TDP-43 and apolipoprotein E (APOE; a major risk factor for AD) (Yamazaki et al., 2019; Serrano-Pozo et al., 2021; Jackson et al., 2024), (2) cellular level: astrocyte, microglia, oligodendrocyte, T-cell and neutrophil (Huang et al., 2022; Gericke et al., 2023), (3) tissue-organ level: cortex, hippocampus, hypothalamus, liver, pancreas, kidney and gut (Wang et al., 2017; Xu et al., 2025), and (4) individual level: infection (Moir et al., 2018; Vojtechova et al., 2022), sleep-circadian failure (Ju et al., 2014; van Erum et al., 2018), cardiovascular diseases (Stampfer, 2006; Tini et al., 2020), diabetes (Sims-Robinson et al., 2010; Takeda et al., 2010; Arnold et al., 2018) and epilepsy (Palop and Mucke, 2009; Horváth et al., 2016).

Interestingly, many mammalian species spontaneously exhibit the amyloid plaque as they age (McKean et al., 2021; Sharma et al., 2023; Ferrer, 2024; Supplementary material). Several mammalian species also naturally present the tau pathology (McKean et al., 2021) and some show symptoms as well (Osella et al., 2007; Prpar Mihevc and Majdić, 2019). The most surprising fact is that dogs, a mammalian species which is evolutionarily divergent from human in mammals (Figure 1), can naturally develop AD-like disorder without any interventions: it is canine cognitive dysfunction (CCD) (Osella et al., 2007; Landsberg et al., 2017; Prpar Mihevc and Majdić, 2019). CCD dogs can exhibit the amyloid plaque and tau pathology, and, very surprisingly, they present symptoms exactly like human such as disorientation (Osella et al., 2007). This fact encourages us and evokes idea of naturally onset mammals for AD, implying that key progression pathways of the multifactorial AD may be fundamentally conserved in mammals.

This review provides the summary of the possibility of effective mammalian models for the multifactorial AD. Also, the efficiency including a small body size, a short life span and rapid disease progression of the animals is discussed.

2 Key criteria for efficient and effective animal models for Alzheimer's disease

Animal models are expected to be efficient and effective. For the efficiency, the AD models need to have a small body size (low maintenance costs), a short lifespan (short generation time) and rapid disease progression: models are particularly required to exhibit the A β accumulation and plaques as early as possible with preserved effectiveness.

For the effectiveness, animal models for AD are expected to exhibit at least the amyloid and tau pathology, and also co-pathology such as those involving α -synuclein and systemic alterations including direct dysfunction in peripheral tissues, breakdown of blood-brain-barrier (BBB) and disruption of central and peripheral homeostasis. This reflects recent findings indicating that AD is far more multifactorial than previously assumed,

involving co-pathology and systemic alterations. Although α -synuclein can be detected in the brain, particularly as a component of the amyloid plaque (Ueda et al., 1993), it can be detected in peripheral, particularly in gut (Xu et al., 2025). This may construct the gut-to-brain axis of α -synuclein spreading. AD patients also show systemic alterations. Typical systemic alterations associated with AD include the A β and tau accumulations in peripheral tissues and organs (Xu et al., 2025), peripheral inflammation and dysfunction (Wang et al., 2017) and breakdown of BBB (Bowman et al., 2007; Sweeney et al., 2018; Cai et al., 2018). Especially, BBB impairment is thought to be critical in the multifactorial AD: since BBB maintains physiological and immunological homeostasis in central nervous system (CNS) and periphery, and BBB leakage may precede the senile plaque (Ujiie et al., 2003), implying that BBB alterations may be linked to the true initiator(s) of AD. BBB dysfunction is believed to be associated with cerebral amyloid angiopathy (CAA) (Kalaria, 1999; Carrano et al., 2012; Magaki et al., 2018) and epilepsy (van Vliet et al., 2007; Löscher and Friedman, 2020; Greene et al., 2022). APOE may also associate with BBB dysfunction (Montagne et al., 2020; Liu et al., 2022). Finally, sleep and circadian disorders are probably associated with huge numbers of factors in the multifactorial AD including co-pathology and systemic alterations (Ju et al., 2014; Musiek and Holtzman, 2016; Cuddapah et al., 2019; Patke et al., 2020; Nassan and Videncovic, 2022). We need to sleep for our health (Buyse, 2014; Mander et al., 2017): not only for AD, but also for other neurodegenerative diseases (Malhotra, 2018; Husain, 2021).

In summary, for the efficiency, the models are required to exhibit the amyloid and related pathology as early as possible. In addition, a smaller body size is preferred to minimize maintenance costs. The effective models for the multifactorial AD should exhibit at least the amyloid and tau pathology. Also, co-pathology of α -synuclein and related accumulation in the brain and body are expected to be observed. For the systemic abnormalities in AD, the models need to present direct alterations in peripheral tissues, BBB dysfunction and/or related disorders such as CAA and epilepsy that are associated with disrupted peripheral-central homeostasis. Moreover, sleep-circadian failures, which are related to co-pathology and systemic abnormalities, are desirable features to be observed. However, it seems impossible to obtain detailed studies for all aspects of co-pathology, systemic alterations and sleep-circadian failures in underexplored animals. This study then evaluates the effective model as the possibility of either co-pathology or at least one of the systemic alterations, and roughly discusses (1) whether each animal species is diurnal and (2) selective sleep-circadian reports in each species.

3 Efficient and effective mammalian models for Alzheimer's disease

3.1 Non-human primate: common marmosets

Non-human primates (NHPs) including common marmosets (marmoset; *Callithrix jacchus*) are probably promising and effective models for AD (McKean et al., 2021; Stonebarger et al., 2021; Rizzo et al., 2023). In particular, marmosets, a small primate species

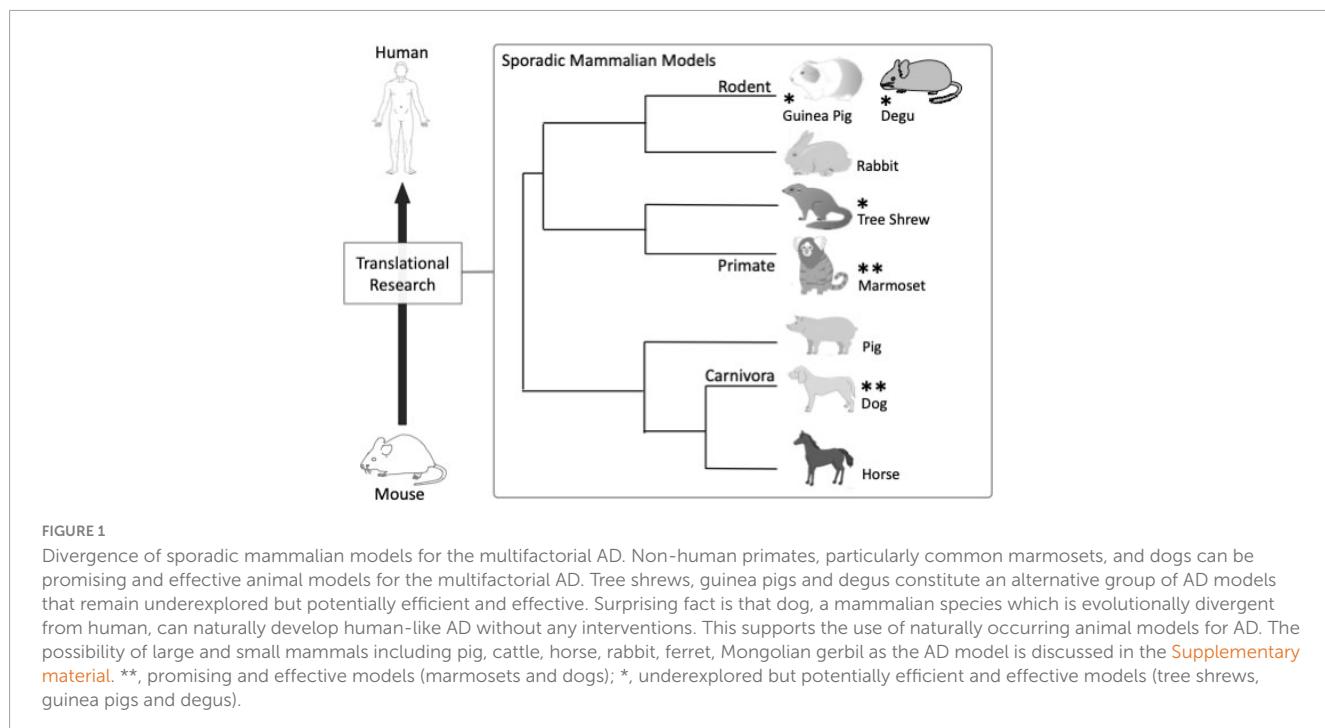


TABLE 1 Comparison of human and selective mammalian models for efficient and effective Alzheimer's disease (AD) research.

