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Characterization and mechanism of action of amphibian-derived wound-healing-promoting peptides

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Wound-healing-promoting peptides are excellent candidates for developing wound-healing agents due to their small size and low production cost. Amphibians are one of the major sources of bioactive peptides, including wound-healing-promoting peptides. So far, a series of wound-healingpromoting peptides have been characterized from amphibians. We hereby summarized the amphibian-derived wound-healing-promoting peptides and their mechanism of action. Among these peptides, two peptides (tylotoin and TK-CATH) were characterized from salamanders, and twenty five peptides were characterized from frogs. These peptides generally have small sizes with 5-80 amino acid residues, nine peptides (tiger17, cathelicidin-NV, cathelicidin-DM, OM-LV20, brevinin-2Ta, brevinin-2PN, tylotoin, Bv8-AJ, and RL-QN15) have intramolecular disulfide bonds, seven peptides (temporin A, temporin B, esculentin-1a, tiger17, Pse-T2, DMS-PS2, FW-1, and FW-2) are amidated at the C-terminus, and the others are linear peptides without modifications. They all efficiently accelerated the healing of skin wounds or photodamage in mice or rats. They selectively promoted the proliferation and migration of keratinocytes and fibroblasts, recruited neutrophils and macrophages to wounds, and regulated the immune response of neutrophils and macrophages in wounds, which were essential for wound healing. Interestingly, MSI-1, Pse-T2, cathelicidin-DM, brevinin-2Ta, brevinin-2PN, and DMS-PS2 were just antimicrobial peptides, but they also significantly promoted the healing of infected wounds by clearing off bacteria. Considering the small size, high efficiency, and definite mechanism, amphibian-derived wound-healing-promoting peptides might be excellent candidates for developing novel wound-healing-promoting agents in future.

KEYWORDS

amphibian, frog, salamander, wound-healing-promoting peptide, antimicrobial peptide

1 Introduction

Skin, including the epidermis and dermis, can form a protective shield from external physical and chemical environmental challenges (Gallo and Hooper, 2012). As a protective barrier against a range of external stress factors, the skin is the most frequently injured one among all tissues (Martin, 1997; Liu et al., 2014a; Eming et al., 2014). Wound healing is essential for all organisms to survive, and for skin, it developed a set of complex mechanisms to protect itself and to restore tissue integrity when damaged (Mangoni et al., 2016). Efficient wound healing requires the coordinated responses of various cell types within an injured

tissue (Enyedi and Niethammer, 2015). Traditionally, acute wound healing includes four overlapping phases known as hemostasis, inflammation, cellular proliferation, and remodeling (Greaves et al., 2013). Following surgery, accidental injury, burns, microbial infection, skin diseases, or metabolic dysfunction, the skin barrier is broken and gives rise to a wound (Kujath and Kujath, 2010). Under normal physiological conditions, restoration of a functional epidermal barrier is highly efficient (Akita, 2019), but the failure of the mechanisms underlying the recovery of damaged tissues can lead to the formation of non-healing chronic wounds (Mustoe, 2004; Demidova-Rice et al., 2012). However, due to lack of therapies for treating non-healing wounds or for speeding up the repair of acute wounds, the number of patients suffering from chronic wounds and impaired healing conditions is reaching epidemic proportions and becoming more burdensome in both human health and economic terms (Sen et al., 2009). In the past decades, a few of growth factors has been developed for woundhealing therapies, including epidermal growth factor (EGF), fibroblast growth factor 2 (FGF-2), vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), keratinocyte growth factor-1 (KGF-1), granulocyte-macrophage colony-stimulating factor (GM-CSF), and granulocyte colonystimulating factor (G-CSF) (13). However, because of the large sizes corresponding to higher production costs, the therapies based on growth factors have not yet proven to be widespread in clinical application (Julier et al., 2017; He et al., 2019). Therefore, it is necessary to develop efficient drugs for curing chronic wounds and impaired healing. Recently, immunomodulatory peptides with small size and potent wound-healing-promoting activities are becoming attractive candidates for treatment of wounds (Mansour et al., 2014; Hancock et al., 2016).

Amphibians are directly exposed to different environments and their skins interact with kinds of environmental factors, such as microorganisms, parasites, predators, and physical factors (Xu and Lai, 2015). For everyday survival, amphibian skins possess excellent wound-healing ability and represent a resource for prospective wound-healing-promoting compounds (Liu et al., 2014b). So far, a series of wound-healing-promoting peptides have been characterized from amphibians. In addition to amphibian-derived wound-healing-promoting peptides, antimicrobial peptides (AMPs) are expressed diversely and abundantly in amphibian skins as the first line in defense against microorganism infection. For the phases of wound healing mentioned previously, in theory, all peptides with a positive effect on inflammation, proliferation, and remodeling may have the wound-healing-promoting ability (Xu and Lai, 2015). Considering the immunomodulatory properties of AMPs on cell migration, survival, proliferation, and induction of antimicrobial and immune mediators, the AMPs are also a huge pool of woundhealing-promoting peptides (Bernard and Gallo, 2011; Niyonsaba et al., 2017). As we know, AMP expression can be induced by infection. Interestingly, AMP expression can also be induced by injury, which indicates that AMPs may act as multifunctional effector molecules in the anti-infective and repair response by triggering cellular responses. The skins of anuran (frogs and toads) amphibians are an extraordinarily rich source of AMPs (Rinaldi, 2002; Ladram and Nicolas, 2016). When the amphibian is stressed or injured, the peptides stored in skin granular glands can be released in high concentrations into skin secretions, including AMP and wound-healing peptides. In this brief review, we summed up the peptides with the wound-healing-promoting effect derived from salamanders and frogs, many of which can also be classified into AMPs. It suggests that AMPs from frogs can be used as a reserve for screening wound-healing-promoting peptides.

2 Wound-healing-promoting peptides from amphibians

2.1 AH90

AH90 is a peptide composed of 24 amino acid residues characterized from the frog Odorrana grahami (Liu et al., 2014a). It was shown to promote wound healing by inducing cell migration, granulation tissue contraction, and fast re-epithelialization process. All the promoting processes induced by AH90 were attributed to the induction of transforming growth factor $\beta 1$ (TGF- $\beta 1$) production in macrophages, and AH90 has a strong capacity to stimulate TGF-B1 secretion. TGF- β is a key factor involved in every phase of wound healing. TGF- β was shown to stimulate keratinocyte migration, angiogenesis, fibroblast proliferation, mvofibroblast differentiation, and matrix deposition (Werner and Grose, 2003). So, AH90 has a strong capacity to promote wound healing by stimulating TGF-β1 production in macrophages.

2.2 CW49

CW49 is a peptide composed of 11 amino acid residues characterized from the frog Odorrana grahami with potential wound-healing ability (Liu et al., 2014b). It was primarily identified by running an angiogenesis screening from a number of small peptides that might have various biological activities. The peptide CW49 showed the most outstanding pro-angiogenic ability among all of the tested peptides (Liu et al., 2014b). Angiogenesis plays a critical role in normal wound healing. However, impaired angiogenesis is often accompanied with some impaired wound healings, such as in diabetic wounds. In addition, many drugs effective in treating the chronic wounds of animal models possess the potent of pro-angiogenic ability, such as VEGF and LL-37 (Koczulla et al., 2003; Galiano et al., 2004; Carretero et al., 2008). CW49 was shown to enhance wound healing in normal and diabetic mice by promoting angiogenesis (Liu et al., 2014b). In addition, preventing an excessive inflammatory response was another underlying mechanism of CW49 in promoting the diabetic wound-healing process.

