



Immunotherapeutic Strategies in Cancer and Atherosclerosis—Two Sides of the Same Coin

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Nettersheim FS, Picard FSR, Hoyer FF and Winkels H (2022) Immunotherapeutic Strategies in Cancer and Atherosclerosis – Two Sides of the Same Coin. Front. Cardiovasc. Med. 8:812702. doi: 10.3389/fcvm.2021.812702 The development and clinical approval of immunotherapies has revolutionized cancer therapy. Although the role of adaptive immunity in atherogenesis is now well-established and several immunomodulatory strategies have proven beneficial in preclinical studies, anti-atherosclerotic immunotherapies available for clinical application are not available. Considering that adaptive immune responses are critically involved in both carcinogenesis and atherogenesis, immunotherapeutic approaches for the treatment of cancer and atherosclerosis may exert undesirable but also desirable side effects on the other condition, respectively. For example, the high antineoplastic efficacy of immune checkpoint inhibitors, which enhance effector immune responses against tumor cells by blocking co-inhibitory molecules, was recently shown to be constrained by substantial proatherogenic properties. In this review, we outline the specific role of immune responses in the development of cancer and atherosclerosis and discuss whether anti-atherosclerotic immunotherapies may similarly have an impact on carcinogenesis.

Keywords: tumor, atherogenesis, cardiovascular disease, immunotherapy, immunity, T cell, checkpoint inhibition, co-stimulatory molecule

INTRODUCTION

Although prevention strategies and therapeutic opportunities have been significantly improved during the past decades, atherosclerotic cardiovascular diseases (CVD) and cancer still represent the two most common causes of death worldwide (1). As already recognized by Rudolph Virchow in the nineteenth century (2, 3), the critical role of inflammatory processes in atherogenesis and carcinogenesis is now well-established and has prompted investigation of strategies to combat these deadly diseases by modulating underlying immune responses (4–10). Several anti-cancer immunotherapies, such as cytokines, antibodies targeting immune cell receptors, or immune checkpoints, dendritic cell therapy, and chimeric antigen receptor (CAR) T cell therapy, already found their way into clinical practice and thereby revolutionized cancer treatment (9, 11). In stark contrast, clinically approved immunotherapies for CVD are still not available [except for antibodies targeting proprotein convertase subtilisin/kexin 9 (PCSK9) to lower low-density lipoprotein (LDL) cholesterol, representing an immunotherapeutic approach in a broader sense (12, 13)]. In 2017, the CANTOS trial demonstrated that administration of an antibody directed

against the pro-inflammatory cytokine interleukin-1 β (IL-1 β) reduced cardiovascular events in patients with coronary artery disease (CAD), thereby providing first evidence for effectiveness of an immunotherapy in CVD (14). Yet, this therapy increased the risk of fatal infections and did not reduce mortality, which consequently prevented its approval for treatment of CVD (14). CANTOS illustrated the central dilemma of many immunomodulatory strategies: Broad interventions in the immune system can have detrimental side effects. In general, anti-atherosclerotic strategies are geared toward suppression of vascular inflammation (5, 8, 15), whereas immune-based cancer treatments aim at enhancing immune responses against tumor cells (7, 9). The therapeutic efficacy of several anti-cancer immunotherapies is constrained by their proimmunogenic (and thus proatherogenic) properties, increasing the risk to develop CVD in patients (16). Particularly, immune checkpoint inhibitors directly aggravate atherosclerotic plaque growth in patients (17). Whereas, cancer survival has dramatically improved over the past few decades (18), the exposure of cancer survivors to therapy-induced cardiovascular risk represents an emerging problem, which leads to excess cardiovascular mortality and thus significantly affects long-term prognosis (19-22). This problem is relevant, as the global cancer burden is expected to increase by ${\sim}47\%$ within the next 20 years and to reach more than 28 million cases in 2040 (23).

In recent years, vaccination strategies aiming to either induce immune responses against tumor-specific neoantigens (4) or to suppress immunity against atherosclerosis-related autoantigens (6) have emerged. Immunization strategies are promising as they enable specific immunomodulation without impairing host defense responses or accelerating progression of atherosclerosis. Whereas, anti-atherosclerotic vaccination strategies are still in their infancy (6), therapeutic cancer vaccines are already being investigated in clinical trials (4).

In this review, we will provide an overview of current immunomodulatory concepts for treatment of cancer and atherosclerosis with a focus on their reciprocal interactions and consequences. Finally, we will highlight the potential of immunization strategies against cancer and CVD that enable targeted, antigen-specific immunity without affecting the immune system.

INFLAMMATION AND ADAPTIVE IMMUNITY IN ATHEROGENESIS

Atherosclerosis involves formation of lipid-laden plaques in large and medium-sized arteries (24), which may rupture or erode and give rise to acute thrombotic vessel occlusion (25). Plaque formation primarily occurs in regions with disturbed blood flow and low endothelial shear stress (26). Such hemodynamic alterations induce a cascade of endothelial dysfunction, subendothelial accumulation and subsequent oxidation of lipoproteins, and finally an inflammatory response that is characterized by monocyte infiltration and foam cell formation (25, 26). Extensive research during the past decades has indicated that plaque-related inflammation is not simply a passive process but is rather orchestrated by an adaptive immune response involving T cells and humoral immunity (27).

T cells derive from hematopoietic progenitor cells and undergo a complex maturation and selection process in the thymus, which is characterized by development of a unique, antigen-specific T cell receptor (TCR) through random genetic recombination (28) and elimination of cells that are either nonfunctional or bind self-antigens with too high-affinity, which are potentially dangerous for the host (29). The high prevalence of autoimmune disorders indicates the insufficiency of this process. Eventually, the TCR and one of its co-receptors CD4 or CD8 are expressed on the T cell surface, which is released into the periphery and circulates through the body to encounter its cognate antigen (30). Activation of a naïve T cell requires two signals. First, the TCR must be bound by its cognate antigen: CD4⁺ T cell activation requires presentation of an antigenic peptide-sequence, the so-called epitope, on major histocompatibility complex class II (MHC-II) molecules, which are exclusively expressed by professional antigen-presenting cells (APCs), such as dendritic cells, macrophages and B cells. CD8⁺ T cells recognize antigens presented on MHC-I molecules, which are expressed by all nucleated cells (31). Second, the T cell must simultaneously receive a proper co-stimulatory signal, that is binding of a specific receptor (such as CD28) by its ligand expressed on the APC (32). Once activated, T cells proliferate and CD8⁺ T cells become cytotoxic, whereas CD4⁺ T cells can differentiate into a variety of different subtypes, which are characterized by expression of specific surface markers, transcription factors (TFs) and cytokines (33). For example, T helper 1 (T_H1) cells, which are characterized by expression of the TF T-box expressed in T cells (T-bet), exert proinflammatory effects through production of interferon gamma (IFN- γ) (**Figure 1**). In contrast, regulatory T cells (T_{regs}), which are characterized by expression of the TF forkhead box protein P3 (FoxP3), produce the anti-inflammatory cytokines IL-10 and transforming growth factor beta (TGF- β) and thus ensure immune tolerance. For a thorough overview of different T cell subtypes and their role in atherosclerosis the interested reader is referred to Saigusa et al. (34).

Presence of T cells in atherosclerotic plaques was firstly described by Hansson and colleagues more than 30 years ago (35). T cells within the plaques were activated (36) and in vitro work showed lesional CD4⁺ T cells responding to oxidized lowdensity lipoprotein (oxLDL), which established the theory of T cells contributing to plaque formation (37). By now, CD4⁺ and CD8⁺ T cell responses against plaque-associated autoantigens have been identified to modulate atherogenesis (34, 38, 39). Whereas, T cell reactivity against LDL was originally thought to be induced by oxidation-dependent generation of neoepitopes representing "altered self" (37), more recent work has identified CD4⁺ T cells responding to peptides of native Apolipoprotein B (ApoB), the core protein of LDL, chylomicrons, and other lipoprotein particles. Several unmodified ApoB-peptides have been found to bind murine and human MHC-II molecules with high affinity and thereby evoke a CD4⁺ T cell response (40-43). Such ApoB-reactive (ApoB⁺) CD4⁺ T cells mainly comprise T_{regs}, which confer atheroprotective properties in



healthy humans, but coexpress TFs typical of proatherogenic $T_{\rm H}1$ or $T_{\rm H}17$ cells in individuals with subclinical atherosclerosis as determined by carotid ultrasound (42). Preclinical studies further elucidated that CD4⁺ $T_{\rm regs}$, particularly those reactive to ApoB, gradually acquire proatherogenic $T_{\rm H}1/T_{\rm H}17$, $T_{\rm H}1/T_{\rm reg}$ or T follicular helper ($T_{\rm FH}$) phenotypes during atherogenesis (43–45). Whereas, therapeutic interventions that aim to stabilize and/or expand ApoB⁺ $T_{\rm regs}$ hold promise for atherosclerosis prevention and treatment, immunomodulatory therapies causing destabilization of $T_{\rm regs}$ naturally aggravate progression of atherosclerotic lesions (46).

Besides T cells, humoral immune responses against plaqueassociated autoantigens have been implicated in atherogenesis (38, 47). Antibodies directed against oxLDL are detectable in human plaques (48) and in plasma samples of humans with our without atherosclerotic CVD (49, 50). Accordingly, B cells can be found in healthy and atherosclerotic vessels, especially in arterial tertiary lymphoid organs located in the adventitia (51).

Autoantibodies against oxLDL were shown to block uptake of oxLDL by macrophages (52, 53) and to confer atheroprotection (54). Genetic B-cell depletion aggravated atherosclerosis in LDLreceptor-deficient $(Ldlr^{-/-})$ mice (55). Yet, depletion of mature B cells through administration of a CD20 monoclonal antibody was unexpectedly atheroprotective in Apolipoprotein E-deficient $(Apoe^{-/-})$ and $Ldlr^{-/-}$ mice (56, 57). This treatment preserved the production of natural IgM antibodies directed against oxLDL but reduced anti-oxLDL IgG antibodies (56). Adoptive transfer of B2 B cells, but not B1 B cells, to lymphocyte or B cell deficient $Apoe^{-/-}$ mice was proatherogenic (57). B cells mainly consist of B2 B cells (which are thus termed conventional B cells) that undergo maturation in the spleen and can produce highaffinity IgG antibodies after receiving T cell help (58). B1 B cells represent a specialized B cell subpopulation: They develop in the fetal and neonatal period, harbor the capability of selfrenewal, mainly reside in body cavities and are characterized by the production of so called "natural" IgM antibodies (59).

Taken together, different B cell subsets and antibody subtypes may exert diametral functions in atherogenesis and therapeutic modulation of humoral immune responses could represent an attractive anti-atherosclerotic strategy but also promote atherosclerosis progression.

INFLAMMATION AND ADAPTIVE IMMUNITY IN CANCER

In the nineteenth century, the German physicians Rudolph Virchow, Wilhelm Busch and Friedrich Fehleisen independently hypothesized that inflammation may affect carcinogenesis (2, 60). Whereas, Virchow assumed that leukocyte infiltrates represented an underlying cause of cancer (2), Busch and Fehleisen suggested that inflammation may reverse tumorigenesis (60). After independently observing involution of malignancies in patients with erysipelas, they demonstrated tumor regression in cancer patients upon intentional infection with bacteria isolated from erysipelas (61, 62). Later, the American surgeon William Coley reported disappearance of tumors in patients with inoperable sarcoma or other types of cancer after treatment with heat-inactivated bacteria which was termed "Coley's Toxin" (63). In 1909, Paul Ehrlich suggested that cellular immunity may recognize neoplastic cells and protect from tumor development, although he was not able to experimentally substantiate this hypothesis (64). First experimental proof for anti-tumor immune responses was provided by Gross and Foley around 1950 (65, 66) and Paul Ehrlich's concept was adopted by Lewis Thomas and Sir Frank Macfarlane Burnet who proposed that lymphocytes recognize and target cancer cells through their expression of tumor-specific antigens, similar to homograft rejection (known as immunosurveillance) (7, 67). Yet, this theory was abandoned after immunologically impaired animals, such as athymic nude mice, showed similar susceptibility to experimentally induced tumors (68, 69). Several limitations of these experiments became evident: Nude mice-despite lack of T and B cells-are not completely immunocompromised and especially susceptible to 3-methylcholanthrene, the chemical carcinogen which was used for tumor induction (due to expression of highly active enzyme isoforms involved in biotransformation of the chemical) (68). Novel immunocompromised mouse models with pure genetic backgrounds demonstrated that lymphocyte deficiency (70), lack of perforin (an important component of cytotoxic T lymphocyte granules) (71, 72), and ablation of proinflammatory cytokine signaling (70-73) increased tumor susceptibility in mice, which led to the renaissance of the immunosurveillance theory (68). This preclinical evidence was supported by studies reporting an increased cancer risk in immunocompromised patients (74-76) and that tumor lymphocyte infiltration predicts better outcome (77, 78). Despite overall proof in support of the immunosurveillance theory was provided, subsequent work demonstrated that immunity may also exert tumor-sculpting effects (68): Tumors derived from immunocompromised mice were rejected more frequently when transplanted into immunocompetent recipients than tumors derived from wild-type controls (70, 72, 79, 80). Thus, the immune system of the wild-type donors must have shaped tumors to become less immunogenic and more resistant to the hosts (uncompromised) immune response.

To account for the dual role of the immune system in tumor development, G.P Dunn and R.D. Schreiber proposed the groundbreaking "immunoediting" or "three E's" theory in 2002 (68), which is the current explanation of tumor-related immune responses (81). The theory involves three processes: (1) In the elimination phase, which conforms to the original immunosurveillance theory, tumor cells are targeted by innate and, subsequently, adaptive immune cells including tumor antigen-specific CD4⁺ and CD8⁺ T cells. If the immune system is successful in destroying all tumor cells, progression to subsequent phases is prevented. (2) In the equilibrium phase, that may last for years, tumor cells that have survived the initial elimination process and the ongoing immune response are in balance. Although tumor growth is still under immunological control, the immune system fails in eliminating all tumor cells, and thus causes selection pressure on the surviving variants. (3) In the escape phase, tumor cells that have undergone extensive immunoediting, evade immunological control, and expand rapidly, resulting in development of clinically apparent disease (68).

More recently, immunological processes underlying the three phases of immunoediting have been characterized in greater detail, which has led to development of immunotherapies that efficiently enhance anti-tumor immune responses (82, 83). Modern technologies have enabled identification of tumorspecific, MHC-I and -II restricted neoantigens and detection of CD8⁺ and CD4⁺ T cells responding to such neoantigens (82, 83). These technologies include deep-sequencing approaches to determine the "mutanome," that is the entirety of tumorspecific mutations, followed by *in-silico* prediction algorithms to identify mutation-specific epitopes capable of binding to MHC-I or -II molecules (84-88). In a second step, immunogenicity of identified epitopes is verified through T cell restimulation assays of peripheral blood mononuclear cells from the sequenced patient (84, 86-89) or MHC-I (85, 90) and -II (91, 92) tetramers or multimers detecting tumor-neoantigen-specific CD8⁺ and CD4⁺ T cells, respectively. Finally, multimer-selected neoepitope-specific CD8⁺ and CD4⁺ T cells can be phenotyped by flow-cytometry or (single-cell) RNA-sequencing approaches (90-92). Whereas, initial studies were focused on the role of CD8⁺ T cells in mediating anti-tumor immunity (85, 87, 93), subsequent work established that the immunogenic mutanomeagainst former expectations- predominantly induced a CD4⁺ T cell response in mice and humans (86, 89-91, 94). Tumor neoepitope-reactive CD4⁺ T cells were crucially involved in generation of potent anti-tumor CD8⁺ T cell responses (92, 95). This T cell help is mainly mediated by interactions of CD40 ligand (CD40L), which is expressed on the surface of activated CD4⁺ T cells, and CD40 on the surface of APCs (96). Additionally, CD4⁺ T cells may exert direct anti-neoplastic activity through production of pro-inflammatory cytokines or execution of cytotoxic signals on tumor cells and aid in B cell mediated humoral anti-tumor responses through CD40L signaling (97).

Neoepitope-specific CD4⁺ T cells of the T_H1 subtype are involved in anti-tumor responses (92). Adoptive transfer of neoepitope-specific CD4⁺ T_H1 cells led to tumor regression in a patient with metastatic cholangiocarcinoma (98). In line with this, high levels of circulating tumor-antigen-specific T_H1 CD4⁺ T cells and low levels of CD4⁺ cells co-expressing the immune-checkpoints programmed cell death protein 1 (PD-1) and T cell immunoglobulin and mucin-domain containing-3 (Tim-3) predict better survival in lung cancer patients (99). In contrast, high levels of tumor-infiltrating T_{regs} , which can be found in various cancer types, are associated with poor prognosis (100, 101). Animal studies identified tumor-induced conversion of CD4⁺ non-T_{regs} into T_{regs} as an important mechanism of immune escape (102) and, accordingly, circulating tumor-antigen-specific T_{regs} can be detected in cancer patients but not in healthy individuals (103). Other T helper cell subsets, such as T_H2 and T_H17 cells, can also be found in the tumor microenvironment, but their specific role in tumor immunity and prognostic importance are still under debate (104, 105).

In conclusion, $CD4^+$ T cells responding to tumorspecific neoepitopes play an important role in mediating anti-tumor immune responses. Yet, tumor cells may engage various escape mechanisms to acquire resistance to this response, which include induction of $CD4^+$ T cell phenotype switching from proinflammatory anti-neoplastic T_H1 cells into immunosuppressive and thus tumor growth-promoting T_{regs} (105).

