



CD44: A Multifunctional Cell Surface Adhesion Receptor Is a Regulator of Progression and Metastasis of Cancer Cells

Linda T. Senbanjo and Meenakshi A. Chellaiah*

Department of Oncology and Diagnostic Sciences, Dental School, University of Maryland, Baltimore, MD, USA

CD44 is a cell surface adhesion receptor that is highly expressed in many cancers and regulates metastasis via recruitment of CD44 to the cell surface. Its interaction with appropriate extracellular matrix ligands promotes the migration and invasion processes involved in metastases. It was originally identified as a receptor for hyaluronan or hyaluronic acid and later to several other ligands including, osteopontin (OPN), collagens, and matrix metalloproteinases. CD44 has also been identified as a marker for stem cells of several types. Beside standard CD44 (sCD44), variant (vCD44) isoforms of CD44 have been shown to be created by alternate splicing of the mRNA in several cancer. Addition of new exons into the extracellular domain near the transmembrane of sCD44 increases the tendency for expressing larger size vCD44 isoforms. Expression of certain vCD44 isoforms was linked with progression and metastasis of cancer cells as well as patient prognosis. The expression of CD44 isoforms can be correlated with tumor subtypes and be a marker of cancer stem cells. CD44 cleavage, shedding, and elevated levels of soluble CD44 in the serum of patients is a marker of tumor burden and metastasis in several cancers including colon and gastric cancer. Recent observations have shown that CD44 intracellular domain (CD44-ICD) is related to the metastatic potential of breast cancer cells. However, the underlying mechanisms need further elucidation.

Keywords: CD44, cancer, metastasis, hyaluronic acid, migration, angiogenesis, invasion, CD44-ICD

CD44 INTRODUCTION

CD44 is a transmembrane glycoprotein also referred to as P-glycoprotein 1. It is encoded by a single gene on chromosome locus 11p13 (Underhill, 1992; Iczkowski, 2010). CD44 is ubiquitously expressed throughout the body and has a molecular weight of 85–200 kDa (Basakran, 2015). The standard CD44 (sCD44) is the conserved form with a molecular weight of about 85–90 kDa protein which is made of transcription of exons 1–5 and 16–20 that are spliced together (Rall and Rustgi, 1995; Rudzki and Jothy, 1997). The primary domains of CD44 are the extracellular domain (or ectodomain), the transmembrane domain, and the intracellular domain/cytoplasmic domain (Iczkowski, 2010). The extracellular domain interacts with the external microenvironment and senses stimuli in the external microenvironment (Underhill, 1992). The transmembrane domain provides an avenue for interacting with co-factors and adaptor proteins as well as directing lymphocyte homing (Underhill, 1992; Williams et al., 2013). CD44 intracellular domain (CD44-ICD) has a short-tail and long-tail configuration with functions in nuclear localization and

OPEN ACCESS

Edited by:

Mitsugu Fujita, Kindai University, Japan

Reviewed by:

Sumit Sahni, University of Illinois at Chicago, USA Sigrid A. Langhans, Alfred I. duPont Hospital for Children, USA

*Correspondence:

Meenakshi A. Chellaiah mchellaiah@umaryland.edu

Specialty section:

This article was submitted to Cell Adhesion and Migration, a section of the journal Frontiers in Cell and Developmental Biology

> Received: 03 January 2017 Accepted: 21 February 2017 Published: 07 March 2017

Citation:

Senbanjo LT and Chellaiah MA (2017) CD44: A Multifunctional Cell Surface Adhesion Receptor Is a Regulator of Progression and Metastasis of Cancer Cells. Front. Cell Dev. Biol. 5:18. doi: 10.3389/fcell.2017.00018 transcription mediation (Okamoto et al., 2001; Williams et al., 2013). Our current understanding of the dual role of CD44 in cancer progression is summarized in **Figure 1** below.

ISOFORMS OF CD44

Multiple isoforms of CD44 can be generated due to insertion of alternative exons at specific sites within the extracellular domain (Cichy and Puré, 2003). The variant isoforms of CD44 (CD44v) comprises of exon 6-15 spliced at various sites between exons 5 and 16 of the standard isoform (Goodison et al., 1999; Zeilstra et al., 2014). The expression of distinct CD44 isoforms appears to be necessary for the progression of human tumors (Günthert et al., 1995; Wang et al., 2009). One or multiple splice variants and standard CD44 may be expressed in cancer cells. There is an increased chance of expressing larger isoforms like that of CD44v8-10 in pancreatic cancers (Rall and Rustgi, 1995) and CD44v6 in colorectal cancer (Yamane et al., 1999). The expression of CD44v6 is well-known as a useful marker of tumor progression and prognosis in colorectal cancer (Yamane et al., 1999). In cultured supernatants from prostate cancer cell lines derived from bone metastasis (PC3), soluble CD44, and variant 6 isoform (v6) was identified, however, it was not identified in lymph node metastatic prostate cancer cell line (LNCaP, Stevens et al., 1996; Desai et al., 2009; Gupta et al., 2012). The switch from standard CD44 to CD44v6 improved survival and adhesion in prostate cancer (PC3) cells (Gupta et al., 2013b). The expression of CD44v6 in colorectal cancer is enhanced by cancer stem cells expression (Todaro et al., 2014).

