



Epithelial-Mesenchymal Transition and Metabolic Switching in Cancer: Lessons From Somatic Cell Reprogramming

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Epithelial-mesenchymal transition (EMT) and its critical roles during cancer progression have long been recognized and extensively reviewed. Recent studies on the generation of induced pluripotent stem cells (iPSCs) have established the connections among EMT, energy metabolism, DNA methylation, and histone modification. Since energy metabolism, DNA methylation, and histone modification are important for cancer development and there are common characteristics between cancer cells and stem cells, it is reasonable to identify mechanisms that have been established during both reprogramming and cancer progression. In the current review, we start from a brief review on EMT and related processes during cancer progression, and then switch to the EMT during somatic cell reprogramming. We summarize the connection between EMT and metabolic switch during reprogramming, and further review the involvements of DNA methylation and cell proliferation. The connections between EMT and mesenchymal-epithelial transition (MET) and cellular aspects including DNA methylation, histone modification and energy metabolism may provide potential new targets for cancer diagnosis and treatment.

Keywords: EMT, cancer, reprogramming, energy metabolism, glycolysis, OXPHOS

INTRODUCTION

Epithelial-mesenchymal transition (EMT) is defined as a biological process in which epithelial cells lose their characteristics and acquire mesenchymal features. During EMT, epithelial cells lose cell-cell junctions, apical-basal polarity, epithelial markers, and acquire cell motility, a spindle-cell shape, and mesenchymal markers. The concept of EMT was initially proposed as the epithelial-mesenchymal transformation by Elizabeth Hay in 1968 (Hay, 1968) as to describe the important cell changes in embryogenesis; it was later renamed EMT to distinguish it from neoplastic transformation (Nieto et al., 2016; Yang et al., 2020). EMT and its reverse process mesenchymal-epithelial transition (MET), display fundamental principles in diversified physiological and pathological progresses. During metazoan development, cells may sequentially undergo rounds of EMT and MET, as is seen in somite formation and heart development. EMT also occurs during

wound healing in adults. Additional evidences related to development and wound healing has previously been reviewed extensively (Thiery et al., 2009; Lim and Thiery, 2012; Nieto et al., 2016). EMT also plays important roles in cancer progression and tissue fibrosis (Nieto et al., 2016; Pastushenko and Blanpain, 2019; Williams et al., 2019). Interestingly, during the processes of embryonic stem cells (ESCs) differentiation and induced pluripotent stem cells (iPSCs) formation, EMT, and MET are highly relevant to the loss and acquisition of pluripotency (Pei et al., 2019). EMT and MET are widely involved in various biological scenarios and display highly plastic and dynamic manners during cell fate transitions (**Figure 1**).

Epithelial-mesenchymal transition is regulated at different levels by multiple factors, including cell signaling, transcriptional control, epigenetic modification, and post-translational modifications (**Figure 1**). Epithelial cells receive EMT-inducing signals from their niches. For example, cytokines such as transforming growth factor- β (TGF- β), fibroblast growth factor (FGF) family, epidermal growth factor (EGF), and hepatocyte growth factor (HGF) can induce or promote the EMT process (Zavadil and Bottinger, 2005; Thiery and Sleeman, 2006; Nieto et al., 2016; Williams et al., 2019). These EMT-inducing signals up-regulate specific transcription factors called EMT-TFs (e.g., SNAI, TWIST, and ZEB; Thiery and Sleeman, 2006; Nieto et al., 2016; Williams et al., 2019). EMT-TFs usually cooperate with miRNAs as well as epigenetic and/or post-translational regulators to control EMT (Ye and Weinberg, 2015; Nieto et al., 2016). For example, the miR-200 family, which includes miR-200a, miR-200b, miR-200c, miR-141, and miR-429 play important roles in repressing EMT by repressing the translation of ZEB1 and ZEB2 (Korpala et al., 2008; Park et al., 2008). Moreover, overexpression of miR-200 family members can upregulate *E-cadherin* expression (Park et al., 2008). miR-200 family expression is regulated by DNA methylation; hypermethylation of miR-200 loci lead to the silencing of miR-200 family expression, which promotes the EMT process and causes human tumors (Davalos et al., 2012). The miR-34 family is also known as to regulate EMT and suppress early phases of tumor metastasis. Ectopic expression of miR-34a prevents TGF- β -induced EMT, and miR-34a/b/c regulate SNAI1 expression by its 3'-UTR. miR-34a also suppresses SLUG and ZEB1. Conversely, SNAI1, and ZEB1 also regulate miR-34a/b/c expression by binding their promoters (Siemens et al., 2011). Additional details on EMT regulation by miRNA have been reviewed in multiple papers (Zaravinos, 2015; Suzuki, 2018; Asadzadeh et al., 2019).

Metabolic changes occur during development and cancer progression. For example, both ESCs and tumor cells prefer glycolysis, this similarity between stem cells and cancer cells indicate the intrinsic mechanism of stemness maintenance and metabolism. More recently, metabolic pathways including glycolysis, the TCA cycle, and amino acid and lipid metabolism have been reported as being involved in EMT, especially in tumor progression (Kang et al., 2019; Georgakopoulos-Soares et al., 2020; Sun and Yang, 2020). During tumor progression, cells prefer to obtain energy by increasing glucose conversion into lactate, even in oxygen-rich condition (Fischer and Bavister,

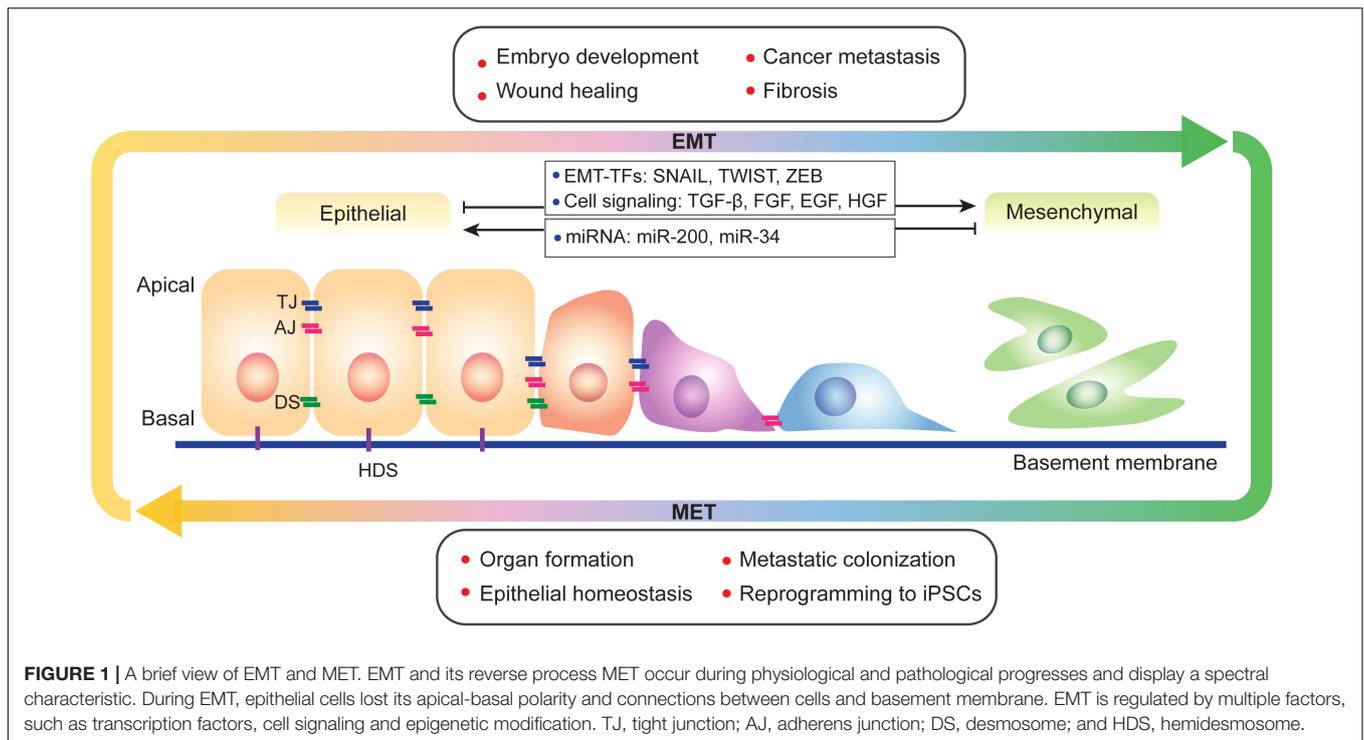
1993; Zhou et al., 2012; Setty et al., 2016). This specific energy metabolism pathway is called aerobic glycolysis. It was first observed in tumors and described by Otto Warburg in the 1920s, and thus, was named the Warburg effect (Warburg, 1956). This metabolic reprogramming depends upon an increase in glucose uptake and highly activated glycolysis. Glucose transporters GLUT1 and GLUT3 can be induced by hypoxia-inducible factor 1 α (HIF-1 α), which contributes to glucose uptake and promote EMT and tumor progression (Macheda et al., 2005). Enzymes participate in glycolysis, such as hexokinase 2 (HK2), phosphofructokinase (PFK), and pyruvate kinase M2 (PKM2) participate in glycolysis, play positive roles in glycolysis flux, and induce EMT (Luo et al., 2011; Patra et al., 2013; Kim et al., 2017). Tumor cells also show abnormal lipid metabolism, such as increased lipogenesis (Swinnen et al., 2006). Enzymes that participate in lipogenesis, such as acetyl-CoA carboxylase (ACC), fatty acid synthase (FASN), and acyl-CoA synthetase long chain family member (ACSL), all show a close relationship with cancer and EMT (Georgakopoulos-Soares et al., 2020). For example, FASN, which synthesizes of palmitate from acetyl-CoA and malonyl-CoA, has been widely reported in various types of cancer. FASN reportedly promotes EMT through TGF- β signaling in lung cancer (Yang et al., 2016) and ErbB receptors in breast cancer (Zaravinos, 2015). Lipids are also important components of the plasma membrane. CTP-phosphocholine cytidyltransferase (CTT), which is involved in phosphatidylcholine synthesis, is contributes to EMT in intestinal epithelial cells (Arsenault et al., 2013). Furthermore, amino acid metabolism is critical in EMT progression. Glutamine also plays important roles in energy supply. Glutaminase 1 (GLS1) and GLS2, which are involved in glutaminolysis can act as positive regulators of Snail (Choi and Park, 2018; Eiriksson et al., 2018). The important role of metabolism-related enzymes in cancer and EMT has resulted in many being selected as therapeutic targets. Additionally, metabolites from the pathways mentioned above can also regulate EMT through EMT-TFs and other epigenetic regulators, as we will discuss in this review.

This review discusses EMT progress and its regulators during cancer progression and iPSCs formation. We will also explore the relationship between metabolism and epigenetics in connection with EMT.

