Check for updates

OPEN ACCESS

EDITED BY Lars Norgren, Örebro University, Sweden

REVIEWED BY Li Lubin, Yantai Yuhuangding Hospital, China Junjun Liu, The Affiliated Hospital of Qingdao University, China

*CORRESPONDENCE Hongyu Wang Ize dr.hongyuwang@foxmail.com

[†]These authors have contributed equally to this work

RECEIVED 14 October 2024 ACCEPTED 04 April 2025 PUBLISHED 24 April 2025

CITATION

Wen H, Su B, Liu J and Wang H (2025) Novel insights into the association between organ damage and inflammatory response in preoperative abdominal aortic aneurysms. Front. Cardiovasc. Med. 12:151112. doi: 10.3389/fcvm.2025.151112

COPYRIGHT

© 2025 Wen, Su, Liu and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Novel insights into the association between organ damage and inflammatory response in preoperative abdominal aortic aneurysms

Huan Wen^{1,2†}, Bo Su^{3†}, Jinbo Liu^{1,2} and Hongyu Wang^{1,2,4,5,6,7,8}*

¹Department of Vascular Medicine, Peking University Shougang Hospital, Beijing, China, ²Beijing Shijingshan District Key Clinical Specialty of Vascular Medicine, Beijing, China, ³Institute of Clinical Pharmacology, Peking University First Hospital, Beijing, China, ⁴Center for Vascular Health Research, Peking University School of Medicine, Beijing, China, ⁶National Key Laboratory of Vascular Homeostasis and Remodeling, Peking University, Beijing, China, ⁶Center for Heart and Vascular Health, Peking University Clinical Research Institute, Beijing, China, ⁷Heart and Vascular Health Research Center of Chengdu Medical College (HVHRC-CMC), Chengdu, China, ⁸Intelligent Heart and Vascular Health Digital Management Research Center, National Institute of Health Data Science At Peking University, Beijing, China

Background: Abdominal aortic aneurysm (AAA) is a life-threatening condition in the elderly population. The insidious nature of AAA onset makes early detection difficult. Currently, there are few studies on changes in laboratory parameters during AAA development.

Methods: This study included 55 elderly patients with AAA who were admitted to the Department of Vascular Medicine, Shougang Hospital, Peking University 2021–2022. Propensity score matching (PSM) in a 1:1 ratio was performed to match the 55 patients and 1,031 controls. In this population of AAA, correlation and regression analyses were used to explore the association between the level of inflammation and each laboratory parameter.

Results: Compared to the control group, significant differences in inflammatory markers, transaminase and bilirubin levels, blood urea nitrogen (BUN) and creatinine (Cr) levels, and ankle-brachial index were found in the aneurysm group. After PSM, the differences between the two groups for each parameter remained statistically significant. Correlation and regression analyses showed a weak positive correlation between the inflammatory index and the BUN and Cr levels (correlation coefficient = 0.22).

Conclusions: Our study demonstrates the presence of a highly inflammatory state and damage to various organs in patients with AAA. This hyperinflammatory state may be associated with kidney injury and is a cause of concern.

KEYWORDS

abdominal aortic aneurysm, inflammation, blood urea nitrogen, creatinine, kidney injury

Introduction

Abdominal aortic aneurysm (AAA) is a serious vascular disease with a prevalence that increases with age and is particularly significant in the elderly population (1, 2). The incidence of AAA is higher in males than in females, and tends to increase annually (3, 4). Smoking, hypertension, coronary heart disease (CHD), and genetics are known risk factors for AAA (5, 6). An aneurysm is formed when the abdominal aorta

undergoes permanent restrictive dilatation with an increase in vessel diameter greater than 50%, and the larger the diameter of the aneurysm, the greater the risk of rupture (2, 7). AAA has no obvious symptoms in the early stages, but pain, nausea, and vomiting may occur when the aneurysm increases in size (8, 9). The diagnosis of AAA is dependent on imaging, and early detection and diagnosis are key to its treatment (10).

Inflammation is the defensive response of the body to stimuli (11, 12). Inflammation plays an important role in the development of cardiovascular disease (13). Laboratory tests, including routine blood tests, C-reactive protein, blood sedimentation, and other inflammatory indicators, can assess the inflammatory status of the body (14). When the body releases large amounts of inflammatory mediators and cytokines, this hyperinflammatory state triggers a series of clinical signs and symptoms (15). Previous studies have shown that a more severe preoperative inflammatory response in patients with AAA can lead to an increase in postoperative complications, thereby affecting patient prognosis (16, 17). However, although studies have focused on the relationship between preoperative inflammatory response and prognosis in AAA, there is still a relative lack of studies on the association between preoperative inflammatory response and changes in organ function in AAA. This study aimed to investigate the relationship between preoperative inflammatory markers of AAA and various laboratory parameters and to assess functional changes in the organ.