2.3 Temporins A and B

Temporin A (Ta) and temporin B (Tb) are short, linear, and mildly cationic AMPs with 13 amino acid residues identified from the frog *Rana temporaria* (Di Grazia et al., 2014). Both peptides efficiently induced HaCaT cell migration by promoting HaCaT cell proliferation via the EGFR signaling pathway. In addition, Ta was found to chemoattract monocytes by using formyl peptide receptorlike 1 as a receptor (Chen et al., 2004). In addition, their antimicrobial efficacy also played an important role in the infection stage of wound healing. In particular, they could kill *Staphylococcus aureus* and multi-drug-resistant *S. aureus* (MRSA) strains, which could be internalized by human epidermal cells. Furthermore, their small sizes should allow a low production cost, which provides the possibility for their wide application.

2.4 Esculentin-1a (1-21) NH₂

Esculentin-1a (1-21) NH₂ is a derivative of the AMP esculentin-1a, which was isolated from the skin of Pelophylax lessonae/ridibundus (formerly known as Rana esculenta) and possessed a wide spectrum of antimicrobial activities with high efficacy against Pseudomonas aeruginosa (Di Grazia et al., 2015). The authors showed that all-L but not all-D Esc(1-21) peptide significantly stimulated keratinocyte (HaCaT cells) migration, rather than keratinocyte proliferation to promote re-epithelialization of wound healing (Di Grazia et al., 2015). In addition, the research indicated that Esc (1-21)-promoted HaCaT cell migration required EGFR activation and involved STAT3 phosphorylation, which is similar to LL-37 (Tokumaru et al., 2005). However, all-L Esc(1-21) was more efficient than LL-37 in promoting migration of HaCaT cells in vitro and exhibits a wider "therapeutic" window. As described in previous research, EGFR-activated cell migration is predominantly mediated by the STAT pathway in keratinocytes, and STAT3 signaling played an essential role in skin remodeling and wound healing (Sano et al., 1999; Andl et al., 2004). Furthermore, Esc(1-21) preserved the capability of antibacterial activity even at high salt concentrations (Luca et al., 2013) as well as in the presence of serum and tears (Kolar et al., 2015). This significant property was absent in the key human AMPs, such as hBD-2 and LL-37, the antimicrobial efficacy of which was completely lost at the high ionic strength (Bals et al., 1998) or drastically reduced in biological fluids (Huang et al., 2007). The antimicrobial ability of Esc(1-21) can kill microbes, without harming mammalian cells (Luca et al., 2013; Kolar et al., 2015), which makes Esc(1-21) a particularly attractive candidate for wound-healing promoter, especially in the management of chronic, often Pseudomonas super-infected, human skin ulcers (Wolcott and Dowd, 2008; Percival et al., 2012).

2.5 Tiger17

Tiger17 was synthesized based on tigerinins, which was identified from the skin secretions of frog *Fejervarya cancrivora* (Tang et al., 2014). Tiger17 significantly promoted the proliferation of keratinocytes (HaCaT cells) and human skin fibroblasts (HSFs) in a concentration-dependent manner. An *in vitro* cell scratch assay showed that tiger17 significantly increased the migration rate of keratinocytes. A full-thickness skin wound-healing model confirmed that tiger17 accelerated the healing of full-thickness wounds in mice. The histological analysis revealed that tiger17 promoted regeneration of neo-epidermal tissues and restoration of dermal tissues in the wound of mice. In addition, immunohistochemical analysis indicated that many macrophages

were recruited to the unwounded skin margin closed to the neoepidermis and neo-dermis in tiger17-treated skin wound. In addition, the productions of TGF- β 1 and interleukin-6 (IL-6) were significantly increased by tiger17, and the induced expression of TGF- β 1 significantly promoted fibroblast-tomyofibroblast transition, which played important roles in wound healing (Haroon et al., 2000; Tredget et al., 2005; Luckett-Chastain and Gallucci, 2009).

2.6 MSI-1

MSI-1 was truncated from the N-terminus of pexiganan (Ma et al., 2020). Pexiganan is a synthetic analog of magainin 2 isolated from Xenopus laevis, and pexiganan was primarily engineered with high potency against many bacteria (Lamb and Wiseman, 1998; Ge et al., 1999; Ge et al., 2002; Lipsky et al., 2008; Wong et al., 2009; Ma et al., 2020). In vitro, pexiganan had an antimicrobial activity against most clinical bacterial isolates cultured from infected diabetic foot ulcers (Fuchs et al., 1998; Ge et al., 1999; Ge et al., 2002). In the study, the application of pexiganan in clinical trials showed that, for mildly infected diabetic foot ulcers, topical pexiganan was clinically comparable to an oral antibiotic (Lipsky et al., 2008). However, for its erythrolysis and poor biological stability, it was not more effective than already approved treatments for diabetic foot ulcers (Lavery et al., 2007). Therefore, a series of peptides were designed by truncating N-terminus of pexiganan (Ma et al., 2020). Among the designed peptides, MSI-1 exerted rapid bactericidal activity and low hemolysis.

Interestingly, MSI-1 efficiently improved the survival rate and wound closure in penicillin-resistant *Escherichia coli*infected mice by eliminating bacterial counts in mouse organs or subeschar (Ma et al., 2020). In addition, MSI-1 strongly reduced the expression of tumor necrosis factor- α (TNF- α) and IL-6 in skin lesion homogenate but not in serum. Moreover, hematoxylin and eosin (H&E) staining showed that the granulation tissues were much thicker in the MSI-1 (10 mg/kg) group, indicating that the congestion and inflammatory cell infiltration could also be attenuated by the peptide (Ma et al., 2020). So, MSI-1 significantly improved wound closure in the *E. coli*-infected burn-wound model by decreasing bacterial counts and inhibiting the bacterium from spreading systemically to relieve the systemic inflammatory response induced by local infection (Ma et al., 2020).

2.7 Cathelicidin-OA1

Cathelicidin-OA1 was identified from the skin of *Odorrana andersonii*, which was a member of cathelicidin AMPs (Cao et al., 2018). Unlike other cathelicidins, cathelicidin-OA1 had no direct antimicrobial activity, acute toxicity, and hemolytic activity, but it did exhibit antioxidant activity. Interestingly, cathelicidin-OA1 significantly accelerated wound healing by inducing HaCaT cell proliferation and HSF cell migration as well as accelerating reepithelialization and granulation tissue formation by enhancing the recruitment of macrophages to the wound site.

2.8 Cathelicidin-NV and antioxidin-NV

Another cathelicidin family peptide, named cathelicidin-NV, was identified from the frog skin of Nanorana ventripunctata (Wu et al., 2018). Cathelicidin-NV significantly promoted wound healing in a fullthickness dermal wound model in mice. The in vitro assay showed that cathelicidin-NV accelerated re-epithelialization by enhancing the proliferation of keratinocytes and fibroblasts and migration of keratinocytes. Cathelicidin-NV also induced the differentiation of fibroblasts to myofibroblasts and collagen production in fibroblasts, which played important roles in wound contraction and repair processes. Furthermore, all the aforementioned wound-healing processes including migration, proliferation, and differentiation were in need of some factors to drive. In this study, the release of essential factors such as monocyte chemoattractant protein-1 (MCP-1), TNF-a, VEGF, and TGF-\u03c61 were proved to be induced by cathelicidin-NV both in vivo and in vitro. In addition, cathelicidin-NV showed no cytotoxicity and hemolytic activity toward mammalian cells, and cathelicidin-NV did not have any antimicrobial activities against the tested bacterial strains at a relatively high concentration.