EMT IN CANCER AND STEM CELLS

EMT Plays Critical Roles in the Cancer Metastatic Process

The important roles of EMT during tumorigenesis and metastasis have been demonstrated for decades. Most lethal human malignancies are derived from epithelial tissues, including the breast, colon, pancreas, and liver (Ye and Weinberg, 2015). Approximately about 90% of cancer-associated deaths are caused by metastatic disease rather than primary tumors (Lambert et al., 2017). The EMT program confers upon these epithelial cells properties critical to invasion and metastatic dissemination including notably increased motility, invasiveness, and the ability



to degrade components of the extracellular matrix (ECM) components (Nieto et al., 2016). These complex metastatic cascades are orchestrated and coordinated by a series of master EMT-TFs that have been extensively explored (Craene and Berx, 2013; Lamouille et al., 2014). EMT-TFs facilitate the EMT process, but it is unclear whether EMT is indispensable for migration. Inhibiting EMT by overexpressing miR-200 did not affect lung metastasis when using an EMT lineage-tracing system in a spontaneous breast-to-lung metastasis model (Fischer et al., 2015). Furthermore, suppressing EMT by deleting *Snai* or *Twist* in a primary tumor did not decrease the invasion and metastasis of pancreatic carcinoma cells (Zheng et al., 2015). However, two subsequent reports pointed out the shortcomings in these experiments (Aiello et al., 2017; Ye et al., 2017). These opposing opinions reflect the complexity of the EMT processes that must to be carefully investigated both *in vivo* and *in vitro*.

One of the difficulties in studying EMT arises because the transitions between epithelial and mesenchymal states are not binary. Instead, carcinoma cells often exhibit a spectrum of epithelial-mesenchymal characteristics (Jolly et al., 2017; Mathieu et al., 2017; Zhang and Weinberg, 2018). E-cadherin, occludins, and cytokeratins are the most commonly used markers for the epithelial traits, and N-cadherin and vimentin are the most commonly used for the mesenchymal state (Thiery et al., 2009). Recent studies have shown that some cancer cells, such as breast, pancreatic, renal, lung, and colorectal cancers, express both epithelial and mesenchymal markers (Bronsert et al., 2014; Sampson et al., 2014; Zhang et al., 2014; Schliekelman et al., 2015; Hiew et al., 2018). In fact, many cancer cells may not undergo a complete EMT process, but rather steadily acquire these intermediate, or hybrid epithelial/mesenchymal E/M phenotypes.

The partial EMT or hybrid E/M phenotypes have multiple advantages compared to the complete EMT phenotype. It is suggested that these collectively migrating cells are the primary actors of metastasis (Jolly et al., 2015). To form distant metastases, cancer cells dissociate from the primary tumor, invade adjacent tissues, and intravasate into lymphatic and blood vessels to later colonize lymph nodes and distant organs (Nieto et al., 2016). Circulating tumor cells (CTCs) are defined as cancer cells that are released from a solid tumor and enter the peripheral blood; they are considered as biomarker of the metastatic process (Barcellos-Hoff et al., 2013). Most CTCs show hybrid E/M markers, suggesting an incomplete EMT. In addition, although EMT plays important roles in tumor progression, its reverse process, MET, also is significant in terms of dissemination. The last step of the invasion-metastasis cascade is termed colonization and largely depends on MET (Dongre and Weinberg, 2019).

Relationship Between Cancer Stem Cell Plasticity, EMT, and Metabolic Reprogramming

The concept of cancer stem cells (CSCs) is based on the observation that not all cells in tumors are equal. A small number of tumor cells display two key features that distinguish them from the others: self-renewal and differentiation potential (Foster and Archer, 1979; Zhang and Wang, 2008). With these features, CSCs are hypothesized to play essential roles in tumor metastasis, heterogeneity, drug resistance, and recurrence. Since EMT and the metastatic cascade are inextricably linked, the relationship between EMT and CSCs has been investigated. A number of studies have now shown that EMT programs

induce cancer cell stemness in many kinds of tissues (Dongre and Weinberg, 2019) and that EMT-TFs displayed the capacity to promote CSCs stemness in mouse and human models (Ye et al., 2015). Development of the cancer stem cell concept has been reviewed in several papers (Clevers, 2011; Batlle and Clevers, 2017). The isolation of CSCs from cancer cells and subsequent xenotransplantation assays provide evidence of the existence of CSCs. In the 1990s, CSCs were identified in acute myeloid leukemia (AML; Lapidot et al., 1994; Uckun et al., 1995; Bonnet and Dick, 1997). In 2003, CSCs were proved to be able to isolate from a solid tumor by a xenograft assay of small numbers of CD44⁺CD24^{-/low} cells which isolated from human breast cancer (Al-Hajj et al., 2003). Interestingly, cancer cells are observed exhibiting a plasticity that demonstrates the ability to switch between CSCs and non-CSCs in different cancers (Thankamony et al., 2020). For example, human basal breast cancer cells can switch from non-CSCs to CSCs by regulating the ZEB1 promoter (Chaffer et al., 2013). Another example for CSC plasticity is from studies on colorectal cancer. LGR5 is a marker of colorectal CSCs; a recent colorectal cancer study showed that LGR5 negative cells can become CSCs and lead to metastasis (Fumagalli et al., 2020).

Epithelial-mesenchymal transition-TFs are often found accompanied by features of stemness. *Slug* (also known as *Snai2*) is proved to be a key inducer of stemness (Guo et al., 2012; Nassour et al., 2012). Other EMT-TFs, such as *Twist*, *Snai1*, and *Six1* also induce stemness in various breast cancer models (McCoy et al., 2009; Morel et al., 2012; Ye et al., 2015). ZEB1 is required for stemness in pancreatic cancer; it inhibits members of the miR-200 family to maintain a stem-like phenotype (Wellner et al., 2009). EMT-related cell signaling factors, such as TGF- β , can be secreted in tumor stromal cells. This boosts metastatic potential and causes a poor-prognosis in colorectal cancer (Calon et al., 2015). Although the EMT process is associated with CSC characteristics, the relationship between EMT and stemness is still a controversial issue in tumorigenesis. Uncoupling EMT and stemness has been discussed in several studies and explained by the two events may occur in parallel (Nieto et al., 2016; Batlle and Clevers, 2017). EMT-TFs which critical in both EMT and CSCs are also provided evidence of uncoupling of the two events. For example, a study on Twist1 shows it is essential for initiate of skin tumorigenesis, however, Twist1 controlled tumor stemness independently of its EMT function (Beck et al., 2015). Recent studies have shown that cells with a hybrid E/M feature are much more likely to gain stemness (Bierie et al., 2017; Pastushenko et al., 2018). These studies may provide new insights for cancer therapy.

Metabolic reprogramming is one of the hallmarks of tumor progression. As discussed in the introduction, cancer cells change cell metabolism in diverse ways to gain energy and meet the requirements for proliferation. Glucose metabolism is considered the most studied metabolic change in CSCs. However, the glucose metabolism phenotype of CSCs, whether glycolysis or oxidative phosphorylation (OXPHOS), depends on both the tumor origin, and microenvironment (Thankamony et al., 2020). CSCs also show an interplay between glycolysis and OXPHOS (Yu et al., 2017). These metabolic characteristics increase the difficulty in

providing effective cancer therapies; other metabolic pathways and metabolites must be investigated further.

EMT-MET Processes in Physiological and Pathological Stem Cells

Stemness or stem cells are functionally defined as cells with the ability to self-renew and differentiate. In addition to cancer stem cells, other kinds of stem cells are isolated during development and in adults. One well accepted classification is that of ESCs and adult stem cells (ASCs). ESCs are derived from the inner cell mass of blastocysts and can differentiate into three germ layers and form chimeras (Evans and Kaufman, 1981; Martin, 1981), while ASCs are isolated from adult tissues in special niches and are capable of differentiating into several cell types depending on their origins (Shyh-Chang et al., 2013; Clevers, 2015). Although ESCs, ASCs, and CSCs are all considered as stem cells, their stemness is displayed in different ways. For example, both ESCs and ASCs show their differentiation abilities by forming normal tissues or cells. However, CSCs displayed their differentiation potential by promoting tumor heterogeneity (Clarke et al., 2006). These stem cells differ in terms of their gene expression profiles, transcriptional regulatory networks, and epigenetic modifications; however, they show their stemness in their consecutive EMT-MET changes. During embryonic development, early mesoderm cells are formed by EMT and migrate to generate intermediate mesoderm, chordamesoderm, paraxial mesoderm, and lateral plate mesoderm. These mesoderm cells then undergo MET to give rise to the urogenital system, notochord, somite, and somatopleure (Nieto, 2013). In contrast, CSCs caused by EMT undergo MET upon reaching an appropriate niche to become a new tumor (Nieto, 2013). Intriguingly, this EMT-MET progress has also been reported in pluripotency setup (Liu et al., 2013). Mouse embryonic fibroblasts (MEFs) can be reprogrammed into iPSCs by introducing exogenous *Oct4*, *Sox2*, *Klf4*, and *c-Myc*. Furthermore, reprogramming efficiency can be improved by sequentially expressing these transcriptional factors. All these phenomena imply that epithelial or even mesenchymal cells can reach a more active mesenchymal state to promote their plasticity.

EMT IN REPROGRAMMING

Early EMT Promotes Reprogramming

Direct reprogramming of somatic cells into iPSCs by defined factors is known as reprogramming (Takahashi and Yamanaka, 2006). A noticeable change that occurs during reprogramming is the transformation of MEF cells from a mesenchymal morphology into tightly packed colonies (Shu and Pei, 2014). This transition is defined as MET and is a critical step in acquiring pluripotency (Li et al., 2010; Samavarchi-Tehrani et al., 2010). Exogenous transcription factors *Oct4*, *Sox2*, and *c-Myc* are orchestrated to suppress *Snai1*, *Tgfb*, and *Tgfbr* to retain the non-mesenchymal characteristics of MEFs; *Klf4* induces epithelial properties by directly up-regulating *E-cadherin* (Li et al., 2010). Activating BMP signaling or repressing TGF- β signaling is

beneficial for iPSC generation by promoting MET progress (Li et al., 2010; Samavarchi-Tehrani et al., 2010). Not long after the discovery of the essential role of MET during reprogramming, another publication revealed that a sequential EMT–MET mechanism at the beginning of reprogramming can enhance reprogramming efficiency (Liu et al., 2013). Briefly, the sequential introduction of *Oct4* and *Klf4* first, followed by *c-Myc*, then *Sox2* (OK+M+S), can induce *Snai2* up-regulation and inhibit *E-cadherin* expression in the early stage of reprogramming. Further analysis revealed that this temporary EMT can generate iPSCs with a high efficiency of about 600% of basal level (Liu et al., 2013). Coincidentally, transient exogenous C/EBP α expression followed by the activation of Yamanaka factors (OSKM) in mouse primary B cells is capable of initiating EMT, which induces a 100-fold increase in reprogramming efficiency (Stefano et al., 2014). Hence, although MET is an essential event for reprogramming, having a more mesenchymal state at the early stage may provide a more facilitated state for cell fate transition.

EMT and Metabolic Regulation Play Multiple Roles During Reprogramming

The generation of iPSCs is a multi-step process. In addition to MET, another important change is the metabolic switch that occurs during reprogramming. Unlike somatic cells that use OXPHOS to gain energy, pluripotent stem cells, including ESCs and iPSCs, and prefer glycolysis (Zhang et al., 2012). Using small molecules to activate glycolysis and inhibit OXPHOS or upregulate glycolytic gene expression can promote the metabolic switch from oxidative phosphorylation to glycolysis (OGS) and accelerate reprogramming (Folmes et al., 2011; Zhang et al., 2012; Prigione et al., 2014; Cao et al., 2015). Since both OKSM-induced EMT–MET processes and facilitated OGS accelerate reprogramming, it remains an open question whether EMT and OGS cooperate to regulate iPSCs.

