

Jan Johansson^{1,2*} and Anna Rising^{1,2*}

¹ Division for Neurogeriatrics, Department of Neurobiology, Care Sciences and Society (NVS), Center for Alzheimer Research, Karolinska Institutet, Huddinge, Sweden

² Department of Anatomy, Physiology and Biochemistry, Swedish University of Agricultural Sciences, Uppsala, Sweden

*Correspondence: janne.johansson@ki.se; anna.rising@slu.se

Edited by:

Janina Burk, University of Leipzig, Germany

Reviewed by:

Celeste Scotti, IRCCS Istituto Ortopedico Galeazzi, Italy Janina Burk, University of Leipzig, Germany

Keywords: biomaterial, scaffold, matrix, cell-binding motif, RGD, cell culture, compatibility

A commentary on

Recombinant spider silk with cell binding motifs for specific adherence of cells

by Widhe M, Johansson U, Hillerdahl CO, and Hedhammar M. Biomaterials (2013) **34**:8223–34. doi:10.1016/j. biomaterials.2013.07.058

The ideal scaffold for engineering and regeneration of tissues would be a replica of the extracellular matrix (ECM), which is unique for each tissue type. The scaffold should mimic the mechanical properties of the targeted tissue and serve as matrix for adhesion, growth, migration, and differentiation of endogenous and/or implanted cells. Recent research has highlighted the potential of targeting also the environment of the intermediate states that are formed during tissue repair, since progenitor cells that contribute to tissue formation in a regenerative niche exist in an environment that is different from the final tissue (e.g., the fracture callus that is formed during osteogenesis is softer than mature bone tissue) (Polo-Corrales et al., 2014). In addition, the scaffold should not evoke inappropriate immune responses and should be degradable. To improve cell interactions, ECM-derived cell-binding peptide motifs have been extensively used (Sengupta and Heilshorn, 2010; Maia et al., 2013).

For improving cell attachment to biomaterials, the RGD (Arg-Gly-Asp) peptide motif is commonly used. This peptide is found in fibronectin, vitronectin, bone sialoprotein (BSP), and collagen VI and is recognized by the $\alpha\nu\beta3$ integrin (Arnaout et al., 2005). The RGD motif exhibits its full binding activity to integrins only when its mobility is restricted in a loop conformation, which can be accomplished *in vitro* by the incorporation of RGD in a cyclic peptide structure (Kumagai et al., 1991; Mohri et al., 1991). In contrast, if the RGD motif is flexible and lacks a stable conformation, it has a much lower affinity to the $\alpha\nu\beta3$ integrin (Pfaff et al., 1994).

RGD-based peptides have been covalently linked to homopolymers (Brandley and Schnaar, 1988; Kuo and Lauffenburger, 1993; Cook et al., 1997; Bolley et al., 2013), but also covalently incorporated in proteins produced in heterologous hosts, including spider silk-derived proteins (Bini et al., 2006; Wohlrab et al., 2012). Artificial spider silk made from recombinant proteins can form various two- and threedimensional matrices that hold promise for culture of cells for tissue engineering (Bini et al., 2006; Wohlrab et al., 2012; Wu et al., 2014). These matrices are promising but to realize their full potential, they have to be assembled in a controlled and reproducible way. The recently determined physiological and molecular events that control spider silk formation (Andersson et al., 2014; Kronqvist et al., 2014) have made this realistic for the first time. Functionalization of the silk matrices with cell adhesion motifs via genetic engineering may be a great advantage of the material that likely allows incorporation of cues for adhesion of specific cells, but this has not been fully investigated.