EFFECTS OF CLINICALLY APPROVED CANCER IMMUNOTHERAPIES ON ATHEROGENESIS

Several immunotherapeutic strategies aim at preserving or restoring anti-tumor immune responses. Yet, the opposing roles of adaptive immunity in atherosclerosis and cancer development (**Figure 1**) implicate that such therapeutic approaches might involve proatherogenic side effects (17), especially if they are not antigen-specific but affect the immune system as a whole. In contrast, B cell depleting antibodies and antibodies targeting growth factor receptors overexpressed by tumor cells may confer atheroprotection. In the following section we will discuss effects of clinically approved cancer immunotherapies on atherogenesis and delineate their mechanistic background (an overview of these effects is given in **Table 1**).

Immune Checkpoint Inhibitors (ICIs)

Immune checkpoints refer to a variety of regulatory pathways that exert inhibitory actions on adaptive immune cells and beyond and are thus critical for preservation self-tolerance and prevention of exaggerated immune responses (130). The Nobel prize winning discoveries of James P. Allison and Tasuku Honjo, who unraveled that tumor cells may engage immune-checkpoint pathways to escape from anti-tumor immune responses, have paved the way for the development of monoclonal antibodies against these molecules—immune checkpoint inhibitors (ICIs) (131, 132). Ipilimumab inhibits the cytotoxic T lymphocyte antigen 4 (CTLA-4) and was shown to improve overall survival in patients with metastatic melanoma (133), which made it the first ICI approved by the Food and Drug Administration (FDA) in 2011 (134). Subsequently, four antibodies (pembrolizumab, nivolumab, cemiplimab, and dostarlimab) targeting the coinhibitory programmed cell death protein 1 (PD-1) and three antibodies (atezolizumab, durvalumab, and avelumab) directed against the programmed cell death ligand 1 (PD-L1) were demonstrated to effectively improve survival in several malignancies (134, 135) which led to the FDA-approval for treatment of 19 different cancer types and two tissueagnostic conditions [that is a tumor with a specific genetic alteration regardless of the cancer type and location (136)]. ICIs have become a cornerstone of modern cancer therapy and nowadays more than 40% of cancer patients are eligible for ICI treatment (137).

Given that immune checkpoints represent important regulators of physiological immune responses, ICI therapy can naturally involve inflammatory side effects, which are referred to as immune-related adverse events (IRAEs) (138). Although the precise pathomechanisms of such IRAEs are not yet fully clear, unconstrained activation of autoreactive T cells is suggested to play a dominant role (138). CTLA-4 and PD-1 are co-inhibitory molecules expressed on the cell surface of CD4⁺ and CD8⁺ T cells (139, 140). When bound by their ligands-CD80/CD86 and PD-L1/PD-L2-CTLA-4- and PD-1 suppress activation of T cells (140). As mentioned above, T cell activation requires simultaneous engagement of the TCR by its cognate antigen and proper costimulatory signals (32). Activation of CD28, the prototype co-stimulatory molecule, by its ligands CD80 or CD86 induces high T cell surface expression of the co-inhibitory molecule CTLA-4 (141). CTLA-4 binds CD80/CD86 with much higher affinity than CD28. However, in contrast to CD28, CTLA-4 does not exert stimulatory but inhibitory signals and thus attenuates T cell activation (141). Given that CD80/CD86 are expressed on the surface of APCs, CTLA-4 inhibits T cell activation mainly in the priming phase. Prolonged TCR stimulation during an ongoing immune response induces PD-1 expression on the cell surface of T cells (141). When bound by its ligands PD-L1 or PD-L2, which can be expressed by tumor cells, PD-1 attenuates TCR-signaling and thus reduces T cell proliferation and cytokine production. Thus, PD-1 mediates T cell inhibition in the effector phase and is used as a marker of T cell exhaustion (141). Consequently, antibodymediated inhibition of CTLA-4, PD-1 and PD-L1 enhances T cell activation. IRAEs can affect almost every organ and mostly occur within 2-16 weeks after treatment initiation (138, 142). According to a recent meta-analysis including 36 phase II and III randomized controlled trials (RCTs), the pooled incidence of all IRAEs ranges between 54 and 76% (143). Whereas, the incidences of specific IRAEs depend on the ICI used and several other factors, the integumentary, gastrointestinal, endocrine, hepatic, and pulmonary systems are overall most commonly affected (143, 144). In a meta-analysis of 112 trials including 19,217 patients, IRAE-associated fatality rates ranged between 0.36% for anti-PD-1 mono-therapy and 1.23% for PD1/PD-L1

TABLE 1 | Pro-atherogenic and athero-protective effects of current cancer immunotherapies.

Type of immunotherapy	Specific approach/substance	Compounds	Effect on atherosclerosis in clinical trials	Effect on atherosclerosis in animal studies	Potential mechanisms
Immune checkpoint inhibitors	Anti-CTLA4-Abs	lpilimumab	↑ (17, 106)	↑ (107–111)	 Increased plaque-infiltration by CD4⁺ and CD8⁺ T cells Higher expression of proinflammatory cytokines (IFN-γ and TNF-α) by T cells Enhanced T cell activation
	Anti-PD1-Abs	Pembrolizumab, Nivolumab, Cemiplimab, Dostarlimab			
	Anti-PD-L1-Abs	Atezolizumab, Durvalumab, Avelumab			
Monoclonal antibodies	Anti-CD20-Abs	Rituximab, Obinutuzumab, Ofatumumab	↓ (112–114)	↓ (56, 57)	- Depletion of mature B cells and reduction of anti-oxLDL IgG antibodies
	VEGF inhibitors	Bevacizumab, Ramucirumab	↑ (115–117)	↑ (118)	 Induction of an inflammatory endothelial cell phenotype and impairment of endothelial function Reduction and functional impairment of T_{regs} and induction of proinflammatory T_H1 cells
	EGFR targeting Abs	Cetuximab, Necitumumab, Panitumumab	-	↓ (119–121)	 Reduced accumulation of macrophages in plaques Reduced lipid uptake by macrophages and reduced foam cell formation Reduced CD4⁺ T cell activation, proliferation and plaque infiltration Reduced pro-inflammatory cytokine production Reduced SMC proliferation
Cytokines	IFN-α	Interferon alfa	↑ (122, 123)*	↑ (124)	 Increased plasma cholesterol and triglyceride levels Induction of lipid uptake by macrophages and increased foam cell formation Inhibition of T_{reg} activation and proliferation Direct stimulation of cytotoxic CD4⁺ T cell function Sensitization of antigen-presenting cells toward pathogen-derived TLR4 ligands
	IL-2	Aldesleukin	High-dose: ↑ (125) Low-dose: ?**	High-dose: ↑ (126) Low-dose***: ↓ (127)	 High-dose: unspecific expansion of T cells Low-dose: selective expansion of functional T_{regs}
Antifolate	DHFR inhibition	Methotrexate	- (128)	↓ (129)****	 Attenuation of monocyte maturation and recruitment Modulation of lipoprotein transcellular transport Reduction of pro-inflammatory cytokine production

 \uparrow = proatherogenic effect. \downarrow = atheroprotective effect. *These studies demonstrated an association between plasma IFN-α levels and atherosclerosis. A direct effect of IFN-α administration on atherosclerosis has not yet been shown in clinical trials. **Currently investigated in the "Low-dose interleukin-2 in patients with stable ischemic heart disease and acute coronary syndromes (LILACS)" trial. ***and complexed with a specific anti-IL2-mAB (JES6-1A12).****delivered via nanoparticles.

Abs, monoclonal antibodies; CTLA4, cytotoxic T lymphocyte antigen 4; DHFR, dihydrofolate reductase; EGFR, epidermal growth factor receptor; IFN-α, interferon alpha; IFN-γ, interferon gamma; IL2, interfeukin 2; mAB, monoclonal antibody; oxLDL, oxidized low density lipoprotein; PD-1, programmed cell death protein 1; PD-L1, programmed cell death ligand 1; SMC, smooth muscle cell; TLR4, toll-like-receptor 4; TNF-α, tumor necrosis factor alpha; VEGF, vascular endothelial growth factor.

plus CTLA-4 combinational therapy and were most commonly caused by colitis, pneumonitis, hepatitis, myocarditis and neurotoxic effects (145). Cardiovascular IRAEs, which include myocarditis, pericardial diseases, heart failure, dyslipidemia, myocardial infarction, and cerebral arterial ischemia, are, overall, relatively rare with an incidence ranging between \sim 3 and 20 per 1,000 patients (146). Yet, cardiovascular toxicities are severe in over 80% of cases (147) and myocarditis, which carries the

highest fatality risk of all IRAEs (40–50%), is of particular prognostic relevance (145, 147).

Besides acutely occurring cardiovascular IRAEs, recent evidence has suggested that ICI therapy may promote atherogenesis (148, 149). In a retrospective analysis of 1,215 patients treated with ICIs, atherosclerotic cardiovascular events (CVE) occurred in 1% within a follow-up period of 6 months (150). In three meta-analyses, the ICI-related incidence of myocardial infarction and stroke ranged from 0.4 to 1.0% and 1.1 to 2.0%, respectively (149). Yet, the majority of studies included in these meta-analyses were not specifically designed to assess CVE and may thus underestimate incidences (149). To evaluate the ICI-related risk of atherosclerotic CVE (defined as the composite of myocardial infarction, coronary revascularization, and ischemic stroke), Drobni et al. analyzed event-rates in 2,842 patients treated with ICIs and matched controls (17). Additionally, a case-crossover analysis was performed, in which event rates within the 2 years before (control period) and the 2 years after (at-risk period) initiation of ICI therapy were compared. ICI therapy was associated with a 3-fold and almost 5-fold higher risk of atherosclerotic CVE in the matched-control study and case-crossover analysis, respectively (17). In a nested imaging substudy including 40 patients, a 3-fold increase in aortic atherosclerotic plaque volume progression from 2.1%/year before to 6.7%/year after ICI initiation could be detected (17). Another recent study retrospectively analyzed 2-[18F]fluorodeoxyglucose (FDG) positron emission tomography/computed tomography scans, which had been performed in 20 melanoma patients before and during ICI treatment (mean time interval: 4.4 months) (106). A significantly increased FDG uptake in large arteries after ICI treatment initiation could be detected, pointing toward an ICI-related induction of arterial inflammation (106). In accordance to these clinical findings, a series of animal studies reported enhanced plaque inflammation and accelerated atherogenesis in $LDLr^{-/-}$ mice genetically deficient for or treated with inhibitory antibodies against PD-1, PD-L1 and CTLA-4 (148, 149). This was accompanied by an increased number of plaque-infiltrating CD4⁺ and CD8⁺ T cells (107-111), higher expression of proinflammatory cytokines [IFN- γ and tumor necrosis factor alpha (TNF- α)] by T cells (107, 111), and enhanced T cell activation (108, 110, 111).

Collectively these data emphasize that ICI therapy promotes atherogenesis and substantially increases the risk of atherosclerotic CVE. Presumably, ICIs exert their proatherogenic effects—at least in part—through disinhibition of T cells responding to plaque-associated autoantigens. Atherosclerosis is a slowly progressing disease and all above-mentioned clinical studies were limited by relatively short follow-up periods. As indications for ICI therapy are rapidly expanding and cancer-survival has dramatically improved in recent years, the detrimental impact of ICIs on atherogenesis will, therefore, likely become a more relevant health issue in the future.

Antibody Therapy

Since the FDA approval of muromonab-CD3, a monoclonal antibody targeting the T cell co-receptor CD3, for the prevention of transplant rejection in 1986, more than 100 therapeutic

antibodies have been included in clinical practice (135). Rituximab, a monoclonal antibody targeting the B cell receptor CD20, was approved for treatment of follicular lymphoma in 1997, which opened the door for the use of antibodies in cancer therapy (151). Cancer has emerged as the most common condition for antibody therapy with currently over 40 FDA-approved antibodies (including ICIs) for treatment of several cancer types (135). Antibodies can target cancer through several mechanisms, including direct tumor cell killing, immunemediated tumor cell-killing, and inhibition of neovascularization or stroma cells (152, 153). Direct tumor cell killing can be achieved through eliciting agonistic activity to apoptosispromoting receptors, inhibiting growth factor receptor signaling, neutralizing key enzymes, or delivering cytotoxic agents into the cell (152, 153). Mechanisms of immune-mediated tumor cell killing include induction of phagocytosis, complementactivation and cellular toxicity (152, 153). Besides ICIs, the use of several other monoclonal antibodies in cancer therapy is constrained by their cardiovascular side effects, which include myocarditis, heart failure, arrhythmia, orthostatic dysregulation and atherosclerotic cardiovascular events (16). Fortunately, the latter complication is rare and some antibodies can even confer atheroprotective effects.

Antibody-Mediated B Cell Depletion

During the past two decades, B cell depleting strategies have been used for treatment of B cell lymphoma and several autoimmune diseases, including rheumatoid arthritis, systemic lupus erythematosus (SLE) and multiple sclerosis (MS) (151, 154). In addition to rituximab and other antibodies targeting CD20 (e.g., obinutuzumab and ofatumumab), antibodies directed against the B cell surface proteins CD19 (blinatumomab), CD22 (inotuzumab ozogamicin and moxetumomab pasudotox), CD38 (daratumumab and isatuximab) and CD319 (elotuzumab) have been approved to treat these conditions (153).

As mentioned above, depletion of B cells through administration of a CD20-specific antibody ameliorated atherogenesis in Apoe^{-/-} and $Ldlr^{-/-}$ mice (56, 57). In line with this, treatment of $Apoe^{-/-}$ mice with a monoclonal antibody targeting B cell activating factor-receptor (BAFFR) to selectively deplete mature B2 cells while sparing B1 cells conferred atheroprotection (155). Similarly, antibody-mediated inhibition of the cytokine B cell-activating factor (BAFF) reduced atherosclerosis in $Apoe^{-/-}$ and $Ldlr^{-/-}$ mice (156). These findings were recently confirmed in a clinical study: Patients who received rituximab therapy after kidney transplantation had a significantly lower rate of atherosclerotic CVE during 8 years of follow-up as compared to propensity-matched controls (112). Accordingly, rituximab therapy was shown to reduce carotid intima media thickness (113) and to improve flow mediated dilation of the brachial artery, a non-invasive marker of endothelial function (114). The effect of other B-cell-depleting antibodies (including those targeting receptors predominantly expressed on antibody-secreting plasma cells, such as CD39 and CD319) on atherosclerosis has not yet been investigated and the role of plasma cells in atherogenesis is not yet clear (157). Depletion of IgG-producing plasma cells reduced atherosclerotic plaque development in $Apoe^{-/-}$ and $Ldlr^{-/-}$ mice (158, 159) but associated with plaque instability, which may have deleterious consequences in patients with preexisting atherosclerosis (158). Evidence from preclinical and clinical studies indicate that antibodies targeting CD20 may confer atheroprotection, but these early findings will have to be confirmed in larger clinical trials.

Antibodies Targeting Vascular Endothelial Growth Factor

Bevacizumab was the first clinically approved monoclonal antibody targeting vascular endothelial growth factor (VEGF) (160). Originally thought to exert antineoplastic actions exclusively via inhibition of tumor angiogenesis, VEGF-targeted therapies have been demonstrated to arrest tumor growth through a variety of mechanisms, which are not yet fully understood (160). In addition to bevacizumab, an antibody targeting VEGF receptor 2 (ramucirumab) and small-molecules inhibiting VEGF receptor tyrosine kinases (sorafenib and sunitinib) have been FDA-approved (153). A major drawback of VEGF inhibitors is their tendency to induce atherosclerotic CVE. Recent meta-analyses including up to 22 studies reported a \approx 1.4- to 2.5-fold higher risk of arterial ischemia in patients treated with bevacizumab (115-117). High-dose bevacizumab therapy was even associated with a 4.4- and 6.7-fold higher risk of cardiac and cerebral ischemia, respectively (115). A preclinical study confirmed and mechanistically substantiated these findings by demonstrating that administration of a VEGF-targeting antibody impaired endothelial function and increased atherosclerotic lesions by 33% in Apoe^{-/-} mice (118). Accordingly, VEGF inhibitors were shown to induce an inflammatory phenotype in cultured human coronary artery endothelial cells (161). Besides affecting endothelial function, VEGF inhibitors may decrease the number of T_{regs} and impair their suppressive capacity, reduce expression of co-inhibitory T cell molecules, and thus induce proinflammatory T_H1 responses (162). Although experimental proof is missing, these immunological effects might contribute to the proatherogenic properties of VEGF-inhibiting antibodies (162).

Antibodies Directed Against Epidermal Growth Factor Receptors

Receptors of the epidermal growth factor receptor family, such as epidermal growth factor receptor (EGFR) or human epidermal growth factor receptor 2 (HER2/neu), may be overexpressed by tumor cells of several cancer types which can thus acquire the capability of autonomous and uncontrolled proliferation (163, 164). Overexpression of EGFR or HER2/neu is a strong predictor of a negative prognosis in a variety of malignancies (165, 166) and the development of monoclonal antibodies targeting such receptors has advanced cancer treatment. Early clinical studies and large-scale phase 3 trials showed improved outcome in patients with metastatic breast cancer and gastric cancer treated with trastuzumab (targeting HER2/neu) and patients with metastatic colorectal cancer and head and neck cancer treated with cetuximab (directed against EGFR) (167–170). Further HER2/neu and EGFR targeting antibodies have been clinically approved (153). A major drawback of growth factor receptor targeting antibodies (especially trastuzumab) is their potential to induce heart failure, which occurs in up to 20% of all cases (167, 171) and is 1.7 to 4 times more frequently compared to standard chemotherapy (172–174). Accordingly, mice lacking Her2/neu were demonstrated to develop dilated cardiomyopathy (175).

