



Low α 2-Plasmin Inhibitor Antigen Levels on Admission Are Associated With More Severe Stroke and Unfavorable Outcomes in Acute Ischemic Stroke Patients Treated With Intravenous Thrombolysis

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Background: Intravenous administration of recombinant tissue plasminogen activator (rt-PA) fails to succeed in a subset of acute ischemic stroke (AIS) patients, while in approximately 6–8% of cases intracerebral hemorrhage (ICH) occurs as side effect.

Objective: Here, we aimed to investigate α 2-plasmin inhibitor (α 2-PI) levels during thrombolysis and to find out whether they predict therapy outcomes in AIS patients.

Patients/Methods: In this prospective, observational study, blood samples of 421 AIS patients, all undergoing i.v. thrombolysis by rt-PA within 4.5 h of their symptom onset, were taken before and 24 h after thrombolysis. In a subset of patients ($n = 131$), blood was also obtained immediately post-lysis. α 2-PI activity and antigen levels were measured by chromogenic assay and an in-house ELISA detecting all forms of α 2-PI. α 2-PI Arg6Trp polymorphism was identified in all patients. Stroke severity was determined by NIHSS on admission and day 7. Therapy-associated ICH was classified according to ECASSII. Long-term outcomes were defined at 3 months post-event by the modified Rankin Scale (mRS).

Results: Median α 2-PI activity and antigen levels showed a significant drop immediately post-lysis and increased to subnormal levels at 24 h post-event. Admission α 2-PI levels showed a significant negative stepwise association with stroke severity. Patients with favorable long-term outcomes (mRS 0–1) had significantly higher admission α 2-PI antigen levels (median:61.6 [IQR:55.9–70.5] mg/L) as compared to patients with poor outcomes (mRS 2–5: median:59.7 [IQR:54.5–69.1] and mRS 6: median:56.0 [IQR:48.5–61.0] mg/L, $p < 0.001$). In a Kaplan–Meier survival analysis, patients with an α 2-PI antigen in the highest quartile on admission showed significantly better long-term survival as

compared to those with α 2-PI antigen in the lowest quartile (HR: 4.54; 95%CI:1.92–10.8, $p < 0.001$); however, in a multivariate analysis, a low admission α 2-PI antigen did not prove to be an independent risk factor of poor long-term outcomes. In patients with therapy-related ICH ($n = 34$), admission α 2-PI antigen levels were significantly, but only marginally, lower as compared to those without hemorrhage.

Conclusions: Low α 2-PI antigen levels on admission were associated with more severe strokes and poor long-term outcomes in this cohort. Our results suggest that in case of more severe strokes, α 2-PI may be involved in the limited efficacy of rt-PA thrombolysis.

Keywords: α 2-plasmin inhibitor, acute ischemic stroke, fibrinolysis, intracerebral hemorrhage (ICH), outcome, recombinant tissue plasminogen activator (rt-PA), thrombolysis

INTRODUCTION

Stroke is a leading cause of death and the major cause of adult disability worldwide, representing a substantial socio-economic burden (1). Eighty percent of all strokes are of ischemic origin, when the patient's only chance of recovery depends on the successful opening of the occluded vessel within a relatively short time frame. As of today, two evidence-based therapies are available for this purpose. Currently, the cornerstone pharmacological therapy for the restoration of blood flow is intravenous recombinant tissue-type plasminogen activator (rt-PA), which must be administered within 4.5 h of symptom onset (2–4). Endovascular therapy known as mechanical thrombectomy is now the standard care for patients in case of large vessel occlusions (LVO) who present within 24 h of stroke symptom onset (5, 6). However, only ~20% of stroke patients have LVO, and delivering treatment to such patients within a rapid time frame is challenging, as the procedure can only be performed in highly specialized centers. Consequently, intravenous thrombolysis using rt-PA remains the mainstay therapy of AIS (2–4). Despite the unquestionable benefit of rt-PA in AIS treatment, this therapy is not a remedy for all. Thrombolysis is only effective in ~30–40% of patients, and its efficacy decreases greatly in case of LVO (7). Besides recanalization failure, about 6–8% of patients develop intracranial hemorrhage (ICH) as side effect, despite taking all precautionary steps to minimize bleeding risk (8). As of today, inefficacy of treatment or therapy-associated ICH cannot be foreseen at the initiation of thrombolysis and their occurrence remains unexplained (9). As the fibrinolytic system provides the very basis of the mechanism of thrombolysis, it can be surmised that individual factors altering lysis susceptibility are likely to be key contributors to recanalization failure and bleeding complications. Surprisingly, our knowledge to date regarding this aspect is limited to a few reports (10–15).

The process of fibrinolysis is tightly regulated on multiple levels by activators, inhibitors, cofactors, and receptors (12, 16, 17). The action of rt-PA during thrombolysis is to generate plasmin from its precursor, plasminogen. The inhibition of rt-PA mainly occurs by the action of plasminogen activator inhibitor 1 (PAI-1). The effect of plasmin itself is regulated by α 2-plasmin inhibitor (α 2-PI) that binds rapidly and irreversibly to plasmin,

forming a stable 1:1 complex, in which the activity of the enzyme is effectively neutralized (18–20). During clot formation, when factor XIII (FXIII) becomes activated, it cross-links α 2-PI into the fibrin clot, where it can directly and effectively inhibit plasmin (16, 17, 21, 22). Additional regulatory proteins also exist that influence the ability of plasminogen to bind to fibrin and therefore hinder fibrinolysis (i.e., thrombin activatable fibrinolysis inhibitor: TAFI). We and others in previous clinical studies have looked at the relation of specific elements of fibrinolysis (e.g., PAI-1, plasminogen, TAFI, and FXIII) and AIS thrombolysis outcome, but these studies could not identify the essential factors or mechanisms driving recanalization failure or post-lysis hemorrhage based on admission blood samples (11, 12, 14, 23, 24). Interestingly, although α 2-PI is a key regulator of fibrinolysis, a potential association between α 2-PI levels and thrombolysis outcomes has not been investigated in a relatively large AIS patient cohort, as yet. Moreover, changes in α 2-PI levels during AIS thrombolysis and their effect on outcomes have not been tested in a cohort of more than 100 AIS patients as yet, despite the fact that α 2-PI activity is expected to decline rapidly during the administration of rt-PA (25, 26).

In the circulation, α 2-PI undergoes both amino-terminal (N-terminal) and carboxy-terminal (C-terminal) proteolytic modifications, leading to a variety of circulating α 2-PI molecules with modified activities (20). Approximately 70% of circulating α 2-PI is N-terminally cleaved by antiplasmin cleaving enzyme (APCE, also known as soluble fibroblast activation protein: sFAP) (27, 28). The N-terminal cleavage has been shown to affect the extent of α 2-PI cross-linking to fibrin by activated FXIII (FXIIIa) to a great extent (17, 21, 29). The N-terminal heterogeneity of α 2-PI is affected by a common polymorphism of the SERPINF2 gene (p.Arg6Trp, rs2070863) (30). The C-terminus of α 2-PI is also posttranslationally modified, although the enzyme responsible for the cleavage has not been identified as yet. The uncleaved, longer form that can bind plasminogen (PB- α 2-PI) constitutes about 65% of circulating α 2-PI, while the other form fails to bind plasminogen (NPB- α 2-PI) (31). Importantly, it has been shown that cross-linking of α 2-PI to fibrin by FXIIIa primarily involves PB- α 2-PI and C-terminal cleavage of the circulating protein regulates α 2-PI activity (20). It has been known for decades that congenital or acquired α 2-PI deficiency leads to a bleeding disorder, while increased levels of α 2-PI have been more

recently shown to be associated with the increased risk of AIS and venous thrombosis (32). Based on this, one can surmise that α 2-PI could play an important regulatory role in the outcome of AIS thrombolysis by rt-PA.

