

Theranostic Nanomedicine for Malignant Gliomas

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Brain tumors mainly originate from glial cells and are classified as gliomas. Malignant gliomas represent an incurable disease; indeed, after surgery and chemotherapy, recurrence appears within a few months, and mortality has remained high in the last decades. This is mainly due to the heterogeneity of malignant gliomas, indicating that a single therapy is not effective for all patients. In this regard, the advent of theranostic nanomedicine, a combination of imaging and therapeutic agents, represents a strategic tool for the management of malignant brain tumors, allowing for the detection of therapies that are specific to the single patient and avoiding overdosing the non-responders. Here, recent theranostic nanomedicine approaches for glioma therapy are described.

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THERANOSTIC TECHNOLOGY

Theranostics is the combination of the two terms "Therapeutics" and "Diagnostics," referring to technologies that include both diagnostic and therapeutic applications (**Figure 1**). The interest in personalized medicine, and thus, theranostic approaches used for individualized diagnosis and treatment are gaining increasing attention (Sun, 2010; Kelkar and Reineke, 2011; Kievit and Zhang, 2011; Ahmed et al., 2012; Wang Y. et al., 2014). This technology allows us to save time and decrease costs but, notably, also allows us to contain side effects of a single strategy (Lammers et al., 2010), obtaining better patient outcomes (Duncan, 2003; Peer et al., 2007).

Nanoplatforms (NPS) are nanoparticles combined with drug and molecular imaging probes, including metal nanoparticles, polymer-drug conjugates, polymer micelles, liposomes, and dendrimers. NPS show several advantages over conventional formulations, allowing for the conjugation or entrapment of drugs (Peer et al., 2007). Nanoparticles (NPs) are complex drug delivery systems, which can be structurally divided into the internal layer (core), and external layer (shell). Nanodimensions ensure that nanoparticles are able to increase drug solubility, mitigate cytotoxicity, and improve drug pharmacokinetic profiles. The creation of nanoplatforms, combining drugs with molecular probes, increases the drug half-life in the circulatory system, and specifically, delivers anticancer drugs to target tissues, controlling the drug release through detectors responsive to different stimuli such as pH, temperature, light, ultrasound, and enzymatic activities, thus improving the delivering of the required drug concentration to the area of interest (Tong and Cheng, 2007). By means of improving the circulating half-life, nanomedicines can accumulate in tumors through the Enhanced Permeability and Retention (EPR) effect (Golombek et al., 2018). In the last years, it has been reported that EPR varies among mouse models and patients, between tumor types of the same origin, and also among tumors and metastases of the

same patient (Harrington et al., 2001; Tanaka et al., 2017), thus explaining the heterogeneous outcomes of nanomedicine clinical trials. To overcome this issue, efforts should be focused on the use of methods that can be employed to individualize and improve nanomedicine treatments.

Despite the advantages offered by the EPR effect, the passive targeting approach offers a limited benefit in the treatment of gliomas and other CNS disorders. In these situations, the BBB remains impenetrable for different nanostructures that tend to accumulate in off-target tissues that also have vasculature gaps, such as the liver, or lymph nodes (Nam et al., 2018).

Since theranostic approaches require the use of molecular imaging tools, a combination of drug delivery systems with imaging techniques, such as computed tomography (CT), magnetic resonance imaging (MRI), optical and ultrasound (US) imaging, positron emission tomography (PET), and single photon emission computed tomography (SPECT), are currently under study. All these imaging techniques, using sensible and specific probes, can, in fact, assess drug efficacy during the drug development procedures (Cai et al., 2006; Pysz et al., 2010; Ai, 2011; Ang et al., 2014), optimizing the right choice of imaging tools and agents (Jokerst and Gambhir, 2011) and defining the best combination for specific therapeutic applications.

MULTIFUNCTIONALIZED NANOPLATFORMS

Multifunctionalized NPS are promising therapeutic approaches in cancer therapy (**Table 1**). Indeed, they offer numerous advantages over conventional agents, including specific targets, the higher ability to solubilize hydrophobic or labile drugs, lower systemic toxicity (resulting in better pharmacokinetics and higher potential to image), and better treatment and prediction of a therapeutic response. NPS utilize nanostructures, such as nanoparticles, made from soluble or colloidal aqueous solutions

and with sizes ranging between 10 and 100 nm (Bhojani et al., 2010). The small size allows them to pass via blood capillaries and reach the specific tumor cells (Bhojani et al., 2010). They have the advantages of modifying the nature and the number of linkers on and within the surface of a nanoparticle and its dimensions, thus leading to the control of the loading/releasing of the entrapped or covalently linked drugs. The NPS can also ameliorate the efficacy of current drugs or tracers, triggering a selective delivery. Among NPS, those based on nanovesicles are also biocompatible, thus increasing the maximum tolerated dose of the drug with low toxicity. This leads to an increased concentration of the agent inside the tumor and a simultaneous decrease in side effects (Liong et al., 2008; Bhojani et al., 2010). The entrapment of the drug with nanoplatforms reduces the limit for the use of poorly soluble or poorly absorbed agents by encapsulating them in the matrix of the NPS during the design and synthetic processes. Furthermore, the encapsulation prevents premature degradation of drugs or inactivation during plasma transport. Being a multidelivery system represents one of the most advantageous characteristics of NPS. They can carry imaging tracers, targeting ligands, therapeutic agents, and "cloaking" agents that avoid interference with the immune system (reviewed in Bhojani et al., 2010; Mendes et al., 2018).

Cancer researchers have shown high interest in theranostic approaches, particularly to detect and develop a solid nanosystem strategy for cancer treatment and diagnosis that can be translated into clinical practice (Cole et al., 2011).

Typical examples of the design of biocompatible nanoplatforms used as theranostic agents are based on magnetic nanoparticles, polymers, vesicles nanoparticles, and dendrimers.

Magnetic nanoparticles have been prepared using, for example, IONPs (iron oxide nanoparticles) coated with a human serum albumin. This formulation is referred as a biocompatible material for a chemotherapeutic agent, photosensitizers, and NPS (reviewed in Choi et al., 2012). Polymeric conjugates for drug delivery, biodistribution, and drug efficiency were extensively



	Nanoplatforms	Biosafety	Size	Loading capacity	References
Magnetic Nanoparticle	DOX IONPS	Low toxicity	10– 50 nm	High	Choi et al., 2012
Polymer- conjugates	DOX-GEM GADOLINIUM HPMA	Low toxicity	20– 70 nm	Good	Vilos and Velasquez, 2012
Nanovesicles	β(CD) SPIO Polymeric micelles	Really low toxicity	10– 70 nm	High	Liong et al., 2008; Bhojan et al., 2010
Dendrimers	asODN MNP PAMAM	Potential toxicity	10– 40 nm	High	Pan et al., 2007

Different nanoplatforms and the respective biosafety, size, and loading capacity.

DOX-conjugated, doxorubicin; PAMAM, poly(amidoamine); IONPs, iron oxide nanoparticles; DOX-GEM, gemcitabine (GEM), and doxorubicin (DOX); HPMA, N-2hydroxypropylmethacrylamide; asODN, antisense oligodeoxynucleotides; MNPs, Magnetic Nanoparticles; SPIO, superparamagnetic iron oxide.

investigated (Vilos and Velasquez, 2012). Lammers et al. (2010) synthesized a simultaneous delivery of doxorubicin and gemcitabine, and they were labeled with a gadolinium HPMA (N-2-hydroxypropylmethacrylamide) copolymer to investigate the biodistribution of nanotheranostics using an MRI (Lammers et al., 2010). This investigation reported that tumor-targeted polymeric drug vectors could be utilized to deliver two different chemotherapeutic drugs to tumors concurrently (Lammers et al., 2010).

Polymeric micelles, nano core/shell structures constituted by amphiphilic copolymers, were thoroughly tested as theranostic carriers and imaging probes as well. The amphiphilic block copolymers captured the superparamagnetic iron oxide (SPIO) or Mn-SPIO nanoparticles and are employed for the MRI (Lu et al., 2009; Liu et al., 2011; Xie et al., 2011; Su et al., 2013). β-cyclodextrin (β-CD) has been successfully used to encapsulate SPIO nanoparticles and small molecule anticancer drugs (Su et al., 2013). Several multifunctional polymeric micelles for tumor drug delivery and distribution have been designed, with particular attention to the creation of a well-controlled nanostructure. The use of polymeric micelles is advantageous because they can entrap an elevated number of hydrophobic drugs and contrast agents, maintaining their hydrophilic feature as a carriers, compared to liposomes or soluble polymers. Polymeric micelles are recognized as multifunctional delivery systems that are able to maximize therapeutic efficacy (Vilos and Velasquez, 2012).

Liposomes are already approved by the FDA since they are able to incorporate drugs, such as chemotherapeutics (Al-Jamal and Kostarelos, 2011). Approved formulations are liposomal doxorubicin and pegylated liposomal doxorubicin, which show low toxicity, cardiac safety, and less alopecia, myelosuppression, nausea, and vomiting when compared to conventional anthracyclines. The difficulty in releasing the encapsulated drug in the target area is caused by a limit in the liposome system. To overcome this issue, new liposome systems have been designed that are able to induce a pH and temperature response or the activation of certain enzymes on liposome cavities, thus improving the drug release in the targeted area (Lindner and Hossann, 2010; Wang D. et al., 2014). Recently, multifunctional theranostic nanoplatforms, using contrast agents encapsulated with liposomes, have been developed for the simultaneous diagnosis of early stage of disease and drug delivery, utilizing liposomes that encapsulate contrast agents, resulting in the creation of multifunctional NPS (Kenny et al., 2011; Na et al., 2011; Petersen et al., 2012). A theranostic nanoparticles inside the liposomes has been developed (Fattahi et al., 2011). Thus, multifunctional theranostic liposomes are widely used in treatment and for the detection of diseases, and they represent a valid carrier to further improve the diagnostic and therapeutic efficacy.

Furthermore, dendrimers are gaining increasing importance in the theranostic field as they can, due to their size, encapsulate several drugs or imaging tracers with high efficiency. For instance, dendrimers can bind non-covalently or covalently to chemotherapeutic drugs, imaging agents, and other biologically active targeting moieties, such peptides, monoclonal antibodies, and folates (Boas as and Heegaard, 2004; Mintzer and Grinstaff, 2011; Lo et al., 2013). The characteristic structure of dendrimers can stabilize the hydrophobic nanoparticles through the ligand exchange reaction method. Recently, multifunctional doxorubicin (DOX)-conjugated poly(amidoamine) (PAMAM) dendrimers have been developed with a specific platform for targeted chemotherapy that uses pH to release the This multifunctional dendrimer drug to tumor cells. excellent biocompatibility, biodistribution, presented and satisfactory cancer imaging results (Chang et al., 2011). Dendrimers represent promising structures for functionalization and also for conjugation with drugs (chemotherapeutics and imaging tracers) and DNA/RNA (Pan et al., 2007; Merkel et al., 2010; Zottel et al., 2019).

