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Case report: Characterization of a rare pathogenic variant associated with loss of COL3A1 expression in vascular Ehlers Danlos syndrome

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The vascular subtype of Ehlers Danlos Syndrome (vEDS) is a rare connective tissue disorder characterized by spontaneous arterial, bowel or organ rupture. The diagnosis of vEDS is established in a proband by identification of a heterozygous pathogenic variant in the alpha-1 gene of type III collagen (*COL3A1*) by molecular analysis. In this report, we present a case of vEDS with life threatening, spontaneous arterial dissections in association with an uncharacterized rare variant of *COL3A1*, *exon19:c.1340G* > *A*. Primary culture of patient skin fibroblasts followed by immunofluorescence revealed a complete absence of COL3A1 protein expression as well as altered morphology. Electron microscopy of the cultured fibroblasts showed abnormal vacuoles in the cytoplasm suggestive of a secretory defect. In this study, we have performed functional characterization of the *COL3A1 exon19:c.1340G* > *A* variant for the first time and this may now be classified as likely pathogenic in vEDS.

KEYWORDS

vascular Ehlers Danlos syndrome, COL3A1 pathogenic variant, clinical fibroblast testing, exome sequencing, stroke, hepatic artery dissection

Introduction

Vascular Ehlers Danlos Syndrome (vEDS; OMIM130050) is a rare connective tissue disorder caused by mutations in the alpha-1 chain of type III collagen (COL3A1) polypeptide leading to serious risk of arterial or organ rupture and premature death (1). It is dominantly inherited in 50% cases and the other 50% present with *de novo* pathogenic, somatic variants in *COL3A1* (2). The most common *COL3A1* pathogenic variant is a heterozygous missense substitution for glycine in the (Gly-X-Y) repeating

sequence of collagen triple helix which disrupts the assembly of type III homotrimeric collagen (3). This leads to defective type III collagen synthesis and assembly, manifesting as loss of mechanical strength in arteries and other hollow organs. The natural course of vEDS and associated clinical phenotype of patients are both reported to be influenced by the type of *COL3A1* variant (3, 4). Therefore, establishing a diagnosis of vEDS should ideally always include molecular genetic testing for *COL3A1* pathogenic variants and biochemical analysis to ascertain the pathogenicity of unreported variants (5).

Due to limited genetic testing and unavailability of clinical fibroblast culture testing, the prevalence and burden of vEDS in India is underestimated and underreported. vEDS is defined based on one major and several minor diagnostic criteria (1) which highlight the plethora of different physical signs that may constitute the clinical phenotype and cause confusion during diagnosis. Considering the unpredictable, life threatening complications of vEDS with a low median survival of around 48 years (2), it becomes imperative to employ genetic testing early during the management of a suspected vEDS patient especially when the phenotype is indistinguishable from other inherited connective tissue disorders.

Case description

A 48-year-old male with no past history of any relevant disease presented with the sudden onset of severe headache followed by right sided hemiparesis. On examination the patient had a thin face with prominent nose and lobeless ears (Figure 1A) along with fragile and thin skin over the extremities suggestive of Acrogeria (Figure 1B). Presence of varicose veins with some ecchymotic patches were observed over the lower limbs (Figure 1C). Chest veins were also found to be prominent (Figure 1A). Hypermobility of small joints of the hands (Figure 1D) and clubbing of feet were observed (Figure 1B). However, hyper elasticity of skin and marfanoid features were not present. Bilateral pitting edema was present in the lower limbs. All other vital parameters were within normal range.

Digital Subtraction Angiography (DSA) was suggestive of dissection with sub occlusive narrowing in the left internal carotid artery in the skull base region (**Figures 1E,F**). Patient was started on anticoagulation therapy with vitamin K antagonist (VKA) and 5 mg of warfarin, dose titrated to maintain a target INR (International Normalized Ratio) of 2–3. Hemiparesis gradually subsided and warfarin was stopped.