Direct effects of antibodies targeting growth factor receptors on atherogenesis have not yet been reported in clinical trials. Nevertheless, EGFR was detected in human atherosclerotic plaques (176) and increased HER2/neu plasma levels were shown to be associated with a higher risk of CAD (177). In line with this, evidence from preclinical studies indicated that inhibition of growth factor signaling may confer atheroprotection (119-121). In two elegant studies, Zeboudj, Ait-Oufella and colleagues demonstrated that cell-specific depletion of EGFR either in myeloid cells (119) or in $CD4^+$ T cells (120) protected $Ldlr^{-/-}$ mice from atherosclerosis. EGFR deficiency in myeloid cells limited macrophage accumulation within plaques and lipid uptake by macrophages, whereas CD4⁺ T cell-specific depletion of EGFR reduced CD4⁺ T cell activation, proliferation and infiltration in atherosclerotic lesions. Both cell-specific EGFR deletions were accompanied by reduced pro-inflammatory cytokine production (119, 120). Despite these promising findings, some uncertainties regarding the mechanistic implication of EGFR and its ligands in atherogenesis remain (178), beyond EGFR's profound immunomodulatory role systems-wide must be taken into account (179). Whether atheroprotective effects of growth factor receptor targeting antibodies also apply to humans is still unclear.

Cytokine Therapy

A variety of cytokines may exert significant anti-neoplastic effects either by directly inhibiting proliferation and inducing apoptosis of tumor cells or by stimulating anti-tumor immune responses (180, 181). Despite promising findings in early preclinical studies, utilization of cytokines as cancer therapeutics was later demonstrated to involve several limitations which hindered broad translation of this treatment approach into clinical practice (180, 181). Nevertheless, IFN- α and IL-2 were clinically approved for the treatment of different malignancies such as hairy cell leukemia, follicular non-Hodgkin lymphoma, melanoma, and Kaposi's sarcoma (IFN- α) or renal cell carcinoma and melanoma (IL-2) (180, 181).

Interferon Alpha (IFN-α)

Clinical application of IFN- α is particularly limited by its proatherogenic properties (182, 183). $Ldlr^{-/-}$ mice treated with IFN- α had accelerated atherosclerosis and increased plasma cholesterol and triglyceride levels (124). Several other proatherogenic effects of IFN- α have been reported, such as induction of lipid uptake by macrophages and foam cell formation (184, 185), inhibition of T_{reg} activation and proliferation (186, 187), direct stimulation of antigen-presenting cells toward pathogen-derived toll-like receptor 4 (TLR4) ligands (189). Clinical studies demonstrated that plasma type I IFN (IFN- α and - β) levels are associated with atherosclerosis development

in patients with SLE (122) and human immunodeficiency virus-1 (HIV-1) infection (123). Experimental evidence suggested that IFN- α directly promotes atherogenesis by impairing vascular repair (190, 191) or inducing endothelial dysfunction (192) and may thus causally contribute to the highly increased risk of atherosclerotic CVE in SLE patients, which is not adequately explained by traditional risk factors (193). For a thorough review on the impact of IFN- α on different atherosclerosis-associated cell types and clinical implications the interested reader is referred to Chen et al. (182).

Interleukin 2 (IL-2)

IL-2 was originally termed T cell growth factor as it was first identified as a component of T cell culture fluids that induced proliferation of antigen-activated T cells (194, 195). It was thought to act as a crucial mediator in T cell immune responses and to play an important role in host response and tumor control, which led to test high-dose IL-2 as a novel cancer treatment in the mid 1980s (196). Although limited by toxicities such as capillary leak syndrome, fever, chills, malaise and arthralgias, this approach facilitated significant tumor regression and emerged as the first effective immunotherapy for human cancer (196). Yet, IL-2 deficient mice developed severe lymphoproliferation and autoimmunity which pointed toward an additional important role of the cytokine in maintaining self-tolerance (197, 198). Subsequent studies revealed that T_{reg} generation is dependent on IL-2 (195). Tregs express increased levels of the highaffinity IL-2 receptor alpha chain (IL-2R α , also known as CD25) compared to effector T cells (Teff cells) and are thus more sensitive for IL-2 (199). Accordingly, daily low-dose IL-2 therapy stimulated selective expansion of functional Tregs through increased proliferation, thymic export and resistance to apoptosis (while only minimally affecting conventional CD4⁺ T cells) and thus led to a substantial clinical improvement in patients with active chronic graft-vs.-host disease (200, 201).

The specific role of IL-2 in atherogenesis has not yet been fully clarified. Increased IL-2 serum levels were shown to be associated with carotid artery intima-media thickness (202), a sonographic marker of atherosclerosis, and CAD (203). An early clinical study reported atherosclerotic CVE in 3.8% (angina or ischemic changes in 2.6% and myocardial infarction in 1.2%) of patients who received IL-2 for cancer therapy (125). Accordingly, IL-2 administration $(2 \times 10^4 \text{ units twice weekly})$ for a period of 6 weeks) accelerated atherogenesis in $Apoe^{-/-}$ mice, whereas administration of an antibody targeting IL-2 was atheroprotective (126). Treatment of $Ldlr^{-/-}$ (204) and $Apoe^{-/-}$ mice (127) with low-dose IL-2 complexed with a specific IL-2 monoclonal antibody (JES6-1A12) conferred significant atheroprotection by inducing a substantial expansion of T_{regs} in atherosclerotic lesions and several other tissues. Neither IL-2 nor the anti-IL2 antibody alone affected atherogenesis (127) and the observed anti-atherogenic efficacy depended on the antibody clone used. Administration of IL-2 complexed with another IL-2 antibody clone (SAB6) induced expansion of natural killer (NK) and CD8⁺ T cells (205). A subsequent study unraveled the mechanism by which the two different antibody complexes selectively induce expansion of T_{regs} or T_{eff} cells: (1) JES6-1

sterically blocks the interaction of IL-2 with IL-2RB and IL- $2R\gamma$ and allosterically disrupts binding of IL2 to IL- $2R\alpha$, thereby favoring activation of T_{regs} with high IL-2R α expression; (2) S4B6 sterically hinders IL-2/IL-2Rα interaction and enhances IL-2/IL-2Rβ interaction, thus stimulating all IL-2-responsive T cells (206). More recently, a human anti-IL2-antibody (F5111.2) was developed that selectively promotes Treg expansion when complexed with human IL-2 by inducing similar conformational changes (207). Administration of IL2-F5111.2 complexes yielded substantial therapeutic efficacy in humanized animal models of different autoimmune diseases, such as type 1 diabetes, autoimmune encephalomyelitis or xenogeneic graft-vs.-host disease (207). Whether such approach might be translatable into clinical practice has not yet been determined. The randomized, double-blind, placebo-controlled LILACS trial (NCT03113773) examined whether solely administering low-dose IL-2 is safe and effective in patients with stable ischemic heart disease and acute coronary syndrome (208). The study has been completed and its results are awaited for publication.

In conclusion, current evidence suggests that high-dose IL-2 therapy promotes atherogenesis through induction of proinflammatory T_{eff} cell responses, whereas administration of low-dose IL-2 might confer atheroprotection by selectively stimulating expansion of T_{regs} . Complexing IL-2 with specific anti-IL-2 antibodies might even enhance the latter effect through augmenting the selectivity to IL-2R α , which is highly expressed on T_{regs} .

Methotrexate

Methotrexate (MTX) is a structural analog of folic acid (Vitamin B9) that inhibits enzymes involved in folate metabolism, such as dihydrofolate reductase (DHFR), and thus limits cellular division (209). DHFR catalyzes conversion of dihydrofolate to tetrahydrofolate, which acts as an important coenzyme in synthesis of pyrimidine and purine. In 1948 Farber et al. firstly reported that treatment with the folate analog aminopterin enabled temporary remission in childhood leukemia (210). Subsequently, MTX (initially termed amethopterin) was found to have better pharmacological properties than aminopterin (211) and thus emerged as one of the most extensively used chemotherapy agents for a variety of cancer types (209, 212). Besides anti-neoplastic properties, MTX exerts potent antiinflammatory actions (213). Several studies reported efficacy of low-dose MTX in the treatment of rheumatoid arthritis (RA) (214, 215). The compound has become a mainstay in therapy of RA and other autoimmune diseases, although its immunosuppressive mechanisms of action have not yet been fully clarified (213). Observational data revealed that lowdose MTX therapy associated with a lower risk for CVD and cardiovascular mortality (216, 217). The Cardiovascular Inflammation Reduction Trial (CIRT) included 4,786 patients with CAD and additional metabolic risk factors (type 2 diabetes or metabolic syndrome) randomly assigned to receive lowdose MTX or placebo (128). After a median follow-up of 2.3 years, MTX neither reduced pro-inflammatory biomarkers [IL-1β, IL-6, and C-reactive protein (CRP)] nor CVE. MTX was associated with modest elevations in liver enzymes, reductions

in leukocyte counts and hematocrit levels, and an increased incidence of non-basal-cell skin cancers. Although the study had some methodical limitations (e.g., patients were not screened for an increased inflammatory risk), the data overall discouraged further investigations on MTX therapy for CVD prevention. In a recent murine study, nanoparticle-formulated MTX conferred substantial atheroprotection through modulating lipoprotein transcellular transport, reducing expression of proinflammatory cytokines and attenuating monocyte maturation and recruitment (129).

Although several immunotherapeutic approaches have yielded promising results in preclinical CVD models, only few of these strategies have proven beneficial in clinical studies (8, 218). Canakinumab (14), a monoclonal antibody targeting IL-1 β , and colchicine (219, 220), an ancient drug traditionally used for gout therapy which exerts anti-inflammatory effects (among other potential mechanisms) through inhibition of the NOD-, LRR-, and pyrin domain-containing protein 3 (NLRP3) inflammasome in leukocytes (221), were demonstrated to reduce cardiovascular events in large-scale clinical trials. Although not targeting inflammatory pathways, monoclonal antibodies [evolocumab (13) and alirocumab (12)] or a siRNA-based compound [inclisiran (222, 223)] targeting PCSK9 to lower LDL cholesterol levels have proven clinically beneficial.

Considering that a pro-inflammatory immune response is critically involved in early elimination of mutated cells (68), immunotherapies exerting systemic anti-inflammatory effects may mitigate anti-tumor immunity and thus increase the risk of cancer development. In the following section we discuss recent evidence on the impact of above-mentioned immunotherapeutic strategies for CVD on cancer incidence (an overview is given in **Table 2**).

Canakinumab

Although canakinumab reduced CVE in patients with CAD, several limitations, such as a significant impairment of host defense and high costs, prevented its clinical approval for secondary prevention of CVD (14). Recent evidence suggests that IL-1 β has a dual role in cancer development: On the one hand, it initiated pro-inflammatory anti-tumor immune responses by activating tumor antigen-specific T_H1 and T_H17 cells and facilitated tumor regression (233). On the other hand, IL-1ß can promote carcinogenesis by inducing chronic inflammation, endothelial cell activation, angiogenesis, or development of immunosuppressive cells, such as tumorassociated macrophages and myeloid-derived suppressor cells (233). In contrast to anti-tumor immunity, which is critically involved in cancer elimination, chronic inflammation may drive tumorigenesis through several mechanisms including inhibition of antineoplastic immune responses, modulation of the tumor microenvironment to become more tumorpermissive, and direct tumor-promoting actions on epithelial and cancer cells (234). In line with this, IL-1\beta-deficient mice transplanted with melanoma cells were protected from development of local tumor and metastases (225) and canakinumab significantly reduced incidence of fatal cancer in the CANTOS trial (14). An exploratory analysis further revealed that canakinumab treatment especially reduced the incidence of lung cancer and lung cancer mortality (224). These findings motivated the initiation of three randomized phase III trials, CANOPY-A (NCT03447769), CANOPY-1 (NCT03631199), and CANOPY-2 (NCT03626545) (235), and a phase II trial, CANOPY-N (NCT03968419) (236) to investigate the potential of canakinumab in the treatment of non-small cell lung cancer (NSCLC). Although most trials are still ongoing, the CANOPY-2 study, which evaluated canakinumab in a second- or third-line treatment setting (that is in patients with locally advanced or metastatic NSCLC and tumor progression after or during platinum-based chemotherapy and PD-(L)1 inhibitor therapy) failed to meet the primary endpoint of overall survival (237).

Colchicine

Colchicine is derived from the autumn crocus, which has been used for gout therapy since ancient times (238). Until today, colchicine represents a first-line drug for gout treatment (239). More recently, the compound has emerged as a promising candidate for secondary prevention of CVD. In two largescale RCTs, the COLCOT (219) and LoDoCo 2 (220) trials, collectively enrolling more than 10,000 patients with recent myocardial infarction or chronic coronary syndrome, lowdose colchicine therapy significantly reduced the incidence of CVE. In contrast to canakinumab, colchicine did not increase the risk of fatal infections, although in COLCOT non-fatal pneumonia occurred more often in colchicine-treated patients. Nevertheless, colchicine therapy did not significantly reduce cardiovascular mortality in these trials and was associated with an almost significant increase in non-cardiovascular mortality in LoDoCo2 (220). A smaller RCT, enrolling 795 patients, reported a significantly higher rate of all-cause mortality (mainly due to non-cardiovascular deaths) in patients with acute coronary syndrome when colchicine was added to standard therapy (240). Recent meta-analyses confirmed that low-dose colchicine therapy in patients with CAD was associated with a significantly reduced risk of CVE (myocardial infarction, stroke, and the need for coronary revascularization) and a non-significant reduction of cardiovascular deaths, which was counterbalanced by a non-significant increase in noncardiovascular deaths (241, 242). Considering the robust reduction of CVE observed in these studies as well as the low price and wide availability of the compound, low-dose colchicine might become an option for secondary prevention in high-risk CVD patients, but the increased non-cardiovascular death rates need further investigation.

Despite the increase in non-cardiovascular mortality, colchicine was not associated with higher rates of cancer in any of the above-mentioned studies (219, 220, 240). Preclinical evidence showed that colchicine may reduce tumor growth of several cancer types, such as prostate (229), hypopharyngeal (230) and liver cancer (231) as well as tumor implantation of pressure-activated colon carcinoma cells (232). The incidence of all-cause cancers was significantly reduced in 13,679 male gout patients (at least temporarily) treated with colchicine compared with 10,371 control gout patients (228). Besides reducing tumor cell proliferation and inducing apoptosis (243),

TABLE 2 Effects of anti-atherosclerotic immunothe	erapies (with clinically proven efficacy)	on cancer.
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Type of immunotherapy	Target	Compounds	Effect on cancer in clinical trials	Effect on cancer in animal studies	Potential mechanisms
Monoclonal antibodies	IL-1β	Canakinumab	↓ (224)	↓ (225)*	- Reduction of tumor-promoting chronic inflammation
	PCSK9	Alirocumab	-	↓ (226)	 Increased MHC I expression on tumor cell surface and enhanced tumor infiltration by cytotoxic T cells
		Evolocumab			
	CD3	Teplizumab Foralumab	-	↓ (227)	- Induction of leukemic cell apoptosis
Natural anti-tubulin agent	NLRP3 inflammasome**	Colchicine	↓ (228)	↓ (229–232)	 Direct antiproliferative effects on tumo cells and induction of apoptosis Enhancement of CD4⁺ and CD8⁺ T-cell-mediated anti-tumor immunity

 \uparrow = pro-carcinogenic effect. \downarrow = anti-carcinogenic effect. * In this study IL-1 β knockout mice were shown to be protected from tumor development, but antibody-mediated IL-1 β depletion was not investigated. **besides other anti-inflammatory mechanisms.

IL-1β, interleukin 1 beta; NLRP3, NOD-, LRR-, and pyrin domain-containing protein 3; PCSK9, proprotein convertase subtilisin/kexin 9.

colchicine enhances CD4⁺ and CD8⁺ T-cell-mediated antitumor immunity by promoting dendritic cell maturation and antigen presentation (244). Colchicine toxicity at high doses prevents its application as anti-tumor drug (243). Whether low-dose colchicine might confer clinically relevant anti-cancer effects has yet to be determined.

PCSK9 Inhibitors

PCSK9, a protein which is primarily expressed in the liver, counteracts clearance of LDL cholesterol by inducing degradation of internalized LDL receptor in hepatocytes leading to increased plasma LDL cholesterol levels (245). Inhibition of PCSK9 has emerged as a highly effective second-line cholesterol lowering strategy, which has received class I recommendations by current guidelines (246). Two monoclonal antibodies targeting PCSK9, evolocumab (13) and alirocumab (12), and a siRNA-based compound, inclisiran (222, 223), are approved for clinical use in selected high-risk patients.

Cholesterol is a driving force in atherogenesis, yet its role in cancer is less clear: Both positive and negative correlations as well as absence of any associations between cholesterol levels and cancer development have been reported by clinical studies (247). Likewise, several meta-analyses concluded that statin therapy reduces cancer incidence or improves prognosis (248-250), whereas others found no such correlations (251, 252). PCSK9 inhibitors have not been reported to affect cancer incidence in clinical trials (12, 13). However, in a recent study PCSK9 inhibition could potentiate anti-tumor immune responses and thus substantially reduced tumor growth in murine cancer models (226), which was independent of cholesterol-lowering. PCSK9 induced lysosomal degradation of MHC-I and disrupted its recycling to the cell surface. PCSK9 inhibition, either through genetic deletion or administration of monoclonal antibodies, increased MHC-I expression on the tumor cell surface and thus enhanced tumor infiltration by cytotoxic T cells. Besides limiting tumor growth when administered alone, anti-PCSK9 antibodies significantly enhanced anti-tumor efficacy of ICI therapy (anti-PD-1) (226). Another recent study demonstrated that a nanoliposomal anti-PCSK9 vaccine limited tumor progression and improved survival in a murine model of colon carcinoma (253). Concluding, these preclinical data warrant further exploration of PCSK9 inhibitors as cancer therapeutics in clinical trials.