GLIOMAS

Glioma is a common type of tumor arising from gliasupporting neurons. About 33% of all brain tumors are gliomas and show different malignancy and differentiation grades. Symptoms depend on the area of the brain affected and by the degree of malignancy; they include headaches, nausea, or vomiting, speech difficulties, vertigo, and motor alteration. In its advanced stages, seizures may be a common manifestation. Gliomas are classified on the basis of the glial type but also on the genetic signature that predicts the outcome and the response to treatment. Gliomas are classified, according to the World Health Organization, as astrocytoma, anaplastic astrocytoma and glioblastoma, oligodendrogliomas, ependymomas, and mixed gliomas (Wesseling and Capper, 2018). Glioblastoma (GB) multiforme is the most malignant and common (more than 60%) type of primary astrocytomas (Rock et al., 2012). Despite the modern therapies to treat GB, it is still a deadly disease with an extremely poor prognosis. Patients usually have a median survival of \sim 14–15 months from the diagnosis (Thakkar et al., 2014). The standard treatment for GB is the resection of the tumor by neurosurgery, followed by radiation, and chemotherapy administration. However, these therapies are often ineffective, having a high rate of recurrence and drug resistance over time, accompanied by severe neurological deterioration of the affected patient (Silantyev et al., 2019).

The surgical approach is often not efficient due to the frequent persistence of tumoral foci; this leads to the recurrence of the disease (Alphandéry et al., 2015) thanks to the high proliferative rate and invasive behavior of GB cells. In this regard, several studies have reported the crucial role of bulk removal in increasing life expectancy and patient outcome (Silantyev et al., 2019). However, even bulk removal is not completely efficient since it is, generally, followed by relapses. For these reasons, GB is considered a not treatable disease. Temozolomide (TMZ) is currently the gold standard treatment for GB. Its metabolites form a complex with alkyl guanine alkyl transferase (O6 MGMT- DNA repair protein), leading to DNA damage; however, some patients show resistance to TMZ. Thus, many studies have reported the efficacy of the combination of TMZ with different compounds, such as curcumin, resveratrol, O6benzylguanine, valproic acid, anti-epilectic drugs, interferon 1β, mesenchymal stem cells, and anti-malarial drugs [extensively reviewed in Bahadur et al. (2019)], with reduced resistance and increased treatment efficacy. In particular, it has been reported that the combined administration of bone marrowderived mesenchymal cells (MSCs), interferon β (IF- β), and TMZ significantly decreased tumor progression in vitro and increased the survival of patients following synergistic effects in vivo (Park et al., 2015). More recently, the simultaneous administration of the inducer of autophagy, sirolimus, the inhibitor of autophagy, Chloroquine, and TMZ on glioblastoma cells was investigated in order to obtain lysosome disruption and apoptotic death (Hsu et al., 2018). In the same way, several new molecules were proposed to enhance TMZ activity in glioblastoma both in vitro and in vivo (extensively reviewed in Bahadur et al., 2019).

The diagnostic tools to detect brain tumors are represented by imaging tests, mainly MRI, including different specialized MRI scan components, including functional MRI, perfusion MRI, and magnetic resonance spectroscopy. These tools help us to understand tumor size and to plan treatment. Other imaging exams may include PET, a computerized tomography (CT) scan, and a cerebral angiogram. Molecular testing of the tumor could also be recommended for the identification of specific proteins, genes, and other factors (i.e., tumor markers) distinctive to the tumor. Indeed, some biomarkers may help in determining a patient's prognosis, increasing the chance of recovery. For the final and definite diagnosis, a biopsy of the tumor's tissue is usually necessary in order for it to be analyzed by a pathologist (Piquer et al., 2014; Tandel et al., 2019).

The first occurrence in tumor transformation is not completely clarified. However, it seems that the genetic signature is different in grade II gliomas, astrocytoma, and

oligodendroglioma. All tumors initially show the same invasive phenotype, making it difficult to develop a unique therapy. Progression-associated genetic modifications target cell cyclecontrolling pathways and growth promoting, causing focal hypoxia, necrosis, and angiogenesis. Retinoblastoma protein (Rb) mutation was identified in 20% of malignant gliomas (Behin et al., 2003), although gliomas may also contain mutations in other molecules involved in Rb signaling, including cyclin-dependent kinase (CDK) and the cell cycle regulator cyclin-dependent kinase inhibitor 2A, multiple tumor suppressor 1 (p16^{INK4A}). Most of the anaplastic astrocytoma show homozygous mutation, deletion, and promoter hypermethylation in the INK4A/ARF locus that encodes two tumor suppressors [p16^{INK4a} and an alternate reading frame tumor suppressor, p14ARF (Yamanaka, 2008)]. Moreover, it has been shown that PDGF (plateletderived growth factor) and platelet-derived growth factor receptor (PDGFR) signaling are involved at the beginning of the progression from astrocytoma to GB. In fact, elevated levels of PDGFRα have been reported in all types of gliomas, particularly in GB. Also, gliomas induce the overexpression of other mitogens, including IGF-1 (Insulin like Growth factor) and EGF (Epidermal growth factor) as well (Wong et al., 1992; Chakravarti et al., 2002; Nicholas et al., 2006; Puputti et al., 2006; Newton, 2010). Their receptors are present as constitutively active mutant forms in gliomas (Wong et al., 1992), leading to the activation of numerous pathways, including PI3K/AKT PBK, phospholipase protein C, and RAS/mitogen-activated protein kinase. In turn, these pathways control invasion, cell proliferation, apoptosis, and differentiation processes (Schlessinger, 2000). A common alteration (20-40%) identified in glioblastoma that affects the PI3K-Akt pathway is the genetic loss or mutation of the tumor suppressor gene PTEN (Phosphatase and Tensin homolog deleted on chromosome ten). Indeed, PTEN is a key negative regulator of the PI3K/Akt pathway (Stambolic et al., 1998; Cantley and Neel, 1999). In addition, gliomas display the upregulation of angiogenic factors, such as the FGF (fibroblast growth factor), TGF (transforming growth factor), Interleukin 8 (IL-8), and Vascular-Endothelial Growth Factor (VEGF) (Benoy et al., 2004; Slettenaar and Wilson, 2006; Xiao et al., 2018). The combination of the genetic alteration of these factors triggers a malignant glioma with an aggressive phenotype and that is resistant to intensive therapies. In this tumorigenic process, glioma stem cells exert a leading role (Uchida et al., 2000; Gaya et al., 2002; Kondo et al., 2004; Gürsel et al., 2011). Since glioma stem cells are able to selfpropagate, in order to avoid recurrence, it is fundamental to target specifically them (Kroonen et al., 2008). The new possibility to isolate GBM stem cells allows for new therapeutic approaches, among which are gene replacement, knockdown, or silencing (Kroonen et al., 2008). Since each GB patient shows a peculiar molecular profile, the response at radioand chemotherapies is different. On this basis, different GB cell lines may show a different response to Cdk inhibitors (Caracciolo et al., 2012; Cimini et al., 2017).

GB, and other solid tumors as well, encounter metabolic reprogramming; thus, the tumor is able to survive in hypoxic conditions and sustain angiogenesis and hyperproliferation

(Kroemer and Pouyssegur, 2008; Tennant et al., 2009; Fidoamore et al., 2016; Antonosante et al., 2018).

In particular, tumor cells activate the glycolytic pathway, also in the presence of oxygen (Warburg effect) (Frezza and Gottlieb, 2009). Indeed, tumor cells exploit the glycolytic signaling intermediates for anabolic reactions (Gatenby and Gillies, 2004). Only the cells subjected to these alterations are able to survive in the tumor environment, suggesting the presence of a selection of those with the altered metabolic phenotype (Tennant et al., 2009). The progress in the genetic biology of gliomas, and the recent insertion of manipulable experimental models, allows for the development of effective targeted therapy.

TARGETED THERANOSTIC NANOPLATFORMS FOR BRAIN CANCER THERAPY AND IMAGING

The human brain is an extremely complicated organ, which simultaneously regulates and supervises several functions. Successful therapy in brain cancers is restricted because the administered therapeutic entity cannot reach the targeted area after systemic administration (Cheng et al., 2014), and this is the main obstacle for the transport of the therapeutic agents represented by the blood-brain barrier (BBB). The BBB consists of a physical barrier, composed by vascular endothelial cells, and is held together by tight junctions, transporters, receptors, enzymes, and the ATP-dependent, 170-kDa efflux pump Pglycoprotein (Sonali et al., 2016a,b,c). The BBB retains the passage of agents with a molecular weight >500 Da but also of the majority of small sized molecules (Wei et al., 2014; Agrawal et al., 2017a,b). ATP-binding P-gp at the same time exerts the efflux function for xenobiotics, and their strong expression inhibits the passage of substrates through the BBB. The majority of the chemotherapeutics are hydrophobic and larger in molecular size; thus, they cannot cross the BBB spontaneously. Also, chemotherapeutics are substrates of multidrug-resistant drug efflux pumps, which are active on both tumor vascular cells and the BBB (Zong et al., 2014).

Brain cancers are difficult to detect and treat during the primary stages. The diagnosis and the detection of the volume of the brain cancers are complex because an accumulation of extracellular fluid (Koo et al., 2006) surrounding the tumor region is generally present. Since the 1970s, the primary modality to treat brain cancer includes surgical resection and/or chemotherapy or radiotherapy (Koo et al., 2006).

Conventional diagnostic and therapeutic agents showed improper bio-distribution and modest pharmacokinetics, leading to insufficient dissemination into tumors (Muthu et al., 2014a,b). In addition, they are non-specific and can accumulate in healthy organs, resulting in high toxicity. To overcome these issues, the nanotheranostic approach could be very useful. Different effective nanotheranostics brain cancer therapies have been recognized, but they need further investigation (Lakka and Rao, 2008; Xie et al., 2010; Keunen et al., 2011; Fan et al., 2014; Nance et al., 2014; Arranja et al., 2017). For instance, nanoparticle-enhanced imaging of the CNS at the subcellular level localizes more precisely the intracranial neoplasms area (Figure 2) (Bhojani et al., 2010). Also, nanoparticle-enhanced neuroimaging is very useful to understand physiological processes, including apoptosis, ischemia, inflammation, cell differentiation, and mitosis, representing the main tool for further research studies in neurodegenerative diseases and stroke (reviewed in Mattei and Rehman, 2015). To study physiological processes, many microscopic and macroscopic imaging modalities have been established. Microscopic methods require the invasive harvesting of tissues and imaging by cell-based assays (i.e., for apoptosis TUNEL, Annexin V, and Caspase Substrate Based Assays) (Cen et al., 2008). Macroscopic imaging modalities, by contrast, visualize apoptosis in living subjects in non-invasive modality. To date, to study these physiological processes, various in vivo molecular imaging technologies have been used, including Radiolabeled Small Molecular Probes, optical imaging probes, MRI agents, and multiple-modality methods. Microscopic and macroscopic imaging strategies improved the understanding of various physiological processes, or pathologies in preclinical and clinical studies (Zeng et al., 2015).

Unfortunately, the BBB still represents a limitation for nanotheranostic delivery (Wilhelm and Krizbai, 2014). To facilitate in vitro studies of drug delivery to the brain, promising in vitro BBB models have been developed based on primary or immortalized cells or on the culture of brain endothelial cells (Wilhelm and Krizbai, 2014; Helms et al., 2016). Valid models can be obtained using primary porcine brain endothelial cells and rodent co-culture models, which are characterized by low paracellular permeability and functional efflux transporters, mimicking the in vivo physiological complexity of the in vivo BBB. These include triple co-culture (brain endothelial cells with pericytes and astrocytes), dynamic, and microfluidic models, but these models are not suitable for rapid analyses. Great efforts have been made to deliver diagnostic agents and drugs into the brain. Thanks to recent advances in BBB research, new approaches have been exploited. Strategies able to pass through the BBB and reach the brain include viral vectors (characterized by high gene transfection efficiency), exosomes, brain permeability enhancers, and nanoparticles (Dong, 2018).