Bini et al. compared films made from a recombinantly produced segment derived

from the repetitive part of major ampullate spidroin (MaSp) from Nephila clavipes with or without an RGD motif incorporated, and analyzed Ca²⁺ deposition as a measure of cell response and human mesenchymal stem (hMSC) growth (Bini et al., 2006). The authors found that the matrix containing RGD did not show enhanced function as regard cell outcomes and speculated that the lack of RGD effect may be caused by insufficient exposure on the films. Wohlrab et al. studied growth of the cell line BALB/3T3 derived from mouse fibroblasts on films made from a recombinant protein segment from MaSp2 from Araneus diadematus (Wohlrab et al., 2012). RGD was either genetically incorporated into the silk protein, or added by linking a cyclic RGD peptide to Cys residues incorporated into the MaSp-derived protein. In both cases, adhesion and proliferation was improved compared to the wildtype protein as well as to a variant in which genetically incorporated RGD was replaced with RGE, a control motif, which binds significantly weaker to integrins. Widhe et al. reported that genetic incorporation of RGD into artificial silk, based on the minispidroin 4RepCT derived from Euprosthenops australis MaSp1, improved adherence and growth of human primary fibroblasts compared to 4RepCT films. Incorporation of the control motif RGE was reported to result in decreased attachment and about 50% slower growth rate compared to the RGD-4RepCT (Widhe et al., 2013). However, growth curves for RGDand RGE-modified matrices were not analvzed.





Wu et al., in contrast, found that the RGD-4RepCT films did not improve attachment of human embryonic stem cells (hESC) compared to wild-type 4RepCT, while incorporation of RGD within the longer vitronectin-derived peptide motif PQVTRGDVFTM resulted in efficient hESC attachment and growth. The effect of the vitronectin-derived peptide motif required integrin binding to the RGD motif, as evidenced by a competition experiment (Wu et al., 2014). The differences between Wohlrab et al. and Widhe et al. on the one hand and Wu et al. on the other hand may depend on that hESC are demanding to culture in vitro, while cell lines and fibroblast are robust and will attach to and grow on a wide range of supports and matrices. To test this hypothesis, we performed the same experiments as in Widhe et al. but analyzed the growth curves for human primary fibroblasts on RGDand RGE-containing films. Surprisingly, we found that human fibroblasts grow equally well on RGD- and RGE-functionalized 4RepCT films (Figure 1).

The different results as regard the functional importance of RGD incorporated into spider silk-derived matrices for cell culture described above suggest that care must be taken when evaluating different silk-derived biomaterials. The cells used for culture must certainly be chosen with respect to intended eventual applications of the material, but the robustness of the cells, in particular their sensitivity to growth conditions, must also be considered. Cell types that are more insensitive to culture conditions, like immortalized cell lines or fibroblasts, may be unsuitable for critical testing of novel biomaterials. It is also important to compare matrices that are functionalized with motifs with presumed biological effects, like RGD, with negative controls, as indirect effects of incorporating novel peptide segments, e.g., mediated by altered general properties of the biomaterial, cannot be excluded.

REFERENCES

- Andersson, M., Chen, G., Otikovs, M., Landreh, M., Nordling, K., Kronqvist, N., et al. (2014). Carbonic anhydrase generates CO₂ and H+ that drive spider silk formation via opposite effects on the terminal domains. *PLoS Biol.* 12:e1001921. doi:10.1371/ journal.pbio.1001921
- Arnaout, M. A., Mahalingam, B., and Xiong, J. P. (2005). Integrin structure, allostery, and bidirectional signaling. *Annu. Rev. Cell Dev. Biol.* 21, 381–410. doi:10.1146/annurev.cellbio.21.090704. 151217
- Bini, E., Foo, C. W., Huang, J., Karageorgiou, V., Kitchel, B., and Kaplan, D. L. (2006). RGDfunctionalized bioengineered spider dragline silk biomaterial. *Biomacromolecules* 7, 3139–3145. doi: 10.1021/bm0607877
- Bolley, J., Lalatonne, Y., Haddad, O., Letourneur, D., Soussan, M., Perard-Viret, J., et al. (2013).

