



The Emerging Role of Neutrophil Extracellular Traps in Arterial, Venous and Cancer-Associated Thrombosis

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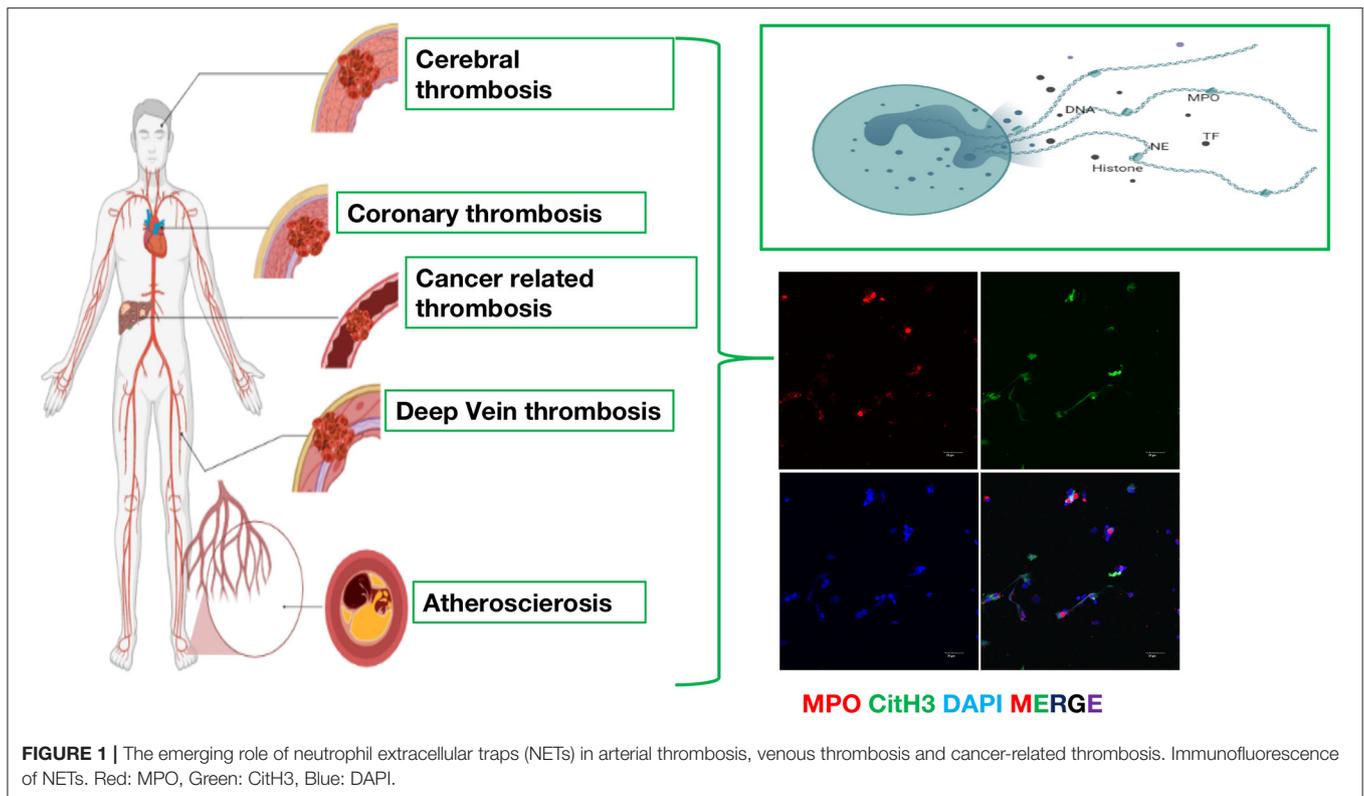
Neutrophils play a vital role in the formation of arterial, venous and cancer-related thrombosis. Recent studies have shown that in a process known as NETosis, neutrophils release proteins and enzymes complexed to DNA fibers, collectively called neutrophil extracellular traps (NETs). Although NETs were originally described as a way for the host to capture and kill bacteria, current knowledge indicates that NETs also play an important role in thrombosis. According to recent studies, the destruction of vascular microenvironmental homeostasis and excessive NET formation lead to pathological thrombosis. *In vitro* experiments have found that NETs provide skeletal support for platelets, red blood cells and procoagulant molecules to promote thrombosis. The protein components contained in NETs activate the endogenous coagulation pathway to promote thrombosis. Therefore, NETs play an important role in the formation of arterial thrombosis, venous thrombosis and cancer-related thrombosis. This review will systematically summarize and explain the study of NETs in thrombosis in animal models and *in vivo* experiments to provide new targets for thrombosis prevention and treatment.

Keywords: neutrophil, neutrophil extracellular traps, thrombosis, deep vein thrombosis (DVT), cancer thrombosis

INTRODUCTION

The role of neutrophil extracellular traps (NETs) in inflammation and thrombosis has been controversial for decades (1, 2). Traditionally, thrombosis is considered only a blood vessel or blood disease (2). Inflammation and thrombosis are two independent pathological processes (3). However, with the progress of immunology research, researchers have discovered that thrombosis is an inflammatory process (4). A previous study observed neutrophil exudation in the blood vessel wall at the early stage of thrombosis induction, followed by monocytes and lymphocytes (5). Based on accumulating evidence, neutrophils play a key role in the process of thrombosis (5–8). The depletion of neutrophils has been shown to reverse experimental thrombosis (9, 10). Another study reported that neutrophils produce tissue factor (TF) and contribute to the formation of thrombosis *in vivo* and *in vitro* (11).

In 2004, Brinkmann et al. found that neutrophils form a structure that separates from the cell itself after stimulation called neutrophil extracellular traps (NETs), which provided new ideas for studies of the interaction between neutrophils in the pathway of inflammation and thrombosis (12). NETs were discovered as extracellular strands of decondensed DNA in complex with histones and granule proteins, which were expelled from dying neutrophils. NETs are composed of circulating markers (myeloperoxidase, neutrophil elastase, etc.) and kill bacteria during general inflammation.



Additionally, NETs play vital roles in arterial, venous and cancer-related thrombosis formation. For arterial and venous thrombosis, activated components are composed of DNA and histones, which provide a structure for red cells and platelets and contribute to the formation of thrombosis. For cancer-related thrombosis, tumor cells can stimulate the formation of NETs and then promote the metastasis of cancer. Therefore, understanding the different mechanisms of thrombosis has become an important direction for the study of thrombotic diseases (Figure 1). The relevant research progress in recent years is summarized below.

THE ROLE OF NETs IN ATHEROSCLEROSIS AND ARTERIAL THROMBOSIS

Chronic inflammation plays an important role in the occurrence and development of atherosclerosis and thrombosis, during which the status of neutrophils has received increasing attention (13). Many studies have confirmed an important role for NETs in the processes of atherosclerosis, coronary artery disease (CAD) and ischemic stroke (14–16). However, most of these studies are based on *in vitro* and animal experiments (Table 1). Few studies have assessed the clinical value of NETs in arterial thrombosis.

NETs in Atherosclerosis

NETs have been detected in patients with atherosclerosis and in animal models, and NETs are related to various pathogenesises of

atherosclerosis. NETs play a role in all stages of atherosclerosis, from early endothelial dysfunction to atherosclerotic plaque rupture and atherosclerotic thrombosis (25, 26). As a scaffold for cells and various coagulation factors, NETs not only exist in plaques and thrombi but also induce oxidative stress, induce the activation of endothelial cells, antigen-presenting cells and platelets, increase the expression of coagulation factors, and lead to proinflammatory reactions (Figure 2A). These structures play a role in the pathogenesis of atherosclerotic plaque formation and thrombosis (3, 4, 27). Compared to the network structures produced by other cells, NETs are mainly produced in the early stage of thrombosis, and most of them are formed in the acute stage of the disease (17, 28). NETs also induce the death of smooth muscle cells, leading to reduced plaque stability (22). At the same time, NETs also promote the abnormal activation of macrophages and upregulate the levels of IL-8 and inflammasomes, thereby further amplifying the role of NETs and accelerating the progression of atherosclerosis (23, 29–31). Macrophages are more common in the types of lesions that are prone to rupture, but studies have shown that the infiltration of neutrophils is more important for these erosion-prone lesions (32). Oxidized low-density lipoprotein (oxLDL), which easily accumulates in macrophages, is used as an atherosclerosis-inducing molecule. Many studies have shown that it stimulates neutrophils to form NETs (33). The specific deletion of peptidyl arginine deiminase-4 (PAD4) reduces the formation of NETs and significantly alleviates atherosclerosis, complications, and inflammation caused by macrophages (34, 35).

TABLE 1 | NETs in atherosclerosis and arterial thrombosis.

Objects	Main findings	References
Patients with STEMI	NETs were dominated in early thrombosis	(17)
Patients with AMI or AIS	The formation and abundance of NETs were associated with prognosis	(18)
Patients with STEMI	Plasma levels of NETs markers increased and were positively correlated with infarct size and left ventricular dysfunction	(19)
Patients with COVID-19	NETs were present in coronary thrombus of patients with COVID-19 and myocardial infarction	(20)
Patients with diabetes	Elevated levels of NETs markers (e.g., citH3) were associated with severe coronary atherosclerosis in patients with diabetes	(21)
Patients with CAD or AIS	citH3 was observed in almost all thrombi	(2)
Mouse/atherosclerosis	NETs lysed smooth muscle cells, leading to the destabilization of plaques.	(22)
Mouse/diabetes	NETs promoted macrophage inflammation and inhibited atherosclerosis resolution	(23)
Integrin $1\alpha^{-/-}$ mouse	Decreased levels of NETs resulted in decreased platelet aggregation, cathepsin-G secretion, and arterial thrombosis	(24)

NETs, neutrophil extracellular traps; STEMI, ST-elevation myocardial infarction; AMI, acute myocardial infarction; AIS, acute ischemic stroke; CAD, coronary artery disease.