For example, Pilkington et al., in one of these in vitro BBB models, tested the properties of chitosyme nanoparticulate structures on BBB integrity, analyzing the tight junction proteins (ZO-1, occludin) and effects on the extra cellular matrix (Pilkington et al., 2014). In additional studies, paclitaxel (PTX) were constructed with a cyclic Arg-Gly-Asp (RGD) peptide as a targeting ligand to pass through the BBB by a targeting method. The nanocarriers were tested on a 3D glioma spheroid of glioma cells grown on agarose and showed targeted accumulation into tumor spheroids and excellent infiltration compared with conventional nanocarriers, suggesting a potential use in therapeutic approaches (Jiang et al., 2013). The theranostic nanosystems are combined with the targeting agent that identifies definite targets of brain cancer cells and binds to and internalizes via a specific mechanism. Several nanomaterials, including Gold Nanoparticles (AuNPs) and Quantum Dots (QDs) have intrinsic



diagnostic/therapeutic properties (Muthu et al., 2014a; Sonali et al., 2018).

GOLD NANOPARTICLES

Gold Nanoparticles designed from gold cores represent a new system for theranostic systems (Table 2). They are biocompatible and are usually prepared as spheres, wire, rods, cubes, and cages. AuNPs, like other inorganic nanoparticles, trigger oxidative stress and subsequent cytotoxic effects. The spheroidal AuNPs ultraviolet (UV) absorption is at 520 nm, while the gold nanorods absorption is in the Infrared radiation (690-900 nm). These intrinsic optical characteristic allow AuNPs to be utilized as multifarious theranostic drugs for clinical applications (Xie et al., 2010; Kumar et al., 2013). AuNPs showed a diagnostic property, tunable core size, low toxicity, surface plasmon absorption and ease of fabrication, and light-scattering properties (Fan et al., 2014). AuNPs have been widely studied; for example, Melancon et al. formulated multi-utility gold-based nanoshells with optical and magnetic activities, which were additionally conjugated with targeting moiety and studied as an approach for head and neck cancer (Melancon et al., 2011). It has been shown that AuNPs improve the treatment of gliomas; for instance, the use of AuNPs-combined radiotherapy promoted long-term survival with respect to radiation therapy alone (Hainfeld et al., 2013; Joh et al., 2013). In another study, a theranostic system for cancer treatment, which was able to reduce the cytotoxic effect on normal cells, has been developed based on the use of gold nanoparticles surface-functionalized with a paclitaxel drug and biotin receptor. Two categories, AuNPs-4 and AuNPs-5, were investigated for their peculiar interaction with cancer cells. These nanoparticles were tested against the immortalized NIH3T3 cells, and it was suggested that the AuNPs-5 was more efficient (Heo et al., 2012). In addition, AuNPs represent an encouraging candidate for tumor margin detection, improving the surgery resection of brain cancers (Tzeng and Green, 2013). An in-vitro study on brain tumor cell lines showed a strong amelioration in uptake studies of targeted particles with respect to non-targeted formulations (Dixit et al., 2015b). Recently, matrix metalloproteinase-2sensitive gold-gelatin nanoparticles were developed; RGD and octarginine were used as targeting ligands to pass through the BBB, allowing a pH-triggered release to the glioma-specific area. Indeed, it has been reported that gold nanotheranostic

TABLE 2 | Nanoparticles examples with some characteristics.

Nanoparticles	Biosafety	Size	References
Gold nanoparticles	Low cytotoxicity	2–60 nm	Fan et al., 2014
Magnetic nanoparticles	Potential cytotoxicity	7–20 nm	Alphandéry et al., 2015
Quantum dots	Potential cytotoxicity	2–50 nm	Onoshima et al., 2015
Carbon nanotubes	Potential cytotoxicity	0.4–40 nm	Wang et al., 2012
Mesoporous silica nanoparticles	Low cytotoxicity	20–25 nm	Wang et al., 2015

Different nanoparticles and the respective biosafety and size.

targeted specific tumor areas since it is able to co-localize within neovessels (Ruan et al., 2015). Gold theranostic micelles coated with polyethylene glycol-polycaprolactone (PEG-PCL) exhibited radiosensitizing efficacy for GBM therapy and can be used as a novel contrasting agent for both MRI and CT studies (Sun et al., 2016).

MAGNETIC NANOPARTICLES

Recently, Magnetic Nanoparticles (MNPs) have been introduced as potential nanocarriers in targeted drug delivery at the tumor area, having the further benefit of MRI traceability (Table 2). The magnetic response (iron oxide core) ameliorates the magnetic targeted delivery (Pankhurst et al., 2003; Frimpong and Hilt, 2010). Interestingly, it has been shown that intravenous administration of these particles is able to reach the cancer site in an animal model. Recently, Chertok et al. reported that magnetic nanoparticles are a useful tool for magnetically enhanced accumulation in brain tumors and for non-invasive MRI screening. This accumulation can be sharply improved with magnetic targeting, as confirmed by MRI (Chertok et al., 2008). A recent in-vivo study suggested the potential clinical application of these nanotheranostics since MNPs overpass the BBB in a reversible way, and the substance can reach the targeted site (Lammers et al., 2015; Tabatabaei et al., 2015). Since 2013, NanoTherm[®] therapy has been established as a new procedure for the focal treatment of solid tumors (Rivera Gil et al., 2010). In this procedure, magnetic nanoparticles are introduced in the tumor or in the resection cavity wall. These particles are then heated by an alternating magnetic field, determining cancer cells death. Nanoparticles are particles of iron oxide, suspended in water, with a diameter of about 15 nm. After the in vivo engraftment, they agglomerate and remain the tissue to be treated. An alternating magnetic field then induces the particles to generate heat. Depending on the temperatures reached in the tumor site or in individual remaining cancer cells in the resection cavity wall, and the length of treatment, cancer cells are destroyed or made more sensitive to concomitant radiotherapy or chemotherapy¹ (Alphandéry et al., 2015).

QUANTUM DOTS

Quantum Dots (QDs) are nanoscale (<10 nm) inorganic semiconductor nanocrystals, which represent a potential candidate for theranostic purposes (Table 2). They emit light which wavelength can be tuned on the basis of their shape, composition, and size. Cadmium selenide/Zinc sulfide-based QDs are the most used nanomaterials for diagnostic purposes. They have a CdSe core that is overcoated with layers of ZnS (Zhang et al., 2017). Furthermore, to gain affinity and target the cancer site, the surface of the QDs can be covalently or non-covalently conjugated with targeting probes, such as various antibodies, peptides, nucleic acids, folate aptamers, and other small molecules. One of the most suitable methods for conjugating targeting molecules on the surface of QDs is represented by the technique of avidin-biotin cross-linking (Tian et al., 2011; Onoshima et al., 2015). QDs can be conjugated with cancer cell-specific ligands, including HER2 (Ahmed et al., 2017), highly expressed in glioblastoma, CD44, proteins, antibodies, folic acid, and so on. Interestingly, QDs can be combined into paramagnetic liposomal designs containing RGD peptides and utilized as a diagnostic tool in tumor angiogenesis using MRI (Volkov, 2015). QDs in theranostic showed a clinical potential limit, due to their potential toxicity in humans (Derfus et al., 2004; Kirchner et al., 2005). To overcome this problem, further investigation is necessary to design biocompatible, excretable, surface-modified QDs (Onoshima et al., 2015).

CARBON NANOTUBES

Carbon Nanotubes (CNTs) are composed of different layers of graphene sheets, which form a cylindrical shape (Table 2). CNTs can be considered as allotropes of carbon with poor biocompatibility and slow biodegradation (Singh et al., 2016). CNTs are useful for theranostic applications since they can ameliorate the effect of chemotherapeutic agents and are translatable to clinical applications (Shapira et al., 2011; Singh et al., 2016). Once CNTs reach the targeted cells, they can interact with DNA and proteins, altering cellular signaling, or the mechanism of other therapeutic approaches (Ren et al., 2012; Chakrabarti et al., 2015). The intrinsic NIR light-absorption characteristic of CNTs has been exploited to eliminate tumor cells in vitro, whereas their NIR photoluminescence property has been utilized in cell imaging. In an interesting study, it has been reported that *i.v.* administration of single-walled carbon nanotubes (SWCNTs) as photo luminescent probes is a valid tool for in vivo tumor imaging, suggesting that SWCNTs could be used for theranostic applications. Moreover, CNTs are able to improve the chemotherapy effect in brain tumors (Robinson et al., 2010). In fact, recently, gold-coated surface-modified CNTs were established as optical nanotheranostic probes, which exhibited high potential as imaging tracers but had poor clinical potential due to slower biodegradation (and subsequent toxicity), as shown in in vivo nanotheranostic studies (Wang et al., 2012). However, CNTs may trigger adverse effects, such as lipid peroxidation, that induce inflammation and cell damage (Shapira et al., 2011; Singh et al., 2016).

 $^{^1}NanoTherm (\ensuremath{\mathbb{R}}\xspace$ therapy. Available online at: https://www.magforce.com/en/home/our_therapy/

MESOPOROUS SILICA NANOPARTICLES

Mesoporous Silica Nanoparticles (MSNPs) are also emerging drug delivery systems. MSNPs are thoroughly investigated and used in diagnostics because of their tunable shape and size and since their wide surface area facilitates a high drug loading (Table 2). Numerous drugs, including paclitaxel, camptothecin, methotrexate, colchicine, and cysteine, have been encapsulated in MSNPs. These encapsulated anticancer drugs are able to precisely cause the death of tumor cells (Gary-Bobo et al., 2012; Mamaeva et al., 2013). Thanks to the hexagonal structure, MSNPs can incorporate numerous functional components of an ideal theranostic approach in a single object, with different regions for the contrasting agent, therapeutic moiety, and biomolecular ligand. In addition, MSNPs are identified as safe materials by the FDA and are approved for evaluation in clinical studies. Scientists have utilized silica to integrate both IONPs and QDs, in order to create a hybrid with both optical and magnetic properties. MSNPs are biocompatible and biodegradable materials for nanotheranostic applications. MSNPs that dissolved silica can be absorbed by the biological system, metabolize, and be excreted through urine in the form of silicic acid or oligomeric silica species (Chen et al., 2013; Wang et al., 2015). Biomolecular targeting agents, proteins and peptides, are conjugated to the surface of MSNPs to ameliorate cancer treatment efficacy. Indeed, the surface of MSNPs was conjugated with a Tf peptide to enhance the detection of brain glioma cells (Cheng et al., 2010; Feng et al., 2016). Due to their efficient drug-loading capability, rugged nature, elevated biodegradation in the body, and diverse functionalization, MSNPs are widely used as tracers in MRIs or contrast agents in ultrasounds for accurate targeting, and they show positive results for brain cancer detection (Feng et al., 2016). In an interesting study, mesenchymal stem cells were engineered with MSNPs to treat and diagnose orthotopic glioblastomas. In particular, the intracerebral injection of engineered stem cells significantly improved the survival of rats with U87MG xenografts. This effect was concomitant with a reduction in tumor growth and proliferation and microvascular density. In GSC1 xenografts, intra-tumoral injections of AdhMSCs depleted the tumor cell population and induced migration of resident microglial cells (Pacioni et al., 2017). Nanotheranostics therapy was administered systemically to the mice and allowed in vivo imaging via MRI, NIR fluorescence, and PET; moreover, it exhibited high specificity for the glioma site (Huang et al., 2013).

