



OPEN ACCESS

EDITED BY

Theodoros Soldatos,
Other, Germany

REVIEWED BY

Zaida Ann Luthey-Schulten,
University of Illinois at Urbana-Champaign,
United States
Ludovic Emmanuel Autin,
The Scripps Research Institute, United States
Congzhou Mike Sha,
The Pennsylvania State University,
United States

*CORRESPONDENCE

Maxime Maria,
✉ maxime.maria@unilim.fr
Matthieu Montes,
✉ matthieu.montes@cnam.fr

RECEIVED 06 March 2025

ACCEPTED 19 May 2025

PUBLISHED 06 June 2025

CITATION

Maria M, Guillaume V, Guionnière S,
Dacquay N, Plateau–Holleville C, Larroque V,
Lardé J, Naimi Y, Piquemal J-P, Levieux G,
Lagarde N, Mérillou S and Montes M (2025)
Interactive visualization of large molecular
systems with VTX: example with a minimal
whole-cell model.
Front. Bioinform. 5:1588661.
doi: 10.3389/fbinf.2025.1588661

COPYRIGHT

© 2025 Maria, Guillaume, Guionnière,
Dacquay, Plateau–Holleville, Larroque, Lardé,
Naimi, Piquemal, Levieux, Lagarde, Mérillou
and Montes. This is an open-access article
distributed under the terms of the [Creative
Commons Attribution License \(CC BY\)](#). The
use, distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Interactive visualization of large molecular systems with VTX: example with a minimal whole-cell model

Maxime Maria^{1*}, Valentin Guillaume², Simon Guionnière²,
Nicolas Dacquay², Cyprien Plateau–Holleville¹,
Vincent Larroque^{1,3}, Jean Lardé^{2,3}, Yassine Naimi³,
Jean-Philip Piquemal^{4,5,6}, Guillaume Levieux⁷,
Nathalie Lagarde², Stéphane Mérillou¹ and Matthieu Montes^{2,6*}

¹XLIM, UMR CNRS 7252, Université de Limoges, Limoges, France, ²Laboratoire GBCM, EA 7528, Conservatoire National des Arts et Métiers, Paris, France, ³Qubit Pharmaceuticals SAS, Paris, France, ⁴LCT, UMR 7616 CNRS, Sorbonne Université, Paris, France, ⁵Department of Biomedical Engineering, University of Texas at Austin, Austin, TX, United States, ⁶Institut Universitaire de France, Paris, France, ⁷Laboratoire CEDRIC, EA 4626, Conservatoire National des Arts et Métiers, Paris, France

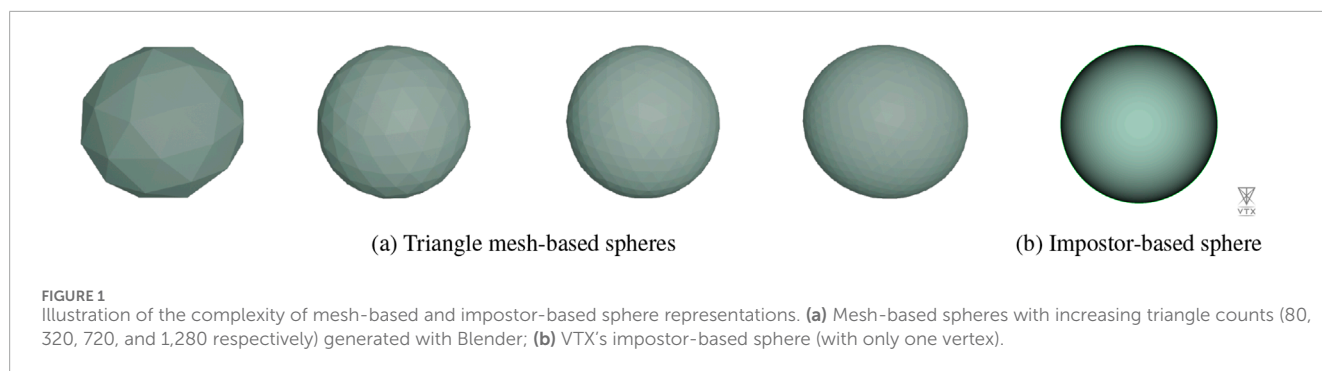
VTX is an open-source molecular visualization software designed to overcome the scaling limitations of existing real-time molecular visualization software when handling massive molecular datasets. VTX employs a meshless molecular graphics engine utilizing impostor-based techniques and adaptive level-of-detail (LOD) rendering. This approach significantly reduces memory usage and enables real-time visualization and manipulation of large molecular systems. Performance benchmarks against VMD, PyMOL, and ChimeraX using a 114-million-bead Martini minimal whole-cell model demonstrate VTX's efficiency, maintaining consistent frame rates even under interactive manipulation on standard computer hardware. VTX incorporates features such as Screen-Space Ambient Occlusion (SSAO) for enhanced depth perception and free-fly navigation for intuitive exploration of large molecular systems. VTX is open-source and free for non-commercial use. Binaries for Windows and Ubuntu Linux are available at <http://vtx.drugdesign.fr>. VTX source code is available at <https://github.com/VTX-Molecular-Visualization>.