NETs in Coronary Thrombosis

NETs promote coronary microvascular thrombosis and affect heart function (36). The content of NETs in cardiogenic thrombosis is higher than that in thrombosis with other causes (37). NETs carrying tissue factor are often observed at the site of coronary thrombosis, and studies have shown that NETs are a potential marker of arterial thrombosis in clinical specimens and animal models (38). The structure of NETs has been detected in thrombi of ST-segment elevation myocardial infarction (STEMI) and non-ST-segment elevation myocardial infarction (NSTEMI) (39), and the myocardial infarction score (18) and circulating markers (citrullinated histone 3; myeloperoxidase, neutrophil elastase, etc.) of NETs are significantly reduced after treatment (40). The activation of platelets and neutrophils increases the risk of major adverse cardiovascular events (MACEs) after acute myocardial infarction (41). Moreover, NETs stimulate fibroblasts, affect heart remodeling after STEMI and are important mediators of fiber remodeling (19). Recent studies have also shown a major role for NETs in the onset of STEMI in patients with COVID-19, and NETs have been detected in thrombus samples from all patients with COVID-19 (20).

NETs in Cerebral Thrombosis

NETs are also an important component of cerebral thrombosis, are cytotoxic to endothelial cells, and together with von Willebrand factor (VWF), promote the hypercoagulable state (42, 43). Abundant NET structures have been observed in almost all thrombi removed from patients with ischemic stroke (37, 44), and patients with cardiogenic embolism have higher levels of NETs (45). In cerebral ischemia, neutrophils are

the first type of cells that migrate to damaged brain tissue. They produce NETs in the brain parenchyma and cerebral blood vessels, thereby aggravating inflammation and brain tissue damage (46). Accumulating evidence indicates that NETs may fight against tissue-type plasminogen activator (t-PA) by promoting coagulation and stabilizing clot-induced thrombosis, which is a problem often encountered in the treatment of patients with stroke (47–49). NETs may be potential biomarkers and therapeutic targets for recurrent stroke in patients with severe carotid artery stenosis (50). Deoxyribonuclease (DNase) degradation of NETs or treatment with PAD4 inhibitors to prevent NET formation significantly inhibits arterial thrombosis in the brains of ischemic mice and improves stroke prognosis, such as by reducing the site of infarction involvement and maintaining basic blood flow levels (51).

NETs as Circulating Markers of Arterial Thrombotic Diseases

The components of various NETs in plasma have been suggested to be used to predict the severity of diseases such as CAD and ischemic stroke. For example, PAD4 levels are very high in carotid plaques (52), and citrullinated histone H3 (CitH3), double-stranded DNA (dsDNA), neutrophil elastase (NE) levels, the myocardial infarction area, and left ventricular dysfunction are related to the poor prognosis of patients with coronary atherosclerosis and myocardial infarction (19, 21, 41). The plasma levels of the MPO-DNA complex and cell-free DNA (cfDNA) are directly proportional to immune thrombosis in patients with COVID-19 complicated with acute respiratory distress syndrome (6, 53). NET-related tissue factors may also be used as markers (54). In patients with type 2 diabetes, circulating markers of NETs are related to thrombosis and a low fibrinolytic status and can be used as biomarkers for the stratification of patients with diabetes who present a higher risk of vascular complications (55, 56). Immunofluorescence staining showed the presence of NETs in thrombus samples from animal models. In integrin $1\alpha^{-/-}$ mice, the proportion of neutrophils releasing NETs is reduced, and arterial thrombosis is significantly reduced (24). In the mouse myocardial infarction model, the inhibition of PAD4 activity by an intraperitoneal injection of a specific drug reduces the infarct size and improves the prognosis of cardiac ischemia (57).

The Therapeutic Potential of DNase in Patients With Arterial Thrombosis

Lower DNase activity is related to infarct size (58), and increased DNase activity reduces the risk of host tissue damage and thrombosis induced by NETs (59). DNase treatment reduces the content of NETs in plaques and the level of macrophage inflammation, promotes disease remission and improves prognosis (23). The use of t-PA for thrombolytic therapy is the basis for the treatment of thrombotic diseases. The addition of DNase targeting NETs to the standard t-PA treatment regimen increases the therapeutic effect of thrombolytic therapy and improves the prognosis of the disease (37, 51). Taken together, these data indicate that DNase improves the prognosis of subjects with cardiac and cerebral ischemia (37, 51). The role of NETs in arterial thrombosis indicates that DNase

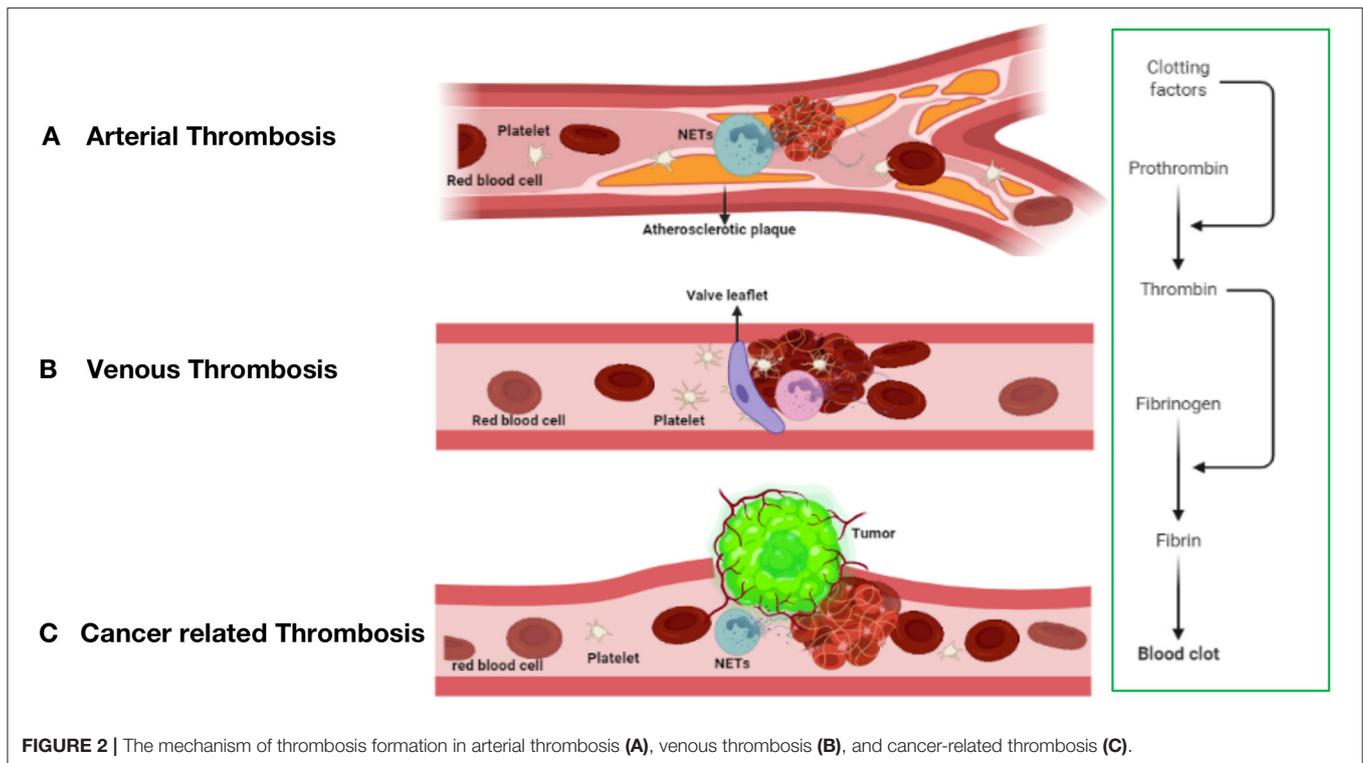


FIGURE 2 | The mechanism of thrombosis formation in arterial thrombosis (A), venous thrombosis (B), and cancer-related thrombosis (C).

degradation of NETs may become a new treatment direction and improve the effectiveness of thrombolytic therapy. In the future, large-scale research should use NETs and their components as disease markers and potential therapeutic targets to reduce atherosclerosis and prevent thrombosis. At the same time, further research is needed to explore the effectiveness and safety of DNase in the treatment of arterial thrombotic diseases.

THE ROLE OF NETs IN VENOUS THROMBOSIS

Unlike arterial thrombosis, venous thrombosis is not caused by endothelial rupture and is mostly due to slow venous blood flow (Figure 2B), a hypercoagulable blood state and venous intima injury (60, 61). Prolonged lack of exercise, pregnancy, and chronic venous blood supply are the main causes of impaired venous blood flow and are associated with an increased risk of DVT development (62). Venous thrombosis is rich in fibrin and red blood cells, and a large amount of white blood cell infiltration is observed (63).

NETs in Deep Vein Thrombosis

The annual incidence of venous thromboembolism (VTE) in the United States is $\sim 1/1000$ (64). Stasis of blood flow in the veins is one of the main causes of DVT, and this process often leads to immune thrombosis. A study compared 150 symptomatic patients with DVT with a control group that was clinically suspected of having DVT but had a negative objective test and

found that compared with the control group, patients with DVT had higher levels of circulating nucleosomes and activated neutrophils. The increase in the levels of the two parameters indicates that the risk of DVT is increased approximately 3 times (65). Deep vein thrombosis (DVT) is the most common complication in patients with traumatic fractures. In patients with traumatic fractures, the levels of citrullinated histone H3 (H3Cit), cfDNA and nucleosome NET biomarkers in plasma were detected, and H3Cit and cfDNA assisted in the diagnosis of DVT in patients with traumatic fractures (66). The thrombus in a patient with microscopic polyangiitis (MPA) complicated with deep vein thrombosis (DVT) was confirmed to be rich in neutrophils (67). Furthermore, NETs exist in the thrombus tissue of patients with venous thromboembolism, which is related to the maturation of human thrombi (61). The formation of NETs may be short-lived, occurring when neutrophils are recruited into the thrombus, and as the thrombus matures, extracellular NETs are degraded. Therefore, NETs are rarely found in a mature thrombus. NETs have been used as human DVT biomarkers (68).