Species		Body size* ¹	Lifespan (y: years)	Plaque (y: years)	Tau	Alpha-synuclein	Systemic abnormalities* ²	Diurnal
Human	(<i>Homo sapiens</i>)	Large	80 y	> 50–60 y	Yes	Yes	Yes	Yes
Marmoset	(<i>Callithrix jacchus</i>)	Small	12 y	> 7–10 y	Yes	Probable	Probable	Yes
Dog	(<i>Canis lupus familiaris</i>)	Middle	15 y	> 8–10 y	Yes	Probable	Probable	Yes
Tree shrew	(<i>Tupaia belangeri</i>)	Small	6–8 y	> 5–6 y ³	Yes	Probable	Possible	Yes
Guinea pig	(<i>Cavia porcellus</i>)	Small	5–7 y	> 4 y	Yes	Possible	Probable	Yes
Degu	(<i>Octodon degus</i>)	Small	5–7 y	> 3 y ⁴	Yes	ND	Probable	Possible
Rat	(<i>Rattus norvegicus</i>)	Small	2 y	ND	ND	ND	Possible ⁵	No
Mouse	(<i>Mus musculus</i>)	Tiny	2 y	ND	ND	ND	Possible ⁵	No

Body size, lifespan and the timing of plaque deposition are associated with the efficiency. Others are related to the effectiveness for the multifactorial Alzheimer's disease (AD) regarding the main pathology (plaque and tau), co-pathology, systemic alterations and diurnality. Yes/No/Probable/Possible is determined by their natural and wild situation, at most with diet/metabolic supplementation (without any drug-induced and/or genetic manipulation). ND means "not detected." *1 Body size: tiny (< 100 g), small (< 1 kg), middle (< 30 kg), and large (> 30 kg). *2 "Probable/Possible" refers possibility of direct alterations in peripheral tissues, blood-brain-barrier (BBB) disruption and related diseases such as cerebral amyloid angiopathy (CAA) and epilepsy that are associated with disrupted peripheral-central homeostasis. *3 In tree shrews, the presence of the amyloid plaque is thought to be relatively rare. *4 Degus with APOE4 allele and wild-captured or early generations after the wild-capture may frequently develop the amyloid pathology. *5 BBB characteristic and dysfunction with aging have been studied in mice (Poduslo et al., 2001; Daneman et al., 2010; Braniste et al., 2014; Goodall et al., 2018) and rats (Yang et al., 1994; Enerson and Drewes, 2006; Hawkins et al., 2007).

(~300 g body weight in wild), are an emerging model for AD and other neurodegenerative diseases (Rizzo et al., 2023; Pérez-Cruz and Rodríguez-Callejas, 2023; Huhe et al., 2025). Marmosets are considered to be aged in 8–10 years and have average lifespan of 12 years (Table 1) (Pérez-Cruz and Rodríguez-Callejas, 2023). Sporadic amyloid plaques are observable as early as 7 years (Rizzo et al., 2023), and aged marmosets exhibit both 3-repeat (3R)

and 4-repeat (4R) tau isoforms in their brain, implying highly toxic patterns of tau expressions similar to human (Huhe et al., 2025). Aggregation of α-synuclein in the marmoset brain and body is thought to be possible (Shimozawa et al., 2017): the natural aggregation of α-synuclein in the olfactory bulb of 6 years old marmoset (without injections of toxic seeds) has been reported (Kobayashi et al., 2016). Also, colitis may be associated with

the alteration in α -synuclein expression and phosphorylation in the myenteric plexus of marmosets (Resnikoff et al., 2019). CCA pathology (Rizzo et al., 2023) and epilepsy (Yang et al., 2022) which are associated with BBB disruption are naturally observed in marmosets: direct BBB characteristics of marmosets have also been well-studied (Hoshi et al., 2013; Parks et al., 2023). The marmoset is diurnal, and the sleep and circadian rhythm of marmosets has been well-studied (Erkert, 1989; Crofts et al., 2001; Koshiba et al., 2021; Bukhtiyarova et al., 2022).

3.2 Companion animal (divergent from primates): dogs

Dogs, a Carnivora species, are another candidate of promising and effective models for AD (Cotman and Head, 2008; Prpar Mihevc and Majdič, 2019; Ambrosini et al., 2019). Dog is evolutionarily divergent from human in mammals (more distant than rodents; Figure 1), but can naturally develop AD-like disorder without any interventions. Severe cognitive decline can be observed in aged dog: it is referred as CCD. Aged dogs can naturally exhibit both amyloid plaque (over 8 to 10 years-old) and tau pathology (Schmidt et al., 2015; Ozawa et al., 2016; Smolek et al., 2016; Table 1), and, very surprisingly, present symptoms exactly like human such as disorientation (Osella et al., 2007). Accumulation of α -synuclein has been presented in spinal cord and hippocampus of 10–12 years old beagle dogs (Ahn et al., 2012; Ahn et al., 2013), although it may be not very frequent and/or breed-genetical specific (Uchida et al., 2003). CAA including micro-bleeding in the brain and epilepsy (probable BBB disruption) seems relatively frequent in aged dogs (Wisniewski et al., 1996; Ozawa et al., 2016; Nešić et al., 2017). Direct association studies between epilepsy and BBB dysfunction in dogs have been reported (Hanael et al., 2019; Hanael et al., 2024). Sleep-circadian cycle has been widely studied in dogs (Adams and Johnson, 1993; Bódizs et al., 2020; Reicher et al., 2021).

3.3 Close to primates: tree shrews

Tree shrews (northern tree shrew; *Tupaia belangeri*), a species in the order Scandentia and widely distributed in South and Southeast Asia, are a possible efficient and effective model for AD (Fan et al., 2018; Li et al., 2023). The advantages of tree shrew as model animals are a small body weight (100–150 g), a short lifespan (6–8 years) and low maintenance costs (Table 1). Tree shrew has a much closer genetic and physiological affinities to primates than those of rodents (Figure 1) and has been used as models for basic science and many types of diseases including brain development, infection (particularly hepatitis viruses), depression, social stress and aging (Fan et al., 2018; Li et al., 2023; Yao et al., 2024). A β aggregates, plaque-like structures and increased phosphorylated tau protein have been detected in the brain of 5–6 years-old tree shrews (Yamashita et al., 2012; Fan et al., 2018; Li et al., 2023), although the plaque deposition may be rare (Pawlak et al., 1999; Yamashita et al., 2010; Li et al., 2023). The α -synuclein protein sequence of tree shrew is 97.1% identical to that of human, implying the tree shrew's α -synuclein might have similar functions compared to human (Wu et al., 2015). A higher expression and aggregates

of α -synuclein has been observed in the brain of tree shrews (Wu et al., 2019). Although CAA, epilepsy and related BBB leakages have not been reported, gut-to-brain axis in cognition (Guo et al., 2021; Wang et al., 2023) and circadian rhythm (Meijer et al., 1990; Legros et al., 2007; Coolen et al., 2012; Luo et al., 2020; Dimanico et al., 2021) of tree shrews has been studied well.

3.4 Rodents: guinea pigs and degus

Rodents other than mice and rats can be other candidates of the efficient and effective models for AD. The order Rodentia (rodents) is divided into three suborders: Sciuroomorpha (squirrel-like) and Myomorpha (mouse and rat-like), and Hystricomorph (porcupine-like). In particular, certain hystricomorph rodents, including guinea pigs (*Cavia porcellus*: Sharman et al., 2013; Wahl et al., 2022) and degus (*Octodon degus*: Inestrosa et al., 2005; Hurley et al., 2018), are emerging AD model rodents. A relatively smaller body size and a shorter lifespan of such rodents than other mammalian models suggest their potential as one of the most efficient models for AD. Moreover, such rodents can be effective and are expected to bridge the translational gap between mouse to human, since they are rodents like mice and rats, but have potential to naturally onset AD.