Methods

Data source and study design

The population for this analysis was derived from patients with AAA admitted to the Department of Vascular Medicine at Shougang Hospital of Peking University, and the diagnosis of AAA was confirmed after rigorous imaging evaluation. AAA was diagnosed in patients with more than 50% increase in abdominal aortic diameter on the imaging report, and 55 patients who came to the hospital from February 2021 to December 2022 were finally included. More than 90% of these patients had taken antihypertensive and lipid-lowering medications prior to diagnosis. The control population for AAA, also from this vascular department, was selected from those who came to the clinic between 2011 and 2015 but whose imaging assessment was not an aneurysm, and a total of 1,031 were included. The Ethics Committee of Shougang Hospital of Peking University approved the study protocol. All the participants provided written informed consent.

Variable collection

The medical registration of patients with AAA and the control population was verified by two physicians to collect information about the participants' sex, age, smoking habits, and previous illnesses. Current smoking was defined as having smoked > 100 cigarettes in a lifetime and currently still smoked daily (18). The

diagnosis of previous diseases, such as hypertension, diabetes, and CHD, depended on the participant being told by a doctor or being on relevant medication themselves (19).

Anthropometric measurements were performed by physicians in the department. Body mass index (BMI) was determined by dividing the participant's weight in kilograms by the square of height in meters. Brachial and ankle artery systolic blood pressure (SBP) measurements were performed for each participant using a mercury sphygmomanometer, and the anklebrachial index (ABI) was determined by dividing the highest ankle artery SBP by the highest brachial artery SBP. Before performing the measurements, the participants were asked to rest in the supine position for at least 10 min while ABI was measured on the right and left sides. The ABI of the participants in the study was calculated as the mean of the left and right sides. ABI reflects the peripheral vascular function (20).

After overnight fasting, venous blood samples were collected from the participants to measure various biochemical markers, including C-reactive protein (CRP), alanine transaminase (ALT), aspartate transaminase (AST), blood urea nitrogen (BUN), creatinine (Cr), total bilirubin (TBil), and direct bilirubin (DBil). CRP, ALT, AST, BUN, Cr, TBil, and DBil levels were measured in the laboratory of the Shougang Hospital of Peking University using standard methods. ALT, AST, TBil, and DBil levels reflect liver function. BUN and Cr levels reflect renal function. The venous blood was divided into different tubes according to the requirements of the laboratory, and then tested using a fully automated biochemical analyzer (Hitachi 7,170) manufactured by Hitachi, Tokyo, Japan, based on the principle of spectrophotometry. Platelet, neutrophil, and lymphocyte counts were obtained from all participants. Systemic immune inflammatory index SII = platelet count $(\times 10^{-9}/L) \times absolute$ neutrophil count ($\times 10^{-9}/L$)/absolute lymphocyte count ($\times 10^{-9}/L$) (21). CRP and SII levels reflect the systemic inflammatory status of the participants (22).

Statistical analysis

Data processing and statistical analyses were performed using IBM SPSS version 27 and R software version 4.3.2. Plotting was performed using the R software and GraphPad Prism version 9.5.1. Categorical variables were expressed as frequencies and percentages. Continuous variables showing a normal distribution were expressed as mean ± standard deviation, while continuous variables showing a skewed distribution were expressed as medians and interquartile ranges. Because SII showed a skewed distribution, all SII values were converted to logarithmic values with a base of 10 and expressed using LogSII, which is normally distributed. The chi-square test was used to compare categorical variables. In addition, the independent t-test and Mann–Whitney test were used to compare normally and non-normally distributed continuous variables, respectively.

In the present study, propensity score matching (PSM) was performed in a 1:1 ratio without replacement using the MatchIt R software package (version 11). Matching was performed using the nearest-neighbor principle, which matches patients with aneurysm and control populations based on propensity scores calculated from a combination of variables. This rigorous approach ensured that the case and control groups were effectively balanced in terms of their propensity scores. Variables used to calculate the propensity score included sex, age, BMI, smoking history, and history of previous disease (23), thus accounting for key factors that may have influenced the results of the study.

After PSM, histograms and densitograms were used to compare the levels of inflammation between the case and control groups. Correlation and regression analyses were used to analyze the association between inflammatory indicators and each laboratory parameter in the case group. Statistical p-value < 0.05.

Results

Characteristics of the study population

The clinical and laboratory characteristics of the AAA and control groups are shown in Table 1. 55 patients had a mean age of 75.0 ± 8.2 years, whereas 1,031 controls had a mean age of 64.2 ± 12.1 years, and the proportion of males was higher in the case group than in the control group. This is consistent with epidemiological data on AAA. History of smoking and hypertension was statistically different between the two groups.

TABLE 1 Bas	ic characteristics	of the study	population	according to	state	
of disease.						

