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Nanomaterials against intracellular bacterial infection: from drug delivery to intrinsic biofunction

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Fighting intracellular bacteria with strong antibiotics evading remains a longstanding challenge. Responding to and regulating the infectious microenvironment is crucial for treating intracellular infections. Sophisticated nanomaterials with unique physicochemical properties exhibit great potential for precise drug delivery towards infection sites, along with modulating infectious microenvironment via their instinct bioactivity. In this review, we first identify the key characters and therapeutic targets of intracellular infection microenvironment. Next, we illustrate how the nanomaterials physicochemical properties, such as size, charge, shape and functionalization affect the interaction between nanomaterials, cells and bacteria. We also introduce the recent progress of nanomaterial-based targeted delivery and controlled release of antibiotics in intracellular infection microenvironment. Notably, we highlight the nanomaterials with unique intrinsic properties, such as metal toxicity and enzyme-like activity for the treatment of intracellular bacteria. Finally, we discuss the opportunities and challenges of bioactive nanomaterials in addressing intracellular infections.

KEYWORDS

intracellular infection, drug delivery, antibacterial, reactive oxygen species, bioactive nanomaterials

1 Introduction

In the territory of infectious disease, chronic and persistent infections caused by intracellular bacteria pose a thorny threat to public health (Kamaruzzaman et al., 2017). In these contexts, pathogens such as *Staphylococcus aureus* (*S. aureus*), *Mycobacterium tuberculosis* (*M. tuberculosis*), *Salmonella* and *Listeria* are able to nestle in professional phagocytic cells, particularly macrophages, which not only shield them from the host immune system's eradication but also from antibacterial agents. The most prominent intracellular infections in clinic are associated with *M. tuberculosis*, which can require prolonged and substantial antibiotic treatments. Over extended periods, intracellular bacteria can act as a 'Trojan horse', resulting in a secondary relapsing infection primarily due to their ability to survive and multiply rapidly within host cells. This

category includes obligate intracellular bacteria that can reproduce both inside and outside their cellular hosts, as well as facultative intracellular bacteria that depend on host cells for their reproduction (Briones et al., 2008).

To date, various families of antibiotics, such as rifampin, isoniazid, and linezolid, are commonly used in clinical settings to treat intracellular bacterial infections. However, these antibiotics are often ineffective in completely eradicating intracellular pathogens. This difficulty in treating intracellular infections is largely due to two factors: the inability of sufficient antibiotics to penetrate infected cells, and the various mechanisms by which bacteria can escape host cells. On the one hand, many antibiotics are hydrophilic and have poor intracellular permeability, which limits their effectiveness in treating intracellular infections. On the other hand, even antibiotics are able to diffuse into cells, they may be inactivated by several factors within cells, such as degradation by acidic, redox, or multienzymatic microenvironment, or discharge by efflux pumps (Wright, 2005). Accordingly, the restricted cellular penetration and intracellular instability of antibiotics cause sub-therapeutic concentrations within cells, resulting in the failure of antiintracellular bacteria and the long-lasting persistence of pathogens.

Phagocytic systems are not only capable of killing invading pathogens, but also acting as a natural shield in some cases, preventing bacteria from being eliminated by antibiotics. However, intracellular bacteria develop some mechanisms to evade the innate immune response. These mechanisms include escaping from endosomal/lysosomal/phagolysosomal compartments to the cytoplasm, preventing the fusion of phagosomes and lysosomes, or developing resistance to the microenvironments bactericidal found in lysosomes/ phagolysosomes. For instance, Salmonella enterica is usually located in late endosomes (Brouillette et al., 2003), while Mycobacterium tuberculosis, the typical intracellular bacterium, survives within phagosomes (Peng et al., 2016). This leads to those antibiotics that can penetrate the cell membranes are unable to eliminate evasive M. tuberculosis due to their failure to concentrate in phagosomes (Onyeji et al., 1994). Taken together, bacteria with different escape mechanisms survive in distinct cell compartments, making it difficult for antibiotics to locate within the appropriate compartments and resulting in inactive antibacterial effects.

Compared with the deficiency of traditional antibiotics against intracellular bacteria, targeted drug delivery systems show promising potentials for the management of intracellular infections through improving the cellular uptake and distribution of antibiotics. With the advances in nanotechnology, a wide variety of nanomaterials have been designed for controlled delivery of antibacterial agents, achieving maximal therapeutic efficiency along with minimizing potential adverse effects. Furthermore, the unique intracellular microenvironment at infected sites, with a low pH, redox potential, abundant H2O2, bacterial and cellular enzymes, can serve as responsive stimulus to realize spatiotemporal drug release of nanomaterials. Thus, exploring sophisticated nanomaterials with unique bioactivities in response to the factors or cues of intracellular microenvironment offers great potentials in eliciting specific responses and functions to kill intracellular bacteria. In these contexts, a deep understanding of the interactions between cells, intracellular bacteria and nanomaterials may provide guidance to develop intelligent nanomaterials with precise environmental responsiveness for efficient and safe management of intracellular infections.

In this review, we identify the key characters and therapeutic targets of intracellular infection microenvironment. We illustrate how the nanomaterials physicochemical properties, such as size, charge, shape and surface functionalization affect the interaction between nanomaterials, cells and bacteria. We introduce the recent progress of nanomaterial-based targeted delivery and controlled release of antibiotics in intracellular infection microenvironment. We highlight the nanomaterials with unique intrinsic properties, such as metal toxicity and enzyme-like activity for the treatment of intracellular bacteria (Figure 1). We discuss the opportunities and challenges of bioactive nanomaterials in addressing intracellular infections.

2 Intracellular bacteria and microenvironment

2.1 Host bactericidal mechanisms

When pathogens invade, macrophages, as one of the major immune cells, fight against infection by expressing a series of receptors to trigger innate immunity. This surveillance recognition is achieved through sensors called pattern recognition receptors (PRRs) that detect pathogen-associated molecular patterns (PAMPs) and damage-related molecular patterns (DAMPs) (Plüddemann et al., 2011; Broz and Monack, 2013). After the detection of pathogens, phagocytosis plays an essential role in anti-bacterial host defense, which is mainly manifested by the effective internalization of pathogens. In such a conversion, the engulfment of pathogens by macrophages and a series of sequent membrane remodeling leads to the formation of a membrane-bound vesicle named phagosome. Following the internalization, it is a clearance process in which the phagosome acquires bactericidal and degradative functions termed phagosomal maturation. At the terminal stage of its maturation, the microenvironment of the phagosome becomes highly acidic, degradative, and oxidative, which all contribute to clearing the invaded pathogens. The low pH in phagosomes is related to the progressive acidification within the vacuole, which is realized by the V-ATPase-mediated proton pump (Flannagan et al., 2009). Meanwhile, this acidic condition favors the subsequent formation of phagolysosomes and optimal enzymatic activity of hydrolases in lysosomes.

Another mechanism to kill bacteria is the delivery of molecules with degradative functions, such as defensins, cathelicidins, lysozyme and hydrolases, into phagosomes. Defensins are able to permeabilize bacteria membranes due to the formation of ion transport channels. Cathelicidins, on the other hand, induce permeabilization acting on the cell wall as well as both the outer and inner membranes of bacteria. In addition, hydrolases targeting carbohydrates and lipids also exist in phagosomes, which degrade a wide range of invading bacterial components (Flannagan et al., 2009).

Phagocytes could also act through a large amount of reactive oxygen species (ROS) and reactive nitrogen species (RNS) produced by the NOX2 NADPH oxidase (Quinn and Gauss, 2004; Minakami

and Sumimoto, 2006) and NOS2 nitric oxide synthase (Fang, 2004). The transfer of electrons from NADPH to molecular oxygen results in the formation of superoxide radicals $(O_2, \overline{})$, which are released into phagosomes (Quinn and Gauss, 2004; Mizushima et al., 2011). After that, O2.- in phagosomes react with H2O2 to produce hydroxyl radicals (·OH) and singlet oxygen (¹O₂) (Minakami and Sumimoto, 2006). It is worth noting that the fusion of the lysosomes and vacuoles leads to partial disruption of the membrane connection, releasing the contents, such as myeloperoxidase, into the phagosomes. Myeloperoxidase within the phagosomes can catalyze abundant H₂O₂ and halogen ions to highly bactericide hypochlorous acid (HClO) and chloramines (Thomas, 1979; Shepherd, 1986). RNS is also essential antimicrobial effectors, which reacts with ROS to destroy pathogens, causing nitrosative stress. The production of RNS starts with NOS catalyzing L-arginine and citrulline to produce nitric oxide (NO), which begins with superoxide to form peroxynitrite (ONOO-), which is a highly reactive species that can directly act with several biological targets and cell components, including lipids, amino acid residues, and DNA bases (Webb et al., 2001). Besides, peroxynitrite also has the ability to get across cell membranes to some subcompartments like phagosomes through anion channel (Fang, 2004).