The guinea pig, a hystricomorph rodent with an average lifespan of 5–7 years and a body weight of 700 to 1,000 g, is an emerging sporadic model for AD (Sharman et al., 2013; Wahl et al., 2022; Table 1). Guinea pigs have been used in research for over 200 years (Harkness et al., 2002), including more recent studies of cerebral cortices (Hatakeyama et al., 2017), infectious diseases (Connolly et al., 1999; Lowen et al., 2006; Padilla-Carlin et al., 2008) and pharmacological, environmental, and dietary interventions (Rakic et al., 1989; Kim et al., 2017; Li et al., 2021). Guinea pigs have the identical A β ₄₂ sequence to human (Salazar et al., 2016) and express both 3R and 4R tau isoforms (Sharman et al., 2013). The A β aggregates (Wahl et al., 2022) and plaques (Bates et al., 2014) can be observed at over 1 and 4 years old, respectively. Although this study has not found any report of the aggregates and/or deposition of α -synuclein in the brain of guinea pigs, the aggregation has been observed in the gut (Sharrad et al., 2013; Sharrad et al., 2017): notice that two possible pathways (brain-first and body (gut)-first) for the synuclein spreading have been reported (Borghammer and Van Den Berg, 2019; Nuzum et al., 2022). The BBB of guinea pigs has been well-studied (Rakic et al., 1989; Uva et al., 2008), including the transport of A β at BBB (Martel et al., 1996). No CAA researches were found in this study. Pharmacologically-induced epilepsy in guinea pig have been widely investigated, and the association between epilepsy and BBB permeability using induced-epilepsy model of the guinea pig has been studied (Uva et al., 2008). Also a gut-to-brain (microbiome-hypothalamus) axis in guinea pigs has been investigated (Li et al., 2021; Nuzum et al., 2022). Guinea pigs are diurnal, and extensive studies have been reported related to sleep and circadian system in guinea pigs (Kurumiya and Kawamura, 1988; Akita et al., 2001; Liu et al., 2020).

The degu, another hystricomorph rodent from central Chile with an average lifespan of 5 to 7 years and a body weight of less than 300 g, has the potential as one of the most efficient and effective mammalian models for AD (Inestrosa et al., 2005;

Cisternas et al., 2018; Hurley et al., 2018; Tan et al., 2022; Table 1). Degus are highly social (Rivera et al., 2016) and thought to have advanced cognitive abilities (Kumazawa-Manita et al., 2013), although they are a small rodent. Degus are the emerging candidate of multimorbidity-systemic models, since recent studies have reported that degus naturally develop visual impairments (Datiles and Fukui, 1989; Szabadfi et al., 2015; Hurley et al., 2018), endocrinological and metabolic dysfunctions including diabetes (Datiles and Fukui, 1989; Rivera et al., 2018; Hurley et al., 2018) and neoplasia (Anderson et al., 1990; Lester et al., 2005; Švara et al., 2020; Ikeda et al., 2024). Interestingly, related to AD pathology, degus spontaneously represent the accumulation of A β and phosphorylated tau in the brain: it may start between 1 and 3 years old (Ardiles et al., 2012). The amyloid plaque can be observed at 3–5 years old (Cisternas et al., 2018). Although some studies have reported contradictory results regarding the potential of degus as a model for AD research (Steffen et al., 2016; Bourdenx et al., 2017), degus with the higher risk APOE4 allele and wild-captured or early generations after the wild-capture may frequently develop the AD pathology (Hurley et al., 2022). No α -synuclein researches in degus were found in this study. However, degus can be one of the best (efficient and effective) models for investigating systemic alterations in AD, because they can naturally exhibit CAA (van Groen et al., 2011) and epilepsy (Ikai et al., 2021) (like marmosets and dogs), implying a higher risk for BBB dysfunction. Degus can be diurnal (Kas and Edgar, 1999a; Lee, 2004; Bonmati-Carrion et al., 2017), and sleep and circadian functions including the association between sleep deprivation and cognitive decline have been studied well (Kas and Edgar, 1999b; Ocampo-Garcés et al., 2013; Tarragon et al., 2014; Estrada et al., 2015).

4 Conclusion and perspective

Interestingly, many mammalian species spontaneously exhibit the amyloid plaques as they age (McKean et al., 2021; Sharma et al., 2023; Ferrer, 2024; Supplementary material). This study updated the information of mammalian models for the multifactorial AD, including co-pathology and systemic alterations. However, it was impossible to investigate prevalence and frequency of such new issues for relatively underexplored animals. This study was based on case reports of mammals. This is a limitation of this review.

Although further investigation is required, NHPs, particularly marmosets, and dogs are candidates of promising and effective AD models. Previous studies have showed that marmosets and dogs can present the amyloid plaque at 7–10 years old. Tree shrews, guinea pigs and degus constitute an alternative group of AD models that remain underexplored but potentially efficient and effective. Particularly, in guinea pigs and degus, the amyloid plaque can be detected in 3–4 years: this seems the earliest (the most efficient) so far in naturally onset mammals. However the possibility of co-pathology and systemic alterations in the three species warrants further investigation for more robust and effective AD models.

Emerging evidences suggest that the key progression pathways of the multifactorial AD may be fundamentally conserved in several mammalian species. The efficient and effective mammalian model provides opportunities to investigate the long-term and spatio (systemic)-temporal observations that are potentially crucial in

current AD research but are highly resource-intensive and time-consuming to implement in epidemiological studies of NHPs (except marmosets) and human. The mammalian models in this study, together with hypothesis-driven mouse models and also advances in data science technologies including omics and imaging analyses, may bridge the translational gap between mouse and human and lead to breakthroughs in AD research.

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Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fnagi.2025.1652754/full#supplementary-material>

