



OPEN ACCESS

Thordur Oskarsson, Moffitt Cancer Center, United States

REVIEWED BY

Suchandrima Saha. Stony Brook Medicine, United States Yueju Li, National Taiwan University Hospital, Taiwan

*CORRESPONDENCE

Nydia Tejeda-Muñoz, ☑ Nydia-TejedaMunoz@ouhsc.edu

RECEIVED 02 May 2025 ACCEPTED 04 August 2025 PUBLISHED 22 August 2025

Flores-Hernández E, Binder G, Mei K-C and Tejeda-Muñoz N (2025) Targeting Wnt-driven metabolic adaptations in cancer: integrating glycolysis, glutaminolysis, IDO1-mediated immune evasion, and therapeutic delivery strategies.

Front. Cell Dev. Biol. 13:1622218. doi: 10.3389/fcell.2025.1622218

© 2025 Flores-Hernández, Binder, Mei and Tejeda-Muñoz. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Targeting Wnt-driven metabolic adaptations in cancer: integrating glycolysis, glutaminolysis, IDO1-mediated immune evasion, and therapeutic delivery strategies

Eric Flores-Hernández¹, Grace Binder², Kuo-Ching Mei^{3,4} and Nydia Tejeda-Muñoz^{5,6}*

¹Departamento de Biología Celular, Facultad de Ciencias, Universidad Nacional Autónoma de México (UNAM), Mexico City, Mexico, ²Department of Biological Chemistry, David Geffen School of Medicine, University of California, Los Angeles, Los Angeles, CA, United States, ³Department of Pharmaceutical Sciences, School of Pharmacy and Pharmaceutical Sciences, State University of New York at Binghamton, Binghamton, NY, United States, ⁴Department of Molecular Pharmaceutics, College of Pharmacy, University of Utah, Salt Lake City, UT, United States, ⁵Department of Oncology Science, University of Oklahoma Health Sciences Center, Oklahoma City, OK, United States, ⁶OU Health Stephenson Cancer Center, University of Oklahoma Health Sciences Center, Oklahoma City, OK, **United States**

The Wnt pathway is an evolutionarily conserved signaling cascade that regulates a wide range of fundamental cellular processes, including proliferation, differentiation, polarity, migration, metabolism, and survival. Due to its central regulatory roles, Wnt signaling is critically involved in the pathophysiology of numerous human diseases. Aberrant activation or insufficient inhibition of this pathway has been causally linked to cancer, degenerative disorders, metabolic syndromes, and developmental abnormalities. Wnt signaling drives cancer progression by reprogramming metabolism and promoting immune evasion. Wnt-driven tumors exhibit enhanced aerobic glycolysis (the Warburg effect), glutaminolysis, and macropinocytosis, which support rapid proliferation and help maintain redox homeostasis under nutrient-limited or nutrientdeprived conditions. These metabolic adaptations sustain tumor survival and contribute to immune suppression, as seen in the Wnt5a-indoleamine 2,3dioxygenase 1 (IDO1) axis, which fosters regulatory T-cell expansion and an immunosuppressive microenvironment. The interplay among glycolysis, glutamine metabolism, and immune escape renders Wnt-driven cancers highly adaptable and resistant to conventional therapies. Targeting metabolic enzymes, such as pyruvate dehydrogenase kinase 1 (PDK1), lactate dehydrogenase A (LDHA), glutaminase (GLS), and monocarboxylate transporters (MCT-1), alongside immune checkpoint inhibitors or IDO1 blockade, presents a promising strategy for overcoming metabolic plasticity and immune evasion in Wnt-driven malignancies, thereby enhancing therapeutic efficacy and improving patient survival in otherwise refractory tumor types. Combining glycolysis and glutaminolysis inhibitors with T-cell activating therapies may disrupt tumor metabolic plasticity and restore anti-tumor immunity. Additionally, advanced drug delivery systems, including lipid nanoparticles (LNPs), polymeric nanocarriers, and exosome-based platforms, enhance

the targeted accumulation of metabolic inhibitors and immunomodulatory agents while minimizing systemic toxicity. This review examines the metabolic and immune adaptations of Wnt-driven cancers, with a focus on glycolysis, glutaminolysis, and macropinocytosis. We highlight emerging therapeutic targets and nanomedicine-based delivery strategies to counteract metabolic adaptation and immune suppression. By integrating metabolic and immune-targeting with precision nano-delivery platforms, future treatment paradigms may improve outcomes for aggressive and therapy-resistant Wnt-driven cancers.

KEYWORD

Wnt signaling, macropinocytosis, metabolic reprogramming, colorectal cancer, membrane trafficking, targeted cancer therapies

Introduction

A significant shift occurs in the metabolic state of cells during cancer progression which involves the metabolism of carbohydrates, lipids, and amino acids (Kimmelman and White, 2017). This metabolic reprogramming enables the tumor to supply itself with the necessary nutrients and energy to drive proliferation, angiogenesis, and overall tumor progression (Martínez-Reyes and Chandel, 2021). One of the key metabolic adaptations in tumor cells is the Warburg effect, also known as aerobic glycolysis, which is characterized by an increased glycolytic index, elevated lactate production, and decreased oxidative phosphorylation, even in the presence of oxygen and fully functional mitochondria (Warburg, 1956). While glycolysis is less efficient in adenosine triphosphate (ATP) production than mitochondrial respiration, it facilitates biosynthetic processes essential for tumor progression including nicotinamide adenine dinucleotide phosphate (NADPH) production, lipid synthesis, and the generation of non-essential amino acids (Vaupel and Multhoff, 2021).

In colorectal cancer (CRC), Wnt signaling is a key driver of tumor progression, with its dysregulation commonly occurring due to Adenomatous Polyposis Coli (APC) mutations that stabilize β-catenin, leading to the constitutive activation of Wnt target genes (Fu et al., 2018; Liu and Yin, 2017). Beyond its role in proliferation and stemness, Wnt signaling has emerged as a central regulator of metabolic reprogramming in cancer cells (Table 1). Activating canonical (β-catenin-dependent) Wnt signaling enhances glycolysis by upregulation of key enzymes such as pyruvate dehydrogenase kinase 1 (PDK1), pyruvate kinase M2 (PKM2), lactate dehydrogenase A (LDH-A), monocarboxylate transporters (MCT-1), and glucose transporters (GLUT) (Roche et al., 2001). Inhibition of Wnt signaling, by targeting PDK1 with Peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC1α) or Chibby (a β-catenin antagonist) has been shown to reduce glycolytic activity, leading to impaired tumor growth and altered metabolic dependencies (Pate et al., 2014; Zuo et al., 2021; Cai et al., 2018). However, in addition to glycolysis, Wnt-driven tumors exhibit metabolic plasticity, shifting towards glutaminolysis to sustain biosynthetic needs. Glutaminolysis supports Tricarboxylic Acid (TCA) cycle replenishment, redox balance, and epigenetic modifications that enhance tumor progression (Dang et al., 2010; Seltzer et al., 2010). This dual reliance on glycolysis and glutaminolysis makes Wnt-driven tumors metabolically adapt, allowing them to thrive even under fluctuating nutrient conditions (Park et al., 2023; Vallée et al., 2021).

Beyond conventional nutrient uptake mechanisms, Wnt signaling has been shown to regulate macropinocytosis. This nutrient-scavenging process enables tumor cells to engulf extracellular proteins and break them down into amino acids via lysosomal degradation (Rainero, 2024; Commisso et al., 2013). While glycolysis and glutaminolysis serve as primary metabolic pathways, macropinocytosis acts as an alternative nutrient source under metabolic stress, particularly in hypoxic or glucose-deprived tumor microenvironments (Lambies et al., 2024). Wnt-driven activation of Rac1 and Mechanistic Target of Rapamycin Complex 2 (mTORC2) enhances actin cytoskeleton remodeling, promoting macropinocytic uptake (Tsujimura et al., 2016; Esen et al., 2013). This adaptation allows cancer cells to maintain biosynthesis and proliferation even when key metabolic pathways are restricted (Xu et al., 2025). Although macropinocytosis is not a primary metabolic pathway, its activation in Wnt-driven cancers enhances tumor survival under nutrient-poor conditions (Choi et al., 2017). This has led to studies in therapeutically targeting macropinocytosis alongside glycolysis and glutaminolysis to disrupt tumor metabolic flexibility and inhibit cancer progression (Li et al., 2025; Tejeda-Muñoz et al., 2024). Another emerging link between Wnt signaling and immune evasion is the Wnt5adriven upregulation of indoleamine 2,3-dioxygenase 1 (IDO1) (Sun G. et al., 2021; Holtzhausen et al., 2015; Zhao et al., 2018). IDO1 is an immunosuppressive enzyme that catabolizes tryptophan into kynurenine, leading to T-cell suppression and immune escape (Zhai et al., 2020). Notably, kynurenine-mediated tryptophan depletion also triggers metabolic shifts, reinforcing glutaminolysis and macropinocytosis as compensatory pathways in Wnt-driven tumors. This metabolic-immune interplay enhances tumor progression and resistance to immunotherapy. Targeting IDO1 in combination with metabolic inhibitors offers a promising therapeutic avenue, as IDO1 blockade (using Epacadostat, Navoximod, Indoximod, etc.) may restore anti-tumor immunity while disrupting metabolic adaptation. This highlights the need for integrated therapeutic strategies which simultaneously targets metabolism and immune evasion in Wnt-driven cancers. Furthermore, the Wnt pathway has been shown to be modulated by key metabolic enzymes and intermediates, sometimes as part of a positive feedback loops (Liang et al., 2023; Nath et al., 2015; Lai et al., 2017; Li et al., 2019; Coant et al., 2010; Madsen et al., 2024;

TABLE 1 Metabolic effects of Wnt/ β -catenin signaling pathway modulation.

Cause	Metabolic proteins involved	Effect	References
Blocking of Wnt/β-catenin signaling by dnLEF/TCFs or XAV939	PDK1	Reduces lactate production, increases ATP production, reduction in glucose consumption and increase in the rate of oxidative phosphorylation relative to glycolysis (OCR/ECAR ratio) in colorectal cancer cells. Formation of small and poorly perfused tumors.	Pate et al. (2014)
Blocking of Wnt/β-catenin signaling by PGC1α-PPARγ	PDK1	Increases basal and maximal OCR and decreases ECAR; increases cellular ATP levels, glucose uptake, and decreases extracellular lactate levels in hepatocellular carcinoma cells. Reversion of the migratory and invasive activity (Zuo et al., 2021).	Zuo et al. (2021)
Blocking of Wnt/ β -catenin signaling by Chibby	PDK1	Reduces cellular glucose uptake and lactate production, decreases cellular ATP levels, and increases cellular O ₂ consumption rate in nasopharyngeal cancer cells. Reduces cell proliferation both <i>in vitro</i> and <i>in vivo</i> .	Cai et al. (2018)
Activation of Wnt/ β -catenin/c-Myc by EHD1/14-3-3 ζ	HK2, PFKL, PGK1, ENO1, LDHA and PDK1	Higher glucose uptake, ECAR and SUVmax. EHD1/14-3-3ζ/β-catenin/c-Myc positive feedback circuit potentiates non-small cell lung cancer cells proliferation <i>in vitro</i> and <i>in vivo</i> .	Huang et al. (2021)
Blocking of Wnt/ β -catenin signaling by SLC25A18	PKM2, LDHA	Reduces glucose uptake, lactate production and ATP generation. Upregulation of SLC25A18 inhibits cell proliferation and its downregulation promotes cell growth.	Liang et al. (2020)
Blocking of Wnt/β-catenin pathway by SMAR1	GLUT1, PFK	Reduces levels in G6P, glucose consumption, lactate, ATP production, OCR, ECAR and glycolytic flux. Reduction of the viability, proliferation, migration, invasion and EMT of bladder cancer cells.	Bao et al. (2023)
Activation of Wnt/β-catenin pathway by PDLIM1	НК2	Promotion of Warburg effect. PDLIM1 inhibition reduces gastric cancer cell proliferation, migration and invasion, and promotes cell apoptosis.	Lei et al. (2024)
Activation of Wnt/ β -catenin/c-Myc pathway	PDK1, MCT-1	MCT-1/SLC16A1 is regulated by Wnt/β-catenin signaling in colon cancer cells. MCT-1 can export lactate, pyruvate as well as a glycolysis-targeting cancer drug, 3-bromopyruvate (3-BP).	Sprowl-Tanio et al. (2016)
Blocking of Wnt/β-catenin pathway by PRI-724 and IWP-O1	PFKM, PKM2 and LDHA	Decreases the expression of glycolytic enzymes and attenuates the survival of tongue carcinoma cells, reduces glucose absorption and lactate release.	Kleszcz et al. (2023)

(Continued on the following page)

TABLE 1 (Continued) Metabolic effects of Wnt/β-catenin signaling pathway modulation.

Cause	Metabolic proteins involved	Effect	References
Blocking of Wnt/ β -catenin pathway by KYA1797K or APC $^{min/+}$	PKM2, LDHA	APC-loss causes the increased expression of metabolic genes including PKM2, LDHA and increases glucose consumption and lactate secretion. Warburg effect and growth of xenografted tumors-induced by APC-mutated-colorectal cancer cells were suppressed by PKM2-depletion.	Cha et al. (2021)
Activation of Wnt/β-catenin by Rspo2-LGR4	GCK, PKM, PFKL, G6PD and LDHA	Increases glucose consumption and lactate production. LGR4 could promote hepatocellular carcinoma formation in mouse model induced by DEN and CCl4.	Bi et al. (2024)
Activation of non-canonic Wnt3a-LRP5-RAC1-mTORC2	GLUT1, HK2, PFK1, PFKFB3 and LDHA	Increases the concentration of lactate, glucose consumption and ECAR but not OCR. Reprogramming of glucose metabolism specifically contributes to Wnt-induced osteoblast differentiation.	Esen et al. (2013)
Activation of non-canonic Wnt3a-PI3K/AKT-PFKP	PFKP	Promotion of the Warburg effect, cell proliferation, colony formation and the migratory ability of cancer cells.	Jeon et al. (2021)

This table outlines how specific interventions in Wnt signaling, including inhibitors and activators, modulate the expression of key metabolic enzymes and transporters involved in glycolysis, oxidative phosphorylation, and lactate metabolism. These changes result in altered energy production, glucose uptake, lactate secretion, and metabolic reprogramming, with direct implications for tumor growth, cell proliferation, migration, invasion, and apoptosis. The data presented are supported by experimental evidence from studies conducted across various cancer cell types and models, as referenced.

Li et al., 2021; Park et al., 2016; Lu et al., 2021; Wang Z. et al., 2024). Its activation or inhibition depends on the metabolic state of the cell, and this regulation influences both normal physiological processes and altered pathological states (Table 2).

Given the metabolic plasticity of Wnt-driven tumors, singleagent metabolic inhibitors often fail to induce durable responses. To address this challenge, nanomedicine offers an opportunity to codeliver multiple metabolic inhibitors with enhanced selectivity and bioavailability (Liu P. et al., 2022). Liposomes, lipid nanoparticles (LNPs), polymeric nanoparticles, and exosome-based drug carriers have shown potential in tumor-targeted delivery of glycolysis inhibitors (Lactate Dehydrogenase A or LDHA, PDK1) (Wang et al., 2020), glutaminolysis inhibitors (Glutaminase or GLS) (Wang Q. et al., 2024), macropinocytosis inhibitors, and IDO1 blockade agents (Jiang et al., 2024). By exploiting tumor-specific metabolic markers (e.g., Glucose Transporter protein type 1 or GLUT1, macropinocytic vesicles), nanocarriers can improve drug accumulation in the tumor microenvironment while minimizing systemic toxicity (Lo et al., 2022). In this review, we discuss the interplay among Wnt signaling, metabolic reprogramming, and immune evasion in colorectal cancer. These insights open new avenues for multi-pronged therapeutic strategies targeting these converging pathways.