Accordingly, those produced free radicals and oxidation-state components contribute to protein denaturation and lipid peroxidation by oxidative damage, which leads to irreversible damage to the invading bacteria (Boyle and Randow, 2013). This natural defense process provides ideas for biomimetic strategies to eliminate intracellular bacteria.

2.2 Bacterial defensive mechanisms

Unlike extracellular bacteria, intracellular bacteria that can survive and replicate in host cells, especially macrophages, adapt to challenging intracellular microenvironment and evolve intelligent mechanisms to evade host clearance. Those intracellular bacteria are typically divided into two categories: phagosomal and cytosolic bacteria. Most intracellular bacteria studied to date are stored in phagosomes. In detail, the phagosomes provide a safe haven for bacteria, shielding them from immune system detection, and facilitating their replication using the components within the phagosomes (Cullinane et al., 2008; Lamkanfi and Dixit, 2010). Alternatively, cytosolic bacteria benefit from rich nutrient conditions and a relatively spacious microenvironment, enabling them to survive in the host cell cytoplasm despite the presence of immune defenses. In general, treatments that can eliminate bacteria in phagosomes can also act on cytosolic bacteria. It is much thornier to treat phagosome bacteria than cytosolic ones, so our review focus on phagosome bacteria.

Survival of intracellular bacteria presents three main challenges: evading immune system surveillance, resisting the microenvironment, and evading the phagolysosomal pathway. Numerous strategies, including actin-based cell-to-cell spread, low expression of flagellin, avoidance, blockage and adaptation to the phagolysosomal pathway, are utilized by intracellular pathogens.

Different species of bacteria have their own intracellular lifestyles. The majority of phagosome bacteria have abilities to

prevent phagosomes maturation, the terminal stage before lysosomes fusion (Ray et al., 2009). For instance, some bacteria have evolved metabolic pathways to prevent acidification in phagosomes, or express specific proteins to withstand low pH microenvironment (Park et al., 1996; Vandal et al., 2008; Huang et al., 2009; Martinez et al., 2011). In addition, some bacteria are able to express detoxifying enzymes (Schmidtchen et al., 2002) like catalase or superoxide dismutase to balance ROS/RNS levels within phagosomes (John et al., 2001; Ng et al., 2004), or interfere with the biological function of enzymes that catalyze ROS/RNS production (Mott et al., 2002). As a result, bacteria protect themselves from being destroyed and eliminated by ROS/RNS (Rudel et al., 2010; Ashida et al., 2011).

Other bacteria escape into the cytoplasm, which constitutes a wild and favorable microenvironment, through sophisticated mechanisms, such as escaping from vesicles, and permeabilizing phagosomes. For those bacteria, it is essential to escape the phagosome subcompartments as early as possible after internalization to avoid fusion with lysosomes. In this process, protein secretion plays a key role in allowing bacteria to cross the cytoplasmic membranes, cell walls, vacuole and host cell membranes, to achieve both intracellular vacuole spread and cellto-cell spread. Among them, the four major protein secretion ssystems are the type III secretion system (T3SS) (Cornelis, 2006), type IV secretion systems (T4SSs) (Vogel et al., 1998; Christie and Cascales, 2005), type VI secretion systems (T6SSs) (Coulthurst, 2013; Kudryashev et al., 2015) and type VII secretion system (T7SS) (Abdallah et al., 2007), which hold the potential to serve as inhibitory targets for treatment design.

With these findings in mind, a comprehensive understanding of the mechanisms by which intracellular pathogens evade host cells and respond to the innate immune systems are critical to elucidate the pathogenesis of intracellular infections. Moreover, systematically deciphering the interactions between host cells and intracellular pathogens may provide clues for designing advanced nanotherapeutics against intracellular infections.

3 Interactions between bacteria, cells, and nanomaterials

As discussed above, most intracellular bacteria spend their whole lifestyles within phagosomes. Internalization and phagosomal maturation are essential to keep phagocytosis effective. After being detected and taken up by phagocytes, bacteria undergo a series of membrane fusions and interactions, resulting in their entrapments in sub-compartments such as phagosomes or vacuoles. However, since bacteria may be located in different phagosomes, tailored designed nanomaterials against intracellular pathogens require coexisting in the same compartments. Therefore, a deep understanding of the interactions among bacteria, phagocyte cells, and nanomaterials is essential for the principle of material design.

Different nanomaterials may be located in different sites relative to cells, such as the inner or outer cell membranes, and can influence cell proliferation, apoptosis, and migration. Therefore, understanding the relevant parameters in nanomaterials interactions with cell membranes is essential in regulating nanomaterials internalization. Nanomaterials can be internalized into cells via phagocytosis, diffusion, and fluid phase endocytosis (He et al., 2009). The endocytosis refers to the process by which the plasma membrane invaginates and forms vesicles, thereby transporting extracellular compositions into cells. It is categorized into four major types, including clathrin/caveolae-dependent endocytosis, phagocytosis, pinocytosis, and macropinocytosis (Doherty and McMahon, 2009; Howes et al., 2010; Sahay et al., 2010; Sandvig et al., 2011). Most nanoparticles enter cells through endocytosis, while relatively few enter via other mechanisms. Several factors, including size, shape, surface charge and functionalization, have influence on the uptake of materials by cells.

3.1 Size-dependent cellular uptake

Unlike non-phagocytes, which tend to engulf spherical nanoparticles in the 20–50 nm range, phagocytes preferentially take up micro scale particles (González et al., 1996; Champion et al., 2008; Jiang et al., 2008). Experimental results for silver nanoparticles have shown that well-dispersed 20–200 nm particles were internalized better by non-phagocytes than phagocytes (Lankoff et al., 2012), whereas aggregated silver particles were more likely to be internalized by phagocytes (Wang et al., 2012). The same phenomenon has been found for smaller nanoparticles like iron oxide particles, where small iron oxide particles exhibited a higher level of phagocytic accumulation than ultra-small particle size particles (Raynal et al., 2004).

This difference in internalization may be attributed to the influence of size on internalization pathways. Normally ultrasmall particles are not recognized as exogenous agents by macrophages and can enter directly into cells based on pores in the cell membranes, whereas microscale particles are more likely to be absorbed by the reticuloendothelial system. Besides, smaller sizes possess larger surface areas, which contributes to particles diffuse into cells. When the diameter of the nanospheres is less than 200 nm, their penetration is mainly regulated by the clathrin pathway, but when the size increases to 500 nm, their internalization is mainly mediated by caveolae pathway (Rejman et al., 2004). The effect of size is also significant in the uptake of different sized anionic polystyrene particles, with smaller particles being taken up mainly through clathrin-independent cavelaeindependent pathways, while larger particles are uptaken via clathrin-mediated endocytosis (Lai et al., 2007). This distinction can be explained by the fact that the clathrin-mediated pathway has a higher uptake rate than clathrin-independent cavelae-independent pathways, resulting in particles internalized through this pathway exhibiting faster accumulation within cells.

3.2 Charge-dependent cellular uptake

Surface charges also play a key role in cellular uptake. It is mainly manifested in the fact that the neutral surface charge nanomaterials have a lower plasma protein adsorption rate, accompanied by a longer blood circulation time, which results in a higher cell uptake due to a longer margin from the phagocytes. Positively charged nanomaterials bind to negatively charged cell membranes through electrostatic interactions (Tahara et al., 2009; Duceppe and Tabrizian, 2010). A mass of works has demonstrated that positively charged particles were internalized into cells at a higher degree than their respective anionic particles, such as gold and silver particles, iron oxide particles, silicon dioxide, chitosan, liposome, and polymers. Besides, for negatively charged nanomaterials, the internalization decreases with increasing surface charge, while positively ones, on the contrary, performs a positive correlation with a certain range of surface charge. Surprisingly, a representative example is antibacterial silver nanoparticles coated with chitosan (CS-AgNPs) with enhanced antibacterial effect (Jena et al., 2012). The designed CS-AgNPs exhibited a significantly improved therapeutic effect on intracellular bacteria mainly due to their strong cell internalization.