Optimized multimodal nanoplatforms for targeting alpha(v)beta3 integrins. *Nanoscale* 5, 11478–11489. doi:10.1039/c3nr03763k

- Brandley, B. K., and Schnaar, R. L. (1988). Covalent attachment of an Arg-Gly-Asp sequence peptide to derivatizable polyacrylamide surfaces: support of fibroblast adhesion and long-term growth. *Anal. Biochem.* 172, 270–278. doi:10.1016/0003-2697(88)90442-3
- Cook, A. D., Hrkach, J. S., Gao, N. N., Johnson, I. M., Pajvani, U. B., Cannizzaro, S. M., et al. (1997). Characterization and development of RGD-peptide-modified poly(lactic acid-co-lysine) as an interactive, resorbable biomaterial. *J. Biomed. Mater. Res.* 35, 513–523. doi:10.1002/(SICI)1097-4636(19970615)35: 4<513::AID-JBM11>3.0.CO;2-C
- Kronqvist, N., Otikovs, M., Chmyrov, V., Chen, G., Andersson, M., Nordling, K., et al. (2014). Sequential pH-driven dimerization and stabilization of the N-terminal domain enables rapid spider silk formation. *Nat. Commun.* 5, 3254. doi:10.1038/ ncomms4254
- Kumagai, H., Tajima, M., Ueno, Y., Giga-Hama, Y., and Ohba, M. (1991). Effect of cyclic RGD peptide on cell adhesion and tumor metastasis. *Biochem. Biophys. Res. Commun.* 177, 74–82. doi:10.1016/0006-291X(91)91950-H
- Kuo, S. C., and Lauffenburger, D. A. (1993). Relationship between receptor/ligand binding affinity and adhesion strength. *Biophys. J.* 65, 2191–2200. doi:10.1016/S0006-3495(93)81277-3
- Maia, F. R., Bidarra, S. J., Granja, P. L., and Barrias, C. C. (2013). Functionalization of biomaterials with small osteoinductive moieties. *Acta Biomater.* 9, 8773–8789. doi:10.1016/j.actbio.2013. 08.004
- Mohri, H., Hashimoto, Y., Ohba, M., Kumagai, H., and Ohkubo, T. (1991). Novel effect of cyclicization of the Arg-Gly-Asp-containing peptide on vitronectin binding to platelets. *Am. J. Hematol.* 37, 14–19. doi:10.1002/ajh.2830370105
- Pfaff, M., Tangemann, K., Muller, B., Gurrath, M., Muller, G., Kessler, H., et al. (1994). Selective recognition of cyclic RGD peptides of NMR defined conformation by alpha IIb beta 3, alpha V beta 3, and alpha 5 beta 1 integrins. *J. Biol. Chem.* 269, 20233–20238.
- Polo-Corrales, L., Latorre-Esteves, M., and Ramirez-Vick, J. E. (2014). Scaffold design for bone regeneration. J. Nanosci. Nanotechnol. 14, 15–56. doi:10. 1166/jnn.2014.9127
- Sengupta, D., and Heilshorn, S. C. (2010). Proteinengineered biomaterials: highly tunable tissue engineering scaffolds. *Tissue Eng. Part B Rev.* 16, 285–293. doi:10.1089/ten.teb.2009.0591
- Widhe, M., Johansson, U., Hillerdahl, C. O., and Hedhammar, M. (2013). Recombinant spider silk with cell binding motifs for specific adherence of cells. *Biomaterials* 34, 8223–8234. doi:10.1016/j. biomaterials.2013.07.058
- Wohlrab, S., Muller, S., Schmidt, A., Neubauer, S., Kessler, H., Leal-Egana, A., et al. (2012). Cell adhesion and proliferation on RGD-modified recombinant spider silk proteins. *Biomaterials* 33, 6650–6659. doi:10.1016/j.biomaterials.2012.05. 069
- Wu, S., Johansson, J., Damdimopoulou, P., Shahsavani, M., Falk, A., Hovatta, O., et al. (2014).

Spider silk for xeno-free long-term self-renewal and differentiation of human pluripotent stem cells. *Biomaterials* 35, 8496–8502. doi:10.1016/j. biomaterials.2014.06.039

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 24 September 2014; accepted: 22 October 2014; published online: 06 November 2014.

Citation: Johansson J and Rising A (2014) Evaluation of functionalized spider silk matrices: choice of cell types and controls are important for detecting specific effects. Front. Bioeng. Biotechnol. 2:50. doi: 10.3389/fbioe.2014.00050 This article was submitted to Tissue Engineering and Regenerative Medicine, a section of the journal Frontiers in Bioengineering and Biotechnology. Copyright © 2014 Johansson and Rising. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.