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Symmetrical peripheral gangrene: potential mechanisms and therapeutic approaches in severe COVID-19

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Symmetrical peripheral gangrene is a rare condition that is characterized by ischemic damage and tissue death (gangrene) in the extremities. Recent reports have shed light on SPG in patients with severe COVID-19. This condition presents with symmetrical cyanosis of the extremities and common COVID-19 symptoms and what the most frightening is within a few days, cutaneous necrosis occurred and patients died. Skin biopsy results have shown the presence of microthrombi in small vessels. The formation of SPG in COVID-19 patients results from immunothrombosis, endothelial dysfunction, and procoagulant platelets, leading to a hypercoagulation state and microvascular thrombosis. Thrombotic microangiopathy, shock, disseminated intravascular coagulation, and anticoagulant depletion promote the development of SPG in COVID-19. At the early stage, SPG patients with COVID-19 exhibit similar clinical manifestations. TMA causes early damage to microvasculature in SPG, and the shock state further exacerbates the ischemic injury due to local hypo-perfusion. The disturbed procoagulant-anticoagulant balance caused by DIC and anticoagulant depletion, combined with the pre-ischemic state brought on by TMA and shock, leads to the rapid formation of extensive microthrombi in the late stage of COVID-19 associated SPG. This review will delve into the clinical features, possible mechanisms, and potential therapeutic managements for COVID-19 associated SPG.

KEYWORDS

COVID-19, symmetrical peripheral gangrene, thrombotic microangiopathy, shock, disseminated intravascular coagulation

1. Introduction

The coronavirus responsible for the outbreak of COVID-19, known as severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), was initially detected in Wuhan city, Hubei province, China, in December 2019 (1). In early March 2020, the World Health Organization formally declared the global pandemic status of COVID-19. Despite primarily affecting the respiratory system, COVID-19 has been documented to cause a range of secondary effects, including myocarditis, acute kidney injury, hepatic dysfunction, and cutaneous manifestations.

The pandemic of COVID-19 has revealed an abnormal pattern of elevated thrombotic events, encompassing venous thromboembolism (VTE), arterial thrombosis, and thrombosis within the microvasculature. The underlying mechanisms behind the increased incidence of thrombosis in COVID-19 patients are not fully clarified but are thought to be related to a cytokine storm, hypoxia, endothelial dysfunction, hypercoagulability, and an increase in platelet activity (2–5).

Recent reports have described a unique cutaneous manifestation known as Symmetrical Peripheral Gangrene (SPG) in severe COVID-19 patients which present with symmetrical cyanosis of the extremities, along with symptoms such as fever, cough, hypoxemia, and hypotension. Unfortunately, all the COVID-19 patients eventually succumbed of SPG (6–8). Skin biopsy of patients with SPG reveals microthrombi in small vessels (6). This review aims to present comprehensive observations of the emerging clinical findings of SPG in COVID-19 patients to summarize the potential mechanisms, and describe the potential managements of COVID-19 associated SPG.

2. Case study of COVID-19 associated SPG

COVID-19 associated SPG presents with the formation of microthrombi and four main features: hypoxia, hypotension, DIC and AT depletion. Four cases of COVID-19 associated SPG have been reported (6–8) (Table 1). The patients consisted of three men and one woman, with ages ranging from 37 to 60. Of the two older patients, both had a history of hypertension. The presenting symptoms of the patients were fever, cough, and dyspnea. Upon admission, the patients were found to be hypoxic and hypotensive, with tachycardia and displayed cyanosis which

was symmetrical at the upper and lower extremities. Polymerase chain reaction (PCR) tests, using the reverse transcription method for detecting SARS-CoV-2, yielded positive results from samples taken from nasopharyngeal and/or oropharyngeal swabs. Laboratory tests indicated notable extensions in both prothrombin time (PT) and activated partial thromboplastin time (APTT), alongside increased levels of D-dimer and fibrin degradation products (FDPs). Neutrophilic leukocytosis and lymphopenia were also observed. One patient with COVID-19-associated SPG had a low level of antithrombin III protein (ATIII) at 53% (normal range: 80%–120%) (7). During hospitalization, the cyanosis of the extremities rapidly progressed to cutaneous necrosis. Skin biopsy of the SPG patients revealed the presence of microthrombi in small vessels, with no signs of vasculitis or vasculopathy (6).

3. The pathogenesis and etiologies of SPG

SPG is a rare complication that shows symmetrical distal ischemic damage and dry gangrene. The necrosis of SPG is predominantly acral and bilateral at least two locations including the toes, fingers, scrotum, and earlobes, without major vascular occlusive disease, while the term "purpura fulminans" (PF) refers to the occurrence the non-acral necrosis (9). Significant coldness, paleness, and bluish discoloration of the extremities are the precursors to suspect SPG, which advance rapidly to acrocyanosis and result in severe gangrene. Documented evidence shows a significant mortality rate, reaching up to 35%, and a substantial risk of multiple limb amputations, which occurs in up to 70% of those who survive SPG (10).

Sepsis is the underlying disorders of SPG, with some studies suggesting an estimated frequency of 2%-6% in sepsis (11).

TABLE 1 Clinical manifestations, laboratory parameters, treatment, and outcome of four COVID-19 associated SPG patients.

	Patient 1	Patient 2	Patient 3	Patient 4
Age	37	42	78	60
Sex	Male	Male	Female	Male
Comorbidities	None	None	Hypertension	Hypertension
Clinical manifestations	Fever, cough, dyspnea, hypoxia (89%)	Fever, cough, dyspnea, hypoxia (84%)	Fever, cough, dyspnea, progressive confusion, hypoxia	NA
Pattern of gangrene	Symmetrical peripheral gangrene appearing in limb	Dry gangrene appearing in toes of both feet with ulceration	Cyanosis at the extremity, in particular at nose area, hands and feet fingers	Gangrene relating with toes of feet
BP (mmHg)	89/50	104/60	85/40	Shock
WBC (cells/ul)	14,000	12,860	15,560	NA
ALC (cells/ul)	1,050	990	490	NA
PLT (cells/ul)	56,000	87,640	58,000	NA
D-dimer(ng/ml)	11,562	9,356	>40,000	Elevated
DIC (ISTH)	Yes	Yes	Yes	NA
ATIII	NA	NA	53%	NA
Skin lesion pathology	Microthrombi	NA	NA	NA
Treatment received	Enoxaparin, methyl-prednisolone, supportive care	Enoxaparin, dexamethasone, supportive care	Antiviral agent, antibiotics, supportive care	Conservative supportive care
Outcomes	Death (after 12 days of hospitalization)	Death (after 10 days of hospitalization)	Death	Death (after 20 days of hospitalization)

BP, blood pressure; WBC, white blood cell; ALC, absolute lymphocyte count; PLT, platelets; DIC, disseminated intravascular coagulation; ATIII, antithrombin III protein; NA, not available.

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Coagulopathy induced by sepsis leading to disseminated intravascular coagulation (DIC), a severe fatal disorder, which is also recognized as one of the pathological features of SPG. DIC occurred in at least 90% of SPG patients (12), nonetheless, SPG develops in only a limited subset of patients who experience shock and DIC (13). Low-flow state, circulatory shock, were observed in SPG. In critically ill patients, the emergence of SPG typically lags, appearing no sooner than two days after the onset of shock and DIC, hinting at a temporal element in the development of this condition. In over 90% of cases, individuals with SPG experience "shock liver", a precursor state to the development of ischemic necrosis in the limbs. The emergence of SPG is linked to a gradual decline in the concentrations of natural anticoagulants produced by the liver, especially antithrombin (AT) (14). Microthrombi in the capillary lumen, along with fibrin deposition and red blood cells extravasation, are observed in biopsy specimens of SPG patients (15). However, no evidence of vasculitis or inflammatory infiltrates was found on the vascular walls. Doppler ultrasonography of the peripheral arteries reveals that no large arteries are involved in the thrombosis (10).

The etiological factors leading to SPG include septic and nonseptic factors, as shown in Table 2.

TABLE 2 Etiology of symmetrical peripheral gangrene.

Septic factors	Other factors	
Bacterial	Cardiac disorders	
Escherichia coli	Low output cardiac failure	
Klebsiella pneumoniae	Third-degree atrioventricular block	
Serratia marcescens	Ventricular tachycardia	
Acinetobacter	Ventricular pseudoaneurysm	
Pseudomonas	Myocardial infarction	
Capnocytophaga canimorsus	Vasopressor drugs	
Proteus mirabilis	Norpin	
Salmonella paratyphi	Dopamin	
Proteus vulgaris	Epinephrine	
Pasteurella multocida	Noradrenaline	
Dermabacter jinjuensis	Various autoimmune conditions	
Neisseria meningitidis	Antiphospholipid syndrome	
Streptococcus pneumoniae	Cryoglobulinemia	
Staphylococcus aureus	Acquired hemolytic anemia	
Mycobacterium tuberculosis	Systemic lupus erythematosus	
Parasitic	Polymyalgia rheumatica	
Plasmodium vivax	Malignancies	
Plasmodium falciparum	Small cell lung cancer	
Leptospira	Hodgkin's lymphoma	
Viral	Drugs or poisons	
Dengue virus	Phenylephrine	
Rubeola virus	Sulphamezathine	
Varicella zoster virus	Penicillin	
Human immunodeficiency virus	Propylthiouracil	
	Warfarin	
	Snake venom	
	Therapy-related	
	Extracorporeal membrane	
	oxygenation	
	Idiopathic SPG	

4. Exploring the manifestations and pathophysiology of COVID-19 and SPG

4.1. The early damage to microvasculature of SPG is potentially caused by COVID-19-induced thrombotic microangiopathy

Thrombotic microangiopathy (TMA), a frequently observed form of microvascular thrombosis, constitutes a pathological characteristic of COVID-19 (5, 16). The presence of retiform purpura and livedo racemose, which are relatively infrequent dermatological manifestations of COVID-19, are distinguished by pauci-inflammatory TMA findings in skin biopsy outcomes (17, 18). All hospitalized patients diagnosed with retiform purpura exhibited acute respiratory distress syndrome in 82% of cases, with bilateral lesions on their extremities and buttocks showing varying degrees of severity (17, 19). Some SPG patients exhibit additional symptoms such as neurological complications, renal dysfunction, thrombocytopenia, and significant increase in lactate dehydrogenase (LDH) (6, 7). Retiform purpura and livedo racemose-like lesions have often been seen in patients with apparent coagulopathy, therefore, based on the comparable clinical presentations and pathological alterations, SPG may represent a more severe and advanced type of retiform purpura/ livedo racemose in COVID-19.