References

- Adams, G. J., and Johnson, K. G. (1993). Sleep-wake cycles and other night-time behaviours of the domestic dog (*Canis familiaris*). *Appl. Anim. Behav. Sci.* 36, 233–248. doi: 10.1016/0168-1591(93)90013-F
- Ahn, J. H., Choi, J. H., Park, J. H., Yan, B. C., Kim, I. H., Lee, J. C., et al. (2012). Comparison of alpha-synuclein immunoreactivity in the spinal cord between the adult and aged beagle dog. *Lab. Anim. Res.* 28, 165–170. doi: 10.5625/lar.2012.28.3.165
- Ahn, J. H., Park, J. H., Yan, B. C., Lee, J. C., Choi, J. H., Lee, C. H., et al. (2013). Comparison of alpha-synuclein immunoreactivity in the hippocampus between the adult and aged beagle dogs. *Cell. Mol. Neurobiol.* 33, 75–84. doi: 10.1007/s10571-012-9873-8
- Akita, M., Ishii, K., Kuwahara, M., and Tsubone, H. (2001). The daily pattern of heart rate, body temperature, and locomotor activity in guinea pigs. *Exp. Anim.* 50, 409–415. doi: 10.1538/expanim.50.409
- Ambrosini, Y. M., Borchering, D. C., Kanthasamy, A. G., Kim, H., Willette, A. A., Jergens, A. E., et al. (2019). The gut–brain axis in neurodegenerative diseases and relevance of the canine model: A review. *Front. Aging Neurosci.* 11:130. doi: 10.3389/fnagi.2019.00130
- Anderson, W. I., Steinberg, H., and King, J. M. (1990). Bronchioalveolar carcinoma with renal and hepatic metastases in a degu (*Octodon degus*). *J. Wildl. Dis.* 26, 129–131. doi: 10.7589/0090-3558-26.1.129
- Ardiles, Á. O., Tapia-Rojas, C. C., Mandal, M., Alexandre, F., Kirkwood, A., Inestrosa, N. C., et al. (2012). Postsynaptic dysfunction is associated with spatial and object recognition memory loss in a natural model of Alzheimer's disease. *Proc. Natl. Acad. Sci. U.S.A.* 109, 13835–13840. doi: 10.1073/pnas.1201209109
- Arnold, S. E., Arvanitakis, Z., Macauley-Rambach, S. L., Koenig, A. M., Wang, H. Y., Ahima, R. S., et al. (2018). Brain insulin resistance in type 2 diabetes and Alzheimer disease: Concepts and conundrums. *Nat. Rev. Neurol.* 14, 168–181. doi: 10.1038/nrneuro.2017.185
- Bates, K., Vink, R., Martins, R., Harvey, A., and Morganti-Kossmann, M. C. (2014). Aging, cortical injury and Alzheimer's disease-like pathology in the guinea pig brain. *Neurobiol. Aging* 35, 1345–1351. doi: 10.1016/j.neurobiolaging.2013.11.020
- Bloom, G. S. (2014). Amyloid- β and tau: The trigger and bullet in Alzheimer disease pathogenesis. *JAMA Neurol.* 71, 505–508. doi: 10.1001/jamaneurol.2013.5847
- Böldizs, R., Kis, A., Gácsi, M., and Topál, J. (2020). Sleep in the dog: comparative, behavioral and translational relevance. *Curr. Opin. Behav. Sci.* 33, 25–33. doi: 10.1016/j.cobeha.2019.12.006
- Bonmatí-Carrion, M. A., Baño-Otalora, B., Madrid, J. A., and Rol, M. A. (2017). Light color importance for circadian entrainment in a diurnal (*Octodon degus*), and a nocturnal (*Rattus norvegicus*) rodent. *Sci. Rep.* 7:8846. doi: 10.1038/s41598-017-08691-7
- Borghammer, P. and Van Den Berge, N. (2019). Brain-first versus gut-first Parkinson's disease: a hypothesis. *J. Parkinsons Dis.* 9, S281–S295. doi: 10.3233/JPD-191721
- Bourdenx, M., Dovero, S., Thiolat, M. L., Bezard, E., and Dehay, B. (2017). Lack of spontaneous age-related brain pathology in *Octodon degus*: A reappraisal of the model. *Sci. Rep.* 7:45831. doi: 10.1038/srep45831
- Bowman, G. L., Kaye, J. A., Moore, M., Waichunas, D., Carlson, N. E., and Quinn, J. F. (2007). Blood–brain barrier impairment in Alzheimer disease: Stability and functional significance. *Neurology* 68, 1809–1814. doi: 10.1212/01.wnl.0000262031.18018.1a
- Braniste, V., Al-Asmakh, M., Kowal, C., Anuar, F., Abbaspour, A., Tóth, M. et al. (2014). The gut microbiota influences blood–brain barrier permeability in mice. *Sci. Transl. Med.* 6:263ra158. doi: 10.1126/scitranslmed.3009759
- Bukhtiyarova, O., Chauvette, S., Seigneur, J., and Timofeev, I. (2022). Brain states in freely behaving marmosets. *Sleep* 45:zsac106. doi: 10.1093/sleep/zsac106
- Buyssse, D. J. (2014). Sleep health: Can we define it? Does it matter? *Sleep* 37, 9–17. doi: 10.5665/sleep.3298
- Cai, Z., Qiao, P. F., Wan, C. Q., Cai, M., Zhou, N. K., and Li, Q. (2018). Role of blood–brain barrier in Alzheimer's disease. *J. Alzheimer's Dis.* 63, 1223–1234. doi: 10.3233/JAD-180098
- Carrano, A., Hoozemans, J. J. M., van der Vies, S. M., van Horssen, J., de Vries, H. E., and Rozemuller, A. J. M. (2012). Neuroinflammation and blood–brain barrier changes in capillary amyloid angiopathy. *Neurodegener. Dis.* 10, 329–331. doi: 10.1159/000335183
- Cheng, Y., Tian, D.-Y., and Wang, Y.-J. (2020). Peripheral clearance of brain-derived A β in Alzheimer's disease: Pathophysiology and therapeutic perspectives. *Transl. Neurodegener.* 9:16. doi: 10.1186/s40035-020-00195-1
- Cisternas, P., Zolezzi, J. M., Lindsay, C., Rivera, D. S., Martinez, A., Bozinovic, F., et al. (2018). New insights into the spontaneous human Alzheimer's disease-like model *Octodon degus*: Unraveling amyloid- β peptide aggregation and age-related amyloid pathology. *J. Alzheimer's Dis.* 66, 1145–1163. doi: 10.3233/JAD-180729
- Cline, E. N., Bicca, M. A., Viola, K. L., Klein, W. L., Watterson, D. M., and Rogers, J. (2018). The Amyloid- β oligomer hypothesis: Beginning of the third decade. *J. Alzheimer's Dis.* 64, S567–S610. doi: 10.3233/JAD-179941
- Connolly, B. M., Steele, K. E., Davis, K. J., Geisbert, T. W., Kell, W. M., Jaax, N. K., et al. (1999). Pathogenesis of experimental Ebola virus infection in guinea pigs. *J. Infect. Dis.* 179, S203–S217. doi: 10.1086/514305
- Coolen, A., Hoffmann, K., Barf, R. P., Fuchs, E., and Meerlo, P. (2012). Telemetric study of sleep architecture and sleep homeostasis in the day-active tree shrew *Tupaia belangeri*. *Sleep* 35, 879–888. doi: 10.5665/sleep.1894
- Cotman, C. W. and Head, E. (2008). The canine (dog) model of human aging and disease: Dietary, environmental and immunotherapy approaches. *J. Alzheimer's Dis.* 15, 685–707. doi: 10.3233/JAD-2008-15413
- Crofts, H. S., Wilson, S., Muggleton, N. G., Nutt, D. J., Scott, E. A., and Pearce, P. C. (2001). Investigation of the sleep electrocorticogram of the common marmoset (*Callithrix jacchus*), using radiotelemetry. *Clin. Neurophysiol.* 112, 2265–2273. doi: 10.1016/S1388-2457(01)00699-X
- Cuddapah, V. A., Zhang, S. L., and Sehgal, A. (2019). Regulation of the blood–brain barrier by circadian rhythms and sleep. *Trends Neurosci.* 42, 500–510. doi: 10.1016/j.tins.2019.05.001
- Daneman, R., Zhou, L., Agalliu, D., Cahoy, J. D., Kaushal, A., and Barres, B. A. (2010). The mouse blood–brain barrier transcriptome: A new resource for understanding the development and function of brain endothelial cells. *PLoS One* 5:e13741. doi: 10.1371/journal.pone.0013741
- Datiles, M. B. and Fukui, H. (1989). Cataract prevention in diabetic *Octodon degus* with Pfizer's sorbinil. *Curr. Eye Res.* 8, 233–237. doi: 10.3109/02713688908997564
- Dimanico, M. M., Klaassen, A. L., Wang, J., KAESER, M., Harvey, M., Rasch, B., et al. (2021). Aspects of tree shrew consolidated sleep structure resemble human sleep. *Commun. Biol.* 4:722. doi: 10.1038/s42003-021-02234-7
- Enerson, B. E. and Drewes, L. R. (2006). The rat blood–brain barrier transcriptome. *J. Cereb. Blood Flow Metab.* 26, 959–973. doi: 10.1038/sj.jcbfm.9600249
- Erkert, H. G. (1989). Characteristics of the circadian activity rhythm in common marmosets (*Callithrix jacchus*). *Am. J. Primatol.* 17, 271–286. doi: 10.1002/ajp.1350170403
- Estrada, C., López, D., Conesa, A., Fernández-Gómez, F. J., Gonzalez-Cuello, A., Toledo, F., et al. (2015). Cognitive impairment after sleep deprivation rescued by transcranial magnetic stimulation application in *Octodon degus*. *Neurotox. Res.* 28, 361–371. doi: 10.1007/s12640-015-9544-x
- Fan, Y., Luo, R., Su, L. Y., Xiang, Q., Yu, D., Xu, L., et al. (2018). Does the genetic feature of the Chinese tree shrew (*Tupaia belangeri chinensis*) support its potential as a viable model for Alzheimer's disease research? *J. Alzheimer's Dis.* 61, 1015–1028. doi: 10.3233/JAD-170594
- Ferrer, I. (2024). Alzheimer's disease neuropathological change in aged non-primate mammals. *Int. J. Mol. Sci.* 25:8118. doi: 10.3390/ijms25158118
- Gericke, C., Kirabali, T., Flury, R., Mallone, A., Rickenbach, C., Kulic, L., et al. (2023). Early β -amyloid accumulation in the brain is associated with peripheral T cell alterations. *Alzheimers Dement.* 19, 5642–5662. doi: 10.1002/alz.13136
- Goodall, E. F., Wang, C., Simpson, J. E., Baker, D. J., Drew, D. R., Heath, P. R., et al. (2018). Age-associated changes in the blood–brain barrier: Comparative studies in human and mouse. *Neuropathol. Appl. Neurobiol.* 44, 328–340. doi: 10.1111/nan.12408
- Greene, C., Hanley, N., Reschke, C. R., Reddy, A., Mäe, M. A., Connolly, R., et al. (2022). Microvascular stabilization via blood–brain barrier regulation prevents seizure activity. *Nat. Commun.* 13:2003. doi: 10.1038/s41467-022-29657-y
- Guo, Y., Wang, L., Lu, J., Jiao, J., Yang, Y., Zhao, H., et al. (2021). GinsenosideRg1 improves cognitive capability and affects the microbiota of large intestine of tree shrew model for Alzheimer's disease. *Mol. Med. Rep.* 23:291. doi: 10.3892/mmr.2021.11931
- Hanael, E., Baruch, S., Altman, R. K., Chai, O., Rapoport, K., Peery, D., et al. (2024). Blood–brain barrier dysfunction and decreased transcription of tight junction proteins in epileptic dogs. *J. Vet. Intern. Med.* 38, 2237–2248. doi: 10.1111/jvim.17099
- Hanael, E., Veksler, R., Friedman, A., Bar-Klein, G., Senatorov, V. V., Kaufer, D., et al. (2019). Blood–brain barrier dysfunction in canine epileptic seizures detected by dynamic contrast-enhanced magnetic resonance imaging. *Epilepsia* 60, 1005–1016. doi: 10.1111/epi.14739
- Hardy, J. and Selkoe, D. J. (2002). The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science* 297, 353–356. doi: 10.1126/science.1072994
- Harkness, J. E., Murray, K. A., and Wagner, J. E. (2002). Biology and diseases of guinea pigs. *Lab. Anim. Sci.* 203–246. doi: 10.1016/B978-012263951-7/50009-0
- Hatakeyama, J., Sato, H., and Shimamura, K. (2017). Developing guinea pig brain as a model for cortical folding. *Dev. Growth Differ.* 59, 286–301. doi: 10.1111/dgd.12371