The Wnt signaling pathway is highly conserved across all metazoans and regulates critical processes during embryonic development as well as in adult tissue homeostasis (Holstein et al., 2011; Loh et al., 2016; Adamska et al., 2010; Hobmayer et al., 2000; Holstein, 2012). The Wnt1 gene, originally named Int-1, was first identified in 1982 by Nusse and Varmus as a proto-oncogene (Nusse and Varmus, 1982). Since then, at least 19 Wnt

family proteins have been identified in mammals. These proteins are essential for early developmental events, including primary body axis formation (Shi, 2024), morphogenetic movements, and germ layer specification during gastrulation (Bubna-Litic et al., 2024). The importance of Wnt signaling in embryogenesis is underscored by the fact that its disruption leads to severe developmental defects or early embryonic lethality (McMahon and Moon, 1989; Zeng et al., 1997; Zylkiewicz et al., 2014; Haegel et al., 1995; Huelsken et al., 2000; Liu et al., 1999). Beyond development, Wnt signaling is involved in the maintenance and regeneration of multiple adult tissues, including cardiac (Li et al., 2022), hepatic (Zou and Park, 2023), follicular (Fu et al., 2024), pulmonary (Raslan and Yoon, 2020), bone (Leucht et al., 2019), dental (Kornsuthisopon et al., 2022), and intestinal tissues (Cordero and Sansom, 2012). It is also critical for maintaining both stem cell identity and tumorigenic phenotypes (Nusse and Varmus, 2012; Mohammed et al., 2016; Yang et al., 2016). Moreover, Wnt signaling regulates genomic stability (Augustin et al., 2017; Malki et al., 2020; Arnovitz et al., 2022), cell fate and differentiation (Bagchi and MacDougald, 2021; Wang J. et al., 2024), proliferation (Lou et al., 2023; Liu et al., 2023; Gaowa et al., 2024), motility (Guo et al., 2021; VanderVorst et al., 2023; Zhang et al., 2022), as well as cell death pathways such as apoptosis and autophagy (Ma et al., 2023).

The Wnt pathway is initiated by the binding of secreted Wnt ligands—cysteine-rich, glycosylated, and acylated proteins of ~40 kDa (Tanaka et al., 2002; Liu Y. et al., 2022; Barrott et al., 2011; Biechele et al., 2011; Coombs et al., 2010)—to receptor complexes on the target cell surface. These complexes typically consist of a seven-pass transmembrane Frizzled (FZD) receptor and a co-receptor, Low Density Lipoprotein Receptor-Related Protein

TABLE 2 Effects of metabolic intermediates and enzymes on Wnt/β-catenin signaling.

Enzymes and metabolic intermediates	Effect on Wnt	Mechanism	References
Butyrate	Inhibits Wnt	Butyrate reduced nuclear β-catenin	Liang et al. (2023)
FAA (High)	Activates Wnt	Stabilizes β -catenin. Increases Wnt gene expression.	Nath et al. (2015)
MUFAs (High)	Activates Wnt	MUFA, generated by Wnt-dependent SCD, provides a positive feedback loop to stabilize β -catenin	Lai et al. (2017)
ROS (High)	Inhibits Wnt	Targeting GLS1 triggers an increase of ROS, attenuates nuclear translocation of $\beta\mbox{-catenin}$	Li et al. (2019)
ROS (High)	Inhibits Wnt	Oxidative stress causes dissociation of NRX from Dvl, which enables Dvl to activate the downstream Wnt signalling pathway.	Coant et al. (2010)
AMPK	Activates Wnt	Inhibition of AMPK stabilizes β -catenin.	Madsen et al. (2024)
AMPK	Inhibits Wnt	Increased AMPK activity reduces GSK3β activity.	Li et al. (2021)
АМРК	Inhibits Wnt	Increased AMPK activity reduces nuclear localization of $\boldsymbol{\beta}\text{-catenin.}$	Park et al. (2016)
PKM2 (Piruvate Kinase M2)	Activates Wnt	$\beta\text{-}catenin$ and the downstream target genes/proteins c-Myc and Cyclin-D1 were upregulated after overexpression of PKM2	Lu et al. (2021)
PC (Piruvate Carboxylase)	Activates Wnt	Overexpression of PC stimulates Wnt/ β -catenin pathway. The translocation of β -catenin into the nucleus was prevented by PC knockdown	Wang et al. (2024a)

The Wnt pathway has been shown to be regulated by various branches of cellular metabolism, including intermediates from lipid, mitochondrial, glycolytic, and anaplerotic metabolism. These modulations sometimes form part of a positive feedback mechanism, in which the intermediate or enzyme activates Wnt signaling to increase its concentration or activity, respectively. These changes result in alterations in energy production, glucose uptake, and maintenance of the stem-like phenotype, with direct implications for fibrosis, tumor growth, cell proliferation, migration, invasion, and epithelial-mesenchymal transition.

(LRP) 5 or 6 (Carmon and Loose, 2010; Dijksterh et al., 2015; Hsieh et al., 1999; Voloshanenko et al., 2017). Additionally, Wnt ligands may bind to alternative receptors such as the Receptor Tyrosine Kinase-like Orphan Receptors ROR1 and ROR2 (Mas et al., 1992; Masiakowski and Yancopoulos, 1998; Xu and Nusse, 1998; Yu et al., 2016). In humans, at least 10 distinct FZD receptors have been characterized (Chae and Bothwell, 2018), all featuring an extracellular N-terminal domain with a conserved cysteine-rich domain (CRD), seven transmembrane domains, and an intracellular C-terminal domain (Zheng and Sheng, 2024; Liu et al., 2024). The CRD (approximately 120 amino acids) serves as a key ligand-binding region for Wnt proteins (Janda et al., 2012).

Ligand-receptor specificity largely depends on the particular FZD involved (Hsieh et al., 1999; Voloshanenko et al., 2017). However, mapping definitive Wnt-FZD interactions remains a challenge due to the presence of 19 Wnt and 10 FZD paralogs in mammals and the observation that individual Wnt ligands can bind multiple FZDs, and *vice versa*. The LRP5/6 co-receptor facilitates ligand-receptor complex stabilization and appears to contribute to ligand specificity (He et al., 2004). ROR1 and ROR2 are single-pass transmembrane proteins that contain an extracellular immunoglobulin-like domain, a CRD, and a Kringle domain (KRD); in which the CRD is the principal domain mediating Wnt ligand interaction (Mas et al., 1992; Menck et al., 2021; Saldanha et al., 1998).

Wnt signaling is broadly categorized into two branches: the β -catenin-dependent (canonical) pathway and β -catenin-independent (non-canonical) pathway (Figure 1). The non-canonical branch includes the Planar Cell Polarity (PCP) pathway and the Wnt/Ca²⁺ pathway (Hayat et al., 2022; Qin et al., 2023). The canonical Wnt/ β -catenin pathway is defined by the stabilization and nuclear translocation of β -catenin following Wnt ligand binding (Moon et al., 2004; Rim et al., 2022). In contrast, the PCP pathway governs cellular and tissue polarity along the body axis (Shi, 2022), while the Wnt/Ca²⁺ pathway involves intracellular calcium flux and activation of calcium-sensitive signaling molecules (De, 2011).

Although these pathways were initially considered distinct, recent evidence suggests that Wnt signals may activate multiple pathways concurrently and even antagonistically (Thrasivoulou et al., 2013; Tufail and Wu, 2023). Traditionally, Wnt1, Wnt3a, Wnt8, and Wnt8b have been classified as canonical ligands, whereas Wnt4, Wnt5a, and Wnt11 are linked to non-canonical signaling. However, this classification is now viewed as oversimplified, as growing evidence shows that the same Wnt ligand can activate both canonical and non-canonical pathways depending on the context (Flores-Hernández et al., 2020). We explore the interplay among Wnt signaling, metabolic reprogramming, and immune evasion in colorectal cancer, focusing on key metabolic pathways and delivery strategies that may enhance therapeutic efficacy.

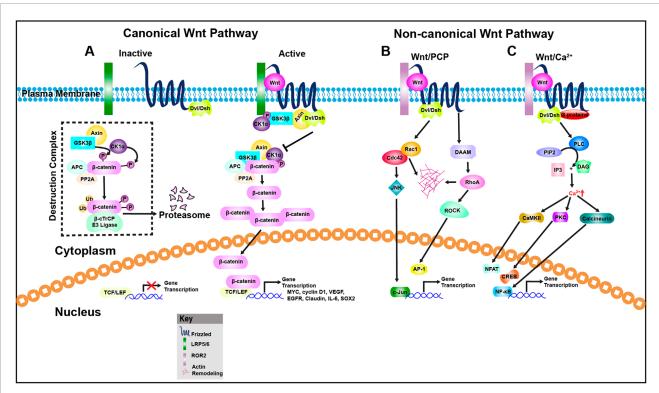


FIGURE 1
What signaling pathways. (A) Canonical What signaling pathway: In the absence of What ligands, β-catenin is targeted for degradation by a destruction complex composed of Axin, APC, CK1α, and GSK3β. This complex phosphorylates β-catenin, marking it for ubiquitination by the β-TrCP E3 ligase and subsequent proteasomal degradation. As a result, β-catenin levels remain low in the cytoplasm, and TCF/LEF transcription factors are bound to corepressors, preventing What target gene expression. Upon What ligand binding to Frizzled (Fz) receptors and co-receptors LRP5/6, Dishevelled (DVL) is recruited to the membrane complex, leading to inhibition of GSK3β and stabilization of β-catenin. Accumulated β-catenin translocates into the nucleus, where it displaces co-repressors and interacts with TCF/LEF to drive transcription of What-responsive genes such as MYC, Cyclin D1, VEGF, EGFR, Claudin, IL-β, and SOX2. (B) Non-canonical Wht/Planar Cell Polarity (PCP) Pathway: This β-catenin-independent pathway regulates cytoskeletal organization and cell polarity. What ligands such as What engage Fz receptors and ROR1/2 or RYK co-receptors, activating DVL and downstream small GTPases like Cdc42, Rac1, and RhoA. Rac1 activates JNK signaling, promoting actin remodeling and transcription via AP-1 and c-Jun. RhoA activates ROCK (Rho-associated kinase), contributing to cytoskeletal dynamics and potentially transcriptional regulation. (C) Non-canonical What/Calcium (Ca²⁺) Pathway: This pathway is triggered by What ligands such as What5a, What11, or What4, which bind Fz or ROR receptors and activate heterotrimeric β proteins. These in turn stimulate phospholipase β (PLC), leading to hydrolysis of PIP2 into IPβ and DAG. IPβ induces β release from the endoplasmic reticulum, which activates CaMKII and calcineurin. Calcineurin dephosphorylates NFAT, promoting its nuclear import and transcriptional activity. DAG activates PKC, which influences other transcription factors including CREB and NF-β contr

Canonical Wnt/β-catenin pathway

In the absence of Wnt ligands, the pathway remains inactive due to the continuous activity of the β -catenin destruction complex, which keeps cytoplasmic and nuclear β -catenin levels low (Figure 1A). This multiprotein complex includes the scaffold protein Axin, the tumor suppressor Adenomatous Polyposis Coli (APC), and the serine/threonine kinases CK1 α (Casein Kinase 1 alpha) and Glycogen Synthase Kinase 3 beta (GSK3 β). β -catenin is sequentially phosphorylated at specific residues (Ser33, Ser37, and Thr41), which marks it for recognition by the E3 ubiquitin ligase complex component β -TrCP. Ubiquitinated β -catenin is then targeted for proteasomal degradation (Hayat et al., 2022). Upon Wnt ligand binding to the FZD-LRP5/6 receptor complex, the intracellular protein Dishevelled (Dvl) is recruited to the membrane and activated. Concurrently, the LRP5/6 co-receptor is

phosphorylated. These events lead to the assembly of a receptor-associated "signalosome" that sequesters components of the β -catenin destruction complex, thereby inhibiting GSK3 β activity. As a result, β -catenin escapes degradation, accumulates in the cytoplasm, and translocated into the nucleus. In the nucleus, β -catenin binds to T-cell factor/lymphoid enhancer-binding factor (TCF/LEF) family transcription factors to activate Wnt target gene expression involved in proliferation, differentiation, and stem cell maintenance (Hayat et al., 2022).

Non-canonical Wnt/planar cell polarity (PCP) pathway

The PCP pathway represents a β -catenin-independent branch of Wnt signaling that is highly conserved and essential for directing cellular orientation and movement during morphogenesis (Goodrich and Strutt, 2011; Koca et al., 2022). "Planar polarity"

refers to the coordinated alignment of cells within the plane of a tissue orthogonal to the apical-basal axis. The PCP pathway is critical in numerous developmental processes such as gastrulation, neural tube closure, and organogenesis (Williams and Solnica-Krezel, 2020; Henders et al., 2018; Matsuda and Sokol, 2021; Shi, 2023). Mutations in PCP genes are associated with developmental disorders including spina bifida (Bosoi et al., 2011), hearing loss (Sun H. et al., 2021), cystic kidney disease (Richards et al., 2019), and limb malformations (Zhu et al., 2023).

PCP signaling modulates cytoskeletal dynamics through downstream effectors including RhoA, Rac1, JNK, and PKC (Figure 1B), influencing cellular morphology and motility (Wallingford, 2012; Adler and Wallingford, 2017). The pathway relies on a group of evolutionarily conserved core PCP proteins: Frizzled (FZD), Dishevelled (Dvl), Celsr, Vangl, Prickle, and Ankrd6 (Strutt and Strutt, 2009). These proteins form asymmetric complexes on opposite sides of a cell, establishing polarity before any visible morphological changes occur. Wnt ligands such as Wnt5a, Wnt5b, and Wnt11 activate PCP signaling by binding to FZD receptors in complex with ROR1/2 or RYK co-receptors (Gao, 2012). Downstream effectors can be broadly classified into two groups: (1) the planar polarity effectors (PPE), including Daam1, small Rho GTPases, and JNK; and (2) the ciliogenesis and planar polarity effectors (CPLANE), such as Intu, Fuz, and Wdpcp, which are less well characterized but contribute to ciliary assembly and polarity (Goodrich and Strutt, 2011; Yang and Mlodzik, 2015).

Non-canonical Wnt/calcium pathway

The Wnt/Ca²⁺ pathway is another β -catenin-independent signaling route that regulates intracellular calcium flux and associated cellular functions (Slusarski et al., 1997; Anastas and Moon, 2013). Upon ligand binding—typically by Wnt5a or Wnt11—G proteins are activated, leading to the stimulation of phospholipase C (PLC). This, in turn, generates inositol trisphosphate (IP₃), which induces the release of calcium from intracellular stores. The resulting calcium surge activates several calcium-dependent effectors, including protein kinase C (PKC) and Ca²⁺/calmodulin-dependent kinase II (CaMKII) (Kühl et al., 2000; Wang et al., 2010), thereby influencing cell migration, polarity, and fate determination (Figure 1C).