NANOPARTICLES FOR GB TREATMENT

Numerous nanostructured delivery systems have been established for brain tumor delivery, and, on the basis of their composition and nature, they can be divided into organic NPS and inorganic NPS (Kumar et al., 2016; Di Martino et al., 2017). Organic NPS (i.e., liposomes, polymeric nanoparticles, lipid nanoparticles, and micelles), compared with the "free" drugs, were able to efficiently cross the BBB, with favored distribution in the brain, in both *in vitro* and *in vivo* studies (Danhier et al., 2015; Chen et al., 2016; Kuo and Cheng, $\ensuremath{\mathsf{TABLE 3}}\xspace$] Examples of clinical trials performed using nanoparticles drugs for gliomas.

Drugs	Diseases	Phase	Clinical Trial
ABI-009 (nab-rapamycin)	Recurrent high-grade glioma; Newly diagnosed glioblastoma	II	NCT03463265
NL CPT-11 (Nanoliposomal CPT-11)	Recurrent high-grade glioma	I completed	NCT00734682
Ferumoxytol	Recurrent high-grade glioma	I	NCT00769093
9-ING-41	Glioblastoma	П	NCT03678883
Pegylated Liposomal Doxorubicine + Temozolomide	Glioblastoma And diffuse intristic pontine glioma	II completed	NCT00944801
SGT-53	Recurrent glioblastoma	П	NCT02340156
Myocet	Refractory or relapsed malignant glioma in children/adolescent	I	NCT02861222

Drugs, disease, and clinical trials with relative phase.

2016; Liu et al., 2016; Qu et al., 2016; Wu et al., 2016; Belhadj et al., 2017; Chai et al., 2017; Graverini et al., 2018; Jhaveri et al., 2018) (Table 3). The main advantages of inorganic NPS (mesoporous silica nanoparticles, gold nanoparticles, iron oxide nanoparticles, and quantum dots) are their resistance to enzymatic degradation, robustness, and interesting intrinsic characteristics (Nam et al., 2013). For the treatment of GB, different kind of NPS (lipidic, magnetic, liposomal, fluorescent, and polymeric) have already been designed in order to cross the BBB, and these take into account active, passive, and stimuli-targeting perspectives (Cheng et al., 2014; Saraiva et al., 2016; Miranda et al., 2017; Aparicio-Blanco and Torres-Suárez, 2018). Theranostic nanoparticles represent a new technological concept that provides a combination of inorganic and organic nanoparticles to acquire synergistic characteristics in a single nanoparticle, exploiting the drug delivery by organic NPS and imaging by inorganic NPS. Theranostic nanoparticles can be used to limit toxicity due to a high and invasive dosage, improving patient outcomes. Recently, a combined chemo-photothermal targeted treatment of gliomas within one nanoparticle was developed. A targeting peptide was synthesized and characterized. In particular, as a therapeutic component, Doxorubicin was chosen, and, as a drug and diagnostic delivery system, a modified mesoporous silica-coated graphene nanosheet (GSPI) was chosen. The doxorubicin-loaded GSPI-based system showed heat-stimulating, pH-responsive, and sustained release properties. The in vitro results were encouraging; glioma cells showed a higher rate of death and strong GSPI accumulation (Lee et al., 2011). Targeting AuNPs with two or more receptor-binding peptides for glioblastoma treatment have been established (Dixit et al., 2015a). AuNPs conjugated with peptides (EGF and transferrin) and loaded with the photosensitizer phthalocyanine 4 (Pc 4) displayed

synergistic effects in human glioma cells, concomitant with a high accumulation in the brain tumor area compared to AuNPs alone (Dixit et al., 2015a). Many in vitro studies reported positive effects; however, in vivo investigations based on theranostic NPS concepts are necessary to translate into clinical practice (Schmieder et al., 2008; Jokerst and Gambhir, 2011; Sailor and Park, 2012). In vitro and in vitro studies for GB treatment have been reported, and include gold nanoparticles, curcumin-loaded RDP-liposomes, curcuminloaded PLGA-DSPE-PEG nanoparticles, chitosan-based nanoparticles, iron oxide nanoparticles coated with a chitosan-PEG-polyethyleneimine copolymer, hyaluronic acid conjugated liposomes, and others (Dixit et al., 2015a,b; Orunoglu et al., 2017; Zhao et al., 2018). Finally, magnetic nanoparticles and Nanotherm theranostic technology have been successfully applied in glioblastoma patients in 27 different European countries with double median survival in 59 patients (reviewed in Xie et al., 2018).

All these studies reported high efficiency against glioblastoma also in *in vivo* investigations, thus indicating a promising application in diagnosis and concomitantly in therapeutic approaches, which results from significant accumulation in the brain tumor regions. NPS are poorly investigated in clinical trials (Andronescu and Grumezescu, 2017). The main limitation for using nanotechnology to diagnose and treat cancer is due to its inability to effectively contain and regulate the activity of NPS inside the body, comprising toxicity, biodistribution, and pharmacokinetics (Wicki et al., 2015; Bregoli et al., 2016; Hare et al., 2017).

CONCLUSION AND PERSPECTIVES

In the last years, the field of "theranostic medicine" has gained increasing interest in order to improve diagnostic and therapeutic interventions by nanotechnology resources that exploit a combined approach (**Figure 1**). A nanoparticle should contain a therapeutic agent combined with a tracer to help monitor the effect of the therapy as well. Theranostic is considered a potential candidate for targeted therapy and personalized medicine because you can follow the specific behavior of each tumor concomitant with a substantial increase in the efficacy of the anticancer drugs. This monitoring during the treatment course is a non-invasive method that, through the use of theranostic strategies, allows for the implementation of the individualization of therapeutic regimens based on each patient's response.

Overall, on light of the published investigations, nanotechnology research may be a potential and valid treatment of CNS pathologies, especially brain cancers (**Figure 2**), helping to address the main issues encountered: unclear tumoral margins, neurotoxicity of adjuvant therapies, fibrosis and immunological responses to intracranial devices, vascular anastomosis, multidrug resistance, BBB blockage, and tumor cell-specificity response to pharmacological treatments. Nanotheranostics, indeed, have shown to be a valid option for malignant brain cancers. Thus, in this review, we reported that nanotheranostics may represent a solid approach to be adopted in brain cancer management. The field of theranostics is pretty new, but considerable efforts have been made in order to develop theranostic nanoparticles for cancer therapy and targeted imaging. Advantages of theranostic nanoparticles include high biosafety, prolonged half-life into the circulatory system, concomitant loading of therapeutic and contrast agents, small size, high surface functionalization, and the ability to perform concomitantly diagnosis/monitoring and therapeutic approaches in real-time. Theranostic NPS allow a specific release of cargo in the affected site, targeting overexpressed proteins and receptors on brain cancer cells. These functions can facilitate the progress of innovative drugs in both preclinical and clinical phases.

Recently, multifunctional applications and combined approaches with personalized medicine applications have increased the hope in a successful clinical translation. Currently, as mentioned above, the only theranostic tool approved for use in the clinical treatment of GBM in Europe is NanoTherm[®] (Shi et al., 2017).

Overall, the goal is that multifunctional nanomedicine is an efficient, targeted *in vivo* drug delivery without systemic toxicity, and the therapeutic efficacy and the dosage can be precisely measured with low or absent invasivity.

However, in order to translate the experimental studies to clinical trials, further investigations are necessary, particularly to understand the low drug-loading capacity and to optimize the drug concentrations that reach the targeted area, and many factors need to be optimized simultaneously for the best clinical outcome.

In vitro and *in vivo* studies optimized to correctly evaluate toxicity, biodistribution, and pharmacokinetics of NPS are strongly requested to test the safety and efficacy of these nanomaterials in clinical studies. Moreover, the complexity of some nanoparticle designs and the high production costs contribute to the lower clinical uptake of NPS (Hare et al., 2017). The major efforts in the field of NPS should be directed toward bridging the gap between preclinical studies and the clinical phase.

The goal would be a better outcome for the patients thanks to the constant monitoring prior and during treatment, which allows for personalized cancer planning with predictable side effects. These multifunctional modern applications may increase a patient's life expectancy and life quality. It is highly probably that, in the near future, the field of nanotheranostics will emerge and become part of the conventional therapy and diagnostic approaches for brain cancer and other type of cancers.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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REFERENCES

- Agrawal, P., Singh, R. P., Sonali, Kumari, L., Sharma, G., Koch, B., et al. (2017a). TPGS-chitosan cross-linked targeted nanoparticles for effective brain cancer therapy. *Mater. Sci. Eng. C Mater. Biol. Appl.* 74, 167–176. doi: 10.1016/j.msec.2017.02.008
- Agrawal, P., Sonali, Singh, R. P., Sharma, G., Mehata, A. K., Singh, S., et al. (2017b). Bioadhesive micelles of d-α-tocopherol polyethylene glycol succinate 1000: synergism of chitosan and transferrin in targeted drug delivery. *Colloids Surf. B Biointerfaces* 152, 277–288. doi: 10.1016/j.colsurfb.2017.01.021
- Ahmed, N., Brawley, V., Hegde, M., Bielamowicz, K., Kalra, M., Landi, D., et al. (2017). HER2-specific chimeric antigen receptor-modified virus-specific t cells for progressive glioblastoma: a phase 1 dose-escalation trial. *JAMA Oncol.* 3, 1094–1101. doi: 10.1001/jamaoncol.2017.0184
- Ahmed, N., Fessi, H., and Elaissari, A. (2012). Theranostic applications of nanoparticles in cancer. *Drug Discov. Today* 17, 928–934. doi: 10.1016/j.drudis.2012.03.010
- Ai, H. (2011). Layer-by-layer capsules for magnetic resonance imaging and drug delivery. Adv. Drug Deliv. Rev. 63, 772–788. doi: 10.1016/j.addr.2011. 03.013
- Al-Jamal, W. T., and Kostarelos, K. (2011). Liposomes: from a clinically established drug delivery system to a nanoparticle platform for theranostic nanomedicine. *Acc. Chem. Res.* 44, 1094–1104. doi: 10.1021/ar200105p
- Alphandéry, E., Grand-Dewyse, P., Lefèvre, R., Mandawala, C., and Durand-Dubief, M. (2015). Cancer therapy using nanoformulated substances: scientific, regulatory and financial aspects. *Expert Rev. Anticancer Ther.* 15, 1233–1255. doi: 10.1586/14737140.2015.1086647
- Andronescu, E., and Grumezescu, A. M. (2017). *Nanostructures for Drug Delivery*. Bucharest: Micro and Nano Technologies.
- Ang, C. Y., Tan, S. Y., Lu, Y., Bai, L., Li, M., Li, P., et al. (2014). "Turn-on" fluorescence probe integrated polymer nanoparticles for sensing biological thiol molecules. *Sci. Rep.* 4:7057. doi: 10.1038/srep07057
- Antonosante, A., d'Angelo, M., Castelli, V., Catanesi, M., Iannotta, D., Giordano, A., et al. (2018). The involvement of PPARs in the peculiar energetic metabolism of tumor cells. *Int. J. Mol. Sci.* 19:1907. doi: 10.3390/ijms19071907
- Aparicio-Blanco, J., and Torres-Suárez, A.-I. (2018). Towards tailored management of malignant brain tumors with nanotheranostics. *Acta Biomater*. 73, 52–63. doi: 10.1016/j.actbio.2018.04.029
- Arranja, A. G., Pathak, V., Lammers, T., and Shi, Y. (2017). Tumortargeted nanomedicines for cancer theranostics. *Pharmacol. Res.* 115, 87–95. doi: 10.1016/j.phrs.2016.11.014
- Bahadur, S., Sahu, A. K., Baghel, P., and Saha, S. (2019). Current promising treatment strategy for glioblastoma multiform: a review. Oncol. Rev. 13:417. doi: 10.4081/oncol.2019.417
- Behin, A., Hoang-Xuan, K., Carpentier, A. F., and Delattre, J.-Y. (2003). Primary brain tumours in adults. *Lancet* 361, 323–331. doi: 10.1016/S0140-6736(03)12328-8
- Belhadj, Z., Zhan, C., Ying, M., Wei, X., Xie, C., Yan, Z., et al. (2017). Multifunctional targeted liposomal drug delivery for efficient glioblastoma treatment. Oncotarget 8, 66889–66900. doi: 10.18632/oncotarget.17976
- Benoy, I. H., Salgado, R., Van Dam, P., Geboers, K., Van Marck, E., Scharpé, S., et al. (2004). Increased serum interleukin-8 in patients with early and metastatic breast cancer correlates with early dissemination and survival. *Clin. Cancer Res.* 10, 7157–7162. doi: 10.1158/1078-0432.CCR-04-0812
- Bhojani, M. S., Van Dort, M., Rehemtulla, A., and Ross, B. D. (2010). Targeted imaging and therapy of brain cancer using theranostic nanoparticles. *Mol. Pharm.* 7, 1921–1929. doi: 10.1021/mp100298r
- Boas, U., and Heegaard, P. M. H. (2004). Dendrimers in drug research. *Chem. Soc. Rev.* 33, 43–63. doi: 10.1039/b309043b
- Bregoli, L., Movia, D., Gavigan-Imedio, J. D., Lysaght, J., Reynolds, J., and Prina-Mello, A. (2016). Nanomedicine applied to translational oncology: a future perspective on cancer treatment. *Nanomedicine*. 12, 81–103. doi: 10.1016/j.nano.2015.08.006
- Cai, W., Rao, J., Gambhir, S. S., and Chen, X. (2006). How molecular imaging is speeding up antiangiogenic drug development. *Mol. Cancer Ther.* 5, 2624–2633. doi: 10.1158/1535-7163.MCT-06-0395
- Cantley, L. C., and Neel, B. G. (1999). New insights into tumor suppression: PTEN suppresses tumor formation by restraining the phosphoinositide