Antioxidin-NV was another peptide with wound-healingpromoting activity isolated from the skin from *N. ventripunctata* (Feng et al., 2021). Antioxidin-NV could rapidly clear reactive oxygen species induced by UVB irradiation, attenuate DNA damage by inhibiting p-histone H2A.X expression, reduce cell apoptosis via inhibiting the expression of cleaved caspase 3, decrease inflammatory response by inhibiting Toll-like receptor 4 (TLR4)/p38/JNK/ NF- κ B signaling pathway, and finally, provide protection against skin photoaging induced by UVB irradiation.

2.9 Cathelicidin-DM

Cathelicidin-DM was also a cathelicidin family peptide, which was isolated from the skin of *Duttaphrynus melanostictus* (Shi et al., 2020). *In vitro*, it efficiently promoted HaCaT, HSF, and HUVEC proliferation and induced HSF, HUVEC, and RAW.264.7 cell migration. In addition, cathelicidin-DM significantly activated the MAPK signaling pathway, including inducing the phosphorylation of ERK, JNK, and p38. *In vivo*, it accelerated the healing of non-infected skin wounds in mice by enhancing re-epithelialization, granulation tissue formation, α -SMA expression, and collagen I deposition. Due to its powerful antimicrobial activity, cathelicidin-DM also effectively accelerated the healing of infected skin wounds in mice (Shi et al., 2020; Wang et al., 2022).

2.10 OM-LV20

OM-LV20 was identified from the skin secretion of odorous frog, *Odorrana margaretae* (Li et al., 2018). OM-LV20 accelerated the healing of in a full-thickness dermal wound model in mice. An *in vitro* experiment showed that OM-LV20 promoted wound healing by inducing keratinocytes (HaCaT) and HSF migration. Additionally, it induced the proliferation of HaCaT but not HSF cells. In addition, OM-LV20 showed no direct antimicrobial activity, hemolytic activity, or acute toxicity.

2.11 OA-GL21 and OA-GL17

OA-GL21 was isolated from the skin secretion of Odorrana andersonii (Bian et al., 2018). OA-GL21 significantly promoted wound healing of human keratinocytes (HaCaT) and human fibroblasts in a dose- and time-dependent manner. However, OA-GL21 had no significant effect on the proliferation of these two cell lines. OA-GL21 obviously promoted wound healing in the full-thickness skin wound model in mice in dose-dependent and scar-free manners. Further studies showed that OA-GL21 had no direct antibacterial, hemolytic, and acute toxic activity, and it had weak antioxidant activity but high stability. OA-GL17 was another wound-healing-promoting peptide isolated from O. andersonii (Zhang et al., 2022). It effectively enhanced the proliferation and migration of keratinocytes and accelerated the repair of skin full-thickness wounds and scald wounds in mice. Furthermore, it upregulated the expression of transforming growth factor-\u00c61 and activated the TGF-\u00b61/Smad signaling pathway by decreasing the level of miR-663a.

2.12 Brevinin-2Ta

Brevinin-2Ta (B-2Ta) was isolated from the skin secretion of the European frog, Pelophylax kl. esculentus with wound-healingpromoting activity (Liu et al., 2017). B-2Ta was also a member of AMPs. The antimicrobial and hemolytic assay revealed that B-2Ta showed broad antimicrobial activities against S. aureus, E. coli, and Candida albicans with low cytotoxicity to erythrocytes. B-2Ta effectively promoted wound closure and restrained the bacterial infection in Klebsiella pneumoniae-infected wounds in SD rats. The histological study showed that B-2Ta improved skin reepithelization, granulation, and collagen deposition in the wound area and enhanced epithelial migration and angiogenesis. On the cellular level, reduction in IL-10 demonstrated that B-2Ta could relieve inflammation by inhibiting the bacterial infection. These results indicated that B-2Ta reduced the continuous phase of inflammation and accelerated wound healing in K. pneumoniainfected rats via modulating re-epithelialization.

2.13 Brevinin-2PN

Brevinin-2PN is an antimicrobial peptide identified from *Pelophylax nigromaculatus* and showed potent antibacterial activity by destroying the bacterial membrane and targeting bacterial genomic DNA (Fan et al., 2022). In addition, brevinin-2PN also showed wound-healing-promoting activity. It effectively promoted the healing of HSF cell scratches by inducing cell migration and regulating the expression of growth factors.

2.14 Pse-T2

Pse-T2 is a truncated analog of pseudin-2, which was primarily isolated from the skin of the frog *Pseudis paradoxa* (Kang et al., 2018). Pseudin-2 is a 24-amino acid AMP that exhibits potent antibacterial activity against Gram-negative bacteria, but its cytotoxicity limited the application in clinic. As mentioned

previously, it is more likely to modify a known peptide to enhance activity and reduce toxicity. The authors next assayed the activities of 14 modified peptides based on pseudin-2 (Kang et al., 2018). The data indicated that pseudin-2-truncated analogs (particularly Pse-T2) were the most effective antimicrobial agents against pathogenic bacteria in the modified peptides and exhibited little hemolytic activity. The *in vivo* assay showed that Pse-T2 also enhanced *P. aeruginosa*-infected wound closure via its antibacterial and antibiofilm activities.

2.15 Tylotoin

Tylotoin was identified from the skin of salamander *Tylototriton verrucosus* with potential wound-healing-promoting activity (Mu et al., 2014). Tylotoin was a member of the cathelicidin antimicrobial peptide family, but it had no antimicrobial activities against tested bacteria. Interestingly, it showed comparable wound-healing-promoting ability with EGF in a murine model of full-thickness dermal wounds. Further research revealed that tylotoin directly enhanced the motility and proliferation of keratinocytes, proliferation of vascular endothelial cells and their tube formation, and proliferation of fibroblasts. Tylotoin significantly accelerated re-epithelialization and granulation tissue formation in the wound site. Tylotoin also promoted the release of TGF- β 1 and IL-6, which were essential for wound healing.

2.16 TK-CATH

TK-CATH was identified from the salamander skin *Tylototriton kweichowensis*, which belonged to the cathelicidin antimicrobial peptide family (Luo et al., 2021). Intriguingly, TK-CATH is an anionic cathelicidin antimicrobial peptide that lacks direct antimicrobial activity. However, it effectively accelerated the healing of full-thickness skin wound in mice. It was demonstrated that TK-CATH elicited the production of cytokines, chemokines, and growth factors in that promoted cell migration, cell proliferation, and wound healing.