Several other researches have shown some contradictory results, which might contribute to negatively charged nanoparticles can promote cellular uptake via regulating the formation of aggregation and cluster after initial electrostatic repulsion. Taking the uptake of liposome with different surface charges as an example, a negatively charged PLGA-lipid hybrid system performed better uptake behavior compared to a positively charged system (Maghrebi et al., 2020). Another hypothesis is that bacterial surfaces also present a negative charge, and phagocytes may preferentially take up anionic particles (Fröhlich, 2012). Moreover, some surface groups with negative charges, such as citrate groups, can improve the stability of nanoparticles in culture media and increase their affinity for cell membranes (Kolosnjaj-Tabi et al., 2013).

3.3 Shape-dependent cellular uptake

Shape is another important factor influencing the internalization of nanomaterials, especially when endocytosis pathways of nanomaterials need to be mediated by receptors. Although the surface area of rod-shaped nanomaterials is smaller than that of spherical particles, the limited binding sites on its surface can more efficiently recognize and bind to target cell surface receptors due to its aspect ratio. This allows rod-shaped nanomaterials to exhibit higher cell adhesion efficiency than spherical ones (Kolosnjaj-Tabi et al., 2013; Shao et al., 2017). On the other hand, nanomaterials with sharp shapes such as spines are able to locate in the cytoplasm due to their better ability to penetrate membranes, which enables them to remain in cells benefiting from low exocytosis (Chu et al., 2014).

3.4 Surface functionalization-dependent cellular uptake

Surface functionalization of nanomaterials not only regulates surface charge, but also improves properties such as hydrophobicity and softness. The most common strategy is surface polyethylene glycolation (PEGylation), which is mainly used to reduce the hydrophobicity of nanomaterials and alter their biological characteristics. PEGylation forms a hydrated layer that reduces the adsorption of serum proteins, increases the hydrophilicity, and reduces macrophage uptake, which is often referred to the "stealth effect" (Figures 2A, B) (Yang et al., 2014; Sanchez et al.,



FIGURE 1

Schematic of interactions between cells, intracellular bacteria and nanomaterials during employing multifunctional nanomaterials against intracellular infections. nanomaterials. For example, nanoparticles modified with polysaccharides could interact with specific receptors on cell membranes, resulting in active targeting for more precise internalization. Surface arginine-modified mesoporous silica nanoparticle (MSN) was able to co-localize with intracellular *Salmonella*, leading to efficient antibiotic delivery (Figures 2C–E) (Mudakavi et al., 2017).

4 Nanoparticulate materials against intracellular bacteria

Due to the particularity of intracellular bacteria, three subjects, cells, bacteria and nanomaterials included, should be taken into consideration at the same time for nano-therapy design. For instance, positively charged materials have a higher likelihood of being taken up by cells and can also bind to negatively charged bacterial surfaces through electrostatic interactions. Yang et al. utilized this strategy by designing surface cation-targeted peptidemodified MSN to deliver gentamicin, and the results proved the



Schematic illustration of the surface arginine-modified MSN co-localized with various grating densities by differentiated numan THP-1 Cells. (C) Schematic illustration of the surface arginine-modified MSN co-localized with intracellular *Salmonella*, resulting in efficient antibiotic delivery. (D) Cellular trafficking and mechanism of endocytosis of Arg-MSN in RAW 264.7 cells. (E) Cellular uptake of Arg-MSN particles in the presence of pharmacological inhibitors of endocytosis. Reproduced with permission from Yang et al. (2014) (Copyright 2014 American Chemical Society) and Mudakavi et al. (2017).

2017). Additionally, the difference in uptake may also be related to the change in particle softness by functionalized modification, where softer particles are more likely to be taken up by macrophages. Surface functionalization also enables active targeting of ability of nanoparticles to specifically target *S. aureus* and internalize into RAW 264.7 cells, indicating potentials for fighting against intracellular infections (Yang et al., 2018). Similarly, Maya et al. developed *o*-carboxymethyl-coated chitosan to achieve tetracycline



intracellular MRSA. (E) The pH-sensitive release profile of DDS. (F) The intracellular bactericidal effect of different treatments. Reproduced with permission from Wang et al. (2020) (Copyright 2020 American Chemical Society) and Feng et al. (2022).

targeted delivery to the infectious sites of intracellular *S. aureus* (Maya et al., 2012).

Currently, the strategies of targeting infected cells involve nonspecific electrostatic interactions and specific receptor-ligand interactions, the most representative of which are mannose receptor, CD44 receptor, tuftsin receptor, and hyaluronic acid ligand. On the other hand, direct targeting of bacteria is mainly achieved through non-specific electrostatic interactions, ligandreceptor recognition, and antigen-antibody specific binding.

4.1 Nanocarriers

According to the above discussion, bacteria invade cells and locate in different sub-compartments, making it crucial for nanotherapeutics to deliver antibiotics in on-demand manner. This approach can improve treatment efficacy and reduce toxicity to normal tissues. Furthermore, bacterial invasion creates a unique infection microenvironment characterized by low pH, related enzyme secretion, and slight temperature changes. These conditions can be leveraged as stimulus to achieve drug targeting and release.

4.1.1 pH-responsive release

Progressive acidification of phagosomes and the acidic microenvironment of lysosomes are the most common stimulus for antibiotic release (Ninan et al., 2016; Pei et al., 2017; Pei et al., 2017; Lu et al., 2018; Qu et al., 2018; Su et al., 2018; Mohebali and Abdouss, 2020; Wang et al., 2020). Platensimycin (PTM), a promising natural product

drug, was designed to be loaded in pH responsive release polymers, which demonstrate significantly reduce residual methicillin-resistant Staphylococcus aureus (MRSA) in macrophage cells (Figures 3A-C). Antituberculosis drug-loaded MSNs equipped with a pH-sensitive valve (\beta-cyclodextrin) were constructed to optimize loading and achieve specific intracellular delivery of drug for the treatment of tuberculosis (TB) (Clemens et al., 2012). Greater therapeutic efficacy was achieved which may be attributed to the fact that it can only be released in acidified phagosomal conditions. Other drug delivery systems (DDS) were synthesized to realize cascade release of rifampicin after Schiff base cleavage in acidic phagolysosome (Figures 3D-F) (Feng et al., 2022). This DDS in a cascade manner performed outstanding targeting and killing activity against MRSA inside macrophages. In addition, some nanomaterials with tailored surface charge have been prepared. This particular type of nanoparticles typically perform positive surface charges only under acidic conditions, which allows them to better cross the cell membranes and bind to bacteria with negatively charged surfaces. In such a scenario, nanomaterials not only exhibit efficient internalization, but also have lower toxicity to normal cells. This is because cationic particles are more likely to cause hemolysis and cytotoxicity, which can lead to lysosomal and mitochondrial damage and cell membranes destruction.

4.1.2 Enzymes-responsive release

As mentioned earlier, during the process of phagosomes maturation, various enzymes like lipase (Jaeger et al., 1994; Jaeger and Reetz, 1998), phosphatase or phospholipase (DeVinney et al., 2000) are secreted and recruited into phagosomes. Accordingly,



FIGURE 4

(A) Schematics of Gen@MSN-LU, in which the outer layer of liposomes can be degraded by the bacterium-secreted toxins, resulting in the Gen release. (B) The release profile of Gen from Gen@MSN-LU in different treatments (A control; B in the presence of *S. aureus*; C with lipase; D with lipase inhibitors in the presence of *S. aureus*). (C) Schematics of bacterial lipase triggered release of vancomycin to treat the bacterial infections. (D,E) Cumulative release of vancomycin from vancomycin-loaded TLN in different treatments. (F,G) The behavior of the redox-triggered release of Moxifloxacin. Reproduced with permission from Yang et al. (2018) (Copyright 2018 American Chemical Society), Xiong et al. (2012a) (Copyright 2012 American Chemical Society), and Lee et al. (2016).

these cellular enzymes (Alkekhia et al., 2022), bacterial enzymes (Sunnapu et al., 2022) and bacterium-secreted toxins (Figures 4A, B) (Yang et al., 2018) can also be used as stimulators for corresponding release. Peng et al. (2020) provided a practicable strategy to realize intracellular rapid release of antibiotics triggered by both enzymes and acid, thus promoting efficiency against intracellular infections. A rationally designed polymer, which was able to be degraded by bacterial lipase, significantly increased intracellular concentration of ciprofloxacin, thereby enhancing its therapeutic efficacy.