3-kinase/AKT pathway. Proc. Natl. Acad. Sci. U.S.A. 96, 4240–4245. doi: 10.1073/pnas.96.8.4240

- Caracciolo, V., Laurenti, G., Romano, G., Carnevale, V., Cimini, A. M., Crozier-Fitzgerald, C., et al. (2012). Flavopiridol induces phosphorylation of AKT in a human glioblastoma cell line, in contrast to siRNA-mediated silencing of Cdk9: implications for drug design and development. *Cell Cycle* 11, 1202–1216. doi: 10.4161/cc.11.6.19663
- Cen, H., Mao, F., Aronchik, I., Fuentes, R. J., and Firestone, G. L. (2008). DEVD-NucView488: a novel class of enzyme substrates for real-time detection of caspase-3 activity in live cells. *FASEB J.* 22, 2243–2252. doi: 10.1096/fj.07-099234
- Chai, Z., Hu, X., Wei, X., Zhan, C., Lu, L., Jiang, K., et al. (2017). A facile approach to functionalizing cell membrane-coated nanoparticles with neurotoxin-derived peptide for brain-targeted drug delivery. J. Control. Release 264, 102–111. doi: 10.1016/j.jconrel.2017.08.027
- Chakrabarti, M., Kiseleva, R., Vertegel, A., and Ray, S. K. (2015). Carbon nanomaterials for drug delivery and cancer therapy. J. Nanosci. Nanotechnol. 15, 5501–5511. doi: 10.1166/jnn.2015.10614
- Chakravarti, A., Loeffler, J. S., and Dyson, N. J. (2002). Insulin-like growth factor receptor I mediates resistance to anti-epidermal growth factor receptor therapy in primary human glioblastoma cells through continued activation of phosphoinositide 3-kinase signaling. *Cancer Res.* 62, 200–207.
- Chang, Y., Meng, X., Zhao, Y., Li, K., Zhao, B., Zhu, M., et al. (2011). Novel water-soluble and pH-responsive anticancer drug nanocarriers: doxorubicin-PAMAM dendrimer conjugates attached to superparamagnetic iron oxide nanoparticles (IONPs). J. Colloid Interface Sci. 363, 403–409. doi: 10.1016/j.jcis.2011.06.086
- Chen, N.-T., Cheng, S.-H., Souris, J. S., Chen, C.-T., Mou, C.-Y., and Lo, L.-W. (2013). Theranostic applications of mesoporous silica nanoparticles and their organic/inorganic hybrids. J. Mater. Chem. B 1:3128. doi: 10.1039/c3tb20249f
- Chen, Z., Lai, X., Song, S., Zhu, X., and Zhu, J. (2016). Nanostructured lipid carriers based temozolomide and gene co-encapsulated nanomedicine for gliomatosis cerebri combination therapy. *Drug Deliv.* 23, 1369–1373. doi: 10.3109/10717544.2015.1038857
- Cheng, S.-H., Lee, C.-H., Chen, M.-C., Souris, J. S., Tseng, F.-G., Yang, C.-S., et al. (2010). Tri-functionalization of mesoporous silica nanoparticles for comprehensive cancer theranostics—the trio of imaging, targeting and therapy. *J. Mater. Chem.* 20:6149. doi: 10.1039/c0jm00645a
- Cheng, Y., Morshed, R. A., Auffinger, B., Tobias, A. L., and Lesniak, M. S. (2014). Multifunctional nanoparticles for brain tumor imaging and therapy. *Adv. Drug Deliv. Rev.* 66, 42–57. doi: 10.1016/j.addr.2013.09.006
- Chertok, B., Moffat, B. A., David, A. E., Yu, F., Bergemann, C., Ross, B. D., et al. (2008). Iron oxide nanoparticles as a drug delivery vehicle for MRI monitored magnetic targeting of brain tumors. *Biomaterials* 29, 487–496. doi: 10.1016/j.biomaterials.2007.08.050
- Choi, K. Y., Liu, G., Lee, S., and Chen, X. (2012). Theranostic nanoplatforms for simultaneous cancer imaging and therapy: current approaches and future perspectives. *Nanoscale* 4, 330–342. doi: 10.1039/C1NR11277E
- Cimini, A., d'Angelo, M., Benedetti, E., D'Angelo, B., Laurenti, G., Antonosante, A., et al. (2017). Flavopiridol: an old drug with new perspectives? Implication for development of new drugs. J. Cell. Physiol. 232, 312–322. doi: 10.1002/jcp.25421
- Cole, A. J., Yang, V. C., and David, A. E. (2011). Cancer theranostics: the rise of targeted magnetic nanoparticles. *Trends Biotechnol.* 29, 323–332. doi: 10.1016/j.tibtech.2011.03.001
- Danhier, F., Messaoudi, K., Lemaire, L., Benoit, J.-P., and Lagarce, F. (2015). Combined anti-Galectin-1 and anti-EGFR siRNA-loaded chitosanlipid nanocapsules decrease temozolomide resistance in glioblastoma: *in vivo* evaluation. *Int. J. Pharm.* 481, 154–161. doi: 10.1016/j.ijpharm.2015.01.051
- Derfus, A. M., Chan, W. C. W., and Bhatia, S. N. (2004). Probing the cytotoxicity of semiconductor quantum dots. *Nano Lett.* 4, 11–18. doi: 10.1021/nl03 47334
- Di Martino, A., Guselnikova, O. A., Trusova, M. E., Postnikov, P. S., and Sedlarik, V. (2017). Organic-inorganic hybrid nanoparticles controlled delivery system for anticancer drugs. *Int. J. Pharm.* 526, 380–390. doi: 10.1016/j.ijpharm.2017.04.061
- Dixit, S., Miller, K., Zhu, Y., McKinnon, E., Novak, T., Kenney, M. E., et al. (2015a). Dual receptor-targeted theranostic nanoparticles for localized delivery

and activation of photodynamic therapy drug in glioblastomas. *Mol. Pharm.* 12, 3250–3260. doi: 10.1021/acs.molpharmaceut.5b00216