CD44 EXPRESSION IN NORMAL AND TUMOR CELLS

The ubiquitous transmembrane cell surface molecule CD44 is widely distributed in normal adult and fetal tissues. CD44 standard isoform was originally isolated from hematopoietic cells but now it is found in a variety of tissues e.g., central nervous system, lung, and epidermis. In comparison, the distribution of CD44 variant isoforms is restricted and expressed on a selection of epithelial cells (Sneath and Mangham, 1998). The isoforms with restricted distribution and exon sequence may have different functions as compared to the standard isoform of CD44. Keratinocytes, macrophages, and select epithelial cells express the variant CD44 (CD44v) isoforms and are present on tissues at various stages of development (Sneath and Mangham, 1998). In normal tissues, the importance of CD44 is vital in the regulation of hyaluronic metabolism, activation of lymphocytes, and release of cytokines. However, targeting of CD44 resulting in its loss leads to the disruption of hyaluronic metabolism, wound healing, and keratinocyte proliferation (Yu and Stamenkovic, 1999). Among many of CD44 functions, one is to make cell lines that are non-metastatic become more metastatic (Heider et al., 1993). The details that confer CD44 metastatic potential in human malignancies is the subject of further elucidation. Prostate cells that are benign express higher CD44 variant 5 isoforms (CD44v5), whereas neoplastic prostate cells express higher levels of CD44s (Dhir et al., 1997; Desai et al., 2009; Gupta et al., 2013a).



Various breast cancer cells show abnormal expression of CD44 including heterogeneously expressing CD44 isoforms (Basakran, 2015).

CD44 RECEPTOR-LIGAND INTERACTION

CD44 is known to interact with various ligands and this interaction is crucial for its many cellular functions (Goodison et al., 1999). There are several well-known ligands of CD44 including hyaluronic acid (HA), osteopontin (OPN), collagens, and matrix metalloproteinases (MMPs) (Goodison et al., 1999). CD44 effects on cell migration and growth are dependent on its specificity to ligands (Weber et al., 1996).

Hyaluronan (HA)

HA is a glycosaminoglycan that is a ubiquitous component of the extracellular membrane. It is considered the major ligand for CD44 and can bind CD44v isoforms that are ubiquitously expressed. Through binding of CD44, HA can activate cytoskeleton and matrix metalloproteinases (MMPs) signaling involved in tumor progression (Bourguignon et al., 2014). Multiple regions of the cytoplasmic domain of CD44 can promote enhancement of HA binding, however, the role of the cytoplasmic domain in mediating the binding does not require a specific amino acid sequence in T-lymphoma cells (Perschl et al., 1995). HA can exist in high molecular weight or low molecular weight form due to cleavage into varying sizes. In breast cancer cell lines, high molecular weight HA is involved in tumorigenesis, antiangiogenic and anti-inflammatory responses. However, low molecular weight HA has been shown to promote cell motility, CD44 cleavage and angiogenesis. Therefore, the size of HA ligand is important for the biological function (Louderbough and Schroeder, 2011).

Osteopontin (OPN)

Several studies have demonstrated an elevated expression of OPN in highly invasive metastatic human cancers (Tuck et al., 2007). Integrin $\alpha v\beta 3$ and CD44 are receptors for OPN and can interact with $\alpha_{\nu}\beta_3$ through its functional arginine-glycineaspartic acid (RGD) cell binding sequence (Thalmann et al., 1999; Desai et al., 2007). Prostate cancer growth and progression is shown to be mediated by paracrine and autocrine signaling of OPN (Thalmann et al., 1999). CD44-OPN interaction induces cell migration out of the bloodstream to sites of inflammation. The migration of cells and subsequent invasion at distant sites involves a complex sequence of events (Weber et al., 1996). Variant CD44 isoforms bind to OPN independent of RGD sequences present at the N-terminal domain of CD44 (Katagiri et al., 1999). OPN binding to CD44 variants/beta1-containing integrin promotes cell spreading, motility, and chemotactic behavior in rat pancreatic carcinoma (Katagiri et al., 1999). OPN increases surface expression of standard CD44 (sCD44) in osteoclasts and both sCD44 and variant isoforms in human melanoma and PC3 cells (Chellaiah et al., 2003; Samanna et al., 2006; Desai et al., 2007). Osteopontin regulation of surface expression of CD44v6 and sCD44 was observed in breast and hepatocellular cancer cells (Gao et al., 2003; Khan et al., 2005).

Matrix Metalloproteases (MMPs)

Matrix metalloproteinases (MMPs) are important extracellular matrix proteins that are involved in degradation of the extracellular matrix. They are also important during development, wound healing, bone resorption, and angiogenesis (Paiva and Granjeiro, 2014). There is evidence suggesting that MMP-9 and CD44 associate in mouse and human tumor cells resulting in MMP9 activity localization on the cell surface (Yu and Stamenkovic, 1999; Gupta et al., 2013a). The interaction of CD44 and proteolytic form of MMP-9 is particularly involved in the invasion of prostate cancer cells (PC3) derived from bone metastases (Desai et al., 2007). Therefore, the ability of CD44 to localize proteolytically active MMP-9 to the tumor cell surface is important for tumor invasion (Yu and Stamenkovic, 1999).