KEYWORDS

molecular visualization, molecular graphics, usability, molecular modeling, molecular dynamics, high-performance computing, protein structure, structural biology

Introduction

Recent advances in the determination of atomic resolution molecular structures and assemblies using CryoEM (Jumper et al., 2021), in protein structure prediction methods like AlphaFold (Yip et al., 2020; Varadi et al., 2021), and the growing availability of molecular dynamics simulations (Amaro Åqvist et al., 2025; Hospital and Orozco, 2024), have significantly increased the volume of molecular structural biology data. Additionally, modern high-performance computing hardware has enabled the simulation of larger molecular systems (Jung et al., 2019; Stevens et al., 2023; Coshic et al., 2024; Gilbert et al., 2023), further amplifying data



production. This massive molecular data poses a challenge for existing software solutions in terms of data processing, visualization, and storage.

In this work, we demonstrate the performance of VTX (Maria et al., 2025) on large molecular datasets by visualizing a 114 million Martini-beads minimal whole cell model published in 2023 (Stevens et al., 2023), which pre-figures future massive molecular systems for molecular dynamics simulation. We describe the solutions implemented in VTX to efficiently handle large all-atom molecular data while ensuring high-quality and informative real-time rendering. This serves as a valuable benchmark for future applications of massive molecular dynamics simulation visualization, where memory management and high-quality rendering remain key challenges.

Methods

VTX (Maria et al., 2025) is an open-source molecular visualization software designed for efficient real-time handling of large molecular simulations datasets. It achieves this through a notably high-performance, meshless molecular graphics engine coupled with a minimalistic, task-oriented GUI. Here, we illustrate VTX's features and performance on a massive molecular system: the 2023 Martini minimal whole cell model (Stevens et al., 2023).

Molecular graphics engine

Meshless representations

All molecular representation can be described using a triangular mesh. To draw a triangular mesh, each triangle requires the XYZ coordinates of its three points, resulting in at least 36 bytes of memory per triangle.

In order to produce images of acceptable quality, many triangles are needed to accurately define the geometry and have smooth surfaces (see Figure 1a). Even if GPUs are designed for fast triangle rendering, the number of required triangles is prohibitive for real-time visualization.

To address this issue, *Ball and Stick* and *Van der Waals* representations are based on quadrics such as spheres or cylinders that are described implicitly, using atom coordinates and radii exclusively. To achieve this, the rendering engine employs impostor-based techniques (Bagur et al., 2012). For each primitive, a simple

quad is rasterized. Then, ray-casting is used to evaluate the implicit equation of the primitive, generating the final shape. This approach enables fast and pixel-perfect rendering (see Figure 1b) while minimizing both bandwidth and memory usage, which is crucial for handling large molecular structures or dynamics data.

Cartoon representation

Cartoon representations in VTX use an adaptive level-of-detail (LOD) approach with tessellation shaders (Hermosilla et al., 2015). Triangles are dynamically computed for each frame based on the atom coordinates, enabling fast rendering of secondary structure cartoons without requiring any preprocess. This allows updates at each trajectory step with minimal computational cost.

Solvent excluded surface (SES)

As is common in molecular visualization software, the surface is simply computed using the marching cubes algorithm (Lorensen and Cline, 1987). This approach requires balancing surface smoothness and the number of generated triangles to ensure both visual quality and computational efficiency.

No instancing

Instancing is a widely used technique for rendering multiple copies of the same object in a scene simultaneously, significantly improving performance when dealing with many identical objects. By reducing the number of draw calls, it saves both CPU and GPU resource usage. Massive static molecular scenes, containing up to billion atoms, as displayed with YASARA (Krieger and Vriend, 2014) or Mol* (Sehnel et al., 2021; Rose et al., 2024) effectively leverage instancing. However, instancing assumes that all copies share the same conformation, making it unsuitable for molecular dynamics trajectories where structures evolve independently. In this work, which pre-figures future applications for visualizing molecular dynamics trajectories of massive molecular systems, no instancing is used to display the 2023 Martini minimal whole cell model (Stevens et al., 2023).