Here, we aimed to test α 2-PI activity and antigen levels during the course of intravenous thrombolysis and a potential association between α 2-PI levels, stroke severity, and treatment outcomes. We also aimed to test the effect of α 2-PI Arg6Trp polymorphism on α 2-PI levels and on thrombolysis outcomes. We report that low α 2-PI antigen levels on admission were associated with more severe strokes and poor long-term outcomes in the tested cohort of 421 AIS patients. Our results suggest that in case of more severe strokes, α 2-PI may be involved in the limited efficacy of rt-PA thrombolysis. α 2-PI Arg6Trp polymorphism had no effect on α 2-PI activity or antigen levels and showed no association with stroke severity, etiology, or outcomes.

MATERIALS AND METHODS

Patients

In this prospective observational study, adult AIS patients, all within 4.5 h of stroke onset and eligible for intravenous thrombolysis, were enrolled (Department of Neurology, Faculty of Medicine, University of Debrecen, Hungary). Patient enrollment was initiated in March 2011 and was completed in April 2019. Intravenous thrombolytic therapy was applied according to the 2008 European Stroke Organization (ESO) guidelines for rt-PA (Alteplase, Boehringer Ingelheim, Germany) administration (33). The inclusion and exclusion criteria of patients were identical to standard criteria of thrombolysis as described in the ESO 2008 guideline. None of the patients included in the cohort were eligible for mechanical thrombectomy: thrombectomy was either unavailable at the time of enrollment, or the patient was unsuitable to perform the procedure. The presence of AIS was diagnosed based on clinical symptoms and imaging using non-contrast computerized tomography (CT) scan and CT angiography (CTA). CT images taken on admission and 24 h post-lysis were analyzed simultaneously by three independent investigators, and the Alberta Stroke Program Early CT Scores (ASPECTS) were calculated (34). For each patient, the time of symptom onset, demographic and clinical characteristics (age, sex, BMI, previous medications, history of cerebrovascular and cardiovascular diseases, and cerebrovascular risk factors including smoking) were registered on admission. Stroke severity was determined by the National Institutes of Health Stroke Scale (NIHSS) on admission and day 7 after therapy (35). Trial of ORG 10172 in Acute Stroke Treatment (TOAST) criteria was used to identify the etiology of stroke (36). Patients were followed, and long-term functional outcomes were determined at 3 months after the stroke event using the modified Rankin Scale (mRS) (37).

The following outcomes and safety endpoint were registered in this study: 1. Short-term outcome at 7 days post-event: a decrease in NIHSS score by at least 4 points or to 0 was defined as favorable outcome (neurologic improvement), while an increase in NIHSS score by at least 4 points was defined

as unfavorable outcome (3, 37). 2. Long-term outcome at 90 days post-event: mRS 0–1 was defined as favorable long-term outcome. 3. Therapy-associated intracerebral hemorrhage (ICH): symptomatic (SICH) or asymptomatic (aSICH) using the European Cooperative Acute Stroke Study (ECASS) II criteria, as observed on CT scans at 24 h post-lysis (38).

Informed Consent

The study design was in accordance with the guiding principles of the Declaration of Helsinki and was approved by the Institutional Ethics Committee of the University of Debrecen and the Ethics Committee of the National Medical Research Council. All patients or their relatives provided written informed consent.

Blood Sampling and Laboratory Measurements

Peripheral blood samples were taken from all patients on admission, before the initiation of rt-PA infusion, and at 24 h post-lysis. In a subset of patients ($n = 131$), blood was also obtained immediately after administering full dose of rt-PA. Routine laboratory tests (ions, glucose level, renal and liver function tests, high-sensitivity C-reactive protein measurement, and complete blood count) were measured by standard laboratory methods (Roche Diagnostics, Mannheim, Germany, and Sysmex Europe GmbH, Hamburg, Germany). For studying specific hemostasis tests, blood samples were collected to vacutainer tubes containing 0.109 M sodium citrate (Becton Dickinson, Franklin Lane, NJ) and were processed immediately (centrifugation twice at 1,500 g, room temperature for 15 min). Screening tests of coagulation (prothrombin time, activated partial thromboplastin time, and thrombin time) and the determination of fibrinogen levels according to Clauss were performed on a BCS coagulometer using standard methods (Siemens Healthcare Diagnostic Products, Marburg, Germany). For the measurements of α 2-PI activity and antigen levels, aliquots of citrated plasma were labeled with a unique code and stored at -80°C until analysis in batches. The buffy coat of blood samples was used to extract genomic DNA based on standard protocols (QIAamp DNA Blood Mini Kit, Qiagen, Hilden, Germany). The α 2-PI Arg6Trp (rs2070863) polymorphism was identified by real-time PCR using fluorescence resonance energy transfer detection and melting curve analysis on a LightCycler® 480 instrument as described previously (Roche Diagnostics GmbH, Mannheim, Germany; primers are available from the authors upon request) (10). α 2-PI activity and antigen level assays as well as the determination of α 2-PI Arg6Trp genotype were carried out by investigators blinded to patient identification and clinical data.

α 2-Plasmin Inhibitor Activity and Antigen Assays

Functional α 2-PI activity was measured from stored plasma samples using the Berichrom α 2-PI activity chromogenic assay on a BCS coagulometer following the manufacturer's instructions (Siemens Healthcare Diagnostic Products, Marburg, Germany). The assay principle is the following: patient plasma containing α 2-PI is incubated with the reagent that contains plasmin

in excess. Plasmin becomes rapidly inactivated by α 2-PI, and residual plasmin activity is measured by an amidolytic assay based on the cleavage of the plasmin-specific chromogenic substrate (D-norvalil-cyclohexylalanil-lysyl-nitroanilid). α 2-PI activity inversely correlates with the changes in absorption at 405 nm.

Total α 2-PI antigen levels were measured by an in-house ELISA as previously described (39). This assay detects all forms of α 2-PI and is not influenced by the presence of plasmin-antiplasmin complexes (reference range of plasma α 2-PI antigen levels: 48–85 mg/L).

Statistical Analysis

Statistical analysis was performed using the Statistical Package for Social Sciences (SPSS, Version 26.0, Chicago, IL) and GraphPad Prism 8.0 (GraphPad Prism Inc., La Jolla, CA). Shapiro–Wilk test was used to assess the normality of the data. Mann–Whitney U-test was performed for two-group analyses. In case of paired data, Wilcoxon signed-rank test was applied. Depending on the normality of data, ANOVA with Bonferroni *post-hoc* test or Kruskal–Wallis analysis with Dunn–Bonferroni *post-hoc* analysis was applied for multiple group comparisons. Spearman's correlation coefficient was used to determine the strength of correlation between continuous variables. Differences between categorical variables were assessed by χ^2 test or by Fisher's exact where appropriate. The Kaplan–Meier method was applied to plot survival vs. non-survival of patients based on their α 2-PI levels. Survival curves were compared using the log-rank test. Binary backward logistic regression models were used to determine independent predictors of mortality and long-term functional outcomes. Adjustments of the models were based on the results of univariate statistical analyses of baseline characteristics between groups (Mann–Whitney U-test, χ^2 test, or Fisher's exact), previous literature, and methodological principles. The results of the logistic regression analysis were expressed as odds ratio (OR) and 95% confidence interval (CI). A $p < 0.05$ was considered statistically significant.