- Dixit, S., Novak, T., Miller, K., Zhu, Y., Kenney, M. E., and Broome, A.-M. (2015b). Transferrin receptor-targeted theranostic gold nanoparticles for photosensitizer delivery in brain tumors. *Nanoscale* 7, 1782–1790. doi: 10.1039/C4NR04853A
- Dong, X. (2018). Current strategies for brain drug delivery. *Theranostics* 8, 1481–1493. doi: 10.7150/thno.21254
- Duncan, R. (2003). The dawning era of polymer therapeutics. Nat. Rev. Drug Discov. 2, 347–360. doi: 10.1038/nrd1088
- Fan, Z., Fu, P. P., Yu, H., and Ray, P. C. (2014). Theranostic nanomedicine for cancer detection and treatment. J. Food Drug Anal. 22, 3–17. doi: 10.1016/j.jfda.2014.01.001
- Fattahi, H., Laurent, S., Liu, F., Arsalani, N., Vander Elst, L., and Muller, R. N. (2011). Magnetoliposomes as multimodal contrast agents for molecular imaging and cancer nanotheragnostics. *Nanomedicine*. 6, 529–544. doi: 10.2217/nnm.11.14
- Feng, Y., Panwar, N., Tng, D. J. H., Tjin, S. C., Wang, K., and Yong, K.-T. (2016). The application of mesoporous silica nanoparticle family in cancer theranostics. *Coord. Chem. Rev.* 319, 86–109. doi: 10.1016/j.ccr.2016.04.019
- Fidoamore, A., Cristiano, L., Antonosante, A., d'Angelo, M., Di Giacomo, E., Astarita, C., et al. (2016). Glioblastoma stem cells microenvironment: the paracrine roles of the niche in drug and radioresistance. *Stem Cells Int.* 2016:6809105. doi: 10.1155/2016/6809105
- Frezza, C., and Gottlieb, E. (2009). Mitochondria in cancer: not just innocent bystanders. Semin. Cancer Biol. 19, 4–11. doi: 10.1016/j.semcancer.2008.11.008
- Frimpong, R. A., and Hilt, J. Z. (2010). Magnetic nanoparticles in biomedicine: synthesis, functionalization and applications. *Nanomedicine* 5, 1401–1414. doi: 10.2217/nnm.10.114
- Gary-Bobo, M., Hocine, O., Brevet, D., Maynadier, M., Raehm, L., Richeter, S., et al. (2012). Cancer therapy improvement with mesoporous silica nanoparticles combining targeting, drug delivery and PDT. *Int. J. Pharm.* 423, 509–515. doi: 10.1016/j.ijpharm.2011.11.045
- Gatenby, R. A., and Gillies, R. J. (2004). Why do cancers have high aerobic glycolysis? *Nat. Rev. Cancer* 4, 891–899. doi: 10.1038/nrc1478
- Gaya, A., Rees, J., Greenstein, A., and Stebbing, J. (2002). The use of temozolomide in recurrent malignant gliomas. *Cancer Treat. Rev.* 28, 115–120. doi: 10.1053/ctrv.2002.0261
- Golombek, S. K., May, J.-N., Theek, B., Appold, L., Drude, N., Kiessling, F., et al. (2018). Tumor targeting via EPR: strategies to enhance patient responses. *Adv. Drug Deliv. Rev.* 130, 17–38. doi: 10.1016/j.addr.2018.07.007
- Graverini, G., Piazzini, V., Landucci, E., Pantano, D., Nardiello, P., Casamenti, F., et al. (2018). Solid lipid nanoparticles for delivery of andrographolide across the blood-brain barrier: *in vitro* and *in vivo* evaluation. *Colloids Surf. B Biointerfaces* 161, 302–313. doi: 10.1016/j.colsurfb.2017.10.062
- Gürsel, D. B., Shin, B. J., Burkhardt, J.-K., Kesavabhotla, K., Schlaff, C. D., and Boockvar, J. A. (2011). Glioblastoma stem-like cells-biology and therapeutic implications. *Cancers*. 3, 2655–2666. doi: 10.3390/cancers3022655
- Hainfeld, J. F., Smilowitz, H. M., O'Connor, M. J., Dilmanian, F. A., and Slatkin, D. N. (2013). Gold nanoparticle imaging and radiotherapy of brain tumors in mice. *Nanomedicine*. 8, 1601–1609. doi: 10.2217/nnm.12.165
- Hare, J. I., Lammers, T., Ashford, M. B., Puri, S., Storm, G., and Barry, S. T. (2017). Challenges and strategies in anti-cancer nanomedicine development: an industry perspective. *Adv. Drug Deliv. Rev.* 108, 25–38. doi: 10.1016/j.addr.2016.04.025
- Harrington, K. J., Mohammadtaghi, S., Uster, P. S., Glass, D., Peters, A. M., Vile, R. G., et al. (2001). Effective targeting of solid tumors in patients with locally advanced cancers by radiolabeled pegylated liposomes. *Clin. Cancer Res.* 7, 243–254.
- Helms, H. C., Abbott, N. J., Burek, M., Cecchelli, R., Couraud, P.-O., Deli, M. A., et al. (2016). *In vitro* models of the blood-brain barrier: an overview of commonly used brain endothelial cell culture models and guidelines for their use. *J Cerebral Blood Flow Metab.* 36, 862–890. doi: 10.1177/0271678X16630991
- Heo, D. N., Yang, D. H., Moon, H.-J., Lee, J. B., Bae, M. S., Lee, S. C., et al. (2012). Gold nanoparticles surface-functionalized with paclitaxel drug and biotin receptor as theranostic agents for cancer therapy. *Biomaterials* 33, 856–866. doi: 10.1016/j.biomaterials.2011.09.064

- Hsu, S. P. C., Kuo, J. S., Chiang, H.-C., Wang, H.-E., Wang, Y.-S., Huang, C.-C., et al. (2018). Temozolomide, sirolimus and chloroquine is a new therapeutic combination that synergizes to disrupt lysosomal function and cholesterol homeostasis in GBM cells. *Oncotarget* 9, 6883–6896. doi: 10.18632/oncotarget.23855
- Huang, X., Zhang, F., Wang, H., Niu, G., Choi, K. Y., Swierczewska, M., et al. (2013). Mesenchymal stem cell-based cell engineering with multifunctional mesoporous silica nanoparticles for tumor delivery. *Biomaterials* 34, 1772–1780. doi: 10.1016/j.biomaterials.2012.11.032
- Jhaveri, A., Deshpande, P., Pattni, B., and Torchilin, V. (2018). Transferrintargeted, resveratrol-loaded liposomes for the treatment of glioblastoma. J. Control. Release 277, 89–101. doi: 10.1016/j.jconrel.2018.03.006
- Jiang, X., Sha, X., Xin, H., Xu, X., Gu, J., Xia, W., et al. (2013). Integrinfacilitated transcytosis for enhanced penetration of advanced gliomas by poly(trimethylene carbonate)-based nanoparticles encapsulating paclitaxel. *Biomaterials* 34, 2969–2979. doi: 10.1016/j.biomaterials.2012.12.049
- Joh, D. Y., Sun, L., Stangl, M., Al Zaki, A., Murty, S., Santoiemma, P. P., et al. (2013). Selective targeting of brain tumors with gold nanoparticle-induced radiosensitization. *PLoS ONE* 8:e62425. doi: 10.1371/journal.pone.0062425
- Jokerst, J. V., and Gambhir, S. S. (2011). Molecular imaging with theranostic nanoparticles. Acc. Chem. Res. 44, 1050–1060. doi: 10.1021/ar200106e
- Kelkar, S. S., and Reineke, T. M. (2011). Theranostics: combining imaging and therapy. *Bioconjug. Chem.* 22, 1879–1903. doi: 10.1021/bc200151q
- Kenny, G. D., Kamaly, N., Kalber, T. L., Brody, L. P., Sahuri, M., Shamsaei, E., et al. (2011). Novel multifunctional nanoparticle mediates siRNA tumour delivery, visualisation and therapeutic tumour reduction *in vivo. J. Control. Release* 149, 111–116. doi: 10.1016/j.jconrel.2010.09.020
- Keunen, O., Johansson, M., Oudin, A., Sanzey, M., Rahim, S. A. A., Fack, F., et al. (2011). Anti-VEGF treatment reduces blood supply and increases tumor cell invasion in glioblastoma. *Proc. Natl. Acad. Sci. U.S.A.* 108, 3749–3754. doi: 10.1073/pnas.1014480108
- Kievit, F. M., and Zhang, M. (2011). Cancer nanotheranostics: improving imaging and therapy by targeted delivery across biological barriers. *Adv. Mater. Weinheim* 23, H217–247. doi: 10.1002/adma.201102313
- Kirchner, C., Liedl, T., Kudera, S., Pellegrino, T., Muñoz Javier, A., Gaub, H. E., et al. (2005). Cytotoxicity of colloidal CdSe and CdSe/ZnS nanoparticles. *Nano Lett.* 5, 331–338. doi: 10.1021/nl047996m
- Kondo, T., Setoguchi, T., and Taga, T. (2004). Persistence of a small subpopulation of cancer stem-like cells in the C6 glioma cell line. *Proc. Natl. Acad. Sci. U.S.A.* 101, 781–786. doi: 10.1073/pnas.0307618100
- Koo, Y.-E. L., Reddy, G. R., Bhojani, M., Schneider, R., Philbert, M. A., Rehemtulla, A., et al. (2006). Brain cancer diagnosis and therapy with nanoplatforms. *Adv. Drug Deliv. Rev.* 58, 1556–1577. doi: 10.1016/j.addr.2006.09.012
- Kroemer, G., and Pouyssegur, J. (2008). Tumor cell metabolism: cancer's achilles' heel. *Cancer Cell* 13, 472–482. doi: 10.1016/j.ccr.2008.05.005
- Kroonen, J., Nguyen-Khac, M. T., Deprez, M., Rogister, B., and Robe, P. (2008). [Glioblastoma, an example of translational research?]. *Rev. Med. Liege* 63, 251–256.
- Kumar, A., Lee, J.-Y., and Kim, H.-S. (2016). Selective fluorescence sensing of 3,5-dinitrosalicylic acid based on pyrenesulfonamide-functionalized inorganic/organic hybrid nanoparticles. J. Industr. Eng. Chem. 44, 82–89. doi: 10.1016/j.jiec.2016.08.010
- Kumar, A., Zhang, X., and Liang, X.-J. (2013). Gold nanoparticles: emerging paradigm for targeted drug delivery system. *Biotechnol. Adv.* 31, 593–606. doi: 10.1016/j.biotechadv.2012.10.002
- Kuo, Y.-C., and Cheng, S.-J. (2016). Brain targeted delivery of carmustine using solid lipid nanoparticles modified with tamoxifen and lactoferrin for antitumor proliferation. *Int. J. Pharm.* 499, 10–19. doi: 10.1016/j.ijpharm.2015.12.054
- Lakka, S. S., and Rao, J. S. (2008). Antiangiogenic therapy in brain tumors. *Expert Rev. Neurother.* 8, 1457–1473. doi: 10.1586/14737175.8.10.1457
- Lammers, T., Kiessling, F., Hennink, W. E., and Storm, G. (2010). Nanotheranostics and image-guided drug delivery: current concepts and future directions. *Mol. Pharm.* 7, 1899–1912. doi: 10.1021/mp100228v
- Lammers, T., Koczera, P., Fokong, S., Gremse, F., Ehling, J., Vogt, M., et al. (2015). Theranostic USPIO-loaded microbubbles for mediating and monitoring blood-brain barrier permeation. *Adv. Funct. Mater.* 25, 36–43. doi: 10.1002/adfm.201401199