A recent study explored the relationship between early EMT and OGT (Sun et al., 2020). This study used 5C medium, a chemically-defined medium (He et al., 2015), to promote reprogramming by inducing a sequential EMT–MET process. Further analysis revealed that 5C medium increases glycolysis, inhibits OXPHOS, and accelerates the OGS process compared to a serum-containing medium. Early EMT and OGS may help cells overcome epigenetic barriers during reprogramming by upregulating five epigenetic factors, *Bmi1*, *Ctcf*, *Ezh2*, *Kdm2b*, and *Wdr5*, which have been proven to facilitate reprogramming in previous studies (Ang et al., 2011; Moon et al., 2011; Wang et al., 2011, 2017; Onder et al., 2012; **Figure 2**). The barriers that trapped cells at the pre-iPSC stage during reprogramming in serum containing medium (Chen et al., 2013b) may be overcome by the five epigenetic factors present in 5C medium. However, because the positive feedback loop between early EMT and facilitated OGS is too strong, further reprogramming is inhibited at the late stage by the induction of EMT and glycolysis to a level that is too high for further reprogramming and by preventing necessary MET (Sun et al., 2020). These findings are consistent with other works showing that EMT induced at the late stage of reprogramming significantly impairs the generation of

pluripotency (Liu et al., 2013). Furthermore, *HIF2 α* , which is upregulated in 5C medium, represses reprogramming at later stages (Mathieu et al., 2014).

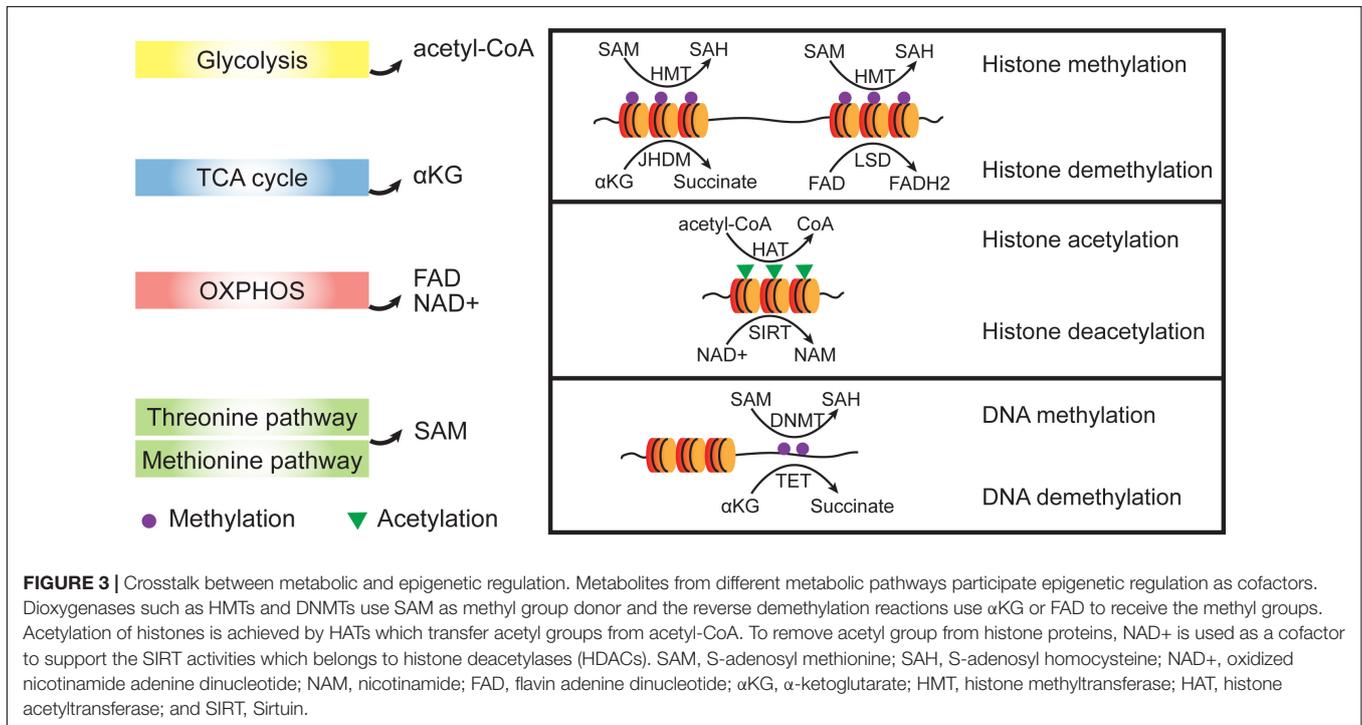
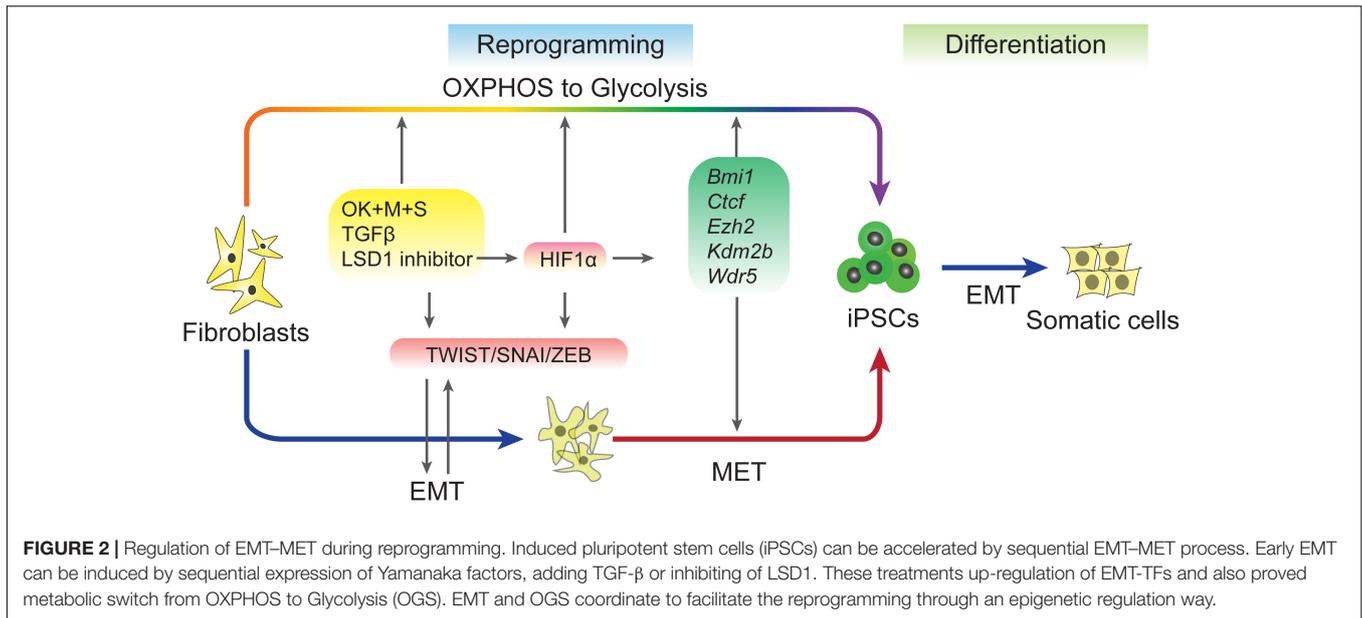
In addition to glycolysis, other metabolic pathways and metabolites are involved in EMT during reprogramming. A recent report showed that phospholipid remodeling is important for acquiring pluripotency by facilitating MET (Wu et al., 2019). This study demonstrated that the CDP-ethanolamine (CDP-Etn) pathway mechanically promotes reprogramming at the early stage through the CDP-Etn-Pebp1 axis to inhibit mesenchymal genes. Since metabolic regulation plays critical roles in EMT regulation and tumor progression, additional metabolic and EMT–MET connections in reprogramming need to be studied.

Metabolism Regulates EMT and Reprogramming Through Histone Modification

As discussed above, while OGS upregulates five epigenetic factors in an HIF1 α -dependent manner, early EMT promotes the expression of EMT–TFs that are enriched by both the five epigenetic factors and glycolytic genes (Sun et al., 2020). These reveal that the mesenchymal state may act as a link between metabolic and epigenetic regulation. Since metabolites generated during multiple metabolic processes are utilized in enzymatic reactions leading to epigenetic modifications and transcriptional regulation as previously reviewed (Ryall et al., 2015), we will focus on the crosstalk between EMT–MET, metabolism and epigenetic regulation during somatic cell reprogramming.

A vital role for metabolism in regulating cell fate has been derived from studies documenting rapid and dynamic changes in substrate utilization. Metabolites, including α -ketoglutarate (α KG), acetyl-coA, S-adenosyl methionine (SAM), and flavin adenine dinucleotide (FAD), regulate many of the important cell fate conversions by epigenetic catalytic reactions (Ryall et al., 2015; Wu et al., 2016; **Figure 3**). For example, glycolysis-produced acetyl-coA have been reportedly controls the early differentiation of ESCs by regulation histone acetylation (Moussaieff et al., 2015).

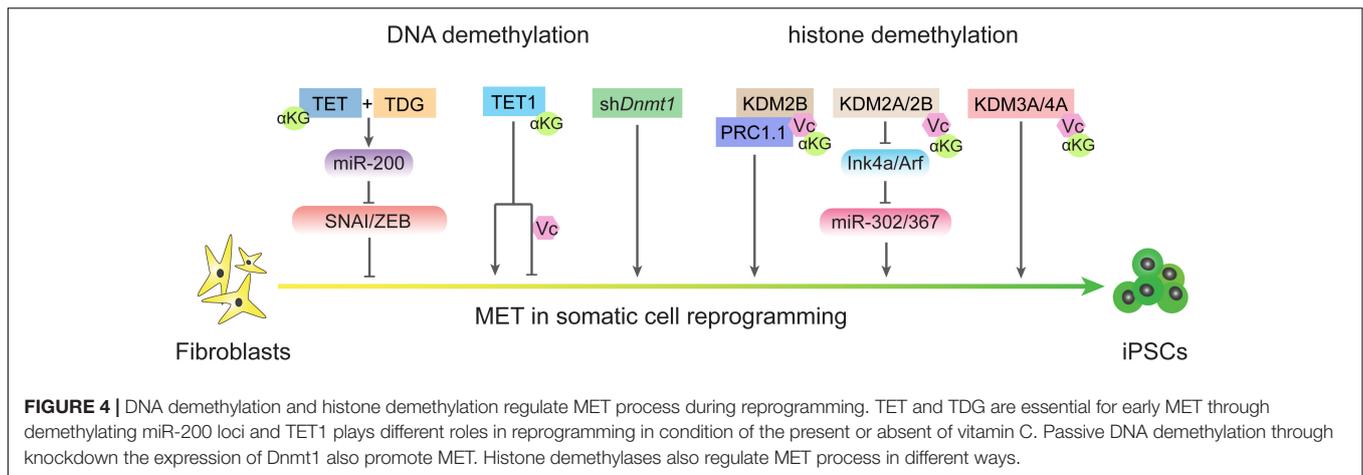
S-adenosyl methionine, which functions as a methyl donor for both DNA and histone methylation, is important for the maintaining of pluripotency in both mouse and human ESCs (Ng et al., 2013; Shiraki et al., 2014). The cellular α KG/succinate ratio contributes to the ability of ESCs to suppress differentiation (Carey et al., 2015). α KG/succinate ratios are involved in the regulation of a large family of α KG-dependent dioxygenases, such as Jumonji C (JmjC)-domain-containing histone demethylases (Carey et al., 2015; **Figure 4**). JHDM1A and JHDM1B (also known as KDM2A and KDM2B, respectively) belong to the Jumonji family of proteins and have been shown to demethylate H3K36me2/3 (He et al., 2008). Early studies showed that JHDM1A/1B inhibit Ink4a/Arf and activate miR-302/367 to promote reprogramming (Wang et al., 2011). A recent investigation showed that JHDM1B works as a component of the PRC1.1 complex and cooperates with BMP signaling to upregulate reprogramming efficiency by promoting MET



(Zhou et al., 2017). Consistently, histone demethylases JMJD1 and JMJD2 (also known as KDM3A and KDM4A, respectively) function as the on/off switch for pre-iPSC fate by regulating H3K9 methylation status at core pluripotency loci (Chen et al., 2013b). In addition, lysine-specific demethylase 1 (LSD1, also known as KDM1A) functions through an FAD-dependent oxidative reaction to specifically catalyze the demethylation of H3K4me1/2 or H3K9me1/2 (Shi et al., 2004; Amente et al., 2013). Accordingly, inhibiting LSD1 by shRNA or a small molecular inhibitor can promote reprogramming at the early

stage by increasing both OGS and exogenous transcriptional factors expression (Sun et al., 2016).