Variables	Aneurysm (<i>N</i> = 55)	Control (N = 1,031)	<i>P</i> -value	
Age, years	75.0 ± 8.2	64.2 ± 12.1	< 0.001	
Men, n (%)	42 (76.4)	468 (45.4)	< 0.001	
BMI, kg/m ²	24.5 ± 3.3	25.3 ± 3.5	0.081	
Current smoking, n (%)	35 (63.6)	154 (14.9)	< 0.001	
Hypertension, n (%)	48 (87.3)	672 (65.2)	< 0.001	
DM, n (%)	18 (32.7)	289 (28.0)	0.451	
CHD, n (%)	31 (56.4)	438 (42.5)	0.043	
CRP, mg/L	15.1 (5.4, 27.8)	1.7 (0.8, 3.5)	< 0.001	
Neutrophils, ×10 ⁹ /L	7.9 ± 2.5	3.9 ± 1.5	< 0.001	
Lymphocytes, ×10 ⁹ /L	1.4 ± 0.3	1.7 ± 0.6	< 0.001	
Platelet, ×10 ⁹ /L	222.3 ± 42.5	201.8 ± 52.4	0.004	
SII, ×10 ⁹ /L	1197.0 (911.8, 1749.4)	444.5 (305.1, 646.0)	< 0.001	
LogSII	3.1 ± 0.2	2.7 ± 0.3	< 0.001	
ALT, U/L	30.9 ± 15.7	21.0 ± 12.7	< 0.001	
AST, U/L	27.5 ± 11.4	24.4 ± 8.4	0.009	
BUN, mmol/L	7.0 ± 1.8	5.5 ± 1.8	< 0.001	
Cr, µmol/L	103.1 ± 35.7	69.3 ± 22.2	< 0.001	
TBil, mmol/L	15.0 ± 4.7	13.3 ± 4.8	0.010	
DBil, mmol/L	5.2 ± 1.7	3.8 ± 1.6	< 0.001	
R-ABI	0.9 ± 0.2	1.1 ± 0.2	< 0.001	
L-ABI	0.9 ± 0.2	1.1 ± 0.2	< 0.001	

The values in the table are presented as the mean \pm standard deviation, median (interquartile range), or *n* (%). BMI, body mass index; DM, diabetes mellitus; CHD, coronary heart disease; CRP, C-reactive protein; SII, systemic immune-inflammation index; ALT, alanine aminotransferase; AST, aspartate aminotransferase; BUN, blood urea nitrogen; Cr, creatinine; TBil, total bilirubin; DBil, direct bilirubin; R-ABI, right ankle brachial index; L-ABI, left ankle brachial index.

CRP and SII levels were compared between the two groups, and inflammation levels were significantly higher in the case group than in the control group. There were statistically significant differences in ALT, AST, BUN, Cr, TBil, DBil levels, and ABI. Compared with the control group, the case group may have impaired liver, kidney, and vascular functions.