A key transcriptional component of the Wnt/Ca²⁺ pathway is the Nuclear Factor of Activated T-cells (NFAT), which is activated via the calcium-dependent phosphatase calcineurin. Upon dephosphorylation, NFAT is translocated to the nucleus where it drives the expression of genes involved in morphogenesis (Farrera-Hernández et al., 2021), mesenchymal-to-epithelial transitions (Dejmek et al., 2006), and immune regulation (Crabtree and Olson, 2002).

Wnt/ β -catenin signaling and aerobic glycolysis

Numerous studies support a relationship between Wnt/ β -catenin signaling and metabolic reprogramming, particularly aerobic glycolysis. Knockdown assays in lung adenocarcinoma cells

have shown that Wnt/β-catenin activation increases c-Myc and PDK1 protein levels, reinforcing its role in enhancing glycolytic flux (Huang et al., 2021). In colon cancer cell lines, overexpression of the mitochondrial glutamate transporter SLC25A18 reduced glucose uptake and downregulated β-catenin, Lactate Dehydrogenase A (LDHA), and Pyruvate Kinase M2 (PKM2), while its deletion had the opposite effect. Notably, the inhibitory effects of SLC25A18 on glucose metabolism, lactate production, ATP generation, and cell proliferation were reversed by the Wnt inhibitor Dickkopfrelated protein 1 (DKK1) (Liang et al., 2020). Similar observations were seen in bladder cancer, where overexpression of SMAR1 (Scaffold/matrix attachment region binding protein 1) reduced glycolysis by inhibiting GLUT1 and phosphofructokinase (PFK) expression. Treatment with XAV-939 (a Wnt/β-catenin inhibitor) reversed the increase in glycolytic activity following SMAR1 silencing, while treatment with LiCl (a Wnt activator) negated the effects of SMAR1 overexpression (Bao et al., 2023). In gastric cancer cells, PDLIM1 (PDZ and LIM domain protein 1) interacted with HK2 (Hexokinase 2) to drive glycolysis, proliferation, migration, and apoptosis resistance via Wnt/β-catenin signaling, particularly under glucose-deprived conditions (Lei et al., 2024). Additionally, monocarboxylate transporter 1 (MCT-1), encoded by SLC16A1, was identified as a direct Wnt target gene, and its Wnt response elements (WREs) in the SLC16A1 promoter were sensitive to Wnt inhibition (Sprowl-Tanio et al., 2016). Further supporting Wnt's role in metabolic control, Wnt inhibitors PRI-724 and IWP-O1 attenuated glucose uptake and lactate production in tongue carcinoma cells while also reducing the expression of Phosphofructokinase, Muscle type (PFKM), PKM2, and LDHA (Kleszcz et al., 2023). Likewise, in colon adenocarcinoma cells, loss of APC increased the expression of PKM2 and LDHA, along with glucose consumption and lactate secretion. Depletion of PKM2 suppressed glycolysis and inhibited xenograft tumor growth in APC-mutant cells (Cha et al., 2021). Beyond APC-driven metabolic shifts, leucine-rich repeatcontaining G protein-coupled receptor 4 (LGR4), a receptor in the Wnt pathway, promoted metabolic reprogramming in hepatocellular carcinoma (HCC) by enhancing glycolysis and lactate production. Notably, LGR4 activation by R-spondin ligands enhanced Wnt/βcatenin signaling, driving cancer progression (Bi et al., 2024; Glinka et al., 2011). LGR5, along with its homologs LGR4 and LGR6, belongs to subgroup B of the leucine-rich repeat-containing G protein-coupled receptors (LGRs), a subset of the GPCR superfamily typically associated with hormone signaling (Hsu et al., 2000). While LGR4/5/6 share a high degree of structural similarity, their expression patterns only partially overlap, and functional studies indicate that they have distinct phenotypic effects (Kriz and Korinek, 2018). Although all three receptors are known to modulate Wnt signaling in cooperation with R-spondins, only LGR4 has been conclusively implicated in metabolic regulation via Wnt pathway interactions so far (Zhang et al., 2023). Additionally, c-Myc and cyclin D1, two downstream Wnt targets, further stimulate aerobic glycolysis (Wise et al., 2008). The integration of Wnt signaling with metabolic reprogramming is crucial for cancer progression, particularly through the regulation of glycolytic enzymes such as 3phosphoinositide-dependent protein kinase 1 (PDK1), PKM2, and LDHA (Rong et al., 2024; Leung and Lee, 2022). The Warburg effect, driven by Wnt signaling, not only enhances energy production but also generates biosynthetic intermediates essential for tumor growth

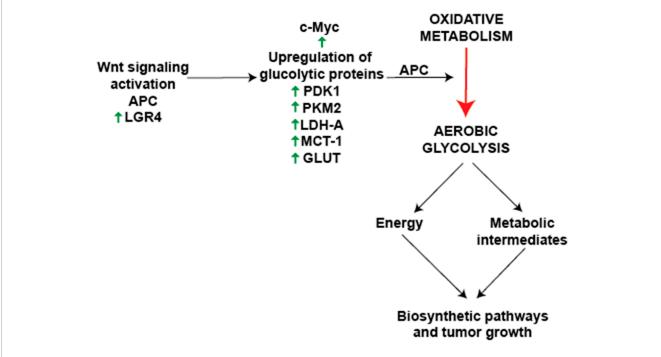


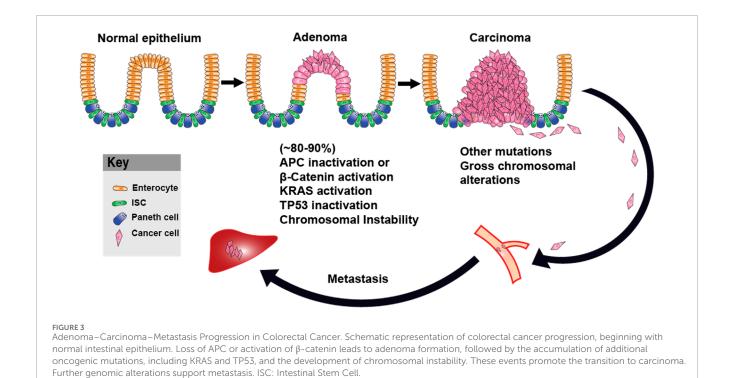
Figure 2
Canonical Wnt signaling and cancer metabolic reprogramming activation of Wnt signaling via the β-catenin pathway leads to the transcriptional regulation of key metabolic enzymes and transporters, including PDK1, LDHA, PKM2, and GLUT1. These molecular changes promote the Warburg effect (aerobic glycolysis), characterized by increased glucose uptake, lactate production, and diversion of metabolic intermediates into biosynthetic pathways critical for tumor growth. The schematic also highlights the inhibition of pyruvate oxidation through the PDK1-mediated inactivation of PDH and its impact on oxidative phosphorylation. Additionally, the role of downstream targets such as c-Myc and LGR4 in supporting glycolytic flux and cell proliferation is depicted. These processes collectively underscore the significance of Wnt signaling in driving metabolic adaptability and cancer progression. This representation summarizes findings on the interrelation between Wnt signaling and cancer metabolism, emphasizing therapeutic opportunities in targeting these pathways.

and survival (Figure 2). This metabolic reprogramming is closely associated with the stepwise progression from normal intestinal epithelium to carcinoma and metastasis, as shown in Figure 3. Loss of APC or activation of β -catenin initiates adenoma formation, followed by mutations in KRAS and TP53, and an increase in chromosomal instability, which collectively drive the transition from adenoma to carcinoma. Additional mutations and chromosomal alterations facilitate the metastasis process. Therapeutically targeting Wnt-driven metabolic enzymes thus offers a compelling strategy for malignancies dependent on metabolic reprogramming.

Non-canonical Wnt pathway and metabolic regulation

β-catenin-independent (non-canonical) Wnt signaling also plays a key role in energy metabolism regulation (Figure 4). Although Wnt3a is classically characterized as a canonical Wnt ligand that primarily activates the β -catenin–dependent signaling cascade, emerging evidence suggests that, in certain cellular contexts, Wnt3a can also trigger non-canonical signaling responses. Studies have shown that Wnt3a increases aerobic glycolysis by upregulating key glycolytic enzymes. Interestingly, this metabolic regulation requires LRP5 but not β -catenin and instead relies on mTORC2-AKT (also known as Protein Kinase B or PKB) signaling downstream of RAC1 (19). In another assay, Wnt3a-induced

EGFR-PI3K/AKT transactivation stabilized phosphofructokinase P (PFKP) via S386 phosphorylation, leading to increased PFKP expression, enhanced glycolysis, as well as cell proliferation and migration, all independent of β -catenin (Jeon et al., 2021). The Wnt pathway also interacts with multiple signaling cascades, including PI3K/Akt, STAT3, and c-Myc-driven transactivation of Hypoxia-Inducible Factor 1-alpha (HIF-1α) (Shih and Mei, 2021). HIF-1α activation promotes aerobic glycolysis, lactate production, and tumor progression, even in normoxic conditions (El-Sahli et al., 2019). Elevated lactate production due to HIF-1a and c-Myc upregulation increases PDK1 and LDHA expression, which enhances glycolytic flux (Liao, 2017). HIF-1α-induced PDK1 expression inhibits the PDH complex, preventing pyruvate entry into the mitochondrial TCA cycle and increasing lactate release. This metabolic shift drives tumor microenvironment remodeling (Gatenby et al., 2006), enhances cell migration (Baumann et al., 2009; Seliger et al., 2013), suppresses the immune response (Fischer et al., 2007; Gottfried et al., 2006), and promotes antiapoptotic signaling (Kondoh et al., 2005). Additionally, c-Myc and PI3K/Akt-mediated activation of HIF-1α inhibits glucose oxidation via PDK1 and hexokinase 2 (HK2) (Kim et al., 2007; Lum et al., 2007). Thus, non-canonical Wnt signaling not only regulates glycolysis but also modulates key oncogenic pathways such as PI3K/Akt-HIF-1α, shaping the metabolic landscape of cancer cells. These mechanisms underscore new therapeutic targets in metabolic



reprogramming, which could complement canonical Wnt inhibition strategies.

Glutaminolysis and additional metabolic interactions

In cancer cells, pyruvate produced during glycolysis is converted into lactate instead of being utilized in the tricarboxylic acid (TCA) cycle. Although mitochondrial ATP dependency decreases in tumor cells, the demand for biosynthetic precursors and NADPH remains unchanged (Frezza and Gottlieb, 2009). To compensate for these changes and maintain the TCA pathway's functionality, cancer cells often rely on increased glutaminolysis (Wise and Thompson, 2010; Medina, 2001; Reitzer et al., 1979; Lu et al., 2010). Glutaminolysis is an anaplerotic pathway involving the deamination of glutamine, converting it into glutamate and subsequently into alpha-ketoglutarate (α-KG) by glutaminase (GLS), GDH, and other enzymes. This process enables ATP production through the TCA cycle and provides nitrogen, sulfur, and carbon atoms needed for the synthesis of biosynthetic precursors essential for cancer cell growth and proliferation (Wise and Thompson, 2010; Medina, 2001; Reitzer et al., 1979; Lu et al., 2010). The relationship between Wnt/β-catenin signaling and glutaminolysis remains partially understood; however, emerging evidence highlights key interactions between these two processes. In hepatocellular carcinoma (HCC), overexpression of glutamine synthase (GS) correlates with Wnt/βcatenin activation, indicating a link between glutamine metabolism and tumor progression (Wong et al., 2020). In KRAS-mutated colorectal cancer (CRC) cells, SLC25A22-mediated glutamate transport promotes succinate accumulation, which in turn activates the Wnt pathway, induces DNA methylation, and upregulates

LGR5. These effects drive stem cell-like properties, proliferation, and chemotherapy resistance (Cadoret et al., 2002). Additionally, inflammatory responses further integrate glutaminolysis with the Wnt signaling pathway. Pulmonary endothelial cells release Rspondin3, a β-catenin activator, which enhances mitochondrial respiration via glutaminolysis. This produces α-ketoglutarate, which modifies DNA hydroxymethylation via TET2 (Ten-Eleven Translocation-2), linking Wnt activity to epigenetic regulation (Zhou et al., 2020). In gastric cancer (GC) cells, circHECTD1 regulates glutaminolysis via the miR-1256/USP5 axis, activating the β-catenin/c-Myc pathway and promoting tumor progression (Cai et al., 2019). Similarly, niclosamide, a Wnt inhibitor, reduces liver fibrosis and metabolic reprogramming, highlighting potential therapeutic applications (El-Ashmawy et al., 2020). These findings underscore the intertwined relationship between glutaminolysis, epigenetic remodeling, and Wnt signaling in cancer progression and therapy resistance. Targeting Wnt-glutaminolysis interactions may offer novel strategies for disrupting tumor metabolism and enhancing therapeutic efficacy.

Wnt signaling and macropinocytosis as an adaptive metabolic strategy

Macropinocytosis is a form of endocytosis characterized by the non-selective internalization of extracellular fluid into large vesicles known as macropinosomes (Mercer and Helenius, 2009; Bloomfield and Kay, 2016). These vesicles are typically 0.2–10 μm in diameter—significantly larger than those formed by other endocytic pathways—which enables the uptake of solutes and nutrients that cannot be internalized through more selective mechanisms such as clathrin-mediated endocytosis (Welliver et al., 2011). Once

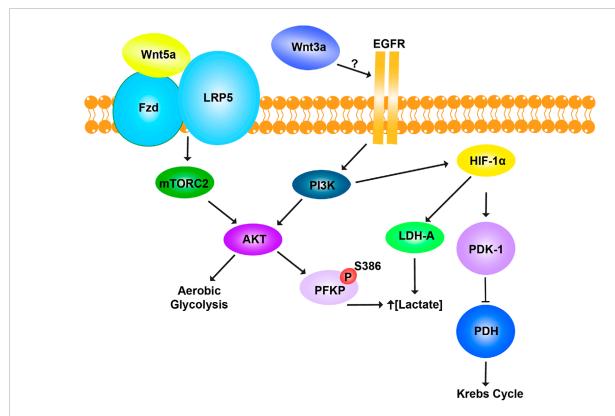
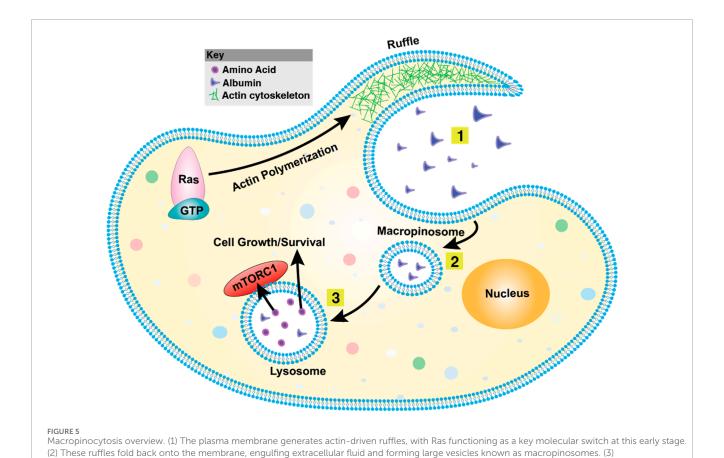


FIGURE 4
Non-canonical Wnt pathway and metabolic regulation. The Wnt pathway regulates glycolysis and mitochondrial respiration in a β -catenin-independent manner. Wnt5a promotes aerobic glycolysis through activation of the mTORC2-AKT pathway, the co-receptor LRP5. Activation of AKT enhances glycolytic metabolism to meet cellular energy demands. Wnt3a regulates cellular metabolism through a distinct β -catenin-independent mechanism by engaging EGFR, though the exact interaction remains unclear. Wnt3a promotes lactate production by phosphorylating phosphofructokinase platelet type (PFKP) at residue S386 and activating lactate dehydrogenase A (LDH-A). This increases lactate concentration, supporting glycolytic flux. Simultaneously, Wnt3a inhibits mitochondrial respiration by upregulating pyruvate dehydrogenase kinase-1 (PDK-1), which phosphorylates and inactivates pyruvate dehydrogenase (PDH). As a result, pyruvate is diverted away from the Krebs cycle and toward fermentation, enhancing glycolysis. Together, Wnt5a and Wnt3a reprogram cellular metabolism to favor glycolysis, bypassing mitochondrial respiration through β -catenin-independent pathways.

formed, macropinosomes traffic through the endocytic system, where their contents are degraded, and the resulting metabolites are absorbed by the cell (Buckley and King, 2017). The process is driven by actin cytoskeletal remodeling and the formation of membrane ruffles that fold back onto the plasma membrane to engulf extracellular fluid (Welliver et al., 2011; Hoon et al., 2012; Bernitt et al., 2015; Valdivia et al., 2017). Unlike receptor-mediated endocytosis, macropinocytosis is a non-selective process that is typically activated by extracellular signals. It plays critical roles in cellular nutrient acquisition (Amyere et al., 2002), immune surveillance and antigen presentation (Sallusto et al., 1995; Bondy-Denomy et al., 2013; Liu and Roche, 2015; Canton, 2018), microbial pathogenesis (Lee C. H. R. et al., 2019; Lee J. H. et al., 2019), and tumor biology (Figure 5).