CD44 ROLE IN MIGRATION/INVASION, ANGIOGENESIS, AND BONE METASTASIS

Migration and Invasion

CD44 receptor has the potential to integrate adhesive and signaling activities to modulate migration/invasion processes during cancer progression (Lokeshwar et al., 1995). The mechanisms by which CD44 receptors mediate migration, proliferation, survival of tumor cells through HA-mediated signaling have been widely studied (Bourguignon et al., 1998, 2001, 2004; Kuniyasu et al., 2001; Wang and Bourguignon, 2006a,b; Wang et al., 2009). Changes in cell shape and formation

of adhesive structures are regulated by the dynamic regulation of the actin cytoskeleton. The dynamic regulation of the actin cytoskeleton and the specialized structures involved in migration are regulated by the temporal and spatial localization of actinbinding proteins (Chellaiah et al., 2000; Linder and Aepfelbacher, 2003; Desai et al., 2008). The surface expression of CD44 along with its interaction with matrix metalloproteinase 9 (MMP9) on the surface of the cell results in secretion of active MMP9, migration, and invasion of PC3 cells (Desai et al., 2007, 2008; Gupta et al., 2013a). Disruption of CD44/MMP9 interaction on the cell surface reduces migration and invasion of PC3 cells. When MMP9 is knockdown, CD44 expression switches to variant 6 (v6) isoform. This results in a less invasive phenotype due to lack of expression of sCD44 and inability to form invadopodia (Gupta et al., 2013a). CD44v6 expression inversely correlates with pathologic stage and disease progression and positively correlates with PSA-free survival in prostate cancer (Ekici et al., 2002). However, expression of CD44v6 in nonmetastatic rat carcinoma cells has been shown to convert them into metastatic cells and promote tumor progression (Günthert et al., 1991; Seiter et al., 1993). Furthermore, CD44v3 has been shown to upregulate the function of cytoskeleton through ankyrin to activate the actomyosin contractile complex in order to mediate cell migration in head and neck squamous carcinoma cell line. Transfection of v3 cDNA into non-expressing cell lines also resulted in a significant increase in cell migration but not proliferation (Franzmann et al., 2001; Wang et al., 2007). CD44 variants have also been shown to function as a co-receptor for the activation of growth-promoting tumor receptor tyrosine kinases (Orian-Rousseau et al., 2002, 2007).

Angiogenesis

The formation of new blood vessels (angiogenesis) is required for tumor cell to disseminate and migrate to distant organs. Past studies have identified CD44 expression on endothelial cells (Liesveld et al., 1994; Xu et al., 1994) and this controls the formation of blood vessels (Trochon et al., 1996; Savani et al., 2001). Inhibition of CD44 therefore results in impaired formation of vessel-like networks (Savani et al., 2001; Cao et al., 2006). Endothelial cells were found in increased numbers in prostate cancer tissues in relation to normal tissues (Wang et al., 2013). When CD44-null mice was used to study in vivo angiogenic responses, wound healing and vascularization were both impaired in matrigel implants. Therefore, metastasis formation is also linked to vascular CD44 expression (Cao et al., 2006). Adhesion of cancer cells to vasculature and enhanced expression of CD44 (CD44s and/or CD44v) by angiogenic factors (e.g., VEGF) produced by tumor cells might lead to facilitated extravasation via angiogenesis. Furthermore, the role of CD44 in tumor angiogenesis is enhanced by its binding to immobilized HA (Griffioen et al., 1997). CD44 variants are shown to have binding domains for various growth factors including vascular endothelial growth factor (VEGF), heparin-binding basic fibroblast growth factor and heparin binding epidermal growth factor (Bourguignon et al., 1998, 1999; Kalish et al., 1999). Analysis of tissue microarray and lysates of prostatic tumor cells showed that OPN and VEGF expression was more pronounced in prostate cancer as compared to benign or normal prostate tissues. It was suggested that an increase in micro vessel number and expression of CD44 might be useful diagnostic markers of metastasis of breast cancer (Ozer et al., 1997).