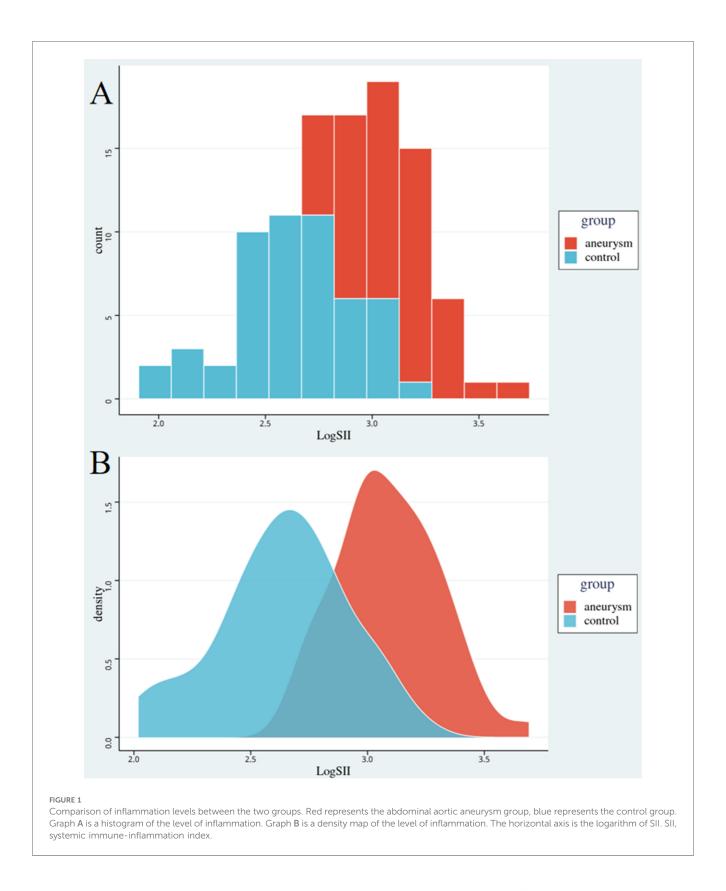
Propensity-matched analysis

Table 2 shows the clinical and laboratory characteristics of the AAA and control groups after the PSM. The mean age of the 52 patients was 74.8 ± 8.2 years, whereas that of the 52 controls was 73.7 ± 9.6 years. Statistically insignificant differences in sex ratio, smoking history, and history of hypertension were observed in the case group compared with the control group, which in turn attenuated the interference of relevant factors in the comparison of inflammation levels between the two groups. Comparison of CRP and SII levels between the two groups showed that the inflammation level in the case group remained significantly higher than that in the control group. Statistically significant differences were found in ALT, AST, Cr, and DBil levels, and ABI. Statistical differences in BUN and TBil levels were attenuated in both the groups. Figure 1 shows LogSII to represent the level of inflammation, demonstrating the hyperinflammatory state of the patient group.

TABLE 2	Basic	characterist	ics of	the	study	population	according	to	state
of diseas	e afte	r propensity	score	ma	tching	(1:1).			

Variables	Aneurysm (<i>N</i> = 52)	Control (<i>N</i> = 52)	<i>P</i> -value
Age, years	74.8 ± 8.2	73.7 ± 9.6	0.511
Men, n (%)	39 (75.0)	37 (71.2)	0.658
BMI, kg/m ²	24.5 ± 3.4	24.0 ± 2.9	0.469
Current smoking, n (%)	32 (61.5)	34 (65.4)	0.684
Hypertension, n (%)	45 (86.5)	45 (86.5)	1.000
DM, n (%)	16 (30.8)	17 (32.7)	0.833
CHD, n (%)	29 (55.8)	29 (55.8)	1.000
CRP, mg/L	15.4 (5.5, 28.3)	1.9 (1.2, 3.6)	< 0.001
Neutrophils, ×10 ⁹ /L	8.0 ± 2.5	4.1 ± 1.5	< 0.001
Lymphocytes, ×10 ⁹ /L	1.4 ± 0.3	1.7 ± 0.6	0.007
Platelet, ×10 ⁹ /L	221.4 ± 43.2	188.1 ± 49.0	< 0.001
SII, ×10 ⁹ /L	1195.9 (913.5, 1743.6)	448.8 (304.5, 655.1)	< 0.001
LogSII	3.1 ± 0.2	2.6 ± 0.3	< 0.001
ALT, U/L	30.3 ± 15.5	17.6 ± 10.8	< 0.001
AST, U/L	27.1 ± 11.4	22.7 ± 6.4	0.016
BUN, mmol/L	7.0 ± 1.9	6.5 ± 2.9	0.243
Cr, µmol/L	104.2 ± 36.4	84.1 ± 37.2	0.006
TBil, mmol/L	14.9 ± 4.7	13.9 ± 5.2	0.334
DBil, mmol/L	5.1 ± 1.7	4.2 ± 1.7	0.008
R-ABI	0.9 ± 0.2	1.0 ± 0.2	0.024
L-ABI	0.9 ± 0.2	1.0 ± 0.2	0.034

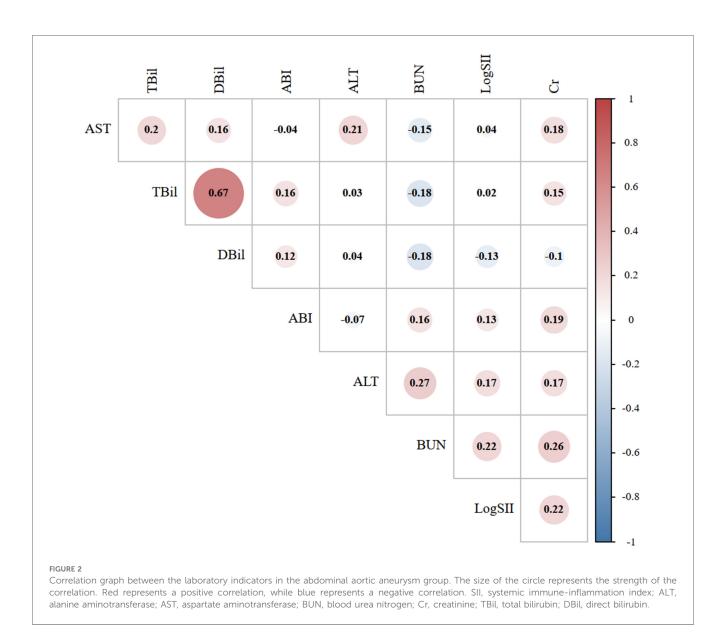
The values in the table are presented as the mean \pm standard deviation, median (interquartile range), or *n* (%). BMI, body mass index; DM, diabetes mellitus; CHD, coronary heart disease; CRP, C-reactive protein; SII, systemic immune-inflammation index; ALT, alanine aminotransferase; AST, aspartate aminotransferase; BUN, blood urea nitrogen; Cr, creatinine; TBil, total bilirubin; DBil, direct bilirubin; R-ABI, right ankle brachial index; L-ABI, left ankle brachial index.