RESULTS

A total of 421 AIS patients receiving intravenous thrombolysis were included in the study. Baseline characteristics of patients and stroke outcomes are shown in **Table 1**. Median age of the cohort was 68 (IQR: 60–77) years, and 57.2% were men. Median NIHSS on admission was 7 (IQR: 4–11). Median time from symptom onset to treatment with rt-PA was 150 (IQR: 115–185) min. The most frequent cerebrovascular risk factor was hypertension (82.2%). Favorable short- and long-term outcome was achieved in 45.2% and 48.6% of patients, respectively, excluding cases with post-lysis ICH. ICH occurred in 34 patients (8.1%), in 14 cases (3.3% of total cohort) it was considered SICH, and in 20 patients (4.8% of total cohort) aSICH.

The Effect of Thrombolysis on α 2-Plasmin Inhibitor Levels

α 2-PI activity and antigen levels before, immediately after, and 24 h after thrombolysis are shown on **Figure 1**. Admission

TABLE 1 | Baseline characteristics and outcomes of enrolled patients.

	<i>n</i>
Number of patients	421
Age, y, median (IQR)	68 (60–77)
Male sex, n (%)	241 (57.2)
NIHSS on admission, median (IQR)	7 (4–11)
Cerebrovascular risk factors, n (%)	
Arterial hypertension	346 (82.2)
Atrial fibrillation	86 (20.4)
Diabetes mellitus	118 (28.0)
Hyperlipidaemia	267 (63.4)
Active smoker	128 (30.4)
Previous stroke, TIA	114 (27.1)
BMI, kg/m ² , median (IQR)	27.4 (24.5–31.7)
Medication at enrollment, n (%)	
Antihypertensive therapy	209 (49.6)
Antiplatelet medication	173 (41.1)
Oral anticoagulant	22 (5.2)
Lipid lowering therapy	110 (26.1)
Antidiabetic therapy	69 (16.4)
Laboratory measurements on admission, median (IQR)	
WBC, G/L	7.9 (6.3–9.7)
Platelet count, G/L	217 (180–262)
Serum glucose, mmol/L	6.5 (5.6–8.1)
hsCRP, mg/L	2.9 (1.6–6.0)
INR	0.98 (0.93–1.02)
APTT, sec	27.8 (25.9–30.7)
α 2-plasmin inhibitor activity, %	102 (93.0–111.0)
α 2-plasmin inhibitor antigen, mg/L	59.6 (54.0–67.3)
Stroke etiology (TOAST), n (%)	
Large-artery atherosclerosis	146 (34.8)
Small-vessel occlusion	76 (18.0)
Cardioembolic	54 (12.8)
Other/undetermined	145 (34.4)
Imaging data, n (%)	
ASPECTS on admission	
0–7	6 (1.4)
8–10	268 (63.7)
ASPECTS at 24 h after thrombolysis	
0–7	50 (11.9)
8–10	225 (53.4)
Thrombolysis (i.v. rt-PA) treatment, median (IQR)	
Duration of thrombolysis, min	60 (60–60)
Time from symptom onset to treatment, min	150 (115–185)
rt-PA dose, mg	68 (58–81)
Outcomes, n (%)	
Short-term outcome (Δ NIHSS, day 7) *	
Favorable (at least –4 points or day 7 NIHSS = 0)	175 (45.2)
Unchanged status (\pm 3 points)	159 (41.0)
Unfavorable (at least +4 points or death)	46 (11.9)
Undetermined	7 (1.8)
Long-term outcome (mRS, day 90) *	

(Continued)

TABLE 1 | Continued

	<i>n</i>
Favorable (mRS 0–1)	188 (48.6)
Unfavorable (mRS 2–6)	157 (40.6)
Undetermined	42 (10.8)
Intracerebral hemorrhage, ICH (ECASS II, 24 h)	
No ICH	387 (91.9)
aSICH	20 (4.8)
SICH	14 (3.3)

Data are medians (interquartile ranges) or *n* (%).

APTT, activated partial thromboplastin time; aSICH, asymptomatic intracerebral hemorrhage; ASPECTS, Alberta Stroke Program Early CT Score; ECASS II, European Cooperative Acute Stroke Study-II; hsCRP, high sensitive C-reactive protein measurement; ICH, intracerebral hemorrhage; INR, international normalized ratio; IQR, interquartile ranges; mRS, modified Rankin Scale; NIHSS, National Institutes of Health Stroke Scale; SICH, symptomatic intracerebral hemorrhage; rt-PA, recombinant tissue plasminogen activator; TIA, transient ischemic attack; TOAST, Trial of ORG 10,172 in Acute Stroke Treatment; WBC, white blood cell count.

*Excluding patients with post-lysis intracerebral hemorrhage (*n* = 34).

α 2-PI activity and antigen levels showed a surprisingly wide distribution, but the majority of patients had α 2-PI levels within the reference range. Both α 2-PI activity and antigen levels showed a highly significant decrease immediately after thrombolysis, indicating a rapid complex formation and consumption of the protein by the generated plasmin during the procedure. As shown in **Figure 1**, α 2-PI activity and antigen levels of all patients were below the lower limit of reference when measured immediately after thrombolysis (α 2-PI activity median: 8 [IQR: 1–29] %; α 2-PI antigen median: 11.3 [IQR: 8.2–17.3] mg/L). Twenty-four hours after thrombolysis, α 2-PI activity and antigen levels were substantially increased but were still below admission values (α 2-PI activity median: 76 [66–86] %; α 2-PI antigen median: 39.4 [IQR: 34.1–46.1] mg/L). Strongest correlation between α 2-PI activity and antigen levels ($r=0.770$, 95%CI: 0.723–0.808, $p < 0.001$) was observed at 24 h after thrombolysis (**Figure 2**). Interestingly, the association between α 2-PI activity and antigen levels on admission was found to be the weakest in this cohort ($r=0.560$, 95%CI: 0.486–0.627, $p < 0.001$). Among the baseline clinical and laboratory parameters (as listed in **Table 1**), both α 2-PI activity and antigen levels showed a modest significant negative correlation with age (Spearman's $r: -0.2244$; 95%CI: -0.3168 to -0.1278 , $p < 0.001$ and $r: -0.3908$; 95%CI: -0.4739 to -0.3008 , $p < 0.001$, respectively). α 2-PI activity, but not α 2-PI antigen, showed a fair significant positive correlation with admission fibrinogen levels ($r: 0.3623$; 95%CI: 0.2723 to 0.4460, $p < 0.001$). α 2-PI activity and antigen levels were significantly higher in active smokers and in women.