- Lee, J. E., Lee, N., Kim, T., Kim, J., and Hyeon, T. (2011). Multifunctional mesoporous silica nanocomposite nanoparticles for theranostic applications. *Acc. Chem. Res.* 44, 893–902. doi: 10.1021/ar2000259
- Lindner, L. H., and Hossann, M. (2010). Factors affecting drug release from liposomes. *Curr. Opin. Drug Discov. Devel.* 13, 111–123.
- Liong, M., Lu, J., Kovochich, M., Xia, T., Ruehm, S. G., Nel, A. E., et al. (2008). Multifunctional inorganic nanoparticles for imaging, targeting, and drug delivery. ACS Nano 2, 889–896. doi: 10.1021/nn800072t
- Liu, G., Wang, Z., Lu, J., Xia, C., Gao, F., Gong, Q., et al. (2011). Low molecular weight alkyl-polycation wrapped magnetite nanoparticle clusters as MRI probes for stem cell labeling and *in vivo* imaging. *Biomaterials* 32, 528–537. doi: 10.1016/j.biomaterials.2010.08.099
- Liu, X., Madhankumar, A. B., Miller, P. A., Duck, K. A., Hafenstein, S., Rizk, E., et al. (2016). MRI contrast agent for targeting glioma: interleukin-13 labeled liposome encapsulating gadolinium-DTPA. *Neuro-oncology* 18, 691–699. doi: 10.1093/neuonc/nov263
- Lo, S.-T., Kumar, A., Hsieh, J.-T., and Sun, X. (2013). Dendrimer nanoscaffolds for potential theranostics of prostate cancer with a focus on radiochemistry. *Mol. Pharm.* 10, 793–812. doi: 10.1021/mp3005325
- Lu, J., Ma, S., Sun, J., Xia, C., Liu, C., Wang, Z., et al. (2009). Manganese ferrite nanoparticle micellar nanocomposites as MRI contrast agent for liver imaging. *Biomaterials* 30, 2919–2928. doi: 10.1016/j.biomaterials.2009.02.001
- Mamaeva, V., Sahlgren, C., and Lindén, M. (2013). Mesoporous silica nanoparticles in medicine–recent advances. Adv. Drug Deliv. Rev. 65, 689–702. doi: 10.1016/j.addr.2012.07.018
- Mattei, T. A., and Rehman, A. A. (2015). "Extremely minimally invasive": recent advances in nanotechnology research and future applications in neurosurgery. *Neurosurg. Rev.* 38, 27–37; discussion 37. doi: 10.1007/s10143-014-0566-2
- Melancon, M. P., Lu, W., Zhong, M., Zhou, M., Liang, G., Elliott, A. M., et al. (2011). Targeted multifunctional gold-based nanoshells for magnetic resonance-guided laser ablation of head and neck cancer. *Biomaterials* 32, 7600–7608. doi: 10.1016/j.biomaterials.2011.06.039
- Mendes, M., Sousa, J. J., Pais, A., and Vitorino, C. (2018). Targeted theranostic nanoparticles for brain tumor treatment. *Pharmaceutics* 10:E181. doi: 10.3390/pharmaceutics10040181
- Merkel, O. M., Mintzer, M. A., Librizzi, D., Samsonova, O., Dicke, T., Sproat, B., et al. (2010). Triazine dendrimers as non-viral vectors for *in vitro* and *in vivo* RNAi: the effects of peripheral groups and core structure on biological activity. *Mol. Pharm.* 7, 969–983. doi: 10.1021/mp100101s
- Mintzer, M. A., and Grinstaff, M. W. (2011). Biomedical applications of dendrimers: a tutorial. *Chem. Soc. Rev.* 40, 173–190. doi: 10.1039/B901839P
- Miranda, A., Blanco-Prieto, M. J., Sousa, J., Pais, A., and Vitorino, C. (2017). Breaching barriers in glioblastoma. Part II: Targeted drug delivery and lipid nanoparticles. *Int. J. Pharm.* 531, 389–410. doi: 10.1016/j.ijpharm.2017. 07.049
- Muthu, M. S., Leong, D. T., Mei, L., and Feng, S.-S. (2014a). Nanotheranostics application and further development of nanomedicine strategies for advanced theranostics. *Theranostics* 4, 660–677. doi: 10.7150/thno.8698
- Muthu, M. S., Mei, L., and Feng, S.-S. (2014b). Nanotheranostics: advanced nanomedicine for the integration of diagnosis and therapy. *Nanomedicine* 9, 1277–1280. doi: 10.2217/nnm.14.83
- Na, K., Lee, S. A., Jung, S. H., and Shin, B. C. (2011). Gadolinium-based cancer therapeutic liposomes for chemotherapeutics and diagnostics. *Colloids Surfaces B Biointerfaces* 84, 82–87. doi: 10.1016/j.colsurfb.2010.12.019
- Nam, J., Won, N., Bang, J., Jin, H., Park, J., Jung, S., et al. (2013). Surface engineering of inorganic nanoparticles for imaging and therapy. Adv. Drug Deliv. Rev. 65, 622–648. doi: 10.1016/j.addr.2012.08.015
- Nam, L., Coll, C., Erthal, L., de la Torre, C., Serrano, D., Martínez-Máñez, R., et al. (2018). Drug delivery nanosystems for the localized treatment of glioblastoma multiforme. *Materials*. 11:779. doi: 10.3390/ma11050779
- Nance, E., Zhang, C., Shih, T.-Y., Xu, Q., Schuster, B. S., and Hanes, J. (2014). Brain-penetrating nanoparticles improve paclitaxel efficacy in malignant glioma following local administration. ACS Nano 8, 10655–10664. doi: 10.1021/nn504210g.
- Newton, H. B. (2010). "Overview of the molecular genetics and molecular chemotherapy of GBM," in *Glioblastoma*, ed S. K. Ray (New York, NY: Springer New York), 1–42. doi: 10.1007/978-1-4419-0410-2_1

- Nicholas, M. K., Lukas, R. V., Jafri, N. F., Faoro, L., and Salgia, R. (2006). Epidermal growth factor receptor - mediated signal transduction in the development and therapy of gliomas. *Clin. Cancer Res.* 12, 7261–7270. doi: 10.1158/1078-0432.CCR-06-0874
- Onoshima, D., Yukawa, H., and Baba, Y. (2015). Multifunctional quantum dotsbased cancer diagnostics and stem cell therapeutics for regenerative medicine. *Adv. Drug Deliv. Rev.* 95, 2–14. doi: 10.1016/j.addr.2015.08.004
- Orunoglu, M., Kaffashi, A., Pehlivan, S. B., Sahin, S., and Söylemezoglu, F., Oguz, K. K., et al. (2017). Effects of curcumin-loaded PLGA nanoparticles on the RG2 rat glioma model. *Mater. Sci. Eng. C* 78, 32–38. doi: 10.1016/j.msec.2017.03.292
- Pacioni, S., D'Alessandris, Q. G., Giannetti, S., Morgante, L., Coccè, V., Bonomi, A., et al. (2017). Human mesenchymal stromal cells inhibit tumor growth in orthotopic glioblastoma xenografts. *Stem Cell Res. Ther.* 8:53. doi: 10.1186/s13287-017-0516-3
- Pan, B., Cui, D., Sheng, Y., Ozkan, C., Gao, F., He, R., et al. (2007). Dendrimermodified magnetic nanoparticles enhance efficiency of gene delivery system. *Cancer Res.* 67, 8156–8163. doi: 10.1158/0008-5472.CAN-06-4762
- Pankhurst, Q. A., Connolly, J., Jones, S. K., and Dobson, J. (2003). Applications of magnetic nanoparticles in biomedicine. J. Phys. D Appl. Phys. 36, R167–R181. doi: 10.1088/0022-3727/36/13/201
- Park, J.-H., Ryu, C. H., Kim, M. J., and Jeun, S.-S. (2015). Combination therapy for gliomas using temozolomide and interferon-beta secreting human bone marrow derived mesenchymal stem cells. J. Korean Neurosurg. Soc. 57, 323–328. doi: 10.3340/jkns.2015.57.5.323
- Peer, D., Karp, J. M., Hong, S., Farokhzad, O. C., Margalit, R., and Langer, R. (2007). Nanocarriers as an emerging platform for cancer therapy. *Nat. Nanotechnol.* 2, 751–760. doi: 10.1038/nnano.2007.387
- Petersen, A. L., Binderup, T., Jølck, R. I., Rasmussen, P., Henriksen, J. R., Pfeifer, A. K., et al. (2012). Positron emission tomography evaluation of somatostatin receptor targeted 64Cu-TATE-liposomes in a human neuroendocrine carcinoma mouse model. J. Control. Release 160, 254–263. doi: 10.1016/j.jconrel.2011.12.038
- Pilkington, G. J., Maherally, Z., Jassam, S., Barbu, E., and Fillmore, H. (2014). An all human 3D *in vitro* model of the blood brain barrier in nanoparticle delivery and cancer metastasis studies. *Neuro-Oncology* 16:iii33. doi: 10.1093/neuonc/nou208.39
- Piquer, J., Llácer, J. L., Rovira, V., Riesgo, P., Rodriguez, R., and Cremades, A. (2014). Fluorescence-guided surgery and biopsy in gliomas with an exoscope system. *Biomed Res. Int.* 2014:207974. doi: 10.1155/2014/207974
- Puputti, M., Tynninen, O., Sihto, H., Blom, T., Mäenp,ää, H., Isola, J., et al. (2006). Amplification of KIT, PDGFRA, VEGFR2, and EGFR in gliomas. *Mol. Cancer Res.* 4, 927–934. doi: 10.1158/1541-7786.MCR-06-0085
- Pysz, M. A., Gambhir, S. S., and Willmann, J. K. (2010). Molecular imaging: current status and emerging strategies. *Clin. Radiol.* 65, 500–516. doi: 10.1016/j.crad.2010.03.011
- Qu, J., Zhang, L., Chen, Z., Mao, G., Gao, Z., Lai, X., et al. (2016). Nanostructured lipid carriers, solid lipid nanoparticles, and polymeric nanoparticles: which kind of drug delivery system is better for glioblastoma chemotherapy? *Drug Deliv.* 23, 3408–3416. doi: 10.1080/10717544.2016.1189465
- Ren, J., Shen, S., Wang, D., Xi, Z., Guo, L., Pang, Z., et al. (2012). The targeted delivery of anticancer drugs to brain glioma by PEGylated oxidized multi-walled carbon nanotubes modified with angiopep-2. *Biomaterials* 33, 3324–3333. doi: 10.1016/j.biomaterials.2012.01.025
- Rivera Gil, P., Hühn, D., del Mercato, L. L., Sasse, D., and Parak, W. J. (2010). Nanopharmacy: inorganic nanoscale devices as vectors and active compounds. *Pharmacol. Res.* 62, 115–125. doi: 10.1016/j.phrs.2010.01.009
- Robinson, J. T., Welsher, K., Tabakman, S. M., Sherlock, S. P., Wang, H., Luong, R., et al. (2010). High performance *in vivo* near-IR (>1 μm) imaging and photothermal cancer therapy with carbon nanotubes. *Nano Res.* 3, 779–793. doi: 10.1007/s12274-010-0045-1
- Rock, K., McArdle, O., Forde, P., Dunne, M., Fitzpatrick, D., O'Neill, B., et al. (2012). A clinical review of treatment outcomes in glioblastoma multiformethe validation in a non-trial population of the results of a randomised Phase III clinical trial: has a more radical approach improved survival? *Br. J. Radiol.* 85, e729–733. doi: 10.1259/bjr/83796755
- Ruan, S., He, Q., and Gao, H. (2015). Matrix metalloproteinase triggered sizeshrinkable gelatin-gold fabricated nanoparticles for tumor microenvironment

sensitive penetration and diagnosis of glioma. Nanoscale 7, 9487-9496. doi: 10.1039/c5nr01408e