Histones can also be acetylated and deacetylated by histone acetyltransferases (HATs) and histone deacetylases (HDACs), respectively. HATs mediate the acetyl group transfer from acetyl-CoA, a metabolite from glycolysis. A higher level of acetyl-CoA is found in mouse ESCs compared to that of differentiated cells (Wang et al., 2020) and reportedly controls the early differentiation of ESCs by regulating histone acetylation (Moussaieff et al., 2015). In addition to histone acetylation,



non-histone acetylation is also related to pluripotency. A recent study showed that the inhibition of CBP/EP300, which is closely related HATs, can repress the expression of mesenchymal transcription factor PRRX1 and promote reprogramming (Ebrahimi et al., 2019). HDACs are grouped into four classes. Class I (HDACs 1–3 and 8), class II (HDACs 4–7, 9, and 10), and class IV (HDAC 11) are zinc-dependent metalloproteins; class III (SIRT1–7) are NAD⁺-dependent proteins (Emmett and Lazar, 2019). Class I HDAC1–HDAC2 can form multiprotein complexes named SIN3A, NuRD, and CoREST, respectively. These HDAC containing complexes play different roles in establishing and maintaining pluripotency in a context-dependent manner. For example, the SIN3A/HDAC2 complex cooperates with NANOG to promote reprogramming (Saunders et al., 2017). However, HDAC2 reportedly acts as a barrier to reprogramming in other studies (Anokye-Danso et al., 2011; Wei et al., 2015). Interestingly, we noticed that miR-302/367 can promote reprogramming by inhibiting HDAC2 (Anokye-Danso et al., 2011), but in a contemporaneous study by Liao et al., miR-302/367 accelerated MET by inhibiting TGFBR2 and promoting E-cadherin expression (Liao et al., 2011). These studies indicate that the relationship between HDACs and EMT, and indeed, deacetylation by HDACs, can occur without histones. Snail interaction with HDAC1 and HDAC2 is dependent upon its SNAG domain and can mediate E-cadherin repression (Peinado et al., 2004). Sirtuins (SIRT1–7) belong to class III HDACs and depend on NAD⁺ to exhibit their enzyme activity. SIRT1 enhances reprogramming via SOX2 hypoacetylation (Mu et al., 2015), as well as through the miR34a–SIRT1–p53 axis (Lee et al., 2012). SIRT7 reportedly reverse metastatic phenotypes in tumors (Malik et al., 2015) and may affect MET during reprogramming (Hsu et al., 2018).

DNA Methylation Regulates EMT-MET During Reprogramming

DNA methylation is another area of crosstalk between metabolism and epigenetics that also plays critical roles in regulating EMT-MET during reprogramming (Figure 2). In mammals, methyl groups supplied by SAM are added at the

5-carbon of cytosines by the catalysis of DNA methyltransferases (DNMTs; Wu and Zhang, 2014). DNA methylation can be removed by both TETs-mediated active DNA demethylation and cell cycle-dependent passive DNA demethylation (Wu and Zhang, 2014, 2017). Since reprogramming occurs along with DNA demethylation in pluripotency-related loci, the mechanisms of DNA demethylation during iPSCs formation has been investigated. TET proteins belong to α KG-dependent dioxygenases and require Fe²⁺ and vitamin C (Vc) as assistant (Ngo et al., 2019). Both TET1 and TET2 promote reprogramming (Doege et al., 2012; Costa et al., 2013; Gao et al., 2013), and under certain conditions, *Tet1* even could replace *Oct4* (Gao et al., 2013). Furthermore, active demethylation promoted by TETs and TDG is essential to reactive the miR-200 family that enables MET by inhibiting SNAI/ZEB in the reprogramming process (Hu et al., 2014). Repressing DNA methylation by *shDnmt1* or promoting passive demethylation by *shp53*-induced proliferation acceleration also promotes MET and facilitates iPSCs generation (He et al., 2017b, 2019). Further analysis has revealed that both *shDnmt1*-induced passive DNA demethylation and TET1-induced active DNA demethylation prefer hemi-methylated CpG sites that are enriched at the loci of core pluripotency genes and epithelial markers (He et al., 2019). The effects of DNA methylation on EMT-TFs were also studied in several types of cancer (Lee and Kong, 2016). Thus, metabolites from diverse metabolic pathways contribute to cell fate conversions by modulating epigenetic properties and show a high correlation with EMT-MET.

Intriguingly, further study of TET1 found that the physiological concentration of vitamin C increases TET1 activity, which in turn impairs the MET process and inhibits reprogramming (Chen et al., 2013a). In the absence of vitamin C, TET1 induces DNA demethylation on the loci of core pluripotency genes and epithelial markers rather than mesenchymal markers, which results in MET and accelerated reprogramming. The DNA demethylation activity of TET1 is enhanced in the presence of vitamin C. Because DNA methylation on the loci of core pluripotency genes and epithelial markers is already at a low level, the expression of core pluripotency genes and epithelial markers is not further

affected. However, increased TET1 activity can induce DNA demethylation on the loci of mesenchymal markers, which in turn suppresses the expression of core pluripotency genes and epithelial markers. In addition to their DNA demethylation function, TET proteins also have important functions that are independent of catalytic activity. For example, TET1 could be recruited by Polycomb repressive complex 2 (PRC2) at H3K27me3 positive regions in ESCs (Neri et al., 2013). TET1 also interacts with SIN3A/HDAC complex (Vella et al., 2013) which is reported to promote pluripotency (Saunders et al., 2017). TET proteins including TET1, TET2, and TET3, are reported associated with the O-GlcNAc transferase (OGT) (Deplus et al., 2013; Vella et al., 2013; Chen et al., 2013c) in ESCs. OGT or O-GlcNAcylation could further recruit SET/COMPASS (Deplus et al., 2013) or modified transcription factors by O-GlcNAc, such as Pou5f1 (Constable et al., 2017), that may promote somatic cell reprogramming. Recent studies show that O-GlcNAcylation effects EMT through multiple pathways and promote cancer metastasis (Lucena et al., 2016; Harosh-Davidovich and Khalaila, 2018; Jiang et al., 2019; Feng et al., 2020), these may set up the relationship between TET and EMT. Tsai et al. reports TET1 could interact with HIF-1 α and HIF-2 α to regulate hypoxia-induced EMT (Tsai et al., 2014).

Therefore, regulation of DNA methylation can affect both EMT and its reverse process, MET.

CONCLUDING REMARKS

Although the field of EMT research has developed rapidly, many critical questions remain unanswered. For example, the driving force initiating EMT and the manner by which dose metabolism influences EMT, as well as the function of the mesenchymal state during the EMT-MET process are still unknown. Since transcription factors are important for cell identity, EMT-TFs are considered critical for losing epithelial characteristics and gaining mesenchymal phenotypes (Nieto, 2013). Hence, the regulation of EMT-TFs cooperating with cell signaling *in vitro* and transcriptional and epigenetic controls *in vivo* need to be carefully considered when studying EMT.

Mesenchymal State Displays Plasticity of Cells

The concept of EMT comes from studies on early embryogenesis and is now widely investigated in cancer and stem cell biology. The transitions between mesenchymal and epithelial states demonstrate the plasticity of the cells. The sequential transition of EMT-MET not only occurs during tumor metastasis and somatic cell reprogramming as discussed above, but is also observed in cell differentiation and transdifferentiation. For example, a sequential EMT-MET drives the differentiation of human ESCs toward hepatocytes (Li et al., 2017). Activin A-induced formation of definitive endoderm (DE) accompanies a synchronous EMT mediated by autocrine TGF- β signaling and activates SNAI1 to initiate EMT followed by MET (Li et al., 2017). In addition, a temporary EMT-MET can increase the conversion efficiency of mouse astrocytes into induced dopamine

neurons (Cervo et al., 2017) and human gastric epithelial cells into multipotent endodermal progenitors (Wang et al., 2016). Interestingly, in the early study of 5C medium, sequential EMT-MET was also observed during the transition of MEFs to neuron-like cells (He et al., 2017a). All of this evidence suggests that the mesenchymal state may be a necessary state for cell fate transition.

Lessons From Somatic Cell Reprogramming Provide New Insights for Cancer Therapy

A recent study revealed the cooperation of EMT-MET and OGS during reprogramming as discussed above (Sun et al., 2020). However, coordinated regulation between EMT-MET and OGS should be investigated in other types of cell fate transition for common regulatory mechanisms. Metabolic shifts can control EMT during tumor metastasis, and metabolites from different metabolic pathways are involved in epigenetic regulation (Ryall et al., 2015; Kang et al., 2019). Both cancer cells and ESCs prefer to obtain energy through glycolysis, implying an intrinsic connection between the two kinds of cells. Furthermore, lessons can be learned from somatic cell reprogramming. For example, AML can be caused by the mutation of isocitrate dehydrogenase (IDH) IDH1 and IDH2 or TET2 (Figuroa et al., 2010; Qivoron et al., 2011). TET2 is a dioxygenase and its activity depends upon α KG, Fe²⁺, and Vc (Blaschke et al., 2013; Yin et al., 2013; Chen et al., 2013a). IDH1 and IDH2 convert isocitrate to α KG, which supports TET2 activity. A mutation in IDH1 or IDH2 causes the accumulation of 2-hydroxyglutarate (2-HG) and inhibits TET2 enzyme activity, resulting increased DNA methylation that drives AML progression (Lu et al., 2012). Previous studies on TETs in ESCs and reprogramming have showed that Vc can activate TETs as a cofactor, and a high dose of Vc has rescued TET2 deletion in a mouse model of leukemia (Agathocleous et al., 2017; Cimmino et al., 2017).