Patient was asymptomatic for 1 year followed by the sudden onset of upper abdominal pain. Computed tomography (CT) angiography showed aneurysmal dilatation of left hepatic artery (Figures 1G,H) along with multiple arterial dissections involving left renal artery (Figures 1I,J) and multiple infarcts in right renal artery and proximal right common iliac artery

(Figures 1K,L). The patient also developed elevated blood pressure which was a new symptom and was managed using anti-hypertensives. A rise in the level of the inflammatory marker, C-reactive protein was observed in serum. Nerve conduction study and nerve biopsy were suggestive of axonal and demyelinating neuropathy in both lower legs. Blood tests for vasculitis markers including antinuclear antibody, perinuclear and cytoplasmic antineutrophil cytoplasmic antibodies (myeloperoxidase and proteinase) were found to be negative. The patient was negative for HIV, hepatitis B and C. Within a few days our patient presented with another episode of acute abdominal pain with bilateral subcostal tenderness. He was diagnosed with hemoperitoneum due to Hepatic Artery rupture with ischemic liver changes, for which aneurysmal coiling was performed. There was no history of weight loss, fever, hemoptysis or melena. Laboratory test results are shown in Supplementary Table 1.

Whole exome sequencing analysis reveals a likely pathogenic *COL3A1* variant of uncertain significance

DNA was isolated from the peripheral blood sample of the patient and sent for whole exome sequencing (WES) to identify the cause of the disease. A total of 34058 variants were called after filtering for the depth of sequencing (minimum 10), of which 9257 were exonic variants, including single nucleotide variants and short indels. Exclusion of synonymous variants and filtering for minor allele frequency of <5% retained 854 variants. Once the non-pathogenic variants were removed using SIFT, Polyphen2-HDIV and Mutation Assessor predictions, 548 variants remained (**Supplementary Data 1**). After inclusion of 28 variants which affected splicing, there were 576 pathogenic variants across 392 different genes, of which 154 were found to be associated with a phenotype in OMIM.

On further evaluation based on the patient's phenotype of 'blood vessel abnormalities AND hypermobility' in OMIM, COL3A1 exon19:c.1340G > A [chr2:g.188994587G A,NM_000090.3:c.1340G>A, > NM_000090.3(COL3A1_i001):p.(Gly447Asp), all descriptions are based on hg38] variant leading to the substitution of Aspartic acid for Glycine at codon 447 (p.G447D) in type III collagen α 1 polypeptide was identified. The variant attributes according to the American College of Medical Genetics and Genomics (ACMG) and the Association for Molecular Pathology (AMP) guidelines (6) for the interpretation of sequence variants were PM2, PP2, and PP3. Visualization of this variant in Integrative Genome Viewer (IGV) suggested heterozygosity (Supplementary Data 1 in Supplementary Figure 1). This variant has been previously found to be associated with vascular subtype of Ehlers Danlos syndrome in one case report (7). Since this variant has not been reported in the 1000 genomes



(red arrow) with club foot (white arrow), (C): Easy bruising and thin skin, varicose veins, (D): Hypermobility of small joints (shown in distal phalange of index finger). Digital subtraction angiography images of bilateral common carotid arteries showing dissection of left internal carotid artery (E,F). Right internal carotid artery was normal. Axial (G) and coronal (H) maximum intensity projection (MIP) CT images showing aneurysmal dilatation of left hepatic artery. Coronal MIP images (I,J) of contrast enhanced CT show dissection involving left renal artery with multiple infarcts of renal parenchyma. Axial (K) and coronal (L) MIP CT images show dissection involving right renal artery and right common iliac artery.

(8), Exome Aggregation Consortium (9) (ExAC) and other databases (10) and no functional data was available regarding this variant, we further proceeded to analyze the pathogenic basis of this variant using molecular techniques.

The Exon19:c. 1340G > A variant leads to absence of COL3A1 protein in skin fibroblasts

Patient and control fibroblasts were cultured from skin biopsy (11) (Figures 2A,B) and analysis of protein expression of COL3A1 was done using immunofluorescence. There was a complete absence of COL3A1 production in the patient fibroblasts compared to the control fibroblasts (Figures 2C,D). The morphology of patient fibroblasts was also observed to be altered as compared to the control fibroblasts.