4.2. Septic shock is the potential cause of SPG in COVID-19 patients

The SARS-CoV-2 virus, being an infectious pathogen, has the potential to induce sepsis or septic shock in individuals with COVID-19. However, the prevalence of shock in adult COVID-19 patients varies greatly (ranging from 1% to 35%), and is contingent upon the population being researched, the disease severity, and the definition of shock (20, 21). Sepsis or septic shock contributes to persistent vasodilation, consequently causing hypotension and hypoxia (22).

All SPG patients in COVID-19 all had low blood pressures and three of them developed shock. The shock state exacerbates the already present ischemic injury by causing local hypoperfusion, leading to multiple organ failures, including liver dysfunction (7).

4.3. Disturbed procoagulant–anticoagulant balance in COVID-19 patients induces rapid formation of extensive microthrombi

The significance of DIC in SPG is supported by pathological studies revealing noninflammatory fibrin microthrombi deposits in small vessels (23). At the decompensated stage, excessively activated coagulation process leads to microthrombus generation and the depletion of clotting factors. According to the DIC score of the International Society on Thrombosis and Haemostasi, APTT and PT were prolonged and the levels of FDPs and D-

dimer are dramatically elevated, while the platelet numbers and fibrinogen levels are already decreased (24).

DIC and anticoagulant depletion greatly disturb the procoagulant–anticoagulant balance and lead to the rapid formation of extensive microthrombi at the late stage of COVID-19 associated SPG. DIC has been recognized in the advanced stage of severe COVID-19. During hospitalization, the criteria of DIC were met by 71.4% of non-survivors, while only 0.6% of survivors (24).

In severe COVID-19 patients, endothelial dysfunction, decreased liver synthesis, degradation through proteolytic cleavage enzymes of neutrophils, and excessive consumption due to DIC contribute to the anticoagulant depletion. Goshua and colleagues demonstrated that an elevation in the shedding of thrombomodulin from endothelial cells in patients with acute COVID-19 has been observed (25), which impairs the anticoagulant activity of protein C (PC). Another crucial component of the natural anticoagulation system, ATIII, is frequently deficient in severe COVID-19, with at least 25% of patients showing ATIII deficiency (26). ATIII level was significantly lower in non-survivors than survivors in COVID-19 (27). Patient 3 presented with low ATIII level (7). In addition, elevated levels of plasminogen activating inhibitor-1 (PAI-1) combined with hypofibrinolysis were found in COVID-19 patients (28). The hypofibrinolytic state suggests more extensive microthrombosis and less frequent hemorrhagic events in COVID-19 related DIC.

5. SARS-COV-2 infection and SPG: insight into underlying molecular machanism

5.1. Infection with SARS-CoV-2 causes direct damage to vascular endothelial cells, potentially causing a procoagulant state

SARS-CoV-2 targets and invades cells lining the respiratory tract, type II pneumocytes in the alveoli, and cells comprising the vascular endothelium by binding to angiotensin-converting enzyme 2 (ACE2) (29). COVID-19 cases reported by Varga et al. found the evidence that SARS-CoV-2 directly infects vascular endothelial cells (30). The integrity of the vascular endothelium is closely related to the hypercoagulable state. Intact endothelium secretes amounts of anticoagulant factors to prevent platelet activation and coagulation, such as nitric oxide, inhibitors of the tissue factor pathway such as TFPI, the coagulation-regulatory protein thrombomodulin, and the endothelial protein C receptor (EPCR). Direct infection of the endothelium by SARS-CoV-2 potentially damages endothelial surface, suppressing the expression of TFPI, TM, and EPCR, which is thought to the leading cause of procoagulant state in COVID-19 patients (31). In an autopsy study of 26 patients who died from COVID-19, researchers explored how SARS-CoV-2 infection damaged human kidneys. Results showed glomerular fibrin thrombosis in COVID-19 patients (32), which means that the SARS-CoV-2 infection may lead to a breakdown of the ECs barrier.

5.2. Activation of immune system caused by SARS-CoV-2 infection induces microvascular thrombosis associated SPG

Microvascular thrombotic events in COVID-19, usually characterized as immunothrombosis, are induced by the activated systemic immune systems, such as cytokine storm, neutrophil extracellular traps (NETs), and activated complementary system (5, 33). SARS-CoV-2 replication and viral particles released by infected host cells cause tissue damages in the lung (5). The pathogen associated molecular patterns (PAMPs) of SARS-CoV-2, including its spike glycoprotein and single-stranded RNA, along with host-derived molecular markers like high-mobility group box 1 (HMGB1) from infected cells, trigger immune responses via pattern recognition receptor (PRR) signaling cascades. This activation results in the synthesis and dissemination of chemokines and pro-inflammatory cytokines, encompassing tumor necrosis factor (TNF), interleukin-1β (IL-1β), IL-2 receptor (IL-2R), IL-6, and IL-8 (34). These chemokines cause the recruitment of neutrophils, macrophages and CD4 positive T lymphocytes in lung tissue (35), aggravating the local inflammatory response. Virus proteins, HMGB1 and cytokines can induce excessive release of NETs (36), which activate the oxygen species (ROS) formation (37). In severe COVID-19 patients, SARS-CoV-2 infection, NET proteins and ROS cause the pulmonary capillary destruction and leakage of proinflammatory mediators, such as HMGB1 and inflammatory cytokines (38-40) (Figure 1). DAMPs and cytokines in circulation further activate the systemic immune systems (41) and trigger intravascular thrombus formation through inducing the tissue factor (TF) expression on vascular ECs and monocytes (42).

The cytokine storm gives rise to lymphopenia which is prevalent in patients with COVID-19 and exists in two cases of COVID-19 associated SPG (6, 7). Lymphopenia impairs the function of the immune system to produce antibodies against virus-specific antigens, as well as reducing the generation of interferon gamma (IFN- γ) by the CD4+ T cells (43). Impaired clearance of the SARS-CoV-2 leads to the viremia in the circulation (44). High plasma levels of SARS-CoV-2 induce the elevation of cytokines such as IL-6 and enhance severe COVID-19 progression (45). The discovery of elevated levels of cytokines, like IL-6, in severe COVID-19 patients could be both a result and a cause of viremia, creating a potentially hazardous cycle that perpetuates itself.

The amount of Circulating NET-forming neutrophil subset (immunotype DEspR+CD11b+) and NETosis induction levels are positively related with the progression, severity, and duration of acute COVID-19 (46–48). The levels of NETosis initiation showed a significant correlation with platelet activation indicators and blood clotting-related factors (47, 49). NETs components cause the degradation of TFPI via neutrophil elastase during thrombotic complications (49), thus activating the coagulation cascade TF pathway (50). The initiation of the intrinsic blood clotting mechanism occurs when NETs carrying a negative charge form a direct association with, and set into action, the coagulation factor FXII. NETs additionally adhere to the Von Willebrand Factor (vWF), offering a foundation for platelets to attach (49). Complexes formed by NETs and platelets serve as frameworks that facilitate the



cytokines.SARS-CoV-2, severe acute respiratory distress syndrome-associated coronavirus-2; DAMPs, damage associated molecular patterns; PAMPs, pathogen associated molecular patterns; PRRs, pattern recognition receptors; ACE2, angiotensin converting enzyme-2; ECs, endothelial cells; NET, neutrophil extracellular trap. (Created with BioRender.com).

attachment of other molecules promoting coagulation, like fibronectin and fibrinogen. Consequently, these complexes capture free-floating blood cells and encourage their clustering, leading to the creation of blood clots and the blockage of blood vessels (51). Elements of NETs, such as histones and dsDNA, contribute to the advancement of thrombosis by enhancing the density, stiffness, and resilience of fibrin strands while obstructing the breakdown of fibrin (52).

By directly interacting with Mannan-binding lectin-associated serine protease-2 (MASP-2), SARS-CoV-2 activates the lectin complement pathway and causes ECs damage through membrane attack complex (MAC) (53). MASPs contribute to thrombus formation by cleaving prothrombin to form activated thrombin (54). Furthermore, the activated complement pathway releases C5a which increases TF activity via C5a receptors on neutrophils and endothelial cells (55).

5.3. Endothelial injury and platelets activation induced by COVID-19 contributes to microvascular thrombosis associated SPG

Activated systemic immune systems, viremia and hypoxemia in severe COVID-19 exert cytotoxic effects on ECs (5, 30). The

injured ECs upregulate the expression of adhesion proteins, including intercellular adhesion molecule-1 (ICAM1) and vascular cell adhesion molecule-1 (VCAM1), chemoattractants like monocyte chemoattractant protein-1 (MCP-1), and inflammatory mediators such as interleukins IL-1 and IL-6 (56), which aggravates systemic inflammatory reactions and immunothrombosis (57). vWF release induced by endothelial injury mediates platelet aggregation through the binding with glycoprotein Ib-IX-V complex and integrin aIIbb3 on platelets (58). Under typical conditions, enzymes present in the plasma like A disintegrin and metalloproteinase with thrombospondin motifs 13 (ADAMTS-13) are responsible for breaking down vWF multimers (59). However, in COVID-19, peptidyl arginine deiminase-4 (PAD4), an enzyme expressed in NETs, changes the structure of ADAMTS-13 and reduces the degrading ability of ADAMTS-13 (60). The increase in the vWF/ADAMTS-13 ratio, which reflects the thrombogenicity of the blood, suggests that hypercoagulable state in COVID-19 patients (61). In COVID-19 cases, the concentration of soluble P-selectin was markedly higher in patients those in the ICU as opposed to those not or healthy individuals (25, 62). P-selectin overexpressed by endothelial injury interacts with its ligand P-selectin glycoprotein ligand-1 (PSGL-1) on almost all leukocytes (63) and the

glycoprotein Ib α (GPIb α) receptor on platelets, which facilitates the attachment of leukocytes and platelets to the injured regions of the endothelium through activation of the PSGL-1 signaling in leukocytes and the GPIb signaling in platelets (64, 65). Furthermore, endothelial injury upregulates the expression of TF (66) which combines with coagulation factor VII (FVII) and promotes extrinsic pathway.