- Hawkins, B. T., Lundein, T. F., Norwood, K. M., Brooks, H. L., and Egletton, R. D. (2007). Increased blood-brain barrier permeability and altered tight junctions in experimental diabetes in the rat: Contribution of hyperglycaemia and matrix metalloproteinases. *Diabetologia* 50, 202–211. doi: 10.1007/s00125-006-0485-z
- Horváth, A., Szűcs, A., Barcs, G., Noebels, J. L., and Kamondi, A. (2016). Epileptic seizures in Alzheimer disease: A review. *Alzheimer Dis. Assoc. Disord.* 30:186–192. doi: 10.1097/WAD.0000000000000134
- Hoshi, Y., Uchida, Y., Tachikawa, M., Inoue, T., Ohtsuki, S., and Terasaki, T. (2013). Quantitative atlas of blood-brain barrier transporters, receptors, and tight junction proteins in rats and common marmoset. *J. Pharm. Sci.* 102, 3343–3355. doi: 10.1002/jps.23575
- Huang, L. T., Zhang, C. P., Wang, Y. B., and Wang, J. H. (2022). Association of peripheral blood cell profile with Alzheimer's disease: A meta-analysis. *Front. Aging Neurosci.* 14:888946. doi: 10.3389/fnagi.2022.888946
- Huhe, H., Shapley, S. M., Duong, D. M., Wu, F., Ha, S. K., Choi, S. H., et al. (2025). Marmosets as model systems for the study of Alzheimer's disease and related dementias: Substantiation of physiological tau 3R and 4R isoform expression and phosphorylation. *Alzheimers Dement.* 21:1. doi: 10.1002/alz.14366
- Hurley, M. J., Deacon, R. M., Beyer, K., Ioannou, E., Ibáñez, A., Teeling, J. L., et al. (2018). The long-lived *Octodon degus* as a rodent drug discovery model for Alzheimer's and other age-related diseases. *Ageing Res. Rev.* 47, 19–35. doi: 10.1016/j.arr.2018.05.004
- Hurley, M. J., Urrea, C., Garduno, B. M., Bruno, A., Kimbell, A., Wilkinson, B., et al. (2022). Genome sequencing variations in the Octodon degus, an unconventional natural model of aging and Alzheimer's disease. *Front. Aging Neurosci.* 14:894994. doi: 10.3389/fnagi.2022.894994
- Husain, M. (2021). Sleep and neurodegenerative diseases. *Brain* 144, 695–696. doi: 10.1093/brain/awab031
- Ikai, Y., Shinohara, A., Nagura-Kato, G., Shichijo, H., and Koshimoto, C. (2021). Evaluation index of epileptiform-like seizures observed in common degu (*Octodon degus*). *Honyurui Kagaku*. 61, 3–11. doi: 10.11238/mammalianscience.61.3
- Ikeda, M., Kondo, H., Hamada, F., Yamashita, T., and Shibuya, H. (2024). Disseminated histiocytic sarcoma in a degu (*Octodon degus*). *J. Vet. Med. Sci.* 86, 529–532. doi: 10.1292/jvms.24-0081
- Inestrosa, N. C., Reyes, A. E., Chacón, M. A., Cerpa, W., Villalón, A., Montiel, J., et al. (2005). Human-like rodent amyloid- β -peptide determines Alzheimer pathology in aged wild-type *Octodon degu*. *Neurobiol. Aging* 26, 1023–1028. doi: 10.1016/j.neurobiolaging.2004.09.016
- Jackson, R. J., Hyman, B. T., and Serrano-Pozo, A. (2024). Multifaceted roles of APOE in Alzheimer disease. *Nat. Rev. Neurol.* 20, 457–474. doi: 10.1038/s41582-024-0988-2
- Ju, Y. E. S., Lucey, B. P., and Holtzman, D. M. (2014). Sleep and Alzheimer disease pathology—a bidirectional relationship. *Nat. Rev. Neurol.* 10, 115–119. doi: 10.1038/nrneurol.2013.269
- Kalaria, R. N. (1999). The blood-brain barrier and cerebrovascular pathology in Alzheimer disease. *Ann. N. Y. Acad. Sci.* 893, 113–125. doi: 10.1111/j.1749-6632.1999.tb07821.x
- Kas, M. J. H. and Edgar, D. M. (1999a). A nonphotic stimulus inverts the diurnal-nocturnal phase preference in *Octodon degus*. *J. Neurosci.* 19, 328–333. doi: 10.1523/JNEUROSCI.19-01-00328.1999
- Kas, M. J. H. and Edgar, D. M. (1999b). Circadian timed wakefulness at dawn opposes compensatory sleep responses after sleep deprivation in *Octodon degus*. *Sleep* 22, 1045–1053. doi: 10.1093/sleep/22.8.1045
- Kim, N. H., Park, J. H., Park, J. S., and Joung, Y. H. (2017). The effect of deoxycholic acid on secretion and motility in the rat and guinea pig large intestine. *J. Neurogastroenterol. Motil.* 23, 606–615. doi: 10.5056/jnm16201
- Kobayashi, R., Takahashi-Fujigasaki, J., Shiozawa, S., Hara-Miyauchi, C., Inoue, T., Okano, H. J., et al. (2016). α -Synuclein aggregation in the olfactory bulb of middle-aged common marmoset. *Neurosci. Res.* 106, 55–61. doi: 10.1016/j.neures.2015.11.006
- Koshiba, M., Watarai-Senoo, A., Karino, G., Ozawa, S., Kamei, Y., Honda, Y., et al. (2021). A susceptible period of photic day-night rhythm loss in common marmoset social behavior development. *Front. Behav. Neurosci.* 14:539411. doi: 10.3389/fnbeh.2020.539411
- Kumazawa-Manita, N., Hama, H., Miyawaki, A., and Iriki, A. (2013). Tool use specific adult neurogenesis and synaptogenesis in rodent (*Octodon degus*). hippocampus. *PLoS One* 8:e58649. doi: 10.1371/journal.pone.0058649
- Kurumiya, S. and Kawamura, H. (1988). Circadian oscillation of the multiple unit activity in the guinea pig suprachiasmatic nucleus. *J. Comp. Physiol. A* 162, 301–308. doi: 10.1007/BF00606118
- Landsberg, G., Mađarić, A., and Žilka, N. (2017). *Canine and feline dementia: molecular basis, diagnostics and therapy*. Cham: Springer International Publishing. doi: 10.1007/978-3-319-53219-6
- Lee, T. M. (2004). *Octodon degus*: A diurnal, social, and long-lived rodent. *ILAR J.* 45, 14–24. doi: 10.1093/ilar.45.1.14
- Legros, C., Chalivoy, S., Gabriel, C., Mocaer, E., Delagrange, P., Fuchs, E., et al. (2007). First evidence of melatonin receptors distribution in the suprachiasmatic nucleus of tree shrew brain. *Neuro Endocrinol. Lett.* 28, 267–273. doi: 10.1159/000112678
- Lester, P. A., Rush, H. G., and Sigler, R. E. (2005). Renal transitional cell carcinoma and choristoma in a degu (*Octodon degus*). *Contemp. Top. Lab Anim. Sci.* 44, 41–44. doi: 10.1093/ilarjournal/ilj040
- Li, H., Xiang, B. L., Li, X., Cong, L., Li, Y., Miao, Y., et al. (2023). Cognitive deficits and Alzheimer's disease-like pathologies in the aged Chinese tree shrew (*Tupaia belangeri chinensis*). *Mol. Neurobiol.* 61, 1892–1908. doi: 10.1007/s12035-023-03663-7
- Li, J., Zhu, S., Lv, Z., Dai, H., Wang, Z., Wei, Q., et al. (2021). Drinking water with saccharin sodium alters the microbiota-gut-hypothalamus axis in guinea pig. *Anim. 11:1875*. doi: 10.3390/ani11071875
- Liu, C. C., Zhao, J., Fu, Y., Inoue, Y., Ren, Y., Chen, Y., et al. (2022). Peripheral apoE4 enhances Alzheimer's pathology and impairs cognition by compromising cerebrovascular function. *Nat. Neurosci.* 25, 1020–1033. doi: 10.1038/s41593-022-01127-0
- Liu, W., Zhang, Y., Chen, Q., Liu, S., Xu, W., and Shang, W. et al. (2020). Melatonin alleviates glucose and lipid metabolism disorders in Guinea pigs caused by different artificial light rhythms. *J. Diabetes Res.* 2020:4927403. doi: 10.1155/2020/4927403
- Löscher, W. and Friedman, A. (2020). Structural, molecular, and functional alterations of the blood-brain barrier during epileptogenesis and epilepsy: A cause, consequence, or both? *Int. J. Mol. Sci.* 21:591. doi: 10.3390/ijms2120591
- Lowen A., Mubareka S., Tumpey T., García-Sastre A., Palese P. (2006). The guinea pig as a transmission model for human influenza viruses. *Proc. Natl. Acad. Sci. U.S.A.* 103, 9988–9992. doi: 10.1073/pnas.0604157103
- Luo, P. H., Shu, Y. M., Ni, R. J., Liu, Y. J., Zhou, J. N., and Yan, J. (2020). A characteristic expression pattern of core circadian genes in the diurnal tree shrew (*Tupaia belangeri*). *Neuroscience* 437, 145–160. doi: 10.1016/j.neuroscience.2020.04.027
- Magaki, S., Tang, Z., Tung, S., Williams, C. K., Lo, D., Yong, W. H., et al. (2018). The effects of cerebral amyloid angiopathy on integrity of the blood-brain barrier. *Neurobiol. Aging* 70, 70–77. doi: 10.1016/j.neurobiolaging.2018.06.004
- Malhotra, R. K. (2018). Neurodegenerative disorders and sleep. *Sleep Med. Clin.* 13, 63–70. doi: 10.1016/j.jsmc.2017.09.006
- Mander, B. A., Winer, J. R., Jagust, W. J., and Walker, M. P. (2017). Sleep and human aging. *Neuron* 94, 19–36. doi: 10.1016/j.neuron.2017.02.004
- Martel, C. L., Mackie, J. B., McComb, J. G., Ghiso, J., and Zlokovic, B. V. (1996). Blood-brain barrier uptake of the 40 and 42 amino acid sequences of circulating Alzheimer's amyloid beta in guinea pigs. *Neurosci. Lett.* 206, 157–160. doi: 10.1016/S0304-3940(96)12462-9
- McKean, N. E., Handley, R. R., and Snell, R. G. (2021). A review of the current mammalian models of Alzheimer's disease and challenges that need to be overcome. *Int. J. Mol. Sci.* 22:13168. doi: 10.3390/ijms222313168
- Meijer, J. H., Daan, S., Overkamp, G. J. F., and Hermann, P. M. (1990). The two-oscillator circadian system of tree shrews (*Tupaia belangeri*) and its response to light and dark pulses. *J. Biol. Rhythms* 5, 1–16. doi: 10.1177/074873049000500101
- Moir, R. D., Lathe, R., Rudolph, E., and Tanzi, R. E. (2018). The antimicrobial protection hypothesis of Alzheimer's disease. *Alzheimers Dement.* 14, 1602–1614. doi: 10.1016/j.jalz.2018.06.3040
- Montagne, A., Nation, D. A., Sagare, A. P., Barisano, G., Sweeney, M. D., Chakraborty, A., et al. (2020). APOE4 leads to blood-brain barrier dysfunction predicting cognitive decline. *Nature* 581, 71–76. doi: 10.1038/s41586-020-2247-3
- Musiek, E. S. and Holtzman, D. M. (2016). Mechanisms linking circadian clocks, sleep, and neurodegeneration. *Science* 354, 1004–1008. doi: 10.1126/science.aaa4968
- Nassan, M. and Videncovic, A. (2022). Circadian rhythms in neurodegenerative disorders. *Nat. Rev. Neurol.* 18:7–24. doi: 10.1038/s41582-021-00577-7
- Nešić, S., Kukolj, V., Marinković, D., Vučićević, I., and Jovanović, M. (2017). Histological and immunohistochemical characteristics of cerebral amyloid angiopathy in elderly dogs. *Vet. Q.* 37:1–7. doi: 10.1080/01652176.2016.1235301
- Nuzum, N. D., Loughman, A., Szymlek-Gay, E. A., Teo, W. P., Hendy, A. M., and Macpherson, H. (2022). To the gut microbiome and beyond: The brain-first or body-first hypothesis in Parkinson's disease. *Front. Microbiol.* 13:791213. doi: 10.3389/fmicb.2022.791213
- Ocampo-Garcés, A., Hernández, F., and Palacios, A. G. (2013). REM sleep phase preference in the crepuscular *Octodon degus* assessed by selective REM sleep deprivation. *Sleep* 36:1247–1256. doi: 10.5665/sleep.2896
- Osella, M. C., Re, G., Odore, R., Girardi, C., Badino, P., Barbero, R., et al. (2007). Canine cognitive dysfunction syndrome: Prevalence, clinical signs and treatment with a neuroprotective nutraceutical. *Appl. Anim. Behav. Sci.* 105, 297–310. doi: 10.1016/j.applanim.2006.11.007