Fibroblasts derived from skin biopsy of patient with exon19:c. 1340G > A variant were larger in size as compared to the control fibroblasts (Figures 2C,D). It has been previously reported that dermal fibroblasts from vEDS have defective collagen biosynthesis/processing, endoplasmic reticulum homeostasis/protein folding, disorganized ECM interactions and may acquire a myofibroblast like phenotype (12). In order to investigate the altered morphology, we performed a Transmission electron microscopy (TEM) analysis. TEM has previously been suggested to be a useful addition to the repertoire of diagnostic tests in vEDS (13). Previous studies have reported an inefficient secretion of mutant collagen molecules which can lead to altered structural anatomy of the endoplasmic reticulum and unfolded protein response (14). We observed lamellar vacuolar bodies in the cytoplasm of vEDS fibroblasts which were not present in control fibroblasts (Figures 2E,F). On in-depth observational analysis, these bodies appeared to be "Amphisomes," organelles of the autophagy pathway resulting from fusion of endosomes and autophagosomes (15). This may explain the altered morphology and size of vEDS fibroblasts.

Sanger sequencing

Genomic DNA was also extracted from cultured dermal fibroblasts from patient and age-matched control and the *COL3A1 exon19:c.1340G* > A variant obtained from Whole exome sequencing was confirmed by Sanger sequencing (Figure 2G). All methodology provided in Supplementary Data 2.

Discussion

Diagnosis of vEDS based on clinical diagnostic criteria alone is often difficult due to significant overlap in clinical presentation with other connective tissue disorders and arteriopathies. vEDS diagnosis should be suspected in individuals presenting with spontaneous ruptures of arteries, uterus or bowel at a young age. Careful interpretation of genetic testing results and variant assessment according to the ACMG/AMP guidelines (6) is essential to confirm the diagnosis, as all COL3A1 variants are not pathogenic (16). Additionally, clinical fibroblast testing and functional analysis can help to accurately report pathogenicity of novel sequence variants.

Classification of *COL3A1* exon19:c.1340G > A as likely pathogenic variant

About 705 unique likely pathogenic/pathogenic COL3A1 variants have been reported, which are listed in the Leiden Open Variation Database (see https://databases.lovd.nl/shared/genes/ COL3A1). Missense mutation involving substitution of a glycine amino acid in the triple helix is the most common type of mutation reported in the database. The second most common type of mutation is a COL3A1 RNA splicing mutation. Large deletions or insertions in COL3A1 are relatively uncommon in vEDS. Most patients with vEDS develop major complications before the age of 30 years. Autosomal dominant inheritance of vEDS shows a penetrance approaching 100% but may also vary according to the age of presentation. Haploinsufficient patients have a lower penetrance of \sim 50% and were observed to survive 10-15 years longer than individuals harboring RNA splicing or glycine mutations (17). Interestingly, a missense mutation with substitution of a valine or an aspartic acid for a glycine was reported to have a poorer prognosis than a substitution of a serine (17). Although the phenotype of patients with amino acid substitutions in COL3A1 may partially overlap with of some haploinsufficient patients (18), there is considerable evidence for genotype-phenotype correlation (3, 16, 17) in vEDS which may be critical for screening and lifelong management of the affected individual and family members.

Our patient presented with stroke along with cranial and abdominal visceral arteries' dissections and rupture. Patient was hypertensive, had an elevated C-reactive protein (CRP) and neuropathy which lead to an initial suspicion of Polyarteritis Nodosa (PAN) (19). However, he did not show constitutional symptoms of fever, fatigue, weight loss, muscle/joint pains, radiological findings or other features defined by American College of Rheumatology classification criteria for PAN. The clinical presentation also favored the differential diagnosis of vEDS, but the presenting age of 48 years was not consistent with this diagnosis. The presence of multiple clinical findings namely acrogeria, progeria, lobeless ear, hypermobile small joints of hands, thin fragile skin, foot clubbing and multiple arterial dissections favored the diagnosis of vEDS. Patient had renal infarcts following renal artery dissection which could have contributed to the rise in blood pressure. It has been reported that delayed clearance of thrombi as well as the vascular insult to arteries in vEDS may lead to a rise in inflammatory markers (20), as observed in this case. Presence of axonal polyneuropathy has also been reported before in vEDS (21, 22). As there was no family history of EDS or related syndromes, in order to establish the diagnosis in a proband it was imperative to perform molecular genetic testing to identify and characterize this variant (2). Corticosteroids are used in PAN to reduce inflammation but they have been reported to trigger arterial dissections by elevating blood pressure and increasing blood vessel fragility by its inhibitory effect on collagen formation and connective tissue strength (23). Therefore, the patient was managed symptomatically till the genetic testing results were obtained.