2.17 Ot-WHP

In our previous work, we also isolated a wound-healingpromoting peptide, designated as Ot-WHP, from Chinese concave-eared frog *Odorrana tormota* (He et al., 2019). In our study, Ot-WHP efficiently promoted wound healing in a murine model of full-thickness wounds. It significantly increased the number of neutrophils and macrophages in wound sites. However, our results showed that Ot-WHP did not act as a chemoattractant for neutrophils and macrophages. Then, we found that Ot-WHP significantly induced chemokine, cytokine, and growth factor production in macrophages by activating mitogen-activated protein kinases (MAPKs) and nuclear factorkB (NF-kB) signaling pathways. It suggested that the chemotactic activity of Ot-WHP depended on inducing chemoattractant production in macrophages. In addition, Ot-WHP directly promoted keratinocyte migration by enhancing integrin expression and cell adhesion. Ot-WHP promoted the proliferation of keratinocytes and fibroblasts, collagen deposition, and α -SMA expression in fibroblasts via activating macrophages, which suggested that Ot-WHP significantly enhanced the crosstalk between macrophages and keratinocytes/fibroblasts, which, in turn, efficiently promoted wound closure in mice. Furthermore, Ot-WHP modestly promoted neutrophil phagocytosis and phorbol myristate acetate (PMA)-induced neutrophil extracellular trap formation, which were extremely critical for clearing off cell debris and microbes in wounds.

2.18 Bv8-AJ

Bv8-AJ was isolated from the skin secretion of *Amolops jingdongensis* and was identified as a homologous of mammalian prokineticins (Chang et al., 2019). Bv8-AJ significantly accelerated wound healing of full-thickness wounds in mice models. The histopathological study indicated that Bv8-AJ accelerated the initiation and cessation of the inflammatory phase of wounds in mice models. Moreover, Bv8-AJ exerted a strong proliferative effect on fibroblasts and keratinocytes isolated from newborn mice by activating the interleukin-1 signaling pathway.

2.19 DMS-PS2

DMS-PS2 was identified from the skin of the waxy monkey tree frog, *Phyllomedusa sauvagei* (Song et al., 2020). It was demonstrated to be a typical cationic antimicrobial peptide belonging to the dermaseptin family and have potent antibacterial activity against Gram-positive bacteria, Gram-negative bacteria, and fungi. DMS-PS2 also showed an inhibitory effect against Gram-positive and Gram-negative bacterial biofilms. Remarkably, DMS-PS2 effectively promoted the healing of methicillinresistant *Staphylococcus aureus* (MRSA)-infected wounds in murine skin.

2.20 RL-10 and RL-QN15

RL-10 was identified from the skin of *Rana limnocharis* (Wang et al., 2021a). It significantly enhanced the migration and proliferation of HaCaT cells *in vitro*. RL-10 also effectively promoted the healing of a full-thickness wound in mice. RL-QN15 was another wound-healing-promoted peptide identified from the skin of *Rana limnocharis* (Wang et al., 2021b) and provided protection against chronic wounds, skin fibrosis, and oral ulcers. The mechanistic study revealed that RL-QN15 obviously induced the activation of MAPK and Smad signaling and selectively regulated the production of cytokines in macrophages.

2.21 FW-1 and FW-2

FW-1 and FW-2 were isolated from the skin of *Hyla annectans*, which were cleaved from one protein precursor containing five copies of FW-1 and four copies of FW-2 (Liu et al., 2021). FW-1 and FW-2 provided protection against ultraviolet B (UVB)-induced skin inflammation in mice. *In vitro*, they significantly inhibited inflammatory responses in UVB-irradiated keratinocytes, modulated

TABLE 1 Information	of amphibian-derived	wound-healing-promoting peptides.

Peptide	Amino acid sequence	Molecular weight (Da)	Modification	AMP	Source
AH90	ATAWDFGPHGLLPIRPIRIRPLCG	2657.2	1	Yes	Odorrana grahami
CW49	APFRMGICTTN	1210.43	1	No	Odorrana grahami
Temporins A and B	Ta: FLPLIGRVLSGIL-NH ₂	1397.77	N-terminal amidation	Yes	Rana temporaria
	Tb: LLPIVGNLLKSLL-NH ₂	1392.79			
Esculentin1a(1-21)NH ₂	GIFSKLAGKKIKNLLISGLKG-NH ₂	2184.72	One N-terminal amidation	Yes	Rana esculenta
Tiger17	WCKPKPRCH-NH ₂	1376.79	N-terminal amidation, one intramolecular disulfide bond	No	Fejervarya cancrivora
MSI-1	GIWKFLKKAKKFWK	1807.36	1	Yes	Xenopus laevis
Cathelicidin-OA1	IGRDPTWSHLAASCLKCIFDDLPKTHN	3038.5	1	Yes	Odorrana andersonii
Cathelicidin-NV	ARGKKECKDDRCRLLMKRGSFSYV	2845.9	One intramolecular disulfide bond	Yes	Nanorana ventripunctata
Antioxidin-NV	GWANTLKNVAGGLCKMTGAA	1963.30	1		Nanorana ventripunctata
Cathelicidin-DM	SSRRKPCKGWLCKLKLRGGYTLIGSATNLNRPTYVRA	4165.9	Intramolecular disulfide bond	Yes	Duttaphrynus melanostictus
OM-LV20	LVGKLLKGAVGDVCGLLPIC	1966.39	One intramolecular disulfide bond	No	Odorrana margaretae
OA-GL21	GLLSGHYGRVVSTQSGHYGRG	2188.0	1	No	Odorrana andersonii
OA-GL17	GLFKWHPRCGEEQSMWT	2092.38	1	No	Odorrana andersonii
Brevinin-2Ta	GILDTLKNLAKTAGKGILKSLVNTASCKLSGQC	3347.01	One intramolecular disulfide bond	Yes	Pelophylax kl. esculentus
Brevinin-2PN	GLMDSLKGLAATAGKTVLQGLLKTASCKLEKTC	3349.04	One intramolecular disulfide bond	Yes	Pelophylax nigromaculatus
Pse-T2	LNALKKVFQKIHEAIKLI-NH ₂	2106.6	N-terminal amidation	Yes	Pseudis paradoxa
Tylotoin	KCVRQNNKRVCK	1473.79	One intramolecular disulfide bond	No	Tylototriton verrucosus
ТК-САТН	GGQDTGKEGETGKKKKSDNWFMNLLNKFLELIGLKEAGDD SEPFCFTCIFDMFSQ	6196.99	/	No	Tylototriton kweichowensis
Ot-WHP	ATAWDLGPHGIRPLRPIRIRPLCG	2666.12	1	No	Odorrana tormota

(Continued on following page)

IABLE I (CONTINUED) IN	I ABLE 1 (<i>Continued</i>) imormation of ampinipian-derived wound-nealing-promoting peptides.				
Peptide	Amino acid sequence	Molecular weight (Da)	Modification	AMP	AMP Source
Bv8-AJ	AVITGACERDVQCGGGTCCAVSLWMQGLRICTPLGRQGENCHPASRKVPFAGLRLHNSCPCQSNLACKTLPNGKYQCMPS 8485.9	8485.9	Five intramolecular disulfide bonds	No	Amolops jingdongensis
DMS-PS2	$ALWKFLLKNVGKAAGKAVLNAVTDMVNQ-NH_2$	2953.52	N-terminal amidation	Yes	Phyllomedusa sauvagei
RL-10	RLFKCWKKDS	1310.58	/	No	Rana limnocharis
RL-QN15	QNSYADLWCQFHYMC	1906.71	One intramolecular disulfide bond	No	Rana limnocharis
FW-1	FWPLI-NH ₂	673.8	N-terminal amidation	No	Hyla annectans
FW-2	FWPMI-NH ₂	691.9	N-terminal amidation	No	Hyla annectans
PM-7	FLNWRRILFLKVVR	1860.32	1	No	Polypedates megacephalus
/, no modification.					