Lighting

Depth perception is essential for localizing and distinguishing molecular entities in a complex scene. For this purpose, VTX uses a Screen-Space Ambient Occlusion (SSAO) (Mitrting, 2007) to enhance the perception of structural details and emphasize the burriedness of atoms, improving the understanding of molecular architectures (see Figure 2).

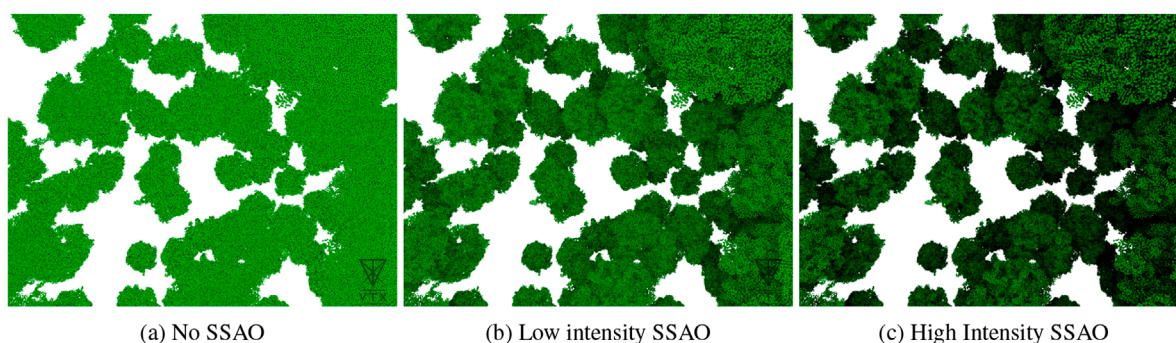


FIGURE 2

Impact of the intensity of the Screen-Space Ambient Occlusion (SSAO) on a close-up rendering of a large molecular system (Stevens et al., 2023): (a) No SSAO; (b) Low intensity SSAO; (c) High intensity SSAO.

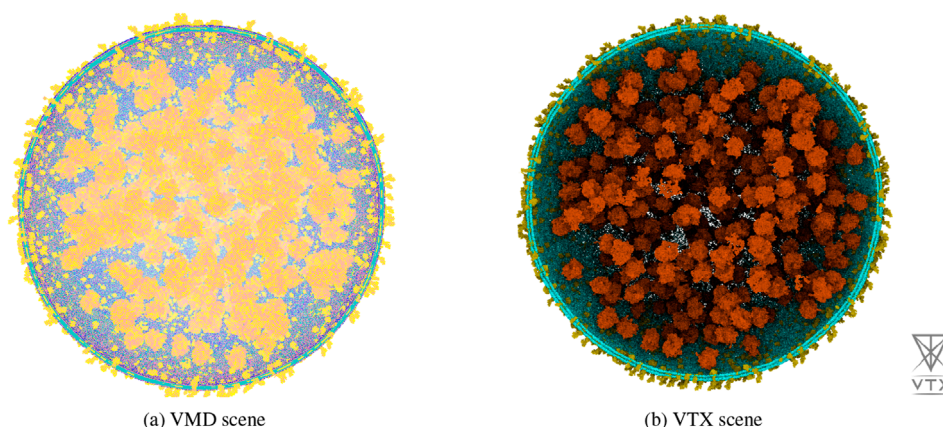


FIGURE 3

Wide-system view of the lipids, membrane proteins, and ribosomes (36,566,468 Martini beads) of the 2023 Martini minimal whole cell model rendered in real-time with VMD (a) on the left, and VTX (b) on the right. These illustrations were captured on a Dell precision 5480 laptop equipped with an Intel i7-13800H, 32 GB of RAM and a NVidia Quadro RTX 3000 GPU. All attempts to improve the rendering with VMD on this setup resulted in a freeze of the application.

Free-fly navigation and interaction

Navigating through large systems composed by multiple proteins, lipids and other heavy molecules poses a new challenge compared to single-protein scale visualization. Traditional trackball navigation, which rotates the camera around a fixed point, can be restrictive for large systems, making it difficult to explore surrounding regions.