Thrombocytopenia was revealed in the patients with COVID-19 associated SPG (6-8). In COVID-19 patients, a significant association between low platelet counts and increased disease severity as well as high mortality rates has been reported (67, 68). In individuals who succumbed to COVID-19, autopsies revealed a substantial presence of megakaryocytes and clots abundant in platelets within the heart, pulmonary, and renal tissues (69, 70). Platelets release procoagulant factors in response to the cytokine storm (71). Furthermore, the engagement between NETs and platelets, coupled with platelet activation driven by the HMGB1-Toll-like receptor (TLR)-4 pathway, impact the initiation of the coagulation process, resulting in the widespread generation of microthrombi (36). Platelets may also be activated by the incorporation of the C5b-9 complex into their surface (72), the binding of C1q to its corresponding receptor on the platelet membrane (73), and their responsiveness to component C3 (74). It has been reported that human and mouse platelets express ACE2, to which the spike protein of SARS-CoV-2 binds, leading to the release of vWF from α - and dense granules (75). Activated platelets express TF and P-selectin on their membranes, which promotes their binding to leukocytes and endothelial cells, enhancing the expression of TF by activating the nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) pathway (76). As a coagulation factor, PF4 released by activated platelets neutralizes heparin-like molecules on the endothelial surface of blood vessels. Increased PF4 levels and enhanced plateletneutrophil aggregates were found in COVID-19 patients (77). Upon activation, platelets discharge vascular endothelial growth factor (VEGF), which triggers endothelial cells to upregulate the expression of TF (66). Additionally, activated platelets exhibit catalytic activities by assembling coagulation factors on their surface (78). The main mechanisms of microvascular thrombosis in COVID-19 associated SPG were summarized in Figure 2.

6. Potential approaches for managing thrombotic events in patients with COVID-19 associated SPG

In the realm of managing COVID-19 associated SPG, a multifaceted therapeutic approach is crucial due to the complex pathophysiology of the disease. The treatment strategies encompass a broad spectrum, ranging from targeting the virus itself to addressing the systemic effects induced by the infection. This comprehensive review elucidates the multifarious treatment options (Figure 3), underlining the necessity of a tailored therapeutic regimen based on individual patient profiles and disease severity in COVID-19 associated SPG.

6.1. Antiviral therapy

During the early phase of infection, when the viral load of SARS-CoV-2 is elevated and the patient exhibits mild to moderate symptoms, medications aimed at inhibiting viral replication may prove more beneficial, as the adaptive immune system has yet to mount a sufficient response. Administering antiviral medications promptly increases their potential effectiveness, particularly for individuals at elevated risk (79-82). Now the common used antiviral drugs include protease inhibitors (like Paxlovid) and RNA-dependent RNA polymerase (RdRp) blockers (like Remdesivir and Molnupiravir). Paxlovid, a combined formulation of Nirmatrelvir and Ritonavir tablets, inhibits the SARS-CoV-2's 3-CL protease during the proteolysis stage, thereby preventing the virus from replicating (83). It was formulated for the management and post-exposure prevention of COVID-19 in patients who are more likely to develop severe symptoms (84). Molnupiravir, a ribonucleoside prodrug of Nhydroxycytidine effectively blocks the RdRp of SARS-CoV-2, thereby preventing the virus's ability to transcribe and replicate its genetic material (85). When administered within 5 days of symptom onset, it has the potential to lower the likelihood of hospitalization and fatality in individuals with COVID-19 who are more susceptible to progressing to a severe form of the disease (86). Remdesivir is another RdRp blocker (87). Being the initial drug to receive FDA approval for COVID-19 treatment, Remdesivir demonstrates strong binding capabilities with the SARS-CoV-2 spike, ACE2, and transmembrane protease serine2 (TMPRSS2), suggesting its potential to block the entry of the virus (88). The research trial indicated that Remdesivir successfully reduced the duration of recovery and hindered the advancement of respiratory illness in patients with COVID-19 (89). By effectively reducing viral load and inhibiting virus entry into host cells, the following inflammatory response, injury to the endothelium, and initiation of coagulation will subsequently diminish, leading to the reduced COVID-19 associated complications, especially the occurrence of thrombosis (Figure 3).

6.2. Anticoagulant therapy

Heparin or low-molecular-weight heparin (LMWH) is recommended as prioritized anticoagulant due to its monitorable anticoagulant effect. Additionally, liver dysfunction and renal failure does not affect heparin clearance. Besides its anticoagulant properties, heparin has anti-inflammatory effects (90). A recent meta-analysis found that patients with sepsis, septic shock, and infection associated DIC those who received heparin (compared with usual care or placebo) had a 12% decrease in mortality rate (90). Fondaparinux sodium, a selectively synthesized inhibitor of factor Xa, is authorized for use in both preventing and treating VTE incidents in patients who are acutely unwell (including those with COVID-19 or cancer) as well as individuals undergoing surgical procedures (91). Additionally, it is applicable



FIGURE 2

The main mechanisms of microvascular thrombosis in COVID-19 associated SPG. SARS-CoV-2 stimulates neutrophils to release NETs via ACE2 and activates the NET/TFPI/TF and NET/FXII pathways. By directly binding to MASP-2, SARS-CoV-2 can activate the lectin complement pathway and cause ECs damage through MAC. C5a which is released by the activated complement pathway increases TF activity via C5a receptors on neutrophils and ECs. SARS-CoV-2 also can bind to ACE2 on platelets, leading to the release of granules, such as vWF, P-selectin, PF4, and VEGF. VEGF triggers ECs to upregulate the expression of TF. Due to the interaction with PAD4 released by NETs, the degrading ability of ADAMTS-13 decreased, aggravating the increase of vWF. vWF multimers and P-selectin released by activated ECs, lead to aggregation of platelets by GPIba receptor. Pselectin on activated platelets induce TF expression of monocytes by PSGL-1 signaling pathway. DAMPs and cytokines inducing the expression of TF on monocytes and vascular ECs. The cytokines also can induce the TF expression and the granules release of platelets. In addition, NETs, DAMPs (such as HMGB1), and complementary system also contribute to the activation of platelets. The coagulation cascades are initiated mainly by TF through the extrinsic pathway. Furthermore, activated platelets can directly activate the intrinsic pathway by assembling coagulation factors on their surface. These mechanisms ultimately lead to the unchecked generation of thrombin, resulting in micro-thrombus formation in COVID-19 associated SPG. SARS-CoV-2, severe acute respiratory distress syndrome-associated coronavirus-2; NETs, neutrophil extracellular traps; ACE2, angiotensin converting enzyme-2; TFPI, tissue factor pathway inhibitor; FXII, factor XII; MASP, Mannan-binding lectin-associated serine protease; ECs, endothelial cells; MAC, membrane attack complex; TF, tissue factor; PF4, platelet factor 4; VEGF, vascular endothelial growth factor; DAMPs, damage associated molecular patterns; HMGB1, high mobility group box 1; PAD4, peptidylarginine deiminase; ADAMTS-13, A disintegrin and metalloproteinase with thrombospondin motifs 13; vWF, von Willebrand factor; GPIba, glycoprotein Iba; PSGL-1, P-selectin glycoprotein ligand 1; PRRs, pattern recognition receptors; MBL, mannan-binding lectin. (Created with BioRender.com).

for use in cases of acute coronary syndromes, bariatric surgical procedures, heparin-induced thrombocytopenia (HIT) (92), and vaccine-induced immune thrombotic thrombocytopenia (VITT) (93). The incidence of venous thromboembolism (VTE) at 6.5% compared to 13.5% (P = 0.36), and bleeding events at 6.5% vs. 4.1% (P = 0.68), did not significantly differ between the group of COVID-19 patients administered fondaparinux and the group given enoxaparin treatment (94). Moreover, fondaparinux presents several clinical benefits for individuals diagnosed with COVID-19, based on its possible antiviral (95) and anti-inflammatory effects (96). In this review, despite receiving appropriate treatments, such as enoxaparin, glucocorticoids, and

antiviral agents, all COVID-19 patients suffered from SPG succumbed of ARDS or multiple organ failure within a few days of hospitalization (6–8). In addition, anticoagulant therapy is controversial in terms of dose and intensity. Accordingly, we need to identify the early signals of COVID-19 associated SPG, such as symmetrical purpura and TMA related indexes, and optimize the timing and dose of anticoagulant therapy (97).

In a clinical trial of patients with severe sepsis, despite the fact that the death rate was lower in AT-treated individuals with DIC who were not administered heparin (98), high-dose AT concentrates did not improve the total mortality (99). In addition, after a large-scale, randomized trial failing to



demonstrate an improvement in survivals of patients with septic shock, recombinant activated protein C was removed from the candidate drugs (100). Employing rTFPI as part of therapeutic approaches targeting the fibrinolysis system might be effective against endothelial damage, inflammation, NETosis, and coagulation disorders induced by SARS-CoV-2 (52).

6.3. Anti-inflammatory therapy

Corticosteroids, recognized medications for addressing inflammation and autoimmune disorders, connect with nuclear subsequently diminishing receptors. the secretion of proinflammatory cytokines (101). Across various laboratory and live organism studies, steroids prompt a decrease in the formation of NETs (102). Steroids additionally diminish the secretion of HMGB1 and its engagement with TLR4 (103). The evidence regarding their application in COVID-19 remains inconclusive (104–107). Recent findings suggested that administering methylprednisolone in the early stages for a short duration is linked to improved clinical results in patients experiencing severe COVID-19 pneumonia, and such treatment should be contemplated prior to the onset of ARDS (108, 109).

Considering the presence of cytokine storm and inflammatory response in sepsis and coagulopathy of COVID-19 patients (38, 39, 110), IL-6 is identified as a potential drug target. RECOVERY trial results indicate that tocilizumab (anti-IL-6 drug), along with corticosteroids, improved the survival in patients with COVID-19 who had hypoxia and systemic symptoms (111). Janus kinase (JAK) inhibitor has shown therapeutic potential in severe COVID-19 by suppressing inflammatory pathway (112) and should be considered in the treatment of SPG patients.