1 (Continued) Information of amphibian-derived wound-healing-promoting peptides

the MAPK and NF- κ B signaling pathways activated by UVB, and reduced the production of reactive oxygen species (ROS) induced by UVB.

2.22 PM-7

PM-7 was a wound-healing-promoting peptide isolated from the skin of *Polypedates megacephalus* (Fu et al., 2022). It efficiently promoted the healing of full-thickness skin wounds in mice and enhanced the proliferation and migration of HSF and HUVECs. In addition, it could induce the activation of MAPK signaling such as ERK and p38 in HUVECs.

3 Discussion and conclusion

We herein summarized amphibian-derived wound-healingpromoting peptides, which were mainly identified from anuran (frogs and toads) amphibians (Table 1). We also compared the mechanisms of these peptides, such as effector cells and signaling pathways (Table 2). Among these peptides, most of them could induce cell proliferation, cell migration, and the production of growth factors related to wound healing. However, the research focus of each peptide was different, and not every amphibian-derived wound-healing-promoting peptide has exactly uncovered its mechanism of action. Taking MSI-1 for example, it improved wound closure in the E. coli-infected burn-wound model mainly by decreasing bacterial counts and relieving the systemic inflammatory response induced by local infection. In addition, the enhanced effects of Pse-T2 and brevinin-2Ta on bacteriainfected wound closure are attributed to their antibacterial activities, which are similar to the MSI-1 peptide. Although the antimicrobial activities of other peptides were not reflected in the process of wound healing, the antimicrobial properties of these peptides play important roles in the process of wound healing. Furthermore, microbial infection is usually associated with wound, and infection is a major factor that hinders wound healing (Demidova-Rice et al., 2012; Mangoni et al., 2016). Microbial infection is one of the most common factors that result in chronic wounds (Mustoe, 2004). Opportunistic pathogens, such as the Gram-negative bacteria P. aeruginosa or the Gram-positive bacteria S. aureus, are able to colonize skin wounds and form biofilms (Mancl et al., 2013; Vyas and Wong, 2016). It is difficult for the host to eradicate microbial infection when microbial cells aggregate in the wound and immobilize in an adhesive matrix of extracellular polymeric substances. The microbial biofilm usually results in the weak penetration of antibiotics or host clearance (Mustoe, 2004; Wolcott et al., 2010; Taylor et al., 2014). Unlike traditional antibiotics, AMPs can efficiently control microbial proliferation and modulate host's immune response to a variety of biological or physical insults, and these properties make AMP good candidates for dealing with the challenge of infectious wounds. As mentioned previously, MSI-1, Pse-T2, and brevinin-2Ta promoted wound healing by inhibiting bacterial infection. Among the listed wound-healing-promoting peptides

TABLE 2 Mechanism of amphibian-derived wound-healing-promoting peptides.

Peptide	Effector cell type						Signaling	
	Immunocyte		Keratinocyte		Fibroblast			pathway
	Recruitment	Cytokine	Proliferation	Migration	Proliferation	Migration	Differentiation	
AH90	1	+	-	+	1	1	+	MAPK, NF-кB, Smad
CW49	+	+	1	/	1	1	1	HIF-1, eNOS and iNOS
Temporins A and B	+	1	+	+	1	1	1	1
Esculentin1a(1-21)NH ₂	1	1	-	+	1	1	1	STAT3
Tiger17	+	+	+	+	+	1	+	MAPK, Smad
MSI-1	1	+	1	+	1	1	1	1
Cathelicidin-OA1	+	+	+	-	-	+	1	1
Cathelicidin-NV	1	+	+	+	+	1	+	МАРК
Antioxidin-NV	1	+	1	/	1	1	+	TLR4/p38/JNK/ NF-κB, TGF-β1/ Smad2
Cathelicidin-DM	+	1	+	+	+	+	+	MAPKs
OM-LV20	1	1	+	+	-	+	/	1
OA-GL21	1	1	-	+	-	+	1	1
OA-GL17	1	1	+	+	1	1	1	miR-663a/TGF- β1/Smad axis
Brevinin-2Ta	1	+	1	1	1	/	1	1
Brevinin-2PN	1	1	1	1	1	+	1	1
Pse-T2	1	+	1	1	1	1	1	1
Tylotoin	+	+	+	+	+	1	+	MAPK, Smad
TK-CATH	1	+	+	+	1	1	1	МАРК
Ot-WHP	+	+	+	+	+	1	+	MAPKs, NF-κB, Smad
Bv8-AJ	1	+	+	1	+	1	1	IL-1/MAPK
DMS-PS2	1	1	1	1	1	/	/	/
RL-10	1	1	+	+	1	1	1	/
RL-QN15	1	+	+	+	1	1	/	MAPK, Smad
FW-1/FW-2	1	+	1	1	1	1	1	MAPK, NF-κB
PM-7	1	1	1	1	+	+	1	MAPK

/, no mentioned; -, had no effect; +, had positive effect.

in Table 1, seven peptides belonged to AMPs or their derivatives. The dual antimicrobial and immunomodulatory activities of AMPs in infectious wound healing make them particularly attractive candidates, possibly superior to conventional antibiotics, for the local treatment of infected skin wounds (Do et al., 2014).

The molecular weights of all the amphibian-derived woundhealing-promoting peptides are less than 3,500 Da (Table 1). Small molecular weight corresponds to low production cost, which makes the peptides possible to be widely applied in clinic. Meanwhile, the small molecular weight of peptide is more suitable for the preparation of emulsion ointment for the local treatment of skin wounds. As a topical agent, high local concentrations in the superficial soft tissues could be achieved easily, surpassing the concentrations that could be reached systemically (Mangoni et al., 2016). Among these peptides, we noticed that many of them were not natural peptides, but were modified by amidating the C-terminus known peptides or truncated from known peptides (Table 1). As we know, if the synthetic peptide is the internal sequence of one protein, the charge of peptide can be removed

by N-terminal acetylation or C-terminal amidation, which will make it easy to maintain the natural structure and enhance the resistance of peptide to external peptidase. In addition, disulfide bonds between two cysteine residues can make peptide cyclization, which is usually an important way to improve the stability of one peptide. In addition, MSI-1 and Pse-T2 were truncated peptides of pexiganan and pseudin-2, respectively, which exhibited better activities and less toxicity. These provide efficient strategies to improve the activity and reduce the toxicity of known woundhealing-promoting peptides.

A series of wound-healing-promoting peptides have been characterized from amphibians, but these peptides had different research foci. Overall, the effects of most amphibian-derived wound-healing-promoting peptides on epithelial cells and immune cells were studied. In addition, some of them were studied on one special focus, such as CW49. The investigation of CW49 mainly focused on its pro-angiogenesisis effect in the wound-healing process and the related signaling pathways. Although different peptides play different roles in wound healing, these amphibian-derived peptides, in general, exhibited high efficiency in promoting wound healing in animal models. Considering that frog and mammalian skin share the same evolutionary ancestry, basic architecture, and conserved mechanisms underlying wound healing (Yoshizato, 2007), it is interesting to find more wound-healing-promoting peptides from amphibians, and the amphibian-derived woundhealing-promoting peptides might also be capable of promoting human skin wound healing and being selected as excellent templates for developing novel wound-healing-promoting agents in future.