To address this issue, VTX implements a free-fly camera mode in addition to the default trackball. Free-fly mode allows to move freely within the 3D space using keyboard inputs for movement and the mouse controls for directions, similar to the first-person navigation in video games. This approach has successfully applied in interactive docking tool UDock (Levieux et al., 2014; Plateau-Holleville et al., 2023), and in the multi-scale molecular visualization software CellVIEW (Le Muzic et al., 2015).

Performance evaluation on a massive molecular system: the 2023 martini minimal whole cell model

We compared the performance of VTX version 0.4.4 (Maria et al., 2025), VMD version 1.9.3 (Humphrey et al., 1996), PyMOL version 3.1 (Schrödinger) and ChimeraX version 1.9 (Pettersen et al., 2021) in terms of system file loading time and frame rate using the 2023 Martini minimal whole cell model (Stevens et al., 2023). This system is a coarse-grained model of 114 million Martini beads available in the Gromacs (Abraham et al., 2024) *gro* file format that pre-figures future massive molecular systems constructed for molecular dynamics simulation. The system, displayed in figure ?? Contains 60,887 soluble proteins, 2,200 membrane proteins, 503 ribosomes, a single 500 kbp circular dsDNA, 1.3 million lipids, 1.7 million metabolites and 14 million ions. For clarity of the visual analysis, we did not display the 447 million water beads.

TABLE 1 Loading time and frame rate (benchmark results on the 2023 Martini minimal whole cell model (114 million Martini beads) (Stevens et al., 2023).

Software	Loading time (s)	Frame rate - close-up (s ⁻¹)	Frame rate - far (s ⁻¹)
ChimeraX	—	—	—
PyMOL3.1	—	—	—
VMD	200.33 ± 16.05	1.36	1.38
VTX	205.00 ± 13.06	11.41	12.82

No values could be obtained with ChimeraX and PyMOL due to a frozen application (ChimeraX) or a crash (PyMOL) during system loading. System file loading time performance has been evaluated in triplicate. Frame rate measurement has been evaluated using the NVidia Frameview software version 1.6. The average frame rate was calculated over a 20-s period for both close-up (as in Figure 2) and far (as in Figure 3) camera views. All performance evaluations were conducted on a Dell precision 5480 laptop equipped with an Intel i7-13800H, 32 GB of RAM and a NVidia Quadro RTX 3000 GPU.

The benchmark results are presented in Table 1. ChimeraX and PyMOL were unable to load the system due respectively to a crash and a freeze. VMD and VTX loaded the system with similar times and successfully displayed it, but VTX achieved significantly higher frame rate. Any attempts to modify VMD rendering settings caused in a severe drop in frame rate, eventually leading to a complete freeze. With a stable interactive frame rate and despite limited computational resource, VTX allowed to interactively manipulate the system, perform precise selections and modify rendering settings.

A video displaying a free-fly real-time navigation inside this system is available as a [Supplementary Material](#).

Perspectives

VMD, ChimeraX and PyMOL are reference molecular visualization tools that are widely used by the community and that, compared to VTX, offer many additional functionalities notably through the use of numerous and diverse plugins developed over time. In the next version of VTX, Python bindings will be supported to allow the development and integration of external scripts and plugins.

The construction and rendering of a Solvent Excluded Surface (SES) for large molecular systems currently exceed the capabilities of existing molecular visualization software due to high memory demands. To address this limitation, we will integrate our analytical SES construction method (Plateau—Holleville et al., 2024) into VTX. This approach will also enable the display of a smooth, pixel-perfect molecular surface.

Visualizing large system trajectories presents a challenge due to the memory demands of loading all atomic coordinates for each frame. Currently, VTX loads all frames to ensure smooth playback, which can cause performance issues on memory-constrained systems. To address this, we will implement a streaming system that limits the number of frames loaded into memory simultaneously, significantly reducing the memory footprint for large systems.

Even if VTX can already render large molecular systems without relying on instancing, further performance gains could be achieved by enabling instancing for identical entities. This would leverage VTX's graphics engine to display systems containing

billions of atoms. For certain applications, such as visualizing repetitive molecular assemblies or large-scale structural models, instancing could significantly enhance rendering efficiency and reduce memory usage.

Finally, we are developing a ray tracing engine specifically designed for molecular scenes, integrating state-of-the-art computer graphics techniques to enable the fast generation of high-quality images and interactive previews that accurately reflect the final rendering.