Bone Metastasis

Breast and prostate cancer cells ability to metastasize to bone is based on their ability to arrest on, adhere to, and extravasate across the bone marrow endothelium into the underlying bone matrix (Draffin et al., 2003). In prostate cancer cells, the selective adhesion of these cells to bone marrow epithelium is based on the role of adhesive properties of integrin receptors. Prostate cancer cells have been involved in strong interaction with the bone marrow endothelial cells (Draffin et al., 2004). There currently exists a dissension between clinical and experimental data in literature regarding the importance of sCD44 in breast cancer disease progression. A recent study suggests that breast cancer models show the expression of CD44 standard and variant isoforms which increase disease-progressing and metastatic behavior (McFarlane et al., 2015). HA and CD44 co-localize in the bone marrow sinusoidal epithelium, which is a site of metastasis of breast cancer. This suggests the contribution of HA-CD44 to the efficiency of distant metastasis to bone in breast cancer cells (McFarlane et al., 2015). Cells producing low levels of CD44 have lower ability to form tumor sphere in vitro. Furthermore, CD44 is a marker for cancer stem cells (Jaggupilli and Elkord, 2012; Cho et al., 2015; Stivarou and Patsavoudi, 2015) and CD44 expressing cancer stem cells increases the likelihood of bone metastases through its interaction with HA. Therefore, CD44-HA interaction could be a potential target for reducing bone metastases. CD44 signaling in prostate cancer cells has also been shown to regulate key proteins (i.e., RANKL and MMP9) involved in osteoclast differentiation and tumor metastasis (Gupta et al., 2012). Runx2 is a master transcription factor with important roles in osteoblast differentiation. Transcription of many osteoblast and bone formation related factors such as OPN, osteocalcin, and collagen type I are regulated by Runx2 (Akech et al., 2010).

ROLE OF CD44 AS A TRANSCRIPTIONAL FACTOR

Proteolytic cleavage that occurs at the extracellular domain releasing soluble CD44 has long been recognized. However, recent studies have shown that CD44 can undergo further sequential proteolytic processing by membrane type 1 matrix metalloproteases (MT1-MMP) and presenilin-1/y-secretase to produce the extracellular domain and intracellular domain (ICD) fragments. Presenilin-1/y-secretase cleavage occurs at the intramembrane site releasing two cleavage products of ~25 and ~16 kDa size. The 12 kDa ICD translocates to the nucleus to

REFERENCES

Akech, J., Wixted, J. J., Bedard, K., van der Deen, M., Hussain, S., Guise, T. A., et al. (2010). Runx2 association with progression of prostate cancer in patients: mechanisms mediating bone osteolysis and osteoblastic metastatic lesions. Oncogene 29, 811–821. doi: 10.1038/onc.2009.389

activate transcription of several proteins including CD44 itself (Okamoto et al., 2001; Nagano and Sava, 2004; Thorne et al., 2004). Consequently, if this cleavage can be inhibited through metalloprotease inhibitors, it can serve as a therapeutic way of preventing tumor progression and metastasis (Nagano and Saya, 2004). The translocation of CD44-ICD to the nucleus initiates the process of transcriptional regulation via it binding to novel promoter response element thereby regulating transcription of several genes that are involved in cell survival during stress, inflammation, oxidative glycolysis, tumor invasion (Okamoto et al., 2001; Miletti-González et al., 2012). This suggests a mechanism for the multifunctional role of CD44 in cancer cell metastasis and metabolism (Miletti-González et al., 2012). Nuclear translocation of the intracellular domain also shown to interact with stemness factors (Cho et al., 2015). CD44-ICD is linked with the regulation of MMP-9 gene in prostate and breast cancer cells through its interaction with the transcriptional factor RUNX2 (Miletti-González et al., 2012).

CONCLUSIONS

The multifunctional glycoprotein CD44 can undergo alternative splicing events to produce CD44 variant isoforms that are more restricted in their distribution as compared to the standard CD44 isoforms (Rall and Rustgi, 1995). The ubiquitously expressed cell surface protein is primarily involved in aggregation, migration, and activation of cells, these functions are mediated through the adhesive properties of CD44 (Heider et al., 1993). Initially described for hematopoietic stem cells, it has since been confirmed as a marker of cancer stem cells (Bourguignon et al., 1998). CD44 interacts with a variety of ligands and can undergo sequential proteolytic processing resulting in the generation of CD44-ICD. CD44-ICD is known to translocate into the nucleus to activate gene transcription (Okamoto et al., 1999). Though the information provided here provides a comprehensive review of the literature thus far, there exists some discourse in the effect of ICD as the main modulator of metastatic events in cancers. To further substantiate CD44's effect in metastasis, research into the specifics will need to be completed to address this.

AUTHOR CONTRIBUTIONS

LS and MC drafted the manuscript and equally contributed in editing and rewriting final contents.

FUNDING

This work was supported by a research grant to MC from the National Institute of Health - National Institute of Arthritis and Musculoskeletal and Skin Diseases (5R01AR066044).

Basakran, N. S. (2015). CD44 as a potential diagnostic tumor marker. *Saudi Med. J.* 36, 273–279. doi: 10.15537/smj.2015.3.9622

Bourguignon, L. Y., Gunja-Smith, Z., Iida, N., Zhu, H. B., Young, L. J., Muller, W. J., et al. (1998). CD44v(3,8-10) is involved in cytoskeletonmediated tumor cell migration and matrix metalloproteinase (MMP-9) association in metastatic breast cancer cells. J. Cell. Physiol. 176, 206–215. doi: 10.1002/(SICI)1097-4652(199807)176:1<206::AID-JCP22>3.0. CO;2-3