- Ozawa, M., Chambers, J. K., Uchida, K., and Nakayama, H. (2016). The relation between canine cognitive dysfunction and age-related brain lesions. *J. Vet. Med. Sci.* 78, 997–1006. doi: 10.1292/jvms.15-0624
- Padilla-Carlin, D. J., McMurray, D. N., and Hickey, A. J. (2008). The guinea pig as a model of infectious diseases. *Comp. Med.* 58, 324–340. doi: 10.4149/CM_2008_058_4_324
- Palop, J. J. and Mucke, L. (2009). Epilepsy and cognitive impairments in Alzheimer disease: A network dysfunction perspective. *Arch. Neurol.* 66, 435–440. doi: 10.1001/Archneurol.2009.15
- Parks, T. V., Szuszupak, D., Choi, S. H., Alikaya, A., Mou, Y., Silva, A. C., et al. (2023). Noninvasive disruption of the blood-brain barrier in the marmoset monkey. *Commun. Biol.* 6:806. doi: 10.1038/s42003-023-05185-3
- Patke, A., Young, M. W., and Axelrod, S. (2020). Molecular mechanisms and physiological importance of circadian clocks. *Nat. Rev. Mol. Cell Biol.* 20, 521–537. doi: 10.1038/s41580-019-0179-2
- Pawlak, M., Fuchs, E., Walker, L. C., Levy, E., Silhol, S., and Calenda, A. (1999). Primate-like amyloid- β sequence but no cerebral amyloidosis in aged tree shrews. *Neurobiol. Aging* 20, 47–51. doi: 10.1016/S0197-4580(99)00017-2
- Pérez-Cruz, C. and Rodríguez-Callejas, J. D. (2023). The common marmoset as a model of neurodegeneration. *Trends Neurosci.* 46, 394–409. doi: 10.1016/j.tins.2023.02.002
- Poduslo, J. F., Curran, G. L., Wengenack, T. M., Malester, B., and Duff, K. (2001). Permeability of proteins at the blood-brain barrier in the normal adult mouse and double transgenic mouse model of Alzheimer's disease. *Neurobiol. Dis.* 8, 555–567. doi: 10.1006/nbdi.2001.9402
- Prpar Mihevc, S. and Majdič, G. (2019). Canine cognitive dysfunction and Alzheimer's disease – two facets of the same disease? *Front. Neurosci.* 13:604. doi: 10.3389/fnins.2019.00604
- Rakic, L. M., Zlokovic, B. V., Davson, H., Segal, M. B., Begley, D. J., Lipovac, M. N., et al. (1989). Chronic amphetamine intoxication and blood-brain barrier permeability to inert polar molecules studied in the vascularly perfused guinea pig brain. *J. Neurol. Sci.* 94, 41–50. doi: 10.1016/0022-510x(89)90216-5
- Reicher, V., Kis, A., Simor, P., Bodizs, R., and Gácsi, M. (2021). Interhemispheric asymmetry during NREM sleep in the dog. *Sci. Rep.* 11:18817. doi: 10.1038/s41598-021-98178-3
- Resnikoff, H., Metzger, J. M., Lopez, M., Bondarenko, V., Mejia, A., Simmons, H. A., et al. (2019). Colonic inflammation affects myenteric alpha-synuclein in nonhuman primates. *J. Inflamm. Res.* 12, 113–126. doi: 10.2147/JIR.S196552
- Rivera, D. S., Inestrosa, N. C., and Bozinovic, F. (2016). On cognitive ecology and the environmental factors that promote Alzheimer disease: Lessons from *Octodon degus* (Rodentia: Octodontidae). *Biol. Res.* 49:10. doi: 10.1186/s40659-016-0074-7
- Rivera, D. S., Lindsay, C. B., Codocedo, J. F., Carreño, L. E., Cabrera, D., Arrese, M. A., et al. (2018). Long-term, fructose-induced metabolic syndrome-like condition is associated with higher metabolism, reduced synaptic plasticity and cognitive impairment in *Octodon degus*. *Mol. Neurobiol.* 55, 9169–9187. doi: 10.1007/s12035-018-0969-0
- Rizzo, S. J., Homanics, G., Schaeffer, D. J., Schaeffer, L., Park, J. E., Olouoch, J., et al. (2023). Bridging the rodent to human translational gap: Marmosets as model systems for the study of Alzheimer's disease. *Alzheimers Dement. Transl. Res. Clin. Interv.* 9:e12417. doi: 10.1002/trc2.12417
- Robinson, J. L., Lee, E. B., Xie, S. X., Rennert, L., Suh, E., Bredenberg, C., et al. (2018). Neurodegenerative disease concomitant proteinopathies are prevalent, age-related and APOE4-associated. *Brain* 141, 2181–2193. doi: 10.1093/brain/awy146
- Salazar, C., Valdivia, G., Ardiles, A. O., Ewer, J., and Palacios, A. G. (2016). Genetic variants associated with neurodegenerative Alzheimer disease in natural models. *Biol. Res.* 49:14. doi: 10.1186/s40659-016-0072-9
- Scheltens, P., de Strooper, B., Kivipelto, M., Frisoni, G. B., Salloway, S., Van der Flier, W. M., et al. (2021). Seminar: Alzheimer's disease – pathophysiology, diagnosis, and treatments. *Lancet* 397, 1577–1590. doi: 10.1016/S0140-6736(20)32205-4
- Schmidt, F., Boltze, J., Jäger, C., Hofmann, S., Willems, N., Seeger, J., et al. (2015). Detection and quantification of β -amyloid, pyroglutamyl A β , and tau in aged canines. *J. Neuropathol. Exp. Neurol.* 74, 912–923. doi: 10.1097/NEN.0000000000000230
- Selkoe, D. J., and Hardy, J. (2016). The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol. Med.* 8, 595–608. doi: 10.15252/emmm.201606210
- Sengupta, U., and Kayed, R. (2022). Amyloid β , Tau, and α -Synuclein aggregates in the pathogenesis, prognosis, and therapeutics for neurodegenerative diseases. *Prog. Neurobiol.* 214:102270. doi: 10.1016/j.pneurobio.2022.102270
- Serrano-Pozo, A., Das, S., and Hyman, B. T. (2021). APOE and Alzheimer's disease: Advances in genetics, pathophysiology, and therapeutic approaches. *Lancet Neurol.* 20, 68–80. doi: 10.1016/S1474-4422(20)30412-9
- Sharma, H., Chang, K. A., Hulme, J., and An, S. S. A. (2023). Mammalian models in Alzheimer's research: An update. *Cells* 12:2459. doi: 10.3390/cells12202459
- Sharman, M. J., Moussavi Nik, S. H., Chen, M. M., Ong, D., Wijaya, L., Laws, S. M., et al. (2013). The guinea pig as a model for sporadic Alzheimer's disease (AD): The impact of cholesterol intake on expression of AD-related genes. *PLoS One* 8:e66235. doi: 10.1371/journal.pone.0066235