First described nearly a century ago (Edwards, 1925; Lewis, 1931), macropinocytosis has since been observed across a wide range of organisms, from amoebae to mammals, suggesting an evolutionarily ancient origin (Guha et al., 2003; Fares and Greenwald, 2001; Ganot et al., 2020). In cancer, macropinocytosis contributes to tumor cell survival, proliferation, and metastasis, particularly under stressful conditions such as hypoxia or nutrient deprivation (Commisso et al., 2013; Kamphorst et al., 2015).

Its activation is commonly driven by oncogenic mutations in genes such as KRAS, components of the PI3K/AKT pathway, and Rho-family GTPases (Overmeyer et al., 2008; Palm et al., 2015; Kim et al., 2018). Furthermore, aberrant activation of signaling pathways including Wnt (Redelman-Sidi et al., 2018; Albrecht et al., 2020), Hippo (King et al., 2020), and the energy sensor AMPK (Kim et al., 2018) can also induce macropinocytosis in specific cancer contexts. Importantly, this process has been implicated in mechanisms of therapeutic resistance, highlighting its relevance in cancer progression and treatment response (Qian et al., 2014). The convergence of Wnt signaling, macropinocytosis, and metabolic reprogramming is a key adaptive strategy in cancer, allowing tumor cells to thrive under nutrient-limited conditions. Although canonical Wnt/β-catenin signaling is primarily recognized for its role in tumor progression, recent studies reveal its involvement in controlling macropinocytosis, a vital mechanism for sustaining cellular growth. (Tejeda-Muñoz et al., 2019). Macropinocytosis enables tumor cells to engulf extracellular proteins, which are subsequently degraded in lysosomes to provide amino acids and metabolic intermediates, thereby supporting biosynthetic processes, proliferation, and redox homeostasis (Tejeda-Muñoz et al., 2019; Tejeda-Muñoz and Mei, 2024). This pathway is particularly



Macropinosomes undergo maturation and can fuse with endosomes or lysosomes, leading to the degradation of internalized material. This process plays a vital role in nutrient acquisition, antigen sampling, and pathogen entry. The mechanistic Target of Rapamycin Complex 1 (mTORC1) is a central

regulator of macropinocytosis, integrating nutrient availability with cell growth and metabolic responses.

critical in hypoxic and nutrient-deprived tumor regions, where conventional nutrient uptake mechanisms are insufficient to meet the metabolic demands of rapidly proliferating cancer cells (Amyere et al., 2002; Tejeda-Muñoz et al., 2019; Tejeda-Muñoz and Mei, 2024). Recent findings suggest that Wnt signaling enhances macropinocytosis through actin cytoskeleton remodeling, facilitating extracellular protein uptake and metabolic adaptation (Figure 5). By utilizing macropinocytosis, tumor cells maintain metabolic flexibility, allowing them to sustain biosynthesis even under nutrient limitations (Tejeda-Muñoz and Robertis, 2022). Additionally, macropinocytosis buffers oxidative stress by providing amino acids essential for redox homeostasis and modulates the tumor immune response, enabling immune evasion (Shen et al., 2018). Wnt-driven macropinocytosis also interacts with lysosomal function, promoting focal adhesion dynamics that enhance tumor cell motility and invasion (Tejeda-Muñoz and Robertis, 2022). Focal adhesions, which link the actin cytoskeleton to the extracellular matrix (ECM), are dynamically regulated during cancer cell migration and metastasis (Shen et al., 2018). Wnt activation, through both canonical and non-canonical pathways, facilitates ECM remodeling, reinforcing the invasive potential of cancer cells

Macropinocytosis is intricately linked to Wnt-regulated metabolic pathways, including the Warburg effect and glutaminolysis, which enable tumors to maintain high proliferative

rates even in hypoxia (Tejeda-Muñoz and Robertis, 2022). The Warburg effect provides ATP through glycolysis, while glutaminolysis supplies TCA cycle intermediates and biosynthetic precursors. Recent studies demonstrate that macropinocytosis and glutaminolysis are co-regulated by Wnt signaling. Specifically, Wnt/ β -catenin signaling regulates the expression of glutamine transporters and enzymes, such as glutaminase (GLS), which catalyzes the conversion of glutamine to glutamate (Shen et al., 2021). This highlights the role of Wnt signaling in orchestrating multiple metabolic pathways to support tumor growth.

Beyond canonical Wnt signaling, non-canonical Wnt pathways also contribute to macropinocytosis regulation. Components such as Rac1 and mTORC2 integrate non-canonical Wnt signaling with metabolic adaptation (Tejeda-Muñoz and Mei, 2024). Rac1, a small GTPase, is essential for actin cytoskeleton remodeling and macropinosome formation, facilitating nutrient scavenging in tumor cells (Buckley et al., 2020). mTORC2, a key metabolic regulator, is activated downstream of non-canonical Wnt signaling (Fu et al., 2021) and influences glycolysis, Warburg metabolism, and protein translation (Esen et al., 2013). This crosstalk between Wnt signaling, Rac1, and mTORC2 underscores the metabolic adaptability of cancer cells, where both canonical and non-canonical pathways converge to drive tumor survival.

Targeting Wnt signaling to disrupt macropinocytosis presents a promising therapeutic approach, as it could restrict nutrient

(Penny et al., 2023; Pham et al., 2022).

availability, impair metabolic flexibility, and inhibit tumor growth. Wnt inhibitors such as PRI-724, which have been investigated for anti-tumor activity, show potential for modulating metabolic pathways and impairing macropinocytosis (Tejeda-Muñoz et al., 2019; Tejeda-Muñoz and Mei, 2024). Cancers with APC mutations or aberrant Wnt signaling may be particularly sensitive to therapies targeting macropinocytosis and associated metabolic rewiring. Additionally, combining Wnt inhibitors with ECM-targeting agents or immune checkpoint inhibitors may enhance therapeutic efficacy by simultaneously disrupting tumor metabolism and the tumor microenvironment. Given that macropinocytosis supports both metabolic adaptation and immune evasion, dual inhibition strategies could provide long-term benefits by overcoming resistance mechanisms.

Wnt5a-IDO1 axis: linking metabolic reprogramming and immune evasion

Wnt signaling not only regulates metabolic reprogramming but also plays a crucial role in modulating immune evasion (Malladi et al., 2016; Katoh and Katoh, 2022). An emerging link between Wnt signaling and immune suppression is the Wnt5adriven upregulation of IDO1, an immunoregulatory enzyme that depletes tryptophan and generates immunosuppressive kynurenine metabolites (Holtzhausen et al., 2015). IDO1 activity is known to promote immune tolerance within the tumor microenvironment, enabling evasion of T-cell-mediated surveillance and resistance to immune checkpoint blockade therapy (Zhao et al., 2018). Recent studies have shown that Wnt5a, a non-canonical Wnt ligand, activates the IDO1 pathway, thereby reinforcing immune suppression while sustaining metabolic adaptations that support tumor survival and proliferation (Holtzhausen et al., 2015; Zhao et al., 2018; DeVito et al., 2021). This shift promotes the expansion of regulatory T cells (Tregs) while suppressing the activation of effector T cells (Thaker et al., 2013). In colorectal cancer, aberrant Wnt5a signaling correlates with elevated IDO1 expression and reduced infiltration of cytotoxic CD8+ T cells, indicating a more immunosuppressive tumor microenvironment (Thaker et al., 2013). Moreover, tryptophan deprivation or depletion further exacerbates metabolic stress within the tumor microenvironment, reinforcing pathways such as glutaminolysis and macropinocytosis as compensatory mechanisms for nutrient acquisition (Thaker et al., 2013). This metabolic-immune crosstalk highlights how Wnt5adriven IDO1 activity facilitates immune escape and sustains metabolic plasticity, thereby supporting tumor progression.

The metabolic effects of IDO1 extend beyond immune evasion, influencing key nutrient utilization pathways that sustain tumor cell survival. IDO1-induced tryptophan deprivation/depletion alters amino acid metabolism, shifting cellular dependence toward glutaminolysis as a compensatory energy source (Odunsi et al., 2022). This adaptation is particularly evident in Wnt-driven tumors, where IDO1 activation leads to the upregulation of glutamine transporters (SLC1A5, SLC7A5) and glutaminase (GLS1), enhancing glutamine metabolism to compensate for the lack of tryptophan-derived intermediates (Yoo and Han, 2022). In addition to glutaminolysis, IDO1 activity influences lipid metabolism and mitochondrial function. Studies suggest that kynurenine

accumulation promotes oxidative phosphorylation (OXPHOS) and fatty acid oxidation (FAO), favoring metabolism pathways that are less dependent on glucose (Zhao et al., 2018). This metabolic plasticity enables tumors to thrive in nutrient-deprived and hypoxic microenvironments, further reinforcing the survival advantage of Wnt5a-IDO1 signaling. The immunosuppressive effects of Wnt5a-IDO1 signaling pose significant challenges for immunotherapy, particularly in tumors resistant to immune checkpoint inhibitors (ICIs) targeting Programmed Cell Death Protein 1 (PD-1), Programmed Death-Ligand 1 (PD-L1), and Cytotoxic T-Lymphocyte-Associated protein 4 (CTLA-4) (Liu et al., 2021). IDO1-mediated kynurenine accumulation has been shown to suppress dendritic cell maturation and impair antigen presentation, leading to reduced activation of tumor-infiltrating lymphocytes (TILs) (Jochems et al., 2016). Furthermore, Wnt5a signaling promotes the expression of PD-L1 on tumor cells and myeloid-derived suppressor cells (MDSCs), reinforcing an immunosuppressive microenvironment that dampens anti-tumor immunity (Douglass et al., 2021). Clinical studies indicate that tumors with high Wnt5a and IDO1 expression exhibit poor responses to immune checkpoint blockade, suggesting that the Wnt5a-IDO1 axis serves as a mechanism of primary or acquired resistance to immunotherapy. Targeting this pathway alongside ICIs could enhance therapeutic efficacy by restoring anti-tumor immunity and reversing metabolic adaptations that fuel tumor progression (Holtzhausen et al., 2015; DeVito et al., 2021; Theivanthiran et al., 2020). Given the dual role of Wnt5a-IDO1 signaling in immune evasion and metabolic adaptation, therapeutic strategies targeting this axis hold promise for overcoming resistance to immunotherapy and metabolic inhibitors. Pharmacological IDO1 inhibitors such as Epacadostat, Navoximod, and Indoximod have shown potential in reversing kynurenine-mediated immune suppression (Prendergast et al., 2017). However, results from clinical trials, such as ECHO-301, demonstrate that IDO1 inhibition alone or in combination with anti-PD1 is insufficient in highly immunosuppressive tumors (Muller et al., 2019; Chen et al., 2021; Smith et al., 2020). These outcomes underscore the need to explore new combination strategies for IDO1 inhibition. Wnt5a signaling can be targeted using small-molecule Wnt inhibitors (e.g., ETC-159, C59) or monoclonal antibodies blocking Wnt5a-Fzd interactions (Chua et al., 2023; Motono et al., 2016). These inhibitors may not only suppress IDO1 expression but also disrupt Wnt5a-driven metabolic adaptations, thereby sensitizing tumors to immunotherapy and metabolic-modulating therapeutics.

Advanced therapeutic delivery strategies for immuno-metabolic reprogramming in Wnt-driven cancers

The intricate interplay among Wnt signaling, metabolic plasticity, and immune evasion in tumors presents a significant challenge for conventional therapies. Wnt-driven cancers exploit aerobic glycolysis (the Warburg effect), glutaminolysis, and macropinocytosis to sustain their biosynthetic needs, ensuring tumor survival in nutrient-deprived conditions while evading immune surveillance. Given these complexities, therapeutic delivery strategies must integrate metabolic and immunomodulatory

interventions to reprogram the tumor microenvironment and improve treatment efficacy. Advances in nanomedicine, biomaterials, and engineered drug delivery platforms offer new opportunities to co-target metabolic and immune pathways, addressing tumor resistance mechanisms at multiple levels. One promising approach involves co-delivering small-molecule or nucleic acid-based metabolic inhibitors alongside immune-modulating agents using lipid nanoparticles (LNPs) (Wei et al., 2024; Nel et al., 2022), polymeric nanocarriers (Dubey et al., 2017; Mei et al., 2016), and liposomes (Shah et al., 2020; Etter et al., 2021). These nanocarriers can be further functionalized to be immune-stimulatory or immune cell-targeted (Miao et al., 2019; Mei et al., 2023; Mei et al., 2024).