Correlation and regression analysis

Figure 2 shows the correlation analysis between LogSII and ALT, AST, BUN, Cr, TBil, DBil, and ABI in the case group. The highest correlation was found between LogSII and BUN and Cr

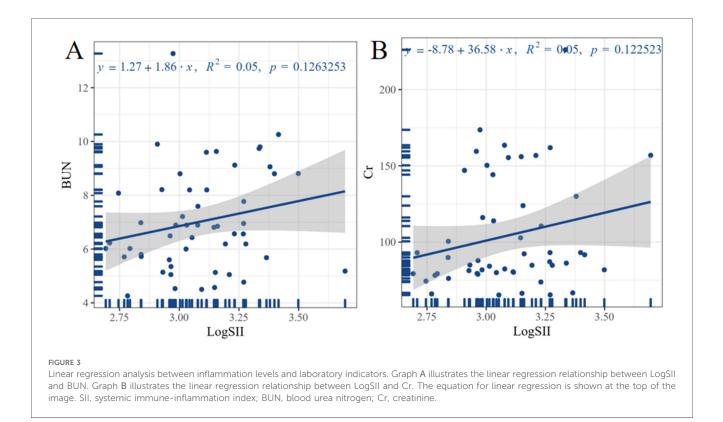
levels, with a correlation coefficient of 0.22, but the correlation remained weak. Linear regression can further quantify the trend between the two variables. Figure 3 shows the linear regression between LogSII and BUN and Cr; as LogSII increased, BUN and Cr both increased to varying degrees.



Discussion

Our study compared the laboratory parameters between the AAA and control groups. Inflammation, transaminase, bilirubin, and Cr levels were significantly higher in the case group than in the control group, and these indices were statistically significant between the two groups before and after PSM. There was a positive correlation between inflammation level and renal function evaluation parameters in the case group, and it is possible that there was an increased risk of kidney injury in the high inflammatory state. However, linear regression analysis of the level of inflammation and renal function evaluation parameters in the case group showed a non-significant relationship, which may be related to the state of AAA where the renal blood supply is affected, among other things.

To the best of our knowledge, this study is the first to investigate the association between preoperative inflammatory status and organ damage in AAA patients. PSM was also used to attenuate the interference of confounding factors between the case and control groups. Few clinical studies have been conducted on AAA. In a study of 79 patients with AAA who underwent endovascular aneurysm repair, Shalaby SY et al. found that patients with preoperative systemic inflammation experienced more postoperative complications (17). Windsor et al. selected 20 patients with AAA and 20 controls for their study and found that a higher intensity of exercise induced a decrease in inflammatory factors in patients with AAA, such as matrix metalloproteinase-9 (MMP-9) and tumor necrosis factor alpha (TNF- α) (24). Golledge J et al. collected data on 997 elderly male patients with AAA and found that the patients had higher levels of the inflammatory mediator angiopoietin-2, and that increased angiopoietin-2 was associated with increased risk of cardiovascular death (25). In a retrospective study of 373 patients undergoing elective endovascular aortic aneurysm repair, King AH et al. found that patients with an elevated preoperative neutrophil-lymphocyte ratio (NLR) had an



increased risk of postoperative mortality (26). A survey of 99 postoperative AAA patients by Muehling et al. found that perioperative advanced nursing care significantly reduced the rates of systemic inflammatory response syndrome (SIRS) and organ failure (27). Our study found that most patients with AAA had a history of hypertension and were on antihypertensive and lipid-lowering medications, and therefore rational control of blood pressure may be beneficial to the stabilization of the patient's condition.

Inflammation is a central factor in the development of AAA (28). Primary inflammation of the vessel wall leads to progressive dilatation of the abdominal aorta (29). After the formation of an AAA, the inflammatory response may be further exacerbated by a series of pathological changes that may occur in the surrounding tissues and vessel walls, such as thickening of the vessel wall, fibrosis, and infiltration of inflammatory cells (30). AAA is a local autoimmune response, and complement activation amplifies the inflammatory response (31). When lipids in the aneurysmal plaque leak into the surrounding tissues, they act as allergens that trigger an immune response, leading to inflammation (32). In addition, embolization of aortic trophoblastic vessels may trigger intima-media damage, accompanied by an inflammatory response (33). Lymphatic stasis and retroperitoneal edema due to lymphatic vessel compression also play roles in the inflammatory response (34). In patients with AAA, the inflammatory response is exacerbated by a history of smoking and hypertension (35).