Prospects for Cancer Therapy

Recent studies on CSCs focusing on their plasticity and resistance to therapy have been discussed in several other reviews (Batlle and Clevers, 2017; Najafi et al., 2019; Thankamony et al., 2020). Firstly, enzymes involved in metabolic reprogramming during tumor progression can be used as biomarkers for the therapeutic targeting of cancers (Sun and Yang, 2020). In addition to IDH, FASN in lipid synthesis and GLS1 in glutamine metabolism have been reported as targets for breast cancer and colorectal cancer, respectively, Sun and Yang (2020). Secondly, regulators involved in EMT can be used biomarkers and for therapeutic targeting (Williams et al., 2019). For example, both the transcriptional and epigenetic regulation of EMT-TFs could be treatment targets (Sun and Yang, 2020). Thirdly, since CSCs contribute to tumor resistance and recurrence, special attention must be paid to CSCs. Therefore, this review has discussed EMT in the context of both cancer and reprogramming, and explores the relationship between EMT, metabolism, and epigenetic regulation. These mechanisms could provide new scenarios for regenerative medicine and cancer therapy.

AUTHOR CONTRIBUTIONS

LG, XL, and QL wrote this manuscript. HZ, JC, FW, and JL helped to improve this manuscript. JC, HZ, and LG approved the final version of this submission. All authors contributed to the article and approved the submitted version.

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REFERENCES

- Agathocleous, M., Meacham, C., Burgess, R., Piskounova, E., Zhao, Z., Crane, G., et al. (2017). Ascorbate regulates haematopoietic stem cell function and leukaemogenesis. *Nature* 549, 476–481. doi: 10.1038/nature23876
- Aiello, N., Brabletz, T., Kang, Y., Nieto, M., Weinberg, R., and Stanger, B. (2017). Upholding a role for EMT in pancreatic cancer metastasis. *Nature* 547, E7–E8. doi: 10.1038/nature22963
- Al-Hajj, M., Wicha, M., Benito-Hernandez, A., Morrison, S., and Clarke, M. (2003). Prospective identification of tumorigenic breast cancer cells. *Proc. Natl. Acad. Sci. U.S.A.* 100, 3983–3988. doi: 10.1073/pnas.0530291100
- Amente, S., Lania, L., and Majello, B. (2013). The histone LSD1 demethylase in stemness and cancer transcription programs. *Biochim. Biophys. Acta* 1829, 981–986. doi: 10.1016/j.bbagr.2013.05.002
- Ang, Y.-S., Tsai, S.-Y., Lee, D.-F., Monk, J., Su, J., Ratnakumar, K., et al. (2011). Wdr5 mediates self-renewal and reprogramming via the embryonic stem cell core transcriptional network. *Cell* 145, 183–197. doi: 10.1016/j.cell.2011.03.003
- Anokye-Danso, F., Trivedi, C. M., Juhr, D., Gupta, M., Cui, Z., Tian, Y., et al. (2011). Highly efficient miRNA-mediated reprogramming of mouse and human somatic cells to pluripotency. *Cell Stem Cell* 8, 376–388. doi: 10.1016/j.stem.2011.03.001
- Arsenault, D. J., Yoo, B. H., Rosen, K. V., and Ridgway, N. D. (2013). ras-Induced up-regulation of CTP:phosphocholine cytidyltransferase alpha contributes to malignant transformation of intestinal epithelial cells. *J. Biol. Chem.* 288, 633–643. doi: 10.1074/jbc.M112.347682
- Asadzadeh, Z., Mansoori, B., Mohammadi, A., Aghajani, M., Haji-Asgarzadeh, K., Safarzadeh, E., et al. (2019). microRNAs in cancer stem cells: biology, pathways, and therapeutic opportunities. *J. Cell. Physiol.* 234, 10002–10017. doi: 10.1002/jcp.27885
- Barcellos-Hoff, M., Lyden, D., and Wang, T. (2013). The evolution of the cancer niche during multistage carcinogenesis. *Nat. Rev. Cancer* 13, 511–518. doi: 10.1038/nrc3536
- Battle, E., and Clevers, H. (2017). Cancer stem cells revisited. *Nat. Med.* 23, 1124–1134. doi: 10.1038/nm.4409
- Beck, B., Lapouge, G., Rorive, S., Drogat, B., Desaedelaere, K., Delafaille, S., et al. (2015). Different levels of Twist1 regulate skin tumor initiation, stemness, and progression. *Cell Stem Cell* 16, 67–79. doi: 10.1016/j.stem.2014.12.002
- Bierie, B., Pierce, S. E., Kroeger, C., Stover, D. G., Pattabiraman, D. R., Thiru, P., et al. (2017). Integrin-beta4 identifies cancer stem cell-enriched populations of partially mesenchymal carcinoma cells. *Proc. Natl. Acad. Sci. U.S.A.* 114, E2337–E2346. doi: 10.1073/pnas.1618298114
- Blaschke, K., Ebata, K., Karimi, M., Zepeda-Martinez, J., Goyal, P., Mahapatra, S., et al. (2013). Vitamin C induces Tet-dependent DNA demethylation and a blastocyst-like state in ES cells. *Nature* 500, 222–226. doi: 10.1038/nature12362
- Bonnet, D., and Dick, J. E. (1997). Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nat. Med.* 3, 730–737. doi: 10.1038/nm0797-730
- Bronsart, P., Enderle-Ammour, K., Bader, M., Timme, S., Kuehs, M., Csanadi, A., et al. (2014). Cancer cell invasion and EMT marker expression: a three-dimensional study of the human cancer-host interface. *J. Pathol.* 234, 410–422. doi: 10.1002/path.4416
- Calon, A., Lonardo, E., Berenguer-Llergo, A., Espinet, E., Hernando-Mombalona, X., Iglesias, M., et al. (2015). Stromal gene expression defines poor-prognosis subtypes in colorectal cancer. *Nat. Genet.* 47, 320–329. doi: 10.1038/ng.3225
- Cao, Y., Guo, W., Tian, S., He, X., Wang, X., Liu, X., et al. (2015). miR-290/371-Mbd2-Myc circuit regulates glycolytic metabolism to promote pluripotency. *EMBO J.* 34, 609–623. doi: 10.15252/embo.201490441
- Carey, B. W., Finley, L. W. S., Cross, J. R., Allis, C. D., and Thompson, C. B. (2015). Intracellular α -ketoglutarate maintains the pluripotency of embryonic stem cells. *Nature* 518, 413–416. doi: 10.1038/nature13981
- Cervo, P. R. D. V., Romanov, R. A., Spigolon, G., Masini, D., Martin-Montanez, E., Toledo, E. M., et al. (2017). Induction of functional dopamine neurons from human astrocytes *in vitro* and mouse astrocytes in a Parkinson's disease model. *Nat. Biotechnol.* 35, 444–452. doi: 10.1038/nbt.3835
- Chaffer, C. L., Marjanovic, N. D., Lee, T., Bell, G., Kleer, C. G., Reinhardt, F., et al. (2013). Poised chromatin at the ZEB1 promoter enables breast cancer cell plasticity and enhances tumorigenicity. *Cell* 154, 61–74. doi: 10.1016/j.cell.2013.06.005
- Chen, J., Guo, L., Zhang, L., Wu, H., Yang, J., Liu, H., et al. (2013a). Vitamin C modulates TET1 function during somatic cell reprogramming. *Nat. Genet.* 45, 1504–1509. doi: 10.1038/ng.2807
- Chen, J., Liu, H., Liu, J., Qi, J., Wei, B., Yang, J., et al. (2013b). H3K9 methylation is a barrier during somatic cell reprogramming into iPSCs. *Nat. Genet.* 45, 34–42. doi: 10.1038/ng.2491
- Chen, Q., Chen, Y., Bian, C., Fujiki, R., and Yu, X. (2013c). TET2 promotes histone O-GlcNAcylation during gene transcription. *Nature* 493, 561–564. doi: 10.1038/nature11742
- Choi, Y. K., and Park, K. G. (2018). Targeting glutamine metabolism for cancer treatment. *Biomol. Ther.* 26, 19–28. doi: 10.4062/biomolther.2017.178
- Cimmino, L., Dolgalev, I., Wang, Y., Yoshimi, A., Martin, G., Wang, J., et al. (2017). Restoration of TET2 function blocks aberrant self-renewal and leukemia progression. *Cell* 170, 1079–1095.e20. doi: 10.1016/j.cell.2017.07.032
- Clarke, M., Dick, J., Dirks, P., Eaves, C., Jamieson, C., Jones, D., et al. (2006). Cancer stem cells—perspectives on current status and future directions: AACR Workshop on cancer stem cells. *Cancer Res.* 66, 9339–9344. doi: 10.1158/0008-5472.CAN-06-3126
- Clevers, H. (2011). The cancer stem cell: premises, promises and challenges. *Nat. Med.* 17, 313–319. doi: 10.1038/nm.2304
- Clevers, H. (2015). STEM CELLS. What is an adult stem cell? *Science* 350, 1319–1320. doi: 10.1126/science.aad7016
- Constable, S., Lim, J., Vaidyanathan, K., and Wells, L. (2017). O-GlcNAc transferase regulates transcriptional activity of human Oct4. *Glycobiology* 27, 927–937. doi: 10.1093/glycob/cwx055
- Costa, Y., Ding, J., Theunissen, T., Faiola, F., Hore, T., Shliha, P., et al. (2013). NANOG-dependent function of TET1 and TET2 in establishment of pluripotency. *Nature* 495, 370–374. doi: 10.1038/nature11925
- Craene, B. D., and Bex, G. (2013). Regulatory networks defining EMT during cancer initiation and progression. *Nat. Rev. Cancer* 13, 97–110. doi: 10.1038/nrc3447
- Davalos, V., Moutinho, C., Villanueva, A., Boque, R., Silva, P., Carneiro, F., et al. (2012). Dynamic epigenetic regulation of the microRNA-200 family mediates