Whole exome sequencing revealed a rare, uncharacterized variant of COL3A1 gene, exon19:c.1340G > A variant. A molecular diagnostic study was carried out to confirm if this variant was responsible for the vEDS phenotype. Dermal Fibroblast culture was established using skin biopsy from the patient and age matched control and COL3A1 protein expression was studied using immunofluorescence. The COL3A1 expression was observed to be absent in the exon19:c.1340G > A variant (Figures 2C,D). This suggests the expression of a mutant collagen with a dominant negative effect leading to a possible degradation after binding with the wild type collagen molecule. The presence of the variant was further confirmed by Sanger sequencing of the PCR amplicon obtained by using DNA extracted from cultured fibroblasts as the template and primers flanking the site of the variant (Figure 2G). We also observed 2 more nonsynonymous variants of COL3A1 (exon 30:c.2092G > A and exon 50:c.4059T > G) during the data analysis of the WES. However, based on their high minor allele frequencies, these were found to be non-deleterious (Table 1). Transmission



Electron Microscopy (TEM) studies of the dermal fibroblasts suggested that exon19:c.1340G > A variant is associated with altered endoplasmic reticular morphology in vEDS which may contribute to the disease mechanism.

According to the ACMG/AMP standards and guidelines for interpretation of sequence variants (6), the following COL3A1 variant criteria may be used to classify exon19:c. 1340G > A variant as pathogenic variant:

 $PS3 \rightarrow$ functional studies supporting damaged or altered gene product, absence of COL3A1 protein expression on immunofluorescence studies on skin fibroblasts and altered morphology of fibroblasts on electron microscopy studies.

 $PM2 \rightarrow variant not reported before in Exome sequencing project 1000 genomes and ExAC.$

 $PP2 \rightarrow$ missense z-scores of COL3A1 gene > 3.09, which was regarded as intolerant to missense variants.

PP3 \rightarrow SIFT (Deleterious), PolyPhen (Damaging) and a CADD-Phred score of 26.1 (pathogenic) support a deleterious effect on the gene or gene product.

Using the ACMG/AMP rules for combining the above criteria (6), $COL3A1 \ exon19$:c.1340G > A variant classification was found to be likely pathogenic (II) (Table 1).

Treatment and management

Diagnosis and management of a rare, life threatening vascular disease condition in absence of any curative therapy is a difficult conundrum for both the patient and the treating physician. Besides symptomatic treatment and regular follow up, psychological therapy and family support is required to accept and live with such a condition. Clinical evaluation of vEDS patients includes regular blood pressure monitoring, non-invasive arterial screening to detect dissections and dilatations such as ultrasonography, computed tomography or magnetic resonance imaging (1, 2). Any invasive procedures like colonoscopy, angiograms or surgeries should be performed by surgeons experienced and familiar with the enhanced caution required due to tissue fragility and its associated complications (24).