Both NETs and inflammasome activation play a role in the development of DVT. The stimulation of neutrophils induces the formation of NETs and activates Caspase-1. Active Caspase-1 requires NETs as an adhesion surface. NETs and their component histones promote the activation of Caspase-1 in platelets. Colocalized NETs and Caspase-1 and platelet recruitment were observed at the site of thrombosis. Pharmacological inhibition of Caspase-1 substantially reduces DVT in mice, and thrombi are still formed without citrulline histone 3. These data indicate an interaction between NETs and inflammasomes *in vitro* and in the

environment of deep vein thrombosis. This interaction may be an important mechanism supporting venous thrombosis (69). IL-1 β regulates the recruitment of neutrophils, and the inflammasome mediates the activation and secretion of IL-1 β . The NLRP-3 inflammasome mainly acts on Caspase-1 and Caspase-11, leading to the cleavage and activation of IL-1 β and IL-18.

The venous endothelium plays an important role in the formation of DVT. Marie-Luise von Brühl et al. confirmed that blood monocytes and neutrophils adhere to the venous endothelium, providing an initial stimulus for the development of DVT (62). This study confirmed that neutrophils are a key trigger for the formation of DVT. Neutrophils form NETs, which induce FXII-dependent coagulation. In addition, platelets can also promote leukocyte accumulation and fibrin formation by enhancing neutrophil-dependent coagulation. Vascular endothelial activation or damage locally activates complement to release allergens and chemokines C3a and C5a. These pathways synergistically trigger the recruitment and activation of platelets, neutrophils, and monocytes.

The role of platelets in venous thrombosis is not as obvious as that in arterial thrombosis. However, thrombocytosis is considered a risk factor for VTE (70). Pathogens and DAMPs stimulate neutrophils to activate the coagulation system, and this interaction mediates immune thrombosis. Platelets release high mobility histone B1 (HMGB1), which triggers the formation of NETs (71, 72).

Mixed lineage kinase-like (MLKL)-driven neutrophil necrosis is related to venous thromboembolism (73). Human inferior vena cava thrombosis is positive for phosphorylated MLKL, and phosphorylated MLKL induces cell necrosis. In mice, MLKL colocalizes with citrulline histone H3, and a genetic defect in MLKL partially prevents clot formation during inferior vena cava ligation in mice. Platelets activated by VTE induce NETosis, resulting in the release of chromatin and DAMPs, which contribute to the formation of clots (74).

Heparin-induced thrombocytopenia (HIT) is an immune-mediated thrombocytopenia associated with a severe prethrombotic state. HIT induction leads to increased neutrophil adhesion to the venous endothelium. In HIT mice, neutrophils migrate in a retrograde manner through a CXCR2-dependent mechanism and accumulate in the thrombus. After PF4 binds to NETs, it compresses itself and resists degradation by DNase. The PF4-NET complex selectively binds to HIT antibodies, further protecting them from nuclease digestion. In HIT mice, inhibition of NET formation by Padi4 gene disruption or DNase treatment limits the size of venous thrombosis. Neutrophil activation promotes HIT venous thrombosis by enhancing neutrophil-endothelial cell adhesion and neutrophil clot infiltration, in which the PF4-NET-HIT antibody complex causes thrombosis to spread (75). Therefore, strategies to prevent venous thrombosis may be to inhibit the adhesion of neutrophil endothelial cells, prevent the recruitment of neutrophil chemokine-dependent neutrophils to thrombi, or inhibit the release of NETs.

Many substances in plasma induce DVT. IFN γ promotes venous thrombosis through the formation of NETs, and NK cells play an important role in this process. The specific consumption of natural killer cells (NK) leads to reduced NET formation

and reduced thrombus formation (76). C5a, the most effective chemotactic complement activating fragment, is released after C5 protein lysis and is considered a key determinant of neutrophil recruitment and the activation of thrombosis. In the mouse venous thrombosis model, the weight of the thrombin-antithrombin complex is closely related to C5a, which indicates that the process triggered during thrombosis promotes C5a production. *In vitro*, the catalytic efficiency of plasmin-mediated C5a production far exceeds that of thrombin or factor Xa and is similar to the recognized complement C5 convertase. C5 activated by plasmin mediates the production of the membrane attack complex (MAC) (77). Antiphospholipid antibodies (aPLs) activate neutrophils to release NETs, thereby inducing arterial and venous thrombosis inherent in antiphospholipid syndrome (APS). Compared with healthy volunteers, patients with primary antiphospholipid syndrome (APS) have higher levels of cell-free DNA and NETs in serum and plasma. Freshly isolated neutrophils from patients with APS tend to release high levels of spontaneous NETs. In addition, the serum of patients with APS, as well as IgG purified from patients with APS, stimulates the release of NETs from controlled neutrophils. Human monoclonal aPLs, particularly those targeting β 2GPI, also enhance the release of NETs. The APS induction of NET production can be eliminated by the formation of reactive oxygen species and TLR4 inhibitors (78).

In addition to classic thrombosis-related substances, other molecules are also involved in the formation of DVT. Resolvin D4 (RvD4) is an SPM (specialized proresolving mediator) that is enriched at the natural beginning of thrombolysis. After administration, the burden of thrombi is significantly reduced, the infiltration of neutrophils in the thrombus is reduced, the number of monocytes increases, and these cells are in the early stage of apoptosis. The number of cells in the apoptotic state increases. Neutrophils of mice treated with RvD4 are less sensitive to the release of NETs (79). Slc44a2 is a ubiquitous transmembrane protein that has been identified as a receptor for a vascular hematoma factor (VWF). The expression of the human neutrophil antigen 3b (HNA-3b) epitope on the Slc44a2 protein is related to the risk of human venous thrombosis (VT). Mice lacking Slc44a2 showed a substantial reduction in neutrophil recruitment in the inflamed mesenteric venules. Slc44a2/HNA-3a plays an important role in the adhesion and activation of neutrophils in veins under inflammation and specific shearing (80).

NETs in Pulmonary Embolism

Acute pulmonary embolism (PE), isolated or combined with deep vein thrombosis (DVT), is the main cause of death or hospitalization due to venous thromboembolism (VTE) (81), accounting for 5–10% of deaths in hospitalized patients. The acute all-cause mortality of patients with venous thromboembolism is 6.6% (82). The risk of death after pulmonary embolism is particularly high, 2.1 times higher than that of deep vein thrombosis (DVT) (83).

VTE is related to the release of NETs. Extracellular chromatin and citrulline histone H3 (citH3) have been observed in deep vein thrombosis in patients with VTE (61). Compared

with those in healthy controls, NET formation markers in symptomatic patients with VTE were significantly increased (84). The increased formation of NETs reflected by the level of citH3 is positively correlated with impaired fibrinolytic function and may be related to the severity of the disease by enhancing inflammation and the prethrombotic state. A high endogenous thrombin potential (ETP) combined with elevated citH3 levels and prolonged clot lysis time (CLT) are associated with an eight-fold increase in the risk of PE-related death (85, 86). Thrombotic fibrillin clot characteristics and enhanced neutrophil extracellular trap formation are associated with a higher risk of early death in patients with acute PE, which suggests the role of these biomarkers in determining prognosis.

Extracellular DNA in human plasma is also called cell-free DNA (cfDNA) (87–91). The accumulation of cfDNA in the circulation is thought to result from increased cell death and/or activation, impaired cfDNA clearance, and/or decreased endogenous DNase enzyme levels (92). DNA is mainly released by neutrophils through NETosis (93). Circulating cfDNA and nucleosomes are considered surrogate markers of NETs in patients with venous thromboembolism. Elevated levels of circulating nucleosomes and neutrophil elastase/ α 1-antitrypsin complexes are associated with a threefold higher risk of DVT (65). The plasma cfDNA level is positively correlated with d-dimer and von Willebrand factor levels, as well as with MPO activity, suggesting that neutrophils are the main source of plasma cfDNA in patients with venous thrombosis. Nucleic acid-binding polymers have been shown to prevent thrombosis in mice without increasing the risk of bleeding. These polymers bind to DNA, RNA and inorganic polyphosphate molecules with high affinity and inhibit the activation of the intrinsic coagulation pathway induced by nucleic acids and polyphosphates (94). The complex of extracellular DNA, histones and neutrophil-derived peptides stimulates the inflammatory response by activating pattern recognition receptors on immune cells (95, 96). Circulating extracellular DNA is an independent predictor of mortality in elderly patients with venous thromboembolism (97).

Many substances are currently confirmed to be related to the treatment of thrombosis. DNase I pretreatment degrades NETs and reduces the incidence of thrombosis in wild-type (WT) mice (62, 98). The addition of deoxyribonuclease I to tissue plasminogen activator (tPA) significantly accelerates the dissolution of thrombi and human lung thrombi (99). Glucocorticoids exert important anti-inflammatory effects and regulate the inflammatory immune response in the body. In an infected state, glucocorticoids inhibit the formation of NETs, and as the concentration of glucocorticoid treatment increases, the inhibitory effect becomes more obvious (99). Because NETs are positively charged, unfractionated heparin (UFH) may be a more effective anticoagulant in patients with acute PE. Patients with acute PE presenting lactic acid concentrations >2 mM have a higher possibility of suppressing strong NETosis, and thus UFH may be the first choice (97). Heparin has been shown to bind to histones and prevent histone-mediated cytotoxicity of endothelial cells. *In vivo*, heparin reduced the mortality of aseptic inflammation and sepsis in a mouse model. The protective effect of heparin is not related to its anticoagulant properties (100).

Heparin also replaces histones from the chromatin backbone of NETs, thereby destroying the stability of NETs (87). However, recent studies have shown that heparin stimulates NET formation *in vitro*, and the ability of enoxaparin to induce NET production is much lower than that of heparin, while pentosan sodium (fondaparinux) does not induce the formation of NETs (101). The hypothesis of the potential benefits of heparin therapy for patients with PE in real life requires further verification.