NETs play a vital role in immunothrombosis after SARS-CoV-2 infection. Continued efforts that prevent NET release could potentially be a successful tactic in managing COVID-19.associated coagulopathy. The release of NETs can be controlled by curbing the generation of ROS and hypochlorous acid (HOCl). This can be achieved either by suppressing NADPH oxidase and/or myeloperoxidase (MPO) activity or by adding antioxidants (113). For instance, both metformin and diphenyleneiodonium (DPI) have the capacity to reduce NETosis, owing to their potential to inhibit the activity of NADPH oxidase and reduce the production of reactive oxygen species from mitochondria (114, 115). Likewise, augmenting with various compounds that neutralize ROS, such as resveratrol, flavonoids, and N-acetylcysteine, can prove efficacious in minimizing the discharge of NETs (116-119). Moreover, individuals with severe or critical COVID-19 exhibited notably reduced levels of NETosis following treatment with dexamethasone (47).

Therapeutic drugs which inhibit the complement process potentially inhibit the process of sepsis and the overactive inflammatory reaction in COVID-19 (18, 110). For example, in the case of COVID-19 patients with severe pneumonia, Diurno et al. noted that after initiating treatment with eculizumab, a complement system inhibitor, there was observable progress in clinical symptoms, lung lesions identified via CT scans, and laboratory test results within the first 48 h (120).

6.4. Endothelial repair and antioxidant therapy

COVID-19 outcomes are adversely affected by preexisting endothelial dysfunction, which is associated with sex, smoking, metabolic syndrome, and existing cardiovascular diseases (121). Severe COVID-19 results in endothelial dysfunction, leading to a shift towards a pro-coagulant state with increased vasoconstriction and inflammation (56). In addition, endothelial injury is a core pathogenesis and the therapeutic target of TMA (122, 123). Agents with endothelial cell-modifying effects, for example, phosphodiesterase inhibitors (PDEi) (124) potentially have therapeutic effect in SPG patients with severe COVID-19.

N-acetylcysteine (NAC), as an antioxidant, increases the biosynthesis of glutathione and reduces the generation of reactive oxygen species (ROS) (125). In recent years, NAC which protects endothelial cells free from ROS attack has been utilized in cases of thrombotic thrombocytopenic purpura and in transplant-associated thrombotic microangiopathy (126, 127). Research showed that NAC protects COVID-19 patients from oxidative stress-mediated endothelial damage (128).

6.5. Antiplatelet and fibrinolytic therapy

Clinically, dipyridamole (DIP), a drug that possesses anti-platelet aggregation properties, has been linked to elevated platelet counts and decreased levels of D-dimer. In both animal and *in vitro* research, DIP has been shown to inhibit the replication of SARS-CoV-2 and stimulate a type I interferon (IFN) response (129). Previous research has reported that DIP has protective effect against the aggravation of endotoxin induced DIC in experimental animals (130). The impact of aspirin use, another anti-platelet aggregation drug, has been examined in cases of COVID-19 infection accompanied by ARDS. Nonetheless, the findings have been varied, with certain studies indicating benefits while others have not shown positive outcomes (131–134). To ascertain the definitive efficacy of antiplatelet therapy in COVID-19, randomized controlled trials involving varied patient groups are necessary.

Wang J et al. documented instances of three individuals with COVID-19-induced ARDS who underwent treatment with Alteplase, an intravenous tissue-type plasminogen activator (tPA). Their findings revealed an initial improvement of PaO₂/ FiO₂ ratio ranging from 38% to 100% in all cases. In addition, the administration of tPA systemically was linked to a decrease in mortality [47.6% (tPA) vs. 71.0% (no tPA)] for COVID-19 patients who had a refractory PaO2/FiO2 of less than 60 mmHg (135). Nonetheless, the widespread administration of fibrinolytic agents carries the hazard of possibly lethal bleeding events. Indeed, nearly 7% patients subjected to such agents require transfusion of blood products, while approximately 1% succumb to the effects of hemorrhages (136). Administering the treatment locally through nebulization presents a compelling alternative, possibly offering enhanced effectiveness while minimizing the risk of bleeding. The nebulized form of recombinant tPA may aid in promoting localized clot breakdown within the alveolar space and enhance oxygen levels (137). The improvement of hypoxia status is also beneficial for improving COVID-19 related lung injury and other complications. Finally, enhanced-fibrinolytic-type DIC occurs at advanced stage of COVID-19, which means fibrinolytic therapy is only effective in the early stages of COVID-19 (138). Therefore, early recognition of SPG before the onset of enhanced-fibrinolytic-type DIC is critical.

6.6. Plasma exchange

In patients with COVID-19 suffering from SPG, plasma exchange (PEX) may serve as a therapeutic intervention, ideally initiated within the first 48 h after shock liver onset (14). PEX operates by a mechanism akin to that seen in other TMAs, wherein it clears detrimental substances such as endotoxins and inflammatory cytokines while replenishing depleted components of the coagulation system, including natural anticoagulants and cofactors, thus inhibiting fibrin production throughout various stages of the coagulation pathway (139).

6.7. Recovered plasma and intravenous immunoglobulins

Administering serum from individuals who have healed and tested PCR-negative, enriched with IgG antibodies against SARS-CoV-2 [also termed hyperimmune IgG-containing plasma (HIgCP)], offers a treatment strategy for recently infected individuals, drawing from past learnings associated with various viral ailments. Utilizing HIgCP could offer therapeutic advantages for the management or prevention of ARDS caused by SARS-CoV-2 (140, 141). Nonetheless, the requirement for a compatible blood type between the donor and recipient, coupled with the potential danger of contracting additional viral infections, renders HIgCP less apt for widespread use. Intravenous immunoglobulins (IVIg) have been influential in adjusting immune reactions. IVIg reduces NET formation (142), cytokine and DAMP production (143). Moreover, IVIg offers protection from cell death initiated by HMGB1, influencing the expression of TLR and RAGE (144). The efficacy of IVIG treatment in managing COVID-19 is still a matter of debate. The outcomes of a meta-analysis suggested that IVIg treatment did not have a discernible impact on either the death rate or the duration of hospitalization (145). While other study showed that IVIG might decrease the death rate in comparison to the control group in severely ill COVID-19 individuals (146). So IVIG has shown to be clinically effective on critical ill patients which indicated its use in SPG patients with severe COVID-19.

6.8. Vascular dilation therapy

Cases of peripheral gangrene have been documented subsequent to the use of several vasoconstrictive medications such as vasopressin, dopamine, and noradrenaline, and traditionally, it occurs after consuming ergot (147–151). Vasodilators such as prostaglandin (152), epoprostenol (150, 153) and iloprost (154) may minimize tissue loss with regression of necrotic lesions in SPG, especially applied in early stage (150). However, the use of vasodilators is mainly reported in case reports, with the combination of other treatments, such as tissue plasminogen activator (153) and heparin (154). To verify their healing impact on SPG, rigorous clinical studies must be conducted.

7. Conclusion

SPG is a frequently overlooked complication of severe COVID-19 that arises from a systemic disorder rather than a localized vascular disease. COVID-19 associated SPG presented with formation of microthrombi and four main features: hypoxia, hypotension, DIC and AT depletion. Immunothrombosis, endothelial dysfunction and procoagulant platelets contribute to the formation of microvascular thrombosis. TMA, circulatory shock, DIC and anticoagulant depletion enhance the development of COVID-19 associated SPG. Ultimately, SPG occurs in COVID-19 due to the procoagulant-anticoagulant imbalance caused by the greatly disturbed microvasculature at risk, the DIC state, and anticoagulant depletion. Managing strategies are to treat patients suffering SPG from the perspective of anticoagulation benefiting or without affecting the treatment of COVID-19. Searching for early indicators from the progression of COVID-19 are still the keys to manage SPG.

Author contributions

JL: Conceptualization, Funding acquisition, Supervision, Writing – original draft, Writing – review & editing. MW: Conceptualization, Writing – original draft, Writing – review &

References

1. Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A novel coronavirus from patients with pneumonia in China, 2019. *N Engl J Med.* (2020) 382(8):727–33. doi: 10. 1056/NEJMoa2001017

2. Bilaloglu S, Aphinyanaphongs Y, Jones S, Iturrate E, Hochman J, Berger JS. Thrombosis in hospitalized patients with COVID-19 in a New York City health system. *JAMA*. (2020) 324(8):799–801. doi: 10.1001/jama.2020.13372

 Jimenez D, Garcia-Sanchez A, Rali P, Muriel A, Bikdeli B, Ruiz-Artacho P, et al. Incidence of VTE and bleeding among hospitalized patients with coronavirus disease 2019: a systematic review and meta-analysis. *Chest.* (2021) 159(3):1182–96. doi: 10. 1016/j.chest.2020.11.005

4. Cheruiyot I, Kipkorir V, Ngure B, Misiani M, Munguti J, Ogeng'o J. Arterial thrombosis in coronavirus disease 2019 patients: a rapid systematic review. *Ann Vasc Surg.* (2021) 70:273–81. doi: 10.1016/j.avsg.2020.08.087

5. Ackermann M, Verleden SE, Kuehnel M, Haverich A, Welte T, Laenger F, et al. Pulmonary vascular endothelialitis, thrombosis, and angiogenesis in COVID-19. *N Engl J Med.* (2020) 383(2):120–8. doi: 10.1056/NEJMoa2015432

6. Sil A, Chakraborty U, Chandra A, Biswas SK. COVID-19 associated symmetrical peripheral gangrene: a case series. *Diabetes Metab Syndr*. (2022) 16(1):102356. doi: 10. 1016/j.dsx.2021.102356

7. Novara E, Molinaro E, Benedetti I, Bonometti R, Lauritano EC, Boverio R. Severe acute dried gangrene in COVID-19 infection: a case report. *Eur Rev Med Pharmacol Sci.* (2020) 24(10):5769–71. doi: 10.26355/eurrev_202005_21369

8. Deraje V, Gujjalanavar RS, Das S, Janardhan R, Rao P. Acute extremity gangrene in COVID-19 patients. *Indian J Plast Surg.* (2021) 54(2):237–9. doi: 10.1055/s-0041-1731853

9. Warkentin TE. Ischemic limb gangrene with pulses. N Engl J Med. (2015) 373 (7):642–55. doi: 10.1056/NEJMra1316259

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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10. Ghosh SK, Bandyopadhyay D, Ghosh A. Symmetrical peripheral gangrene: a prospective study of 14 consecutive cases in a tertiary-care hospital in eastern India. *Eur Acad Dermatol Venereol.* (2010) 24(2):214–8. doi: 10.1111/j.1468-3083.2009.03329.x