In addition, some nanogels were designed to respond to enzymes (Chen et al., 2020). Xiong et al. prepared two different feasible nanogels, a triple-layered polymer nanogel (Figures 4C–E) (Xiong et al., 2012a) and a core-shell mannose-modified nanogel (Xiong et al., 2012b), to inhibit the growth of intracellular *S. aureus*. In two systems, bacteria-secreting lipase and phosphatase or phospholipase were acted as triggers to release antibiotics, respectively.

4.1.3 Redox-responsive release

Nanomaterials that can respond to the redox environment could be a potential therapeutic strategy for the treatment of intracellular infections. A functionalized MSN was used to selectively release drugs intracellularly in response to the GSH/GSSG species (reduced glutathione/oxidized glutathione disulfide), which were commonly found intracellularly (Figures 4F, G) (Lee et al., 2016). The reducing microenvironment can also be utilized to break the disulfide bonds of red blood cell (RBC) nanogels, resulting in the rapid release of antibiotics (Zhang et al., 2017).

4.2 Bioactive nanomaterials

Beyond the application as drug delivery carriers, some nanomaterials with intrinsic antimicrobial bioactivities have recently shown promising aspects for potential clinical applications against intracellular bacteria.

4.2.1 Metal ion-based nanobactericides

Given that metals are traditionally used as drug delivery carriers, recent studies have also shown that some biomaterials with intrinsic antimicrobial bioactivity hold promise against intracellular infections. For example, gold nanomaterials can reduce the attachment of tRNA to ribosome units, decrease membrane viability and cause bacterial death. Copper ions can inhibit bacterial DNA replication and induce bacterial death through ROS production and lipid peroxidation. The release of metal ions, such as Ag^+ , Cu^{2+} , and Fe^{2+} , also can enhance the antibacterial effect. For instance, ZnO has the ability to inhibit the formation of biofilms and catalyze the production of ROS. Besides, the release of Zn^{2+} can also increase the permeability and degradability of membranes.

The most widely used metal for antibacterial activity is silver, which exerts its effects through a variety of mechanisms, such as lipid peroxidation, ROS generation, interference with cell wall synthesis, and increasing membrane permeability. Besides, Ag^+ hydrolyzes bacterial macromolecules by driving the generation of hydroxyl radicals. Aurore et al. (2018) designed Ag-NPs with synergistic antimicrobial effects against *S. aureus* in human



(A) Schematic illustration of the bioactive nanoparticles with collaborative bactericidal function depends on both ferrous iron and polysulfide. (B) Schematic illustration of the bacteriostatic effects of IONzymes with enzymes-like activity and mechanism responsible for the antibacterial activity. (C) Schematic illustration of the peptide-chlorophyll-based photodynamic therapy (PDT) agents to enhance the PDT effect and active targeting property to eliminate intracellular infections. (D) Confocal images of intracellular ROS production by MPepP₁₈(Cu²⁺) in *S. aureus* infected macrophage. (E) Schematic illustration of the biodegradable multi-metallic particles (MMPs), containing Ag NPs and ZnO NPs, and its pulmonary delivery of antituberculous drugs to the endosomal system of *M.tb*-infected macrophages. (F) MMPs embedded with Ag NPs and ZnO NPs within the endosome of an *M.tb*-infected alveolar macrophage. Reproduced with permission from Shen et al. (2020); Shi et al. (2018); Cai et al. (2018) (Copyright 2018 American Chemical Society) and Ellis et al. (2018).

osteoclasts. The promoted bactericidal activity not only came from the direct toxicity of silver itself but also from the ROS production in osteoclasts induced by Ag-NPs. Taken together, these surprising results indicate that silver nanoparticles can be used as an effective treatment for chronic long-term infections caused by intracellular bacteria, where conventional antibiotics are difficult to achieve equivalent therapeutic effects to their extracellular effects.

Ferrous ions are another type of metal ions that have shown great promise in terms of their antimicrobial biological activity, as they play a critical role in ferroptosis. In a recent study by Shen et al., discovered a multi-nanomaterials combined with ferrous iron and polysulfide (Fe(II)S_naq) (Figure 5A) (Shen et al., 2020). The nanomaterials were found to be effective in killing both extracellular and intracellular bacteria, while only having slight toxicity towards host cells. In this system, the sulfur atoms were replaced by oxygen atoms to trigger the release of polysulfides and iron. The collaborative function depended on both ferrous iron and polysulfide. Ferrous irons were able to trigger lipid peroxidation and depress the respiratory chain, which induced ferroptosis-like death within bacteria. At the same time, polysulfide species prevented oxidation of ferrous irons, and demonstrated the ability to oxidize

glutathione into GSSG, following with GSH depletion, which leads to DNA degradation and bacteria death.

4.2.2 Nanozymes

The natural process of scavenging pathogens by macrophages involves converting a large amount of hydrogen peroxide in phagosomes into more toxic ROS, catalyzed by enzymes such as oxidase, myeloperoxidase, lipid peroxidase, and other peroxidases. Inspired by this process, there are increased efforts devoted to build artificial enzymes with natural enzyme-like activity to combat extracellular bacteria. Based on their own instinct oxidase enzyme activity, nanozymes can provide various bactericidal ROS, including singlet oxygen, superoxide anion, hypochlorous acid and other oxidation-state components with non- or lowtoxic substrates. These excess ROS are capable of damaging bacterial DNA, protein, or nucleic acid, along with breaking cell membranes. Nevertheless, development of nanozymes for efficient production of ROS against intracellular bacteria remains a significant challenge. With these findings in mind, the integration of ROS production and antibiotic delivery may be a promising strategy for efficient elimination both intracellular and extracellular pathogens, especially towards phagosomal bacteria.

Peroxidase is a series of enzymes that catalyze the substrates like hydrogen peroxide into hydroxyl radicals, which are essential in defending pathogens. Presently, enormous nanomaterials have been reported to exhibit the peroxidase-like catalytic activity, including metal, metal sulfide, metal oxide, metal organic frameworks, inorganic materials, and carbon-based materials. Shi et al. designed an iron oxide nanozyme (IONzymes) that can destroy structures, inhibit multiplication, and ultimately cause death of intracellular bacterial (Figure 5B) (Shi et al., 2018). In this work, IONzymes were able to co-localize with intracellular *S. enteritidis* in autophagic vacuoles and regulate ROS levels within acid vacuoles. Moreover, the increasing ROS levels suppressed the survival and reproduction of those pathogens hiding in Leghorn Male Hepatoma-derived cells (LMH).

Oxidases are the major enzymes in peroxisomes, accounting for almost half of the total ones. Generally, the catalytic process of oxidases requires the participation of oxygen, followed by the generation of superoxide anions (Wu et al., 2019). Therefore, nanozymes with oxidase-mimic activity can also be utilized as anti-bacterial materials. Haloperoxidase is another typical category of peroxidase in nature, divided into three subtypes: chloroperoxidase, bromoperoxidase, and iodoperoxidase (ten Brink et al., 2000). Particularly, haloperoxidase can catalyze halide ions in the physiologic environment to hypohalous acid in the presence of acidic and hydrogen peroxide (Hu et al., 2018). Hypohalous acid is a strong oxidizing agent, which can effectively damage bacterial structure (Butler, 1999). Accordingly, haloperoxidase-like nanozymes also have the promising potential for eliminating intracellular bacteria.

Collectively, a vast array of nanozymes present bi-enzymatic and even tri-enzymatic synergism activities, making it preferable to design them as functional components of composite materials.

4.2.3 Photo-active nanomaterials

Photodynamic therapy (PDT) is a method of using photosensitizers remaining in cells to produce singlet oxygen and free radicals to chemically eliminate bacteria. Some biomaterials with PDT effects are used for their active targeting properties to eliminate intracellular infections. Cai et al. (2018) presented a PDT-based strategy to clear *S. aureus* inside macrophages (Figures 5C, D). The dimer coated with peptide–chlorophyll was enabled to active targeting of macrophages, which can generate abundant ROS in infected macrophages with the laser, resulting in high-efficiency photodynamic elimination. This design demonstrated the potential of photodynamic and photothermal effects in intracellular bacterial clearance.

4.3 Multifunctional nanomaterials

From above mentioned findings, these bioactive nanomaterials can serve as alternative treatments to traditional antibiotics due to their multiple mechanisms involving intrinsic enzyme-mimic activities, metal toxicity, or physicochemical properties, making them less prone to cause antibiotic resistance. To achieve more efficient synergistic antibacterial effects, researchers are exploring the application of multifunctional nanomaterials integrating the functions of delivery with antibacterial activities (Chang et al., 2017a; Chang et al., 2017b; Lu et al., 2017; Lu et al., 2018; Zhang et al., 2020; Huo et al., 2021; Li et al., 2021; Liu et al., 2021).