NETs AND CANCER-RELATED THROMBOSIS

In current research on thrombosis, an increasing number of people are paying attention to the effect of the formation of neutrophil extracellular traps (NETs) on thrombosis, including thrombosis from the automatic venous system and acquired thrombosis (102, 103), such as tumor-related thrombosis and other diseases. The formation of tumor-associated thrombosis may be related to thrombosis derived from the arteriovenous system, but the specific mechanism is not yet fully understood. This may result from the interaction of several mechanisms (104). The risk of deep vein thrombosis (DVT) in patients with cancer is 5–7 times higher than that in healthy people. Tumor-related thrombosis increases the risk of death by 47 times and seriously affects the survival and prognosis of patients with cancer (105). The cause may be a source of cytokines, and tissue damage caused by radiotherapy and chemotherapy causes patients with tumors to usually be in a hypercoagulable state or a prethrombotic state (106). The mechanism of VTE formation (Virchow's triad) is composed of three components: blood flow stagnation, endothelial injury, and a hypercoagulable state. The latter includes abnormalities in the coagulation and fibrinolytic pathways and platelet activation. Tumor-related thrombosis has been roughly divided into direct mechanisms and indirect mechanisms, which are described below (Figure 2C).

Direct Mechanism

Tumor cells directly induce platelet activation. Activated platelets stimulate tumor cells or normal cells in the tumor microenvironment (TME) to secrete a large amount of thrombo-activating factors, including tissue factor (TF) and collagen exposure. These factors increase vascular permeability and then start the coagulation cascade (107). The expression of podoplanin (PDPN) directly causes platelet activation and aggregation through calcium-dependent lectin-like receptor 2 (CLEC-2) on platelets (108). Tumor cells also secrete platelet agonists, such as ADP and thrombin, which further promote platelet activation through P2Y₁₂ and protease-activated receptor 1/4 (PAR1/4), respectively (109). Phosphatidylserine (PS) on tumor cells also promotes blood coagulation, and PS also serves as a key surface component of the coagulation complex (110). The coagulant secreted by tumor cells has been shown to directly activate the coagulation pathway by activating factor X, stimulating the local TME and even systemic platelet activation and thrombosis. In addition, plasminogen activation inhibitor-1

(PAI-1) is expressed at high levels in tumor cells, inhibiting the process of fibrinolysis (111).

Indirect Mechanism

Inflammatory cytokines secreted by tumor cells cause platelet activation and promote the procoagulant phenotype of endothelial cells. Platelets also promote tumor metastasis through different mechanisms. Activated platelets protect tumor cells from immune recognition in the circulation, promote tumor cell adhesion, and help tumor cells pass through the blood vessel wall. Platelet α -granules released after platelet activation carry a large amount of growth factors, such as PDGF, VEGF and TGF- β . In the TME, these growth factors are conducive to tumor proliferation, angiogenesis and invasiveness, thereby promoting tumor progression (112). At the same time, these factors cooperate with heparanase and matrix metalloproteinases (MMPs) in the TME to further promote tumor-related thrombosis (113).

Previously, researchers postulated that the blood coagulation process and the immune regulation of neutrophils are two processes that do not interfere with each other, but the latest research shows that they are closely related. Neutrophils infiltrating the TME release cytokines and enzymes, which promote immune thrombosis (114). Studies have shown that NETs cause a prethrombotic state and thrombosis through various mechanisms. NETs cause the release of a large number of histones and proteasomes from neutrophils into the blood. NETs serve as mesh scaffolds to physically capture platelets or interact with NETs. Histones increasingly activate platelets and enhance the adhesion, aggregation and release functions of platelets. These changes ultimately lead to fibrin deposition and red blood cell capture, thereby accelerating the clotting process (115). In addition, growth factors secreted by tumor cells also stimulate neutrophils to release NETs (114). The interaction of NETs and platelets provides a new target for the clinical evaluation and treatment of thrombosis, and effective intervention in the formation of tumor-related thrombosis has important clinical significance.

Research on NETs and Tumor-Related Thrombi in Animal Models

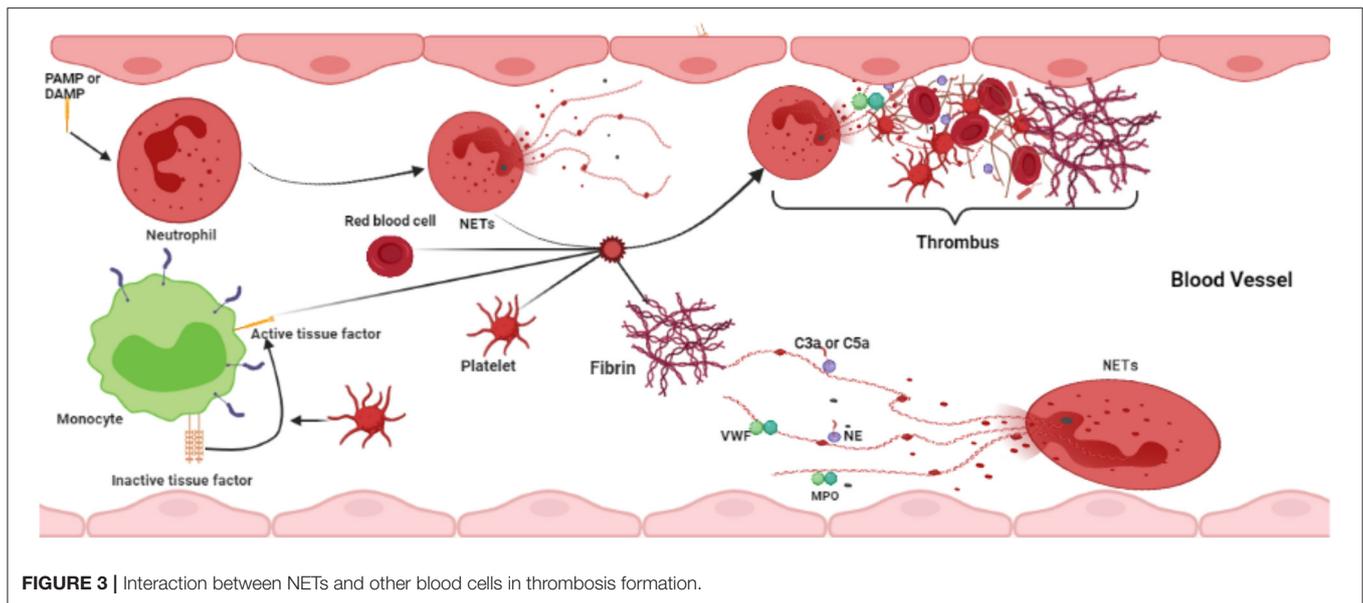
NETs are present at high levels in a variety of malignant tumors, and the close relationship between NETs and tumor-related thrombosis was first confirmed in animal models (116). In mouse models of breast cancer, non-small cell lung cancer and chronic myelogenous leukemia, neutrophils are more likely to induce the formation of NETs (117). In many different types of malignant tumors, increases in plasma free DNA and circulating NET levels are observed, which are closely related to spontaneous thrombosis (118). Compared with tumor-free mice, tumor-bearing mice had increased venous and arterial thrombosis, and citrulline histone H3 (8) was directly detected in the thrombus. Neutrophils isolated from tumor-bearing mice showed higher H3Cit levels upon *in vitro* stimulation, and NETs formed more readily than normal neutrophils (119).

An endotoxin injection into the abdominal cavity of tumor-bearing mice induces the formation of NETs, which in turn induces a prethrombotic state and coagulation dysfunction (47). The accumulation and activation of neutrophils at the site of endothelial injury is considered to be the cause of thrombosis. NETs cause endothelial cell damage. Studies have found that phenyl iodide significantly inhibits this damage, indirectly indicating that endothelial cell damage leads to the formation of thrombi.

Spontaneous DNA-rich thrombi were observed in the lungs of tumor-bearing mice, consistent with the increase in plasma H3Cit and extracellular DNA levels (120). Notably, DNase I pretreatment or depletion of neutrophils completely eliminates arterial thrombosis in tumor-bearing mice and control mice and reduces the size of venous thrombosis in tumor-bearing mice but does not affect venous thrombosis in control mice (8). Based on these studies, DNase I inhibits the formation of NETs or promotes the degradation of NETs, thereby reducing tumor-related thrombosis. Tumor-related neutrophils alter the function and status of host immune cells by producing various chemokines, inflammatory factors and reactive oxygen species. Increased formation and release of NETs were observed when neutrophils were stimulated with tumor-derived granulocyte colony-stimulating factor (G-CSF) (121). Another mechanism by which platelets activate tumor cells to promote the formation of NETs has recently been proposed. Platelets are carriers of tumor-derived exosomes (EVs), which in turn promote the formation of NETs (122). Coincubation of tumor-derived EVs with neutrophils enhanced the formation of NETs, and EVs adhered to the NET complex. The interaction between tumor-derived EVs and neutrophils may lead to tumor-related thrombosis in breast cancer. Similarly, the increase in TF levels in pancreatic tumors accelerates the adhesion of EVs to NETs (118).