Inflammation is the body's defense response to external stimuli, which is beneficial to the body; however, the function of all organs in the body is damaged in a state of high inflammation (36). In a state of high inflammation, a large number of inflammatory and anti-inflammatory factors are distributed to tissues and organs throughout the body via blood circulation (37). These factors act on the capillary wall, leading to damage to capillary endothelial cells and resulting in microcirculation disorders (38). Microstructural changes occur in organs, such as thickening of the vessel wall, narrowing of the lumen, and fibrosis. These changes, in turn, affect the normal function of the organs (37, 38). Kidney damage results in decreased urine output and increased blood Cr levels (39). Liver damage causes jaundice and elevated transaminase levels (40). Therefore, preoperative monitoring of the patient's inflammation level may help to protect the organs, and the development of a timely and dynamic monitoring device in the future will be more useful for the preoperative preparation of the patient.

This study has several limitations. First, as this was a crosssectional survey study, it was not possible to clarify the relationship between dynamic changes in inflammation levels and organ damage. Second, although the interference of relevant confounders, such as smoking history and history of previous diseases, was attenuated by PSM, other confounders, such as drug use, could not be avoided. Third, the values of transaminase, bilirubin, BUN, and Cr used in our assessment of hepatic and renal function were based on the results on the day of the study, which may be confounded by a variety of factors. Fourth, the population we studied was single-center and had a limited sample size, and we did not investigate multicenter data, and the results may have been limited in extensibility.

Conclusions

In conclusion, patients with AAA have a hyperinflammatory state preoperatively, and a hyperinflammatory response may be associated with organ damage. Therefore, preoperative monitoring and attention to inflammatory markers and changes in organ function are advocated.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

The studies involving humans were approved by Ethics Committee of the Peking University Shougang Hospital. The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study. Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

Author contributions

HW: Writing – original draft. BS: Writing – original draft. JL: Writing – original draft. HW: Writing – original draft, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This work was

References

1. Harrison SC, Holmes MV, Burgess S, Asselbergs FW, Jones GT, Baas AF, et al. Genetic association of lipids and lipid drug targets with abdominal aortic aneurysm: a meta-analysis. *JAMA Cardiol.* (2018) 3:26. doi: 10.1001/jamacardio.2017.4293

2. Sweeting MJ, Masconi KL, Jones E, Ulug P, Glover MJ, Michaels JA, et al. Analysis of clinical benefit, harms, and cost-effectiveness of screening women for abdominal aortic aneurysm. *Lancet*. (2018) 392:487–95. doi: 10.1016/S0140-6736(18)31222-4

3. Stepien KL, Bajdak-Rusinek K, Fus-Kujawa A, Kuczmik W, Gawron K. Role of extracellular matrix and inflammation in abdominal aortic aneurysm. *Int J Mol Sci.* (2022) 23(19):11078. doi: 10.3390/ijms231911078

4. Sakalihasan N, Michel JB, Katsargyris A, Kuivaniemi H, Defraigne JO, Nchimi A, et al. Abdominal aortic aneurysms. *Nat Rev Dis Primers*. (2018) 4(1):34. doi: 10.1038/s41572-018-0030-7

5. Sorvillo N, Cherpokova D, Martinod K, Wagner DD. Extracellular DNA NETworks with dire consequences for health. *Circ Res.* (2019) 125(4):470–88. doi: 10. 1161/CIRCRESAHA.119.314581

 Villahoz S, Yunes-Leites PS, Méndez-Barbero N, Urso K, Bonzon-Kulichenko E, Ortega S, et al. Conditional deletion of Rcan1 predisposes to hypertension-mediated intramural hematoma and subsequent aneurysm and aortic rupture. *Nat Commun.* (2018) 9:4795. doi: 10.1038/s41467-018-07071-7 supported by Capital's Funds for Health Improvement and Research (2020-2-6042), Key Medical disciplines/schools of Shijingshan District (Vascular Medicine), and Key Clinical Projects in Peking University Shougang Hospital (2019-Yuan-LC-01; SGYYZ202105).

Acknowledgments

We thank all staff and participants of the study for their invaluable contributions.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

7. Chen P-Y, Qin L, Li G, Malagon-Lopez J, Wang Z, Bergaya S, et al. Smooth muscle cell reprogramming in aortic aneurysms. *Cell Stem Cell.* (2020) 26:542–557.e11. doi: 10.1016/j.stem.2020.02.013

8. Lee JS, Park SC, Kim SD. The relation between metabolic syndrome and aspects of abdominal aortic aneurysm. *Asian J Surg*. (2022) 45(1):307–14. doi: 10.1016/j.asjsur. 2021.05.046

9. Menges AL, D Oria M, Zimmermann A, Dueppers P. Ruptured abdominal aortoiliac aneurysms: diagnosis, treatment, abdominal compartment syndrome, and role of simulation-based training. *Semin Vasc Surg.* (2023) 36(2):163–73. doi: 10.1053/j. semvascsurg.2023.03.002