Dube et al. presented nanoparticles with chitosan-functionalized shells and PLGA cores for the treatment of tuberculosis (Dube et al., 2014). This rationally designed nanoparticle showed a collaborative function between stimulation of ROS/RNS and delivery of rifampicin. In this case, the combined nanoparticles can not only regulate ROS/RNS levels but also modulate pro-inflammatory cytokine secretion. Meanwhile, it acted as a vehicle to transfer rifampicin inside alveolar macrophages. Similarly, Marwa et al. prepared Ag-coated PLGA particles loaded with pexiganan, which exhibited coordinated antibacterial functions, and were specifically uptaken by macrophages, but not by any nonphagocytic cells (Elnaggar et al., 2020). Additionally, Timothy et al. established composite materials containing Ag and ZnO for the targeted delivery of rifampicin (Figures 5E, F) (Ellis et al., 2018). The sophisticated nanoplatforms were developed to destabilize membranes. increase permeabilization of intracellular Mycobacterium tuberculosis and enhance penetration of rifampicin. As a result, multifunctional nanomaterials presented extraordinary bactericidal effects compared to free antibiotics.

5 Conclusion and future perspectives

Nanomaterials can enhance preferential accumulation and controlled release of bactericides within pathogens-infected host cells, resulting in increased therapeutic efficiency and reduced potential adverse effects. Size plays a crucial role in the ability of nanomaterial to permeate cells and reach the therapeutic concentration. Positively charged nanomaterials perform excellent interactions with negative-surface bacteria. Moreover, surfacefunctionalized nanomaterials with special ligands are conducive to an improving targeting of the infected cells. Responsive release at the diseased site can be triggered via pH, enzymes, and redox microenvironment. Beyond providing on-demand delivery of antibiotics, nanomaterials with intrinsic bioactivities, such as metal toxicity, enzyme-like activity and physicochemical properties also show promising potential in combating intracellular pathogens.

However, fabrication of multifunctional nanomaterials for eliminating bacteria is still in infancy, with several fundamental concerns unclear and numerous challenges to be addressed. Positively charged nanomaterials generally present a better cell uptake and adsorption with negative-surface bacteria, but cationic particles tend to cause hemolysis and cytotoxicity. Besides, there are also some examples of negative charged nanomaterials exhibit a higher internalization. Thus, it is worthy to clarify the detailed and wide functions of surface charge in different nanomaterials to balance cell uptake and safety. Despite the effects of size, charge, shape and surface modifications on internalization have been extensively studied, the mechanisms of other properties, such as smoothness and hydrophily, affecting internalization remain indistinct. A comprehensive understanding of the interactions between bacteria, nanomaterials and cellular microenvironment may provide insights into the development of intelligent nanomaterials with precise release property and high level of safety. Besides, nanozyme-based composite materials perform poor selectivity compared with natural enzymes, which might cause undesired toxicity. Additionally, most nanozymes, especially haloperoxidase-mimic ones, are only effective in an acidic microenvironment, limiting their further application. How to

compound bioactive and delivering materials to realize synergistic therapeutic effects is highly desired. Lastly, biosafety remains a major concern that determining the translation of nanomaterials against intracellular bacterial infection in clinic.

In summary, we review recent studies on employing multifunctional nanomaterials in treating intracellular infections through targeted delivery of anti-bacterial agents or utilizing their intrinsic bioactivities. We also discuss the opportunities and challenges that need to be focused on in future work. Broadening understanding of the interactions among macrophages, intracellular pathogens and nanoparticles contributes to inspire the development of the next-generation of nanomaterial-based therapeutics against intracellular bacterial infections.

Author contributions

YC and XH contributed equally to this work. YC and CY: conceptualization. YC and XH: original draft preparation. XH, QC, YH, FC, and CY: review and editing. LW: supervision and review. All authors contributed to the article and approved the submitted version.

References

Abdallah, A. M., Gey van Pittius, N. C., DiGiuseppe Champion, P. A., Cox, J., Luirink, J., Vandenbroucke-Grauls, C. M., et al. (2007). Type VII secretion—Mycobacteria show the way. *Nat. Rev. Microbiol.* 5 (11), 883–891. doi:10.1038/nrmicro1773

Alkekhia, D., LaRose, C., and Shukla, A. (2022). β-Lactamase-Responsive hydrogel drug delivery platform for bacteria-triggered cargo release. *ACS Appl. Mater. Interfaces* 14 (24), 27538–27550. doi:10.1021/acsami.2c02614

Ashida, H., Mimuro, H., Ogawa, M., Kobayashi, T., Sanada, T., Kim, M., et al. (2011). Cell death and infection: A double-edged sword for host and pathogen survival. *J. Cell Biol.* 195 (6), 931–942. doi:10.1083/jcb.201108081

Aurore, V., Caldana, F., Blanchard, M., Hess, S. K., Lannes, N., Mantel, P.-Y., et al. (2018). Silver-nanoparticles increase bactericidal activity and radical oxygen responses against bacterial pathogens in human osteoclasts. *Nanomedicine Nanotechnol. Biol. Med.* 14 (2), 601–607. doi:10.1016/j.nano.2017.11.006

Boyle, K. B., and Randow, F. (2013). The role of 'eat-me'signals and autophagy cargo receptors in innate immunity. *Curr. Opin. Microbiol.* 16 (3), 339–348. doi:10.1016/j.mib. 2013.03.010

Briones, E., Colino, C. I., and Lanao, J. M. (2008). Delivery systems to increase the selectivity of antibiotics in phagocytic cells. *J. Control. Release* 125 (3), 210–227. doi:10. 1016/j.jconrel.2007.10.027

Brouillette, E., Grondin, G., Shkreta, L., Lacasse, P., and Talbot, B. G. (2003). *In vivo* and *in vitro* demonstration that *Staphylococcus aureus* is an intracellular pathogen in the presence or absence of fibronectin-binding proteins. *Microb. Pathog.* 35 (4), 159–168. doi:10.1016/s0882-4010(03)00112-8

Broz, P., and Monack, D. M. (2013). Newly described pattern recognition receptors team up against intracellular pathogens. *Nat. Rev. Immunol.* 13 (8), 551–565. doi:10. 1038/nri3479

Butler, A. (1999). Vanadium haloperoxidases. *Bioinorg. Catal.* 1999, 71-96. doi:10. 1016/S1367-5931(98)80070-7

Cai, Q., Fei, Y., An, H. W., Zhao, X.-X., Ma, Y., Cong, Y., et al. (2018). Macrophageinstructed intracellular *Staphylococcus aureus* killing by targeting photodynamic dimers. ACS Appl. Mater. interfaces 10 (11), 9197–9202. doi:10.1021/acsami.7b19056

Champion, J. A., Walker, A., and Mitragotri, S. (2008). Role of particle size in phagocytosis of polymeric microspheres. *Pharm. Res.* 25, 1815–1821. doi:10.1007/s11095-008-9562-y

Chang, Z., Wang, Z., Lu, M. m., Shao, D., Yue, J., Yang, D., et al. (2017a). Janus silver mesoporous silica nanobullets with synergistic antibacterial functions. *Colloids Surfaces B Biointerfaces* 157, 199–206. doi:10.1016/j.colsurfb.2017.05.079

Chang, Z., Wang, Z., Lu, M., Li, M., Li, L., Zhang, Y., et al. (2017b). Magnetic Janus nanorods for efficient capture, separation and elimination of bacteria. *RSC Adv.* 7 (6), 3550–3553. doi:10.1039/C6RA27296G

Chen, M., He, J., Xie, S., Wang, T., Ran, P., Zhang, Z., et al. (2020). Intracellular bacteria destruction via traceable enzymes-responsive release and deferoxamine-

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Conflict of interest

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mediated ingestion of antibiotics. J. Control. Release 322, 326-336. doi:10.1016/j. jconrel.2020.03.037

Christie, P. J., and Cascales, E. (2005). Structural and dynamic properties of bacterial Type IV secretion systems (Review). *Mol. Membr. Biol.* 22 (1-2), 51–61. doi:10.1080/09687860500063316

Chu, Z., Zhang, S., Zhang, B., Zhang, C., Fang, C.-Y., Rehor, I., et al. (2014). Unambiguous observation of shape effects on cellular fate of nanoparticles. *Sci. Rep.* 4 (1), 4495. doi:10.1038/srep04495