Anti-CD3 Antibody Therapy

Anti-CD3 monoclonal antibodies bind to the CD3-TCR complex on the surface of T cells and thus induce disappearance by shedding or internalization of the receptor complex (254). This process, which is termed antigenic modulation, renders T cells temporarily blind to their cognate antigen and leads to anergy or apoptosis of activated T cells (254). Anti-CD3 antibody therapy induced long-lasting T_{reg}-mediated immune tolerance through increased TGF-β production by apoptotic T cells and phagocytes involved in clearance of apoptotic T cells (255, 256). Clinical application of the murine muromonab-CD3 is limited by high immunogenicity and resulting side effects (e.g., nausea, fever, headaches) (254). Humanized anti-CD3 antibodies (e.g., teplizumab, and otelixizumab) and a fully human anti-CD3 antibody (foralumab) have been developed, that were well-tolerated in initial clinical studies (254). Early clinical evidence suggests reasonable efficacy of such novel anti-CD3 antibodies in the treatment or prevention of autoimmune diseases, such as multiple sclerosis (257), type 1 diabetes (258-261), and inflammatory bowel disease (262). In several preclinical studies, intravenous or oral administration of anti-CD3 antibodies conferred substantial atheroprotection by enhancing TGF-B production and thereby inducing antiatherogenic T_{regs} (263-265). Muromonab-CD3 was reported to significantly decrease CD3⁺ tumor cells in a patient with refractory T cell acute lymphoblastic leukemia (266), but was ineffective in enhancing immune activation in patients with solid tumors when administered in combination with highor low-dose IL-2 (267, 268). However, a recent preclinical study demonstrated high anti-tumor efficacy of teplizumab and foralumab in murine models of T cell acute lymphoblastic leukemia (227). To date, the potential of humanized or fully human anti-CD3 antibodies in the treatment of CVD or cancer has not yet been investigated in clinical trials. Considering the promising data from animal studies these compounds merit further investigation into their clinical application.

ADOPTIVE T CELL TRANSFER IN CANCER AND ATHEROSCLEROSIS

Chimeric antigen receptor (CAR) T cell therapy represents an innovative cancer treatment strategy, in which circulating T cells are isolated and genetically modified in vitro to express a synthetic tumor-antigen-specific receptor (269), which are subsequently expanded and infused back into the patient to attack tumor cells (269). In 2017, CAR T cells directed against CD19 (tisagenlecleucel and axicabtagene ciloleucel) showed substantial anti-tumor activity in patients with refractory large B cell lymphoma and follicular lymphoma (270, 271), which led to their FDA approval. Subsequently, anti-CD19 CAR T cells brexucabtagene autoleucel and lisocabtagene maraleucel were approved for treatment of mantle cell lymphoma and diffuse large B cell lymphoma, respectively (272, 273). Idecabtagene vicleucel targets B cell maturation antigen (BCMA) and is the first clinically approved CAR T cell therapy for multiple myeloma treatment (274). A major drawback of CAR T cell therapy is its association with severe and potentially fatal side effects (275). Cytokine-release syndrome (CRS), that can potentially develop into fulminant haemophagocytic lymphohistiocytosis (HLH), and CAR-T-cell-related encephalopathy syndrome (CRES), are the two most common adverse reactions (275). A recent study reported that CVE, such as new onset of heart failure or arrhythmias, occurred in 12% of 137 patients who received CAR T cell therapy (all events were associated with CRS) (276). Whether CAR T cells affect atherogenesis and increase the cardiovascular risk in the long-term, is unknown.

Adoptive transfer of autologous ex vivo expanded polyclonal T_{regs} has emerged as a promising strategy to treat autoimmune diseases and is currently investigated in clinical trials (277). First evidence suggested safety and efficacy of this approach for the treatment of type 1 diabetes (278, 279), prevention of graft-vs.host-disease (280, 281), or transplant rejection (282). Therapeutic potency may be enhanced by utilization of antigen-specific rather than polyclonal Tregs. Administration of ovalbumin-specific Tregs, which respond to a major component of chicken egg white, was demonstrated to be safe and effective in patients with refractory Crohn's disease (283). Adoptive transfer of both polyclonal (284) and antigen-specific Tregs (responding to heatshock protein 60) (285) conferred atheroprotection in $Apoe^{-/-}$ mice fed with a WD for 6-8 weeks. Yet, administration of ApoB⁺ T_{regs} to WD-fed Apoe^{-/-} mice failed to limit plaque progression during a more extended period of observation (12 weeks) (43). In this study, more than half of all transferred cells lost expression of T_{reg} markers and converted into conventional T cells, which likely accounted for treatment failure. Clarification of the mechanisms underlying such phenotypic conversion and development of strategies to ensure T_{reg} stability are essential for clinical translation of this approach.

VACCINATION STRATEGIES IN THE TREATMENT OF CANCER AND ATHEROSCLEROSIS

Vaccination strategies aiming to induce pro-inflammatory immune responses against tumor-specific antigens or immune tolerance to plaque-associated autoantigens hold great promise for the treatment of cancer and atherosclerosis, respectively (4, 6). Through eliciting antigen-specific immunity, such approaches are highly effective without impairing the host defense against infectious agents and cancer cells or enhancing atherogenesis.

A series of animal studies have indicated the great potential of immunization against ApoB-related antigens for the treatment of atherosclerosis (6). The underlying idea originates from the observation that administration of oxLDL to hypercholesterolemic rabbits reduced atherosclerotic lesions (286). Subsequent studies identified an expansion of ApoB⁺ T_{regs} (42, 287, 288) and a humoral immune response against LDL (289) to account for the observed atheroprotection. Despite these promising preclinical data, several unknowns, such as optimal epitopes, adjuvants, administration route and vaccination scheme, stability of the atheroprotective immune response, and criteria for patient selection, have hitherto hindered translation of anti-atherosclerotic vaccination strategies into clinical practice (6). Recently, utilization of nanoparticle-formulated, nucleosidemodified messenger RNA (mRNA) without addition of adjuvants was demonstrated to enable sustained immune tolerance to MSrelated autoantigens through induction of functional Tregs in mice (290). Considering that mRNA-based vaccines are already in clinical use, this approach has high translational potential for development of a vaccine against atherosclerosis.

Therapeutic cancer vaccines aim to induce a sustained effector immune-response against tumor-specific antigens (291). Initial approaches used self-antigens, which exist in non-malignant tissues, but are abnormally expressed or overexpressed by tumor cells, or applied non-self-antigens of viral origin (4, 291). Technological advances, such as next-generation sequencing, enabled identification of tumor-specific epitopes resulting from mutations (4, 291). These so-called neoepitopes or neoantigens are then evaluated for their capacity to bind human MHC-I or -II molecules. Candidates with high binding capacity can finally be utilized for development of personalized cancer vaccines or in vitro expansion of tumor-antigen specific CD4⁺ and CD8⁺ T cells, which are then transferred into the patient (4, 291). Vaccination with neoepitopes predicted to bind MHC-II or -I was shown to elicit strong CD4⁺ and/or CD8⁺ responses and thereby facilitate tumor rejection in animal models (292). In 2015, Carreno et al. firstly demonstrated the capability of a neoantigen-based vaccine strategy using dendritic cells as vaccine platform to induce a tumor-specific CD8⁺ T cell response in three patients with advanced melanoma (293). Subsequently peptide- and mRNA-based neoantigen vaccines were shown to induce strong CD4⁺ and CD8⁺ T cell responses

alongside with significant tumor regression in melanoma patients (90, 91). Despite the promising results of these early clinical trials, several open questions on how to improve efficacy and feasibility of neoantigen-based tumor vaccines remain, that include identification of optimal antigens, delivery platforms, adjuvants, and routes of administration (4). Several clinical trials investigating the efficacy of neoantigen-based vaccine approaches in different cancer types are underway and will likely help to find answers to these questions (4, 291).

CONCLUSION

Adaptive immunity is critically involved in the pathogenesis of atherosclerotic cardiovascular diseases and cancer, which represent the two most common causes of death worldwide. During the past few decades, novel treatment strategies enhancing anti-tumor immune responses have already found their way into clinical practice, whereas successful translation of strategies targeting atherogenesis-related immune responses into the clinic has not yet been accomplished. Given that some immunotherapies (e.g., CD20-, EGFR-, IL-1β- or PCSK9-targeting antibodies) were shown to protect from both cancer and atherosclerosis, inflammatory processes and immunity underlying carcinogenesis and atherogenesis may be closely interconnected. Further characterization of tumorpromoting and proatherogenic immune responses may help to identify novel pharmacological targets that allow simultaneous treatment of both disease entities. Further characterization of tumor-promoting and proatherogenic immune responses may help to identify novel pharmacological targets that allow simultaneous treatment of both disease entities. In that regard, multimodal sequencing approaches, such as Cellular Indexing of Transcriptomes and Epitopes by Sequencing (CITE-Seq),

REFERENCES

- Roth GA, Abate D, Abate KH, Abay SM, Abbafati C, Abbasi N, et al. Global, regional, and national age-sex-specific mortality for 282 causes of death in 195 countries and territories, 1980–2017: a systematic analysis for the Global Burden of Disease Study 2017. *Lancet.* (2018) 392:1736–88. doi: 10.1016/S0140-6736(18)32203-7
- 2. Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? *Lancet.* (2001) 357:539–45. doi: 10.1016/S0140-6736(00)04046-0
- Libby P. Inflammation in atherosclerosis. Arterioscler Thromb Vasc Biol. (2012) 32:2045–51. doi: 10.1161/ATVBAHA.108.179705
- Blass E, Ott PA. Advances in the development of personalized neoantigenbased therapeutic cancer vaccines. *Nat Rev Clin Oncol.* (2021) 18:215–29. doi: 10.1038/s41571-020-00460-2
- Ait-Oufella H, Lavillegrand J-R, Tedgui A. Regulatory T cellenhancing therapies to treat atherosclerosis. *Cells.* (2021) 10:723. doi: 10.3390/cells10040723
- Nettersheim FS, De Vore L, Winkels H. Vaccination in atherosclerosis. *Cells*. (2020) 9:2560. doi: 10.3390/cells9122560
- Waldman AD, Fritz JM, Lenardo MJ. A guide to cancer immunotherapy: from T cell basic science to clinical practice. *Nat Rev Immunol.* (2020) 20:651–668. doi: 10.1038/s41577-020-0306-5
- Zhao TX, Mallat Z. Targeting the immune system in atherosclerosis: JACC state-of-the-art review. J Am Coll Cardiol. (2019) 73:1691–706. doi: 10.1016/j.jacc.2018.12.083

which allow combined measurement of gene and surfaceprotein expression on a single-cell level, will be valuable tools. Active immunization represents a novel, promising approach for the treatment of cancer and atherosclerosis. Preclinical studies have underscored the great anti-atherosclerotic potential of vaccination against plaque-related autoantigens. Further research is required to optimize this promising approach. Major objectives in this context include identification of optimal vaccine delivery platforms, adjuvants and administration routes. Furthermore, development of clinically feasible approaches to identify eligible patients, to determine expression of target antigens within an individual patient, and to monitor treatment responses will be crucial for broad implementation of this approach into clinical practice. If these obstacles can be overcome, active immunization may prospectively take cancer and atherosclerosis therapy to the next level.

AUTHOR CONTRIBUTIONS

FN wrote the manuscript. FP prepared the figure. FN and HW conceptualized the work. HW and FH supervised the work and provided funding. All authors substantially contributed to data research, critically discussed the content, reviewed the manuscript before submission, and have read and agreed to the published version of the manuscript.

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- 9. Dobosz P, Dzieciatkowski T. The intriguing history of cancer immunotherapy. *Front Immunol.* (2019) 10:2965. doi: 10.3389/fimmu.2019.02965
- Libby P. The changing landscape of atherosclerosis. *Nature*. (2021) 592:524– 33. doi: 10.1038/s41586-021-03392-8
- Dall'Olio FG, Marabelle A, Caramella C, Garcia C, Aldea M, Chaput N, et al. Tumour burden and efficacy of immune-checkpoint inhibitors. *Nat Rev Clin Oncol.* (2021) 2021:1–16. doi: 10.1038/s41571-021-00564-3
- Schwartz GG, Steg PG, Szarek M, Bhatt DL, Bittner VA, Diaz R, et al. Alirocumab and cardiovascular outcomes after acute coronary syndrome. N Engl J Med. (2018) 379:2097–107. doi: 10.1056/NEJMoa1801174
- Sabatine MS, Giugliano RP, Keech AC, Honarpour N, Wiviott SD, Murphy SA, et al. Evolocumab and clinical outcomes in patients with cardiovascular disease. N Engl J Med. (2017) 376:1713–22. doi: 10.1056/NEJMoa1615664
- Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, et al. Antiinflammatory therapy with canakinumab for atherosclerotic disease. N Engl J Med. (2017) 377:1119–31. doi: 10.1056/NEJMoa1707914
- Libby P. Targeting inflammatory pathways in cardiovascular disease: the inflammasome, interleukin-1, interleukin-6 and beyond. *Cells.* (2021) 10:951. doi: 10.3390/cells10040951
- Lobenwein D, Kocher F, Dobner S, Gollmann-Tepeköylü C, Holfeld J. Cardiotoxic mechanisms of cancer immunotherapy – a systematic review. *Int J Cardiol.* (2021) 323:179–87. doi: 10.1016/j.ijcard.2020.08.033
- 17. Drobni ZD, Alvi RM, Taron J, Zafar A, Murphy SP, Rambarat PK, et al. Association between immune checkpoint inhibitors with cardiovascular

events and atherosclerotic plaque. *Circulation.* (2020) 142:2299–311. doi: 10.1161/CIRCULATIONAHA.120.049981

- Arnold M, Rutherford MJ, Bardot A, Ferlay J, Andersson TML, Myklebust TÅ, et al. Progress in cancer survival, mortality, and incidence in seven high-income countries 1995–2014 (ICBP SURVMARK-2): a population-based study. *Lancet Oncol.* (2019) 20:1493–505. doi: 10.1016/S1470-2045(19)30456-5
- Stoltzfus KC, Zhang Y, Sturgeon K, Sinoway LI, Trifiletti DM, Chinchilli VM, et al. Fatal heart disease among cancer patients. *Nat Commun.* (2020) 11:1–8. doi: 10.1038/s41467-020-15639-5
- Sturgeon KM, Deng L, Bluethmann SM, Zhou S, Trifiletti DM, Jiang C, et al. A population-based study of cardiovascular disease mortality risk in US cancer patients. *Eur Heart J.* (2019) 40:3889–97. doi: 10.1093/eurheartj/ehz766
- Wang L, Wang F, Chen L, Geng Y, Yu S, Chen Z. Long-term cardiovascular disease mortality among 160 834 5-year survivors of adolescent and young adult cancer: an American population-based cohort study. *Eur Heart J.* (2020) 42:101–9. doi: 10.1093/eurheartj/ehaa779
- 22. Strongman H, Gadd S, Matthews A, Mansfield KE, Stanway S, Lyon AR, et al. Medium and long-term risks of specific cardiovascular diseases in survivors of 20 adult cancers: a population-based cohort study using multiple linked UK electronic health records databases. *Lancet.* (2019) 394:1041–54. doi: 10.1016/S0140-6736(19)31674-5
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* (2021) 71:209–49. doi: 10.3322/caac.21660
- Wolf D, Ley K. Immunity and inflammation in atherosclerosis. *Circ Res.* (2019) 124:315–27. doi: 10.1161/CIRCRESAHA.118.313591
- Bentzon JF, Otsuka F, Virmani R, Falk E. Mechanisms of plaque formation and rupture. *Circ Res.* (2014) 114:1852–66. doi: 10.1161/CIRCRESAHA.114.302721
- Wentzel JJ, Chatzizisis YS, Gijsen FJH, Giannoglou GD, Feldman CL, Stone PH. Endothelial shear stress in the evolution of coronary atherosclerotic plaque and vascular remodelling: current understanding and remaining questions. *Cardiovasc Res.* (2012) 96:234–43. doi: 10.1093/cvr/cvs217
- Roy P, Orecchioni M, Ley K. How the immune system shapes atherosclerosis: roles of innate and adaptive immunity. *Nat Rev Immunol.* (2021). doi: 10.1038/s41577-021-00584-1. [Epub ahead of print].
- Shah DK, Zúñiga-Pflücker JC. An overview of the intrathymic intricacies of T cell development. J Immunol. (2014) 192:4017–23. doi: 10.4049/jimmunol.1302259
- Klein I., Kyewski B, Allen PM, Hogquist KA. Positive and negative selection of the T cell repertoire: what thymocytes see (and don't see). *Nat Rev Immunol.* (2014) 14:377–91. doi: 10.1038/nri3667
- Van Den Broek T, Borghans JAM, Van Wijk F. The full spectrum of human naive T cells. Nat Rev Immunol. (2018) 18:363–73. doi: 10.1038/s41577-018-0001-y
- Neefjes J, Jongsma MLM, Paul P, Bakke O. Towards a systems understanding of MHC class i and MHC class II antigen presentation. *Nat Rev Immunol.* (2011) 11:823–36. doi: 10.1038/nri3084
- Smith-Garvin JE, Koretzky GA, Jordan MS. T cell activation. Annu Rev Immunol. (2009) 27:591–619. doi: 10.1146/annurev.immunol.021908.132706
- Zhou L, Chong MMW, Littman DR. Plasticity of CD4+ T cell lineage differentiation. *Immunity*. (2009) 30:646–55. doi: 10.1016/j.immuni.2009.05.001
- Saigusa R, Winkels H, Ley K. T cell subsets and functions in atherosclerosis. Nat Rev Cardiol. (2020) 17:387–401. doi: 10.1038/s41569-020-0352-5
- Jonasson L, Holm J, Skalli O, Bondjers G, Hansson GK. Regional accumulations of T cells, macrophages, and smooth muscle cells in the human atherosclerotic plaque. *Arteriosclerosis*. (1986) 6:131–8. doi: 10.1161/01.ATV.6.2.131
- Grivel JC, Ivanova O, Pinegina N, Blank PS, Shpektor A, Margolis LB, et al. Activation of T lymphocytes in atherosclerotic plaques. *Arterioscler Thromb Vasc Biol.* (2011) 31:2929–37. doi: 10.1161/ATVBAHA.111.237081
- 37. Stemme S, Faber B, Holm J, Wiklund O, Witztum JL, Hansson GK. T lymphocytes from human atherosclerotic plaques recognize oxidized

low density lipoprotein. Proc Natl Acad Sci USA. (1995) 92:3893-7. doi: 10.1073/pnas.92.9.3893