After the confirmation of *COL3A1* variant by exome sequencing and biochemical analysis, diagnosis was confirmed as vEDS. Patient was explained about the disease and its prognosis and referred for genetic counseling. It has been reported that Celiprolol improves the biomechanical integrity

Det	Details of the variant	÷.		Minor al	Minor allele frequency		P.	Pathogenicity prediction — <i>in silico</i>	liction — <i>in sil</i>	ico	ACMG attributes	Variant classification
Genomic co-ordinates	AAChange. refGene	Conseq -uence	ExAC_ ALL	1000g 2015 aug_all	gnomAD_ exome_ ALL	gnomAD_ genome_ ALL	SIFT_ pred	Polyphen2_ HDIV_pred	Mutation Taster_pred	CADD_ phred		
chr2:188994587	NM_000090.3	Missense					D	D	D	26.1	PS3	Likely
G > A	(COL3A1_i001):										PM2 PP2 PP3	Pathogenic (II)
chr20188000354	p.(Gly447Asp) NM 000090 3	Missense	0 3204	0.21845	0 2497	0.2118	F	£	đ	9.7 Q	RA1 RD4	Renian (I)
G > A	(COL3A1_i001):	TATISSETTSC	107C'0	01017.0	1647.0	0117.0	-	a	-	6.77		nemgu (1)
	p.(Ala698Thr)											
chr2:189010695	NM_000090.3	Missense	0.9985	0.996605	0.9988	0.9958	Т	В	Р	6.799	BA1 BP4	Benign (I)
T > G	(COL3A1_i001):											
	p.(His1353Gln)											

As research advances further in this field, gene therapy may be a promising option for the treatment of vEDS in the future. However, addition of the defective gene is not applicable for dominant diseases such as vEDS where the defective procollagen gene, COL3A, is a homotrimer of three identical α 1 chains. As most of the patients present with a heterozygous mutation, there is a structural defect in half of the synthesized collagen α 1 fibrils which results in 1/8th normal and 7/8th abnormal COL3A trimers. Interesting approaches such as siRNA mediated inhibition of the mutated allele and strategies to enhance transcriptional activation of the normal allele are being tested *in vivo* for phenotypic correction of vEDS (27, 28).

A recent study has identified PLC/IP3/PKC/ERK pathway (phospholipase C/inositol 1,4,5-triphosphate/protein kinase C/extracellular signal-regulated kinase) as major drivers of vascular pathology in vEDS based on preclinical models (29). AR101 (enzastaurin) is an orally active, small molecule, serine/threonine kinase inhibitor of the PLC/IP3/PKC/ERK pathways which has been previously studied in over 40 human trials including a range of cancers.¹ PREVEnt trial, which will assess the efficacy of enzastaurin in preventing cardiac or arterial events in patients with vEDS confirmed with *COL3A1* gene mutations is scheduled to begin next year and if successful may provide us with a new treatment for the life-threatening complications of vEDS.

Conclusion

We evaluated a rare *COL3A1* variant in vEDS using the ACMG/AMP standards and guidelines. The diagnosis was confirmed by genetic testing and pathogenicity was established by clinical fibroblast testing and functional assays. Based on our observations, *COL3A1 exon19:c.1340G* > *A* variant may now be classified as a likely pathogenic variant for vEDS exome analysis. Further detailed study may be required to understand the exact molecular mechanism of vEDS pathogenesis in this variant.

Data availability statement

The original contributions presented in this study are included in the article/**Supplementary material**, further inquiries can be directed to the corresponding author.

TABLE 1 ACMG/AMP classification criterion: Pathogenic Strong (PS3): Well-established in vitro or in vivo functional studies supportive of a damaging effect on the gene or gene product; Pathogenic

¹ Prevention of Rupture with Enzastaurin in Vascular Ehlers-Danlos Syndrome (https://clinicaltrials.gov/ct2/show/NCT05463679).

Ethics statement

Ethical review and approval was not required for the study on human participants in accordance with the local legislation and institutional requirements. The patients/participants provided their written informed consent to participate in this case study. Written informed consent was obtained from the individual(s) for the publication of this case study and any potentially identifiable images or data included in this article.

Author contributions

JM, LL, and UK conceived and designed the study. LL and UK performed the clinical evaluation. JM, AS, and SS performed and analyzed the experiments related to genetic testing and biochemistry. SG analyzed the radiology findings. JM, LL, and AS wrote the article. All authors proofread the manuscript.