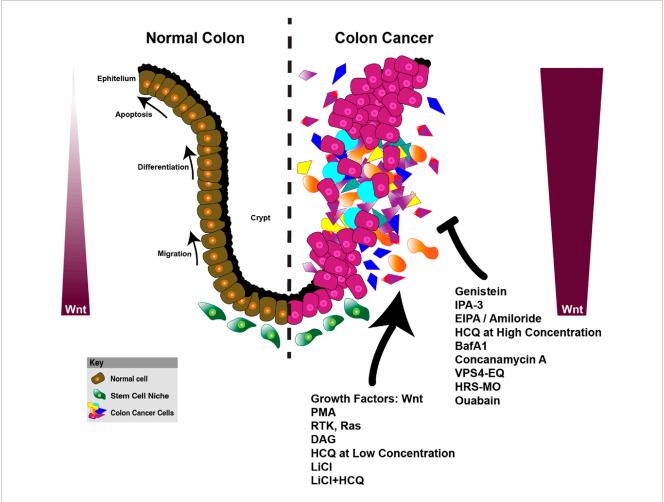
Several metabolic enzymes and pathways have emerged as therapeutic vulnerabilities in Wnt-driven tumors, including lactate dehydrogenase A (LDHA), pyruvate dehydrogenase kinase 1 (PDK1), monocarboxylate transporters (MCTs), glutaminase (GLS), and key regulators of macropinocytosis. Given prior evidence that glycolysis inhibition impairs tumor growth and that IDO1 blockade restores anti-tumor immunity, combining these strategies presents a promising but underexplored approach. LNPs delivering siRNA against IDO1 (Endo et al., 2019), or liposomes delivering IDO1 inhibitors (Mei et al., 2020), combined with glycolytic inhibitors such as LDHA or PDK1 inhibitors (Cheng et al., 2024; Lucero-Acuña et al., 2014) may synergistically disrupt tumor metabolic plasticity while reducing immune suppression. Similarly, nanocarriers loaded with glutaminase (GLS) inhibitors, like CB-839 liposomes, could block glutaminolysis, a critical compensatory pathway in highly glycolytic Wnt-driven tumors (Ni et al., 2023; Shen et al., 2024). Since glutamine metabolism supports nucleotide biosynthesis and redox balance, dual nanoparticle-based inhibition of IDO1 and glutaminolysis may prevent tumors from escaping metabolic stress, thereby enhancing anti-tumor efficacy. Beyond targeting glycolysis and glutaminolysis, macropinocytosis inhibition presents an emerging therapeutic strategy in Wntdriven tumors that rely on nutrient scavenging (Figure 6). Small molecule macropinocytosis inhibitors, such as EIPA (5-(N-Ethyl-N-isopropyl)amiloride) and Rac1/mTORC2 inhibitors, impair the internalization and lysosomal degradation of extracellular proteins, effectively starving tumors that rely on this process (Son et al., 2023). Targeted delivery of macropinocytosis inhibitors in combination with metabolic disruptors could limit both exogenous nutrient uptake and internal metabolic pathways, cutting off key survival mechanisms for tumors. Interestingly, macropinocytosis can also be leveraged as a drug delivery mechanism to enhance therapeutic uptake into tumor cells. Since Wnt-driven tumors exhibit high macropinocytic activity, nanoparticles engineered to mimic extracellular proteins, also known as protein corona (Mei et al., 2018; Li et al., 2024; Mei et al., 2021), or nutrients could facilitate preferential uptake by tumor cells, thereby improving drug bioavailability and selectivity. Albumin-coated nanoparticles could exploit tumor macropinocytosis (Cullis et al., 2017), while PEGylated nanoparticles may evade immune clearance while still being efficiently endocytosed by macropinocytic vesicles (Guo et al., 2023). Charge-modified polymeric nanoparticles can further enhance intracellular retention following micropinocytosis (Means et al., 2022). By hijacking macropinocytosis for selective drug delivery (Jiang et al., 2023; Hyun et al., 2018), nanoparticle formulations could achieve wider tumor accumulation, improve intracellular drug bioavailability, and reduce systemic toxicity. This approach is particularly advantageous for delivering metabolic inhibitors (e.g., LDHA, GLS, or IDO1 blockers) that require efficient cytosolic access to exert their effects.

In addition to nanoparticle-based systems, biomaterial scaffolds and hydrogels offer localized metabolic reprogramming within tumors. Biodegradable hydrogels embedded with IDO1 inhibitors (Zhuang et al., 2023), kynureninase (Wang et al., 2022a), Wnt or GSK-3β antagonists (Wang et al., 2022b), glycolytic inhibitors (LDHA, PDK1) (Meng et al., 2022), and glutaminolysis inhibitors (GLS) (Guo et al., 2025) could provide sustained drug release which gradually alters the tumor microenvironment and preventing metabolic adaptation over time. Stimulus-responsive biomaterials, such as pH- or redox-sensitive hydrogels, can ensure that drug release occurs only in metabolically active, highly glycolytic tumor environments, thereby minimizing off-target effects while maximizing therapeutic efficacy. Another promising approach is exosome-based drug delivery, which offers tumor-specific targeting while minimizing immune clearance. Engineered exosomes can be loaded with siRNA (Amiri et al., 2022), miRNA (Mathiyalagan and Sahoo, 2017), metabolic inhibitors, or Wnt-targeting molecules (e.g., PRR7, $14-3-3\zeta$) (Lee et al., 2018; Zhang et al., 2016), allowing for precise targeting of Wnt-driven tumors. Unlike synthetic nanoparticles, exosomes have natural biocompatibility and immune-modulating properties, making them attractive candidates for disrupting both metabolic plasticity and immune suppression.

Given the metabolic adaptability of Wnt-driven tumors, single-agent metabolic inhibitors are unlikely to yield durable responses. The integration of nanomedicine, biomaterials, and exosome-based drug delivery presents a promising avenue for multi-targeted tumor reprogramming. Future research should focus on optimizing nanoparticle design, engineering exosomes for metabolic inhibition, and refining biomaterial-based delivery platforms to enhance drug selectivity, reduce systemic toxicity, and improve tumor-specific accumulation. Additionally, identifying metabolic biomarkers that predict treatment response could facilitate precision-targeted therapies that consider both tumor metabolism and immune modulation. With continued advancements in precision medicine, the integration of immuno-metabolic reprogramming with cutting-edge drug delivery may pave the way for more effective, durable, and personalized cancer therapies.

Conclusion and future perspectives

Wnt-driven cancers exhibit metabolic plasticity that enables their survival in nutrient-deprived conditions while evading immune surveillance. These tumors rely on aerobic glycolysis (the Warburg effect), glutaminolysis, and macropinocytosis to meet their biosynthetic demands and energy requirements. Key metabolic targets include LDHA, PDK1, HK2, GLUT1, and MCT1, which regulate glycolysis and lactate metabolism; GLS, SLC1A5, and SLC7A5, which drive glutaminolysis and glutamine-dependent biosynthesis; as well as Rac1 and mTORC2, which enhance macropinocytosis and extracellular protein scavenging. Additionally, the Wnt5a-IDO1 axis promotes immune evasion by upregulating tryptophan catabolism through IDO1, resulting



What signaling and its role in colon cancer progression, macropinocytosis, and metabolic reprogramming. This figure compares the organization of the normal colon epithelium and the dysregulated colon cancer microenvironment, highlighting the impact of What signaling on cellular processes. In the normal colon (left), What signaling regulates key biological processes such as apoptosis, differentiation, and migration within the epithelial layer, contributing to homeostasis. The normal stem cell niche is tightly controlled, ensuring proper cellular turnover and balance. On the right, the colon cancer model illustrates the disruption of these processes, with enhanced proliferation, migration, and altered metabolic activity in cancerous cells. What signaling in cancer cells is upregulated, promoting macropinocytosis (phorbol 12-myristate 13-acetate (PMA), RTK, Ras, DAG (diacylglycerol), HCQ at low concentration, LiCl, LiCl+HCQ)) and metabolic reprogramming to support tumor growth. Various compounds targeting What signaling, macropinocytosis (EIPA, Amilloride, Ouabain, Genistein, IPA-3), lysosomes (HCQ at high concentration, Bafilomycin A1, VPS4-EQ (VPS4 ATPase-defective mutant), and hepatocyte growth factor-regulated tyrosine kinase substrate-morpholino oligomers (HRS-MO) are shown as therapeutic interventions that may alter cancer progression. The key shows the differentiation of normal cells (brown), stem cell niche (green), and colon cancer cells (pink and purple), reflecting the complex interplay of signaling pathways and metabolic alterations in cancer progression (Tejeda-Muñoz et al., 2024; Albrecht et al., 2020; Tejeda-Muñoz et al., 2022; Azbazdar et al., 2023; Tejeda-Muñoz et al., 2022b; Tejeda-Muñoz et al., 2022c; Azbazdar et al., 2024).

in kynurenine accumulation and T-cell suppression. Given the compensatory nature of these metabolic pathways, single-agent therapies often fail due to metabolic rewiring. A multipronged therapeutic strategy that simultaneously targets glycolysis, glutaminolysis, macropinocytosis, and immune escape mechanisms may be necessary to achieve durable anti-tumor effects. Several metabolic inhibitors have shown promise in preclinical and early-phase clinical studies. Glycolysis inhibitors such as LDHA inhibitors (FX11), PDK1 inhibitors (DCA, AZD7545), and HK2 inhibitors (3-BrPA) have demonstrated efficacy in limiting glucose metabolism and reducing tumor growth. Glutaminolysis inhibitors, including GLS1 inhibitors (CB-839/Telaglenastat, BPTES, DON derivatives), have been investigated for their ability to impair

glutamine-dependent tumor metabolism. Macropinocytosis inhibitors such as EIPA (5-(N-Ethyl-N-isopropyl)amiloride), Rac1 inhibitors (EHT 1864), and mTORC2 inhibitors (PP242, Torin2) present novel approaches to starve tumor cells by blocking their nutrient scavenging mechanisms. Given the immunosuppressive role of IDO1, IDO1 inhibitors (Epacadostat, Navoximod, Indoximod) are being explored in combination with immune checkpoint inhibitors (ICIs) such as anti-PD-1 and anti-CTLA-4 therapies to enhance T-cell activation and improve immune responses in Wnt-driven cancers. Advanced therapeutic delivery platforms offer a promising approach to co-delivering metabolic inhibitors and immunotherapies, improving tumor specificity while minimizing systemic toxicity. Lipid nanoparticles (LNPs),

polymeric nanocarriers, and exosomes enable the precise delivery of siRNA-based therapies targeting IDO1, PDK1, GLS, or regulators of macropinocytosis. Albumin-coated nanoparticles and PEGylated liposomes can exploit macropinocytosis as a selective uptake mechanism, enhancing drug accumulation in tumors with high macropinocytic activity. Additionally, biomaterial-based hydrogels and tumor-responsive scaffolds offer localized and sustained drug release, thereby enhancing therapeutic efficacy while minimizing off-target effects. Future research could focus on optimizing drug combinations, refining nanocarrier engineering, and identifying metabolic biomarkers to guide the development of precisiontargeted therapies. Given the complex interplay between metabolism and immune evasion, combining metabolic inhibitors with immune checkpoint blockade (ICB), IDO1 inhibitors, or Wnt antagonists may yield synergistic therapeutic effects. Stimuli-responsive biomaterials, such as pH- or redox-sensitive hydrogels, could enable tumor-specific drug activation, further enhancing selectivity and minimizing toxicity. Ultimately, integrating metabolic targeting, immune modulation, and advanced drug delivery technologies may pave the way for more effective, durable, and personalized treatment strategies for Wnt-driven cancers. By leveraging these innovations, it may be possible to overcome metabolic adaptability, restore antitumor immunity, and improve clinical outcomes in aggressive and therapy-resistant malignancies.

Author contributions

EF-H: Writing – original draft. GB: Writing – original draft. K-CM: Writing – original draft, Writing – review and editing. NT-M: Writing – original draft, Writing – review and editing.

Funding

The author(s) declare that no financial support was received for the research and/or publication of this article.

References

Adamska, M., Larroux, C., Adamski, M., Green, K., Lovas, E., Koop, D., et al. (2010). Structure and expression of conserved Wnt pathway components in the demosponge Amphimedon queenslandica: Wnt pathway components in Amphimedon queenslandica. *Evol. Dev.* 12 (5), 494–518. doi:10.1111/j.1525-142X.2010.00435.x

Adler, P. N., and Wallingford, J. B. (2017). From planar cell polarity to ciliogenesis and back: the curious tale of the PPE and CPLANE proteins. *Trends Cell Biol.* 27, 379–390. doi:10.1016/j.tcb.2016.12.001

Albrecht, L. V., Tejeda-Muñoz, N., Bui, M. H., Cicchetto, A. C., Di Biagio, D., Colozza, G., et al. (2020). GSK3 inhibits macropinocytosis and lysosomal activity through the Wnt destruction complex machinery. *Cell Rep.* 32 (4), 107973. doi:10.1016/j.celrep.2020.107973

Amiri, A., Bagherifar, R., Ansari Dezfouli, E., Kiaie, S. H., Jafari, R., and Ramezani, R. (2022). Exosomes as bio-inspired nanocarriers for RNA delivery: preparation and applications. *J. Transl. Med.* 20 (1), 125. doi:10.1186/s12967-022-03325-7

Amyere, M., Mettlen, M., Van Der Smissen, P., Platek, A., Payrastre, B., Veithen, A., et al. (2002). Origin, originality, functions, subversions and molecular signalling of macropinocytosis. *Int. J. Med. Microbiol.* 291 (6-7), 487–494. doi:10.1078/1438-4221-00157

Anastas, J. N., and Moon, R. T. (2013). WNT signalling pathways as therapeutic targets in cancer. *Nat. Rev. Cancer* 13, 11–26. doi:10.1038/nrc3419

Arnovitz, S., Mathur, P., Tracy, M., Mohsin, A., Mondal, S., Quandt, J., et al. (2022). Tcf-1 promotes genomic instability and T cell transformation in response to

Acknowledgments

We are grateful to Dewi Sugiharto for her critical reading of the manuscript. We are thankful for funding from the University of Oklahoma and the Stephenson Cancer Center, as well as from SUNY Binghamton and the University of Utah Faculty Startup Fund.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

aberrant β -catenin activation. *Proc. Natl. Acad. Sci. U. S. A.* 119 (32), e2201493119. doi:10.1073/pnas.2201493119

Augustin, I., Dewi, D. L., Hundshammer, J., Erdmann, G., Kerr, G., and Boutros, M. (2017). Autocrine Wnt regulates the survival and genomic stability of embryonic stem cells. *Sci. Signal* 10 (461), eaah6829. doi:10.1126/scisignal.aah6829

Azbazdar, Y., Tejeda-Munoz, N., Monka, J. C., Dayrit, A., Binder, G., Ozhan, G., et al. (2023). Addition of exogenous diacylglycerol enhances Wnt/β-catenin signaling through stimulation of macropinocytosis. *iScience* 26, 108075. doi:10.1016/j.isci.2023.108075

Azbazdar, Y., Sosa, E. A., Monka, J., Kurmangaliyev, Y. Z., and Tejeda-Muñoz, N. (2024). Interactions between genistein and Wnt pathway in colon adenocarcinoma and early embryos. *Heliyon* 10 (11), e32243. doi:10.1016/j.heliyon.2024. e32243

Bagchi, D. P., and MacDougald, O. A. (2021). Wnt signaling: from mesenchymal cell fate to lipogenesis and other mature adipocyte functions. Diabetes~70~(7),~1419-1430.~doi:10.2337/dbi20-0015

Bao, P., Li, P., Zhou, X., Zhang, H., You, S., Xu, Z., et al. (2023). SMAR1 inhibits proliferation, EMT and Warburg effect of bladder cancer cells by suppressing the activity of the Wnt/ β -catenin signaling pathway. *Cell Cycle* 22 (2), 229–241. doi:10.1080/15384101.2022.2112006

Barrott, J. J., Cash, G. M., Smith, A. P., Barrow, J. R., and Murtaugh, L. C. (2011). Deletion of mouse Porcn blocks Wnt ligand secretion and reveals an

ectodermal etiology of human focal dermal hypoplasia/goltz syndrome. PNAS 108 (31), 12752–12757. doi:10.1073/pnas.1006437108