- Ma SD, Mussbacher M, Galkina E V. Functional role of B cells in atherosclerosis. *Cells*. (2021) 10:270. doi: 10.3390/cells10020270
- Schäfer S, Zernecke A. CD8+ T cells in atherosclerosis. Cells. (2020) 10:37. doi: 10.3390/cells10010037
- Tse K, Gonen A, Sidney J, Ouyang H, Witztum JL, Sette A, et al. Atheroprotective vaccination with MHC-II restricted peptides from ApoB-100. Front Immunol. (2013) 4:493. doi: 10.3389/fimmu.2013.00493
- Kimura T, Tse K, McArdle S, Gerhardt T, Miller J, Mikulski Z, et al. Atheroprotective vaccination with MHC-II-restricted ApoB peptides induces peritoneal IL-10-producing CD4T cells. *Am J Physiol Hear Circ Physiol.* (2017) 312:H781–90. doi: 10.1152/ajpheart.00798.2016
- Kimura T, Kobiyama K, Winkels H, Tse K, Miller J, Vassallo M, et al. Regulatory CD4+ T cells recognize major histocompatibility complex class II molecule-restricted peptide epitopes of apolipoprotein B. *Circulation*. (2018) 138:1130–43. doi: 10.1161/CIRCULATIONAHA.117.031420
- Wolf D, Gerhardt T, Winkels H, Anto Michel N, Pramod AB, Ghosheh Y, et al. Pathogenic autoimmunity in atherosclerosis evolves from initially protective ApoB-reactive CD4 ⁺ T-regulatory cells. *Circulation*. (2020) 142:1279–93. doi: 10.1161/CIRCULATIONAHA.119.042863
- 44. Gaddis DE, Padgett LE, Wu R, McSkimming C, Romines V, Taylor AM, et al. Apolipoprotein AI prevents regulatory to follicular helper T cell switching during atherosclerosis. *Nat Commun.* (2018) 9:1095. doi: 10.1038/s41467-018-03493-5
- Butcher MJ, Filipowicz AR, Waseem TC, McGary CM, Crow KJ, Magilnick N, et al. Atherosclerosis-driven treg plasticity results in formation of a dysfunctional subset of plastic IFNγ+ Th1/tregs. *Circ Res.* (2016) 119:1190– 203. doi: 10.1161/CIRCRESAHA.116.309764
- Klingenberg R, Gerdes N, Badeau RM, Gisterå A, Strodthoff D, Ketelhuth DFJ, et al. Depletion of FOXP3+ regulatory T cells promotes hypercholesterolemia and atherosclerosis. J Clin Invest. (2013) 123:1323–34. doi: 10.1172/JCI63891
- Sage AP, Tsiantoulas D, Binder CJ, Mallat Z. The role of B cells in atherosclerosis. Nat Rev Cardiol. (2019) 16:180–96. doi: 10.1038/s41569-018-0106-9
- Yla-Herttuala S, Palinski W, Butler SW, Picard S, Steinberg D, Witztum JL. Rabbit and human atherosclerotic lesions contain IgG that recognizes epitopes of oxidized LDL. *Arterioscler Thromb.* (1994) 14:32–40. doi: 10.1161/01.ATV.14.1.32
- Orekhov AN, Tertov V V., Kabakov AE, Adamova IY, Pokrovsky SN, Smirnov VN. Autoantibodies against modified low density lipoprotein. Nonlipid factor of blood plasma that stimulates foam cell formation. *Arterioscler Thromb.* (1991) 11:316–26. doi: 10.1161/01.ATV.11.2.316
- Parums D V., Brown DL, Mitchinson MJ. Serum antibodies to oxidized lowdensity lipoprotein and ceroid in chronic periaortitis. *Arch Pathol Lab Med.* (1990) 114:383–387.
- Zernecke A, Winkels H, Cochain C, Williams JW, Wolf D, Soehnlein O, et al. Meta-analysis of leukocyte diversity in atherosclerotic mouse aortas. *Circ Res.* (2020) 127:402–26. doi: 10.1161/CIRCRESAHA.120.316903
- 52. Shaw PX, Hörkkö S, Tsimikas S, Chang MK, Palinski W, Silverman GJ, et al. Human-derived anti-oxidized LDL autoantibody blocks uptake of oxidized LDL by macrophages and localizes to atherosclerotic lesions *in vivo. Arterioscler Thromb Vasc Biol.* (2001) 21:1333–9. doi: 10.1161/hq0801.093587
- Hörkkö S, Bird DA, Miller E, Itabe H, Leitinger N, Subbanagounder G, et al. Monoclonal autoantibodies specific for oxidized phospholipids or oxidized phospholipid–protein adducts inhibit macrophage uptake of oxidized lowdensity lipoproteins. J Clin Invest. (1999) 103:117–28. doi: 10.1172/JCI4533
- 54. Que X, Hung M-Y, Yeang C, Gonen A, Prohaska TA, Sun X, et al. Oxidized phospholipids are proinflammatory and proatherogenic in hypercholesterolaemic mice. *Nature*. (2018) 558:301–6. doi: 10.1038/s41586-018-0198-8
- Major AS, Fazio S, Linton MF. B-lymphocyte deficiency increases atherosclerosis in LDL receptor-null mice. Arterioscler Thromb Vasc Biol. (2002) 22:1892–8. doi: 10.1161/01.ATV.0000039169.47943.EE
- 56. Ait-Oufella H, Herbin O, Bouaziz JD, Binder CJ, Uyttenhove C, Laurans L, et al. B cell depletion reduces the development of

atherosclerosis in mice. J Exp Med. (2010) 207:1579–87. doi: 10.1084/jem. 20100155

- 57. Kyaw T, Tay C, Khan A, Dumouchel V, Cao A, To K, et al. Conventional B2 B cell depletion ameliorates whereas its adoptive transfer aggravates atherosclerosis. J Immunol. (2010) 185:4410–19. doi: 10.4049/jimmunol.1000033
- Cyster JG, Allen CDC. B cell responses: cell interaction dynamics and decisions. *Cell.* (2019) 177:524–40. doi: 10.1016/j.cell.2019.03.016
- Baumgarth N. The double life of a B-1 cell: self-reactivity selects for protective effector functions. *Nat Rev Immunol.* (2011) 11:34–46. doi: 10.1038/nri2901
- Oiseth SJ, Aziz MS. Cancer immunotherapy: a brief review of the history, possibilities, and challenges ahead. J Cancer Metastasis Treat. (2017) 3:250– 61. doi: 10.20517/2394-4722.2017.41
- Busch W. Aus der Sitzung der medicinischen section vom 13 november 1867. Berlin Klin Wochenschr. (1868) 5:137.
- Fehleisen F. Ueber die Züchtung der Erysipelkokken auf künstlichem Nährboden und ihre Uebertragbarkeit auf den Menschen. Dtsch Med Wochenschr. (1882) 8:553–4. doi: 10.1055/s-0029-1196806
- Coley WB. The treatment of inoperable sarcoma by bacterial toxins (the mixed toxins of the streptococcus erysipelas and the Bacillus prodigiosus). *Proc R Soc Med.* (1910) 3:1. doi: 10.1177/003591571000301601
- Ehrlich P. Ueber den jetzigen Stand der Karzinomforschung. Ned Tijdschr Geneeskd. (1909) 53:273–90.
- 65. Gross L. Intradermal immunization of C3H mice against a sarcoma that originated in an animal of the same line. *Cancer Res.* (1943) 3:326–33.
- 66. Foley EJ. Antigenic properties of methylcholanthrene-induced tumors in mice of the strain of origin. *Cancer Res.* (1953) 13:835–7.
- 67. Ribatti D. The concept of immune surveillance against tumors: the first theories. *Oncotarget*. (2017) 8:7175. doi: 10.18632/oncotarget.12739
- Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoediting: from immunosurveillance to tumor escape. *Nat Immunol.* (2002) 3:991–8. doi: 10.1038/ni1102-991
- Stutman O. Tumor development after 3-methylcholanthrene in immunologically deficient athymic-nude mice. *Science*. (1974) 183:534–6. doi: 10.1126/science.183.4124.534
- Shankaran V, Ikeda H, Bruce AT, White JM, Swanson PE, Old LJ, et al. IFNγ and lymphocytes prevent primary tumour development and shape tumour immunogenicity. *Nature*. (2001) 410:1107–11. doi: 10.1038/35074122
- Street SEA, Cretney E, Smyth MJ. Perforin and interferon-γ activities independently control tumor initiation, growth, and metastasis. *Blood.* (2001) 97:192–7. doi: 10.1182/blood.V97.1.192
- Street SEA, Trapani JA, MacGregor D, Smyth MJ. Suppression of lymphoma and epithelial malignancies effected by interferon γ. J Exp Med. (2002) 196:129. doi: 10.1084/jem.20020063
- Kaplan DH, Shankaran V, Dighe AS, Stockert E, Aguet M, Old LJ, et al. Demonstration of an interferon γ-dependent tumor surveillance system in immunocompetent mice. *Proc Natl Acad Sci.* (1998) 95:7556–61. doi: 10.1073/pnas.95.13.7556
- 74. Pham SM, Kormos RL, Landreneau RJ, Kawai A, Gonzalez-Cancel I, Hardesty RL, et al. Solid tumors after heart transplantation: lethality of lung cancer. *Ann Thorac Surg.* (1995) 60:1623–6. doi: 10.1016/0003-4975(95)00120-4
- Penn I. Malignant melanoma in organ allograft recipients. *Transplantation*. (1996) 61:274–8. doi: 10.1097/00007890-199601270-00019
- Birkeland SA, Storm HH, Lamm LU, Barlow L, Blohmé I, Forsberg B, et al. Cancer risk after renal transplantation in the nordic countries, 1964–1986. *Int J Cancer*. (1995) 60:183–9. doi: 10.1002/ijc.2910600209
- Clark WH, Elder DE, Guerry D, Braitman LE, Trock BJ, Schultz D, et al. Model predicting survival in stage I melanoma based on tumor progression. *JNCI J Natl Cancer Inst.* (1989) 81:1893–904. doi: 10.1093/jnci/81. 24.1893
- Clemente CG, Mihm Jr. MC, Bufalino R, Zurrida S, Collini P, Cascinelli N. Prognostic value of tumor infiltrating lymphocytes in the vertical growth phase of primary cutaneous melanoma. *Cancer.* (1996) 77:1303–10.
- Smyth MJ, Thia KYT, Street SEA, Cretney E, Trapani JA, Taniguchi M, et al. Differential tumor surveillance by Natural Killer (Nk) and Nkt cells. J Exp Med. (2000) 191:661–8. doi: 10.1084/jem.191.4.661

- Svane IM, Engel A-M, Nielsen M-B, Ljunggren H-G, Rygaard J, Werdelin O. Chemically induced sarcomas from nude mice are more immunogenic than similar sarcomas from congenic normal mice. *Eur J Immunol.* (1996) 26:1844–50. doi: 10.1002/eji.1830260827
- Fridman WH. From cancer immune surveillance to cancer immunoediting: birth of modern immuno-oncology. J Immunol. (2018) 201:825–6. doi: 10.4049/jimmunol.1800827
- Schumacher TN, Schreiber RD. Neoantigens in cancer immunotherapy. Science. (2015) 348:69–74. doi: 10.1126/science.aaa4971
- O'Donnell JS, Teng MWL, Smyth MJ. Cancer immunoediting and resistance to T cell-based immunotherapy. *Nat Rev Clin Oncol.* (2018) 16:151–67. doi: 10.1038/s41571-018-0142-8
- Castle JC, Kreiter S, Diekmann J, Löwer M, Roemer N van de, Graaf J de, et al. Exploiting the mutanome for tumor vaccination. *Cancer Res.* (2012) 72:1081–91. doi: 10.1158/0008-5472.CAN-11-3722
- Matsushita H, Vesely MD, Koboldt DC, Rickert CG, Uppaluri R, Magrini VJ, et al. Cancer exome analysis reveals a T-cell-dependent mechanism of cancer immunoediting. *Nature*. (2012) 482:400–4. doi: 10.1038/nature10755
- Kreiter S, Vormehr M, van de Roemer N, Diken M, Löwer M, Diekmann J, et al. Mutant MHC class II epitopes drive therapeutic immune responses to cancer. *Nature*. (2015) 520:692–6. doi: 10.1038/nature14426
- Robbins PF, Lu Y-C, El-Gamil M, Li YF, Gross C, Gartner J, et al. Mining exomic sequencing data to identify mutated antigens recognized by adoptively transferred tumor-reactive T cells. *Nat Med.* (2013) 19:747–52. doi: 10.1038/nm.3161
- Balachandran VP, Łuksza M, Zhao JN, Makarov V, Moral JA, Remark R, Herbst B, et al. Identification of unique neoantigen qualities in long-term survivors of pancreatic cancer. *Nature*. (2017) 551:512–6. doi: 10.1038/nature24462
- Linnemann C, van Buuren MM, Bies L, Verdegaal EME, Schotte R, Calis JJA, et al. High-throughput epitope discovery reveals frequent recognition of neoantigens by CD4+ T cells in human melanoma. *Nat Med.* (2014) 21:81–5. doi: 10.1038/nm.3773
- Sahin U, Derhovanessian E, Miller M, Kloke B-P, Simon P, Löwer M, et al. Personalized RNA mutanome vaccines mobilize poly-specific therapeutic immunity against cancer. *Nature*. (2017) 547:222–6. doi: 10.1038/nature23003
- Ott PA, Hu Z, Keskin DB, Shukla SA, Sun J, Bozym DJ, et al. An immunogenic personal neoantigen vaccine for patients with melanoma. *Nature*. (2017) 547:217–21. doi: 10.1038/nature22991
- 92. Alspach E, Lussier DM, Miceli AP, Kizhvatov I, DuPage M, Luoma AM, et al. MHC-II neoantigens shape tumour immunity and response to immunotherapy. *Nature*. (2019) 574:696–701. doi: 10.1038/s41586-019-1671-8
- Lennerz V, Fatho M, Gentilini C, Frye RA, Lifke A, Ferel D, et al. The response of autologous T cells to a human melanoma is dominated by mutated neoantigens. *Proc Natl Acad Sci.* (2005) 102:16013–18. doi: 10.1073/pnas.0500090102
- Brightman SE, Naradikian MS, Miller AM, Schoenberger SP. Harnessing neoantigen specific CD4 T cells for cancer immunotherapy. *J Leukoc Biol.* (2020) 107:625–33. doi: 10.1002/JLB.5RI0220-603RR
- Zander R, Schauder D, Xin G, Nguyen C, Wu X, Zajac A, et al. CD4+ T cell help is required for the formation of a cytolytic CD8+ T cell subset that protects against chronic infection and cancer. *Immunity.* (2019) 51:1028– 42.e4. doi: 10.1016/j.immuni.2019.10.009
- Schoenberger SP, Toes REM, van der Voort EIH, Offringa R, Melief CJM. T-cell help for cytotoxic T lymphocytes is mediated by CD40–CD40L interactions. *Nature*. (1998) 393:480–3. doi: 10.1038/31002
- Tay RE, Richardson EK, Toh HC. Revisiting the role of CD4+ T cells in cancer immunotherapy—new insights into old paradigms. *Cancer Gene Ther.* (2020) 28:5–17. doi: 10.1038/s41417-020-0183-x
- Tran E, Turcotte S, Gros A, Robbins PF, Lu YC, Dudley ME, et al. Cancer immunotherapy based on mutation-specific CD4+ T cells in a patient with epithelial cancer. *Science*. (2014) 344:641–5. doi: 10.1126/science.1251102
- 99. Laheurte C, Dosset M, Vernerey D, Boullerot L, Gaugler B, Gravelin E, et al. Distinct prognostic value of circulating anti-telomerase CD4+ Th1 immunity and exhausted PD-1+/TIM-3+ T cells in lung cancer. *Br J Cancer*. (2019) 121:405–16. doi: 10.1038/s41416-019-0531-5