Clinical Research and Application of NETs and Cancer-Related Thrombi

Clinical data have shown that patients with tumors are more susceptible to the formation of NETs, and NETs play a very important role in the process of tumor-related thrombosis. NETs released from neutrophils isolated from patients with tumors exhibit increased thrombin and fibrin production in the plasma of healthy patients (123). Therefore, NETs are observed in the thrombi of patients with tumors. Pancreatic cancer is one of the tumors most prone to thrombosis. Recently, human-derived pancreatic cancer cells (AsPC-1) were shown to induce rapid NET formation and release when cocultured with neutrophils from healthy humans (124). Similarly, the NET release capacity of neutrophils from patients with gastric cancer is also increased compared to that of neutrophils from patients without tumors (125). Both human lung cancer tissues and osteosarcoma tissues display neutrophil necrosis and NETs, which are closely related to the therapeutic effect and prognosis (126). Interestingly, in a clinical study involving 936 patients with newly diagnosed tumors or progression after remission, plasma H3Cit levels were related to the occurrence



of VTE in patients with lung cancer and pancreatic cancer but not in patients with other tumors, such as breast cancer and brain cancer. Plasma H3Cit levels were unrelated to VTE in patients with colorectal cancer and prostate cancer (127). We documented the colocalization of H3Cit-positive cells and extracellular H3Cit with extracellular DNA, indicating the presence of NETs in the brain, coronary arteries, and lung microthrombi of patients with stroke presenting with potential tumors. The positive correlation between circulating H3Cit levels and the thrombin-antithrombin complex indicates that NETs may promote the activation of coagulation in patients with cancer. This finding suggests the existence and importance of applying NETs as a predictor of the risk of thromboembolism in patients with lung and pancreatic malignant tumors (128). Peptidylarginine deiminase 4 (PAD4) is expressed at high levels in neutrophils that release NETs. The levels of PAD4 and citrullinated histones reflect the level of NETs in the body (129) and evaluate the tendency of patients with cancer to spontaneously form a thrombus. Tumor-related thrombosis is usually accompanied by an increase in circulating hypercitrullinated neutrophil counts and plasma histone hypercitrullination, but this phenomenon does not occur in healthy patients (8). Histone-DNA complexes are abundant in tumor-associated thrombi, but the plasma levels of circulating nucleosomes and extracellular DNA in patients are correlated with cancer-related stroke and D-dimer levels (130).

Tumors are systemic diseases. This change induces neutrophils to release NETs. Although mechanisms related to tumor-associated thrombosis and the formation of NETs have been proposed, the relationship between this interaction and the formation of NETs remains to be studied. Moreover, further large-sample animal experiments and clinical research support for the use of NETs to predict the sensitivity and specificity of tumor-related thrombosis and the determination of the cutoff value are still needed.

CONCLUSION AND PERSPECTIVES

Neutrophil extracellular traps have become undeniable factors in the field of thrombosis and hemostasis. NETs regulate thrombosis through different pathways and are implicated in the pathophysiology of both arterial and venous thrombotic complications (Figure 3). Undoubtedly, future studies will further advance our knowledge of the temporal and spatial processes of neutrophil-driven thrombus formation and maturation. This information will become very valuable for developing novel antithrombotic therapies. Pharmacological disassembly and degradation of NETs in thrombi may enhance acute thrombosis. In addition, strategies preventing the formation of NETs may reduce thrombogenicity, which might be beneficial in thrombosis prevention. Preclinical and clinical studies investigating these new therapeutic opportunities are now needed to fully understand the efficacy and safety of targeting NETs in thrombosis.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding authors.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

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REFERENCES

- Papayannopoulos V. Neutrophil extracellular traps in immunity and disease. *Nat Rev Immunol.* (2018) 18:134–47. doi: 10.1038/nri.2017.105
- Laridan E, Martinod K, De Meyer SF. Neutrophil extracellular traps in arterial and venous thrombosis. *Semin Thromb Hemost.* (2019) 45:86–93. doi: 10.1055/s-0038-1677040
- Moschonas IC, Tselepis AD. The pathway of neutrophil extracellular traps towards atherosclerosis and thrombosis. *Atherosclerosis.* (2019) 288:9–16. doi: 10.1016/j.atherosclerosis.2019.06.919
- Döring Y, Soehnlein O, Weber C. Neutrophil extracellular traps in atherosclerosis and atherothrombosis. *Circ Res.* (2017) 120:736–43. doi: 10.1161/CIRCRESAHA.116.309692
- Skendros P, Mitsios A, Chrysanthopoulou A, Mastellos DC, Metallidis S, Rafailidis P, et al. Complement and tissue factor-enriched neutrophil extracellular traps are key drivers in COVID-19 immunothrombosis. *J Clin Invest.* (2020) 130:6151–7. doi: 10.1172/JCI141374
- Middleton EA, He XY, Denorme F, Campbell RA, Ng D, Salvatore SP, et al. Neutrophil extracellular traps contribute to immunothrombosis in COVID-19 acute respiratory distress syndrome. *Blood.* (2020) 136:1169–79. doi: 10.1182/blood.2020007008
- Tomar B, Anders HJ, Desai J, Mulay SR. Neutrophils and neutrophil extracellular traps drive necroinflammation in COVID-19. *Cells.* (2020) 9:1383. doi: 10.3390/cells9061383
- Hisada Y, Grover SP, Maqsood A, Houston R, Ay C, Noubouossie DF, et al. Neutrophils and neutrophil extracellular traps enhance venous thrombosis in mice bearing human pancreatic tumors. *Haematologica.* (2020) 105:218–25. doi: 10.3324/haematol.2019.217083
- Zhang J, Yu M, Liu B, Zhou P, Zuo N, Wang Y, et al. Neutrophil extracellular traps enhance procoagulant activity and thrombotic tendency in patients with obstructive jaundice. *Liver Int.* (2021) 41:333–47. doi: 10.1111/liv.14725
- Li T, Wang C, Liu Y, Li B, Zhang W, Wang L, et al. Neutrophil extracellular traps induce intestinal damage and thrombotic tendency in inflammatory bowel disease. *J Crohns Colitis.* (2020) 14:240–53. doi: 10.1093/ecco-jcc/ijj132
- Wang C, Yu M, Zhou P, Li B, Liu Y, Wang L, et al. Endothelial damage and a thin intercellular fibrin network promote haemorrhage in acute promyelocytic leukaemia. *EBioMedicine.* (2020) 60:102992. doi: 10.1016/j.ebiom.2020.102992
- Brinkmann V, Reichard U, Goosmann C, Fauler B, Uhlemann Y, Weiss DS, et al. Neutrophil extracellular traps kill bacteria. *Science.* (2004) 303:1532–5. doi: 10.1126/science.1092385
- Noubouossie DF, Reeves BN, Strahl BD, Key NS. Neutrophils: back in the thrombosis spotlight. *Blood.* (2019) 133:2186–97. doi: 10.1182/blood-2018-10-862243
- Döring Y, Libby P, Soehnlein O. Neutrophil extracellular traps participate in cardiovascular diseases: recent experimental and clinical insights. *Circ Res.* (2020). 126:1228–41. doi: 10.1161/CIRCRESAHA.120.315931
- Li T, Zhang Z, Li X, Dong G, Zhang M, Xu Z, et al. Neutrophil extracellular traps: signaling properties and disease relevance. *Mediators Inflamm.* (2020) 2020:9254087. doi: 10.1155/2020/9254087
- Klopf J, Brostjan C, Eilenberg V, Neumayer C. Neutrophil extracellular traps and their implications in cardiovascular and inflammatory disease. *Int J Mol Sci.* (2021) 22:559. doi: 10.3390/ijms22020559
- Pertiwi KR, de Boer OJ, Mackaaij C, Pabittei DR, de Winter RJ, Li X, van der Wal AC. Extracellular traps derived from macrophages, mast cells, eosinophils and neutrophils are generated in a time-dependent manner during atherothrombosis. *J Pathol.* (2019) 247:505–12. doi: 10.1002/path.5212
- Novotny J, Oberdieck P, Titova A, Pelisek J, Chandraratne S, Nicol P, et al. Thrombus NET content is associated with clinical outcome in stroke and myocardial infarction. *Neurology.* (2020) 94:e2346–60. doi: 10.1212/WNL.0000000000009532
- Hofbauer TM, Mangold A, Scherz T, Seidl V, Panzenböck A, Ondracek AS, et al. Neutrophil extracellular traps and fibrocytes in ST-segment elevation myocardial infarction. *Basic Res Cardiol.* (2019) 114:33. doi: 10.1007/s00395-019-0740-3
- Blasco A, Coronado MJ, Hernández-Terciado F, Martín P, Royuela A, Ramil E, et al. Assessment of neutrophil extracellular traps in coronary thrombus of a case series of patients with COVID-19 and myocardial infarction. *JAMA Cardiol.* (2020) 6:1–6. doi: 10.1001/jamacardio.2020.7308
- Chatzigeorgiou A, Mitroulis I, Chrysanthopoulou A, Legaki AI, Ritis K, Tentolouris N, et al. Increased neutrophil extracellular traps related to smoking intensity and subclinical atherosclerosis in patients with type 2 diabetes. *Thromb Haemost.* (2020) 120:1587–9. doi: 10.1055/s-0040-1714371
- Silvestre-Roig C, Braster Q, Wichapong K, Lee EY, Teulon JM, Berrebeh N, et al. Externalized histone H4 orchestrates chronic inflammation by inducing lytic cell death. *Nature.* (2019) 569:236–40. doi: 10.1038/s41586-019-1167-6
- Josefs T, Barrett TJ, Brown EJ, Quezada A, Wu X, Voisin M, et al. Neutrophil extracellular traps promote macrophage inflammation and impair atherosclerosis resolution in diabetic mice. *JCI Insight.* (2020) 5:e134796. doi: 10.1172/jci.insight.134796
- Dhanesha N, Nayak MK, Doddapattar P, Jain M, Flora GD, Kon S, et al. Targeting myeloid-cell specific integrin $\alpha 9\beta 1$ inhibits arterial thrombosis in mice. *Blood.* (2020) 135:857–61. doi: 10.1182/blood.2019002846
- Carbone F, Bonaventura A, Liberale L, Paolino S, Torre F, Dallegrì F, et al. Atherosclerosis in rheumatoid arthritis: promoters and opponents. *Clin Rev Allergy Immunol.* (2020) 58:1–14. doi: 10.1007/s12016-018-8714-z
- Kessler T, Schunkert H, von Hundelshausen P. Novel approaches to fine-tune therapeutic targeting of platelets in atherosclerosis: a critical appraisal. *Thromb Haemost.* (2020) 120:1492–504. doi: 10.1055/s-0040-1714352
- Reyes-García AML, Aroca A, Arroyo AB, García-Barbera N, Vicente V, González-Conejero R, et al. Neutrophil extracellular trap components increase the expression of coagulation factors. *Biomed Rep.* (2019) 10:195–201. doi: 10.3892/br.2019.1187
- de Vries JJ, Hoppenbrouwers T, Martínez-Torres C, Majied R, Özcan B, van Hoek M, et al. Effects of diabetes mellitus on fibrin clot structure and mechanics in a model of acute neutrophil extracellular traps (NETs) formation. *Int J Mol Sci.* (2020) 21:7107. doi: 10.3390/ijms21197107
- An Z, Li J, Yu J, Wang X, Gao H, Zhang W, et al. Neutrophil extracellular traps induced by IL-8 aggravate atherosclerosis via activation NF- κ B signaling in macrophages. *Cell Cycle.* (2019) 18:2928–38. doi: 10.1080/15384101.2019.1662678
- Zhang YG, Song Y, Guo XL, Miao RY, Fu YQ, Miao CF, et al. Exosomes derived from oxLDL-stimulated macrophages induce neutrophil extracellular traps to drive atherosclerosis. *Cell Cycle.* (2019) 18:2674–84. doi: 10.1080/15384101.2019.1654797
- Li L, Yu X, Liu J, Wang Z, Li C, Shi J, et al. Neutrophil extracellular traps promote aberrant macrophages activation in Behçet's disease. *Front Immunol.* (2021) 11:590622. doi: 10.3389/fimmu.2020.590622
- Hofbauer TM, Ondracek AS, Lang IM. Neutrophil extracellular traps in atherosclerosis and thrombosis. In: *Handbook of Experimental Pharmacology*. Heidelberg: Springer (2020)
- Obama T, Itabe H. Neutrophils as a novel target of modified low-density lipoproteins and an accelerator of cardiovascular diseases. *Int J Mol Sci.* (2020) 21:8312. doi: 10.3390/ijms21218312
- Liu Y, Carmona-Rivera C, Moore E, Seto NL, Knight JS, Pryor M, et al. Myeloid-specific deletion of peptidylarginine deiminase 4 mitigates atherosclerosis. *Front Immunol.* (2018) 9:1680. doi: 10.3389/fimmu.2018.01680
- Van Avondt K, Maegdefessel L, Soehnlein O. Therapeutic targeting of neutrophil extracellular traps in atherogenic inflammation. *Thromb Haemost.* (2019) 119:542–52. doi: 10.1055/s-0039-1678664
- Ling S, Xu JW. NETosis as a pathogenic factor for heart failure. *Oxid Med Cell Longev.* (2021) 2021:6687096. doi: 10.1155/2021/6687096
- Laridan E, Denorme F, Desender L, François O, Andersson T, Deckmyn H, et al. Neutrophil extracellular traps in ischemic stroke thrombi. *Ann Neurol.* (2017) 82:223–32. doi: 10.1002/ana.24993
- Elia E, Montecucco F, Portincasa P, Sahebkar A, Mollazadeh H, Carbone F. Update on pathological platelet activation in coronary thrombosis. *J Cell Physiol.* (2019) 234:2121–33. doi: 10.1002/jcp.27575
- Farkas ÁZ, Farkas VJ, Gubucz I, Szabó L, Bálint K, Tenekedjiev K, et al. Neutrophil extracellular traps in thrombi retrieved during interventional