10. Schanzer A, Messina LM, Ghosh K, Simons JP, Robinson WP, Aiello FA, et al. Follow-up compliance after endovascular abdominal aortic aneurysm repair in medicare beneficiaries. J Vasc Surg. (2015) 61:16–22.e1. doi: 10.1016/j.jvs.2014.06.006

11. Giese MA, Hind LE, Huttenlocher A. Neutrophil plasticity in the tumor microenvironment. Blood. (2019) 133:2159-67. doi: 10.1182/blood-2018-11-844548

12. Yang Y, Hozawa A, Kogure M, Narita A, Hirata T, Nakamura T, et al. Dietary inflammatory Index positively associated with high-sensitivity C-reactive protein level in Japanese from NIPPON DATA2010. *Journal of Epidemiology.* (2020) 30:98–107. doi: 10.2188/jea.JE20180156

13. Baumer Y, Dey AK, Gutierrez-Huerta CA, Khalil NO, Sekine Y, Sanda GE, et al. Hyperlipidaemia and IFNgamma/TNFalpha synergism are associated with cholesterol crystal formation in endothelial cells partly through modulation of lysosomal pH and cholesterol homeostasis. *eBioMedicine*. (2020) 59:102876. doi: 10.1016/j.ebiom.2020. 102876

14. Jin Y, Li J, Chen J, Shao M, Zhang R, Liang Y, et al. Tissue-specific autoantibodies improve diagnosis of primary Sjögren's syndrome in the early stage and indicate localized salivary injury. *J Immunol Res.* (2019) 2019:1–8. doi: 10.1155/2019/3642937

15. Wilson CH, Kumar S. Caspases in metabolic disease and their therapeutic potential. *Cell Death Differ*. (2018) 25:1010–24. doi: 10.1038/s41418-018-0111-x

16. Bradley NA, Walter A, Wilson A, Siddiqui T, Roxburgh CSD, McMillan DC, et al. The prognostic value of preoperative systemic inflammation-based scoring in patients undergoing endovascular repair of abdominal aortic aneurysm. *J Vasc Surg.* (2023) 78(2):362–369.e2. doi: 10.1016/j.jvs.2023.04.018

17. Shalaby SY, Foster TR, Hall MR, Brownson KE, Vasilas P, Federman DG, et al. Systemic inflammatory disease and its association with type II endoleak and late interventions after endovascular aneurysm repair. *JAMA Surg.* (2016) 151(2):147–53. doi: 10.1001/jamasurg.2015.3219

18. Chai X, Chen Y, Li Y, Chi J, Guo S. Lower geriatric nutritional risk index is associated with a higher risk of all-cause mortality in patients with chronic obstructive pulmonary disease: a cohort study from the national health and nutrition examination survey 2013–2018. *BMJ Open Resp Res.* (2023) 10:e001518. doi: 10.1136/bmjresp-2022-001518

19. Sun Y-H, Hu N-Q, Huang X-Y, Liu Z-X, Li Q-Y, Li Q-L, et al. Central and peripheral blood pressures in relation to the triglyceride-glucose index in a Chinese population. *Cardiovasc Diabetol.* (2024) 23:3. doi: 10.1186/s12933-023-02068-z

20. Roth GA, Mensah GA, Johnson CO, Addolorato G, Ammirati E, Baddour LM, et al. Global burden of cardiovascular diseases and risk factors, 1990–2019. J Am Coll Cardiol. (2020) 76:2982–3021. doi: 10.1016/j.jacc.2020.11.010

21. Mahemuti N, Jing X, Zhang N, Liu C, Li C, Cui Z, et al. Association between systemic immunity-inflammation Index and hyperlipidemia: a population-based study from the NHANES (2015–2020). *Nutrients*. (2023) 15:1177. doi: 10.3390/ nu15051177

22. Cheng W, Bu X, Xu C, Wen G, Kong F, Pan H, et al. Higher systemic immuneinflammation index and systemic inflammation response index levels are associated with stroke prevalence in the asthmatic population: a cross-sectional analysis of the NHANES 1999–2018. *Front Immunol.* (2023) 14:1191130. doi: 10.3389/fimmu.2023. 1191130

23. Esparham A, Shoar S, Mehri A, Modukuru VR. Bariatric surgery and cardiovascular disease risk in patients with pulmonary hypertension: a propensity score matched analysis of US national inpatient sample. *Obes Surg.* (2023) 33:3230–6. doi: 10.1007/s11695-023-06799-6

24. Windsor MT, Bailey TG, Perissiou M, Greaves K, Jha P, Leicht AS, et al. Acute inflammatory responses to exercise in patients with abdominal aortic aneurysm. *Med Sci Sports Exerc.* (2018) 50:649–58. doi: 10.1249/MSS.000000000001501