Clemens, D. L., Lee, B.-Y., Xue, M., Thomas, C. R., Meng, H., Ferris, D., et al. (2012). Targeted intracellular delivery of antituberculosis drugs to Mycobacterium tuberculosis-infected macrophages via functionalized mesoporous silica nanoparticles. *Antimicrob. agents Chemother.* 56 (5), 2535–2545. doi:10.1128/AAC. 06049-11

Cornelis, G. R. (2006). The type III secretion injectisome. Nat. Rev. Microbiol. 4 (11), 811–825. doi:10.1038/nrmicro1526

Coulthurst, S. J. (2013). The Type VI secretion system-a widespread and versatile cell targeting system. *Res. Microbiol.* 164 (6), 640–654. doi:10.1016/j.resmic.2013.03.017

Cullinane, M., Gong, L., Li, X., Adler, N.-L., Tra, T., Wolvetang, E., et al. (2008). Stimulation of autophagy suppresses the intracellular survival of Burkholderia pseudomallei in mammalian cell lines. *Autophagy* 4 (6), 744–753. doi:10.4161/ auto.6246

DeVinney, R., Steele-Mortimer, O., and Finlay, B. B. (2000). Phosphatases and kinases delivered to the host cell by bacterial pathogens. *Trends Microbiol.* 8 (1), 29–33. doi:10. 1016/s0966-842x(99)01657-1

Doherty, G. J., and McMahon, H. T. (2009). Mechanisms of endocytosis. Annu. Rev. Biochem. 78, 857–902. doi:10.1146/annurev.biochem.78.081307.110540

Dube, A., Reynolds, J. L., Law, W.-C., Maponga, C. C., Prasad, P. N., and Morse, G. D. (2014). Multimodal nanoparticles that provide immunomodulation and intracellular drug delivery for infectious diseases. *Nanomedicine Nanotechnol. Biol. Med.* 10 (4), 831–838. doi:10.1016/j.nano.2013.11.012

Duceppe, N., and Tabrizian, M. (2010). Advances in using chitosan-based nanoparticles for *in vitro* and *in vivo* drug and gene delivery. *Expert Opin. drug Deliv.* 7 (10), 1191–1207. doi:10.1517/17425247.2010.514604

Ellis, T., Chiappi, M., Garcia-Trenco, A., Al-Ejji, M., Sarkar, S., Georgiou, T. K., et al. (2018). Multimetallic microparticles increase the potency of rifampicin against intracellular *Mycobacterium tuberculosis. ACS Nano* 12 (6), 5228–5240. doi:10.1021/acsnano.7b08264

Elnaggar, M. G., Jiang, K., Eldesouky, H. E., Pei, Y., Park, J., Yuk, S. A., et al. (2020). Antibacterial nanotruffles for treatment of intracellular bacterial infection. *Biomaterials* 262, 120344. doi:10.1016/j.biomaterials.2020.120344

Fang, F. C. (2004). Antimicrobial reactive oxygen and nitrogen species: Concepts and controversies. *Nat. Rev. Microbiol.* 2 (10), 820–832. doi:10.1038/nrmicro1004

Feng, W., Li, G., Kang, X., Wang, R., Liu, F., Zhao, D., et al. (2022). Cascade-targeting poly (amino acid) nanoparticles eliminate intracellular bacteria via on-site antibiotic delivery. *Adv. Mater.* 34 (12), 2109789. doi:10.1002/adma.202109789

Flannagan, R. S., Cosío, G., and Grinstein, S. (2009). Antimicrobial mechanisms of phagocytes and bacterial evasion strategies. *Nat. Rev. Microbiol.* 7 (5), 355–366. doi:10. 1038/nrmicro2128

Fröhlich, E. (2012). The role of surface charge in cellular uptake and cytotoxicity of medical nanoparticles. *Int. J. nanomedicine* 7, 5577–5591. doi:10.2147/IJN.S36111

González, O., Smith, R. L., and Goodman, S. B. (1996). Effect of size, concentration, surface area, and volume of polymethylmethacrylate particles on human macrophages *in vitro. J. Biomed. Mater. Res.* 30 (4), 463–473. doi:10.1002/(SICI)1097-4636(199604) 30:4<463:AID-JBM4>3.0.CO;2-N

He, Q., Zhang, Z., Gao, Y., Shi, J., and Li, Y. (2009). Intracellular localization and cytotoxicity of spherical mesoporous silica nano- and microparticles. *Small* 5 (23), 2722–2729. doi:10.1002/smll.200900923

Howes, M. T., Mayor, S., and Parton, R. G. (2010). Molecules, mechanisms, and cellular roles of clathrin-independent endocytosis. *Curr. Opin. cell Biol.* 22 (4), 519–527. doi:10.1016/j.ceb.2010.04.001

Hu, M., Korschelt, K., Viel, M., Wiesmann, N., Kappl, M., Brieger, J. r., et al. (2018). Nanozymes in nanofibrous mats with haloperoxidase-like activity to combat biofouling. ACS Appl. Mater. interfaces 10 (51), 44722–44730. doi:10.1021/acsami. 8b16307

Huang, J., Canadien, V., Lam, G. Y., Steinberg, B. E., Dinauer, M. C., Magalhaes, M. A., et al. (2009). Activation of antibacterial autophagy by NADPH oxidases. *Proc. Natl. Acad. Sci.* 106 (15), 6226–6231. doi:10.1073/pnas.0811045106

Huo, J., Jia, Q., Huang, H., Zhang, J., Li, P., Dong, X., et al. (2021). Emerging photothermal-derived multimodal synergistic therapy in combating bacterial infections. *Chem. Soc. Rev.* 50 (15), 8762–8789. doi:10.1039/D1CS00074H

Jaeger, K.-E., Ransac, S., Dijkstra, B. W., Colson, C., van Heuvel, M., and Misset, O. (1994). Bacterial lipases. *FEMS Microbiol. Rev.* 15 (1), 29–63. doi:10.1111/j.1574-6976. 1994.tb00121.x

Jaeger, K.-E., and Reetz, M. T. (1998). Microbial lipases form versatile tools for biotechnology. Trends Biotechnol. 16 (9), 396-403. doi:10.1016/S0167-7799(98)01195-0

Jena, P., Mohanty, S., Mallick, R., Jacob, B., and Sonawane, A. (2012). Toxicity and antibacterial assessment of chitosancoated silver nanoparticles on human pathogens and macrophage cells. *Int. J. nanomedicine* 7, 1805–1818. doi:10.2147/IJN.S28077

Jiang, W., Kim, B. Y., Rutka, J. T., and Chan, W. C. (2008). Nanoparticle-mediated cellular response is size-dependent. *Nat. Nanotechnol.* 3 (3), 145–150. doi:10.1038/nnano.2008.30

John, G. S., Brot, N., Ruan, J., Erdjument-Bromage, H., Tempst, P., Weissbach, H., et al. (2001). Peptide methionine sulfoxide reductase from *Escherichia coli* and *Mycobacterium tuberculosis* protects bacteria against oxidative damage from reactive nitrogen intermediates. *Proc. Natl. Acad. Sci.* 98 (17), 9901–9906. doi:10.1073/pnas. 161295398

Kamaruzzaman, N. F., Kendall, S., and Good, L. (2017). Targeting the hard to reach: Challenges and novel strategies in the treatment of intracellular bacterial infections. *Br. J. Pharmacol.* 174 (14), 2225–2236. doi:10.1111/bph.13664

Kolosnjaj-Tabi, J., Wilhelm, C., Clément, O., and Gazeau, F. (2013). Cell labeling with magnetic nanoparticles: Opportunity for magnetic cell imaging and cell manipulation. *J. nanobiotechnology* 11 (1), S7–S19. doi:10.1186/1477-3155-11-S1-S7

Kudryashev, M., Wang, R. Y.-R., Brackmann, M., Scherer, S., Maier, T., Baker, D., et al. (2015). Structure of the type VI secretion system contractile sheath. *Cell* 160 (5), 952–962. doi:10.1016/j.cell.2015.01.037

Lai, S. K., Hida, K., Man, S. T., Chen, C., Machamer, C., Schroer, T. A., et al. (2007). Privileged delivery of polymer nanoparticles to the perinuclear region of live cells via a non-clathrin, non-degradative pathway. *Biomaterials* 28 (18), 2876–2884. doi:10.1016/j. biomaterials.2007.02.021