11. Vincent JL, Nadel S, Kutsogiannis DJ, Gibney RT, Yan SB, Wyss VL, et al. Drotrecogin alfa (activated) in patients with severe sepsis presenting with purpura fulminans, meningitis, or meningococcal disease: a retrospective analysis of patients enrolled in recent clinical studies. *Crit Care.* (2005) 9(4):R331–43. doi: 10.1186/cc3538

12. Molos MA, Hall JC. Symmetrical peripheral gangrene and disseminated intravascular coagulation. *Arch Dermatol.* (1985) 121:1057-61. doi: 10.1001/archderm.1985.01660080111027

13. Knight TT Jr., Gordon SV, Canady J, Rush DS, Browder W. Symmetrical peripheral gangrene: a new presentation of an old disease. *Am Surg.* (2000) 66 (2):196–9. doi: 10.1177/000313480006600217

14. Warkentin TE, Ning S. Symmetrical peripheral gangrene in critical illness. *Transfus Apher Sci.* (2021) 60(2):103094. doi: 10.1016/j.transci.2021.103094

15. Davis MD, Dy KM, Nelson S. Presentation and outcome of purpura fulminans associated with peripheral gangrene in 12 patients at mayo clinic. *J Am Acad Dermatol.* (2007) 57(6):944–56. doi: 10.1016/j.jaad.2007.07.039

16. Ahmadian E, Hosseiniyan Khatibi SM, Razi Soofiyani S, Abediazar S, Shoja MM, Ardalan M, et al. COVID-19 and kidney injury: pathophysiology and molecular mechanisms. *Rev Med Virol.* (2021) 31(3):e2176. doi: 10.1002/rmv.2176

17. Freeman EE, McMahon DE, Lipoff JB, Rosenbach M, Kovarik C, Desai SR, et al. The spectrum of COVID-19-associated dermatologic manifestations: an international registry of 716 patients from 31 countries. *J Am Acad Dermatol.* (2020) 83(4):1118–29. doi: 10.1016/j.jaad.2020.06.1016

18. Magro C, Mulvey JJ, Berlin D, Nuovo G, Salvatore S, Harp J, et al. Complement associated microvascular injury and thrombosis in the pathogenesis of severe COVID-19 infection: a report of five cases. *Transl Res.* (2020) 220:1–13. doi: 10.1016/j.trsl.2020.04.007

19. Galvan Casas C, Catala A, Carretero Hernandez G, Rodriguez-Jimenez P, Fernandez-Nieto D, Rodriguez-Villa Lario A, et al. Classification of the cutaneous manifestations of COVID-19: a rapid prospective nationwide consensus study in Spain with 375 cases. *Br J Dermatol.* (2020) 183(1):71–7. doi: 10.1111/bjd.19163

20. Alhazzani W, Møller MH, Arabi YM, Loeb M, Gong MN, Fan E, et al. Surviving sepsis campaign: guidelines on the management of critically ill adults with coronavirus disease 2019 (COVID-19). *Crit Care Med.* (2020) 48(6):e440–e69. doi: 10.1097/CCM. 00000000004363

21. Yang L, Liu S, Liu J, Zhang Z, Wan X, Huang B, et al. COVID-19: immunopathogenesis and immunotherapeutics. *Signal Transduction and Targeted Therapy*. (2020) 5(1):128. doi: 10.1038/s41392-020-00243-2

22. Chang JC. Sepsis and septic shock: endothelial molecular pathogenesis associated with vascular microthrombotic disease. *Thromb J.* (2019) 17:10. doi: 10. 1186/s12959-019-0198-4

23. Musher DM. Cutaneous and soft-tissue manifestations of sepsis due to gramnegative enteric bacilli. *Rev Infect Dis.* (1980) 2(6):854–66. doi: 10.1093/clinids/2.6.854

24. Tang N, Li D, Wang X, Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. *J Thromb Haemost.* (2020) 18(4):844–7. doi: 10.1111/jth.14768

25. Goshua G, Pine AB, Meizlish ML, Chang CH, Zhang H, Bahel P, et al. Endotheliopathy in COVID-19-associated coagulopathy: evidence from a single-centre, cross-sectional study. *The Lancet Haematology.* (2020) 7(8):e575–e82. doi: 10.1016/S2352-3026(20)30216-7

26. Arachchillage DJ, Remmington C, Rosenberg A, Xu T, Passariello M, Hall D, et al. Anticoagulation with argatroban in patients with acute antithrombin deficiency in severe COVID-19. *Br J Haematol.* (2020) 190(5):e286–e8. doi: 10. 1111/bjh.16927

27. Boknas N, Laine C, Hillarp A, Macwan AS, Gustafsson KM, Lindahl TL, et al. Associations between hemostatic markers and mortality in COVID-19– compounding effects of D-dimer, antithrombin and PAP complex. *Thromb Res.* (2022) 213:97–104. doi: 10.1016/j.thromres.2022.03.013

28. Han M, Pandey D. ZMPSTE24 Regulates SARS-CoV-2 spike protein-enhanced expression of endothelial PAI-1. *Am J Respir Cell Mol Biol.* (2021) 65(3):300–8. doi: 10. 1165/rcmb.2020-0544OC

29. Zou X, Chen K, Zou J, Han P, Hao J, Han Z. Single-cell RNA-seq data analysis on the receptor ACE2 expression reveals the potential risk of different human organs vulnerable to 2019-nCoV infection. *Front Med.* (2020) 14(2):185–92. doi: 10.1007/s11684-020-0754-0

30. Varga Z, Flammer AJ, Steiger P, Haberecker M, Andermatt R, Zinkernagel AS, et al. Endothelial cell infection and endotheliitis in COVID-19. *Lancet.* (2020) 395 (10234):1417–8. doi: 10.1016/S0140-6736(20)30937-5

31. Evans PC, Rainger GE, Mason JC, Guzik TJ, Osto E, Stamataki Z, et al. Endothelial dysfunction in COVID-19: a position paper of the ESC working group for atherosclerosis and vascular biology, and the ESC council of basic cardiovascular science. *Cardiovasc Res.* (2020) 116(14):2177–84. doi: 10.1093/cvr/ cvaa230

32. Su H, Yang M, Wan C, Yi LX, Tang F, Zhu HY, et al. Renal histopathological analysis of 26 postmortem findings of patients with COVID-19 in China. *Kidney Int.* (2020) 98(1):219–27. doi: 10.1016/j.kint.2020.04.003

33. Connors JM, Levy JH. COVID-19 and its implications for thrombosis and anticoagulation. *Blood.* (2020) 135(23):2033-40. doi: 10.1182/blood.2020006000

34. Huckriede J, Anderberg SB, Morales A, de Vries F, Hultstrom M, Bergqvist A, et al. Evolution of NETosis markers and DAMPs have prognostic value in critically ill COVID-19 patients. *Sci Rep.* (2021) 11(1):15701. doi: 10.1038/s41598-021-95209-x

35. Yao XH, Li TY, He ZC, Ping YF, Liu HW, Yu SC, et al. A pathological report of three COVID-19 cases by minimal invasive autopsies. *Zhonghua Bing Li Xue Za Zhi*. (2020) 49(5):411–7. doi: 10.3760/cma.j.cn112151-20200312-00193

36. Cicco S, Cicco G, Racanelli V, Vacca A. Neutrophil extracellular traps (NETs) and damage-associated molecular patterns (DAMPs): two potential targets for COVID-19 treatment. *Mediators Inflamm.* (2020) 2020:7527953. doi: 10.1155/2020/7527953

37. Parker H, Dragunow M, Hampton MB, Kettle AJ, Winterbourn CC. Requirements for NADPH oxidase and myeloperoxidase in neutrophil extracellular trap formation differ depending on the stimulus. *J Leukoc Biol.* (2012) 92(4):841–9. doi: 10.1189/jlb.1211601

38. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet.* (2020) 395 (10223):497–506. doi: 10.1016/S0140-6736(20)30183-5

39. Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, Manson JJ, et al. COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet*. (2020) 395(10229):1033-4. doi: 10.1016/S0140-6736(20)30628-0

40. Ruan Q, Yang K, Wang W, Jiang L, Song J. Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China. *Intensive Care Med.* (2020) 46(5):846–8. doi: 10.1007/s00134-020-05991-x

41. Skendros P, Mitsios A, Chrysanthopoulou A, Mastellos DC, Metallidis S, Rafailidis P, et al. Complement and tissue factor-enriched neutrophil extracellular traps are key drivers in COVID-19 immunothrombosis. *J Clin Invest.* (2020) 130 (11):6151–7. doi: 10.1172/JCI141374

42. Kang S, Kishimoto T. Interplay between interleukin-6 signaling and the vascular endothelium in cytokine storms. *Exp Mol Med.* (2021) 53(7):1116–23. doi: 10.1038/ s12276-021-00649-0

43. Pedersen SF, Ho YC. SARS-CoV-2: a storm is raging. J Clin Invest. (2020) 130 (5):2202–5. doi: 10.1172/JCI137647

44. Li Y, Schneider AM, Mehta A, Sade-Feldman M, Kays KR, Gentili M, et al. SARS-CoV-2 viremia is associated with distinct proteomic pathways and predicts COVID-19 outcomes. *J Clin Invest.* (2021) 131(13):e148635. doi: 10.1172/JCI148635

45. Fajnzylber J, Regan J, Coxen K, Corry H, Wong C, Rosenthal A, et al. SARS-CoV-2 viral load is associated with increased disease severity and mortality. *Nat Commun.* (2020) 11(1):5493. doi: 10.1038/s41467-020-19057-5

46. Herrera VLM, Bosch NA, Lok JJ, Nguyen MQ, Lenae KA, deKay JT, et al. Circulating neutrophil extracellular trap (NET)-forming "rogue" neutrophil subset, immunotype [DEspR+CD11b+], mediate multi-organ failure in COVID-19-an observational study. *Transl Med Commun.* (2023) 8(1):12. doi: 10.1186/s41231-023-00143-x

47. Krinsky N, Sizikov S, Nissim S, Dror A, Sas A, Prinz H, et al. NETosis induction reflects COVID-19 severity and long COVID: insights from a 2-center patient cohort study in Israel. *J Thromb Haemost.* (2023) 21(9):2569–84. doi: 10.1016/j.jtha.2023.02.033

48. Bauer A, Pachl E, Hellmuth JC, Kneidinger N, Heydarian M, Frankenberger M, et al. Proteomics reveals antiviral host response and NETosis during acute COVID-19 in high-risk patients. *Biochim Biophys Acta Mol Basis Dis.* (2023) 1869(2):166592. doi: 10.1016/j.bbadis.2022.166592