- Bourguignon, L. Y., Shiina, M., and Li, J. J. (2014). Hyaluronan-CD44 interaction promotes oncogenic signaling, microRNA functions, chemoresistance, and radiation resistance in cancer stem cells leading to tumor progression. Adv. Cancer Res. 123, 255–275. doi: 10.1016/B978-0-12-800092-2.00010-1
- Bourguignon, L. Y., Singleton, P. A., Diedrich, F., Stern, R., and Gilad, E. (2004). CD44 interaction with Na+-H+ exchanger (NHE1) creates acidic microenvironments leading to hyaluronidase-2 and cathepsin B activation and breast tumor cell invasion. J. Biol. Chem. 279, 26991–27007. doi: 10.1074/jbc.M311838200
- Bourguignon, L. Y., Zhu, H., Shao, L., and Chen, Y. W. (2001). CD44 interaction with c-Src kinase promotes cortactin-mediated cytoskeleton function and hyaluronic acid-dependent ovarian tumor cell migration. J. Biol. Chem. 276, 7327–7336. doi: 10.1074/jbc.M006498200
- Bourguignon, L. Y., Zhu, H., Shao, L., Zhu, D., and Chen, Y. W. (1999). Rhokinase (ROK) promotes CD44v(3,8-10)-ankyrin interaction and tumor cell migration in metastatic breast cancer cells. *Cell Motil. Cytoskeleton* 43, 269–287. doi: 10.1002/(SICI)1097-0169(1999)43:4<269::AID-CM1>3.0.CO;2-5
- Cao, G., Savani, R. C., Fehrenbach, M., Lyons, C., Zhang, L., Coukos, G., et al. (2006). Involvement of endothelial CD44 during *in vivo* angiogenesis. *Am. J. Pathol.* 169, 325–336. doi: 10.2353/ajpath.2006.060206
- Chellaiah, M. A., Kizer, N., Biswas, R., Alvarez, U., Strauss-Schoenberger, J., Rifas, L., et al. (2003). Osteopontin deficiency produces osteoclast dysfunction due to reduced CD44 surface expression. *Mol. Biol. Cell* 14, 173–189. doi: 10.1091/mbc.E02-06-0354
- Chellaiah, M., Kizer, N., Silva, M., Alvarez, U., Kwiatkowski, D., and Hruska, K. A. (2000). Gelsolin deficiency blocks podosome assembly and produces increased bone mass and strength. J. Cell Biol. 148, 665–678. doi: 10.1083/jcb.148.4.665
- Cho, Y., Lee, H. W., Kang, H. G., Kim, H. Y., Kim, S. J., and Chun, K. H. (2015). Cleaved CD44 intracellular domain supports activation of stemness factors and promotes tumorigenesis of breast cancer. *Oncotarget* 6, 8709–8721. doi: 10.18632/oncotarget.3325
- Cichy, J., and Puré, E. (2003). The liberation of CD44. J. Cell Biol. 161, 839–843. doi: 10.1083/jcb.200302098
- Desai, B., Ma, T., and Chellaiah, M. A. (2008). Invadopodia and matrix degradation, a new property of prostate cancer cells during migration and invasion. J. Biol. Chem. 283, 13856–13866. doi: 10.1074/jbc.M709401200
- Desai, B., Ma, T., Zhu, J., and Chellaiah, M. A. (2009). Characterization of the expression of variant and standard CD44 in prostate cancer cells: identification of the possible molecular mechanism of CD44/MMP9 complex formation on the cell surface. *J. Cell. Biochem.* 108, 272–284. doi: 10.1002/jcb.22248
- Desai, B., Rogers, M. J., and Chellaiah, M. A. (2007). Mechanisms of osteopontin and CD44 as metastatic principles in prostate cancer cells. *Mol. Cancer* 6:18. doi: 10.1186/1476-4598-6-18
- Dhir, R., Gau, J. T., Krill, D., Bastacky, S., Bahnson, R. R., Cooper, D. L., et al. (1997). CD44 expression in benign and neoplastic human prostates. *Mol. Diagn.* 2, 197–204. doi: 10.1016/S1084-8592(97)80029-X
- Draffin, J. E., Hill, A., Johnston, P. G., and Waugh, D. J. (2003). CD44 Expression on prostate cancer cells correlates with adhesion to bone marrow endothelial cells. *Clin. Cancer Res.* 9, 6181S–6181S.
- Draffin, J. E., McFarlane, S., Hill, A., Johnston, P. G., and Waugh, D. J. (2004). CD44 potentiates the adherence of metastatic prostate and breast cancer cells to bone marrow endothelial cells. *Cancer Res.* 64, 5702–5711. doi: 10.1158/0008-5472.CAN-04-0389
- Ekici, S., Ayhan, A., Kendi, S., and Ozen, H. (2002). Determination of prognosis in patients with prostate cancer treated with radical prostatectomy: prognostic value of CD44v6 score. J. Urol. 167, 2037–2041. doi: 10.1016/S0022-5347(05)65078-1
- Franzmann, E. J., Weed, D. T., Civantos, F. J., Goodwin, W. J., and Bourguignon, L. Y. (2001). A novel CD44 v3 isoform is involved in head and neck squamous cell carcinoma progression. *Otolaryngol. Head Neck Surg.* 124, 426–432. doi: 10.1067/mhn.2001.114674
- Gao, C., Guo, H., Downey, L., Marroquin, C., Wei, J., and Kuo, P. C. (2003). Osteopontin-dependent CD44v6 expression and cell adhesion in HepG2 cells. *Carcinogenesis* 24, 1871–1878. doi: 10.1093/carcin/bgg139
- Goodison, S., Urquidi, V., and Tarin, D. (1999). CD44 cell adhesion molecules. *Mol. Pathol.* 52, 189–196. doi: 10.1136/mp.52.4.189