- epithelial and mesenchymal transitions in human tumorigenesis. *Oncogene* 31, 2062–2074. doi: 10.1038/ncr.2011.383
- Deplus, R., Delatte, B., Schwinn, M., Defrance, M., Mendez, J., Murphy, N., et al. (2013). TET2 and TET3 regulate GlcNAcylation and H3K4 methylation through OGT and SET1/COMPASS. *EMBO J.* 32, 645–655. doi: 10.1038/emboj.2012.357
- Doegre, C., Inoue, K., Yamashita, T., Rhee, D., Travis, S., Fujita, R., et al. (2012). Early-stage epigenetic modification during somatic cell reprogramming by Parp1 and Tet2. *Nature* 488, 652–655. doi: 10.1038/nature11333
- Dongre, A., and Weinberg, R. (2019). New insights into the mechanisms of epithelial-mesenchymal transition and implications for cancer. *Nat. Rev. Mol. Cell Biol.* 20, 69–84. doi: 10.1038/s41580-018-0080-4
- Ebrahimi, A., Sevinc, K., Gurhan Sevinc, G., Cribbs, A. P., Philpott, M., Uyulur, F., et al. (2019). Bromodomain inhibition of the coactivators CBP/EP300 facilitate cellular reprogramming. *Nat. Chem. Biol.* 15, 519–528. doi: 10.1038/s41589-019-0264-z
- Eiriksson, F. F., Rolfsson, O., Ogmundsdottir, H. M., Haraldsson, G. G., Thorsteinsdottir, M., and Halldorsson, S. (2018). Altered plasmalogen content and fatty acid saturation following epithelial to mesenchymal transition in breast epithelial cell lines. *Int. J. Biochem. Cell Biol.* 103, 99–104. doi: 10.1016/j.biocel.2018.08.003
- Emmett, M. J., and Lazar, M. A. (2019). Integrative regulation of physiology by histone deacetylase 3. *Nat. Rev. Mol. Cell Biol.* 20, 102–115. doi: 10.1038/s41580-018-0076-0
- Evans, M. J., and Kaufman, M. H. (1981). Establishment in culture of pluripotential cells from mouse embryos. *Nature* 292, 154–156. doi: 10.1038/292154a0
- Feng, D., Sheng-Dong, L., Tong, W., and Zhen-Xian, D. (2020). O-GlcNAcylation of RAF1 increases its stabilization and induces the renal fibrosis. *Biochim. Biophys. Acta* 1866:165556. doi: 10.1016/j.bbadis.2019.165556
- Figuerola, M., Abdel-Wahab, O., Lu, C., Ward, P., Patel, J., Shih, A., et al. (2010). Leukemic IDH1 and IDH2 mutations result in a hypermethylation phenotype, disrupt TET2 function, and impair hematopoietic differentiation. *Cancer Cell* 18, 553–567. doi: 10.1016/j.ccr.2010.11.015
- Fischer, B., and Bavister, B. D. (1993). Oxygen tension in the oviduct and uterus of rhesus monkeys, hamsters and rabbits. *J. Reprod. Fertil.* 99, 673–679. doi: 10.1530/jrf.0.0990673
- Fischer, K., Durrans, A., Lee, S., Sheng, J., Li, F., Wong, S., et al. (2015). Epithelial-to-mesenchymal transition is not required for lung metastasis but contributes to chemoresistance. *Nature* 527, 472–476. doi: 10.1038/nature15748
- Folmes, C., Nelson, T., Martinez-Fernandez, A., Arrell, D., Lindor, J., Dzeja, P., et al. (2011). Somatic oxidative bioenergetics transitions into pluripotency-dependent glycolysis to facilitate nuclear reprogramming. *Cell Metab.* 14, 264–271. doi: 10.1016/j.cmet.2011.06.011
- Foster, J., and Archer, S. (1979). Birth order and intelligence: an immunological interpretation. *Percept. Mot. Skills* 48, 79–93. doi: 10.2466/pms.1979.48.1.79
- Fumagalli, A., Oost, K. C., Kester, L., Morgner, J., Bornes, L., Bruens, L., et al. (2020). Plasticity of Lgr5-negative cancer cells drives metastasis in colorectal cancer. *Cell Stem Cell* 26, 569–578.e7. doi: 10.1016/j.stem.2020.02.008
- Gao, Y., Chen, J., Li, K., Wu, T., Huang, B., Liu, W., et al. (2013). Replacement of Oct4 by Tet1 during iPSC induction reveals an important role of DNA methylation and hydroxymethylation in reprogramming. *Cell Stem Cell* 12, 453–469. doi: 10.1016/j.stem.2013.02.005
- Georgakopoulos-Soares, I., Chartoumpakis, D. V., Kyriazopoulou, V., and Zaravinos, A. (2020). EMT factors and metabolic pathways in cancer. *Front. Oncol.* 10:499. doi: 10.3389/fonc.2020.00499
- Guo, W., Keckesova, Z., Donaher, J., Shibue, T., Tischler, V., Reinhardt, F., et al. (2012). Slug and Sox9 cooperatively determine the mammary stem cell state. *Cell* 148, 1015–1028. doi: 10.1016/j.cell.2012.02.008
- Harosh-Davidovich, S., and Khalaila, I. (2018). O-GlcNAcylation affects beta-catenin and E-cadherin expression, cell motility and tumorigenicity of colorectal cancer. *Exp. Cell Res.* 364, 42–49. doi: 10.1016/j.yexcr.2018.01.024
- Hay, E. (1968). “Organization and fine structure of epithelium and mesenchyme in the developing chick embryos,” in *Proceedings of the 18th Hahnemann Symposium Epithelial-Mesenchymal Interactions*, eds R. Fleischmajer and R. E. Billingham (Baltimore, Maryland: Williams & Wilkins), 31–55.
- He, J., Kallin, E. M., Tsukada, Y. I., and Zhang, Y. (2008). The H3K36 demethylase Jhdmlb/Kdm2b regulates cell proliferation and senescence through p15Ink4b/Ink4b. *Nat. Struct. Mol. Biol.* 15, 1169–1175. doi: 10.1038/nsmb.1499
- He, S., Chen, J., Zhang, Y., Zhang, M., Yang, X., Li, Y., et al. (2017a). Sequential EMT-MET induces neuronal conversion through Sox2. *Cell Discov.* 3:17017. doi: 10.1038/celldisc.2017.17
- He, S., Guo, Y., Zhang, Y., Li, Y., Feng, C., Li, X., et al. (2015). Reprogramming somatic cells to cells with neuronal characteristics by defined medium both *in vitro* and *in vivo*. *Cell Regen.* 4:12. doi: 10.1186/s13619-015-0027-6
- He, S., Sun, H., Lin, L., Zhang, Y., Chen, J., Liang, L., et al. (2017b). Passive DNA demethylation preferentially up-regulates pluripotency-related genes and facilitates the generation of induced pluripotent stem cells. *J. Biol. Chem.* 292, 18542–18555. doi: 10.1074/jbc.M117.810457
- He, S., Wang, F., Zhang, Y., Chen, J., Liang, L., Li, Y., et al. (2019). Hemimethylated CpG sites connect Dnmt1-knockdown-induced and Tet1-induced DNA demethylation during somatic cell reprogramming. *Cell Discov.* 5:11. doi: 10.1038/s41421-018-0074-6
- Hiew, M., Cheng, H., Huang, C., Chong, K., Cheong, S., Choo, K., et al. (2018). Incomplete cellular reprogramming of colorectal cancer cells elicits an epithelial/mesenchymal hybrid phenotype. *J. Biomed. Sci.* 25:57. doi: 10.1186/s12929-018-0461-1
- Hsu, Y. C., Wu, Y. T., Tsai, C. L., and Wei, Y. H. (2018). Current understanding and future perspectives of the roles of sirtuins in the reprogramming and differentiation of pluripotent stem cells. *Exp. Biol. Med.* 243, 563–575. doi: 10.1177/1535370218759636
- Hu, X., Zhang, L., Mao, S.-Q., Li, Z., Chen, J., Zhang, R.-R., et al. (2014). Tet and TDG Mediate DNA demethylation essential for mesenchymal-to-epithelial transition in somatic cell reprogramming. *Cell Stem Cell* 14, 512–522. doi: 10.1016/j.stem.2014.01.001
- Jiang, M., Xu, B., Li, X., Shang, Y., Chu, Y., Wang, W., et al. (2019). O-GlcNAcylation promotes colorectal cancer metastasis via the miR-101-O-GlcNAc/EZH2 regulatory feedback circuit. *Oncogene* 38, 301–316. doi: 10.1038/s41388-018-0435-5
- Jolly, M., Boaretto, M., Huang, B., Jia, D., Lu, M., Ben-Jacob, E., et al. (2015). Implications of the hybrid epithelial/mesenchymal phenotype in metastasis. *Front. Oncol.* 5:155. doi: 10.3389/fonc.2015.00155
- Jolly, M. K., Ware, K. E., Gilja, S., Somarelli, J. A., and Levine, H. (2017). EMT and MET: necessary or permissive for metastasis? *Mol. Oncol.* 11, 755–769. doi: 10.1002/1878-0261.12083
- Kang, H., Kim, H., Lee, S., Youn, H., and Youn, B. (2019). Role of metabolic reprogramming in epithelial-mesenchymal transition (EMT). *Int. J. Mol. Sci.* 20, 2042. doi: 10.3390/ijms20082042
- Kim, N. H., Cha, Y. H., Lee, J., Lee, S. H., Yang, J. H., Yun, J. S., et al. (2017). Snail reprograms glucose metabolism by repressing phosphofructokinase PFKP allowing cancer cell survival under metabolic stress. *Nat. Commun.* 8:14374. doi: 10.1038/ncomms14374
- Korpal, M., Lee, E. S., Hu, G., and Kang, Y. (2008). The miR-200 family inhibits epithelial-mesenchymal transition and cancer cell migration by direct targeting of E-cadherin transcriptional repressors ZEB1 and ZEB2. *J. Biol. Chem.* 283, 14910–14914. doi: 10.1074/jbc.C800074200
- Lambert, A. W., Pattabiraman, D. R., and Weinberg, R. A. (2017). Emerging Biological Principles of Metastasis. *Cell* 168, 670–691. doi: 10.1016/j.cell.2016.11.037
- Lamouille, S., Xu, J., and Derynck, R. (2014). Molecular mechanisms of epithelial-mesenchymal transition. *Nat. Rev. Mol. Cell Biol.* 15, 178–196. doi: 10.1038/nrm3758
- Lapidot, T., Sirard, C., Vormoor, J., Murdoch, B., Hoang, T., Caceres-Cortes, J., et al. (1994). A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. *Nature* 367, 645–648. doi: 10.1038/367645a0
- Lee, J. Y., and Kong, G. (2016). Roles and epigenetic regulation of epithelial-mesenchymal transition and its transcription factors in cancer initiation and progression. *Cell. Mol. Life Sci.* 73, 4643–4660. doi: 10.1007/s00018-016-2313-z
- Lee, Y. L., Peng, Q., Fong, S. W., Chen, A. C., Lee, K. F., Ng, E. H., et al. (2012). Sirtuin 1 facilitates generation of induced pluripotent stem cells from mouse embryonic fibroblasts through the miR-34a and p53 pathways. *PLoS One* 7:e45633. doi: 10.1371/journal.pone.0045633