- Sharrad, D. F., Chen, B. N., Gai, W. P., Vaikath, N., El-Agnaf, O. M., and Brookes, S. J. H. (2017). Rotenone and elevated extracellular potassium concentration induce cell-specific fibrillation of α -synuclein in axons of cholinergic enteric neurons in the guinea-pig ileum. *Neurogastroenterol. Motil.* 29:e12985. doi: 10.1111/nmo.12985
- Sharrad, D. F., de Vries, E., and Brookes, S. J. H. (2013). Selective expression of α -synuclein-immunoreactivity in vesicular acetylcholine transporter-immunoreactive axons in the guinea pig rectum and human colon. *J. Comp. Neurol.* 521, 657–676. doi: 10.1002/cne.23198
- Shimozawa, A., Ono, M., Takahara, D., Tarutani, A., Imura, S., Masuda-Suzukake, M., et al. (2017). Propagation of pathological α -synuclein in marmoset brain. *Acta Neuropathol. Commun.* 5:12. doi: 10.1186/s40478-017-0413-0
- Sims-Robinson, C., Kim, B., Rosko, A. J., and Feldman, E. L. (2010). How does diabetes accelerate Alzheimer disease pathology? *Nat. Rev. Neurol.* 6, 551–559. doi: 10.1038/nrneurol.2010.130
- Smolek, T., Madari, A., Farbakova, J., Kandrac, O., Jadhav, S., Cente, M., et al. (2016).Tau hyperphosphorylation in synaptosomes and neuroinflammation are associated with canine cognitive impairment. *J. Comp. Neurol.* 524, 874–895. doi: 10.1002/cne.23877
- Stampfer, M. J. (2006). Cardiovascular disease and Alzheimer's disease: Common links. *J. Intern. Med.* 260, 211–223. doi: 10.1111/j.1365-2796.2006.01687.x
- Steffen, J., Krohn, M., Paarmann, K., Schwitlick, C., Brüning, T., Marreiros, R., et al. (2016). Revisiting rodent models: *Octodon degus* as Alzheimer's disease model? *Acta Neuropathol. Commun.* 4:91. doi: 10.1186/s40478-016-0363-y
- Stonebarger, G. A., Bimonte-Nelson, H. A., and Urbanski, H. F. (2021). The rhesus macaque as a translational model for neurodegeneration and Alzheimer's disease. *Front. Aging Neurosci.* 13:734173. doi: 10.3389/fnagi.2021.734173
- Švara, T., Gombač, M., Poli, A., Račnik, J., and Zadravec, M. (2020). Spontaneous tumors and non-neoplastic proliferative lesions in pet degus (*Octodon degus*). *Vet. Sci.* 7:32. doi: 10.3390/vetsci7010032
- Sweeney, M. D., Sagare, A. P., and Zlokovic, B. V. (2018). Blood-brain barrier breakdown in Alzheimer disease and other neurodegenerative disorders. *Nat. Rev. Neurol.* 14, 133–150. doi: 10.1038/nrneurol.2017.188
- Szabadi, K., Estrada, C., Fernandez-Villalba, E., Tarragon, E., Setalo, G., Izura, V., et al. (2015). Retinal aging in the diurnal Chilean rodent (*Octodon degus*): Histological, ultrastructural and neurochemical alterations of the vertical information processing pathway. *Front. Cell Neurosci.* 9:126. doi: 10.3389/fncel.2015.00126
- Takeda, S., Sato, N., Uchio-Yamada, K., Sawada, K., Kunieda, T., and Takeuchi, D. (2010). Diabetes-accelerated memory dysfunction via cerebrovascular inflammation and A β deposition in an Alzheimer mouse model with diabetes. *Proc. Natl. Acad. Sci. U. S. A.* 107, 7036–7041. doi: 10.1073/pnas.1000645107
- Tan, Z., Garduño, B. M., Aburto, P. F., Chen, L., Ha, N., Cogram, P., et al. (2022). Cognitively impaired aged *Octodon degus* recapitulate major neuropathological features of sporadic Alzheimer's disease. *Acta Neuropathol. Commun.* 10:182. doi: 10.1186/s40478-022-01481-x
- Tarragon, E., Lopez, D., Estrada, C., Gonzalez-Cuello, A., Ros, C. M., Lamberty, Y., et al. (2014). Memantine prevents reference and working memory impairment caused by sleep deprivation in both young and aged *Octodon degus*. *Neuropharmacology* 85, 206–214. doi: 10.1016/j.neuropharm.2014.05.023
- Tini, G., Scagliola, R., Monacelli, F., La Malfa, G., Porto, I., and Brunelli, C. (2020). Alzheimer's disease and cardiovascular disease: A particular association. *Cardiol. Res. Pract.* 2020:2617970. doi: 10.1155/2020/2617970
- Uchida, K., Kihara, N., Hashimoto, K., Nakayama, H., Yamaguchi, R., and Tateyama, S. (2003). Age-related histological changes in the canine substantia nigra. *J. Vet. Med. Sci.* 65, 179–185. doi: 10.1292/jvms.65.179
- Ueda, K., Fukushima, H., Masliah, E., Xia, Y., Iwai, A., Yoshimoto, M., et al. (1993). Molecular cloning of cDNA encoding an unrecognized component of amyloid in Alzheimer disease. *Proc. Natl. Acad. Sci. U. S. A.* 90, 11282–11286. doi: 10.1073/pnas.90.23.11282
- Ujiiie, M., Dickstein, D. L., Carlow, D. A., and Jefferies, W. A. (2003). Blood-brain barrier permeability precedes senile plaque formation in an Alzheimer disease model. *Microcirculation* 10, 463–470. doi: 10.1038/sj.mn.7800212
- Uva, L., Librizzi, L., Marchi, N., Noe, F., Bongiovanni, R., Vezzani, A., et al. (2008). Acute induction of epileptiform discharges by pilocarpine in the in vitro isolated guinea-pig brain requires enhancement of blood-brain barrier permeability. *Neuroscience* 151, 303–312. doi: 10.1016/j.neuroscience.2007.10.037
- van Erum, J., Van Dam, D., and De Deyn, P. P. (2018). Sleep and Alzheimer's disease: A pivotal role for the suprachiasmatic nucleus. *Sleep Med. Rev.* 40, 17–27. doi: 10.1016/j.smrv.2017.07.005
- van Groen, T., Kadish, I., Popović, N., Popović, M., Caballero-Bleda, M., Baño-Otálora, B., et al. (2011). Age-related brain pathology in *Octodon degus*: Blood vessel, white matter and Alzheimer-like pathology. *Neurobiol. Aging* 32, 1651–1661. doi: 10.1016/j.neurobiolaging.2009.10.008