- Griffioen, A. W., Coenen, M. J., Damen, C. A., Hellwig, S. M., van Weering, D. H., Vooys, W., et al. (1997). CD44 is involved in tumor angiogenesis; an activation antigen on human endothelial cells. *Blood* 90, 1150–1159.
- Günthert, U., Hofmann, M., Rudy, W., Reber, S., Zöller, M., Haussmann, I., et al. (1991). A new variant of glycoprotein CD44 confers metastatic potential to rat carcinoma cells. *Cell* 65, 13–24. doi: 10.1016/0092-8674(91)90403-L
- Günthert, U., Stauder, R., Mayer, B., Terpe, H. J., Finke, L., and Friedrichs, K. (1995). Are CD44 variant isoforms involved in human tumour progression? *Cancer Surv.* 24, 19–42.
- Gupta, A., Cao, W., and Chellaiah, M. A. (2012). Integrin $\alpha\nu\beta3$ and CD44 pathways in metastatic prostate cancer cells support osteoclastogenesis via a Runx2/Smad 5/receptor activator of NF- κ B ligand signaling axis. *Mol. Cancer* 11:66. doi: 10.1186/1476-4598-11-66
- Gupta, A., Cao, W., Sadashivaiah, K., Chen, W., Schneider, A., and Chellaiah, M. A. (2013a). Promising noninvasive cellular phenotype in prostate cancer cells knockdown of matrix metalloproteinase 9. *ScientificWorldJournal* 2013:493689. doi: 10.1155/2013/493689
- Gupta, A., Zhou, C. Q., and Chellaiah, M. A. (2013b). Osteopontin and MMP9: associations with VEGF expression/Secretion and angiogenesis in PC3 prostate cancer Cells. *Cancers* 5, 617–638. doi: 10.3390/cancers5020617
- Heider, K. H., Hofmann, M., Hors, E., van den Berg, F., Ponta, H., Herrlich, P., et al. (1993). A human homologue of the rat metastasis-associated variant of CD44 is expressed in colorectal carcinomas and adenomatous polyps. *J. Cell Biol.* 120, 227–233. doi: 10.1083/jcb.120.1.227
- Iczkowski, K. A. (2010). Cell adhesion molecule CD44: its functional roles in prostate cancer. Am. J. Transl. Res. 3, 1–7.
- Jaggupilli, A., and Elkord, E. (2012). Significance of CD44 and CD24 as cancer stem cell markers: an enduring ambiguity. *Clin. Dev. Immunol.* 2012:708036. doi: 10.1155/2012/708036
- Kalish, E. D., Iida, N., Moffat, F. L., and Bourguignon, L. Y. (1999). A new CD44V3-containing isoform is involved in tumor cell growth and migration during human breast carcinoma progression. *Front. Biosci.* 4, A1–A8. doi: 10.2741/Kalish
- Katagiri, Y. U., Sleeman, J., Fujii, H., Herrlich, P., Hotta, H., Tanaka, K., et al. (1999). CD44 variants but not CD44s cooperate with beta1-containing integrins to permit cells to bind to osteopontin independently of arginineglycine-aspartic acid, thereby stimulating cell motility and chemotaxis. *Cancer Res.* 59, 219–226.
- Khan, S. A., Cook, A. C., Kappil, M., Günthert, U., Chambers, A. F., Tuck, A. B., et al. (2005). Enhanced cell surface CD44 variant (v6, v9) expression by osteopontin in breast cancer epithelial cells facilitates tumor cell migration: novel post-transcriptional, post-translational regulation. *Clin. Exp. Metastasis* 22, 663–673. doi: 10.1007/s10585-006-9007-0
- Kuniyasu, H., Oue, N., Tsutsumi, M., Tahara, E., and Yasui, W. (2001). Heparan sulfate enhances invasion by human colon carcinoma cell lines through expression of CD44 variant exon 3. *Clin. Cancer Res.* 7, 4067–4072.
- Liesveld, J. L., Frediani, K. E., Harbol, A. W., DiPersio, J. F., and Abboud, C. N. (1994). Characterization of the adherence of normal and leukemic CD34+ cells to endothelial monolayers. *Leukemia* 8, 2111–2117.
- Linder, S., and Aepfelbacher, M. (2003). Podosomes: adhesion hot-spots of invasive cells. *Trends Cell Biol.* 13, 376–385. doi: 10.1016/S0962-8924(03)00128-4
- Lokeshwar, B. L., Lokeshwar, V. B., and Block, N. L. (1995). Expression of CD44 in prostate cancer cells: association with cell proliferation and invasive potential. *Anticancer Res.* 15, 1191–1198.
- Louderbough, J. M., and Schroeder, J. A. (2011). Understanding the dual nature of CD44 in breast cancer progression. *Mol. Cancer Res.* 9, 1573–1586. doi: 10.1158/1541-7786.MCR-11-0156
- McFarlane, S., Coulter, J. A., Tibbits, P., O'Grady, A., McFarlane, C., Montgomery, N., et al. (2015). CD44 increases the efficiency of distant metastasis of breast cancer. *Oncotarget* 6, 11465–11476. doi: 10.18632/oncotarget.3410
- Miletti-González, K. E., Murphy, K., Kumaran, M. N., Ravindranath, A. K., Wernyj, R. P., Kaur, S., et al. (2012). Identification of function for CD44 intracytoplasmic domain (CD44-ICD): modulation of matrix metalloproteinase 9 (MMP-9) transcription via novel promoter response element. J. Biol. Chem. 287, 18995–19007. doi: 10.1074/jbc.M111. 318774
- Nagano, O., and Saya, H. (2004). Mechanism and biological significance of CD44 cleavage. *Cancer Sci.* 95, 930–935. doi: 10.1111/j.1349-7006.2004.tb03179.x