Admission α 2-Plasmin Inhibitor Levels and Stroke Characteristics

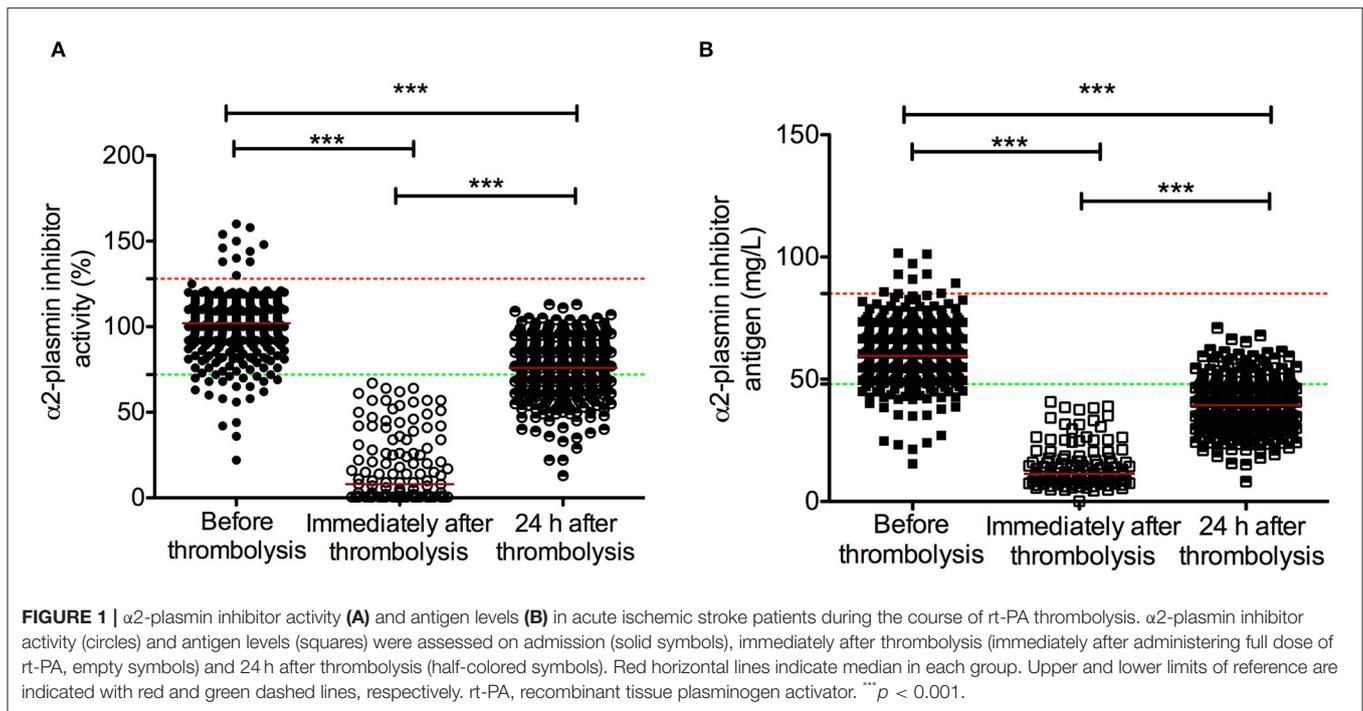
α 2-PI levels on admission showed a significant association with stroke severity (**Figure 3**). Those patients who suffered more severe stroke based on their NIHSS value on admission demonstrated significantly lower α 2-PI levels (**Figures 3A,B**). The stepwise inverse association with stroke severity was found

to be more profound for α 2-PI antigen levels (**Figure 3B**), and a similar but weaker association was found again in samples obtained at 24 h post-lysis (**Figure 3F**). α 2-PI levels after thrombolysis did not show a significant association with stroke severity (**Figures 3C,D**). Admission α 2-PI antigen levels were the highest in case of small vessel infarcts and lowest in strokes of cardio-embolic origin (α 2-PI antigen median: 61.8 [IQR: 56.3–72.8] mg/L vs. 56.6 [IQR: 52.3–64.2] mg/L, respectively, $p = 0.024$).

α 2-Plasmin Inhibitor Levels During Thrombolysis and Stroke Outcomes

Admission α 2-PI antigen levels showed a significant association with short-term outcomes of stroke (**Figure 4**). Significantly lower α 2-PI antigen levels were found on admission in patients with more severe stroke at 7 days post-lysis (**Figure 4A**). In patients who demonstrated unfavorable outcome at 7 days post-lysis, α 2-PI antigen levels on admission were significantly lower as compared to those improving or showing no change in their status (favorable outcome/no change group median: 60.4 [IQR: 54.5–68.8] mg/L vs. unfavorable outcome group median: 58.0 [IQR: 49.6–64.1] mg/L, $p=0.045$), although it must be noted that the difference between the median of the two groups was marginal (**Figure 4B**). α 2-PI antigen levels after thrombolysis and α 2-PI activity as measured at any time points were not associated with short-term outcomes. Similarly, long-term functional outcomes and mortality at 90 days were found to be associated only with α 2-PI antigen levels on admission (**Figure 5A**). Of note, the extent of decrease between pre- and post-thrombolysis α 2-PI activity or antigen levels did not show an association with outcomes (data not shown). Patients who died or had unfavorable long-term outcomes (mRS 2–5) had a significantly lower α 2-PI antigen levels on admission as compared to patients with favorable long-term outcome at 90 days after the event (mRS 0–1 median: 61.6 [IQR: 55.9–70.5] mg/L vs. mRS 2–5 median: 59.7 [IQR: 54.5–69.1] mg/L vs. mRS 6 median: 56.0 [IQR: 48.5–61.0] mg/L, $p < 0.001$). In a Kaplan–Meier survival analysis, those patients who presented with α 2-PI antigen level in the highest quartile on admission showed significantly better survival as compared to those with α 2-PI antigen level in the lowest quartile on admission (HR: 4.54; 95%CI: 1.92–10.8, $p < 0.001$) (**Figure 5B**).

Binary backward logistic regression models (model 1: including hyperlipidemia, BMI, diabetes mellitus, sex, and CRP) revealed that α 2-PI antigen level in the lowest quartile on admission is a significant predictor of unfavorable long-term outcomes (mRS 3–6) (OR: 2.10; 95%CI: 1.21–3.66, $p=0.008$) and death (mRS = 6) 3 months after thrombolysis (OR: 2.22, 95%CI: 1.15–4.31, $p = 0.018$) (**Table 2**). However, when age and NIHSS were also introduced in the models (**Table 2**, Models 2 and 3), the effect of α 2-PI antigen level diminished and only age and NIHSS on admission remained as independent predictors of both outcomes.



In patients with therapy-related ICH ($n = 32$), admission α 2-PI antigen levels were significantly lower as compared to those without hemorrhagic complications (Figure 6A). No such association was observed in the post-lysis samples of patients (Figures 6B,C) and in case of α 2-PI activity at any measured time points (data not shown). However, difference between median α 2-PI antigen levels on admission of those without or with post-lysis ICH was marginal (no ICH median: 59.8 [IQR: 54.0–68.0] mg/L vs. ICH: 57.0 [IQR: 53.4–61.7] mg/L, $p = 0.036$), and no correlation was observed between estimated post-lysis hematoma volumes and α 2-PI antigen levels on admission ($r = 0.207$; 95%CI: -0.176 to 0.536 , $p = 0.272$). α 2-PI antigen or activity levels did not differ in subgroups of SICH or aSICH at any time points investigated (data not shown). As expected, NIHSS was significantly higher in patients with ICH as compared to those without post-lysis hemorrhage (median: 12 [IQR: 7–32] vs. 6.5 [IQR: 4–36], respectively, $p < 0.001$).

α 2-Plasmin Inhibitor Arg6Trp Polymorphism, Stroke Characteristics, and Outcomes

Genotype frequencies of α 2-PI Arg6Trp polymorphism were consistent with Hardy–Weinberg equilibrium in the cohort ($C = 0.8046$; $T = 0.1953$) and were practically identical to allele frequencies reported in the 1000 Genomes project for the European subgroup ($C = 0.8012$; $T = 0.1988$) (40). α 2-PI Arg6Trp polymorphism had no effect on α 2-PI activity or antigen

levels at any measured time points and showed no association with stroke severity, etiology, or outcomes (data not shown).

DISCUSSION

In this prospective, observational study of 421 AIS patients undergoing thrombolysis, low α 2-PI levels on admission were found to be associated with more severe strokes and significantly worse long-term outcomes and mortality. Here, we report that α 2PI activity and antigen levels show a significant drop immediately post-lysis and increase to subnormal levels at 24 h post-event; however, post-lysis α 2-PI levels showed no association with outcomes. Despite the fact that in a multivariate analysis, admission α 2-PI levels did not prove to be an independent predictor of thrombolysis outcome, our results suggest that α 2-PI incorporation into thrombi could be an important mechanism for the limited efficacy of rt-PA thrombolysis.