- Li, Q., Hutchins, A. P., Chen, Y., Li, S., Shan, Y., Liao, B., et al. (2017). A sequential EMT-MET mechanism drives the differentiation of human embryonic stem cells towards hepatocytes. *Nat. Commun.* 3:15166. doi: 10.1038/ncomms15166
- Li, R., Liang, J., Ni, S., Zhou, T., Qing, X., Li, H., et al. (2010). A mesenchymal-to-epithelial transition initiates and is required for the nuclear reprogramming of mouse fibroblasts. *Cell Stem Cell* 7, 51–63. doi: 10.1016/j.stem.2010.04.014
- Liao, B., Bao, X., Liu, L., Feng, S., Zovoilis, A., Liu, W., et al. (2011). MicroRNA cluster 302-367 enhances somatic cell reprogramming by accelerating a mesenchymal-to-epithelial transition. *J. Biol. Chem.* 286, 17359–17364. doi: 10.1074/jbc.C111.235960
- Lim, J., and Thiery, J. P. (2012). Epithelial-mesenchymal transitions: insights from development. *Development* 139, 3471–3486. doi: 10.1242/dev.071209
- Liu, X., Sun, H., Qi, J., Wang, L., He, S., Liu, J., et al. (2013). Sequential introduction of reprogramming factors reveals a time-sensitive requirement for individual factors and a sequential EMT-MET mechanism for optimal reprogramming. *Nat. Cell Biol.* 15, 829–838. doi: 10.1038/ncb2765
- Lu, C., Ward, P., Kapoor, G., Rohle, D., Turcan, S., Abdel-Wahab, O., et al. (2012). IDH mutation impairs histone demethylation and results in a block to cell differentiation. *Nature* 483, 474–478. doi: 10.1038/nature10860
- Lucena, M., Carvalho-Cruz, P., Donadio, J., Oliveira, I., de, Q. R., Marinho-Carvalho, M., et al. (2016). Epithelial mesenchymal transition induces aberrant glycosylation through hexosamine biosynthetic pathway activation. *J. Biol. Chem.* 291, 12917–12929. doi: 10.1074/jbc.M116.729236
- Luo, W., Hu, H., Chang, R., Zhong, J., Knapel, M., O'Meally, R., et al. (2011). Pyruvate kinase M2 is a PHD3-stimulated coactivator for hypoxia-inducible factor 1. *Cell* 145, 732–744. doi: 10.1016/j.cell.2011.03.054
- Macheda, M. L., Rogers, S., and Best, J. D. (2005). Molecular and cellular regulation of glucose transporter (GLUT) proteins in cancer. *J. Cell. Physiol.* 202, 654–662. doi: 10.1002/jcp.20166
- Malik, S., Villanova, L., Tanaka, S., Aonuma, M., Roy, N., Berber, E., et al. (2015). SIRT7 inactivation reverses metastatic phenotypes in epithelial and mesenchymal tumors. *Sci. Rep.* 5:9841. doi: 10.1038/srep09841
- Martin, G. R. (1981). Isolation of a pluripotent cell line from early mouse embryos cultured in medium conditioned by teratocarcinoma stem cells. *Proc. Natl. Acad. Sci. U.S.A.* 78, 7634–7638. doi: 10.1073/pnas.78.12.7634
- Mathieu, J., Zhou, W., Xing, Y., Sperber, H., Ferreccio, A., Agoston, Z., et al. (2014). Hypoxia-inducible factors have distinct and stage-specific roles during reprogramming of human cells to pluripotency. *Cell Stem Cell* 14, 592–605. doi: 10.1016/j.stem.2014.02.012
- Mathieu, J., Zhou, W., Xing, Y., Sperber, H., Ferreccio, A., Agoston, Z., et al. (2017). EMT and MET: Necessary or permissive for metastasis? *Mol. Oncol.* 11, 755–769.
- McCoy, E., Iwanaga, R., Jedlicka, P., Abbey, N., Chodosh, L., Heichman, K., et al. (2009). Six1 expands the mouse mammary epithelial stem/progenitor cell pool and induces mammary tumors that undergo epithelial-mesenchymal transition. *J. Clin. Invest.* 119, 2663–2677. doi: 10.1172/JCI37691
- Moon, J. H., Heo, J. S., Kim, J. S., Jun, E. K., Lee, J. H., Kim, A., et al. (2011). Reprogramming fibroblasts into induced pluripotent stem cells with Bmi1. *Cell Res.* 21, 1305–1315. doi: 10.1038/cr.2011.107
- Morel, A., Hinkal, G., Thomas, C., Fauvet, F., Courtois-Cox, S., Wierinckx, A., et al. (2012). EMT inducers catalyze malignant transformation of mammary epithelial cells and drive tumorigenesis towards claudin-low tumors in transgenic mice. *PLoS Genet.* 8:e1002723. doi: 10.1371/journal.pgen.1002723
- Moussaieff, A., Rouleau, M., Kitsberg, D., Cohen, M., Levy, G., Barasch, D., et al. (2015). Glycolysis-mediated changes in acetyl-CoA and histone acetylation control the early differentiation of embryonic stem cells. *Cell Metab.* 21, 392–402. doi: 10.1016/j.cmet.2015.02.002
- Mu, W. L., Wang, Y. J., Xu, P., Hao, D. L., Liu, X. Z., Wang, T. T., et al. (2015). Sox2 Deacetylation by Sirt1 Is Involved in Mouse Somatic Reprogramming. *Stem Cells* 33, 2135–2147. doi: 10.1002/stem.2012
- Najafi, M., Farhood, B., and Mortezaee, K. (2019). Cancer stem cells (CSCs) in cancer progression and therapy. *J. Cell. Physiol.* 234, 8381–8395. doi: 10.1002/jcp.27740
- Nassour, M., Idoux-Gillet, Y., Selmi, A., Come, C., Faraldo, M., Deugnier, M., et al. (2012). Slug controls stem/progenitor cell growth dynamics during mammary gland morphogenesis. *PLoS One* 7:e53498. doi: 10.1371/journal.pone.0053498
- Neri, F., Incarnato, D., Krepelova, A., Rapelli, S., Pagnani, A., Zecchina, R., et al. (2013). Genome-wide analysis identifies a functional association of Tet1 and Polycomb repressive complex 2 in mouse embryonic stem cells. *Genome Biol.* 14:R91. doi: 10.1186/gb-2013-14-8-r91
- Ng, S.-C., Locasale, J. W., Lyssiotis, C. A., Zheng, Y., Teo, R. Y., Ratanasirintraawot, S., et al. (2013). Influence of threonine metabolism on S-Adenosylmethionine and histone methylation. *Science* 339, 222–226. doi: 10.1126/science.1226603
- Ngo, B., Van, R. J., Cantley, L., and Yun, J. (2019). Targeting cancer vulnerabilities with high-dose vitamin C. *Nat. Rev. Cancer* 19, 271–282. doi: 10.1038/s41568-019-0135-7
- Nieto, M. A. (2013). Epithelial plasticity: a common theme in embryonic and cancer cells. *Science* 342:1234850. doi: 10.1126/science.1234850
- Nieto, M. A., Huang, R. Y., Jackson, R. A., and Thiery, J. P. (2016). Emt: 2016. *Cell* 166, 21–45. doi: 10.1016/j.cell.2016.06.028
- Onder, T. T., Kara, N., Cherry, A., Sinha, A. U., Zhu, N., Bernt, K. M., et al. (2012). Chromatin-modifying enzymes as modulators of reprogramming. *Nature* 483, 598–602. doi: 10.1038/nature10953
- Park, S. M., Gaur, A. B., Lengyel, E., and Peter, M. E. (2008). The miR-200 family determines the epithelial phenotype of cancer cells by targeting the E-cadherin repressors ZEB1 and ZEB2. *Genes Dev.* 22, 894–907. doi: 10.1101/gad.1640608
- Pastushenko, I., and Blanpain, C. (2019). EMT transition states during tumor progression and metastasis. *Trends Cell Biol.* 29, 212–226. doi: 10.1016/j.tcb.2018.12.001
- Pastushenko, I., Brisebarre, A., Sifrim, A., Fioramonti, M., Revenco, T., Boumahdi, S., et al. (2018). Identification of the tumour transition states occurring during EMT. *Nature* 556, 463–468. doi: 10.1038/s41586-018-0040-3
- Patra, K. C., Wang, Q., Bhaskar, P. T., Miller, L., Wang, Z., Wheaton, W., et al. (2013). Hexokinase 2 is required for tumor initiation and maintenance and its systemic deletion is therapeutic in mouse models of cancer. *Cancer Cell* 24, 213–228. doi: 10.1016/j.ccr.2013.06.014
- Pei, D., Shu, X., Gassama-Diagne, A., and Thiery, J. P. (2019). Mesenchymal-epithelial transition in development and reprogramming. *Nat. Cell Biol.* 21, 44–53. doi: 10.1038/s41556-018-0195-z
- Peinado, H., Ballestar, E., Esteller, M., and Cano, A. (2004). Snail mediates E-cadherin repression by the recruitment of the Sin3A/histone deacetylase 1 (HDAC1)/HDAC2 complex. *Mol. Cell Biol.* 24, 306–319. doi: 10.1128/mcb.24.1.306-319.2004
- Prigione, A., Rohwer, N., Hoffmann, S., Mlody, B., Drews, K., Bukowiecki, R., et al. (2014). HIF1 α modulates cell fate reprogramming through early glycolytic shift and upregulation of PDK1-3 and PKM2. *Stem Cells* 32, 364–376. doi: 10.1002/stem.1552
- Quivoron, C., Couronne, L., Della Valle, V., Lopez, C. K., Plo, I., Wagner-Ballon, O., et al. (2011). TET2 inactivation results in pleiotropic hematopoietic abnormalities in mice and is a recurrent event during human lymphomagenesis. *Cancer Cell* 20, 25–38. doi: 10.1016/j.ccr.2011.08.006
- Ryall, J. G., Cliff, T., Dalton, S., and Sartorelli, V. (2015). Metabolic Reprogramming of Stem Cell Epigenetics. *Cell Stem Cell* 17, 651–662. doi: 10.1016/j.stem.2015.11.012
- Samavarchi-Tehrani, P., Golipour, A., David, L., Sung, H. K., Beyer, T. A., Datti, A., et al. (2010). Functional genomics reveals a BMP-driven mesenchymal-to-epithelial transition in the initiation of somatic cell reprogramming. *Cell Stem Cell* 7, 64–77. doi: 10.1016/j.stem.2010.04.015
- Sampson, V., David, J., Puig, I., Patil, P., Herreros, A., Thomas, G., et al. (2014). Wilms' tumor protein induces an epithelial-mesenchymal hybrid differentiation state in clear cell renal cell carcinoma. *PLoS One* 9:e102041. doi: 10.1371/journal.pone.0102041
- Saunders, A., Huang, X., Fidalgo, M., Reimer, M. H. Jr., Faiola, F., Ding, J., et al. (2017). The SIN3A/HDAC corepressor complex functionally cooperates with NANOG to promote pluripotency. *Cell Rep* 18, 1713–1726. doi: 10.1016/j.celrep.2017.01.055
- Schliekelman, M., Taguchi, A., Zhu, J., Dai, X., Rodriguez, J., Celiktas, M., et al. (2015). Molecular portraits of epithelial, mesenchymal, and hybrid States in lung adenocarcinoma and their relevance to survival. *Cancer Res.* 75, 1789–1800. doi: 10.1158/0008-5472.CAN-14-2535
- Setty, M., Tadmor, M. D., Reich-Zeliger, S., Angel, O., Salame, T. M., Kathail, P., et al. (2016). Wishbone identifies bifurcating developmental trajectories from single-cell data. *Nat. Biotechnol.* 34, 637–645. doi: 10.1038/nbt.3569
- Shi, Y. J., Lan, F., Matson, C., Mulligan, P., Whetstone, J. R., Cole, P. A., et al. (2004). Histone demethylation mediated by the nuclear arginine oxidase homolog LSD1. *Cell* 119, 941–953. doi: 10.1016/j.cell.2004.12.012