Lamkanfi, M., and Dixit, V. M. (2010). Manipulation of host cell death pathways during microbial infections. *Cell host microbe* 8 (1), 44–54. doi:10.1016/j.chom.2010. 06.007

Lankoff, A., Sandberg, W. J., Wegierek-Ciuk, A., Lisowska, H., Refsnes, M., Sartowska, B., et al. (2012). The effect of agglomeration state of silver and titanium dioxide nanoparticles on cellular response of HepG2, A549 and THP-1 cells. *Toxicol. Lett.* 208 (3), 197–213. doi:10.1016/j.toxlet.2011.11.006

Lee, B. Y., Li, Z., Clemens, D. L., Dillon, B. J., Hwang, A. A., Zink, J. I., et al. (2016). Redox-triggered release of moxifloxacin from mesoporous silica nanoparticles functionalized with disulfide snap-tops enhances efficacy against pneumonic tularemia in mice. *Small* 12 (27), 3690–3702. doi:10.1002/smll.201600892

Li, H., Li, D., Chen, F., Yang, C., Li, X., Zhang, Y., et al. (2021). Nanosilver-decorated biodegradable mesoporous organosilica nanoparticles for GSH-responsive gentamicin release and synergistic treatment of antibiotic-resistant bacteria. *Int. J. Nanomedicine* 16, 4631–4642. doi:10.2147/IJN.S315067

Liu, F., Sheng, S., Shao, D., Xiao, Y., Zhong, Y., Zhou, J., et al. (2021). Targeting multiple mediators of sepsis using multifunctional tannic acid-Zn2+-gentamicin nanoparticles. *Matter* 4 (11), 3677–3695. doi:10.1016/j.matt.2021.09.001

Lu, M.-m., Ge, Y., Qiu, J., Shao, D., Zhang, Y., Bai, J., et al. (2018). Redox/pH dualcontrolled release of chlorhexidine and silver ions from biodegradable mesoporous silica nanoparticles against oral biofilms. *Int. J. nanomedicine* 13, 7697–7709. doi:10. 2147/IJN.S181168

Lu, M.-m., Wang, Q.-j., Chang, Z.-m., Wang, Z., Zheng, X., Shao, D., et al. (2017). Synergistic bactericidal activity of chlorhexidine-loaded, silver-decorated mesoporous silica nanoparticles. *Int. J. nanomedicine* 12, 3577–3589. doi:10.2147/ IJN.S133846

Maghrebi, S., Jambhrunkar, M., Joyce, P., and Prestidge, C. A. (2020). Engineering PLGA–lipid hybrid microparticles for enhanced macrophage uptake. *ACS Appl. Bio Mater.* 3 (7), 4159–4167. doi:10.1021/acsabm.0c00251

Martinez, J., Almendinger, J., Oberst, A., Ness, R., Dillon, C. P., Fitzgerald, P., et al. (2011). Microtubule-associated protein 1 light chain 3 alpha (LC3)-associated phagocytosis is required for the efficient clearance of dead cells. *Proc. Natl. Acad. Sci.* 108 (42), 17396–17401. doi:10.1073/pnas.1113421108

Maya, S., Indulekha, S., Sukhithasri, V., Smitha, K., Nair, S. V., Jayakumar, R., et al. (2012). Efficacy of tetracycline encapsulated O-carboxymethyl chitosan nanoparticles against intracellular infections of Staphylococcus aureus. *Int. J. Biol. Macromol.* 51 (4), 392–399. doi:10.1016/j.ijbiomac.2012.06.009

Minakami, R., and Sumimoto, H. (2006). Phagocytosis-coupled activation of the superoxide-producing phagocyte oxidase, a member of the NADPH oxidase (nox) family. *Int. J. Hematol.* 84, 193–198. doi:10.1532/JJH97.06133

Mizushima, N., Yoshimori, T., and Ohsumi, Y. (2011). The role of Atg proteins in autophagosome formation. *Annu. Rev. cell Dev. Biol.* 27, 107–132. doi:10.1146/annurev-cellbio-092910-154005

Mohebali, A., and Abdouss, M. (2020). Layered biocompatible pH-responsive antibacterial composite film based on HNT/PLGA/chitosan for controlled release of minocycline as burn wound dressing. *Int. J. Biol. Macromol.* 164, 4193–4204. doi:10. 1016/j.ijbiomac.2020.09.004

Mott, J., Rikihisa, Y., and Tsunawaki, S. (2002). Effects of Anaplasma phagocytophila on NADPH oxidase components in human neutrophils and HL-60 cells. *Infect. Immun.* 70 (3), 1359–1366. doi:10.1128/IAI.70.3.1359-1366. 2002

Mudakavi, R. J., Vanamali, S., Chakravortty, D., and Raichur, A. M. (2017). Development of arginine based nanocarriers for targeting and treatment of intracellular Salmonella. *RSC Adv.* 7 (12), 7022–7032. doi:10.1039/C6RA27868J

Ng, V. H., Cox, J. S., Sousa, A. O., MacMicking, J. D., and McKinney, J. D. (2004). Role of KatG catalase-peroxidase in mycobacterial pathogenesis: Countering the phagocyte oxidative burst. *Mol. Microbiol.* 52 (5), 1291–1302. doi:10.1111/j.1365-2958.2004. 04078.x

Ninan, N., Forget, A., Shastri, V. P., Voelcker, N. H., and Blencowe, A. (2016). Antibacterial and anti-inflammatory pH-responsive tannic acid-carboxylated agarose composite hydrogels for wound healing. ACS Appl. Mater. interfaces 8 (42), 28511–28521. doi:10.1021/acsami.6b10491

Onyeji, C., Nightingale, C., Nicolau, D., and Quintiliani, R. (1994). Efficacies of liposome-encapsulated clarithromycin and ofloxacin against Mycobacterium avium-M. intracellulare complex in human macrophages. *Antimicrob. agents Chemother.* 38 (3), 523–527. doi:10.1128/AAC.38.3.523

Park, Y. K., Bearson, B., Bang, S. H., Bang, I. S., and Foster, J. W. (1996). Internal pH crisis, lysine decarboxylase and the acid tolerance response of *Salmonella typhimurium*. *Mol. Microbiol.* 20 (3), 605–611. doi:10.1046/j.1365-2958.1996. 5441070.x

Pei, Y., Mohamed, M. F., Seleem, M. N., and Yeo, Y. (2017). Particle engineering for intracellular delivery of vancomycin to methicillin-resistant *Staphylococcus aureus* (MRSA)-infected macrophages. *J. Control. Release* 267, 133–143. doi:10.1016/j. jconrel.2017.08.007

Peng, H., Xie, B., Yang, X., Dai, J., Wei, G., and He, Y. (2020). Pillar [5] arene-based, dual pH and enzyme responsive supramolecular vesicles for targeted antibiotic delivery against intracellular MRSA. *Chem. Commun.* 56 (58), 8115–8118. doi:10.1039/D0CC02522D

Peng, X., Jiang, G., Liu, W., Zhang, Q., Qian, W., and Sun, J. (2016). Characterization of differential pore-forming activities of ESAT-6 proteins from *Mycobacterium tuberculosis* and Mycobacterium smegmatis. *FEBS Lett.* 590 (4), 509–519. doi:10. 1002/1873-3468.12072

Plüddemann, A., Mukhopadhyay, S., and Gordon, S. (2011). Innate immunity to intracellular pathogens: Macrophage receptors and responses to microbial entry. *Immunol. Rev.* 240 (1), 11–24. doi:10.1111/j.1600-065X.2010.00989.x

Qu, J., Zhao, X., Ma, P. X., and Guo, B. (2018). Injectable antibacterial conductive hydrogels with dual response to an electric field and pH for localized "smart" drug release. *Acta biomater.* 72, 55–69. doi:10.1016/j.actbio.2018.03.018

Quinn, M. T., and Gauss, K. A. (2004). Structure and regulation of the neutrophil respiratory burst oxidase: Comparison with nonphagocyte oxidases. *J. Leukoc. Biol.* 76 (4), 760–781. doi:10.1189/jlb.0404216

Ray, K., Marteyn, B., Sansonetti, P. J., and Tang, C. M. (2009). Life on the inside: The intracellular lifestyle of cytosolic bacteria. *Nat. Rev. Microbiol.* 7 (5), 333–340. doi:10. 1038/nrmicro2112

Raynal, I., Prigent, P., Peyramaure, S., Najid, A., Rebuzzi, C., and Corot, C. (2004). Macrophage endocytosis of superparamagnetic iron oxide nanoparticles: Mechanisms and comparison of ferumoxides and ferumoxtran-10. *Investig. Radiol.* 39 (1), 56–63. doi:10.1097/01.rli.0000101027.57021.28