References

Akita, S. (2019). Wound repair and regeneration: Mechanisms, signaling. Int. J. Mol. Sci. 20 (24), 6328. doi:10.3390/ijms20246328

Andl, C. D., Mizushima, T., Oyama, K., Bowser, M., Nakagawa, H., and Rustgi, A. K. (2004). EGFR-induced cell migration is mediated predominantly by the JAK-STAT pathway in primary esophageal keratinocytes. *Am. J. Physiol. Gastrointest. Liver Physiol.* 287 (6), G1227–G1237. doi:10.1152/ajpgi.00253.2004

Bals, R., Wang, X., Zasloff, M., and Wilson, J. M. (1998). The peptide antibiotic LL-37/ hCAP-18 is expressed in epithelia of the human lung where it has broad antimicrobial activity at the airway surface. *Proc. Natl. Acad. Sci. U. S. A.* 95 (16), 9541–9546. doi:10. 1073/pnas.95.16.9541

Bernard, J. J., and Gallo, R. L. (2011). Protecting the boundary: The sentinel role of host defense peptides in the skin. *Cell Mol. Life Sci.* 68 (13), 2189–2199. doi:10.1007/s00018-011-0712-8

Bian, W., Meng, B., Li, X., Wang, S., Cao, X., Liu, N., et al. (2018). OA-GL21, a novel bioactive peptide from Odorrana andersonii, accelerated the healing of skin wounds. *Biosci. Rep.* 38 (3). doi:10.1042/BSR20180215

Cao, X., Wang, Y., Wu, C., Li, X., Fu, Z., Yang, M., et al. (2018). Cathelicidin-OA1, a novel antioxidant peptide identified from an amphibian, accelerates skin wound healing. *Sci. Rep.* 8 (1), 943. doi:10.1038/s41598-018-19486-9

Carretero, M., Escamez, M. J., Garcia, M., Duarte, B., Holguin, A., Retamosa, L., et al. (2008). *In vitro* and *in vivo* wound healing-promoting activities of human cathelicidin LL-37. *J. Invest. Dermatol* 128 (1), 223–236. doi:10.1038/sj.jid.5701043

Chang, J., He, X., Hu, J., Kamau, P. M., Lai, R., Rao, D., et al. (2019). Bv8-Like toxin from the frog venom of Amolops jingdongensis promotes wound healing via the interleukin-1 signaling pathway. *Toxins (Basel)* 12 (1), 15. doi:10.3390/toxins12010015

Chen, Q., Wade, D., Kurosaka, K., Wang, Z. Y., Oppenheim, J. J., and Yang, D. (2004). Temporin A and related frog antimicrobial peptides use formyl peptide receptor-like 1 as a receptor to chemoattract phagocytes. *J. Immunol.* 173 (4), 2652–2659. doi:10. 4049/jimmunol.173.4.2652

Demidova-Rice, T. N., Hamblin, M. R., and Herman, I. M. (2012). Acute and impaired wound healing: Pathophysiology and current methods for drug delivery,

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Conflict of interest

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part 2: Role of growth factors in normal and pathological wound healing: Therapeutic potential and methods of delivery. *Adv. Skin. Wound Care* 25 (8), 349–370. doi:10.1097/01.ASW.0000418541.31366.a3

Di Grazia, A., Cappiello, F., Imanishi, A., Mastrofrancesco, A., Picardo, M., Paus, R., et al. (2015). The frog skin-derived antimicrobial peptide esculentin-1a(1-21) NH2 promotes the migration of human HaCaT keratinocytes in an EGF receptor-dependent manner: A novel promoter of human skin wound healing? *PLoS One* 10 (6), e0128663. doi:10.1371/journal.pone.0128663

Di Grazia, A., Luca, V., Segev-Zarko, L. A., Shai, Y., and Mangoni, M. L. (2014). Temporins A and B stimulate migration of HaCaT keratinocytes and kill intracellular *Staphylococcus aureus*. *Antimicrob. Agents Chemother*. 58 (5), 2520–2527. doi:10.1128/AAC.02801-13

Do, N., Weindl, G., Grohmann, L., Salwiczek, M., Koksch, B., Korting, H. C., et al. (2014). Cationic membrane-active peptides - anticancer and antifungal activity as well as penetration into human skin. *Exp. Dermatol* 23 (5), 326–331. doi:10.1111/exd.12384

Eming, S. A., Martin, P., and Tomic-Canic, M. (2014). Wound repair and regeneration: Mechanisms, signaling, and translation. *Sci. Transl. Med.* 6 (265), 265sr6. doi:10.1126/scitranslmed.3009337

Enyedi, B., and Niethammer, P. (2015). Mechanisms of epithelial wound detection. Trends Cell Biol. 25 (7), 398-407. doi:10.1016/j.tcb.2015.02.007

Fan, X. L., Yu, S. S., Zhao, J. L., Li, Y., Zhan, D. J., Xu, F., et al. (2022). Brevinin-2PN, an antimicrobial peptide identified from dark-spotted frog (Pelophylax nigromaculatus), exhibits wound-healing activity. *Dev. Comp. Immunol.* 137, 104519. doi:10.1016/j.dci.2022.104519

Feng, G., Wei, L., Che, H., Shen, Y., Yang, J., Mi, K., et al. (2021). A frog peptide ameliorates skin photoaging through scavenging reactive oxygen species. *Front. Pharmacol.* 12, 761011. doi:10.3389/fphar.2021.761011

Fu, S., Du, C., Zhang, Q., Liu, J., Zhang, X., and Deng, M. (2022). A novel peptide from Polypedates megacephalus promotes wound healing in mice. *Toxins (Basel)* 14 (11), 753. doi:10.3390/toxins14110753

Fuchs, P. C., Barry, A. L., and Brown, S. D. (1998). *In vitro* antimicrobial activity of MSI-78, a magainin analog. *Antimicrob. Agents Chemother.* 42 (5), 1213–1216. doi:10. 1128/AAC.42.5.1213

Gallo, R. L., and Hooper, L. V. (2012). Epithelial antimicrobial defence of the skin and intestine. *Nat. Rev. Immunol.* 12 (7), 503–516. doi:10.1038/nri3228

Galiano, R. D., Tepper, O. M., Pelo, C. R., Bhatt, K. A., Callaghan, M., Bastidas, N., et al. (2004). Topical vascular endothelial growth factor accelerates diabetic wound healing through increased angiogenesis and by mobilizing and recruiting bone marrow-derived cells. *Am. J. Pathology*, 164(6), 1935–1947. doi:10.1016/S0002-9440(10)63754-6

Ge, Y., MacDonald, D., Hait, H., Lipsky, B., Zasloff, M., and Holroyd, K. (2002). Microbiological profile of infected diabetic foot ulcers. *Diabet. Med.* 19 (12), 1032–1034. doi:10.1046/j.1464-5491.2002.00696_1.x

Ge, Y., MacDonald, D., Henry, M. M., Hait, H. I., Nelson, K. A., Lipsky, B. A., et al. (1999). *In vitro* susceptibility to pexiganan of bacteria isolated from infected diabetic foot ulcers. *Diagn Microbiol. Infect. Dis.* 35 (1), 45–53. doi:10.1016/s0732-8893(99) 00056-5

Greaves, N. S., Ashcroft, K. J., Baguneid, M., and Bayat, A. (2013). Current understanding of molecular and cellular mechanisms in fibroplasia and angiogenesis during acute wound healing. *J. Dermatol Sci.* 72 (3), 206–217. doi:10.1016/j.jdermsci. 2013.07.008