- treatment of ischemic arterial diseases. *Thromb Res.* (2019) 175:46–52. doi: 10.1016/j.thromres.2019.01.006
40. Abo-Aly M, George B, Shokri E, Chelvarajan L, El-Helw M, Smyth SS, et al. Cangrelor in addition to standard therapy reduces cardiac damage and inflammatory markers in patients with ST-segment elevation myocardial infarction. *J Thromb Thrombolysis.* (2020). doi: 10.1007/s11239-020-02345-8
 41. Hally KE, Parker OM, Brunton-O'Sullivan MM, Harding SA, Larsen PD. Linking neutrophil extracellular traps and platelet activation: a composite biomarker score for predicting outcomes after acute myocardial infarction. *Thromb Haemost.* (2021). doi: 10.1055/s-0041-1728763
 42. Zhou P, Li T, Jin J, Liu Y, Li B, Sun Q, et al. Interactions between neutrophil extracellular traps and activated platelets enhance procoagulant activity in acute stroke patients with ICA occlusion. *EBioMedicine.* (2020) 53:102671. doi: 10.1016/j.ebiom.2020.102671
 43. Yang J, Wu Z, Long Q, Huang J, Hong T, Liu W, et al. Insights into immunothrombosis: the interplay among neutrophil extracellular trap, von Willebrand factor, and ADAMTS13. *Front Immunol.* (2020) 11:610696. doi: 10.3389/fimmu.2020.610696
 44. Essig F, Kollikowski AM, Pham M, Solymosi L, Stoll G, Haeusler KG, et al. Immunohistological analysis of neutrophils and neutrophil extracellular traps in human thrombemboli causing acute ischemic stroke. *Int J Mol Sci.* (2020) 21:7387. doi: 10.3390/ijms21197387
 45. Genchi A, Semerano A, Gullotta GS, Strambo D, Schwarz G, Bergamaschi A, et al. Cerebral thrombi of cardioembolic etiology have an increased content of neutrophil extracellular traps. *J Neurol Sci.* (2021) 423:117355. doi: 10.1016/j.jns.2021.117355
 46. Deng J, Zhao F, Zhang Y, Zhou Y, Xu X, Zhang X, et al. Neutrophil extracellular traps increased by hyperglycemia exacerbate ischemic brain damage. *Neurosci Lett.* (2020) 738:135383. doi: 10.1016/j.neulet.2020.135383
 47. Kim SW, Lee JK. Role of HMGB1 in the Interplay between NETosis and thrombosis in ischemic stroke: a review. *Cells.* (2020) 9:1794. doi: 10.3390/cells9081794
 48. Heo JH, Nam HS, Kim YD, Choi JK, Kim BM, Kim DJ, et al. Pathophysiologic and therapeutic perspectives based on thrombus histology in stroke. *J Stroke.* (2020) 22:64–75. doi: 10.5853/jos.2019.03440
 49. Locke M, Longstaff C. Extracellular histones inhibit fibrinolysis through noncovalent and covalent interactions with fibrin. *Thromb Haemost.* (2021) 121:464–76. doi: 10.1055/s-0040-1718760
 50. Zhang S, Guo M, Liu Q, Liu J, Cui Y. Neutrophil extracellular traps induce thrombogenicity in severe carotid stenosis. *Immun Inflamm Dis.* (2021) 9:1025–36. doi: 10.1002/iid3.466
 51. Peña-Martínez C, Durán-Laforet V, García-Culebras A, Ostos F, Hernández-Jiménez M, Bravo-Ferrer I, et al. Pharmacological modulation of neutrophil extracellular traps reverses thrombotic stroke tPA (tissue-type plasminogen activator) resistance. *Stroke.* (2019) 50:3228–37. doi: 10.1161/STROKEAHA.119.026848
 52. Shimonaga K, Matsushige T, Takahashi H, Hashimoto Y, Yoshiyama M, Ono C, et al. Peptidylarginine deiminase 4 as a possible biomarker of plaque instability in carotid artery stenosis. *J Stroke Cerebrovasc Dis.* (2021) 30:105816. doi: 10.1016/j.jstrokecerebrovasdis.2021.105816
 53. Zuo Y, Zuo M, Yalavarthi S, Gockman K, Madison JA, Shi H, et al. Neutrophil extracellular traps and thrombosis in COVID-19. *J Thromb Thrombolysis.* (2021) 51:446–53. doi: 10.1007/s11239-020-02324-z
 54. Liberale L, Holy EW, Akhmedov A, Bonetti NR, Nietlispach F, Matter CM, et al. Interleukin-1 β mediates arterial thrombus formation via net-associated tissue factor. *J Clin Med.* (2019) 8:2072. doi: 10.3390/jcm8122072
 55. Bryk AH, Prior SM, Plens K, Konieczynska M, Hohendorf J, Malecki MT, et al. Predictors of neutrophil extracellular traps markers in type 2 diabetes mellitus: associations with a prothrombotic state and hypofibrinolysis. *Cardiovasc Diabetol.* (2019) 18:49. doi: 10.1186/s12933-019-0850-0
 56. Berezin A. Neutrophil extracellular traps: The core player in vascular complications of diabetes mellitus. *Diabetes Metab Syndr.* (2019) 13:3017–23. doi: 10.1016/j.dsx.2018.07.010
 57. Du M, Yang W, Schnull S, Gu J, Xue S. Inhibition of peptidyl arginine deiminase-4 protects against myocardial infarction induced cardiac dysfunction. *Int Immunopharmacol.* (2020) 78:106055. doi: 10.1016/j.intimp.2019.106055
 58. Hofbauer TM, Mangold A, Ondracek AS, Panzenböck A, Scherz T, Müller J, et al. Deoxyribonuclease 1 Q222R single nucleotide polymorphism and long-term mortality after acute myocardial infarction. *Basic Res Cardiol.* (2021) 116:29. doi: 10.1007/s00395-021-00864-w
 59. Dinsdale RJ, Hazeldine J, Al Tarrak K, Hampson P, Devi A, Ermogenous C, et al. Dysregulation of the actin scavenging system and inhibition of DNase activity following severe thermal injury. *Br J Surg.* (2020) 107:391–401. doi: 10.1002/bjs.11310
 60. Reitsma PH, Versteeg HH, Middeldorp S. Mechanistic view of risk factors for venous thromboembolism. *Arterioscler Thromb Vasc Biol.* (2012) 32:563–8. doi: 10.1161/ATVBAHA.111.242818
 61. Savchenko AS, Martinod K, Seidman MA, Wong SL, Borissoff JI, Piazza G, et al. Neutrophil extracellular traps form predominantly during the organizing stage of human venous thromboembolism development. *J Thromb Haemost.* (2014) 12:860–70. doi: 10.1111/jth.12571
 62. von Brühl M, Stark K, Steinhart A, Chandraratne S, Konrad I, Lorenz M, et al. Monocytes, neutrophils, and platelets cooperate to initiate and propagate venous thrombosis in mice *in vivo*. *J Exp Med.* (2012) 209:819–35. doi: 10.1084/jem.20112322
 63. Wakefield TW, Strieter RM, Wilke CA, Kadell AM, Wroblewski SK, Burdick MD, et al. Venous thrombosis-associated inflammation and attenuation with neutralizing antibodies to cytokines and adhesion molecules. *Arterioscler Thromb Vasc Biol.* (1995) 15:258–68. doi: 10.1161/01.ATV.15.2.258
 64. Rao AN. Do neutrophil extracellular traps contribute to the heightened risk of thrombosis in inflammatory diseases? *World J Cardiol.* (2015) 7:829. doi: 10.4330/wjc.v7.i12.829
 65. van Montfoort ML, Stephan F, Lauw MN, Hutten BA, Van Mierlo GJ, Solati S, et al. Circulating nucleosomes and neutrophil activation as risk factors for deep vein thrombosis. *Arterioscler Thromb Vasc Biol.* (2013) 33:147–51. doi: 10.1161/ATVBAHA.112.300498
 66. Liu L, Zhang W, Su Y, Chen Y, Cao X, Wu J. The impact of neutrophil extracellular traps on deep venous thrombosis in patients with traumatic fractures. *CLIN CHIM ACTA.* (2021) 519:231–8. doi: 10.1016/j.cca.2021.04.021
 67. Nakazawa D, Tomaru U, Yamamoto C, Jodo S, Ishizu A. Abundant neutrophil extracellular traps in thrombus of patient with microscopic polyangiitis. *FRONT IMMUNOL.* (2012) 3:333. doi: 10.3389/fimmu.2012.00333
 68. Diaz JA, Fuchs TA, Jackson TO, Kremer HJ, Lammle B, Henke PK, et al. Plasma DNA is elevated in patients with deep vein thrombosis. *J Vasc Surg Venous Lymphat Disord.* (2013) 1:2. doi: 10.1016/j.jvsv.2012.12.002
 69. Campos J, Ponomaryov T, De Prendergast A, Whitworth K, Smith CW, Khan AO, et al. Neutrophil extracellular traps and inflammasomes cooperatively promote venous thrombosis in mice. *Blood Adv.* (2021) 5:2319–24. doi: 10.1182/bloodadvances.2020003377
 70. Simanek R, Vormittag R, Ay C, Alguel G, Dunkler D, Schwarzinger I, et al. High platelet count associated with venous thromboembolism in cancer patients: results from the Vienna Cancer and Thrombosis Study (CATS). *J Thromb Haemost.* (2010) 8:114–20. doi: 10.1111/j.1538-7836.2009.03680.x
 71. Zhang H, Zhou Y, Qu M, Yu Y, Chen Z, Zhu S, et al. Tissue factor-enriched neutrophil extracellular traps promote immunothrombosis and disease progression in sepsis-induced lung injury. *Front Cell Infect Microbiol.* (2021) 11:677902. doi: 10.3389/fcimb.2021.677902
 72. Dyer MR, Chen Q, Haldeman S, Yazdani H, Hoffman R, Loughran P, et al. Deep vein thrombosis in mice is regulated by platelet HMGB1 through release of neutrophil-extracellular traps and DNA. *Sci Rep.* (2018) 8:2068. doi: 10.1038/s41598-018-20479-x
 73. Stark K, Philipp V, Stockhausen S, Busse J, Antonelli A, Miller M, et al. Disulfide HMGB1 derived from platelets coordinates venous thrombosis in mice. *Blood.* (2016) 128:2435–49. doi: 10.1182/blood-2016-04-710632
 74. Nakazawa D, Desai J, Steiger S, Müller S, Devarapu SK, Mulay SR, et al. Activated platelets induce MLKL-driven neutrophil necroptosis and release of neutrophil extracellular traps in venous thrombosis. *Cell Death Discov.* (2018) 4:71. doi: 10.1038/s41420-018-0073-2
 75. Gollomp K, Kim M, Johnston I, Hayes V, Welsh J, Arepally GM, et al. Neutrophil accumulation and NET release contribute to thrombosis in HIT. *JCI Insight.* (2018) 3:99445. doi: 10.1172/jci.insight.99445