Conclusion

VTX is an open-source, high performance molecular visualization software designed for real-time exploration of massive molecular systems. Its graphics engine, leveraging impostor-based rendering and adaptive level-of-detail, achieves interactive frame rates with a 114-million-atom model, outperforming existing tools on standard hardware. VTX integrates features such as Screen-Space Ambient Occlusion for enhanced depth perception and free-fly navigation for intuitive exploration of large systems. Future developments will include analytical Solvent-Excluded Surface construction, a streaming system for handling large molecular dynamics trajectories, instancing for reducing the memory load with multiple rigid copies of the same molecular object, and high-quality path-traced rendering. As an open-source software, VTX nature fosters community-driven advancements in large-scale molecular structure and dynamics visualization.

Data availability statement

All .gro files that constitute the Martini minimal whole cell model are available upon request at the Marrink lab. The procedure to generate the model is available at https://github.com/marrink-lab/Martini_Minimal_Cell.

Author contributions

MM: Conceptualization, Funding acquisition, Investigation, Methodology, Project administration, Resources, Software,

Supervision, Validation, Writing – original draft, Writing – review and editing. VG: Investigation, Software, Writing – review and editing, Visualization, Writing – original draft. SG: Conceptualization, Formal Analysis, Investigation, Methodology, Validation, Writing – review and editing, Software. ND: Data curation, Investigation, Software, Validation, Writing – review and editing. CP-H: Investigation, Methodology, Software, Visualization, Writing – review and editing. VL: Methodology, Software, Visualization, Writing – review and editing. JL: Methodology, Software, Visualization, Writing – review and editing. YN: Methodology, Software, Visualization, Writing – review and editing. J-PP: Methodology, Project administration, Resources, Visualization, Writing – review and editing. GL: Conceptualization, Methodology, Software, Writing – review and editing. NL: Methodology, Validation, Visualization, Writing – review and editing. SM: Methodology, Resources, Supervision, Writing – review and editing. MM: Conceptualization, Formal Analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Software, Supervision, Validation, Visualization, Writing – original draft, Writing – review and editing.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. VTX has received funding from the European Research Council (ERC) under the European Union's Horizon 2020 research and innovation program (grant agreement n°640283) and from the ERC under the Horizon Europe research and innovation program (grant agreement n°101069190). CP-H and VL are supported by institutional grants from the National Research Agency under the Investments for the future program with the reference ANR-18-EURE-0017 TACTIC. VL is recipient of a fellowship from Qubit Pharmaceuticals.

References

- Abraham, M., Alekseenko, A., Basov, V., Bergh, C., Briand, E., Brown, A., et al. (2024). GROMACS 2024.0 Manual.
- Amaro, R. E., Åqvist, J., Bahar, I., Battistini, F., Bellaiche, A., Beltran, D., et al. (2025). The need to implement FAIR principles in biomolecular simulations. *Nat. Methods* 22, 641–645. doi:10.1038/s41592-025-02635-0
- Bagur, P. D., Shivashankar, N., and Natarajan, V. (2012). "Improved quadric surface impostors for large bio-molecular visualization," in *Proceedings of the eighth Indian conference on computer vision, graphics and image processing, ICGIP '12* (New York, NY, USA: Association for Computing Machinery).
- Coshic, K., Maffeo, C., Winogradoff, D., and Aksimentiev, A. (2024). The structure and physical properties of a packaged bacteriophage particle. *Nature* 627, 905–914. doi:10.1038/s41586-024-07150-4
- Gilbert, B. R., Thornburg, Z. R., Brier, T. A., Stevens, J. A., Grünewald, F., Stone, J. E., et al. (2023). Dynamics of chromosome organization in a minimal bacterial cell. *Front. Cell Dev. Biol.* 11 – 2023. doi:10.3389/fcell.2023.1214962
- Hermosilla, P., Guallar, V., Vinacua, A., and Vázquez, P. -P. (2015). *Instant visualization of secondary structures of molecular models*. VCBM '15. Chester, UK: Eurographics Association.
- Hospital, A., and Orozco, M. (2024). MD-DATA: the legacy of the ABC Consortium. *Biophys. Rev.* 16 (3), 269–271. doi:10.1007/s12551-024-01197-3
- Humphrey, W., Dalke, A., and Schulten, K. (1996). VMD: visual molecular dynamics. *J. Mol. Graph.* 14 (1), 33–38. doi:10.1016/0263-7855(96)00018-5
- Jumper, J., Evans, R., Pritzel, A., Green, T., Figurnov, M., Ronneberger, O., et al. (2021). Highly accurate protein structure prediction with AlphaFold. *Nature* 596, 583–589. doi:10.1038/s41586-021-03819-2
- Jung, J., Nishima, W., Daniels, M., Bascom, G., Kobayashi, C., Adedoyin, A., et al. (2019). Scaling molecular dynamics beyond 100,000 processor cores for large-scale biophysical simulations. *J. Comput. Chem.* 40 (21), 1919–1930. doi:10.1002/jcc.25840
- Krieger, E., and Vriend, G. (2014). YASARA view - molecular graphics for all devices - from smartphones to workstations. *Bioinformatics* 30, 2981–2982. doi:10.1093/bioinformatics/btu426
- Le Muzic, M., Autin, L., Parulek, J., and Viola, I. (2015). cellVIEW: a tool for illustrative and multi-scale rendering of large biomolecular datasets. *Eurographics Workshop Vis. Comput. Biomed.* 61–70. doi:10.2312/vcbm.20151209
- Levieux, G., Tiger, G., Mader, S., Zagury, J. F., Natkin, S., and Montes, M. (2014). Udock, the interactive docking entertainment system. *Faraday Discuss.* 169, 425–441. doi:10.1039/c3fd00147d
- Lorensen, W. E., and Cline, H. E. (1987). Marching cubes: a high resolution 3d surface construction algorithm. *SIGGRAPH Comput. Graph.* 21 (4), 163–169. doi:10.1145/37401.37422
- Maria, M., Guionnière, S., Dacquay, N., Plateau-Holleville, C., Guillaume, V., Larroque, V., et al. (2025). VTX: real-time high-performance molecular structure and dynamics visualization software. *Bioinformatics*, btaf295. doi:10.1093/bioinformatics/btaf295