- Shiraki, N., Shiraki, Y., Tsuyama, T., Obata, F., Miura, M., Nagae, G., et al. (2014). Methionine metabolism regulates maintenance and differentiation of human pluripotent stem cells. *Cell Metab.* 19, 780–794. doi: 10.1016/j.cmet.2014.03.017
- Shu, X., and Pei, D. (2014). The function and regulation of mesenchymal-to-epithelial transition in somatic cell reprogramming. *Curr. Opin. Genet. Dev.* 28, 32–37. doi: 10.1016/j.gde.2014.08.005
- Shyh-Chang, N., Daley, G., and Cantley, L. (2013). Stem cell metabolism in tissue development and aging. *Development* 140, 2535–2547. doi: 10.1242/dev.091777
- Siemens, H., Jackstadt, R., Hunten, S., Kaller, M., Menssen, A., Gotz, U., et al. (2011). miR-34 and SNAIL form a double-negative feedback loop to regulate epithelial-mesenchymal transitions. *Cell Cycle* 10, 4256–4271. doi: 10.4161/cc.10.24.18552
- Stefano, B. D., Sardina, J., Oevelen, C. V., Collombet, S., Kallin, E., Vicent, G., et al. (2014). C/EBP α poises B cells for rapid reprogramming into induced pluripotent stem cells. *Nature* 506, 235–239. doi: 10.1038/nature12885
- Sun, H., Liang, L., Li, Y., Feng, C., Li, L., Zhang, Y., et al. (2016). Lysine-specific histone demethylase 1 inhibition promotes reprogramming by facilitating the expression of exogenous transcriptional factors and metabolic switch. *Sci. Rep.* 6:30903. doi: 10.1038/srep30903
- Sun, H., Yang, X., Liang, L., Zhang, M., Li, Y., Chen, J., et al. (2020). Metabolic switch and epithelial-mesenchymal transition cooperate to regulate pluripotency. *EMBO J.* 39:e102961. doi: 10.15252/embj.2019102961
- Sun, N. Y., and Yang, M. H. (2020). Metabolic reprogramming and epithelial-mesenchymal plasticity: opportunities and challenges for cancer therapy. *Front. Oncol.* 10:792. doi: 10.3389/fonc.2020.00792
- Suzuki, H. I. (2018). MicroRNA control of TGF- β signaling. *Int. J. Mol. Sci.* 19:1901. doi: 10.3390/ijms19071901
- Swinnen, J. V., Brusselmans, K., and Verhoeven, G. (2006). Increased lipogenesis in cancer cells: new players, novel targets. *Curr. Opin. Clin. Nutr. Metab. Care* 9, 358–365. doi: 10.1097/01.mco.0000232894.28674.30
- Takahashi, K., and Yamanaka, S. (2006). Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 126, 663–676. doi: 10.1016/j.cell.2006.07.024
- Thankamony, A. P., Saxena, K., Murali, R., Jolly, M. K., and Nair, R. (2020). Cancer stem cell plasticity - a deadly deal. *Front. Mol. Biosci.* 7:79. doi: 10.3389/fmolb.2020.00079
- Thiery, J. P., Acloque, H., Huang, R. Y., and Nieto, M. A. (2009). Epithelial-mesenchymal transitions in development and disease. *Cell* 139, 871–890. doi: 10.1016/j.cell.2009.11.007
- Thiery, J. P., and Sleeman, J. P. (2006). Complex networks orchestrate epithelial-mesenchymal transitions. *Nat. Rev. Mol. Cell Biol.* 7, 131–142. doi: 10.1038/nrm1835
- Tsai, Y., Chen, H., Chen, S., Cheng, W., Wang, H., Shen, Z., et al. (2014). TET1 regulates hypoxia-induced epithelial-mesenchymal transition by acting as a co-activator. *Genome Biol.* 15:513. doi: 10.1186/s13059-014-0513-0
- Uckun, F. M., Sather, H., Reaman, G., Shuster, J., Land, V., Trigg, M., et al. (1995). Leukemic cell growth in SCID mice as a predictor of relapse in high-risk B-lineage acute lymphoblastic leukemia. *Blood* 85, 873–878.
- Vella, P., Scelfo, A., Jammula, S., Chiacchiera, F., Williams, K., Cuomo, A., et al. (2013). Tet proteins connect the O-linked N-acetylglucosamine transferase Ogt to chromatin in embryonic stem cells. *Mol. Cell* 49, 645–656. doi: 10.1016/j.molcel.2012.12.019
- Wang, F., Han, J., Wang, L., Jing, Y., Zhu, Z., Hui, D., et al. (2017). CCCTC-binding factor transcriptionally targets Wdr5 to mediate somatic cell reprogramming. *Stem Cells Dev.* 26, 743–750. doi: 10.1089/scd.2016.0309
- Wang, T., Chen, K., Zeng, X., Yang, J., Wu, Y., Shi, X., et al. (2011). The Histone Demethylases Jhdm1a/1b enhance somatic cell reprogramming in a vitamin-C-dependent manner. *Cell Stem Cell* 9, 575–587. doi: 10.1016/j.stem.2011.10.005
- Wang, Y., Dong, C., and Zhou, B. P. (2020). Metabolic reprogram associated with epithelial-mesenchymal transition in tumor progression and metastasis. *Genes Dis.* 7, 172–184. doi: 10.1016/j.gendis.2019.09.012
- Wang, Y., Qin, J., Wang, S., Zhang, W., Duan, J., Zhang, J., et al. (2016). Conversion of human gastric epithelial cells to multipotent endodermal progenitors using defined small molecules. *Cell Stem Cell* 19, 449–461. doi: 10.1016/j.stem.2016.06.006
- Warburg, O. (1956). On the origin of cancer cells. *Science* 123, 309–314. doi: 10.1126/science.123.3191.309
- Wei, T., Chen, W., Wang, X., Zhang, M., Chen, J., Zhu, S., et al. (2015). An HDAC2-TET1 switch at distinct chromatin regions significantly promotes the maturation of pre-iPS to iPS cells. *Nucleic Acids Res.* 43, 5409–5422. doi: 10.1093/nar/gkv430
- Wellner, U., Schubert, J., Burk, U., Schmalhofer, O., Zhu, F., Sonntag, A., et al. (2009). The EMT-activator ZEB1 promotes tumorigenicity by repressing stemness-inhibiting microRNAs. *Nat. Cell Biol.* 11, 1487–1495. doi: 10.1038/ncb1998
- Williams, E. D., Gao, D., Redfern, A., and Thompson, E. W. (2019). Controversies around epithelial-mesenchymal plasticity in cancer metastasis. *Nat. Rev. Cancer* 19, 716–732. doi: 10.1038/s41568-019-0213-x
- Wu, H., and Zhang, Y. (2014). Reversing DNA methylation: mechanisms, genomics, and biological functions. *Cell* 156, 45–68. doi: 10.1016/j.cell.2013.12.019
- Wu, J., Ocampo, A., and Belmonte, J. (2016). Cellular Metabolism and Induced Pluripotency. *Cell* 166, 1371–1385. doi: 10.1016/j.cell.2016.08.008
- Wu, X., and Zhang, Y. (2017). TET-mediated active DNA demethylation: mechanism, function and beyond. *Nat. Rev. Genet.* 18, 517–534. doi: 10.1038/nrg.2017.33
- Wu, Y., Chen, K., Xing, G., Li, L., Ma, B., Hu, Z., et al. (2019). Phospholipid remodeling is critical for stem cell pluripotency by facilitating mesenchymal-to-epithelial transition. *Sci. Adv.* 5:eaax7525. doi: 10.1126/sciadv.aax7525
- Yang, J., Antin, P., Berx, G., Blanpain, C., Brabletz, T., Bronner, M., et al. (2020). Guidelines and definitions for research on epithelial-mesenchymal transition. *Nat. Rev. Mol. Cell Biol.* 21, 341–352. doi: 10.1038/s41580-020-0237-9
- Yang, L., Zhang, F., Wang, X., Tsai, Y., Chuang, K. H., Keng, P. C., et al. (2016). A FASN-TGF- β 1-FASN regulatory loop contributes to high EMT/metastatic potential of cisplatin-resistant non-small cell lung cancer. *Oncotarget* 7, 55543–55554. doi: 10.18632/oncotarget.10837
- Ye, X., Brabletz, T., Kang, Y., Longmore, G., Nieto, M., Stanger, B., et al. (2017). Upholding a role for EMT in breast cancer metastasis. *Nature* 547, E1–E3. doi: 10.1038/nature22816
- Ye, X., Tam, W. L., Shibue, T., Kaygusuz, Y., Reinhardt, F., Ng Eaton, E., et al. (2015). Distinct EMT programs control normal mammary stem cells and tumour-initiating cells. *Nature* 525, 256–260. doi: 10.1038/nature14897
- Ye, X., and Weinberg, R. A. (2015). Epithelial-mesenchymal plasticity: a central regulator of cancer progression. *Trends Cell Biol.* 25, 675–686. doi: 10.1016/j.tcb.2015.07.012
- Yin, R., Mao, S.-Q., Zhao, B., Chong, Z., Yang, Y., Zhao, C., et al. (2013). Ascorbic acid enhances Tet-Mediated 5-methylcytosine oxidation and promotes DNA demethylation in mammals. *J. Am. Chem. Soc.* 135, 10396–10403. doi: 10.1021/ja4028346
- Yu, L., Lu, M., Jia, D., Ma, J., Ben-Jacob, E., Levine, H., et al. (2017). Modeling the genetic regulation of cancer metabolism: interplay between glycolysis and oxidative phosphorylation. *Cancer Res.* 77, 1564–1574. doi: 10.1158/0008-5472.CAN-16-2074
- Zaravinos, A. (2015). The regulatory role of MicroRNAs in EMT and cancer. *J. Oncol.* 2015:865816. doi: 10.1155/2015/865816
- Zavadil, J., and Bottinger, E. P. (2005). TGF- β and epithelial-to-mesenchymal transitions. *Oncogene* 24, 5764–5774. doi: 10.1038/sj.onc.1208927
- Zhang, H., and Wang, Z. (2008). Mechanisms that mediate stem cell self-renewal and differentiation. *J. Cell. Biochem.* 103, 709–718. doi: 10.1002/jcb.21460
- Zhang, J., Nuebel, E., Daley, G., Koehler, C., and Teitell, M. (2012). Metabolic regulation in pluripotent stem cells during reprogramming and self-renewal. *Cell Stem Cell* 11, 589–595. doi: 10.1016/j.stem.2012.10.005
- Zhang, J., Tian, X., Zhang, H., Teng, Y., Li, R., Bai, F., et al. (2014). TGF- β -induced epithelial-to-mesenchymal transition proceeds through stepwise activation of multiple feedback loops. *Sci. Signal.* 7:ra91. doi: 10.1126/scisignal.2005304
- Zhang, Y., and Weinberg, R. A. (2018). Epithelial-to-mesenchymal transition in cancer: complexity and opportunities. *Front. Med.* 12, 361–373. doi: 10.1007/s11684-018-0656-6
- Zheng, X., Carstens, J., Kim, J., Scheible, M., Kaye, J., Sugimoto, H., et al. (2015). Epithelial-to-mesenchymal transition is dispensable for metastasis but induces

- chemoresistance in pancreatic cancer. *Nature* 527, 525–530. doi: 10.1038/nature16064
- Zhou, W., Choi, M., Margineantu, D., Margaretha, L., Hesson, J., Cavanaugh, C., et al. (2012). HIF1 α induced switch from bivalent to exclusively glycolytic metabolism during ESC-to-EpiSC/hESC transition. *EMBO J.* 31, 2103–2116. doi: 10.1038/emboj.2012.71
- Zhou, Z., Yang, X., He, J., Liu, J., Wu, F., Yu, S., et al. (2017). Kdm2b regulates somatic reprogramming through variant PRC1 complex-dependent function. *Cell Rep.* 21, 2160–2170. doi: 10.1016/j.celrep.2017.10.091

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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