Rejman, J., Oberle, V., Zuhorn, I. S., and Hoekstra, D. (2004). Size-dependent internalization of particles via the pathways of clathrin-and caveolae-mediated endocytosis. *Biochem. J.* 377 (1), 159–169. doi:10.1042/BJ20031253

Rudel, T., Kepp, O., and Kozjak-Pavlovic, V. (2010). Interactions between bacterial pathogens and mitochondrial cell death pathways. *Nat. Rev. Microbiol.* 8 (10), 693–705. doi:10.1038/nrmicro2421

Sahay, G., Alakhova, D. Y., and Kabanov, A. V. (2010). Endocytosis of nanomedicines. J. Control. release 145 (3), 182–195. doi:10.1016/j.jconrel.2010. 01.036

Sanchez, L., Yi, Y., and Yu, Y. (2017). Effect of partial PEGylation on particle uptake by macrophages. *Nanoscale* 9 (1), 288–297. doi:10.1039/C6NR07353K

Sandvig, K., Pust, S., Skotland, T., and van Deurs, B. (2011). Clathrin-independent endocytosis: Mechanisms and function. *Curr. Opin. Cell Biol.* 23 (4), 413–420. doi:10. 1016/j.ceb.2011.03.007

Schmidtchen, A., Frick, I. M., Andersson, E., Tapper, H., and Björck, L. (2002). Proteinases of common pathogenic bacteria degrade and inactivate the antibacterial peptide LL-37. *Mol. Microbiol.* 46 (1), 157–168. doi:10.1046/j. 1365-2958.2002.03146.x

Shao, D., Lu, M.-m., Zhao, Y.-w., Zhang, F., Tan, Y.-f., Zheng, X., et al. (2017). The shape effect of magnetic mesoporous silica nanoparticles on endocytosis, biocompatibility and biodistribution. *Acta biomater.* 49, 531–540. doi:10.1016/j. actbio.2016.11.007

Shen, X., Ma, R., Huang, Y., Chen, L., Xu, Z., Li, D., et al. (2020). Nano-decocted ferrous polysulfide coordinates ferroptosis-like death in bacteria for anti-infection therapy. *Nano Today* 35, 100981. doi:10.1016/j.nantod.2020.100981

Shepherd, V. L. (1986). The role of the respiratory burst of phagocytes in host defense. Semin. Respir. Infect. 1 (2), 99–106.

Shi, S., Wu, S., Shen, Y., Zhang, S., Xiao, Y., He, X., et al. (2018). Iron oxide nanozyme suppresses intracellular Salmonella Enteritidis growth and alleviates infection *in vivo*. *Theranostics* 8 (22), 6149–6162. doi:10.7150/thno.29303

Su, F.-Y., Chen, J., Son, H.-N., Kelly, A. M., Convertine, A. J., West, T. E., et al. (2018). Polymer-augmented liposomes enhancing antibiotic delivery against intracellular infections. *Biomaterials Sci.* 6 (7), 1976–1985. doi:10.1039/c8bm00282g

Sunnapu, O., Khader, R., Dhanka, M., Kumar Vemula, P., and Karuppannan, S. (2022). Enzyme-responsive hydrogel for delivery of the anti-inflammatory agent Zingerone. *ChemNanoMat* 8 (11), e202200334. doi:10.1002/cnma.202200334

Tahara, K., Sakai, T., Yamamoto, H., Takeuchi, H., Hirashima, N., and Kawashima, Y. (2009). Improved cellular uptake of chitosan-modified PLGA nanospheres by A549 cells. *Int. J. Pharm.* 382 (1-2), 198–204. doi:10.1016/j.ijpharm.2009.07.023

ten Brink, H. B., Dekker, H. L., Schoemaker, H. E., and Wever, R. (2000). Oxidation reactions catalyzed by vanadium chloroperoxidase from Curvularia inaequalis. *J. Inorg. Biochem.* 80 (1-2), 91–98. doi:10.1016/s0162-0134(00)00044-1

Thomas, E. L. (1979). Myeloperoxidase-hydrogen peroxide-chloride antimicrobial system: Effect of exogenous amines on antibacterial action against *Escherichia coli*. *Infect. Immun.* 25 (1), 110–116. doi:10.1128/iai.25.1.110-116.1979

Vandal, O. H., Pierini, L. M., Schnappinger, D., Nathan, C. F., and Ehrt, S. (2008). A membrane protein preserves intrabacterial pH in intraphagosomal *Mycobacterium tuberculosis. Nat. Med.* 14 (8), 849–854. doi:10.1038/nm.1795

Vogel, J. P., Andrews, H. L., Wong, S. K., and Isberg, R. R. (1998). Conjugative transfer by the virulence system of *Legionella pneumophila*. *Science* 279 (5352), 873–876. doi:10. 1126/science.279.5352.873

Wang, H., Wu, L., and Reinhard, B. r. M. (2012). Scavenger receptor mediated endocytosis of silver nanoparticles into J774A. 1 macrophages is heterogeneous. *Acs Nano* 6 (8), 7122–7132. doi:10.1021/nn302186n

Wang, Z., Liu, X., Peng, Y., Su, M., Zhu, S., Pan, J., et al. (2020). Platensimycinencapsulated liposomes or micelles as biosafe nanoantibiotics exhibited strong antibacterial activities against methicillin-resistant *Staphylococcus aureus* infection in mice. *Mol. Pharm.* 17 (7), 2451–2462. doi:10.1021/acs.molpharmaceut.0c00194

Webb, J., Harvey, M., Holden, D. W., and Evans, T. (2001). Macrophage nitric oxide synthase associates with cortical actin but is not recruited to phagosomes. *Infect. Immun.* 69 (10), 6391–6400. doi:10.1128/IAI.69.10.6391-6400.2001

Wright, G. D. (2005). Bacterial resistance to antibiotics: Enzymatic degradation and modification. Adv. Drug Deliv. Rev. 57 (10), 1451-1470. doi:10.1016/j.addr.2005.04.002

Wu, J., Wang, X., Wang, Q., Lou, Z., Li, S., Zhu, Y., et al. (2019). Nanomaterials with enzyme-like characteristics (nanozymes): Next-generation artificial enzymes (II). *Chem. Soc. Rev.* 48 (4), 1004–1076. doi:10.1039/C8CS00457A

Xiong, M. H., Bao, Y., Yang, X.-Z., Wang, Y.-C., Sun, B., and Wang, J. (2012a). Lipasesensitive polymeric triple-layered nanogel for "on-demand" drug delivery. *J. Am. Chem. Soc.* 134 (9), 4355–4362. doi:10.1021/ja211279u

Xiong, M. H., Li, Y. J., Bao, Y., Yang, X. Z., Hu, B., and Wang, J. (2012b). Bacteriaresponsive multifunctional nanogel for targeted antibiotic delivery. *Adv. Mater.* 24 (46), 6175–6180. doi:10.1002/adma.201202847

Yang, Q., Jones, S. W., Parker, C. L., Zamboni, W. C., Bear, J. E., and Lai, S. K. (2014). Evading immune cell uptake and clearance requires PEG grafting at densities substantially exceeding the minimum for brush conformation. *Mol. Pharm.* 11 (4), 1250–1258. doi:10.1021/mp400703d

Yang, S., Han, X., Yang, Y., Qiao, H., Yu, Z., Liu, Y., et al. (2018). Bacteria-targeting nanoparticles with microenvironment-responsive antibiotic release to eliminate intracellular *Staphylococcus aureus* and associated infection. *ACS Appl. Mater. interfaces* 10 (17), 14299–14311. doi:10.1021/acsami.7b15678

Zhang, Y., He, Y., Shi, C., Sun, M., Yang, C., Li, H., et al. (2020). Tannic acid-assisted synthesis of biodegradable and antibacterial mesoporous organosilica nanoparticles decorated with nanosilver. *ACS Sustain. Chem.* 8 (3), 1695–1702. doi:10.1021/acssuschemeng.9b07576

Zhang, Y., Zhang, J., Chen, W., Angsantikul, P., Spiekermann, K. A., Fang, R. H., et al. (2017). Erythrocyte membrane-coated nanogel for combinatorial antivirulence and responsive antimicrobial delivery against *Staphylococcus aureus* infection. *J. Control. Release* 263, 185–191. doi:10.1016/j.jconrel.2017.01.016