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References

1. Malfait F, Francomano C, Byers P, Belmont J, Berglund B, Black J, et al. The 2017 international classification of the ehlers-danlos syndromes. *Am J Med Genet C Semin Med Genet.* (2017) 175:8–26.

2. Byers PH. Vascular ehlers-danlos syndrome. In: Adam MP, Ardinger HH, Pagon RA, Wallace SE, Bean LJ, Stephens K editors. *GeneReviews*. Seattle (WA): University of Washington (2019).

3. Frank M, Albuisson J, Ranque B, Golmard L, Mazzella JM, Bal-Theoleyre L, et al. The type of variants at the COL3A1 gene associates with the phenotype and severity of vascular ehlers-danlos syndrome. *Eur J Hum Genet.* (2015) 23:1657–64. doi: 10.1038/ejhg. 2015.32

4. The Ehlers–Danlos Syndromes, Rare Types - Brady - 2017. American Journal of Medical Genetics Part C: Seminars in Medical Genetics - Wiley Online Library. (2020). Available online at: https://onlinelibrary.wiley.com/doi/full/10.1002/ajmg. c.31550 (accessed July 24, 2020).

5. Cortini F, Villa C, Marinelli B, Combi R, Pesatori AC, Bassotti A. Understanding the basis of ehlers-danlos syndrome in the era of the next-generation sequencing. *Arch Dermatol Res.* (2019) 311:265-75. doi: 10.1007/s00403-019-01894-0

6. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the american college of medical genetics and genomics and the

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/ fcvm.2022.939013/full#supplementary-material

SUPPLEMENTARY TABLE 1 Laboratory investigations.

DATASHEET 1

Supplementary Data 1 (Whole exome sequencing data analysis pipeline).

DATASHEET 2

Supplementary Data 2 (Materials and Methods).

association for molecular pathology. Genet Med. (2015) 17:405-24. doi: 10.1038/gim.2015.30

7. Jones K, Choong A, Canham N, Renton S, Pollitt R, Nesbitt M, et al. A combined vascular surgical and clinical genetics approach to diffuse aneurysmal disease. *Ann R Coll Surg Engl.* (2015) 97:e73–6. doi: 10.1308/003588415X14181254790121

8. 1000 Genomes Project Consortium, Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, et al. A global reference for human genetic variation. *Nature*. (2015) 526:68–74.

9. Karczewski KJ, Weisburd B, Thomas B, Solomonson M, Ruderfer DM, Kavanagh D, et al. The ExAC browser: displaying reference data information from over 60 000 exomes. *Nucleic Acids Res.* (2017) 45:D840–5. doi: 10.1093/nar/gkw971

10. gnomAD. (2020). Available online at: https://gnomad.broadinstitute.org/ (accessed July 24, 2020).

11. Vangipuram M, Ting D, Kim S, Diaz R, Schüle B. Skin punch biopsy explant culture for derivation of primary human fibroblasts. *J Vis Exp.* (2013) 77:1437. doi: 10.3791/3779

12. Chiarelli N, Carini G, Zoppi N, Ritelli M, Colombi M. Molecular insights in the pathogenesis of classical ehlers-danlos syndrome from transcriptome-wide expression profiling of patients' skin fibroblasts. *PLoS One.* (2019) 14:e0211647. doi: 10.1371/journal.pone.0211647

13. Angwin C, Ghali N, Baker D, Brady AF, Pope FM, Vandersteen A, et al. Electron microscopy in the diagnosis of ehlers–danlos syndromes: correlation with clinical and genetic investigations. *Br J Dermatol.* (2020) 182:698–707. doi: 10.1111/bjd.18165

14. Chiarelli N, Ritelli M, Zoppi N, Colombi M. Cellular and molecular mechanisms in the pathogenesis of classical, vascular, and hypermobile ehlers-danlos syndromes. *Genes.* (2019) 10:3307. doi: 10.3390/genes1008 0609

15. Meijer AJ, Codogno P. Autophagy: regulation and role in disease. Crit Rev Clin Lab Sci. (2009) 46:210-40. doi: 10.1080/1040836090304 4068