Hancock, R. E., Haney, E. F., and Gill, E. E. (2016). The immunology of host defence peptides: Beyond antimicrobial activity. *Nat. Rev. Immunol.* 16 (5), 321–334. doi:10. 1038/nri.2016.29

Haroon, Z. A., Raleigh, J. A., Greenberg, C. S., and Dewhirst, M. W. (2000). Early wound healing exhibits cytokine surge without evidence of hypoxia. *Ann. Surg.* 231 (1), 137–147. doi:10.1097/0000658-200001000-00020

He, X., Yang, Y., Mu, L., Zhou, Y., Chen, Y., Wu, J., et al. (2019). A frog-derived immunomodulatory peptide promotes cutaneous wound healing by regulating cellular response. *Front. Immunol.* 10, 2421. doi:10.3389/fimmu.2019.02421

Huang, L. C., Jean, D., Proske, R. J., Reins, R. Y., and McDermott, A. M. (2007). Ocular surface expression and *in vitro* activity of antimicrobial peptides. *Curr. Eye Res.* 32 (7-8), 595–609. doi:10.1080/02713680701446653

Julier, Z., Park, A. J., Briquez, P. S., and Martino, M. M. (2017). Promoting tissue regeneration by modulating the immune system. *Acta Biomater*. 53, 13–28. doi:10.1016/j.actbio.2017.01.056

Kang, H. K., Seo, C. H., Luchian, T., and Park, Y. (2018). Pse-T2, an antimicrobial peptide with high-level, broad-spectrum antimicrobial potency and skin biocompatibility against multidrug-resistant *Pseudomonas aeruginosa* infection. *Antimicrob. Agents Chemother.* 62 (12), e01493-18. doi:10.1128/AAC.01493-18

Koczulla, R., von Degenfeld, G., Kupatt, C., Krotz, F., Zahler, S., Gloe, T., et al. (2003). An angiogenic role for the human peptide antibiotic LL-37/hCAP-18. *J. Clin. Investigation* 111 (11), 1665–1672. doi:10.1172/JCI17545

Kolar, S. S. N., Luca, V., Baidouri, H., Mannino, G., McDermott, A. M., and Mangoni, M. L. (2015). Esculentin-1a(1-21)NH2: A frog skin-derived peptide for microbial keratitis. *Cell Mol. Life Sci.* 72 (3), 617–627. doi:10.1007/s00018-014-1694-0

Kujath, P., and Kujath, C. (2010). Complicated skin, skin structure and soft tissue infections - are we threatened by multi-resistant pathogens? *Eur. J. Med. Res.* 15 (12), 544–553. doi:10.1186/2047-783x-15-12-544

Ladram, A., and Nicolas, P. (2016). Antimicrobial peptides from frog skin: Biodiversity and therapeutic promises. *Front. Biosci. (Landmark Ed.* 21, 1341–1371. doi:10.2741/4461

Lamb, H. M., and Wiseman, L. R. (1998). Pexiganan acetate. Drugs 56 (6), 1047–1052. discussion 1053-4 (1998). doi:10.2165/00003495-199856060-00011

Lavery, L. A., Armstrong, D. G., Murdoch, D. P., Peters, E. J., and Lipsky, B. A. (2007). Validation of the Infectious Diseases Society of America's diabetic foot infection classification system. *Clin. Infect. Dis.* 44 (4), 562–565. doi:10.1086/511036

Li, X., Wang, Y., Zou, Z., Yang, M., Wu, C., Su, Y., et al. (2018). OM-LV20, a novel peptide from odorous frog skin, accelerates wound healing *in vitro* and *in vivo*. *Chem. Biol. Drug Des.* 91 (1), 126–136. doi:10.1111/cbdd.13063

Lipsky, B. A., Holroyd, K. J., and Zasloff, M. (2008). Topical versus systemic antimicrobial therapy for treating mildly infected diabetic foot ulcers: A randomized, controlled, double-blinded, multicenter trial of pexiganan cream. *Clin. Infect. Dis.* 47 (12), 1537–1545. doi:10.1086/593185

Liu, H., Duan, Z., Tang, J., Lv, Q., Rong, M., and Lai, R. (2014). A short peptide from frog skin accelerates diabetic wound healing. *FEBS J.* 281 (20), 4633–4643. doi:10.1111/ febs.12968

Liu, H., Guo, X., Yi, T., Zhu, Y., Ren, X., Guo, R., et al. (2021). Frog skin derived peptides with potential protective effects on ultraviolet B-induced cutaneous photodamage. *Front. Immunol.* 12, 613365. doi:10.3389/fimmu.2021.613365

Liu, H., Mu, L., Tang, J., Shen, C., Gao, C., Rong, M., et al. (2014). A potential wound healing-promoting peptide from frog skin. *Int. J. Biochem. Cell Biol.* 49, 32–41. doi:10. 1016/j.biocel.2014.01.010

Liu, S., Long, Q., Xu, Y., Wang, J., Xu, Z., Wang, L., et al. (2017). Assessment of antimicrobial and wound healing effects of Brevinin-2Ta against the bacterium *Klebsiella pneumoniae* in dermally-wounded rats. *Oncotarget* 8 (67), 111369–111385. doi:10.18632/oncotarget.22797

Luca, V., Stringaro, A., Colone, M., Pini, A., and Mangoni, M. L. (2013). Esculentin(1-21), an amphibian skin membrane-active peptide with potent

activity on both planktonic and biofilm cells of the bacterial pathogen *Pseudomonas aeruginosa. Cell Mol. Life Sci.* 70 (15), 2773–2786. doi:10.1007/ s00018-013-1291-7

Luckett-Chastain, L. R., and Gallucci, R. M. (2009). Interleukin (IL)-6 modulates transforming growth factor-beta expression in skin and dermal fibroblasts from IL-6-deficient mice. *Br. J. Dermatol* 161 (2), 237–248. doi:10.1111/j.1365-2133.2009.09215.x

Luo, X., Ouyang, J., Wang, Y., Zhang, M., Fu, L., Xiao, N., et al. (2021). A novel anionic cathelicidin lacking direct antimicrobial activity but with potent anti-inflammatory and wound healing activities from the salamander Tylototriton kweichowensis. *Biochimie* 191, 37–50. doi:10.1016/j.biochi.2021.08.007

Ma, L., Xie, X., Liu, H., Huang, Y., Wu, H., Jiang, M., et al. (2020). Potent antibacterial activity of MSI-1 derived from the magainin 2 peptide against drug-resistant bacteria. *Theranostics* 10 (3), 1373–1390. doi:10.7150/thno.39157

Mancl, K. A., Kirsner, R. S., and Ajdic, D. (2013). Wound biofilms: Lessons learned from oral biofilms. *Wound Repair Regen.* 21 (3), 352–362. doi:10.1111/wrr.12034

Mangoni, M. L., McDermott, A. M., and Zasloff, M. (2016). Antimicrobial peptides and wound healing: Biological and therapeutic considerations. *Exp. Dermatol* 25 (3), 167–173. doi:10.1111/exd.12929

Mansour, S. C., Pena, O. M., and Hancock, R. E. (2014). Host defense peptides: Frontline immunomodulators. *Trends Immunol.* 35 (9), 443–450. doi:10.1016/j.it.2014.07.004