- van Vliet, E. A., da Costa, Araújo, S., Redeker, S., van Schaik, R., Aronica, E., et al. (2007). Blood–brain barrier leakage may lead to progression of temporal lobe epilepsy. *Brain* 130, 521–534. doi: 10.1093/brain/awl318
- Vojtechova, I., Machacek, T., Kristofikova, Z., Stuchlik, A., and Petrasek, T. (2022). Infectious origin of Alzheimer’s disease: Amyloid beta as a component of brain antimicrobial immunity. *PLoS Pathog.* 18:e1010929. doi: 10.1371/journal.ppat.1010929
- Wahl, D., Moreno, J., Santangelo, K. S., Zhang, Q., Afzali, M. F., Walsh, M. A., et al. (2022). Nontransgenic guinea pig strains exhibit hallmarks of human brain aging and Alzheimer’s disease. *J. Gerontol. A Biol. Sci. Med. Sci.* 77, 1766–1774. doi: 10.1093/gerona/glac073
- Wang, J., Gu, B. J., Masters, C. L., and Wang, Y.-J. (2017). A systemic view of Alzheimer disease – insights from amyloid- β metabolism beyond the brain. *Nat. Rev. Neurol.* 13, 612–623. doi: 10.1038/nrneurol.2017.111
- Wang, L., Lu, J., Yang, Y., Zhao, Y., Wang, P., Jiao, J., et al. (2023). Mechanism of cognitive impairment induced by D-galactose and L-glutamate through gut–brain interaction in tree shrews. *Synapse* 77:e22274. doi: 10.1002/syn.22274
- Wisniewski, H. M., Becker, J. T., and Kowall, N. W. (1996). The origin of amyloid in cerebral vessels of aged dogs. *Brain Res.* 715, 151–157. doi: 10.1016/0006-8993(95)01156-0
- Wu, Z. C., Gao, J. H., Du, T. F., Tang, D. H., Chen, N. H., Yuan, Y. H., et al. (2019). Alpha-synuclein is highly prone to distribution in the hippocampus and midbrain in tree shrews, and its fibrils seed Lewy body-like pathology in primary neurons. *Exp. Gerontol.* 116, 37–45. doi: 10.1016/j.exger.2018.12.008
- Wu, Z. C., Huang, Z. Q., Jiang, Q. F., Dai, J. J., Zhang, Y., Gao, J. H., et al. (2015). Human and tree shrew alpha-synuclein: Comparative cDNA sequence and protein structure analysis. *Appl. Biochem. Biotechnol.* 177, 957–966. doi: 10.1007/s12010-015-1789-6
- Xu, B., Lei, X., Yang, Y., Yu, J., Chen, J., Xu, Z., et al. (2025). Peripheral proteinopathy in neurodegenerative diseases. *Transl. Neurodegener.* 14:2. doi: 10.1186/s40035-024-00461-6
- Yamashita, A., Fuchs, E., Taira, M., and Hayashi, M. (2010). Amyloid beta (A β). protein- and amyloid precursor protein (APP)-immunoreactive structures in the brains of aged tree shrews. *Curr. Aging Sci.* 3, 230–238. doi: 10.2174/1874609811003030230
- Yamashita, A., Fuchs, E., Taira, M., Yamamoto, T., and Hayashi, M. (2012). Somatostatin-immunoreactive senile plaque-like structures in the frontal cortex and nucleus accumbens of aged tree shrews and Japanese macaques. *J. Med. Primatol.* 41, 147–157. doi: 10.1111/j.1600-0684.2012.00540.x
- Yamazaki, Y., Zhao, N., Caulfield, T. R., Liu, C.-C., and Bu, G. (2019). Apolipoprotein E and Alzheimer disease: Pathobiology and targeting strategies. *Nat. Rev. Neurol.* 15, 501–518. doi: 10.1038/s41582-019-0228-7
- Yang, G. Y., Betz, A. L., Chenevert, T. L., Brunberg, J. A., and Hoff, J. T. (1994). Experimental intracerebral hemorrhage: Relationship between brain edema, blood flow, and blood–brain barrier permeability in rats. *J. Neurosurg.* 81, 93–102. doi: 10.3171/jns.1994.81.1.0093
- Yang, X., Chen, Z., Wang, Z., He, G., Li, Z., Shi, Y., et al. (2022). A natural marmoset model of genetic generalized epilepsy. *Mol. Brain* 15:16. doi: 10.1186/s13041-022-00901-2
- Yao, Y.-G., Lu, L., Ni, R.-J., Bi, R., Chen, C., Chen, J.-Q., et al. (2024). Study of tree shrew biology and models: A booming and prosperous field for biomedical research. *Zool. Res.* 45, 877–909. doi: 10.24272/j.issn.2095-8137.2024.199