Acknowledgments

We thank Prof. SJ Marrink and J Stevens for kindly providing the dataset.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fbinf.2025.1588661/full#supplementary-material>

- Mittring, M. (2007). "Finding next gen: CryEngine 2," in *ACM SIGGRAPH 2007 courses*, 97–121. SIGGRAPH '07.
- Pettersen, E. F., Goddard, T. D., Huang, C. C., Meng, E. C., Couch, G. S., Croll, T. I., et al. (2021). UCSF ChimeraX: structure visualization for researchers, educators, and developers. *Protein Sci.* 30 (1), 70–82. doi:10.1002/pro.3943
- Plateau-Holleville, C., Guionnière, S., Boyer, B., Jiménez-García, B., Levieux, G., Mérillou, S., et al. (2023). Udock2: interactive real-time multi-body protein–protein docking software. *Bioinformatics* 39 (10), btad609–10. doi:10.1093/bioinformatics/btad609
- Plateau—Holleville, C., Maria, M., Mérillou, S., and Montes, M. (2024). Efficient GPU computation of large protein Solvent-Excluded Surface. *IEEE Trans. Vis. Comput. Graph.*, 1–12. doi:10.1109/TVCG.2024.3380100
- Rose, A., Sehnal, D., Goodsell, D. S., and Autin, L. (2024). Mesoscale explorer: visual exploration of large-scale molecular models. *Protein Sci.* 33, e5177. doi:10.1002/pro.5177
- Schrödinger, L. L. C. The PyMOL molecular graphics system. Available online at: <https://www.pymol.org/>.
- Sehnal, D., Bittrich, S., Deshpande, M., Svobodová, R., Berka, K., Bazgier, V., et al. (2021). Mol*Viewer: modern web app for 3D visualization and analysis of large biomolecular structures. *Nucleic Acids Res.* 49 (W1), W431–W437. doi:10.1093/nar/gkab314
- Stevens, J. A., Grünewald, F., van Tilburg, P. A. M., König, M., Gilbert, B. R., Brier, T. A., et al. (2023). Molecular dynamics simulation of an entire cell. *Front. Chem.* 11, 1106495. doi:10.3389/fchem.2023.1106495
- Varadi, M., Anyango, S., Deshpande, M., Nair, S., Natassia, C., Yordanova, G., et al. (2021). AlphaFold Protein Structure Database: massively expanding the structural coverage of protein-sequence space with high-accuracy models. *Nucleic Acids Res.* 50 (D1), D439–D444. doi:10.1093/nar/gkab1061
- Yip, K. M., Fischer, N., Paknia, E., Chari, A., and Stark, H. (2020). Atomic-resolution protein structure determination by cryo-EM. *Nature* 587, 157–161. doi:10.1038/s41586-020-2833-4