- Baumann, F., Leukel, P., Doerfelt, A., Beier, C. P., Dettmer, K., Oefner, P. J., et al. (2009). Lactate promotes glioma migration by TGF-Beta2-dependent regulation of matrix metalloproteinase-2. *Neuro-Oncology* 11 (4), 368–380. doi:10.1215/15228517-2008-106
- Bernitt, E., Koh, C. G., Gov, N., and Dobereiner, H. G. (2015). Dynamics of actin waves on patterned substrates: a quantitative analysis of circular dorsal ruffles. *PLoS One* 10 (1), e0115857. doi:10.1371/journal.pone.0115857
- Bi, Y., Zhang, L., Song, Y., Sun, L., Mulholland, M. W., Yin, Y., et al. (2024). Rspo2-LGR4 exacerbates hepatocellular carcinoma progression via activation of Wnt/ β -catenin signaling pathway. *Gastroenterol. Hepatol.* 47 (4), 352–365. doi:10.1016/j.gastrohep.2023.05.016
- Biechele, S., Cox, B. J., and Rossant, J. (2011). Porcupine homolog is required for canonical Wnt signaling and gastrulation in mouse embryos. *Dev. Biol.* 355 (2), 275–285. doi:10.1016/j.ydbio.2011.04.029
- Bloomfield, G., and Kay, R. R. (2016). Uses and abuses of macropinocytosis. *J. Cell Sci.* 129 (14), 2697-2705. doi:10.1242/jcs.176149
- Bondy-Denomy, J., Pawluk, A., Maxwell, K. L., and Davidson, A. R. (2013). Bacteriophage genes that inactivate the CRISPR/Cas bacterial immune system. *Nature* 493 (7432), 429–432. doi:10.1038/nature11723
- Bosoi, C. M., Capra, V., Allache, R., Trinh, V. Q., De Marco, P., Merello, E., et al. (2011). Identification and characterization of novel rare mutations in the planar cell polarity gene PRICKLE1 in human neural tube defects. *Hum. Mutat.* 32, 1371–1375. doi:10.1002/humu.21589
- Bubna-Litic, M., Charras, G., and Mayor, R. (2024). Tissue mechanics modulate morphogen signalling to induce the head organiser. *Cells Dev.* 2, 203984. doi:10.1016/j.cdev.2024.203984
- Buckley, C. M., and King, J. S. (2017). Drinking problems: mechanisms of macropinosome formation and maturation. *FEBS J.* 284 (22), 3778–3790. doi:10.1111/febs.14115
- Buckley, C. M., Pots, H., Gueho, A., Vines, J. H., Munn, C. J., Phillips, B. A., et al. (2020). Coordinated ras and rac activity shapes macropinocytic cups and enables phagocytosis of geometrically diverse bacteria. *Curr. Biol.* 30 (15), 2912–2926. doi:10.1016/j.cub.2020.05.049
- Cadoret, A., Ovejero, C., Terris, B., Souil, E., Lévy, L., Lamers, W. H., et al. (2002). New targets of beta-catenin signaling in the liver are involved in the glutamine metabolism. Oncogene 21 (54), 8293–8301. doi:10.1038/sj.onc.1206118
- Cai, C. F., Ye, G. D., Shen, D. Y., Zhang, W., Chen, M. L., Chen, X. X., et al. (2018). Chibby suppresses aerobic glycolysis and proliferation of nasopharyngeal carcinoma via the Wnt/β-catenin-Lin28/let7-PDK1 cascade. *J. Exp. Clin. Cancer Res.* 37 (1), 104. doi:10.1186/s13046-018-0769-4
- Cai, J., Chen, Z., Wang, J., Wang, J., Chen, X., Liang, L., et al. (2019). circHECTD1 facilitates glutaminolysis to promote gastric cancer progression by targeting miR-1256 and activating β -catenin/c-Myc signaling. *Cell Death Dis.* 10 (8), 576. doi:10.1038/s41419-019-1814-8
- Canton, J. (2018). Macropinocytosis: new insights into its underappreciated role in innate immune cell surveillance. *Front. Immunol.* 9, 2286. doi:10.3389/fimmu.2018.02286
- Carmon, K. S., and Loose, D. S. (2010). Development of a bioassay for detection of Wnt-binding affinities for individual frizzled receptors. *Anal. Biochem.* 401 (2), 288–294. doi:10.1016/j.ab.2010.03.009
- Cha, P. H., Hwang, J. H., Kwak, D. K., Koh, E., Kim, K. S., and Choi, K. Y. (2021). APC loss induces Warburg effect via increased PKM2 transcription in colorectal cancer. *Br. J. Cancer.* 124 (3), 634–644. doi:10.1038/s41416-020-01118-7
- Chae, W. J., and Bothwell, A. L. M. (2018). Canonical and non-canonical wnt signaling in immune cells. *Trends Immunol.* 39 (10), 830–847. doi:10.1016/j.it.2018.08.006
- Chen, S., Tan, J., and Zhang, A. (2021). The ups, downs and new trends of IDO1 inhibitors. Bioorg Chem. 110, 104815. doi:10.1016/j.bioorg.2021.104815
- Cheng, Q., Shi, X. L., Li, Q. L., Wang, L., and Wang, Z. (2024). Current advances on nanomaterials interfering with lactate metabolism for tumor therapy. *Adv. Sci. (Weinh)* 11 (3), e2305662. doi:10.1002/advs.202305662
- Choi, J., Kim, H., Bae, Y. K., and Cheong, H. (2017). REP1 modulates autophagy and macropinocytosis to enhance cancer cell survival. *Int. J. Mol. Sci.* 18 (9), 1866. doi:10.3390/ijms18091866
- Chua, K., Sim, A. Y. L., Yeo, E. Y. M., Bin Masroni, M. S., Naw, W. W., Leong, S. M., et al. (2023). ETC-159, an upstream Wnt inhibitor, induces tumour necrosis via modulation of angiogenesis in osteosarcoma. *Int. J. Mol. Sci.* 24 (5), 4759. doi:10.3390/ijms24054759
- Coant, N., Ducroc, R., Mkaddem, S., Pedruzzi, E., Guichard, C., Tréton, X., et al. (2010). NADPH oxidase 1 modulates WNT and NOTCH1 signaling to control the fate of proliferative progenitor cells in the colon. *Mol. Cell Biol.* 30 (11), 2636–2650. doi:10.1128/MCB.01194-09

Colozza, G., Jami-Alahmadi, Y., Dsouza, A., Tejeda-Muñoz, N., Albrecht, L. V., Sosa, E. A., et al. (2020). Wnt-inducible Lrp6-APEX2 interacting proteins identify ESCRT machinery and Trk-fused gene as components of the Wnt signaling pathway. *Sci. Rep.* 10, 21555. doi:10.1038/s41598-020-78019-5

- Commisso, C., Davidson, S. M., Soydaner-Azeloglu, R. G., Parker, S. J., Kamphorst, J. J., Hackett, S., et al. (2013). Macropinocytosis of protein is an amino acid supply route in Ras-transformed cells. *Nature* 497, 633–637. doi:10.1038/nature12138
- Coombs, G. S., Yu, J., Canning, C. A., Veltri, C. A., Covey, T. M., Cheong, J. K., et al. (2010). WLS-dependent secretion of WNT3A requires Ser209 acylation and vacuolar acidification. *J. Cell Sci.* 123 (19), 3357–3367. doi:10.1242/jcs.072132
- Cordero, J. B., and Sansom, O. J. (2012). Wnt signalling and its role in stem cell-driven intestinal regeneration and hyperplasia. *Acta Physiol. (Oxf)* 204 (1), 137–143. doi:10.1111/j.1748-1716.2011.02288.x
- Crabtree, G. R., and Olson, E. N. (2002). NFAT signaling: choreographing the social lives of cells. Cell 109, S67–S79. doi:10.1016/s0092-8674(02)00699-2
- Cullis, J., Siolas, D., Avanzi, A., Barui, S., Maitra, A., and Bar-Sagi, D. (2017). Macropinocytosis of nab-paclitaxel drives macrophage activation in pancreatic cancer. *Cancer Immunol. Res.* 5 (3), 182–190. doi:10.1158/2326-6066.CIR-16-0125
- Dang, L., White, D. W., Gross, S., Bennett, B. D., Bittinger, M. A., Driggers, E. M., et al. (2010). Cancer-associated IDH1 mutations produce 2-hydroxyglutarate. *Nature* 465 (7300), 966. doi:10.1038/nature09132
- De, A. (2011). Wnt/Ca2+ signaling pathway: a brief overview. *Acta Biochim. Biophys. Sin. (Shanghai).* 43 (10), 745–756. doi:10.1093/abbs/gmr079
- Dejmek, J., Säfholm, A., Kamp Nielsen, C., Andersson, T., and Leandersson, K. (2006). Wnt-5a/Ca2+-induced NFAT activity is counteracted by Wnt-5a/Yes-Cdc42-casein kinase 1alpha signaling in human mammary epithelial cells. *Mol. Cell Biol.* 26, 6024–6036. doi:10.1128/MCB.02354-05
- DeVito, N. C., Sturdivant, M., Thievanthiran, B., Xiao, C., Plebanek, M. P., Salama, A. K. S., et al. (2021). Pharmacological Wnt ligand inhibition overcomes key tumor-mediated resistance pathways to anti-PD-1 immunotherapy. *Cell Rep.* 35 (5), 109071. doi:10.1016/j.celrep.2021.109071
- Dijksterhuis, J. P., Baljinnyam, B., Stanger, K., Sercan, H. O., Ji, Y., Andres, O., et al. (2015). Systematic mapping of WNT-FZD protein interactions reveals functional selectivity by distinct WNT-FZD pairs. *J. Biol. Chem.* 290 (11), 6789–6798. doi:10.1074/jbc.M114.612648
- Douglass, S. M., Fane, M. E., Sanseviero, E., Ecker, B. L., Kugel, C. H., Behera, R., et al. (2021). Myeloid-derived suppressor cells are a major source of Wnt5A in the melanoma microenvironment and depend on Wnt5A for full suppressive activity. *Cancer Res.* 81 (3), 658–670. doi:10.1158/0008-5472.CAN-20-1238
- Dubey, R. D., Klippstein, R., Wang, J. T., Hodgins, N., Mei, K. C., Sosabowski, J., et al. (2017). Novel hyaluronic acid conjugates for dual nuclear imaging and therapy in CD44-Expressing tumors in mice *in vivo. Nanotheranostics* 1 (1), 59–79. doi:10.7150/ntno.17896
- Edwards, J. G. (1925). Formation of food-cups in amoeba induced by chemicals. *Biol. Bull.* 48, 236-239. doi:10.2307/1536568
- El-Ashmawy, N. E., Al-Ashmawy, G. M., Fakher, H. E., and Khedr, N. F. (2020). The role of WNT/β-catenin signaling pathway and glutamine metabolism in the pathogenesis of CCl4-induced liver fibrosis: repositioning of niclosamide and concerns about lithium. *Cytokine* 136, 155250. doi:10.1016/j.cyto.2020.155250
- El-Sahli, S., Xie, Y., Wang, L., and Liu, S. (2019). Wnt signaling in cancer metabolism and immunity. *Cancers (Basel)* 11 (7), 904. doi:10.3390/cancers11070904
- Endo, R., Nakamura, T., Kawakami, K., Sato, Y., and Harashima, H. (2019). The silencing of indoleamine 2,3-dioxygenase 1 (IDO1) in dendritic cells by siRNA-loaded lipid nanoparticles enhances cell-based cancer immunotherapy. *Sci. Rep.* 9 (1), 11335. doi:10.1038/s41598-019-47799-w
- Esen, E., Chen, J., Karner, C. M., Okunade, A. L., Patterson, B. W., and Long, F. (2013). WNT-LRP5 signaling induces Warburg effect through mTORC2 activation during osteoblast differentiation. *Cell Metab.* 17 (5), 745–755. doi:10.1016/j.cmet.2013.03.017
- Etter, E. L., Mei, K. C., and Nguyen, J. (2021). Delivering more for less: nanosized, minimal-carrier and pharmacoactive drug delivery systems. *Adv. Drug Deliv. Rev.* 179, 113994. doi:10.1016/j.addr.2021.113994
- Fares, H., and Greenwald, I. (2001). Genetic analysis of endocytosis in Caenorhabditis elegans: coelomocyte uptake defective mutants. *Genetics* 159 (1), 133–145. doi:10.1093/genetics/159.1.133
- Farrera-Hernández, A., Marín-Llera, J. C., and Chimal-Monroy, J. (2021). WNT5A-Ca2+-CaN-NFAT signalling plays a permissive role during cartilage differentiation in embryonic chick digit development. *Dev. Biol.* 469, 86–95. doi:10.1016/j.ydbio.2020.10.003
- Fischer, K., Hoffmann, P., Voelkl, S., Meidenbauer, N., Ammer, J., Edinger, M., et al. (2007). Inhibitory effect of tumor cell-derived lactic acid on human T cells. *Blood* 109 (9), 3812–3819. doi:10.1182/blood-2006-07-035972
- Flores-Hernández, E., Velázquez, D. M., Castañeda-Patlán, M. C., Fuentes-García, G., Fonseca-Camarillo, G., Yamamoto-Furusho, J. K., et al. (2020). Canonical and non-canonical Wnt signaling are simultaneously activated by Wnts in colon cancer cells. *Cell Signal* 72, 109636. doi:10.1016/j.cellsig.2020.109636

Frezza, C., and Gottlieb, E. (2009). Mitochondria in cancer: not just innocent bystanders. Semin. Cancer Biol. 19, 4–11. doi:10.1016/j.semcancer.2008.11.008