- Okamoto, I., Kawano, Y., Murakami, D., Sasayama, T., Araki, N., Miki, T., et al. (2001). Proteolytic release of CD44 intracellular domain and its role in the CD44 signaling pathway. *J. Cell Biol.* 155, 755–762. doi: 10.1083/jcb.2001 08159
- Okamoto, I., Kawano, Y., Tsuiki, H., Sasaki, J., Nakao, M., Matsumoto, M., et al. (1999). CD44 cleavage induced by a membrane-associated metalloprotease plays a critical role in tumor cell migration. *Oncogene* 18, 1435–1446. doi: 10.1038/sj.onc.1202447
- Orian-Rousseau, V., Chen, L., Sleeman, J. P., Herrlich, P., and Ponta, H. (2002). CD44 is required for two consecutive steps in HGF/c-Met signaling. *Genes Dev.* 16, 3074–3086. doi: 10.1101/gad.242602
- Orian-Rousseau, V., Morrison, H., Matzke, A., Kastilan, T., Pace, G., Herrlich, P., et al. (2007). Hepatocyte growth factor-induced Ras activation requires ERM proteins linked to both CD44v6 and F-actin. *Mol. Biol. Cell* 18, 76–83. doi: 10.1091/mbc.E06-08-0674
- Ozer, E., Canda, T., and Kurtodlu, B. (1997). The role of angiogenesis, laminin and CD44 expression in metastatic behavior of early-stage low-grade invasive breast carcinomas. *Cancer Lett.* 121, 119–123. doi: 10.1016/S0304-3835(97)00346-7
- Paiva, K. B., and Granjeiro, J. M. (2014). Bone tissue remodeling and development: focus on matrix metalloproteinase functions. *Arch. Biochem. Biophys.* 561, 74–87. doi: 10.1016/j.abb.2014.07.034
- Perschl, A., Lesley, J., English, N., Trowbridge, I., and Hyman, R. (1995). Role of CD44 cytoplasmic domain in hyaluronan binding. *Eur. J. Immunol.* 25, 495–501. doi: 10.1002/eji.1830250228
- Rall, C. J., and Rustgi, A. K. (1995). CD44 isoform expression in primary and metastatic pancreatic adenocarcinoma. *Cancer Res.* 55, 1831–1835.
- Rudzki, Z., and Jothy, S. (1997). CD44 and the adhesion of neoplastic cells. *Mol. Pathol.* 50, 57–71. doi: 10.1136/mp.50.2.57
- Samanna, V., Wei, H., Ego-Osuala, D., and Chellaiah, M. A. (2006). Alpha-Vdependent outside-in signaling is required for the regulation of CD44 surface expression, MMP-2 secretion, and cell migration by osteopontin in human melanoma cells. *Exp. Cell Res.* 312, 2214–2230. doi: 10.1016/j.yexcr.2006. 03.022
- Savani, R. C., Cao, G., Pooler, P. M., Zaman, A., Zhou, Z., and DeLisser, H. M. (2001). Differential involvement of the hyaluronan (HA) receptors CD44 and receptor for HA-mediated motility in endothelial cell function and angiogenesis. J. Biol. Chem. 276, 36770–36778. doi: 10.1074/jbc.M102273200
- Seiter, S., Arch, R., Reber, S., Komitowski, D., Hofmann, M., Ponta, H., et al. (1993). Prevention of tumor metastasis formation by anti-variant CD44. J. Exp. Med. 177, 443–455. doi: 10.1084/jem.177.2.443
- Sneath, R. J., and Mangham, D. C. (1998). The normal structure and function of CD44 and its role in neoplasia. *Mol. Pathol.* 51, 191–200. doi: 10.1136/mp.51.4.191
- Stevens, J. W., Palechek, P. L., Griebling, T. L., Midura, R. J., Rokhlin, O. W., and Cohen, M. B. (1996). Expression of CD44 isoforms in human prostate tumor cell lines. *Prostate* 28, 153–161. doi: 10.1002/(SICI)1097-0045(199603)28:3<153::AID-PROS2>3.0.CO;2-G
- Stivarou, T., and Patsavoudi, E. (2015). Extracellular molecules involved in cancer cell invasion. *Cancers* 7, 238–265. doi: 10.3390/cancers7010238
- Thalmann, G. N., Sikes, R. A., Devoll, R. E., Kiefer, J. A., Markwalder, R., Klima, I., et al. (1999). Osteopontin: possible role in prostate cancer progression. *Clin. Cancer Res.* 5, 2271–2277.
- Thorne, R. F., Legg, J. W., and Isacke, C. M. (2004). The role of the CD44 transmembrane and cytoplasmic domains in co-ordinating adhesive and signalling events. *J. Cell Sci.* 117(Pt 3), 373–380. doi: 10.1242/jcs.00954
- Todaro, M., Gaggianesi, M., Catalano, V., Benfante, A., Iovino, F., Biffoni, M., et al. (2014). CD44v6 is a marker of constitutive and reprogrammed cancer stem cells driving colon cancer metastasis. *Cell Stem Cell* 14, 342–356. doi: 10.1016/j.stem.2014.01.009