- Sailor, M. J., and Park, J.-H. (2012). Hybrid nanoparticles for detection and treatment of cancer. Adv. Mater. 24, 3779–3802. doi: 10.1002/adma.2012 00653
- Saraiva, C., Praça, C., Ferreira, R., Santos, T., Ferreira, L., and Bernardino, L. (2016). Nanoparticle-mediated brain drug delivery: overcoming blood-brain barrier to treat neurodegenerative diseases. J. Control. Release 235, 34–47. doi: 10.1016/j.jconrel.2016.05.044
- Schlessinger, J. (2000). Cell signaling by receptor tyrosine kinases. *Cell* 103, 211–225. doi: 10.1016/S0092-8674(00)00114-8
- Schmieder, A. H., Caruthers, S. D., Zhang, H., Williams, T. A., Robertson, J. D., Wickline, S. A., et al. (2008). Three-dimensional MR mapping of angiogenesis with alpha5beta1(alpha nu beta3)-targeted theranostic nanoparticles in the MDA-MB-435 xenograft mouse model. *FASEB J.* 22, 4179–4189. doi: 10.1096/fj.08-112060
- Shapira, A., Livney, Y. D., Broxterman, H. J., and Assaraf, Y. G. (2011). Nanomedicine for targeted cancer therapy: towards the overcoming of drug resistance. *Drug Resist. Updat.* 14, 150–163. doi: 10.1016/j.drup.2011.01.003
- Shi, J., Kantoff, P. W., Wooster, R., and Farokhzad, O. C. (2017). Cancer nanomedicine: progress, challenges and opportunities. *Nat. Rev. Cancer* 17, 20–37. doi: 10.1038/nrc.2016.108
- Silantyev, F., Falzone, L., Libra, M., Gurina, O. I., Kardashova, K. S., Nikolouzakis, T. K., et al. (2019). Current and future trends on diagnosis and prognosis of glioblastoma: from molecular biology to proteomics. *Cells* 8:863. doi: 10.3390/cells8080863
- Singh, R. P., Sharma, G., Sonali, Singh, S., Kumar, M., Pandey, B. L., et al. (2016). Vitamin E TPGS conjugated carbon nanotubes improved efficacy of docetaxel with safety for lung cancer treatment. *Colloids Surf. B Biointerfaces* 141, 429–442. doi: 10.1016/j.colsurfb.2016.02.011
- Slettenaar, V. I. F., and Wilson, J. L. (2006). The chemokine network: a target in cancer biology? Adv. Drug Deliv. Rev. 58, 962–974. doi: 10.1016/j.addr.2006.03.012
- Sonali, Agrawal, P., Singh, R. P., Rajesh, C. V., Singh, S., Vijayakumar, M. R., et al. (2016a). Transferrin receptor-targeted vitamin E TPGS micelles for brain cancer therapy: preparation, characterization and brain distribution in rats. *Drug Deliv.* 23, 1788–1798. doi: 10.3109/10717544.2015.1094681
- Sonali, Singh, R. P., Sharma, G., Kumari, L., Koch, B., Singh, S., et al. (2016b). RGD-TPGS decorated theranostic liposomes for brain targeted delivery. *Colloids Surf. B Biointerfaces* 147, 129–141. doi: 10.1016/j.colsurfb.2016.07.058
- Sonali, Singh, R. P., Singh, N., Sharma, G., Vijayakumar, M. R., Koch, B., et al. (2016c). Transferrin liposomes of docetaxel for brain-targeted cancer applications: formulation and brain theranostics. *Drug Deliv.* 23, 1261–1271. doi: 10.3109/10717544.2016.1162878
- Sonali, Viswanadh, M. K., Singh, R. P., Agrawal, P., Mehata, A. K., Pawde, D. M., et al. (2018). Nanotheranostics: emerging strategies for early diagnosis and therapy of brain cancer. *Nanotheranostics* 2, 70–86. doi: 10.7150/ntno.21638
- Stambolic, V., Suzuki, A., de la Pompa, J. L., Brothers, G. M., Mirtsos, C., Sasaki, T., et al. (1998). Negative regulation of PKB/Akt-dependent cell survival by the tumor suppressor PTEN. *Cell* 95, 29–39. doi: 10.1016/S0092-8674(00)81780-8
- Su, H., Liu, Y., Wang, D., Wu, C., Xia, C., Gong, Q., et al. (2013). Amphiphilic starlike dextran wrapped superparamagnetic iron oxide nanoparticle clsuters as effective magnetic resonance imaging probes. *Biomaterials* 34, 1193–1203. doi: 10.1016/j.biomaterials.2012.10.056
- Sun, D. (2010). Nanotheranostics: integration of imaging and targeted drug delivery. *Mol. Pharm.* 7:1879. doi: 10.1021/mp1003652
- Sun, L., Joh, D. Y., Al-Zaki, A., Stangl, M., Murty, S., Davis, J. J., et al. (2016). Theranostic application of mixed gold and superparamagnetic iron oxide nanoparticle micelles in glioblastoma multiforme. *J. Biomed. Nanotechnol.* 12, 347–356. doi: 10.1166/jbn.2016.2173
- Tabatabaei, S. N., Girouard, H., Carret, A.-S., and Martel, S. (2015). Remote control of the permeability of the blood-brain barrier by magnetic heating of nanoparticles: a proof of concept for brain drug delivery. J. Control. Release 206, 49–57. doi: 10.1016/j.jconrel.2015.02.027
- Tanaka, N., Kanatani, S., Tomer, R., Sahlgren, C., Kronqvist, P., Kaczynska, D., et al. (2017). Whole-tissue biopsy phenotyping of three-dimensional tumours reveals patterns of cancer heterogeneity. *Nat. Biomed. Eng.* 1, 796–806. doi: 10.1038/s41551-017-0139-0

- Tandel, G. S., Biswas, M., Kakde, O. G., Tiwari, A., Suri, H. S., Turk, M., et al. (2019). A review on a deep learning perspective in brain cancer classification. *Cancers*. 11:E111. doi: 10.3390/cancers11010111
- Tennant, D. A., Frezza, C., MacKenzie, E. D., Nguyen, Q. D., Zheng, L., Selak, M. A., et al. (2009). Reactivating HIF prolyl hydroxylases under hypoxia results in metabolic catastrophe and cell death. *Oncogene* 28, 4009–4021. doi: 10.1038/onc.2009.250
- Thakkar, J. P., Dolecek, T. A., Horbinski, C., Ostrom, Q. T., Lightner, D. D., Barnholtz-Sloan, J. S., et al. (2014). Epidemiologic and molecular prognostic review of glioblastoma. *Cancer Epidemiol. Biomarkers Prev.* 23, 1985–1996. doi: 10.1158/1055-9965.EPI-14-0275
- Tian, B., Al-Jamal, W. T., Al-Jamal, K. T., and Kostarelos, K. (2011). Doxorubicinloaded lipid-quantum dot hybrids: surface topography and release properties. *Int. J. Pharm.* 416, 443–447. doi: 10.1016/j.ijpharm.2011.01.057
- Tong, R., and Cheng, J. (2007). Anticancer polymeric nanomedicines. Poly. Rev. 47, 345–381. doi: 10.1080/15583720701455079
- Tzeng, S. Y., and Green, J. J. (2013). Therapeutic nanomedicine for brain cancer. *Ther. Deliv.* 4, 687–704. doi: 10.4155/tde.13.38
- Uchida, N., Buck, D. W., He, D., Reitsma, M. J., Masek, M., Phan, T. V., et al. (2000). Direct isolation of human central nervous system stem cells. *Proc. Natl. Acad. Sci. U.S.A.* 97, 14720–14725. doi: 10.1073/pnas.97.26.14720
- Vilos, C., and Velasquez, L. A. (2012). Therapeutic strategies based on polymeric microparticles. J. Biomed. Biotechnol. 2012;672760. doi: 10.1155/2012/6 72760
- Volkov, Y. (2015). Quantum dots in nanomedicine: recent trends, advances and unresolved issues. *Biochem. Biophys. Res. Commun.* 468, 419–427. doi: 10.1016/j.bbrc.2015.07.039
- Wang, D., Lin, B., and Ai, H. (2014). Theranostic nanoparticles for cancer and cardiovascular applications. *Pharm. Res.* 31, 1390–1406. doi: 10.1007/s11095-013-1277-z
- Wang, H., Liu, X., Wang, Y., Chen, Y., Jin, Q., and Ji, J. (2015). Doxorubicin conjugated phospholipid prodrugs as smart nanomedicine platforms for cancer therapy. J. Mater. Chem. B 3, 3297–3305. doi: 10.1039/C4TB01984A
- Wang, X., Wang, C., Cheng, L., Lee, S.-T., and Liu, Z. (2012). Noble metal coated single-walled carbon nanotubes for applications in surface enhanced Raman scattering imaging and photothermal therapy. J. Am. Chem. Soc. 134, 7414–7422. doi: 10.1021/ja300140c
- Wang, Y., Shim, M. S., Levinson, N. S., Sung, H.-W., and Xia, Y. (2014). Stimuliresponsive materials for controlled release of theranostic agents. *Adv. Funct. Mater.* 24, 4206–4220. doi: 10.1002/adfm.201400279
- Wei, X., Chen, X., Ying, M., and Lu, W. (2014). Brain tumor-targeted drug delivery strategies. Acta Pharm. Sin. B 4, 193–201. doi: 10.1016/j.apsb.2014.03.001
- Wesseling, P., and Capper, D. (2018). WHO 2016 classification of gliomas. Neuropathol. Appl. Neurobiol. 44, 139–150. doi: 10.1111/nan.12432
- Wicki, A., Witzigmann, D., Balasubramanian, V., and Huwyler, J. (2015). Nanomedicine in cancer therapy: challenges, opportunities, and clinical applications. *J. Controlled Release* 200, 138–157. doi: 10.1016/j.jconrel.2014.12.030
- Wilhelm, I., and Krizbai, I. A. (2014). *In vitro* models of the blood-brain barrier for the study of drug delivery to the brain. *Mol. Pharm.* 11, 1949–1963. doi: 10.1021/mp500046f
- Wong, A. J., Ruppert, J. M., Bigner, S. H., Grzeschik, C. H., Humphrey, P. A., Bigner, D. S., et al. (1992). Structural alterations of the epidermal growth factor receptor gene in human gliomas. *Proc. Natl. Acad. Sci. U.S.A.* 89, 2965–2969. doi: 10.1073/pnas.89.7.2965
- Wu, M., Fan, Y., Lv, S., Xiao, B., Ye, M., and Zhu, X. (2016). Vincristine and temozolomide combined chemotherapy for the treatment of glioma: a comparison of solid lipid nanoparticles and nanostructured lipid carriers for dual drugs delivery. *Drug Deliv.* 23, 2720–2725. doi: 10.3109/10717544.2015.1058434
- Xiao, Q., Yang, S., Ding, G., and Luo, M. (2018). Anti-vascular endothelial growth factor in glioblastoma: a systematic review and meta-analysis. *Neurol. Sci.* 39, 2021–2031. doi: 10.1007/s10072-018-3568-y
- Xie, J., Lee, S., and Chen, X. (2010). Nanoparticle-based theranostic agents. Adv. Drug Deliv. Rev. 62, 1064–1079. doi: 10.1016/j.addr.2010.07.009
- Xie, J., Liu, G., Eden, H. S., Ai, H., and Chen, X. (2011). Surface-engineered magnetic nanoparticle platforms for cancer imaging and therapy. Acc. Chem. Res. 44, 883–892. doi: 10.1021/ar200044b

- Xie, W., Guo, Z., Gao, F., Gao, Q., Wang, D., Liaw, B., et al. (2018). Shape-, size- and structure-controlled synthesis and biocompatibility of iron oxide nanoparticles for magnetic theranostics. *Theranostics* 8, 3284–3307. doi: 10.7150/thno. 25220
- Yamanaka, R. (2008). Cell- and peptide-based immunotherapeutic approaches for glioma. Trends Mol. Med. 14, 228–235. doi: 10.1016/j.molmed.2008.03.003
- Zeng, W., Wang, X., Xu, P., Liu, G., Eden, H. S., and Chen, X. (2015). Molecular imaging of apoptosis: from micro to macro. *Theranostics* 5, 559–582. doi: 10.7150/thno.11548
- Zhang, B., Yang, C., Gao, Y., Wang, Y., Bu, C., Hu, S., et al. (2017). Engineering quantum dots with different emission wavelengths and specific fluorescence lifetimes for spectrally and temporally multiplexed imaging of cells. *Nanotheranostics* 1, 131–140. doi: 10.7150/ntno. 18989
- Zhao, M., Zhao, M., Fu, C., Yu, Y., and Fu, A. (2018). Targeted therapy of intracranial glioma model mice with curcumin nanoliposomes. *Int. J. Nanomedicine* 13, 1601–1610. doi: 10.2147/IJN.S 157019

- Zong, T., Mei, L., Gao, H., Cai, W., Zhu, P., Shi, K., et al. (2014). Synergistic dualligand doxorubicin liposomes improve targeting and therapeutic efficacy of brain glioma in animals. *Mol. Pharm.* 11, 2346–2357. doi: 10.1021/mp500057n
- Zottel, A., Videtič Paska, A., and Jovčevska, I. (2019). Nanotechnology meets oncology: nanomaterials in brain cancer research, diagnosis and therapy. *Materials*. 12:1588. doi: 10.3390/ma12101588

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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