- 100. Sayour EJ, McLendon P, McLendon R, De Leon G, Reynolds R, Kresak J, et al. Increased proportion of FoxP3+ regulatory T cells in tumor infiltrating lymphocytes is associated with tumor recurrence and reduced survival in patients with glioblastoma. *Cancer Immunol Immunother*. (2015) 64:419–27. doi: 10.1007/s00262-014-1651-7
- Nishikawa H, Sakaguchi S. Regulatory T cells in cancer immunotherapy. Curr Opin Immunol. (2014) 27:1–7. doi: 10.1016/j.coi.2013.12.005
- 102. Valzasina B, Piconese S, Guiducci C, Colombo MP. Tumor-induced expansion of regulatory T cells by conversion of CD4+CD25– lymphocytes is thymus and proliferation independent. *Cancer Res.* (2006) 66:4488–95. doi: 10.1158/0008-5472.CAN-05-4217
- 103. Bonertz A, Weitz J, Pietsch D-HK, Rahbari NN, Schlude C, Ge Y, et al. Antigen-specific Tregs control T cell responses against a limited repertoire of tumor antigens in patients with colorectal carcinoma. *J Clin Invest.* (2009) 119:3311–21. doi: 10.1172/JCI39608
- 104. Schreiber S, Hammers CM, Kaasch AJ, Schraven B, Dudeck A, Kahlfuss S. Metabolic interdependency of Th2 cell-mediated type 2 immunity and the tumor microenvironment. *Front Immunol.* (2021) 12:632581. doi: 10.3389/fimmu.2021.632581
- 105. Li T, Wu B, Yang T, Zhang L, Jin K. The outstanding antitumor capacity of CD4+ T helper lymphocytes. *Biochim Biophys Acta Rev Cancer*. (2020) 1874:188439. doi: 10.1016/j.bbcan.2020.188439
- 106. Calabretta R, Hoeller C, Pichler V, Mitterhauser M, Karanikas G, Haug A, et al. Immune checkpoint inhibitor therapy induces inflammatory activity in large arteries. *Circulation*. (2020) 142:2396–8. doi: 10.1161/CIRCULATIONAHA.120.048708
- 107. Bu D, Tarrio M, Maganto-Garcia E, Stavrakis G, Tajima G, Lederer J, et al. Impairment of the programmed cell death-1 pathway increases atherosclerotic lesion development and inflammation. *Arterioscler Thromb Vasc Biol.* (2011) 31:1100–7. doi: 10.1161/ATVBAHA.111.224709
- 108. Gotsman I, Grabie N, Dacosta R, Sukhova G, Sharpe A, Lichtman AH. Proatherogenic immune responses are regulated by the PD-1/PD-L pathway in mice. J Clin Invest. (2007) 117:2974–82. doi: 10.1172/JCI31344
- 109. Poels K, Leent MMT van, Reiche ME, Kusters PJH, Huveneers S, Winther MPJ de, et al. Antibody-mediated inhibition of CTLA4 aggravates atherosclerotic plaque inflammation and progression in hyperlipidemic mice. *Cells*. (2020) 9:1987. doi: 10.3390/cells9091987
- 110. Poels K, van Leent MMT, Boutros C, Tissot H, Roy S, Meerwaldt AE, et al. Immune checkpoint inhibitor therapy aggravates T cell-driven plaque inflammation in atherosclerosis. *JACC CardioOncology*. (2020) 2:599–610. doi: 10.1016/j.jaccao.2020.08.007
- 111. Cochain C, Chaudhari SM, Koch M, Wiendl H, Eckstein H-H, Zernecke A. Programmed cell death-1 deficiency exacerbates T cell activation and atherogenesis despite expansion of regulatory T cells in atherosclerosisprone mice. *PLoS ONE*. (2014) 9:e93280. doi: 10.1371/journal.pone.0093280
- 112. Kim DG, Lee J, Seo WJ, Lee JG, Kim BS, Kim MS, et al. Rituximab protects against development of atherosclerotic cardiovascular disease after kidney transplantation: a propensity-matched study. *Sci Rep.* (2019) 9:1–8. doi: 10.1038/s41598-019-52942-8
- 113. Novikova DS, Popkova T V., Lukina G V., Luchikhina EL, Karateev DE, Volkov A V., et al. The effects of rituximab on lipids, arterial stiffness and carotid intima-media thickness in rheumatoid arthritis. *J Korean Med Sci.* (2016) 31:202–7. doi: 10.3346/jkms.2016.31.2.202
- 114. Hsue PY, Scherzer R, Grunfeld C, Imboden J, Wu Y, Puerto G del, et al. Depletion of B-cells with rituximab improves endothelial function and reduces inflammation among individuals with rheumatoid arthritis. *J Am Heart Assoc.* (2014) 3:e001278. doi: 10.1161/JAHA.114.001267
- 115. Totzeck M, Mincu RI, Rassaf T. Cardiovascular adverse events in patients with cancer treated with bevacizumab: a meta-analysis of more than 20 000 patients. J Am Heart Assoc. (2017) 6:e006278. doi: 10.1161/JAHA.117.006278
- 116. Schutz FAB, Je Y, Azzi GR, Nguyen PL, Choueiri TK. Bevacizumab increases the risk of arterial ischemia: a large study in cancer patients with a focus on different subgroup outcomes. *Ann Oncol.* (2011) 22:1404–12. doi: 10.1093/annonc/mdq587
- 117. Chen X-L, Lei Y-H, Liu C-F, Yang Q-F, Zuo P-Y, Liu C-Y, et al. Angiogenesis inhibitor bevacizumab increases the risk of ischemic heart disease associated with chemotherapy: a meta-analysis. *PLoS ONE.* (2013) 8:e66721. doi: 10.1371/journal.pone.0066721

- 118. Winnik S, Lohmann C, Siciliani G, Lukowicz T von, Kuschnerus K, Kraenkel N, et al. Systemic VEGF inhibition accelerates experimental atherosclerosis and disrupts endothelial homeostasis implications for cardiovascular safety. *Int J Cardiol.* (2013) 168:2453–61. doi: 10.1016/j.ijcard.2013.03.010
- 119. Zeboudj L, Giraud A, Guyonnet L, Zhang Y, Laurans L, Esposito B, et al. Selective EGFR (Epidermal Growth Factor Receptor) deletion in myeloid cells limits atherosclerosis—brief report. *Arterioscler Thromb Vasc Biol.* (2018) 38:114–19. doi: 10.1161/ATVBAHA.117.309927
- 120. Zeboudj L, Maître M, Guyonnet L, Laurans L, Joffre J, Lemarie J, et al. Selective EGF-receptor inhibition in CD4+ T cells induces anergy and limits atherosclerosis. J Am Coll Cardiol. (2018) 71:160–72. doi: 10.1016/j.jacc.2017.10.084
- 121. Wang L, Huang Z, Huang W, Chen X, Shan P, Zhong P, et al. Inhibition of epidermal growth factor receptor attenuates atherosclerosis via decreasing inflammation and oxidative stress. *Sci Rep.* (2017) 7:1–14. doi: 10.1038/srep45917
- 122. Somers EC, Zhao W, Lewis EE, Wang L, Wing JJ, Sundaram B, et al. Type I interferons are associated with subclinical markers of cardiovascular disease in a cohort of systemic lupus erythematosus patients. *PLoS ONE.* (2012) 7:e37000. doi: 10.1371/journal.pone.0037000
- 123. Santinelli L, Girolamo G De, Borrazzo C, Vassalini P, Pinacchio C, Cavallari EN, et al. Alteration of type I interferon response is associated with subclinical atherosclerosis in virologically suppressed HIV-1-infected male patients. *J Med Virol.* (2021) 93:4930–8. doi: 10.1002/jmv.27028
- 124. Levy Z, Rachmani R, Trestman S, Dvir A, Shaish A, Ravid M, et al. Low-dose interferon-α accelerates atherosclerosis in an LDL receptor-deficient mouse model. *Eur J Intern Med.* (2003) 14:479–83. doi: 10.1016/j.ejim.2003.08.010
- Lee RE, Lotze MT, Skibber JM, Tucker E, Bonow RO, Ognibene FP, et al. Cardiorespiratory effects of immunotherapy with interleukin-2. *J Clin Oncol.* (1989) 7:7–20. doi: 10.1200/JCO.1989.7.1.7
- 126. Upadhya S, Mooteri S, Peckham N, Pai RG. Atherogenic Effect of interleukin-2 and antiatherogenic effect of interleukin-2 antibody in apo-E-deficient mice. *Angiology*. (2016) 55:289–94. doi: 10.1177/000331970405500308
- 127. Dinh TN, Kyaw TS, Kanellakis P, To K, Tipping P, Toh B-H, et al. Cytokine therapy with interleukin-2/anti-interleukin-2 monoclonal antibody complexes expands CD4+CD25+Foxp3+ regulatory T cells and attenuates development and progression of atherosclerosis. *Circulation.* (2012) 126:1256–66. doi: 10.1161/CIRCULATIONAHA.112.099044
- Ridker PM, Everett BM, Pradhan A, MacFadyen JG, Solomon DH, Zaharris E, et al. Low-dose methotrexate for the prevention of atherosclerotic events. *N Engl J Med.* (2019) 380:752–62. doi: 10.1056/NEJMoa1809798
- 129. Di Francesco V, Gurgone D, Palomba R, Ferreira MFMM, Catelani T, Cervadoro A, et al. Modulating lipoprotein transcellular transport and atherosclerotic plaque formation in ApoE-/-mice via nanoformulated lipidmethotrexate conjugates. ACS Appl Mater Interfaces. (2020) 12:37943–56. doi: 10.1021/acsami.0c12202
- Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. Nat Rev Cancer. (2012) 12:252–64. doi: 10.1038/ nrc3239
- Ledford H, Else H, Warren M. Cancer immunologists scoop medicine Nobel prize. Nature. (2018) 562:20–21. doi: 10.1038/d41586-018-06751-0
- 132. Robert C. A decade of immune-checkpoint inhibitors in cancer therapy. *Nat Commun.* (2020) 11:1–3. doi: 10.1038/s41467-020-17670-y
- 133. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved survival with ipilimumab in patients with metastatic melanoma. N Engl J Med. (2010) 363:711–23. doi: 10.1056/NEJMoa1003466
- Twomey JD, Zhang B. Cancer immunotherapy update: FDA-approved checkpoint inhibitors and companion diagnostics. AAPS J. (2021) 23:1–11. doi: 10.1208/s12248-021-00574-0
- Mullard A. FDA approves 100th monoclonal antibody product. Nat Rev Drug Discov. (2021) 20:491–5. doi: 10.1038/d41573-021-00079-7
- Photopoulos J. The future of tissue-agnostic drugs. *Nature*. (2020) 585:S16– 18. doi: 10.1038/d41586-020-02679-6
- 137. Haslam A, Prasad V. Estimation of the percentage of US patients with cancer who are eligible for and respond to checkpoint inhibitor immunotherapy drugs. *JAMA Netw Open.* (2019) 2:e192535. doi: 10.1001/jamanetworkopen.2019.2535

- Postow MA, Sidlow R, Hellmann MD. Immune-related adverse events associated with immune checkpoint blockade. N Engl J Med. (2018) 378:158– 68. doi: 10.1056/NEJMra1703481
- Chen L, Flies DB. Molecular mechanisms of T cell co-stimulation and co-inhibition. *Nat Rev Immunol.* (2013) 13:227–42. doi: 10.1038/nri3405
- 140. Simons KH, de Jong A, Jukema JW, de Vries MR, Arens R, Quax PHA. T cell co-stimulation and co-inhibition in cardiovascular disease: a double-edged sword. *Nat Rev Cardiol.* (2019) 16:325–43. doi: 10.1038/s41569-019-0164-7
- Buchbinder EI, Desai A. CTLA-4 and PD-1 pathways similarities, differences, and implications of their inhibition. *Am J Clin Oncol Cancer Clin Trials*. (2016) 39:98–106. doi: 10.1097/COC.00000000000239
- Ramos-Casals M, Brahmer JR, Callahan MK, Flores-Chávez A, Keegan N, Khamashta MA, et al. Immune-related adverse events of checkpoint inhibitors. *Nat Rev Dis Prim.* (2020) 6:1–21. doi: 10.1038/s41572-020-0160-6
- 143. Xu C, Chen Y-P, Du X-J, Liu J-Q, Huang C-L, Chen L, et al. Comparative safety of immune checkpoint inhibitors in cancer: systematic review and network meta-analysis. *BMJ*. (2018) 363:4226. doi: 10.1136/bmj.k4226
- 144. Martins F, Sofiya L, Sykiotis GP, Lamine F, Maillard M, Fraga M, et al. Adverse effects of immune-checkpoint inhibitors: epidemiology, management and surveillance. *Nat Rev Clin Oncol.* (2019) 16:563–80. doi: 10.1038/s41571-019-0218-0
- 145. Wang DY, Salem J-E, Cohen J V., Chandra S, Menzer C, Ye F, et al. Fatal toxic effects associated with immune checkpoint inhibitors: a systematic review and meta-analysis. *JAMA Oncol.* (2018) 4:1721–8. doi: 10.1001/jamaoncol.2018.3923
- 146. Dolladille C, Akroun J, Morice P-M, Dompmartin A, Ezine E, Sassier M, et al. Cardiovascular immunotoxicities associated with immune checkpoint inhibitors: a safety meta-analysis. *Eur Heart J*. (2021) 48:ehab618. doi: 10.1093/eurheartj/ehab618
- 147. Salem J-E, Manouchehri A, Moey M, Lebrun-Vignes B, Bastarache L, Pariente A, et al. Cardiovascular toxicities associated with immune checkpoint inhibitors: an observational, retrospective, pharmacovigilance study. *Lancet Oncol.* (2018) 19:1579–89. doi: 10.1016/S1470-2045(18)30608-9
- 148. Poels K, Neppelenbroek SIM, Kersten MJ, Antoni ML, Lutgens E, Seijkens TTP. Immune checkpoint inhibitor treatment and atherosclerotic cardiovascular disease: an emerging clinical problem. *J Immunother Cancer*. (2021) 9:e002916. doi: 10.1136/jitc-2021-002916
- Inno A, Chiampan A, Lanzoni L, Verzè M, Molon G, Gori S. Immune checkpoint inhibitors and atherosclerotic vascular events in cancer patients. *Front Cardiovasc Med.* (2021) 8:652186. doi: 10.3389/fcvm.2021.652186
- 150. Bar J, Markel G, Gottfried T, Percik R, Leibowitz-Amit R, Berger R, et al. Acute vascular events as a possibly related adverse event of immunotherapy: a single-institute retrospective study. *Eur J Cancer.* (2019) 120:122–31. doi: 10.1016/j.ejca.2019.06.021
- 151. Pierpont TM, Limper CB, Richards KL. Past, present, and future of rituximab—the world's first oncology monoclonal antibody therapy. *Front* Oncol. (2018) 3:163. doi: 10.3389/fonc.2018.00163
- Scott AM, Wolchok JD, Old LJ. Antibody therapy of cancer. Nat Rev Cancer. (2012) 12:278–87. doi: 10.1038/nrc3236
- Zahavi D, Weiner L. Monoclonal antibodies in cancer therapy. *Antibodies*. (2020) 9:34. doi: 10.3390/antib9030034
- Lee DSW, Rojas OL, Gommerman JL. B cell depletion therapies in autoimmune disease: advances and mechanistic insights. *Nat Rev Drug Discov.* (2020) 20:179–99. doi: 10.1038/s41573-020-00092-2
- 155. Kyaw T, Cui P, Tay C, Kanellakis P, Hosseini H, Liu E, et al. BAFF receptor mAb treatment ameliorates development and progression of atherosclerosis in hyperlipidemic ApoE-/- mice. PLoS ONE. (2013) 8:e60430. doi: 10.1371/journal.pone.0060430
- 156. Tsiantoulas D, Sage AP, Göderle L, Ozsvar-Kozma M, Murphy D, Porsch F, et al. B cell-activating factor neutralization aggravates atherosclerosis. *Circulation.* (2018) 138:2263–73. doi: 10.1161/CIRCULATIONAHA.117.032790
- 157. Porsch F, Binder CJ. Impact of B-cell-targeted therapies on cardiovascular disease. Arterioscler Thromb Vasc Biol. (2019) 39:1705–14. doi: 10.1161/ATVBAHA.119.311996
- 158. Centa M, Jin H, Hofste L, Hellberg S, Busch A, Baumgartner R, et al., et al. Germinal center-derived antibodies promote

atherosclerosis plaque size and stability. *Circulation*. (2019) 139:2466–82. doi: 10.1161/CIRCULATIONAHA.118.038534

- 159. Tay C, Liu Y-H, Kanellakis P, Kallies A, Li Y, Cao A, et al. Follicular B cells promote atherosclerosis via T cell-mediated differentiation into plasma cells and secreting pathogenic immunoglobulin G. *Arterioscler Thromb Vasc Biol.* (2018) 38:e71–84. doi: 10.1161/ATVBAHA.117.310678
- Ellis LM, Hicklin DJ. VEGF-targeted therapy: mechanisms of anti-tumour activity. Nat Rev Cancer. (2008) 8:579–91. doi: 10.1038/nrc2403
- 161. Arnott C, Punnia-Moorthy G, Tan J, Sadeghipour S, Bursill C, Patel S. The vascular endothelial growth factor inhibitors ranibizumab and aflibercept markedly increase expression of atherosclerosis-associated inflammatory mediators on vascular endothelial cells. *PLoS ONE.* (2016) 11:e0150688. doi: 10.1371/journal.pone.0150688
- 162. Seijkens TTP, Lutgens E. Cardiovascular oncology: exploring the effects of targeted cancer therapies on atherosclerosis. *Curr Opin Lipidol.* (2018) 29:381–8. doi: 10.1097/MOL.000000000000538
- Mendelsohn J, Baselga J. The EGF receptor family as targets for cancer therapy. Oncogene. (2000) 19:6550–65. doi: 10.1038/sj.onc.1204082
- Ciardiello F, Tortora G. EGFR antagonists in cancer treatment. N Engl J Med. (2009) 358:1160–74. doi: 10.1056/NEJMra0707704
- 165. Slamon DJ, Clark GM, Wong SG, Levin WJ, Ullrich A, McGuire WL. Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene. *Science*. (1987) 235:182–91. doi: 10.1126/science.3798106
- 166. Nicholson R., Gee JM., Harper M. EGFR and cancer prognosis. Eur J Cancer. (2001) 37:9–15. doi: 10.1016/S0959-8049(01)00231-3
- 167. Slamon DJ, Leyland-Jones B, Shak S, Fuchs H, Paton V, Bajamonde A, et al. Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. N Engl J Med. (2001) 344:783–92. doi: 10.1056/NEJM200103153441101
- Jonker DJ, O'Callaghan CJ, Karapetis CS, Zalcberg JR, Tu D, Au H-J, et al. Cetuximab for the treatment of colorectal cancer. N Engl J Med. (2009) 2:2040–8. doi: 10.1056/NEJMoa071834
- 169. Bang Y-J, Cutsem E Van, Feyereislova A, Chung HC, Shen L, Sawaki A, et al. Trastuzumab in combination with chemotherapy versus chemotherapy alone for treatment of HER2-positive advanced gastric or gastro-oesophageal junction cancer (ToGA): a phase 3, open-label, randomised controlled trial. *Lancet.* (2010) 376:687–97. doi: 10.1016/S0140-6736(10)61121-X
- 170. Vermorken JB, Mesia R, Rivera F, Remenar E, Kawecki A, Rottey S, et al. Platinum-based chemotherapy plus cetuximab in head and neck cancer. N Engl J Med. (2009) 359:1116–27. doi: 10.1056/NEJMoa0802656
- 171. Zamorano JL, Lancellotti P, Rodriguez Muñoz D, Aboyans V, Asteggiano R, Galderisi M, et al. 2016 ESC Position Paper on cancer treatments and cardiovascular toxicity developed under the auspices of the ESC Committee for Practice Guidelines. *Eur Heart J.* (2016) 37:2768–801. doi: 10.1093/eurheartj/ehw211
- 172. Gronich N, Lavi I, Barnett-Griness O, Saliba W, Abernethy DR, Rennert G. Tyrosine kinase-targeting drugs-associated heart failure. *Br J Cancer.* (2017) 116:1366–73. doi: 10.1038/bjc.2017.88
- 173. Bowles EJA, Wellman R, Feigelson HS, Onitilo AA, Freedman AN, Delate T, et al. Risk of heart failure in breast cancer patients after anthracycline and trastuzumab treatment: a retrospective cohort study. *JNCI J Natl Cancer Inst.* (2012) 104:1293–305. doi: 10.1093/jnci/djs317
- 174. Banke A, Fosbøl EL, Ewertz M, Videbæk L, Dahl JS, Poulsen MK, et al. Long-term risk of heart failure in breast cancer patients after adjuvant chemotherapy with or without trastuzumab. *JACC Hear Fail.* (2019) 7:217– 24. doi: 10.1016/j.jchf.2018.09.001
- 175. Crone SA, Zhao Y-Y, Fan L, Gu Y, Minamisawa S, Liu Y, et al. ErbB2 is essential in the prevention of dilated cardiomyopathy. *Nat Med.* (2002) 8:459–65. doi: 10.1038/nm0502-459
- 176. Dreux AC, Lamb DJ, Modjtahedi H, Ferns GAA. The epidermal growth factor receptors and their family of ligands: their putative role in atherogenesis. *Atherosclerosis.* (2006) 186:38–53. doi: 10.1016/j.atherosclerosis.2005.06.038
- 177. Jian W, Wei C-M, Guan J-H, Mo C-H, Xu Y-T, Zheng W-B, et al. Association between serum HER2/ErbB2 levels and coronary artery disease: a case-control study. *J Transl Med.* (2020) 18:1–10. doi: 10.1186/s12967-020-02292-1