76. Bertin FR, Rys RN, Mathieu C, Laurance S, Lemarie CA, Blostein MD. Natural killer cells induce neutrophil extracellular trap formation in venous thrombosis. *J Thromb Haemost.* (2019) 17:403–14. doi: 10.1111/jth.14339
77. Foley JH, Walton BL, Aleman MM, O'Byrne AM, Lei V, Harrasser M, et al. Complement activation in arterial and venous thrombosis is mediated by plasmin. *eBiomedicine.* (2016) 5:175–82. doi: 10.1016/j.ebiom.2016.02.011
78. Yalavarthi S, Gould TJ, Rao AN, Mazza LF, Morris AE, Núñez-Álvarez C, et al. Release of neutrophil extracellular traps by neutrophils stimulated with antiphospholipid antibodies: a newly identified mechanism of thrombosis in the antiphospholipid syndrome. *Arthritis Rheumatol.* (2015) 67:2990–3003. doi: 10.1002/art.39247
79. Cherpokova D, Jouvene CC, Libreros S, DeRoo EP, Chu L, de la Rosa X, et al. Resolvin D4 attenuates the severity of pathological thrombosis in mice. *Blood.* (2019) 134:1458–68. doi: 10.1182/blood.2018886317
80. Zirka G, Robert P, Tilburg J, Tishkova V, Maracle CX, Legendre P, et al. Impaired adhesion of neutrophils expressing Slc44a2/HNA-3b to VWF protects against NETosis under venous shear rates. *Blood.* (2021) 137:2256–66. doi: 10.1182/blood.2020008345
81. Wendelboe AM, Raskob GE. Global burden of thrombosis: epidemiologic aspects. *Circ Res.* (2016) 118:1340–7. doi: 10.1161/CIRCRESAHA.115.306841
82. Spencer FA, Emery C, Lessard D, Anderson F, Emani S, Aragam J, et al. The Worcester Venous Thromboembolism study: a population-based study of the clinical epidemiology of venous thromboembolism. *J Gen Intern Med.* (2006) 21:722–7. doi: 10.1111/j.1525-1497.2006.00458.x
83. Naess IA, Christiansen SC, Romundstad P, Cannegieter SC, Rosendaal FR, Hammerstrom J. Incidence and mortality of venous thrombosis: a population-based study. *J Thromb Haemost.* (2007) 5:692–9. doi: 10.1111/j.1538-7836.2007.02450.x
84. Kimball AS, Obi AT, Diaz JA, Henke PK. The emerging role of NETs in venous thrombosis and immunothrombosis. *Front Immunol.* (2016) 7:236. doi: 10.3389/fimmu.2016.00236
85. Zabczyk M, Natorska J, Janion-Sadowska A, Metzger-Gumiela A, Polak M, Plens K, et al. Prothrombotic fibrin clot properties associated with NETs formation characterize acute pulmonary embolism patients with higher mortality risk. *Sci Rep.* (2020) 10:11433. doi: 10.1038/s41598-020-68375-7
86. Martinod K, Demers M, Fuchs TA, Wong SL, Brill A, Gallant M, et al. Neutrophil histone modification by peptidylarginine deiminase 4 is critical for deep vein thrombosis in mice. *Proc Natl Acad Sci USA.* (2013) 110:8674–9. doi: 10.1073/pnas.1301059110
87. Fuchs TA, Brill A, Duerschmied D, Schatzberg D, Monestier M, Myers DJ, et al. Extracellular DNA traps promote thrombosis. *Proc Natl Acad Sci USA.* (2010) 107:15880–5. doi: 10.1073/pnas.1005743107
88. Zabczyk M, Natorska J, Janion-Sadowska A, Malinowski KP, Janion M, Undas A. Elevated lactate levels in acute pulmonary embolism are associated with prothrombotic fibrin clot properties: contribution of NETs formation. *J Clin Med.* (2020) 9:953. doi: 10.3390/jcm9040953
89. Keshari RS, Jyoti A, Dubey M, Kothari N, Kohli M, Bogra J, et al. Cytokines induced neutrophil extracellular traps formation: implication for the inflammatory disease condition. *PLoS ONE.* (2012) 7:e48111. doi: 10.1371/journal.pone.0048111
90. Li T, Peng R, Wang F, Hua L, Liu S, Han Z, et al. Lysophosphatidic acid promotes thrombus stability by inducing rapid formation of neutrophil extracellular traps: A new mechanism of thrombosis. *J Thromb Haemost.* (2020) 18:1952–64. doi: 10.1111/jth.14839
91. Gould TJ, Lysov Z, Liaw PC. Extracellular DNA and histones: double-edged swords in immunothrombosis. *J Thromb Haemost.* (2015) 13:S82–91. doi: 10.1111/jth.12977
92. Tamkovich SN, Cherepanova AV, Kolesnikova EV, Rykova EY, Pyshnyi DV, Vlassov VV, et al. Circulating DNA and DNase activity in human blood. *Ann N Y Acad Sci.* (2006) 1075:191–6. doi: 10.1196/annals.1368.026
93. Kaplan MJ, Radic M. Neutrophil extracellular traps: double-edged swords of innate immunity. *J Immunol.* (2012) 189:2689–95. doi: 10.4049/jimmunol.1201719
94. Jain S, Pitoc GA, Holl EK, Zhang Y, Borst L, Leong KW, et al. Nucleic acid scavengers inhibit thrombosis without increasing bleeding. *Proc Natl Acad Sci U S A.* (2012) 109:12938–43. doi: 10.1073/pnas.1204928109
95. Semeraro F, Ammollo CT, Morrissey JH, Dale GL, Friese P, Esmon NL, et al. Extracellular histones promote thrombin generation through platelet-dependent mechanisms: involvement of platelet TLR2 and TLR4. *Blood.* (2011) 118:1952–61. doi: 10.1182/blood-2011-03-343061
96. Garcia-Romo GS, Caielli S, Vega B, Connolly J, Allantaz F, Xu Z, et al. Netting neutrophils are major inducers of type I IFN production in pediatric systemic lupus erythematosus. *Sci Transl Med.* (2011) 3:20r-73r. doi: 10.1126/scitranslmed.3001201
97. Jiménez-Alcázar M, Limacher A, Panda R, Méan M, Bitterling J, Peine S, et al. Circulating extracellular DNA is an independent predictor of mortality in elderly patients with venous thromboembolism. *PLoS ONE.* (2018) 13:e191150. doi: 10.1371/journal.pone.0191150
98. Brill A, Fuchs TA, Savchenko AS, Thomas GM, Martinod K, De Meyer SE, et al. Neutrophil extracellular traps promote deep vein thrombosis in mice. *J Thromb Haemost.* (2012) 10:136–44. doi: 10.1111/j.1538-7836.2011.04544.x
99. Meng Z, Sun K, Xie Z, Li Y, Li Y, Song J. Glucocorticoids regulate the activation of neutrophils and inhibit the formation of pulmonary embolism. *Iran J Immunol.* (2020) 17:303–12.
100. Wildhagen KC, Garcia DFP, Reutelingsperger CP, Schrijver R, Areste C, Ortega-Gomez A, et al. Nonanticoagulant heparin prevents histone-mediated cytotoxicity *in vitro* and improves survival in sepsis. *Blood.* (2014) 123:1098–101. doi: 10.1182/blood-2013-07-514984
101. Lelliott PM, Momota M, Shibahara T, Lee M, Smith NI, Ishii KJ, et al. Heparin induces neutrophil elastase-dependent vital and lytic NET formation. *Int Immunol.* (2020) 32:359–68. doi: 10.1093/intimm/dxz084
102. Grilz E, Königsbrügge O, Posch F, Schmidinger M, Pirker R, Lang IM, et al. Frequency, risk factors, and impact on mortality of arterial thromboembolism in patients with cancer. *Haematologica.* (2018) 103:1549–56. doi: 10.3324/haematol.2018.192419
103. Dardiotis E, Aloizou AM, Markoula S, Siokas V, Tsarouhas K, Tzanakakis G, et al. Cancer-associated stroke: pathophysiology, detection and management (Review). *Int J Oncol.* (2019) 54:779–96. doi: 10.3892/ijo.2019.4669
104. Hisada Y, Mackman N. Cancer-associated pathways and biomarkers of venous thrombosis. *Blood.* (2017) 130:1499–506. doi: 10.1182/blood-2017-03-743211
105. Abdol Razak NB, Jones G, Bhandari M, Berndt MC, Metharom P. Cancer-associated thrombosis: an overview of mechanisms, risk factors, and treatment. *Cancers.* (2018) 10:380. doi: 10.3390/cancers10100380
106. Falanga A, Schieppati F, Russo L. Pathophysiology 1. Mechanisms of thrombosis in cancer patients. *Cancer Treat Res.* (2019) 179:11–36. doi: 10.1007/978-3-030-20315-3_2
107. Palacios-Acedo AL, Mège D, Crescence L, Dignat-George F, Dubois C, Panicot-Dubois L. Platelets, thrombo-inflammation, and cancer: collaborating with the enemy. *Front Immunol.* (2019) 10:1805. doi: 10.3389/fimmu.2019.01805
108. Suzuki-Inoue K. Platelets and cancer-associated thrombosis: focusing on the platelet activation receptor CLEC-2 and podoplanin. *Blood.* (2019) 134:1912–8. doi: 10.1182/blood.2019001388
109. Rwbisira Rundinga G, Khan GJ, Kong Y. Protease-activated receptor 4 (PAR4): a promising target for antiplatelet therapy. *Int J Mol Sci.* (2018) 19:573. doi: 10.3390/ijms19020573
110. Birge RB, Boeltz S, Kumar S, Carlson J, Wanderley J, Calianese D, et al. Phosphatidylserine is a global immunosuppressive signal in efferocytosis, infectious disease, and cancer. *Cell Death Differ.* (2016) 23:962–78. doi: 10.1038/cdd.2016.11
111. Lucotti S, Muschel RJ. Platelets and metastasis: new implications of an old interplay. *Front Oncol.* (2020) 10:1350. doi: 10.3389/fonc.2020.01350
112. Mendoza-Almanza G, Burciaga-Hernández L, Maldonado V, Melendez-Zajgla J, Olmos J. Role of platelets and breast cancer stem cells in metastasis. *World J Stem Cells.* (2020) 12:1237–54. doi: 10.4252/wjsc.v12.i11.1237
113. Jayatilake KM, Hulett MD. Heparanase and the hallmarks of cancer. *J Transl Med.* (2020) 18:453. doi: 10.1186/s12967-020-02624-1
114. Selders GS, Fetz AE, Radic MZ, Bowlin GL. An overview of the role of neutrophils in innate immunity, inflammation and host-biomaterial integration. *Regenerat Biomater.* (2017) 4:55–68. doi: 10.1093/rb/rbw041
115. Chen Z, Zhang H, Qu M, Nan K, Cao H, Cata JP, et al. Review: the emerging role of neutrophil extracellular traps in sepsis and