- Fu, H., Gao, H., Qi, X., Zhao, L., Wu, D., Bai, Y., et al. (2018). Aldolase A promotes proliferation and G1/S transition via the EGFR/MAPK pathway in non-small cell lung cancer. *Cancer Commun. (Lond).* 38 (1), 18. doi:10.1186/s40880-018-0290-3
- Fu, Y., Wei, Y., Zhou, Y., Wu, H., Hong, Y., Long, C., et al. (2021). Wnt5a regulates junctional function of sertoli cells through PCP-mediated effects on mTORC1 and mTORC2. *Endocrinology* 162 (10), bqab149. doi:10.1210/endocr/bqab149
- Fu, H., Li, W., Liu, J., Tang, Q., Weng, Z., Zhu, L., et al. (2024). Ellagic acid inhibits dihydrotestosterone-induced ferroptosis and promotes hair regeneration by activating the Wnt/ β -catenin signaling pathway. *J. Ethnopharmacol.* 330, 118227. doi:10.1016/j.jep.2024.118227
- Ganot, P., Tambutté, E., Caminiti-Segonds, N., Toullec, G., Allemand, D., and Tambutté, S. (2020). Ubiquitous macropinocytosis in anthozoans. $\it Elife$ 9, e50022. doi:10.7554/eLife.50022
- Gao, B. (2012). Wnt regulation of planar cell polarity (PCP). Curr. Top. Dev. Biol. 101, 263–295. doi:10.1016/B978-0-12-394592-1.00008-9
- Gaowa, A., Leangpanich, S., Park, E. J., Kawamoto, E., and Shimaoka, M. (2024). Irisin promotes intestinal epithelial cell proliferation via Wnt/ β -catenin and focal adhesion kinase signaling pathways. *Sci. Rep.* 14 (1), 25702. doi:10.1038/s41598-024-76658-6
- Gatenby, R. A., Gawlinski, E. T., Gmitro, A. F., Kaylor, B., and Gillies, R. J. (2006). Acid-mediated tumor invasion: a multidisciplinary study. *Cancer Res.* 66 (10), 5216–5223. doi:10.1158/0008-5472.CAN-05-4193
- Glinka, C., Dolde, N., Kirsch, Y. L., Huang, O., Kazanskaya, D., Ingelfinger, D., et al. (2011). LGR4 and LGR5 are R-spondin receptors mediating Wnt/ β -catenin and Wnt/PCP signalling. *EMBO Rep.* 12 (10), 1055–1061. doi:10.1038/embor.2011.175
- Goodrich, L. V., and Strutt, D. (2011). Principles of planar polarity in animal development. *Development* 138, 1877–1892. doi:10.1242/dev.054080
- Gottfried, E., Kunz-Schughart, L. A., Ebner, S., Mueller-Klieser, W., Hoves, S., Andreesen, R., et al. (2006). Tumor-derived lactic acid modulates dendritic cell activation and antigen expression. *Blood* 107 (5), 2013–2021. doi:10.1182/blood-2005-05-1795
- Guha, A., Sriram, V., Krishnan, K. S., and Mayor, S. (2003). Shibire mutations reveal distinct dynamin-independent and -dependent endocytic pathways in primary cultures of drosophila hemocytes. *J. Cell Sci.* 116 (Pt 16), 3373–3386. doi:10.1242/jcs.00637
- Guo, Q., Xu, J., Huang, Z., Yao, Q., Chen, F., Liu, H., et al. (2021). ADMA mediates gastric cancer cell migration and invasion via Wnt/β-catenin signaling pathway. *Clin. Transl. Oncol.* 23 (2), 325–334. doi:10.1007/s12094-020-02422-7
- Guo, C., Yuan, H., Wang, Y., Feng, Y., Zhang, Y., Yin, T., et al. (2023). The interplay between PEGylated nanoparticles and blood immune system. *Adv. Drug Deliv. Rev.* 200, 115044. doi:10.1016/j.addr.2023.115044
- Guo, Y., Jiang, T., Liang, S., Wang, A., Li, J., et al. (2025). Immunostimulatory hydrogel with synergistic blockage of glutamine metabolism and chemodynamic therapy for postoperative management of glioblastoma. *Adv. Sci. (Weinh)* 12 (15), e2412507. doi:10.1002/advs.202412507
- Haegel, H., Larue, L., Ohsugi, M., Fedorov, L., Herrenknecht, K., and Kemler, R. (1995). Lack of β -catenin affects mouse development at gastrulation. *Development* 121 (11), 3529–3537. doi:10.1242/dev.121.11.3529
- Hayat, R., Manzoor, M., and Hussain, A. (2022). Wnt signaling pathway: a comprehensive review. *Cell Biol. Int.* 46 (6), 863–877. doi:10.1002/cbin.11797
- He, X., Semenov, M., Tamai, K., and Zeng, X. (2004). LDL receptor-related proteins 5 and 6 in Wnt/beta-catenin signaling: arrows point the way. *Development* 131 (8), 1663–1677. doi:10.1242/dev.01117
- Henderson, D. J., Long, D. A., and Dean, C. H. (2018). Planar cell polarity in organ formation. Curr. Opin. Cell Biol. 55, 96–103. doi:10.1016/j.ceb.2018.06.011
- Hobmayer, B., Rentzsch, F., Kuhn, K., Happel, C. M., von Laue, C. C., Snyder, P., et al. (2000). WNT signalling molecules act in axis formation in the diploblastic metazoan hydra. *Nature* 407 (6801), 186–189. doi:10.1038/35025063
- Holstein, T. W. (2012). The evolution of the Wnt pathway. *Cold Spring Harb. Perspect. Biol.* 4 (7), a007922. doi:10.1101/cshperspect.a007922
- Holstein, T. W., Watanabe, H., and Özbek, S. (2011). Signaling pathways and axis formation in the lower metazoa. *Curr. Top. Dev. Biol.* 97, 137–772. doi:10.1016/B978-0-12-385975-4.00012-7
- Holtzhausen, A., Zhao, F., Evans, K. S., Tsutsui, M., Orabona, C., Tyler, D. S., et al. (2015). Melanoma-derived Wnt5a promotes local dendritic-cell expression of IDO and immunotolerance: opportunities for pharmacologic enhancement of immunotherapy. *Cancer Immunol. Res.* 3 (9), 1082–1095. doi:10.1158/2326-6066.CIR-14-0167
- Hoon, J. L., Wong, W. K., and Koh, C. G. (2012). Functions and regulation of circular dorsal ruffles. $Mol.\ Cell.\ Biol.\ 32$ (21), 4246–4257. doi:10.1128/MCB.00551-12
- Hsieh, J. C., Rattner, A., Smallwood, P. M., and Nathans, J. (1999). Biochemical characterization of Wnt-frizzled interactions using a soluble, biologically active vertebrate Wnt protein. *PNAS* 96 (7), 3546–3551. doi:10.1073/pnas.96.7.3546

Hsu, S. Y., Kudo, M., Chen, T., Nakabayashi, K., Bhalla, A., van der Spek, P. J., et al. (2000). The three subfamilies of leucine-rich repeat-containing G protein-coupled receptors (LGR): identification of LGR6 and LGR7 and the signaling mechanism for LGR7. *Mol. Endocrinol.* 14 (8), 1257–1271. doi:10.1210/mend. 14.8.0510

- Huang, J., Tian, F., Song, Y., Cao, M., Yan, S., Lan, X., et al. (2021). A feedback circuit comprising EHD1 and 14-3-3 ζ sustains β -catenin/c-Myc-mediated aerobic glycolysis and proliferation in non-small cell lung cancer. *Cancer Lett.* 520, 12–25. doi:10.1016/j.canlet.2021.06.023
- Huelsken, J., Vogel, R., Brinkmann, V., Erdmann, B., Birchmeier, C., and Birchmeier, W. (2000). Requirement for β -catenin in anterior-posterior axis formation in mice. *J. Cell Biol.* 148, 567–578. doi:10.1083/jcb.148.3.567
- Hyun, H., Park, J., Willis, K., Park, J. E., Lyle, L. T., Lee, W., et al. (2018). Surface modification of polymer nanoparticles with native albumin for enhancing drug delivery to solid tumors. *Biomaterials* 180, 206–224. doi:10.1016/j.biomaterials.2018. 07 074
- Janda, C. Y., Waghray, D., Levin, A. M., Thomas, C., and Garcia, K. C. (2012). Structural basis of Wnt recognition by frizzled. *Science* 337 (6090), 59–64. doi:10.1126/science.1222879
- Jeon, S. M., Lim, J. S., Park, S. H., and Lee, J. H. (2021). Wnt signaling promotes tumor development in part through phosphofructokinase 1 platelet isoform upregulation. *Oncol. Rep.* 46 (5), 234. doi:10.3892/or.2021.8185
- Jiang, G., Wei, C., Chen, Y., Lyu, Y., Huang, J., Chen, H., et al. (2023). Targeted drug delivery system inspired by macropinocytosis. *J. Control Release* 359, 302–314. doi:10.1016/j.jconrel.2023.06.011
- Jiang, K., Wang, Q., Chen, X. L., Wang, X., Gu, X., Feng, S., et al. (2024). Nanodelivery optimization of IDO1 inhibitors in tumor immunotherapy: challenges and strategies. *Int. J. Nanomedicine* 19, 8847–8882. doi:10.2147/IJN.S458086
- Jochems, C., Fantini, M., Fernando, R. I., Kwilas, A. R., Donahue, R. N., Lepone, L. M., et al. (2016). The IDO1 selective inhibitor epacadostat enhances dendritic cell immunogenicity and lytic ability of tumor antigen-specific T cells. *Oncotarget* 7 (25), 37762–37772. doi:10.18632/oncotarget.9326
- Kamphorst, J. J., Nofal, M., Commisso, C., Hackett, S. R., Lu, W., Grabocka, E., et al. (2015). Human pancreatic cancer tumors are nutrient poor and tumor cells actively scavenge extracellular protein. *Cancer Res.* 75, 544–553. doi:10.1158/0008-5472.CAN-14-2711
- Katoh, M., and Katoh, M. (2022). WNT signaling and cancer stemness. *Essays Biochem.* 66 (4), 319–331. doi:10.1042/EBC20220016
- Kim, J., Gao, P., Liu, Y.-C., Semenza, G. L., and Dang, C. V. (2007). Hypoxia-inducible factor 1 and dysregulated c-Myc cooperatively induce vascular endothelial growth factor and metabolic switches hexokinase 2 and pyruvate dehydrogenase kinase 1. *Mol. Cell. Biol.* 27 (21), 7381–7393. doi:10.1128/MCB.00440-07
- Kim, S. M., Nguyen, T. T., Ravi, A., Kubiniok, P., Finicle, B. T., Jayashankar, V., et al. (2018). PTEN deficiency and AMPK activation promote nutrient scavenging and anabolism in prostate cancer cells. *Cancer Discov.* 8 (7), 866–883. doi:10.1158/2159-8290.CD-17-1215
- Kimmelman, A. C., and White, E. (2017). Autophagy and tumor metabolism. Cell Metab. 25 (5), 1037–1043. doi:10.1016/j.cmet.2017.04.004
- King, B., Araki, J., Palm, W., and Thompson, C. B. (2020). Yap/Taz promote the scavenging of extracellular nutrients through macropinocytosis. *Genes Dev.* 34 (19-20), 1345–1358. doi:10.1101/gad.340661.120
- Kleszcz, R., Paluszczak, J., Belka, M., and Krajka-Kuźniak, V. (2023). PRI-724 and IWP-O1 wnt signaling pathway inhibitors modulate the expression of glycolytic enzymes in tongue cancer cell lines. *Curr. Issues Mol. Biol.* 45 (12), 9579–9592. doi:10.3390/cimb45120599
- Koca, Y., Collu, G. M., and Mlodzik, M. (2022). Wnt-frizzled planar cell polarity signaling in the regulation of cell motility. *Curr. Top. Dev. Biol.* 150, 255–297. doi:10.1016/bs.ctdb.2022.03.006
- Kondoh, H., Lleonart, M. E., Gil, J., Wang, J., Degan, P., Peters, G., et al. (2005). Glycolytic enzymes can modulate cellular life span. *Cancer Res.* 65 (1), 177–185. doi:10.1158/0008-5472.177.65.1
- Kornsuthisopon, C., Photichailert, S., Nowwarote, N., Tompkins, K. A., and Osathanon, T. (2022). Wnt signaling in dental pulp homeostasis and dentin regeneration. *Arch. Oral Biol.* 134, 105322. doi:10.1016/j.archoralbio.2021.105322
- Kriz, V., and Korinek, V. (2018). Wnt, RSPO and hippo signalling in the intestine and intestinal stem cells. Genes~(Basel)~9~(1), 20.~doi:10.3390/genes9010020
- Kühl, M., Sheldahl, L. C., Malbon, C. C., and Moon, R. T. (2000). Ca2+/calmodulin-dependent protein kinase II is stimulated by Wnt and frizzled homologs and promotes ventral cell fates in xenopus. *J. Biol. Chem.* 275, 12701–12711. doi:10.1074/jbc.275.17.12701
- Lai, K. K. Y., Kweon, S. M., Chi, F., Hwang, E., Kabe, Y., Higashiyama, R., et al. (2017). Stearoyl-CoA desaturase promotes liver fibrosis and tumor development in mice via a wnt positive-signaling loop by stabilization of low-density lipoprotein-receptor-related proteins 5 and 6. *Gastroenterology* 152 (6), 1477–1491. doi:10.1053/j.gastro.2017.01.021

Lambies, G., Lee, S. W., Duong-Polk, K., Aza-Blanc, P., Maganti, S., Galapate, C. M., et al. (2024). Cell polarity proteins promote macropinocytosis in response to metabolic stress. *Nat. Commun.* 15 (1), 10541. doi:10.1038/s41467-024-54788-9

- Lee, S. H., Shin, S. M., Zhong, P., Kim, H. T., Kim, D. I., Kim, J. M., et al. (2018). Reciprocal control of excitatory synapse numbers by Wnt and Wnt inhibitor PRR7 secreted on exosomes. *Nat. Commun.* 9 (1), 3434. doi:10.1038/s41467-018-05858-2
- Lee, C. H. R., Mohamed Hussain, K., and Chu, J. J. H. (2019a). Macropinocytosis dependent entry of Chikungunya virus into human muscle cells. *PLoS Negl. Trop. Dis.* 13 (8), e0007610. doi:10.1371/journal.pntd.0007610
- Lee, J. H., Pasquarella, J. R., and Kalejta, R. F. (2019b). Cell line models for human cytomegalovirus latency faithfully mimic viral entry by macropinocytosis and endocytosis. *J. Virol.* 93 (21), e01021-19. doi:10.1128/JVI.01021-19
- Lei, Y., He, L., Li, Y., Hou, J., and Zhang, H. (2024). PDLIM1 interacts with HK2 to promote gastric cancer progression through enhancing the Warburg effect via Wnt/ β -catenin signaling. *Cell Tissue Res.* 395 (1), 105–116. doi:10.1007/s00441-023-03840-z
- Leucht, P., Lee, S., and Yim, N. (2019). Wnt signaling and bone regeneration: can't have one without the other. *Biomaterials* 196, 46–50. doi:10.1016/j.biomaterials.2018.03.029
- Leung, R. W. H., and Lee, T. K. W. (2022). Wnt/ β -Catenin signaling as a driver of stemness and metabolic reprogramming in hepatocellular carcinoma. *Cancers (Basel)* 14 (21), 5468. doi:10.3390/cancers14215468
- Lewis, W. H. (1931). Pinocytosis. Bull. Johns Hopkins Hosp. 49, 17-27.
- Li, B., Cao, Y., Meng, G., Qian, L., Xu, T., Yan, C., et al. (2019). Targeting glutaminase 1 attenuates stemness properties in hepatocellular carcinoma by increasing reactive oxygen species and suppressing Wnt/beta-catenin pathway. *EBioMedicine* 39, 239–254. doi:10.1016/j.ebiom.2018.11.063
- Li, M., Liu, Q., He, S., Kong, X., Lin, J., Huang, Y., et al. (2021). Icaritin inhibits skin fibrosis through regulating AMPK and Wnt/ β -catenin signaling. *Cell Biochem. Biophys.* 79 (2), 231–238. doi:10.1007/s12013-020-00952-z
- Li, D., Sun, J., and Zhong, T. P. (2022). Wnt signaling in heart development and regeneration. Curr. Cardiol. Rep. 24 (10), 1425–1438. doi:10.1007/s11886-022-01756-8
- Li, Y. T., Mei, K. C., Liam-Or, R., Wang, J. T., Faruqu, F. N., Zhu, S., et al. (2024). Graphene oxide nanosheets toxicity in mice is dependent on protein corona composition and host immunity. ACS Nano 18 (33), 22572–22585. doi:10.1021/acsnano.4c08561
- Li, C., Liu, Y., Liu, C., Chen, F., Xie, Y., Zeh, H. J., et al. (2025). AGER-dependent macropinocytosis drives resistance to KRAS-G12D-targeted therapy in advanced pancreatic cancer. *Sci. Transl. Med.* 17 (783), eadp4986. doi:10.1126/scitranslmed.adp4986
- Liang, L., Chen, Y., Yu, Y., Pan, W., Cui, Y., Xu, X., et al. (2020). SLC25A18 has prognostic value in colorectal cancer and represses Warburg effect and cell proliferation via Wnt signaling. *Am. J. Cancer Res.* 10 (5), 1548–1567.
- Liang, Y., Rao, Z., Du, D., Wang, Y., and Fang, T. (2023). Butyrate prevents the migration and invasion, and aerobic glycolysis in gastric cancer via inhibiting Wnt/ β -catenin/c-Myc signaling. *Drug Dev. Res.* 84 (3), 532–541. doi:10.1002/ddr.22043
- Liao, Y. (2017). Cancer metabolism as we know it today: a prologue to a special issue of cancer metabolism. $Genes\ Dis.\ 4\ (1),\ 4-6.\ doi:10.1016/j.gendis.2017.02.001$
- Liu, Z., and Roche, P. A. (2015). Macropinocytosis in phagocytes: regulation of MHC class-II-restricted antigen presentation in dendritic cells. *Front. Physiol.* 6, 1. doi:10.3389/fphys.2015.00001
- Liu, T., and Yin, H. (2017). PDK1 promotes tumor cell proliferation and migration by enhancing the warburg effect in non-small cell lung cancer. *Oncol. Rep.* 37 (1), 193–200. doi:10.3892/or.2016.5253
- Liu, P., Wakamiya, M., Shea, M. J., Albrecht, U., Behringer, R. R., and Bradley, A. (1999). Requirement for Wnt3 in vertebrate axis formation. *Nat. Genet.* 22 (4), 361–365. doi:10.1038/11932
- Liu, X., Jiang, J., Liao, Y. P., Tang, I., Zheng, E., Qiu, W., et al. (2021). Combination chemo-immunotherapy for pancreatic cancer using the immunogenic effects of an Irinotecan silicasome nanocarrier plus Anti-PD-1. *Adv. Sci. (Weinh)* 8 (6), 2002147. doi:10.1002/advs.202002147
- Liu, P., Chen, G., and Zhang, J. (2022a). A review of liposomes as a drug delivery system: current status of approved products, regulatory environments, and future perspectives. *Molecules* 27 (4), 1372. doi:10.3390/molecules27041372
- Liu, Y., Qi, X., Donnelly, L., Elghobashi-Meinhardt, N., Long, T., Zhou, R. W., et al. (2022b). Mechanisms and inhibition of Porcupine-mediated Wnt acylation. *Nature* 607 (7920), 816–822. doi:10.1038/s41586-022-04952-2
- Liu, Y., Lei, P., Samuel, R. Z., Kashyap, A. M., Groth, T., Bshara, W., et al. (2023). Cadherin-11 increases tumor cell proliferation and metastatic potential via Wnt pathway activation. *Mol. Oncol.* 17 (10), 2056–2073. doi:10.1002/1878-0261.13507
- Liu, H. Y., Sun, X. J., Xiu, S. Y., Zhang, X. Y., Wang, Z. Q., Gu, Y. L., et al. (2024). Frizzled receptors (FZDs) in Wnt signaling: potential therapeutic targets for human cancers. *Acta Pharmacol. Sin.* 45 (8), 1556–1570. doi:10.1038/s41401-024-01270-3