49. Ghanbari EP, Jakobs K, Puccini M, Reinshagen L, Friebel J, Haghikia A, et al. The role of NETosis and complement activation in COVID-19-associated coagulopathies. *Biomedicines*. (2023) 11(5):1371. doi: 10.3390/biomedicines11051371

50. Folco EJ, Mawson TL, Vromman A, Bernardes-Souza B, Franck G, Persson O, et al. Neutrophil extracellular traps induce endothelial cell activation and tissue factor production through interleukin-1alpha and cathepsin G. *Arterioscler Thromb Vasc Biol.* (2018) 38(8):1901–12. doi: 10.1161/ATVBAHA.118.311150

51. Fuchs TA, Brill A, Duerschmied D, Schatzberg D, Monestier M, Myers DD Jr., et al. Extracellular DNA traps promote thrombosis. *Proc Natl Acad Sci U S A*. (2010) 107(36):15880–5. doi: 10.1073/pnas.1005743107

52. Bhargavan B, Kanmogne GD. SARS-CoV-2 spike proteins and cell-cell communication induce P-selectin and markers of endothelial injury, NETosis, and inflammation in human lung microvascular endothelial cells and neutrophils: implications for the pathogenesis of COVID-19 coagulopathy. *Int J Mol Sci.* (2023) 24(16):12585. doi: 10.3390/ijms241612585

53. Gao T, Hu M, Zhang X, Li H, Zhu L, Liu H, et al. Highly pathogenic coronavirus N protein aggravates lung injury by MASP-2-mediated complement over-activation. *medRxiv.* (2020) 03(29):20041962. doi: 10.1038/s41392-022-01133-5

54. Krarup A, Wallis R, Presanis JS, Gal P, Sim RB. Simultaneous activation of complement and coagulation by MBL-associated serine protease 2. *PLoS One.* (2007) 2(7):e623. doi: 10.1371/journal.pone.0000623

55. Ritis K, Doumas M, Mastellos D, Micheli A, Giaglis S, Magotti P, et al. A novel C5a receptor-tissue factor cross-talk in neutrophils links innate immunity to coagulation pathways. J Immunol. (2006) 177(7):4794–802. doi: 10.4049/jimmunol.177.7.4794

56. Zhang J, Tecson KM, McCullough PA. Endothelial dysfunction contributes to COVID-19-associated vascular inflammation and coagulopathy. *Rev Cardiovasc Med.* (2020) 21(3):315–9. doi: 10.31083/j.rcm.2020.03.126

57. Brinjikji W, Kallmes DF, Virmani R, de Meyer SF, Yoo AJ, Humphries W, et al. Endotheliitis and cytokine storm as a mechanism of clot formation in COVID-19 ischemic stroke patients: a histopathologic study of retrieved clots. *Interv Neuroradiol.* (2023):15910199231185804. doi: 10.1177/15910199231185804

58. Sugimoto M, Miyata S. Functional property of von Willebrand factor under flowing blood. Int J Hematol. (2002) 75(1):19–24. doi: 10.1007/BF02981974

59. Cao W, Krishnaswamy S, Camire RM, Lenting PJ, Zheng XL. Factor VIII accelerates proteolytic cleavage of von willebrand factor by ADAMTS13. *Proc Natl Acad Sci U S A*. (2008) 105(21):7416–21. doi: 10.1073/pnas.0801735105

60. Morrell CN, Hilt ZT, Pariser DN, Maurya P. PAD4 and von Willebrand factor link inflammation and thrombosis. *Circ Res.* (2019) 125(5):520–2. doi: 10.1161/CIRCRESAHA.119.315601

61. Mancini I, Baronciani L, Artoni A, Colpani P, Biganzoli M, Cozzi G, et al. The ADAMTS13-von Willebrand factor axis in COVID-19 patients. *J Thromb Haemost.* (2021) 19(2):513–21. doi: 10.1111/jth.15191

62. Karsli E, Sabirli R, Altintas E, Canacik O, Sabirli GT, Kaymaz B, et al. Soluble Pselectin as a potential diagnostic and prognostic biomarker for COVID-19 disease: a case-control study. *Life Sci.* (2021) 277:119634. doi: 10.1016/j.lfs.2021.119634

63. Grobler C, Maphumulo SC, Grobbelaar LM, Bredenkamp JC, Laubscher GJ, Lourens PJ, et al. COVID-19: the rollercoaster of fibrin(ogen), D-dimer, von Willebrand factor, P-selectin and their interactions with endothelial cells, platelets and erythrocytes. *Int J Mol Sci.* (2020) 21(14):5168. doi: 10.3390/jijms21145168 64. Polgar J, Matuskova J, Wagner DD. The P-selectin, tissue factor, coagulation triad. J Thromb Haemost. (2005) 3(8):1590-6. doi: 10.1111/j.1538-7836.2005.01373.x

65. Agrati C, Sacchi A, Tartaglia E, Vergori A, Gagliardini R, Scarabello A, et al. The role of P-selectin in COVID-19 coagulopathy: an updated review. *Int J Mol Sci.* (2021) 22(15):7942. doi: 10.3390/ijms22157942

66. Pober JS, Sessa WC. Evolving functions of endothelial cells in inflammation. Nat Rev Immunol. (2007) 7(10):803–15. doi: 10.1038/nri2171

67. Lippi G, Plebani M, Henry BM. Thrombocytopenia is associated with severe coronavirus disease 2019 (COVID-19) infections: a meta-analysis. *Clin Chim Acta*. (2020) 506:145–8. doi: 10.1016/j.cca.2020.03.022

68. Cicco S, Vacca A, Albanese F, Susca N, Desantis V, Magistro A, et al. Immune disturbance leads to pulmonary embolism in COVID-19 more than classical risk factors: a clinical and histological study. *Intern Emerg Med.* (2023) 18(7):1981–93. doi: 10.1007/s11739-023-03383-9

69. Ranucci M, Ballotta A, Di Dedda U, Baryshnikova E, Dei Poli M, Resta M, et al. The procoagulant pattern of patients with COVID-19 acute respiratory distress syndrome. *J Thromb Haemost.* (2020) 18(7):1747-51. doi: 10.1111/jth.14854

70. Rapkiewicz AV, Mai X, Carsons SE, Pittaluga S, Kleiner DE, Berger JS, et al. Megakaryocytes and platelet-fibrin thrombi characterize multi-organ thrombosis at autopsy in COVID-19: a case series. *EClinicalMedicine*. (2020) 24:100434. doi: 10.1016/j.eclinm.2020.100434

71. Bautista-Vargas M, Bonilla-Abadia F, Canas CA. Potential role for tissue factor in the pathogenesis of hypercoagulability associated with in COVID-19. J Thromb Thrombolysis. (2020) 50(3):479–83. doi: 10.1007/s11239-020-02172-x

72. Sims PJ, Faioni EM, Wiedmer T, Shattil SJ. Complement proteins C5b-9 cause release of membrane vesicles from the platelet surface that are enriched in the membrane receptor for coagulation factor va and express prothrombinase activity. *J Biol Chem.* (1988) 263(34):18205–12. doi: 10.1016/S0021-9258(19)81346-7

73. de Bont CM, Boelens WC, Pruijn GJM. NETosis, complement, and coagulation: a triangular relationship. *Cell Mol Immunol.* (2019) 16(1):19–27. doi: 10.1038/s41423-018-0024-0

74. Polley MJ, Nachman RL. Human platelet activation by C3a and C3a des-arg. J Exp Med. (1983) 158(2):603-15. doi: 10.1084/jem.158.2.603

75. Zhang S, Liu Y, Wang X, Yang L, Li H, Wang Y, et al. SARS-CoV-2 binds platelet ACE2 to enhance thrombosis in COVID-19. *J Hematol Oncol.* (2020) 13 (1):120. doi: 10.1186/s13045-020-00954-7

76. Levi M, van der Poll T. Two-way interactions between inflammation and coagulation. *Trends Cardiovasc Med.* (2005) 15(7):254–9. doi: 10.1016/j.tcm.2005. 07.004

77. Middleton EA, He XY, Denorme F, Campbell RA, Ng D, Salvatore SP, et al. Neutrophil extracellular traps contribute to immunothrombosis in COVID-19 acute respiratory distress syndrome. *Blood.* (2020) 136(10):1169–79. doi: 10.1182/blood. 2020007008

78. van der Meijden PEJ, Heemskerk JWM. Platelet biology and functions: new concepts and clinical perspectives. *Nat Rev Cardiol.* (2019) 16(3):166–79. doi: 10. 1038/s41569-018-0110-0

79. Jiang Y, Rubin L, Zhou Z, Zhang H, Su Q, Hou ST, et al. Pharmacological therapies and drug development targeting SARS-CoV-2 infection. *Cytokine Growth Factor Rev.* (2022) 68:13–24. doi: 10.1016/j.cytogfr.2022.10.003

80. Gottlieb RL, Vaca CE, Paredes R, Mera J, Webb BJ, Perez G, et al. Early remdesivir to prevent progression to severe COVID-19 in outpatients. *N Engl J Med.* (2022) 386(4):305–15. doi: 10.1056/NEJM0a2116846

81. Butler CC, Hobbs FDR, Gbinigie OA, Rahman NM, Hayward G, Richards DB, et al. Molnupiravir plus usual care versus usual care alone as early treatment for adults with COVID-19 at increased risk of adverse outcomes (PANORAMIC): an open-label, platform-adaptive randomised controlled trial. *Lancet.* (2023) 401(10373):281–93. doi: 10.1016/S0140-6736(22)02597-1

82. Mahase E. COVID-19: pfizer's paxlovid is 89% effective in patients at risk of serious illness, company reports. *Br Med J.* (2021) 375:n2713. doi: 10.1136/bmj. n2713

83. Reina J, Iglesias C. Nirmatrelvir plus ritonavir (paxlovid) a potent SARS-CoV-2 3CLpro protease inhibitor combination. *Rev Esp Quimioter*. (2022) 35(3):236–40. doi: 10.37201/req/002.2022

84. Drozdzal S, Rosik J, Lechowicz K, Machaj F, Szostak B, Przybycinski J, et al. An update on drugs with therapeutic potential for SARS-CoV-2 (COVID-19) treatment. *Drug Resist Updat.* (2021) 59:100794. doi: 10.1016/j.drup.2021.100794