16. Kuivaniemi H, Tromp G. Type III collagen (COL3A1): gene and protein structure, tissue distribution, and associated diseases. *Gene.* (2019) 707:151-71. doi: 10.1016/j.gene.2019.05.003

17. Pepin MG, Schwarze U, Rice KM, Liu M, Leistritz D, Byers PH. Survival is affected by mutation type and molecular mechanism in vascular ehlers-danlos syndrome (EDS type IV). *Genet Med.* (2014) 16:881–8. doi: 10.1038/gim.2014.72

 Schwarze U, Schievink WI, Petty E, Jaff MR, Babovic-Vuksanovic D, Cherry KJ, et al. Haploinsufficiency for one COL3A1 allele of type III procollagen results in a phenotype similar to the vascular form of ehlers-danlos syndrome. ehlersdanlos syndrome type IV. Am J Hum Genet. (2001) 69:989–1001. doi: 10.1086/32 4123

19. Lightfoot RW, Michel BA, Bloch DA, Hunder GG, Zvaifler NJ, McShane DJ, et al. The american college of rheumatology 1990 criteria for the classification of polyarteritis nodosa. *Arthr Rheum.* (1990) 33:1088–93. doi: 10.1002/art.17803 30805

20. Milewicz DM, Reid A, Cecchi A. Vascular ehlers danlos syndrome: exploring the role of inflammation in arterial disease. *Circ Cardiovasc Genet.* (2014) 7:5–7. doi: 10.1161/CIRCGENETICS.114.000507

21. Voermans NC, Drost G, van Kampen A, Gabreëls-Festen AA, Lammens M, Hamel BC, et al. Recurrent neuropathy associated with ehlers-danlos syndrome. *J Neurol.* (2006) 253:670–1. doi: 10.1007/s00415-005-0056-0

22. Castori M, Voermans N. Neurological manifestations of ehlers-danlos syndrome(s): a review. *Iran J Neurol.* (2014) 13:190–208.

23. Ohara N, Miyata T, Sato O, Oshiro H, Shigematsu H. Aortic aneurysm in patients with autoimmune diseases treated with corticosteroids. *Int Angiol.* (2000) 19:270–5. doi: 10.1053/ejvs.1999.0982

24. Eagleton MJ. Arterial complications of vascular ehlers-danlos syndrome. J Vasc Surg. (2016) 64:1869–80. doi: 10.1016/j.jvs.2016.06.120

25. Dubacher N, Münger J, Gorosabel MC, Crabb J, Ksiazek AA, Caspar SM, et al. Celiprolol but not losartan improves the biomechanical integrity of the aorta in a mouse model of vascular ehlers-danlos syndrome. *Cardiovasc Res.* (2020) 116:457–65. doi: 10.1093/cvr/cvz095

26. Frank M, Adham S, Seigle S, Legrand A, Mirault T, Henneton P, et al. Vascular ehlers-danlos syndrome: long-term observational study. *J Am Coll Cardiol.* (2019) 73:1948–57. doi: 10.1016/j.jacc.2019.01.058

27. Müller GA, Hansen U, Xu Z, Griswold B, Talan MI, McDonnell NB, et al. Allele-specific siRNA knockdown as a personalized treatment strategy for vascular ehlers-danlos syndrome in human fibroblasts. *FASEB J.* (2012) 26:668–77. doi: 10.1096/fj.11-182162

28. Watanabe A, Wada T, Tei K, Hata R, Fukushima Y, Shimada T. 618. a novel gene therapy strategy for vascular ehlers-danlos syndrome by the combination with RNAi mediated inhibition of a mutant allele and transcriptional activation of a normal allele. *Mol Ther.* (2005) 11:S240. doi: 10.1016/j.ymthe.2005.07.158

29. Bowen CJ, Calderón Giadrosic JF, Burger Z, Rykiel G, Davis EC, Helmers MR, et al. Targetable cellular signaling events mediate vascular pathology in vascular ehlers-danlos syndrome. *J Clin Invest.* (2020) 130:686–98. doi: 10.1172/JCI 130730