Understanding the reasons of thrombolysis failure is critical for improving acute stroke care. Although α 2-PI is among the key regulators of fibrinolysis, little is known about its role in AIS thrombolysis outcome as yet. In the literature, only a few studies have been published on the potential association between α 2-PI levels and AIS thrombolysis outcomes, with controversial results (12, 25, 26, 41, 42). In a study including 63 AIS patients, baseline levels of α 2-PI activity correlated well with the rate of recanalization (42). Patients who recanalized had lower α 2-PI activity levels, and α 2-PI level was described to be the only predictive variable of recanalization, although it did not show an association with long-term outcomes. On the contrary, in

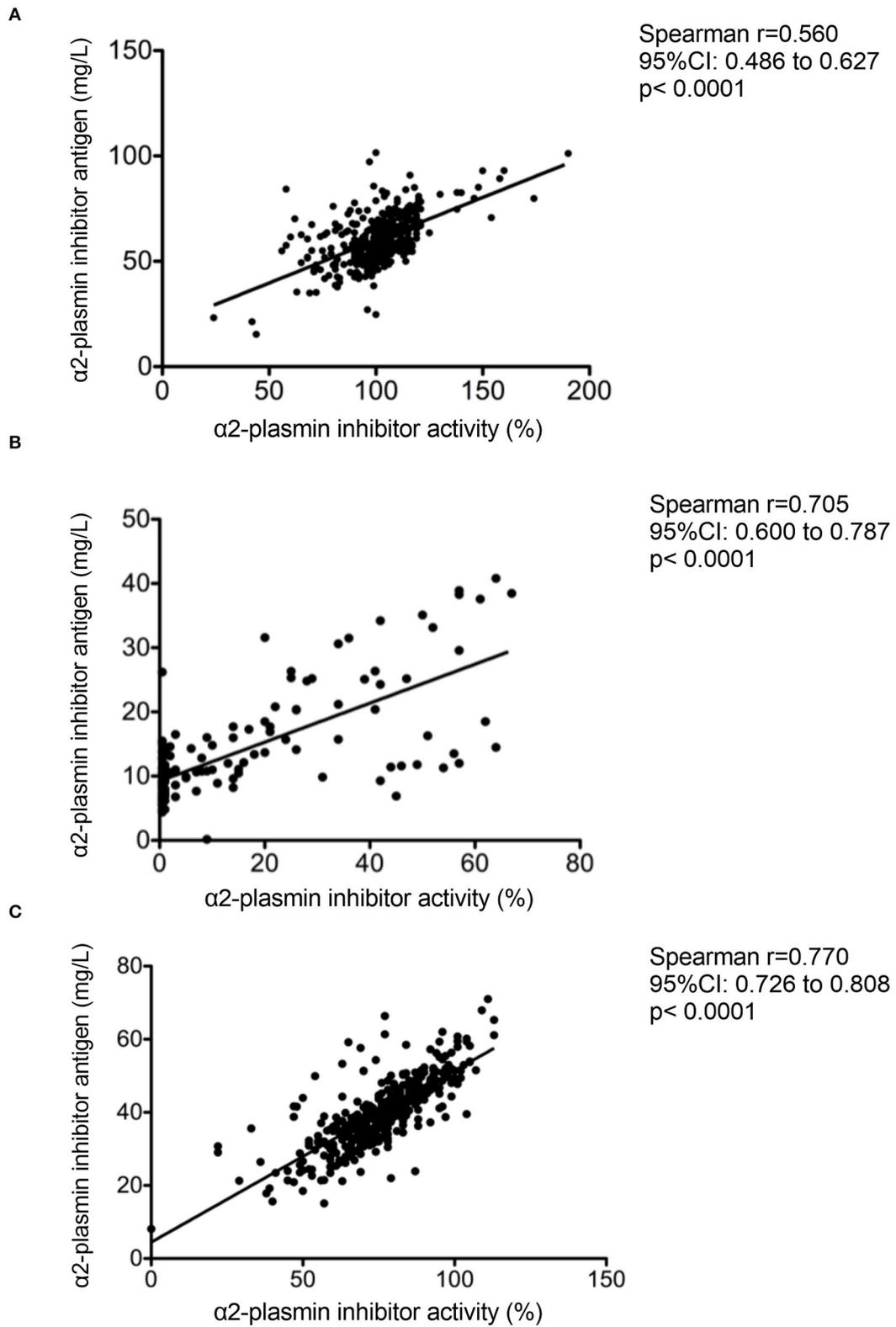


FIGURE 2 | Correlation between α 2-plasmin inhibitor activity and antigen levels as assessed on admission **(A)** immediately after thrombolysis **(B)** and 24 h after rt-PA thrombolysis **(C)** in acute ischemic stroke patients.