- sepsis-associated thrombosis. *Front Cell Infect Microbiol.* (2021) 11:653228. doi: 10.3389/fcimb.2021.653228
116. Shinde-Jadhav S, Mansure JJ, Rayes RF, Marcq G, Ayoub M, Skowronski R, et al. Role of neutrophil extracellular traps in radiation resistance of invasive bladder cancer. *Nat Commun.* (2021) 12:2776. doi: 10.1038/s41467-021-23086-z
 117. Snoderly HT, Boone BA, Bennowitz MF. Neutrophil extracellular traps in breast cancer and beyond: current perspectives on NET stimuli, thrombosis and metastasis, and clinical utility for diagnosis and treatment. *Breast Cancer Res.* (2019) 21:145. doi: 10.1186/s13058-019-1237-6
 118. Leal AC, Mizurini DM, Gomes T, Rochael NC, Saraiva EM, Dias MS, et al. Tumor-derived exosomes induce the formation of neutrophil extracellular traps: implications for the establishment of cancer-associated thrombosis. *Sci Rep.* (2017) 7:6438. doi: 10.1038/s41598-017-06893-7
 119. Li M, Lin C, Leso A, Nefedova Y. Quantification of citrullinated histone H3 bound DNA for detection of neutrophil extracellular traps. *Cancers.* (2020) 12:3424. doi: 10.3390/cancers12113424
 120. Thälén C, Hisada Y, Lundström S, Mackman N, Wallén H. Neutrophil extracellular traps: villains and targets in arterial, venous, and cancer-associated thrombosis. *Arterioscler Thromb Vasc Biol.* (2019) 39:1724–38. doi: 10.1161/ATVBAHA.119.312463
 121. Singel KL, Segal BH. Neutrophils in the tumor microenvironment: trying to heal the wound that cannot heal. *Immunol Rev.* (2016) 273:329–43. doi: 10.1111/imr.12459
 122. Zarà M, Guidetti GF, Camera M, Canobbio I, Amadio P, Torti M, et al. Biology and role of extracellular vesicles (EVs) in the pathogenesis of thrombosis. *Int J Mol Sci.* (2019) 20:2840. doi: 10.3390/ijms20112840
 123. Mutua V, Gershwin LJ, A. Review of neutrophil extracellular traps (NETs) in disease: potential anti-NETs therapeutics. *Clin Rev Allergy Immunol.* (2021) 61:194–211. doi: 10.1007/s12016-020-08804-7
 124. Abdol Razak N, Elaskalani O, Metharom P. Pancreatic cancer-induced neutrophil extracellular traps: a potential contributor to cancer-associated thrombosis. *Int J Mol Sci.* (2017) 18:487. doi: 10.3390/ijms18030487
 125. Zhu T, Zou X, Yang C, Li L, Wang B, Li R, et al. Neutrophil extracellular traps promote gastric cancer metastasis by inducing epithelial mesenchymal transition. *Int J Mol Med.* (2021) 48:127. doi: 10.3892/ijmm.2021.4960
 126. Lecot P, Sarabi M, Pereira Abrantes M, Mussard J, Koenderman L, Caux C, et al. Neutrophil heterogeneity in cancer: from biology to therapies. *Front Immunol.* (2019) 10:2155. doi: 10.3389/fimmu.2019.02155
 127. Mauracher LM, Posch F, Martinod K, Grilz E, Däullary T, Hell L, et al. Citrullinated histone H3, a biomarker of neutrophil extracellular trap formation, predicts the risk of venous thromboembolism in cancer patients. *J Thromb Haemost.* (2018) 16:508–18. doi: 10.1111/jth.13951
 128. Thälén C, Demers M, Blomgren B, Wong SL, von Arbin M, von Heijne A, et al. NETosis promotes cancer-associated arterial microthrombosis presenting as ischemic stroke with troponin elevation. *Thromb Res.* (2016) 139:56–64. doi: 10.1016/j.thromres.2016.01.009
 129. Raup-Konsavage WM, Wang Y, Wang WW, Feliers D, Ruan H, Reeves WB. Neutrophil peptidyl arginine deiminase-4 has a pivotal role in ischemia/reperfusion-induced acute kidney injury. *Kidney Int.* (2018) 93:365–74. doi: 10.1016/j.kint.2017.08.014
 130. Bang OY, Chung JW, Cho YH, Oh MJ, Seo WK, Kim GM, et al. Circulating DNAs, a marker of neutrophil extracellular traps and cancer-related stroke: the OASIS-cancer study. *Stroke.* (2019) 50:2944–7. doi: 10.1161/STROKEAHA.119.026373

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