- Mindur JE, Swirski FK. Growth factors as immunotherapeutic targets in cardiovascular disease. Arterioscler Thromb Vasc Biol. (2019) 39:1275–87. doi: 10.1161/ATVBAHA.119.311994
- 179. Hoyer FF, Naxerova K, Schloss MJ, Hulsmans M, Nair A V., Dutta P, et al. Tissue-specific macrophage responses to remote injury impact the outcome of subsequent local immune challenge. *Immunity*. (2019) 51:899–914.e7. doi: 10.1016/j.immuni.2019.10.010
- Berraondo P, Sanmamed MF, Ochoa MC, Etxeberria I, Aznar MA, Pérez-Gracia JL, et al. Cytokines in clinical cancer immunotherapy. *Br J Cancer*. (2018) 120:6–15. doi: 10.1038/s41416-018-0328-y
- Conlon KC, Miljkovic MD, Waldmann TA. Cytokines in the treatment of cancer. J Interf Cytokine Res. (2019) 39:6. doi: 10.1089/jir.2018.0019
- Chen H-J, Tas SW, de Winther MPJ. Type-I interferons in atherosclerosis. J Exp Med. (2020) 217:e20190459. doi: 10.1084/jem.20190459
- Boshuizen MCS, de Winther MPJ. Interferons as essential modulators of atherosclerosis. Arterioscler Thromb Vasc Biol. (2015) 35:1579–88. doi: 10.1161/ATVBAHA.115.305464
- 184. Li J, Fu Q, Cui H, Qu B, Pan W, Shen N, et al. Interferon-α priming promotes lipid uptake and macrophage-derived foam cell formation: a novel link between interferon-α and atherosclerosis in lupus. *Arthritis Rheum*. (2011) 63:492–502. doi: 10.1002/art.30165
- 185. Lai J-H, Hung L-F, Huang C-Y, Wu D-W, Wu C-H, Ho L-J. Mitochondrial protein CMPK2 regulates IFN alpha-enhanced foam cell formation, potentially contributing to premature atherosclerosis in SLE. *Arthritis Res Ther.* (2021) 23:1–12. doi: 10.1186/s13075-021-02470-6
- 186. Srivastava S, Koch MA, Pepper M, Campbell DJ. Type I interferons directly inhibit regulatory T cells to allow optimal antiviral T cell responses during acute LCMV infection. J Exp Med. (2014) 211:961–74. doi: 10.1084/jem.20131556
- 187. Gangaplara A, Martens C, Dahlstrom E, Metidji A, Gokhale AS, Glass DD, et al. Type I interferon signaling attenuates regulatory T cell function in viral infection and in the tumor microenvironment. *PLoS Pathog.* (2018) 14:e10069855. doi: 10.1371/journal.ppat.1006985
- 188. Niessner A, Sato K, Chaikof EL, Colmegna I, Goronzy JJ, Weyand CM. Pathogen-sensing plasmacytoid dendritic cells stimulate cytotoxic T-cell function in the atherosclerotic plaque through interferon-α. *Circulation*. (2006) 114:2482–9. doi: 10.1161/CIRCULATIONAHA.106.642801
- 189. Niessner A, Shin MS, Pryshchep O, Goronzy JJ, Chaikof EL, Weyand CM. Synergistic proinflammatory effects of the antiviral cytokine interferon- α and toll-like receptor 4 ligands in the atherosclerotic plaque. *Circulation*. (2007) 116:2043–52. doi: 10.1161/CIRCULATIONAHA.107.697789
- 190. Denny MF, Thacker S, Mehta H, Somers EC, Dodick T, Barrat FJ, et al. Interferon- α promotes abnormal vasculogenesis in lupus: a potential pathway for premature atherosclerosis. *Blood.* (2007) 110:2907–15. doi: 10.1182/blood-2007-05-089086
- 191. Thacker SG, Berthier CC, Mattinzoli D, Rastaldi MP, Kretzler M, Kaplan MJ. The detrimental effects of IFN-α on vasculogenesis in lupus are mediated by repression of IL-1 pathways: potential role in atherogenesis and renal vascular rarefaction. *J Immunol.* (2010) 185:4457–69. doi: 10.4049/jimmunol.1001782
- 192. Buie JJ, Renaud LL, Muise-Helmericks R, Oates JC. IFN-α negatively regulates the expression of endothelial nitric oxide synthase and nitric oxide production: implications for systemic lupus erythematosus. *J Immunol.* (2017) 199:1979–88. doi: 10.4049/jimmunol.1600108
- 193. Esdaile JM, Abrahamowicz M, Grodzicky T, Li Y, Panaritis C, Du Berger R, et al. Traditional framingham risk factors fail to fully account for accelerated atherosclerosis in systemic lupus erythematosus. *Arthritis Rheum.* (2001) 44:2331–7. doi: 10.1002/1529-0131(200110)44:10<2331::AID-ART395>3.0.CO;2-I
- Malek TR. The biology of interleukin-2. Annu Rev Immunol. (2008) 26:453– 79. doi: 10.1146/annurev.immunol.26.021607.090357
- Malek TR, Bayer AL. Tolerance, not immunity, crucially depends on IL-2. Nat Rev Immunol. (2004) 4:665–74. doi: 10.1038/nri1435
- Rosenberg SA. IL-2: the first effective immunotherapy for human cancer. J Immunol. (2014) 192:5451–8. doi: 10.4049/jimmunol.1490019
- 197. Sadlack B, Merz H, Schorle H, Schimpl A, Feller AC, Horak I. Ulcerative colitis-like disease in mice with a disrupted interleukin-2 gene. *Cell.* (1993) 75:253–61. doi: 10.1016/0092-8674(93)80067-O

- 198. Sadlack B, Löhler J, Schorle H, Klebb G, Haber H, Sickel E, et al. Generalized autoimmune disease in interleukin-2-deficient mice is triggered by an uncontrolled activation and proliferation of CD4+ T cells. *Eur J Immunol.* (1995) 25:3053–9. doi: 10.1002/eji.1830251111
- 199. Abbas AK, Trotta E, Simeonov DR, Marson A, Bluestone JA. Revisiting IL-2: biology and therapeutic prospects. *Sci Immunol.* (2018) 3:eaat1482. doi: 10.1126/sciimmunol.aat1482
- 200. Koreth J, Matsuoka KI, Kim HT, McDonough SM, Bindra B, Alyea EP, et al. Interleukin-2 and regulatory T cells in graft-versus-host disease. N Engl J Med. (2011) 365:2055–66. doi: 10.1056/NEJMoa1108188
- 201. Matsuoka KI, Koreth J, Kim HT, Bascug G, McDonough S, Kawano Y, et al. Low-dose interleukin-2 therapy restores regulatory T cell homeostasis in patients with chronic graft-versus-host disease. *Sci Transl Med.* (2013) 5:179ra43. doi: 10.1126/scitranslmed.3005265
- 202. Elkind MSV, Rundek T, Sciacca RR, Ramas R, Chen HJ, Boden-Albala B, et al. Interleukin-2 levels are associated with carotid artery intima-media thickness. *Atherosclerosis.* (2005) 180:181–7. doi: 10.1016/j.atherosclerosis.2004.11.015
- 203. Ding R, Gao W, Ostrodci DH, He Z, Song Y, Ma L, et al. Effect of interleukin-2 level and genetic variants on coronary artery disease. *Inflammation*. (2013) 36:1225–31. doi: 10.1007/s10753-013-9659-2
- 204. Foks AC, Frodermann V, ter Borg M, Habets KLL, Bot I, Zhao Y, et al. Differential effects of regulatory T cells on the initiation and regression of atherosclerosis. *Atherosclerosis.* (2011) 218:53–60. doi: 10.1016/j.atherosclerosis.2011.04.029
- 205. Tomala J, Chmelova H, Mrkvan T, Rihova B, Kovar M. *In vivo* expansion of activated naive CD8+ T cells and NK cells driven by complexes of IL-2 and anti-IL-2 monoclonal antibody as novel approach of cancer immunotherapy. *J Immunol.* (2009) 183:4904–12. doi: 10.4049/jimmunol.0900284
- 206. Spangler JB, Tomala J, Luca VC, Jude KM, Dong S, Ring AM, et al. Antibodies to interleukin-2 elicit selective T cell subset potentiation through distinct conformational mechanisms. *Immunity*. (2015) 42:815–25. doi: 10.1016/j.immuni.2015.04.015
- 207. Trotta E, Bessette PH, Silveria SL, Ely LK, Jude KM, Le DT, et al. A human anti-IL-2 antibody that potentiates regulatory T cells by a structure-based mechanism. *Nat Med.* (2018) 24:1005–14. doi: 10.1038/s41591-018-0070-2
- 208. Zhao TX, Kostapanos M, Griffiths C, Arbon EL, Hubsch A, Kaloyirou F, et al. Low-dose interleukin-2 in patients with stable ischaemic heart disease and acute coronary syndromes (LILACS): protocol and study rationale for a randomised, double-blind, placebo-controlled, phase I/II clinical trial. *BMJ Open.* (2018) 8:e022452. doi: 10.1136/bmjopen-2018-022452
- 209. Kozmiński P, Halik PK, Chesori R, Gniazdowska E. Overview of dual-acting drug methotrexate in different neurological diseases, autoimmune pathologies and cancers. Int J Mol Sci. (2020) 21:3483. doi: 10.3390/ijms21103483
- 210. Farber S, Diamond LK, Mercer RD, Sylvester RFJ, Wolff JA. Temporary remissions in acute leukemia in children produced by folic acid antagonist, 4-aminopteroyl-glutamic acid (Aminopterin). *N Engl J Med.* (1948) 238:787– 93. doi: 10.1056/NEJM194806032382301
- 211. Goldin A, Venditti JM, Humphreys SR, Dennis D, Mantel N, Greenhouse SW. a quantitative comparison of the antileukemic effectiveness of two folic acid antagonists in mice. *JNCI J Natl Cancer Inst.* (1955) 15:1657–64.
- WHO. WHO Model List of Essential Medicines 22nd List, 2021. (2021). Available online at: https://www.who.int/publications/i/item/WHO-MHP-HPS-EML-2021.02 (accessed December 16, 2021).
- Cronstein BN, Aune TM. Methotrexate and its mechanisms of action in inflammatory arthritis. Nat Rev Rheumatol. (2020) 16:145–54. doi: 10.1038/s41584-020-0373-9
- 214. Weinblatt ME, Coblyn JS, Fox DA, Fraser PA, Holdsworth DE, Glass DN, et al. Efficacy of low-dose methotrexate in rheumatoid arthritis. *N Engl J Med.* (2010) 16:287–95. doi: 10.1056/NEJM198503283 121303
- Giannini EH, Brewer EJ, Kuzmina N, Shaikov A, Maximov A, Vorontsov I, et al. Methotrexate in resistant juvenile rheumatoid arthritis. N Engl J Med. (2010) 326:1043–9. doi: 10.1056/NEJM199204163261602
- Choi HK, Hernán MA, Seeger JD, Robins JM, Wolfe F. Methotrexate and mortality in patients with rheumatoid arthritis: a prospective study. *Lancet.* (2002) 359:1173–7. doi: 10.1016/S0140-6736(02)08213-2

- 217. Micha R, Imamura F, Wyler Von Ballmoos M, Solomon DH, Hernán MA, Ridker PM, et al. Systematic review and meta-analysis of methotrexate use and risk of cardiovascular disease. *Am J Cardiol.* (2011) 108:1362–70. doi: 10.1016/j.amjcard.2011.06.054
- Lutgens E, Atzler D, Döring Y, Duchene J, Steffens S, Weber C. Immunotherapy for cardiovascular disease. *Eur Heart J.* (2019) 40:3937–46. doi: 10.1093/eurheartj/ehz283
- Tardif J-C, Kouz S, Waters DD, Bertrand OF, Diaz R, Maggioni AP, et al. Efficacy and safety of low-dose colchicine after myocardial infarction. *N Engl J Med.* (2019) 381:2497–505. doi: 10.1056/NEJMoa1912388
- Nidorf SM, Fiolet ATL, Mosterd A, Eikelboom JW, Schut A, Opstal TSJ, et al. Colchicine in patients with chronic coronary disease. N Engl J Med. (2020) 383:1838–47. doi: 10.1056/NEJMoa2021372
- 221. Imazio M, Andreis A, Brucato A, Adler Y, De Ferrari GM. Colchicine for acute and chronic coronary syndromes. *Heart.* (2020) 106:1555–60. doi: 10.1136/heartjnl-2020-317108
- Raal FJ, Kallend D, Ray KK, Turner T, Koenig W, Wright RS, et al. Inclisiran for the treatment of heterozygous familial hypercholesterolemia. N Engl J Med. (2020) 382:1520–30. doi: 10.1056/NEJMoa1913805
- Ray KK, Wright RS, Kallend D, Koenig W, Leiter LA, Raal FJ, et al. Two phase 3 trials of inclisiran in patients with elevated LDL cholesterol. *N Engl J Med.* (2020) 382:1507–19. doi: 10.1056/NEJMoa1912387
- 224. Ridker PM, MacFadyen JG, Thuren T, Everett B, Libby P, Glynn R, et al. Effect of interleukin-1β inhibition with canakinumab on incident lung cancer in patients with atherosclerosis: exploratory results from a randomised, double-blind, placebo-controlled trial. *Lancet.* (2017) 390:1833–42. doi: 10.1016/S0140-6736(17)32247-X
- 225. Voronov E, Shouval DS, Krelin Y, Cagnano E, Benharroch D, Iwakura Y, et al. IL-1 is required for tumor invasiveness and angiogenesis. *Proc Natl Acad Sci.* (2003) 100:2645–50. doi: 10.1073/pnas.0437939100
- 226. Liu X, Bao X, Hu M, Chang H, Jiao M, Cheng J, et al. Inhibition of PCSK9 potentiates immune checkpoint therapy for cancer. *Nature*. (2020) 588:693–8. doi: 10.1038/s41586-020-2911-7
- 227. Quang CT, Zaniboni B, Humeau R, Lengliné E, Dourthe ME, Ganesan R, et al. Preclinical efficacy of humanized, non–FcyR-binding anti-CD3 antibodies in T-cell acute lymphoblastic leukemia. *Blood.* (2020) 136:1298–302. doi: 10.1182/blood.2019003801
- Kuo MC, Chang SJ, Hsieh MC. Colchicine significantly reduces incident cancer in gout male patients: a 12-year cohort study. *Medicine*. (2015) 94:e1570. doi: 10.1097/MD.00000000001570
- Fakih M, Replogle T, Lehr JE, Pienta KJ, Yagoda A. Inhibition of prostate cancer growth by estramustine and colchicine. *Prostate*. (1995) 26:310–15. doi: 10.1002/pros.2990260606
- Cho JH, Joo YH, Shin EY, Park EJ, Kim SM. Anticancer effects of colchicine on hypopharyngeal cancer. *Anticancer Res.* (2017) 37:6269 LP-80. doi: 10.21873/anticanres.12078
- Lin ZY, Wu CC, Chuang YH, Chuang WL. Anti-cancer mechanisms of clinically acceptable colchicine concentrations on hepatocellular carcinoma. *Life Sci.* (2013) 93:323–8. doi: 10.1016/j.lfs.2013.07.002
- 232. Craig DH, Owen CR, Conway WC, Walsh MF, Downey C, Basson MD. Colchicine inhibits pressure-induced tumor cell implantation within surgical wounds and enhances tumor-free survival in mice. J Clin Invest. (2008) 118:3170–80. doi: 10.1172/JCI34279
- 233. Bent R, Moll L, Grabbe S, Bros M. Interleukin-1 beta—a friend or foe in malignancies? *Int J Mol Sci.* (2018) 19:2155. doi: 10.3390/ijms19082155
- Greten FR, Grivennikov SI. Inflammation and cancer: triggers, mechanisms, and consequences. *Immunity*. (2019) 51:27–41. doi: 10.1016/j.immuni.2019.06.025
- 235. Barlesi F, Cho BC, Castro G, Marchi PD, Felip E, Goto Y, et al. The CANOPY program: Canakinumab in patients (pts) with nonsmall cell lung cancer (NSCLC). J Clin Oncol. (2019) 37:TPS9124. doi: 10.1200/JCO.2019.37.15_SUPPL.TPS9124
- 236. Garrido P, Pujol J-L, Kim ES, Lee JM, Tsuboi M, Gómez-Rueda A, et al. Canakinumab with and without pembrolizumab in patients with resectable non-small-cell lung cancer: CANOPY-N study design. *Fut Oncol.* (2021) 17:1459–72. doi: 10.2217/fon-2020-1098
- 237. Novartis Provides Update on Phase III Study Evaluating Canakinumab (ACZ885) as Second or Third-Line Treatment in Combination With

Chemotherapy in Non-Small Cell Lung Cancer. Novartis. Available online at: https://www.novartis.com/news/media-releases/novartis-providesupdate-phase-iii-study-evaluating-canakinumab-acz885-second-or-thirdline-treatment-combination-chemotherapy-non-small-cell-lung-cancer (accessed October 13, 2021).