85. Kabinger F, Stiller C, Schmitzova J, Dienemann C, Kokic G, Hillen HS, et al. Mechanism of molnupiravir-induced SARS-CoV-2 mutagenesis. *Nat Struct Mol Biol.* (2021) 28(9):740-6. doi: 10.1038/s41594-021-00651-0

86. Khiali S, Khani E SBR, Entezari-Maleki T. Comprehensive review on molnupiravir in COVID-19: a novel promising antiviral to combat the pandemic. *Future Microbiol.* (2022) 17(5):377–91. doi: 10.2217/fmb-2021-0252

87. Kokic G, Hillen HS, Tegunov D, Dienemann C, Seitz F, Schmitzova J, et al. Mechanism of SARS-CoV-2 polymerase stalling by remdesivir. *Nat Commun.* (2021) 12(1):279. doi: 10.1038/s41467-020-20542-0 88. Eweas AF, Alhossary AA, Abdel-Moneim AS. Molecular docking reveals ivermectin and remdesivir as potential repurposed drugs against SARS-CoV-2. *Front Microbiol.* (2020) 11:592908. doi: 10.3389/fmicb.2020.592908

89. Lee TC, Murthy S, Del Corpo O, Senecal J, Butler-Laporte G, Sohani ZN, et al. Remdesivir for the treatment of COVID-19: a systematic review and meta-analysis. *Clin Microbiol Infect*. (2022) 28(9):1203–10. doi: 10.1016/j.cmi.2022.04.018

90. Zarychanski R, Abou-Setta AM, Kanji S, Turgeon AF, Kumar A, Houston DS, et al. The efficacy and safety of heparin in patients with sepsis: a systematic review and metaanalysis. *Crit Care Med.* (2015) 43(3):511–8. doi: 10.1097/CCM.000000000000763

91. Bauersachs RM. Fondaparinux sodium: recent advances in the management of thrombosis. *J Cardiovasc Pharmacol Ther*. (2023) 28:10742484221145010. doi: 10. 1177/10742484221145010

92. Bauer KA, Hawkins DW, Peters PC, Petitou M, Herbert JM, van Boeckel CA, et al. Fondaparinux, a synthetic pentasaccharide: the first in a new class of antithrombotic agents—the selective factor Xa inhibitors. *Cardiovasc Drug Rev.* (2002) 20(1):37–52. doi: 10.1111/j.1527-3466.2002.tb00081.x

93. Alam W. COVID-19 vaccine-induced immune thrombotic thrombocytopenia: a review of the potential mechanisms and proposed management. *Sci Prog.* (2021) 104 (2):368504211025927. doi: 10.1177/00368504211025927

94. Russo V, Cardillo G, Viggiano GV, Mangiacapra S, Cavalli A, Fontanella A, et al. Thromboprofilaxys with fondaparinux vs. Enoxaparin in hospitalized COVID-19 patients: a multicenter Italian observational study. *Front Med (Lausanne)*. (2020) 7:569567. doi: 10.3389/fmed.2020.569567

95. Ertan-Bolelli T, Bolelli K, Elci SD, Belen-Apak FB. Promising drug fondaparinux for the treatment of COVID-19: an in silico analysis of low molecular weight heparin, direct oral anticoagulant, and antiplatelet drug interactions with host protease furin. *Cardiovasc Drugs Ther.* (2022):1–8. doi: 10.1007/s10557-022-07406-z

96. Cardillo G, Viggiano GV, Russo V, Mangiacapra S, Cavalli A, Castaldo G, et al. Antithrombotic and anti-inflammatory effects of fondaparinux and enoxaparin in hospitalized COVID-19 patients: the FONDENOXAVID study. *J Blood Med.* (2021) 12:69–75. doi: 10.2147/JBM.S285214

97. Wada H, Thachil J, Di Nisio M, Mathew P, Kurosawa S, Gando S, et al. Guidance for diagnosis and treatment of DIC from harmonization of the recommendations from three guidelines. *J Thromb Haemost.* (2013) 11(4):761–67. doi: 10.1111/jth.12155

98. Kienast J, Juers M, Wiedermann CJ, Hoffmann JN, Ostermann H, Strauss R, et al. Treatment effects of high-dose antithrombin without concomitant heparin in patients with severe sepsis with or without disseminated intravascular coagulation. *J Thromb Haemost.* (2006) 4(1):90–7. doi: 10.1111/j.1538-7836.2005.01697.x

99. Warren BL, Eid A, Singer P, Pillay SS, Carl P, Novak I, et al. Caring for the critically ill patient. High-dose antithrombin III in severe sepsis: a randomized controlled trial. *JAMA*. (2001) 286(15):1869–78. doi: 10.1001/jama.286.15.1869

100. Ranieri VM, Thompson BT, Barie PS, Dhainaut JF, Douglas IS, Finfer S, et al. Drotrecogin alfa (activated) in adults with septic shock. *N Engl J Med.* (2012) 366 (22):2055–64. doi: 10.1056/NEJMoa1202290

101. de Brabander J, Michels EHA, van Linge CCA, Chouchane O, Douma RA, Reijnders TDY, et al. Association between dexamethasone treatment and the host response in COVID-19 patients admitted to the general ward. *Respir Res.* (2022) 23 (1):145. doi: 10.1186/s12931-022-02060-3

102. Zhang G, Zhang X, Huang H, Ji Y, Li D, Jiang W. Saquinavir plus methylprednisolone ameliorates experimental acute lung injury. *Braz J Med Biol Res.* (2018) 51(10):e7579. doi: 10.1590/1414-431x20187579

103. Zhao S, Yang J, Liu T, Zeng J, Mi L, Xiang K. Dexamethasone inhibits NF- κ Bp65 and HMGB1 expression in the pancreas of rats with severe acute pancreatitis. *Mol Med Rep.* (2018) 18(6):5345–52. doi: 10.3892/mmr.2018.9595

104. Veronese N, Demurtas J, Yang L, Tonelli R, Barbagallo M, Lopalco P, et al. Use of corticosteroids in coronavirus disease 2019 pneumonia: a systematic review of the literature. *Front Med (Lausanne)*. (2020) 7:170. doi: 10.3389/fmed.2020.00170

105. Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. *JAMA*. (2020) 323(11):1061–9. doi: 10.1001/jama.2020.1585

106. Ling Y, Xu SB, Lin YX, Tian D, Zhu ZQ, Dai FH, et al. Persistence and clearance of viral RNA in 2019 novel coronavirus disease rehabilitation patients. *Chin Med J (Engl).* (2020) 133(9):1039-43. doi: 10.1097/CM9.00000000000774

107. Wu C, Chen X, Cai Y, Xia J, Zhou X, Xu S, et al. Risk factors associated with acute respiratory distress syndrome and death in patients with coronavirus disease 2019 pneumonia in Wuhan, China. *JAMA Intern Med.* (2020) 180(7):934–43. doi: 10.1001/jamainternmed.2020.0994

108. Fadel R, Morrison AR, Vahia A, Smith ZR, Chaudhry Z, Bhargava P, et al. Early short-course corticosteroids in hospitalized patients with COVID-19. *Clin Infect Dis.* (2020) 71(16):2114–20. doi: 10.1093/cid/ciaa601

109. Wang Y, Jiang W, He Q, Wang C, Wang B, Zhou P, et al. A retrospective cohort study of methylprednisolone therapy in severe patients with COVID-19 pneumonia. *Signal Transduct Target Ther.* (2020) 5(1):57. doi: 10.1038/s41392-020-0158-2

110. van der Poll T, van de Veerdonk FL, Scicluna BP, Netea MG. The immunopathology of sepsis and potential therapeutic targets. *Nat Rev Immunol.* (2017) 17(7):407–20. doi: 10.1038/nri.2017.36

111. Group RC. Tocilizumab in patients admitted to hospital with COVID-19 (RECOVERY): a randomised, controlled, open-label, platform trial. *Lancet.* (2021) 397(10285):1637-45. doi: 10.1016/S0140-6736(21)00676-0

112. Richardson P, Griffin I, Tucker C, Smith D, Oechsle O, Phelan A, et al. Baricitinib as potential treatment for 2019-nCoV acute respiratory disease. *Lancet*. (2020) 395(10223):e30-e1. doi: 10.1016/S0140-6736(20)30304-4

113. Hallberg LAE, Barlous K, Hawkins CL. Antioxidant strategies to modulate NETosis and the release of neutrophil extracellular traps during chronic inflammation. *Antioxidants (Basel)*. (2023) 12(2):478. doi: 10.3390/antiox12020478

114. Ostafin M, Pruchniak MP, Ciepiela O, Reznick AZ, Demkow U. Different procedures of diphenyleneiodonium chloride addition affect neutrophil extracellular trap formation. *Anal Biochem.* (2016) 509:60–6. doi: 10.1016/j.ab.2016.05.003

115. Menegazzo L, Scattolini V, Cappellari R, Bonora BM, Albiero M, Bortolozzi M, et al. The antidiabetic drug metformin blunts NETosis in vitro and reduces circulating NETosis biomarkers in vivo. *Acta Diabetol.* (2018) 55(6):593–601. doi: 10.1007/s00592-018-1129-8

116. Zawrotniak M, Kozik A, Rapala-Kozik M. Selected mucolytic, antiinflammatory and cardiovascular drugs change the ability of neutrophils to form extracellular traps (NETs). *Acta Biochim Pol.* (2015) 62(3):465–73. doi: 10.18388/ abp.2015_1055

117. de Souza Andrade MM, Leal VNC, Fernandes IG, Gozzi-Silva SC, Beserra DR, Oliveira EA, et al. Resveratrol downmodulates neutrophil extracellular trap (NET) generation by neutrophils in patients with severe COVID-19. *Antioxidants (Basel).* (2022) 11(9):1690. doi: 10.3390/antiox11091690

118. Zhang Y, Cartland SP, Henriquez R, Patel S, Gammelgaard B, Flouda K, et al. Selenomethionine supplementation reduces lesion burden, improves vessel function and modulates the inflammatory response within the setting of atherosclerosis. *Redox Biol.* (2020) 29:101409. doi: 10.1016/j.redox.2019.101409

119. Kirchner T, Hermann E, Moller S, Klinger M, Solbach W, Laskay T, et al. Flavonoids and 5-aminosalicylic acid inhibit the formation of neutrophil extracellular traps. *Mediators Inflamm.* (2013) 2013:710239. doi: 10.1155/2013/710239