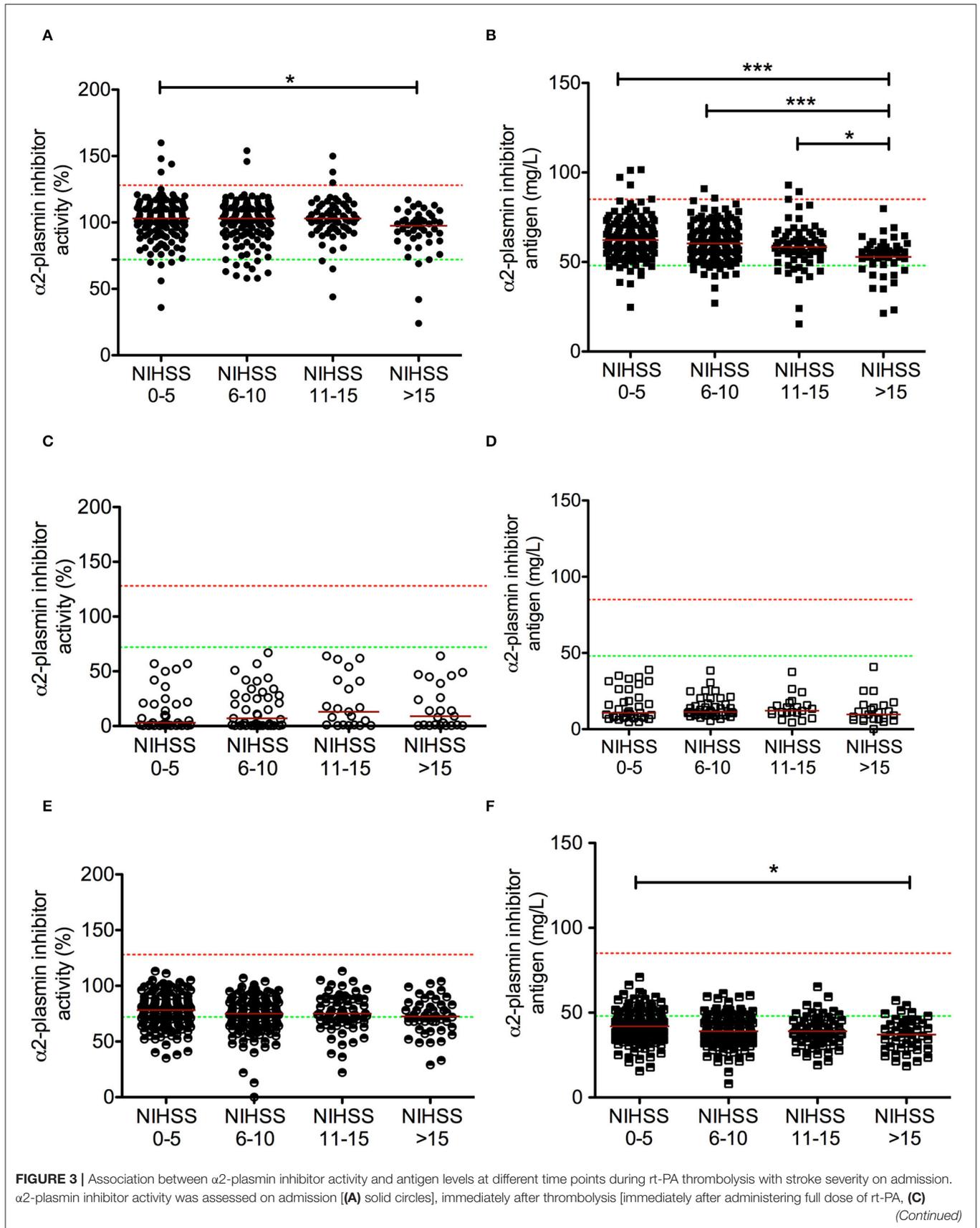
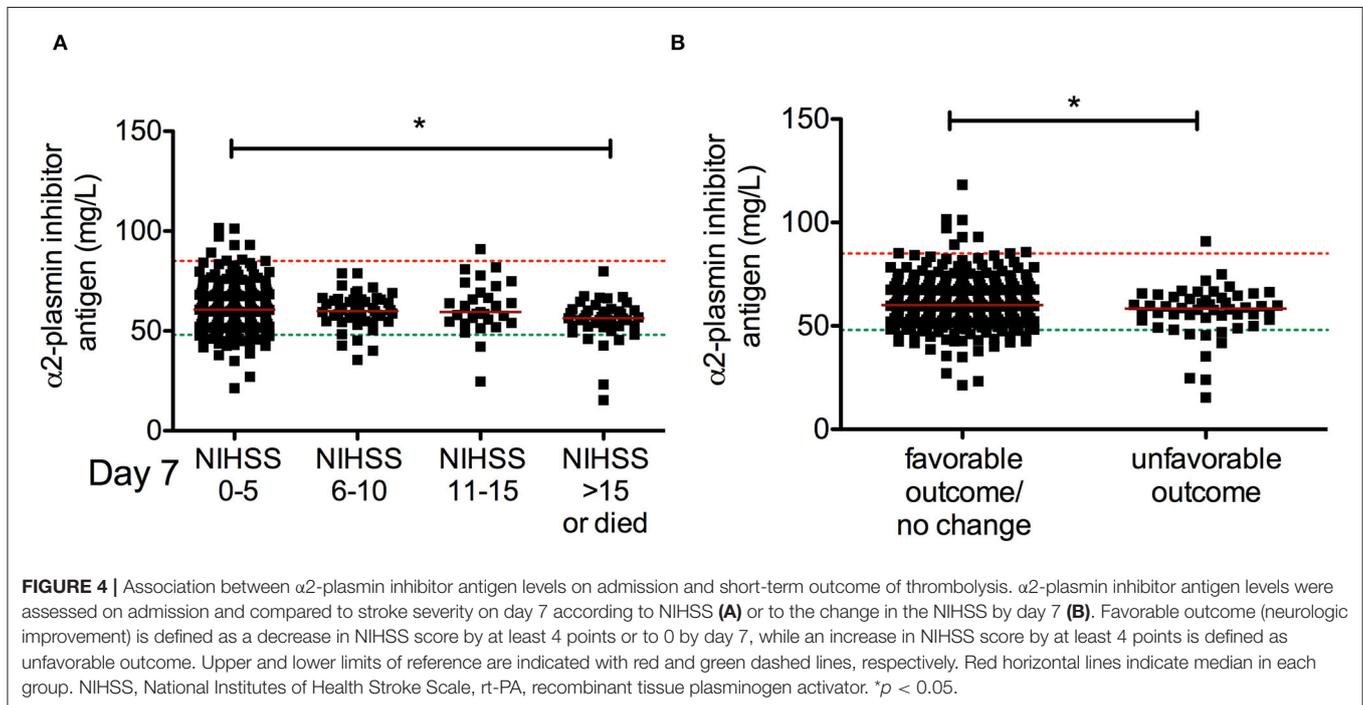


FIGURE 3 | empty circles], and 24 h after thrombolysis [(E) half-colored circles]. α 2-plasmin inhibitor antigen levels were assessed on admission [(B) solid squares], immediately after thrombolysis [immediately after administering full dose of rt-PA, (D) empty squares], and 24 h after thrombolysis [(F) half-colored squares]. Red horizontal lines indicate median in each group. Stroke severity on admission was grouped according to NIHSS. Upper and lower limits of reference are indicated with red and green dashed lines, respectively. NIHSS, National Institutes of Health Stroke Scale, rt-PA, recombinant tissue plasminogen activator. * $p < 0.05$, *** $p < 0.001$.



studies by others, although a decrease in α 2-PI levels was found after thrombolysis, α 2-PI levels showed no association with therapy outcomes (25, 26, 41). As it was pointed out by a recent systematic review, controversial data on markers of hemostasis and fibrinolysis and the outcome in AIS thrombolysis may arise from numerous methodological issues (15). First, most studies include relatively few patients lacking statistical power and only a handful of papers are published in the literature where at least 100 patients are included. Another critical factor is the time interval between stroke onset and blood sampling, as in the majority of studies a baseline blood sample is collected within 24 h after stroke onset, that is, a fairly wide time interval. Moreover, in patients receiving thrombolysis, therapy is provided during this period; thus, it is essential to differentiate between results gained from pre- or post-thrombolysis blood samples. Ideally, a hemostasis biomarker should be assessed before the initiation of thrombolysis treatment in AIS patients, despite the fact that sample collection might be a challenging task due to the short time window of treatment. Finally, depending on the research question, more than one sampling time point could be necessary to draw sound conclusions. In this prospective observational study, we were able to enroll 421 AIS patients undergoing thrombolysis, which makes this cohort one of the largest published as yet. Blood samples were collected from all patients before and 24 h after thrombolysis, while in a subset of patients

we also aimed at collecting samples immediately post-lysis. α 2-PI levels were tested at all time points by a functional test and an antigen assay detecting all four isoforms. Due to the natural heterogeneity of this protein, data derived from both assays could shed light on the action of α 2-PI during AIS thrombus formation and provide information on the inhibitory effect of α 2-PI on the excess plasmin generated during rt-PA therapy. Similarly to previous reports, α 2-PI activity and antigen results showed good correlation in this cohort (43). Interestingly, on admission, only moderately strong correlation was found between the activity and antigen assay results, which then improved post-lysis. This might arise from the fact that the incorporation of α 2-PI into the thrombus by FXIIIa primarily involves PB- α 2-PI (10, 43). It has been suggested by previous reports that the commercially available chromogenic α 2-PI activity assay used in this study is mainly sensitive to the levels of PB- α 2-PI, the more active and kinetically faster plasmin inhibitor (44, 45). On the contrary, the α 2-PI antigen test used captures all forms of α 2-PI to an equal extent; therefore, it is surmised to fully demonstrate the extent of α 2-PI consumption. This might be the reason for the stronger association between α 2-PI antigen levels and stroke severity observed in this cohort as compared to α 2-PI activity and might explain the link between admission α 2-PI antigen levels and outcomes. Further studies are required to understand the extent of incorporation of various α 2-PI isoforms into thrombi