Martin, P. (1997). Wound healing-aiming for perfect skin regeneration. *Science* 276 (5309), 75–81. doi:10.1126/science.276.5309.75

Mu, L., Tang, J., Liu, H., Shen, C., Rong, M., Zhang, Z., et al. (2014). A potential wound-healing-promoting peptide from salamander skin. *FASEB J.* 28 (9), 3919–3929. doi:10.1096/fj.13-248476

Mustoe, T. (2004). Understanding chronic wounds: A unifying hypothesis on their pathogenesis and implications for therapy. *Am. J. Surg.* 187 (5A), 65S–70S. doi:10.1016/S0002-9610(03)00306-4

Niyonsaba, F., Kiatsurayanon, C., Chieosilapatham, P., and Ogawa, H. (2017). Friends or foes? Host defense (antimicrobial) peptides and proteins in human skin diseases. *Exp. Dermatol* 26 (11), 989–998. doi:10.1111/exd.13314

Percival, S. L., Hill, K. E., Williams, D. W., Hooper, S. J., Thomas, D. W., and Costerton, J. W. (2012). A review of the scientific evidence for biofilms in wounds. *Wound Repair Regen.* 20 (5), 647–657. doi:10.1111/j.1524-475X.2012.00836.x

Rinaldi, A. C. (2002). Antimicrobial peptides from amphibian skin: An expanding scenario. *Curr. Opin. Chem. Biol.* 6 (6), 799-804. doi:10.1016/s1367-5931(02)00401-5

Sano, S., Itami, S., Takeda, K., Tarutani, M., Yamaguchi, Y., Miura, H., et al. (1999). Keratinocyte-specific ablation of Stat3 exhibits impaired skin remodeling, but does not affect skin morphogenesis. *EMBO J.* 18 (17), 4657–4668. doi:10.1093/emboj/18.17.4657

Sen, C. K., Gordillo, G. M., Roy, S., Kirsner, R., Lambert, L., Hunt, T. K., et al. (2009). Human skin wounds: A major and snowballing threat to public health and the economy. *Wound Repair Regen.* 17 (6), 763–771. doi:10.1111/j.1524-475X.2009.00543.x

Shi, Y., Li, C., Wang, M., Chen, Z., Luo, Y., Xia, X. S., et al. (2020). Cathelicidin-DM is an antimicrobial peptide from Duttaphrynus melanostictus and has woundhealing therapeutic potential. *ACS Omega* 5 (16), 9301–9310. doi:10.1021/ acsomega.0c00189

Song, X., Pan, H., Wang, H., Liao, X., Sun, D., Xu, K., et al. (2020). Identification of new dermaseptins with self-assembly tendency: Membrane disruption, biofilm eradication, and infected wound healing efficacy. *Acta Biomater*. 109, 208–219. doi:10.1016/j.actbio.2020.03.024

Tang, J., Liu, H., Gao, C., Mu, L., Yang, S., Rong, M., et al. (2014). A small peptide with potential ability to promote wound healing. *PLoS One* 9 (3), e92082. doi:10.1371/journal.pone.0092082

Taylor, P. K., Yeung, A. T., and Hancock, R. E. (2014). Antibiotic resistance in *Pseudomonas aeruginosa* biofilms: Towards the development of novel anti-biofilm therapies. *J. Biotechnol.* 191, 121–130. doi:10.1016/j.jbiotec.2014.09.003

Tokumaru, S., Sayama, K., Shirakata, Y., Komatsuzawa, H., Ouhara, K., Hanakawa, Y., et al. (2005). Induction of keratinocyte migration via transactivation of the epidermal growth factor receptor by the antimicrobial peptide LL-37. *J. Immunol.* 175 (7), 4662–4668. doi:10.4049/jimmunol.175.7.4662

Tredget, E. B., Demare, J., Chandran, G., Tredget, E. E., Yang, L., and Ghahary, A. (2005). Transforming growth factor-beta and its effect on reepithelialization of partialthickness ear wounds in transgenic mice. *Wound Repair Regen.* 13 (1), 61–67. doi:10. 1111/j.1067-1927.2005.130108.x

Vyas, K. S., and Wong, L. K. (2016). Detection of biofilm in wounds as an early indicator for risk for tissue infection and wound chronicity. *Ann. Plast. Surg.* 76 (1), 127–131. doi:10.1097/SAP.00000000000440

Wang, G., Chen, Z., Tian, P., Han, Q., Zhang, J., Zhang, A. M., et al. (2022). Wound healing mechanism of antimicrobial peptide cathelicidin-DM. *Front. Bioeng. Biotechnol.* 10, 977159. doi:10.3389/fbioe.2022.977159

Wang, S., Feng, C., Yin, S., Feng, Z., Tang, J., Liu, N., et al. (2021). A novel peptide from the skin of amphibian Rana limnocharis with potency to promote skin wound repair. *Nat. Prod. Res.* 35 (20), 3514–3518. doi:10.1080/14786419.2019.1710702

Wang, Y., Feng, Z., Yang, M., Zeng, L., Qi, B., Yin, S., et al. (2021). Discovery of a novel short peptide with efficacy in accelerating the healing of skin wounds. *Pharmacol. Res.* 163, 105296. doi:10.1016/j.phrs.2020.105296

Werner, S., and Grose, R. (2003). Regulation of wound healing by growth factors and cytokines. *Physiol. Rev.* 83 (3), 835–870. doi:10.1152/physrev.2003. 83.3.835

Wolcott, R. D., and Dowd, S. E. (2008). A rapid molecular method for characterising bacterial bioburden in chronic wounds. *J. Wound Care* 17 (12), 513–516. doi:10.12968/ jowc.2008.17.12.31769

Wong, W. R., Shyu, W. L., Tsai, J. W., Hsu, K. H., and Pang, J. H. (2009). Intense pulsed light effects on the expression of extracellular matrix proteins and transforming growth factor beta-1 in skin dermal fibroblasts cultured within contracted collagen lattices. *Dermatol Surg.* 35 (5), 816–825. doi:10.1111/j.1524-4725.2009.01138.x

Wolcott, R. D., Rhoads, D. D., Bennett, M. E., Wolcott, B. M., Gogokhia, L., Costerton, J. W., et al. (2010). Chronic wounds and the medical biofilm paradigm. *J. Wound Care*, 19(2), 45–46. doi:10.12968/jowc.2010.19.2.46966

Wu, J., Yang, J., Wang, X., Wei, L., Mi, K., Shen, Y., et al. (2018). A frog cathelicidin peptide effectively promotes cutaneous wound healing in mice. *Biochem. J.* 475 (17), 2785–2799. doi:10.1042/BCJ20180286

Xu, X., and Lai, R. (2015). The chemistry and biological activities of peptides from amphibian skin secretions. *Chem. Rev.* 115 (4), 1760–1846. doi:10.1021/cr4006704

Yoshizato, K. (2007). Molecular mechanism and evolutional significance of epithelialmesenchymal interactions in the body- and tail-dependent metamorphic transformation of anuran larval skin. *Int. Rev. Cytol.* 260, 213–260. doi:10.1016/S0074-7696(06)60005-3

Zhang, Y., Wang, Y., Zeng, L., Liu, Y., Sun, H., Li, S., et al. (2022). Amphibian-derived peptide homodimer OA-GL17d promotes skin wound regeneration through the miR-663a/ TGF-β1/Smad axis. *Burns Trauma* 10, tkac032. doi:10.1093/burnst/tkac032