120. Diurno F, Numis FG, Porta G, Cirillo F, Maddaluno S, Ragozzino A, et al. Eculizumab treatment in patients with COVID-19: preliminary results from real life ASL Napoli 2 Nord experience. *Eur Rev Med Pharmacol Sci.* (2020) 24(7):4040–7. doi: 10.26355/eurrev_202004_20875

121. Liu PP, Blet A, Smyth D, Li H. The science underlying COVID-19: implications for the cardiovascular system. *Circulation*. (2020) 142(1):68–78. doi: 10.1161/CIRCULATIONAHA.120.047549

122. Gloude NJ, Khandelwal P, Luebbering N, Lounder DT, Jodele S, Alder MN, et al. Circulating dsDNA, endothelial injury, and complement activation in thrombotic microangiopathy and GVHD. *Blood.* (2017) 130(10):1259–66. doi: 10. 1182/blood-2017-05-782870

123. Rotz SJ, Dandoy CE, Davies SM. ST2 and endothelial injury as a link between GVHD and microangiopathy. *N Engl J Med.* (2017) 376(12):1189–90. doi: 10.1056/ NEJMc1700185

124. Yasmeen S, Akram BH, Hainsworth AH, Kruuse C. Cyclic nucleotide phosphodiesterases (PDEs) and endothelial function in ischaemic stroke. A review. *Cell Signal.* (2019) 61:108–19. doi: 10.1016/j.cellsig.2019.05.011

125. Rushworth GF, Megson IL. Existing and potential therapeutic uses for Nacetylcysteine: the need for conversion to intracellular glutathione for antioxidant benefits. *Pharmacol Ther*. (2014) 141(2):150–9. doi: 10.1016/j.pharmthera.2013.09.006

126. Cabanillas G, Popescu-Martinez A. N-Acetylcysteine for relapsing thrombotic thrombocytopenic Purpura: more evidence of a promising drug. *Am J Ther.* (2016) 23 (5):e1277–9. doi: 10.1097/MJT.0000000000386

127. Pan T, Qi J, Tang Y, Yao Y, Chen J, Wang H, et al. N-acetylcysteine as prophylactic therapy for transplantation-associated thrombotic microangiopathy: a randomized, placebo-controlled trial. *Transplant Cell Ther.* (2022) 28(11):764.e1-e7. doi: 10.1016/j.jtct.2022.07.029

128. Guglielmetti G, Quaglia M, Sainaghi PP, Castello LM, Vaschetto R, Pirisi M, et al. "War to the knife" against thromboinflammation to protect endothelial function of COVID-19 patients. *Crit Care.* (2020) 24(1):365. doi: 10.1186/s13054-020-03060-9

129. Liu X, Li Z, Liu S, Sun J, Chen Z, Jiang M, et al. Potential therapeutic effects of dipyridamole in the severely ill patients with COVID-19. *Acta Pharm Sin B*. (2020) 10 (7):1205–15. doi: 10.1016/j.apsb.2020.04.008

130. Yoshikawa T, Murakami M, Furukawa Y, Takemura S, Kondo M. Effect of dipyridamole on experimental disseminated intravascular coagulation in rats. *Thromb Res.* (1983) 29(6):619–25. doi: 10.1016/0049-3848(83)90216-5

131. Chow JH, Khanna AK, Kethireddy S, Yamane D, Levine A, Jackson AM, et al. Aspirin use is associated with decreased mechanical ventilation, intensive care unit admission, and in-hospital mortality in hospitalized patients with coronavirus disease 2019. *Anesth Analg.* (2021) 132(4):930–41. doi: 10.1213/ANE.00000000005292

132. Chow JH, Yin Y, Yamane DP, Davison D, Keneally RJ, Hawkins K, et al. Association of prehospital antiplatelet therapy with survival in patients hospitalized

with COVID-19: a propensity score-matched analysis. J Thromb Haemost. (2021) 19(11):2814–24. doi: 10.1111/jth.15517

133. Osborne TF, Veigulis ZP, Arreola DM, Mahajan SM, Roosli E, Curtin CM. Association of mortality and aspirin prescription for COVID-19 patients at the veterans health administration. *PLoS One.* (2021) 16(2):e0246825. doi: 10.1371/journal.pone.0246825

134. Dalbeni A, Susca N, Daidone M, Rossi I, Giontella A, Cimellaro A, et al. Low dose aspirin and clinical outcomes in patients with SARS-CoV-2 pneumonia: a propensity score-matched cohort analysis from the national SIMI-COVID-19 registry. *Intern Emerg Med.* (2023) 18(8):2311–19. doi: 10.1007/s11739-023-03432-3

135. Wang J, Hajizadeh N, Moore EE, McIntyre RC, Moore PK, Veress LA, et al. Tissue plasminogen activator (tPA) treatment for COVID-19 associated acute respiratory distress syndrome (ARDS): a case series. *J Thromb Haemost.* (2020) 18 (7):1752–5. doi: 10.1111/jth.14828

136. Global Use of Strategies to Open Occluded Coronary Arteries I. A comparison of reteplase with alteplase for acute myocardial infarction. *N Engl J Med.* (1997) 337 (16):1118–23. doi: 10.1056/NEJM199710163371603

137. Chowdary P, Agarwal B, Peralta MR, Bhagani S, Lee S, Goldring J, et al. Nebulized recombinant tissue plasminogen activator (rt-PA) for acute COVID-19induced respiratory failure: an exploratory proof-of-concept trial. *J Clin Med.* (2023) 12(18):5848. doi: 10.3390/jcm12185848

138. Asakura H, Ogawa H. Perspective on fibrinolytic therapy in COVID-19: the potential of inhalation therapy against suppressed-fibrinolytic-type DIC. *J Intensive Care.* (2020) 8:71. doi: 10.1186/s40560-020-00491-y

139. Saha M, McDaniel JK, Zheng XL. Thrombotic thrombocytopenic purpura: pathogenesis, diagnosis and potential novel therapeutics. *J Thromb Haemost.* (2017) 15(10):1889–900. doi: 10.1111/jth.13764

140. Ni L, Ye F, Cheng ML, Feng Y, Deng YQ, Zhao H, et al. Detection of SARS-CoV-2-specific humoral and cellular immunity in COVID-19 convalescent individuals. *Immunity*. (2020) 52(6):971–7.e3. doi: 10.1016/j.immuni.2020.04.023

141. Jawhara S. Could intravenous immunoglobulin collected from recovered coronavirus patients protect against COVID-19 and strengthen the immune system of new patients? *Int J Mol Sci.* (2020) 21(7):2272. doi: 10.3390/ijms21072272

142. Uozumi R, Iguchi R, Masuda S, Nishibata Y, Nakazawa D, Tomaru U, et al. Pharmaceutical immunoglobulins reduce neutrophil extracellular trap formation and ameliorate the development of MPO-ANCA-associated vasculitis. *Mod Rheumatol.* (2020) 30(3):544–50. doi: 10.1080/14397595.2019.1602292

143. Murakami K, Suzuki C, Fujii A, Kobayashi F, Nakano A, Kamizono A. Intravenous immunoglobulin preparation prevents the production of proinflammatory cytokines by modulating NFxB and MAPKs pathways in the human monocytic THP-1 cells stimulated with procalcitonin. *Inflamm Res.* (2014) 63 (9):711–8. doi: 10.1007/s00011-014-0744-2

144. Lok KZ, Basta M, Manzanero S, Arumugam TV. Intravenous immunoglobulin (IVIg) dampens neuronal toll-like receptor-mediated responses in ischemia. *J Neuroinflammation*. (2015) 12:73. doi: 10.1186/s12974-015-0294-8

145. Marcec R, Dodig VM, Radanovic I, Likic R. Intravenous immunoglobulin (IVIg) therapy in hospitalised adult COVID-19 patients: a systematic review and meta-analysis. *Rev Med Virol.* (2022) 32(6):e2397. doi: 10.1002/rmv.2397

146. Xiang HR, Cheng X, Li Y, Luo WW, Zhang QZ, Peng WX. Efficacy of IVIG (intravenous immunoglobulin) for corona virus disease 2019 (COVID-19): a metaanalysis. *Int Immunopharmacol.* (2021) 96:107732. doi: 10.1016/j.intimp.2021.107732

147. Kwon JW, Hong MK, Park BY. Risk factors of vasopressor-induced symmetrical peripheral gangrene. *Ann Plast Surg.* (2018) 80(6):622–7. doi: 10.1097/SAP.000000000001314

148. Twiford TW Jr., Granmayeh M, Tucker MJ. Gangrene of the feet associated with mesenteric intraarterial vasopressin. *AJR Am J Roentgenol.* (1978) 130 (3):558–60. doi: 10.2214/ajr.130.3.558

149. Winkler MJ, Trunkey DD. Dopamine gangrene. Association with disseminated intravascular coagulation. *Am J Surg.* (1981) 142(5):588–91. doi: 10.1016/0002-9610 (81)90432-3

150. Hayes MA, Yau EH, Hinds CJ, Watson JD. Symmetrical peripheral gangrene: association with noradrenaline administration. *Intensive Care Med.* (1992) 18 (7):433–6. doi: 10.1007/BF01694349

151. Kempczinski RF, Buckley CJ, Darling RC. Vascular insufficiency secondary to ergotism. *Surgery*. (1976) 79(5):597–600. doi: 10.5555/uri:pii:0039606076903731

152. Albano MN, Brazao SG, Caroco TV, Louro JM, Coelho MI, Costa Almeida CE, et al. Rare case of symmetrical peripheral gangrene due to septic shock, disseminated intravascular coagulation and inotropic use. *Ann Med Surg (Lond)*. (2018) 35:103–7. doi: 10.1016/j.amsu.2018.09.025

153. Denning DW, Gilliland L, Hewlett A, Hughes LO, Reid CD. Peripheral symmetrical gangrene successfully treated with epoprostenol and tissue plasminogen activator. *Lancet.* (1986) 2(8520):1401–2. doi: 10.1016/S0140-6736(86)92046-5

154. Smaoui F, Koubaa M, Rekik K, Mejdoub Y, Mezghani S, Maaloul I, et al. Symmetrical peripheral gangrene: 4 cases. *Ann Dermatol Venereol.* (2018) 145 (2):95–9. doi: 10.1016/j.annder.2017.09.592