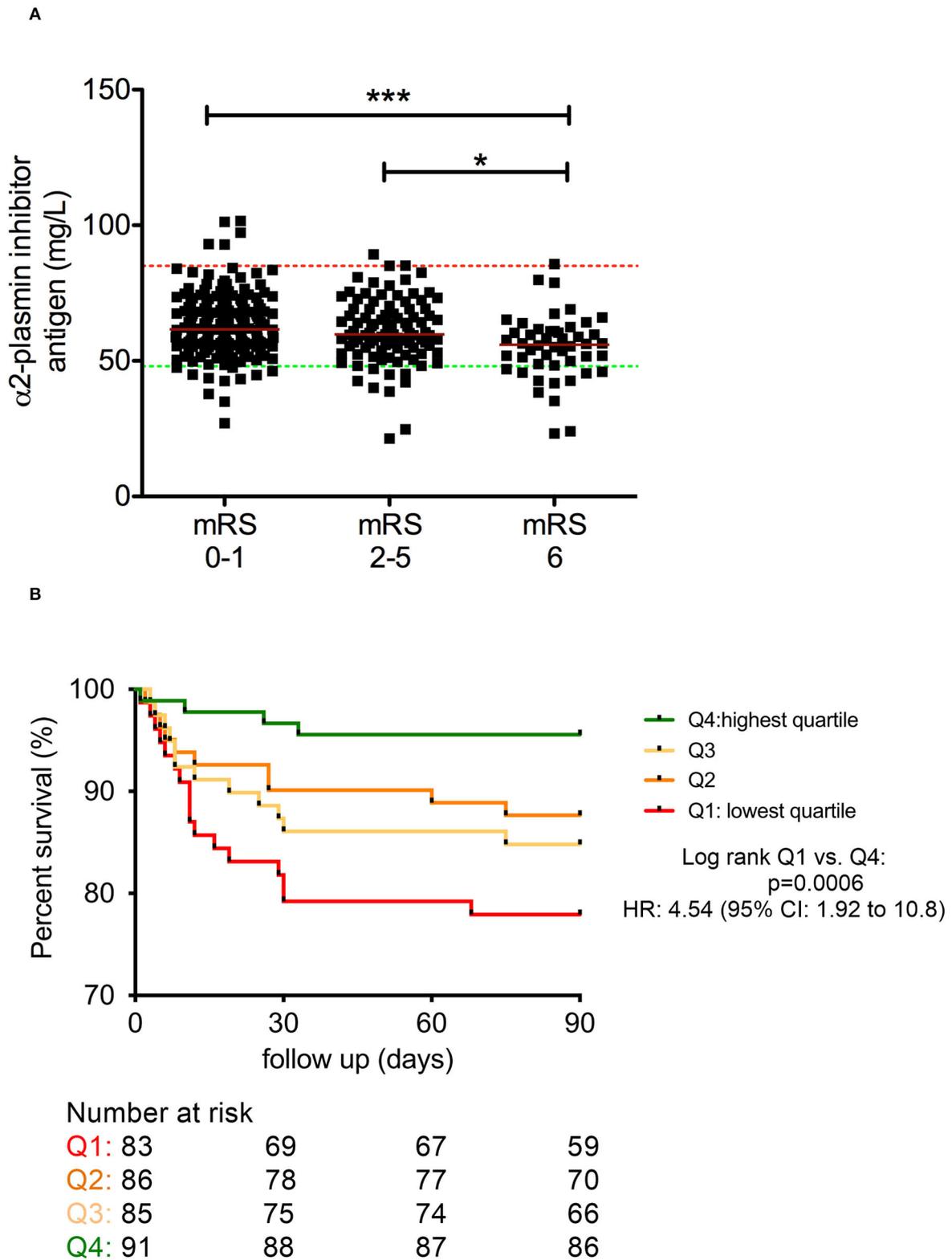


FIGURE 5 | Association between α 2-plasmin inhibitor antigen levels on admission and long-term outcome of thrombolysis **(A)**. Kaplan–Meier survival curves of patients based on quartiles of α 2-plasmin inhibitor antigen levels as measured on admission **(B)**. α 2-plasmin inhibitor antigen levels were assessed on admission (solid squares) and compared to long-term functional outcomes of thrombolysis based on the modified Rankin scale (mRS) **(A)**. Red horizontal lines indicate median

(Continued)

FIGURE 5 | in each group. Upper and lower limits of reference are indicated with red and green dashed lines, respectively. mRS 0–1 was considered as favorable outcome, and mRS 6 indicates mortality. Kaplan–Meier survival curves of patients are indicated according to quartiles of α 2-plasmin inhibitor antigen levels on admission (**B**). The number at risk is indicated below the graph. Upper quartile (Q4, green line), middle quartiles (Q3 and Q2, light orange and orange lines, respectively), and lowest quartile (Q1, red line) results were grouped based on the following cutoff values: 67.4 mg/L, 60 mg/L, and 54.1 mg/L α 2-plasmin inhibitor antigen level, respectively. HR, hazard ratio; mRS, modified Rankin Scale. * $p < 0.05$, *** $p < 0.001$.

TABLE 2 | Independent predictors of unfavorable outcome (mRS 2–6) and mortality (mRS 6) at 90 days post-lysis.

	Unfavorable outcome (mRS 0–1 vs. mRS 2–6)			Mortality at 90 days post-lysis (mRS 0–5 vs. mRS 6)		
	OR	95%CI	<i>p</i>	OR	95%CI	<i>p</i>
Model 1[§]						
α 2-plasmin inhibitor antigen lowest quartile	2.10	1.21–3.66	0.008	2.22	1.15–4.31	0.018
Diabetes mellitus	2.18	1.29–3.68	0.004			
CRP (per 1 mg/L)				1.02	0.99–1.04	0.099
Model 2[#]						
Diabetes mellitus	2.10	1.22–3.62	0.008			
Age (per 1 year)	1.05	1.03–1.07	<0.001	1.11	1.07–1.15	<0.001
CRP (per 1 mg/L)				1.03	1.00–1.05	0.028
Model 3[†]						
Age (per 1 year)	1.04	1.02–1.06	<0.001	1.08	1.04–1.27	<0.001
NIHSS (per 1 point)	1.17	1.10–1.24	<0.001	1.17	1.09–1.25	<0.001
Diabetes mellitus	2.05	1.16–3.62	0.014			

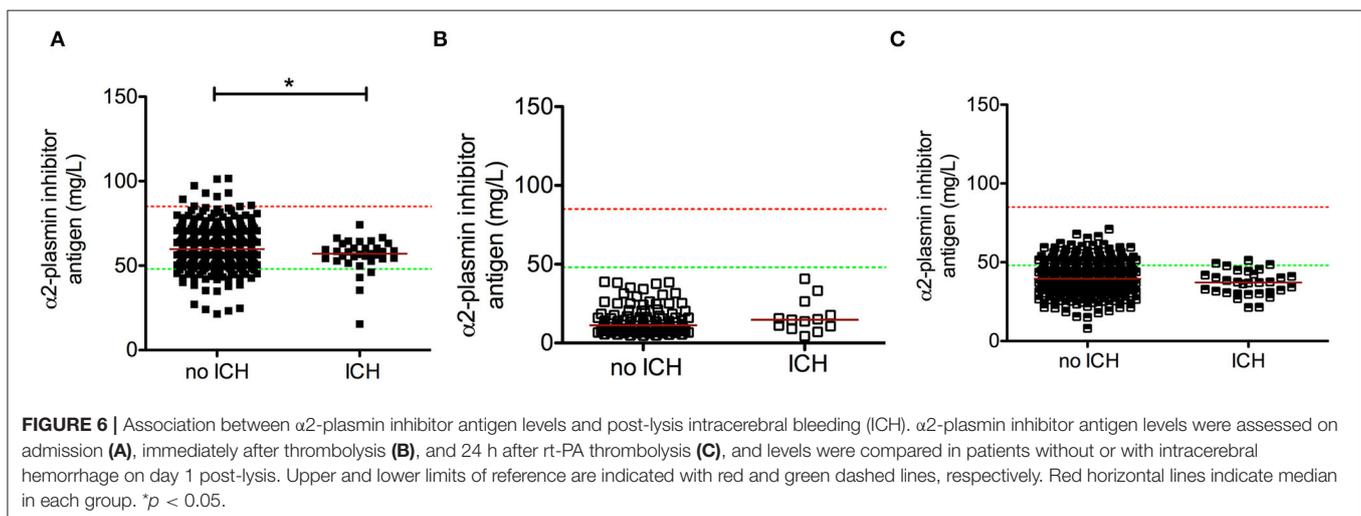
Last step of backward multiple regression analysis is provided.