- Dasgeb B, Kornreich D, McGuinn K, Okon L, Brownell I, Sackett DL. Colchicine: an ancient drug with novel applications. *Br J Dermatol.* (2018) 178:350–6. doi: 10.1111/bjd.15896
- 239. FitzGerald JD, Dalbeth N, Mikuls T, Brignardello-Petersen R, Guyatt G, Abeles AM, et al. 2020 American College of Rheumatology guideline for the management of gout. *Arthritis Care Res.* (2020) 72:744–60. doi: 10.1002/acr.24180
- 240. Tong DC, Quinn S, Nasis A, Hiew C, Roberts-Thomson P, Adams H, et al. Colchicine in patients with acute coronary syndrome. *Circulation*. (2020) 142:1890–900. doi: 10.1161/CIRCULATIONAHA.120.050771
- 241. Fiolet ATL, Collaboration the CCT, Opstal TSJ, Collaboration the CCT, Mosterd A, Collaboration the CCT, et al. Efficacy and safety of lowdose colchicine in patients with coronary disease: a systematic review and meta-analysis of randomized trials. *Eur Heart J.* (2021) 42:2765–75. doi: 10.1093/eurheartj/ehab115
- 242. Kofler T, Kurmann R, Lehnick D, Cioffi GM, Chandran S, Attinger-Toller A, et al. Colchicine in patients with coronary artery disease: a systematic review and meta-analysis of randomized trials. *J Am Heart Assoc.* (2021) 10:21198. doi: 10.1161/JAHA.121.021198
- Kumar A, Sharma PR, Mondhe DM. Potential anticancer role of colchicinebased derivatives: an overview. *Anticancer Drugs*. (2016) 28:250–62. doi: 10.1097/CAD.00000000000464
- 244. Wen C-C, Chen H-M, Chen S-S, Huang L-T, Chang W-T, Wei W-C, et al. Specific microtubule-depolymerizing agents augment efficacy of dendritic cell-based cancer vaccines. *J Biomed Sci.* (2011) 18:1–15. doi: 10.1186/1423-0127-18-44
- Hess CN, Wang CCL, Hiatt WR. PCSK9 Inhibitors: mechanisms of action, metabolic effects, and clinical outcomes. *Annu Rev Med.* (2018) 69:133–45. doi: 10.1146/annurev-med-042716-091351
- 246. Mach F, Baigent C, Catapano AL, Koskinas KC, Casula M, Badimon L, et al. 2019 ESC/EAS Guidelines for the management of dyslipidaemias: lipid modification to reduce cardiovascular risk. *Eur Heart J.* (2020) 41:111–88. doi: 10.15829/1560-4071-2020-3826
- 247. Kuzu OF, Noory MA, Robertson GP. The role of cholesterol in cancer. *Cancer Res.* (2016) 76:2063–70. doi: 10.1158/0008-5472.CAN-15-2613
- 248. Shi M, Zheng H, Nie B, Gong W, Cui X. Statin use and risk of liver cancer: an update meta-analysis. *BMJ Open.* (2014) 4:e005399. doi: 10.1136/bmjopen-2014-005399
- 249. Pradelli D, Soranna D, Zambon A, Catapano A, Mancia G, Vecchia C La, et al. Statins use and the risk of all and subtype hematological malignancies: a meta-analysis of observational studies. *Cancer Med.* (2015) 4:770–80. doi: 10.1002/cam4.411
- 250. Ying Y, Ling Y, Yang L, Huang H, Hu X, Zhao C, et al. Prognostic significance of statin use in colorectal cancer. *Medicine*. (2015) 94:e908. doi: 10.1097/MD.000000000000908
- 251. Dale KM, Coleman CI, Henyan NN, Kluger J, White CM. Statins and cancer risk: a meta-analysis. *JAMA*. (2006) 295:74–80. doi: 10.1001/jama.295.1.74
- 252. Tan P, Wei S, Tang Z, Gao L, Zhang C, Nie P, et al. LDL-lowering therapy and the risk of prostate cancer: a meta-analysis of 6 randomized controlled trials and 36 observational studies. *Sci Rep.* (2016) 6:1–9. doi: 10.1038/srep24521
- 253. Momtazi-Borojeni AA, Jaafari MR, Badiee A, Sahebkar A. Longterm generation of antiPCSK9 antibody using a nanoliposomebased vaccine delivery system. *Atherosclerosis.* (2019) 283:69–78. doi: 10.1016/j.atherosclerosis.2019.02.001
- 254. Kuhn C, Weiner HL. Therapeutic anti-CD3 monoclonal antibodies: from bench to bedside. *Immunotherapy*. (2016) 8:889–906. doi: 10.2217/imt-2016-0049
- 255. Belghith M, Bluestone JA, Barriot S, Mégret J, Bach JF, Chatenoud L. TGFβ-dependent mechanisms mediate restoration of self-tolerance induced by antibodies to CD3 in overt autoimmune diabetes. *Nat Med.* (2003) 9:1202–8. doi: 10.1038/nm924
- 256. Perruche S, Zhang P, Liu Y, Saas P, Bluestone JA, Chen WJ. CD3-specific antibody-induced immune tolerance involves transforming growth factor-β

from phagocytes digesting apoptotic T cells. Nat Med. (2008) 14:528-35. doi: 10.1038/nm1749

- 257. Weinshenker BG, Bass B, Karlik S, Ebers GC, Rice GPA. An open trial of OKT3 in patients with multiple sclerosis. *Neurology*. (1991) 41:1047. doi: 10.1212/WNL.41.7.1047
- Keymeulen B, Vandemeulebroucke E, Ziegler AG, Mathieu C, Kaufman L, Hale G, et al. Insulin needs after CD3-antibody therapy in new-onset type 1 diabetes. N Engl J Med. (2005) 352:2598–608. doi: 10.1056/NEJMoa043980
- 259. Hagopian W, Ferry RJ, Sherry N, Carlin D, Bonvini E, Johnson S, et al. Teplizumab preserves C-peptide in recent-onset type 1 diabetes: Two-year results from the randomized, placebo-controlled Protégé trial. *Diabetes*. (2013) 62:3901–8. doi: 10.2337/db13-0236
- 260. Herold KC, Gitelman SE, Ehlers MR, Gottlieb PA, Greenbaum CJ, Hagopian W, et al. Teplizumab (Anti-CD3 mAb) treatment preserves C-peptide responses in patients with new-onset type 1 diabetes in a randomized controlled trial: metabolic and immunologic features at baseline identify a subgroup of responders. *Diabetes*. (2013) 62:3766–74. doi: 10.2337/db13-0345
- 261. Herold KC, Bundy BN, Long SA, Bluestone JA, DiMeglio LA, Dufort MJ, et al. An anti-CD3 antibody, teplizumab, in relatives at risk for type 1 diabetes. *N Engl J Med.* (2019) 381:603–13. doi: 10.1056/NEJMoa1902226
- 262. Plevy S, Salzberg B, Van Assche G, Regueiro M, Hommes D, Sandborn W, et al. A phase I study of visilizumab, a humanized anti-CD3 monoclonal antibody, in severe steroid-refractory ulcerative colitis. *Gastroenterology*. (2007) 133:1414–22. doi: 10.1053/j.gastro.2007.08.035
- 263. Kita T, Yamashita T, Sasaki N, Kasahara K, Sasaki Y, Yodoi K, et al. Regression of atherosclerosis with anti-CD3 antibody via augmenting a regulatory T-cell response in mice. *Cardiovasc Res.* (2014) 102:107–17. doi: 10.1093/cvr/cvu002
- 264. Sasaki N, Yamashita T, Takeda M, Shinohara M, Nakajima K, Tawa H, et al. Oral anti-CD3 antibody treatment induces regulatory t cells and inhibits the development of atherosclerosis in mice. *Circulation.* (2009) 120:1996–2005. doi: 10.1161/CIRCULATIONAHA.109.863431
- 265. Steffens S, Burger F, Pelli G, Dean Y, Elson G, Kosco-Vilbois M, et al. Short-term treatment with anti-CD3 antibody reduces the development and progression of atherosclerosis in mice. *Circulation*. (2006) 114:1977–84. doi: 10.1161/CIRCULATIONAHA.106.627430
- 266. Gramatzki M, Burger R, Strobel G, Trautmann U, Bartram CR, Helm G, et al. Therapy with OKT3 monoclonal antibody in refractory T cell acute lymphoblastic leukemia induces interleukin-2 responsiveness. *Leukemia*. (1995) 9:382–90.
- 267. Sosman JA, Kefer C, Fisher RI, Jacobs CD, Pumfery P, Ellis TM. A phase IA/IB trial of anti-CD3 murine monoclonal antibody plus lowdose continuous-infusion interleukin-2 in advanced cancer patients. J Immunother. (1995) 17:171–80. doi: 10.1097/00002371-199504000-00006
- 268. Sosman JA, Weiss GR, Margolin KA, Aronson FR, Sznol M, Atkins MB, et al. Phase IB clinical trial of anti-CD3 followed by high-dose bolus interleukin-2 in patients with metastatic melanoma and advanced renal cell carcinoma: clinical and immunologic effects. J Clin Oncol. (1993) 11:1496– 505. doi: 10.1200/JCO.1993.11.8.1496
- 269. Feins S, Kong W, Williams EF, Milone MC, Fraietta JA. An introduction to chimeric antigen receptor (CAR) T-cell immunotherapy for human cancer. *Am J Hematol.* (2019) 94:S3–9. doi: 10.1002/ajh.25418
- 270. Schuster SJ, Svoboda J, Chong EA, Nasta SD, Mato AR, Anak Ö, et al. Chimeric antigen receptor T cells in refractory B-cell lymphomas. N Engl J Med. (2017) 377:2545–54. doi: 10.1056/NEJMoa1708566
- Neelapu SS, Locke FL, Bartlett NL, Lekakis LJ, Miklos DB, Jacobson CA, et al. Axicabtagene ciloleucel CAR T-cell therapy in refractory large B-cell lymphoma. *N Engl J Med.* (2017) 377:2531–44. doi: 10.1056/NEJMoa1707447
- 272. Wang M, Munoz J, Goy A, Locke FL, Jacobson CA, Hill BT, et al. KTE-X19 CAR T-cell therapy in relapsed or refractory mantle-cell lymphoma. *N Engl J Med.* (2020) 382:1331–42. doi: 10.1056/NEJMoa1914347
- 273. Abramson JS, Palomba ML, Gordon LI, Lunning MA, Wang M, Arnason J, et al. Lisocabtagene maraleucel for patients with relapsed or refractory large B-cell lymphomas (TRANSCEND NHL 001): a multicentre seamless design study. *Lancet.* (2020) 396:839–52. doi: 10.1016/S0140-6736(20) 31366-0

- Munshi NC, Anderson LD, Shah N, Madduri D, Berdeja J, Lonial S, et al. Idecabtagene vicleucel in relapsed and refractory multiple myeloma. *N Engl J Med.* (2021) 384:705–16. doi: 10.1056/NEJMoa2024850
- 275. Neelapu SS, Tummala S, Kebriaei P, Wierda W, Gutierrez C, Locke FL, et al., Lin Y, Jain N, Daver N, et al. Chimeric antigen receptor T-cell therapy assessment and management of toxicities. *Nat Rev Clin Oncol.* (2017) 15:47–62. doi: 10.1038/nrclinonc.2017.148
- 276. Alvi RM, Frigault MJ, Fradley MG, Jain MD, Mahmood SS, Awadalla M, et al. Cardiovascular events among adults treated with chimeric antigen receptor T-cells (CAR-T). J Am Coll Cardiol. (2019) 74:3099–108. doi: 10.1016/j.jacc.2019.10.038
- 277. Bayati F, Mohammadi M, Valadi M, Jamshidi S, Foma AM, Sharif-Paghaleh E. The therapeutic potential of regulatory T cells: challenges and opportunities. *Front Immunol.* (2021) 11:3455. doi: 10.3389/fimmu.2020.585819
- 278. Marek-Trzonkowska N, Myśliwiec M, Dobyszuk A, Grabowska M, Derkowska I, Juścińska J, et al. Therapy of type 1 diabetes with CD4+CD25highCD127-regulatory T cells prolongs survival of pancreatic islets — results of one year follow-up. *Clin Immunol.* (2014) 153:23–30. doi: 10.1016/j.clim.2014.03.016
- 279. Bluestone JA, Buckner JH, Fitch M, Gitelman SE, Gupta S, Hellerstein MK, et al. Type 1 diabetes immunotherapy using polyclonal regulatory T cells. *Sci Transl Med.* (2015) 7:315ra189. doi: 10.1126/scitranslmed.aad4134
- 280. Brunstein CG, Miller JS, Cao Q, McKenna DH, Hippen KL, Curtsinger J, et al. Infusion of *ex vivo* expanded T regulatory cells in adults transplanted with umbilical cord blood: safety profile and detection kinetics. *Blood.* (2011) 117:1061–70. doi: 10.1182/blood-2010-07-293795
- 281. Brunstein CG, Miller JS, McKenna DH, Hippen KL, DeFor TE, Sumstad D, et al. Umbilical cord blood-derived T regulatory cells to prevent GVHD: kinetics, toxicity profile, and clinical effect. *Blood.* (2016) 127:1044–51. doi: 10.1182/blood-2015-06-653667
- 282. Mathew JM, H-Voss J, LeFever A, Konieczna I, Stratton C, He J, et al. A phase I clinical trial with *ex vivo* expanded recipient regulatory T cells in living donor kidney transplants. *Sci Rep.* (2018) 8:1–12. doi: 10.1038/s41598-018-25574-7
- Desreumaux P, Foussat A, Allez M, Beaugerie L, Hébuterne X, Bouhnik Y, et al. Safety and efficacy of antigen-specific regulatory T-cell therapy for patients with refractory Crohn's disease. *Gastroenterology.* (2012) 143:1207–17.e2. doi: 10.1053/j.gastro.2012.07.116
- Ait-Oufella H, Salomon BL, Potteaux S, Robertson A-KL, Gourdy P, Zoll J, et al. Natural regulatory T cells control the development of atherosclerosis in mice. *Nat Med.* (2006) 12:178–80. doi: 10.1038/nm1343
- 285. Yang K, Li D, Luo M, Hu Y. Generation of HSP60-specific regulatory T cell and effect on atherosclerosis. *Cell Immunol.* (2006) 243:90–95. doi: 10.1016/j.cellimm.2007.01.002
- Palinski W, Miller E, Witztum JL. Immunization of low density lipoprotein (LDL) receptor-deficient rabbits with homologous malondialdehydemodified LDL reduces atherogenesis. *Proc Natl Acad Sci USA*. (1995) 92:821– 5. doi: 10.1073/pnas.92.3.821
- 287. Klingenberg R, Lebens M, Hermansson A, Fredrikson GN, Strodthoff D, Rudling M, et al. Intranasal immunization with an apolipoprotein B-100 fusion protein induces antigen-specific regulatory T cells and reduces atherosclerosis. *Arterioscler Thromb Vasc Biol.* (2010) 30:946–52. doi: 10.1161/ATVBAHA.109.202671
- 288. Herbin O, Ait-Oufella H, Yu W, Fredrikson GN, Aubier B, Perez N, et al. Regulatory T-cell response to apolipoprotein B100-derived peptides reduces the development and progression of atherosclerosis in mice. *Arterioscler Thromb Vasc Biol.* (2012) 32:605–12. doi: 10.1161/ATVBAHA.111.242800
- 289. Gisterå A, Klement ML, Polyzos KA, Mailer RKW, Duhlin A, Karlsson MCI, et al. Low-density lipoprotein-reactive T cells regulate plasma cholesterol levels and development of atherosclerosis in humanized hypercholesterolemic mice. *Circulation*. (2018) 138:2513–26. doi: 10.1161/CIRCULATIONAHA.118.034076
- 290. Krienke C, Kolb L, Diken E, Streuber M, Kirchhoff S, Bukur T, et al. A noninflammatory mRNA vaccine for treatment of experimental autoimmune encephalomyelitis. *Science.* (2021) 371:145–53. doi: 10.1126/science.aay3638

- 291. Saxena M, van der Burg SH, Melief CJM, Bhardwaj N. Therapeutic cancer vaccines. Nat Rev Cancer. (2021) 21:360–78. doi: 10.1038/s41568-021-00346-0
- Sahin U, Türeci Ö. Personalized vaccines for cancer immunotherapy. *Science*. (2018) 359:1355–60. doi: 10.1126/science.aar7112
- 293. Carreno BM, Magrini V, Becker-Hapak M, Kaabinejadian S, Hundal J, Petti AA, et al. A dendritic cell vaccine increases the breadth and diversity of melanoma neoantigen-specific T cells. *Science*. (2015) 348:803–8. doi: 10.1126/science.aaa3828

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