25. Golledge J, Clancy P, Yeap BB, Hankey GJ, Norman PE. Increased serum angiopoietin-2 is associated with abdominal aortic aneurysm prevalence and cardiovascular mortality in older men. *Int J Cardiol.* (2013) 167:1159–63. doi: 10.1016/j.ijcard.2012.03.120

26. King AH, Schmaier AH, Harth KC, Kumins NH, Wong VL, Zidar DA, et al. Elevated neutrophil-lymphocyte ratio predicts mortality following elective

endovascular aneurysm repair. J Vasc Surg. (2020) 72(1):129–37. doi: 10.1016/j.jvs. 2019.10.058

27. Muehling BM, Ortlieb L, Oberhuber A, Orend KH. Fast track management reduces the systemic inflammatory response and organ failure following elective infrarenal aortic aneurysm repair. *Interact Cardiovasc Thorac Surg.* (2011) 12:784–8. doi: 10.1510/icvts.2010.262337

28. Yuan Z, Lu Y, Wei J, Wu J, Yang J, Cai Z. Abdominal aortic aneurysm: roles of inflammatory cells. *Front Immunol.* (2021) 11:609161. doi: 10.3389/fimmu.2020. 609161

29. Wagenhäuser MU, Mulorz J, Krott KJ, Bosbach A, Feige T, Rhee YH, et al. Crosstalk of platelets with macrophages and fibroblasts aggravates inflammation, aortic wall stiffening, and osteopontin release in abdominal aortic aneurysm. *Cardiovasc Res.* (2024) 120(4):417–32. doi: 10.1093/cvr/cvad168

30. Filiberto AC, Spinosa MD, Elder CT, Su G, Leroy V, Ladd Z, et al. Endothelial pannexin-1 channels modulate macrophage and smooth muscle cell activation in abdominal aortic aneurysm formation. *Nat Commun.* (2022) 13(1):1521. doi: 10. 1038/s41467-022-29233-4

31. Lu S, White JV, Nwaneshiudu I, Nwaneshiudu A, Monos DS, Solomides CC, et al. Human abdominal aortic aneurysm (AAA): evidence for an autoimmune antigen-driven disease. *Autoimmun Rev.* (2022) 21(10):103164. doi: 10.1016/j.autrev. 2022.103164

32. Irimia A, Sarkar A, Stanfield RL, Wilson IA. Crystallographic identification of lipid as an integral component of the epitope of HIV broadly neutralizing antibody 4E10. *Immunity.* (2016) 44:21–31. doi: 10.1016/j.immuni.2015.12.001

33. Jolly SS, Cairns JA, Yusuf S, Rokoss MJ, Gao P, Meeks B, et al. Outcomes after thrombus aspiration for ST elevation myocardial infarction: 1-year follow-up of the prospective randomised TOTAL trial. *Lancet.* (2016) 387:127–35. doi: 10.1016/S0140-6736(15)00448-1

34. Ha CWY, Martin A, Sepich-Poore GD, Shi B, Wang Y, Gouin K, et al. Translocation of viable gut Microbiota to mesenteric adipose drives formation of creeping fat in humans. *Cell.* (2020) 183:666–683.e17. doi: 10.1016/j.cell.2020.09.009

35. Zhu F, Willette-Brown J, Song N-Y, Lomada D, Song Y, Xue L, et al. Autoreactive T cells and chronic fungal infection drive esophageal carcinogenesis. *Cell Host Microbe.* (2017) 21:478–493.e7. doi: 10.1016/j.chom.2017.03.006

36. Lobry T, Miller R, Nevo N, Rocca CJ, Zhang J, Catz SD, et al. Interaction between galectin-3 and cystinosin uncovers a pathogenic role of inflammation in kidney involvement of cystinosis. *Kidney Int.* (2019) 96(2):350–62. doi: 10.1016/j. kint.2019.01.029

37. Popescu NI, Silasi R, Keshari RS, Girton A, Burgett T, Zeerleder SS, et al. Peptidoglycan induces disseminated intravascular coagulation in baboons through activation of both coagulation pathways. *Blood.* (2018) 132(8):849–60. doi: 10.1182/blood-2017-10-813618

38. Park S-Y, Matte A, Jung Y, Ryu J, Anand WB, Han E-Y, et al. Pathologic angiogenesis in the bone marrow of humanized sickle cell mice is reversed by blood transfusion. *Blood.* (2020) 135(23):2071–84. doi: 10.1182/blood.2019002227

39. Oliveira BDD, Xu K, Shen TH, Callahan M, Kiryluk K, D'Agati VD, et al. Molecular nephrology: types of acute tubular injury. *Nat Rev Nephrol.* (2019) 15(10):599–612. doi: 10.1038/s41581-019-0184-x

40. Khoury HJ, Gambacorti-Passerini C. Practical management of toxicities associated with bosutinib in patients with Philadelphia chromosome-positive chronic myeloid leukemia. *Ann Oncol.* (2018) 29(3):578–87. doi: 10.1093/annonc/mdy019