[§] Model 1 included sex, hyperlipidemia, BMI, diabetes mellitus, CRP, and α 2-plasmin inhibitor antigen in the lowest quartile (threshold: <54.1 mg/L).

[#] Model 2 included Model 1 and age.

[†] Model 3 included Model 2 and NIHSS.

95%CI, 95% confidence interval; CRP, C-reactive protein measurement; NIHSS, National Institutes of Health Stroke Scale; OR, odds ratio; mRS, modified Rankin Scale.



and their relation to thrombus burden and treatment outcomes in AIS patients.

Based on our results, there is a considerable consumption of α 2-PI in patients with more severe strokes, as a significant stepwise negative association was found between α 2-PI antigen levels and stroke severity. Significantly lower admission α 2-PI antigen levels were found in patients with unfavorable short- and

long-term outcomes; moreover, admission α 2-PI antigen levels in the lowest quartile showed a significant association with the long-term mortality of patients. However, in a multivariate analysis including all relevant factors determining long-term outcomes, the significant effect of α 2-PI was diminished and only age and NIHSS remained in the model as most important predictors of long-term outcomes. These results suggest that although the

incorporation of α 2-PI into intracerebral thrombi and thus the consumption of the protein is associated with stroke severity and most probably with thrombus burden as well, apparently, α 2-PI cannot be considered an independent biomarker of therapy outcome. Nevertheless, these results suggest that α 2-PI could play a role in the pathophysiology of thrombolysis failure and may be an important factor contributing to the inefficacy of therapy in case of severe strokes. Consistent with this finding, we have shown in previous work that the extent of α 2-PI incorporation into fibrin clots was the lowest in AIS patients with favorable outcomes after thrombolysis (10). These results are in line with another recent work from our group where we showed that together with age and NIHSS, thrombus burden is the most important indicator of thrombolysis outcome, and the levels of a number of key hemostasis parameters measured on admission or 24 h later showed no association with treatment outcomes in a multivariate model (46).

Although it is biologically plausible that low α 2-PI levels post-lysis could be responsible for therapy-associated ICH events, such association was not found in this cohort. α 2-PI levels immediately post-lysis were below the lower limit of reference in all individuals tested in this cohort, yet bleeding complications were not directly related to levels of α 2-PI at this time point. These results are in line with experimental studies suggesting that targeting α 2-PI may be a novel paradigm for the dissolution of thrombi without compromising safety (47). Admission α 2-PI levels were indeed significantly lower in patients with post-lysis ICH; however, difference between groups was very modest. As patients with post-lysis ICH presented with more severe stroke on admission, it can be surmised that the significant difference observed between the α 2-PI levels of the two groups is related to differences in stroke severity. As α 2-PI levels in the groups with or without ICH showed a considerable overlap on admission, moreover, α 2-PI activity levels did not differ among groups, and it is unlikely that low α 2-PI would be the only responsible factor for post-lysis ICH. α 2-PI had no effect on the enlargement of the hematoma as α 2-PI levels did not show a significant correlation with the estimated post-lysis hematoma volumes. On the contrary, it has been known from the literature that more severe strokes are associated with not only lysis failure, but also with a higher chance of bleeding complications, although the reasons for this association have not been clarified (48). We have shown in a previous report that based on the *in vitro* incorporation of α 2-PI into fibrin clots, higher stroke severity was found to be associated with lower susceptibility to thrombolysis, together with a potentially higher risk of bleeding due to increased consumption of α 2-PI. The balance of fibrinolysis in these cases might be shifted toward bleeding near the thrombus site, which, together with other factors, could result in less protection against the rt-PA induced hemostatic challenge.

Among the parameters influencing α 2-PI heterogeneity, α 2-PI Arg6Trp was tested in the cohort. Given the relatively large sample size, the effect of α 2-PI Arg6Trp polymorphism on α 2-PI levels, stroke characteristics, and treatment outcomes was assessed, but no associations were found. Only a few studies have investigated the relation between α 2-PI Arg6Trp

polymorphism and the risk or outcomes of arterial thrombotic events. Limited data based on these reports suggest that the polymorphism has no effect on the risk or outcomes of ischemic events. As we have shown most recently, besides α 2-PI Arg6Trp polymorphism, an important modifying factor of the extent of α 2-PI N-terminal cleavage is circulating APCE (sFAP) level (43). It can be surmised that the effect of α 2-PI Arg6Trp should be considered in light of APCE levels; however, this measurement was out of the scope of our study.

LIMITATIONS

The results of this study should be interpreted in the context of its limitations and strengths. Although a more complex picture on the pathophysiology of α 2-PI during thrombolysis may have been gained by the measurement of α 2-PI isoforms, this was out of the scope of this study. Due to the single-centered study design, the study had the advantage of uniform patient and sample handling; moreover, the proportion of patients lost to follow-up (1.8 and 10.8% for short-term and long-term outcome, respectively) was relatively low as compared to other studies of similar study design (15). On the contrary, patients receiving mechanical thrombectomy were not included in the study and therefore the results cannot be directly translated to current treatment practices of AIS patients with LVO.

CONCLUSIONS

In this cohort of AIS patients, levels of α 2-PI changed dramatically during thrombolysis, but post-lysis α 2-PI levels showed no association with treatment outcomes or safety. Significantly lower α 2-PI levels were found on admission in AIS patients presenting with more severe stroke. Significantly lower admission α 2-PI antigen levels were found in patients with unfavorable short- and long-term outcomes; however, in a multivariate analysis including all relevant factors determining long-term outcomes, the significant effect of α 2-PI was diminished and only age and NIHSS remained in the model as most important predictors of long-term outcomes. Taken together, these data suggest that although α 2-PI is a dominant inhibitor of physiologic fibrinolysis, its levels on admission are strongly related to stroke severity and are affected by age to some extent, and they are not independent predictors of outcomes. It can be surmised, however, that in case of more severe strokes, the extent of α 2-PI incorporation into thrombi is higher, which could be an important mechanism for the limited efficacy of rt-PA thrombolysis in these cases.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary materials, further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by the Institutional Ethics Committee of the University of Debrecen and the Ethics Committee of the National Medical Research Council, Hungary. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

ES and RO-K collected clinical samples, performed experiments, and analyzed and interpreted data. IS, KC-K, NV, IF, KF, and EB collected clinical data and analyzed and interpreted data. BB and LL performed preanalytical sample care and performed

experiments. ÉK, LO, and LC analyzed and interpreted the data. ZB designed the research, analyzed and interpreted the data, and wrote the manuscript. All authors have read and approved the final manuscript.

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