- Trochon, V., Mabilat, C., Bertrand, P., Legrand, Y., Smadja-Joffe, F., Soria, C., et al. (1996). Evidence of involvement of CD44 in endothelial cell proliferation, migration and angiogenesis *in vitro*. *Int. J. Cancer* 66, 664–668. doi: 10.1002/(SICI)1097-0215(19960529)66:5<664::AID-IJC14>3.0.CO;2-4
- Tuck, A. B., Chambers, A. F., and Allan, A. L. (2007). Osteopontin overexpression in breast cancer: knowledge gained and possible implications for clinical management. J. Cell. Biochem. 102, 859–868. doi: 10.1002/jcb.21520
- Underhill, C. (1992). CD44: the hyaluronan receptor. J. Cell. Sci. 103(Pt 2), 293–298.
- Wang, S. J., and Bourguignon, L. Y. (2006a). Hyaluronan and the interaction between CD44 and epidermal growth factor receptor in oncogenic signaling and chemotherapy resistance in head and neck cancer. *Arch. Otolaryngol. Head Neck Surg.* 132, 771–778. doi: 10.1001/archotol.132.7.771
- Wang, S. J., and Bourguignon, L. Y. (2006b). Hyaluronan-CD44 promotes phospholipase C-mediated Ca2+ signaling and cisplatin resistance in head and neck cancer. Arch. Otolaryngol. Head Neck Surg. 132, 19–24. doi: 10.1001/archotol.132.1.19
- Wang, S. J., Wong, G., de Heer, A. M., Xia, W., and Bourguignon, L. Y. (2009). CD44 variant isoforms in head and neck squamous cell carcinoma progression. *Laryngoscope* 119, 1518–1530. doi: 10.1002/lary.20506
- Wang, S. J., Wreesmann, V. B., and Bourguignon, L. Y. (2007). Association of CD44 V3-containing isoforms with tumor cell growth, migration, matrix metalloproteinase expression, and lymph node metastasis in head and neck cancer. *Head Neck* 29, 550–558. doi: 10.1002/hed.20544
- Wang, X., Lee, S. O., Xia, S., Jiang, Q., Luo, J., Li, L., et al. (2013). Endothelial cells enhance prostate cancer metastasis via IL-6→androgen receptor→TGF-β→MMP-9 signals. *Mol. Cancer Ther.* 12, 1026–1037. doi: 10.1158/1535-7163.MCT-12-0895
- Weber, G. F., Ashkar, S., Glimcher, M. J., and Cantor, H. (1996). Receptor-ligand interaction between CD44 and osteopontin (Eta-1). *Science* 271, 509–512. doi: 10.1126/science.271.5248.509
- Williams, K., Motiani, K., Giridhar, P. V., and Kasper, S. (2013). CD44 integrates signaling in normal stem cell, cancer stem cell and (pre)metastatic niches. *Exp. Biol. Med.* 238, 324–338. doi: 10.1177/1535370213480714
- Xu, Y., Swerlick, R. A., Sepp, N., Bosse, D., Ades, E. W., and Lawley, T. J. (1994). Characterization of expression and modulation of cell adhesion molecules on an immortalized human dermal microvascular endothelial cell line (HMEC-1). *J. Invest. Dermatol.* 102, 833–837. doi: 10.1111/1523-1747.ep12382086
- Yamane, N., Tsujitani, S., Makino, M., Maeta, M., and Kaibara, N. (1999). Soluble CD44 variant 6 as a prognostic indicator in patients with colorectal cancer. *Oncology* 56, 232–238. doi: 10.1159/000011970
- Yu, Q., and Stamenkovic, I. (1999). Localization of matrix metalloproteinase 9 to the cell surface provides a mechanism for CD44-mediated tumor invasion. *Genes Dev.* 13, 35–48. doi: 10.1101/gad.13.1.35
- Zeilstra, J., Joosten, S. P., van Andel, H., Tolg, C., Berns, A., Snoek, M., et al. (2014). Stem cell CD44v isoforms promote intestinal cancer formation in Apc(min) mice downstream of Wnt signaling. *Oncogene* 33, 665–670. doi: 10.1038/onc.2012.611

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Senbanjo and Chellaiah. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.