Lo, Y. L., Wang, T. Y., Chen, C. J., Chang, Y. H., and Lin, A. M. (2022). Two-in-One nanoparticle formulation to deliver a tyrosine kinase inhibitor and microRNA for targeting metabolic reprogramming and mitochondrial dysfunction in gastric cancer. *Pharmaceutics* 14 (9), 1759. doi:10.3390/pharmaceutics14091759

- Loh, K. M., van Amerongen, R., and Nusse, R. (2016). Generating cellular diversity and spatial form: Wnt signaling and the evolution of multicellular animals. *Dev. Cell* 38 (6), 643–655. doi:10.1016/j.devcel.2016.08.011
- Lou, Y. X., Gu, J., Zhu, L., Sun, S. Q., Hao, X. L., Chen, J. P., et al. (2023). TC2N promotes cell proliferation and metastasis in hepatocellular carcinoma by targeting the Wnt/ β -Catenin signaling pathway. *Lab. Invest* 103 (12), 100260. doi:10.1016/j.labinv.2023.100260
- Lu, W., Pelicano, H., and Huang, P. (2010). Cancer metabolism: is glutamine sweeter than glucose? Cancer Cell 18, 199–200. doi:10.1016/j.ccr.2010.08.017
- Lu, L., Wang, H., Liu, X., Tan, L., Qiao, X., Ni, J., et al. (2021). Pyruvate kinase isoform M2 impairs cognition in systemic lupus erythematosus by promoting microglial synaptic pruning via the β -catenin signaling pathway. *J. Neuroinflammation* 18 (1), 229. doi:10.1186/s12974-021-02279-9
- Lucero-Acuña, A., Jeffery, J. J., Abril, E. R., Nagle, R. B., Guzman, R., Pagel, M. D., et al. (2014). Nanoparticle delivery of an AKT/PDK1 inhibitor improves the therapeutic effect in pancreatic cancer. *Int. J. Nanomedicine* 9, 5653–5665. doi:10.2147/IJN.S68511
- Lum, J. J., Bui, T., Gruber, M., Gordan, J. D., DeBerardinis, R. J., Covello, K. L., et al. (2007). The transcription factor HIF-1alpha plays a critical role in the growth factor-dependent regulation of both aerobic and anaerobic glycolysis. *Genes Dev.* 21 (9), 1037–1049. doi:10.1101/gad.1529107
- Ma, Q., Yu, J., Zhang, X., Wu, X., and Deng, G. (2023). Wnt/ β -catenin signaling pathway-a versatile player in apoptosis and autophagy. *Biochimie* 211, 57–67. doi:10.1016/j.biochi.2023.03.001
- Madsen, J. F., Ernst, E. H., Amoushahi, M., Dueholm, M., Ernst, E., and Lykke-Hartmann, K. (2024). Dorsomorphin inhibits AMPK, upregulates Wnt and Foxo genes and promotes the activation of dormant follicles. *Commun. Biol.* 7 (1), 747. doi:10.1038/s42003-024-06418-9
- Malki, A., ElRuz, R. A., Gupta, I., Allouch, A., Vranic, S., and Al Moustafa, A. E. (2020). Molecular mechanisms of Colon cancer progression and metastasis: recent insights and advancements. *Int. J. Mol. Sci.* 22 (1), 130. doi:10.3390/ijms22010130
- Malladi, S., Macalinao, D. G., Jin, X., He, L., Basnet, H., Zou, Y., et al. (2016). Metastatic latency and immune evasion through autocrine inhibition of WNT. *Cell* 165 (1), 45–60. doi:10.1016/j.cell.2016.02.025
- Martínez-Reyes, I., and Chandel, N. S. (2021). Cancer metabolism: looking forward. Nat. Rev. Cancer 21 (10), 669–680. doi:10.1038/s41568-021-00378-6
- Masiakowski, P., and Carroll, R. D. (1992). A novel family of cell surface receptors with tyrosine kinase-like domain. *J. Biol. Chem.* 267 (36), 26181–26190. doi:10.1016/s0021-9258(18)35733-8
- Masiakowski, P., and Yancopoulos, G. D. (1998). The wnt receptor CRD domain is also found in MuSK and related orphan receptor tyrosine kinases. *Curr. Biol.* 8, R407. doi:10.1016/s0960-9822(98)70263-5
- Mathiyalagan, P., and Sahoo, S. (2017). Exosomes-based gene therapy for MicroRNA delivery. $Methods\ Mol.\ Biol.\ 1521,\ 139-152.\ doi:10.1007/978-1-4939-6588-5_9$
- Matsuda, M., and Sokol, S. Y. (2021). Xenopus neural tube closure: a vertebrate model linking planar cell polarity to actomyosin contractions. Curr. Top. Dev. Biol. 145, 41–60. doi:10.1016/bs.ctdb.2021.04.001
- McMahon, A. P., and Moon, R. T. (1989). Ectopic expression of the proto-oncogene int-1 in xenopus embryos leads to duplication of the embryonic axis. *Cell* 58 (6), 1075–1084. doi:10.1016/0092-8674(89)90506-0
- Means, N., Elechalawar, C. K., Chen, W. R., Bhattacharya, R., and Mukherjee, P. (2022). Revealing macropinocytosis using nanoparticles. *Mol. Asp. Med.* 83, 100993. doi:10.1016/j.mam.2021.100993
- Medina, M. A. (2001). Glutamine and cancer. J. Nutr. 131, 2539S-2541S. doi:10.1093/jn/131.9.2539S
- Mei, K. C., Bai, J., Lorrio, S., Wang, J. T., and Al-Jamal, K. T. (2016). Investigating the effect of tumor vascularization on magnetic targeting $in\ vivo$ using retrospective design of experiment. $Biomaterials\ 106,\ 276-285.$ doi:10.1016/j.biomaterials.2016.08.030
- Mei, K. C., Ghazaryan, A., Teoh, E. Z., Summers, H. D., Li, Y., Ballesteros, B., et al. (2018). Protein-Corona-by-Design in 2D: a reliable platform to decode bio-nano interactions for the next-generation quality-by-design nanomedicines. *Adv. Mater* 30, e1802732. doi:10.1002/adma.201802732
- Mei, K. C., Liao, Y. P., Jiang, J., Chiang, M., Khazaieli, M., Liu, X., et al. (2020). Liposomal delivery of mitoxantrone and a cholesteryl Indoximod Prodrug provides effective chemo-immunotherapy in multiple solid tumors. *ACS Nano* 14 (10), 13343–13366. doi:10.1021/acsnano.0c05194
- Mei, K. C., Costa, P. M., Kreuzer, M., and Al-Jamal, K. T. (2021). Interpreting 2D materials bio-nano interactions: influence of aggregation status, protein Corona, cell culture media, and cell types. *Adv. Mater. Interfaces.* 8 (17), 2100251. doi:10.1002/admi.202100251

Mei, K. C., Stiepel, R. T., Bonacquisti, E., Jasiewicz, N. E., Chaudhari, A. P., Tiwade, P. B., et al. (2023). Single-tailed heterocyclic carboxamide lipids for macrophage immune-modulation. *Biomater. Sci.* 11 (8), 2693–2698. doi:10.1039/d2bm01804g

- Mei, K. C., Thota, N., Wei, P. S., Yi, B., Bonacquisti, E. E., and Nguyen, J. (2024). Calreticulin P-domain-derived "Eat-me" peptides for enhancing liposomal uptake in dendritic cells. *Int. J. Pharm.* 653, 123844. doi:10.1016/j.ijpharm.2024.123844
- Menck, K., Heinrichs, S., Baden, C., and Bleckmann, A. (2021). The WNT/ROR pathway in cancer: from signaling to the rapeutic intervention. $Cells\ 10\ (1),\ 142.$ doi:10.3390/cells 10010142
- Meng, X., Lu, Z., Lv, Q., Jiang, Y., Zhang, L., and Wang, Z. (2022). Tumor metabolism destruction via metformin-based glycolysis inhibition and glucose oxidase-mediated glucose deprivation for enhanced cancer therapy. *Acta Biomater.* 145, 222–234. doi:10.1016/j.actbio.2022.04.022
- Mercer, J., and Helenius, A. (2009). Virus entry by macropinocytosis. *Nat. Cell Biol.* 11 (5), 510–520. doi:10.1038/ncb0509-510
- Miao, L., Li, L., Huang, Y., Delcassian, D., Chahal, J., Han, J., et al. (2019). Delivery of mRNA vaccines with heterocyclic lipids increases anti-tumor efficacy by STING-mediated immune cell activation. *Nat. Biotechnol.* 37 (10), 1174–1185. doi:10.1038/s41587-019-0247-3
- Mohammed, M. K., Shao, C., Wang, J., Wei, Q., Wang, X., Collier, Z., et al. (2016). Wnt/ β -catenin signaling plays an ever-expanding role in stem cell self-renewal, tumorigenesis and cancer chemoresistance. *Genes Dis.* 3 (1), 11–40. doi:10.1016/j.gendis.2015.12.004
- Moon, R. T., Kohn, A. D., De Ferrari, G. V., and Kaykas, A. (2004). WNT and beta-catenin signalling: diseases and therapies. *Nat. Rev. Genet.* 5 (9), 691–701. doi:10.1038/nrg1427
- Motono, M., Ioroi, Y., Ogura, T., and Takahashi, J. (2016). WNT-C59, a small-molecule WNT inhibitor, efficiently induces anterior cortex that includes cortical motor neurons from human pluripotent stem cells. *Stem Cells Transl. Med.* 5 (4), 552–560. doi:10.5966/sctm.2015-0261
- Muller, A. J., Manfredi, M. G., Zakharia, Y., and Prendergast, G. C. (2019). Inhibiting IDO pathways to treat cancer: lessons from the ECHO-301 trial and beyond. *Semin. Immunopathol.* 41 (1), 41–48. doi:10.1007/s00281-018-0702-0
- Nath, A., Li, I., Roberts, L. R., and Chan, C. (2015). Elevated free fatty acid uptake via CD36 promotes epithelial-mesenchymal transition in hepatocellular carcinoma. *Sci. Rep.* 5, 14752. doi:10.1038/srep14752
- Nel, A. E., Mei, K. C., Liao, Y. P., and Liu, X. (2022). Multifunctional lipid bilayer nanocarriers for cancer immunotherapy in heterogeneous tumor microenvironments, combining immunogenic cell death stimuli with immune modulatory drugs. *ACS Nano* 16 (4), 5184–5232. doi:10.1021/acsnano.2c01252
- Ni, R., Li, Z., Li, L., Peng, D., Ming, Y., et al. (2023). Rethinking glutamine metabolism and the regulation of glutamine addiction by oncogenes in cancer. *Front. Oncol.* 13, 1143798. doi:10.3389/fonc.2023.1143798
- Nusse, R., and Varmus, H. E. (1982). Many tumors induced by the mouse mammary tumor virus contain a provirus integrated in the same region of the host genome. *Cell* $31\ (1)$, 99-109. doi:10.1016/0092-8674(82)90409-3
- Nusse, R., and Varmus, H. (2012). Three decades of wnts: a personal perspective on how a scientific field developed. *EMBO J.* 31 (12), 2670–2684. doi:10.1038/emboj.2012.146
- Odunsi, K., Qian, F., Lugade, A. A., Yu, H., Geller, M. A., Fling, S. P., et al. (2022). Metabolic adaptation of ovarian tumors in patients treated with an IDO1 inhibitor constrains antitumor immune responses. *Sci. Transl. Med.* 14 (636), eabg8402. doi:10.1126/scitranslmed.abg8402
- Overmeyer, J. H., Kaul, A., Johnson, E. E., and Maltese, W. A. (2008). Active ras triggers death in glioblastoma cells through hyperstimulation of macropinocytosis. *Mol. Cancer Res.* 6, 965–977. doi:10.1158/1541-7786.MCR-07-2036
- Palm, W., Park, Y., Wright, K., Pavlova, N. N., Tuveson, D. A., and Thompson, C. B. (2015). The utilization of extracellular proteins as Nutrients is suppressed by mTORC1. *Cell.* 162, 259–270. doi:10.1016/j.cell.2015.06.017
- Park, S. Y., Lee, Y. K., Kim, H. J., Park, O. J., and Kim, Y. M. (2016). AMPK interacts with β -catenin in the regulation of hepatocellular carcinoma cell proliferation and survival with selenium treatment. *Oncol. Rep.* 35 (3), 1566–1572. doi:10.3892/or.2015.4519
- Park, S. J., Kim, J. H., Oh, S., and Lee, D. Y. (2023). Metabolome-wide reprogramming modulated by Wnt/ β -Catenin signaling pathway. *J. Microbiol. Biotechnol.* 33 (1), 114–122. doi:10.4014/jmb.2211.11013
- Pate, K. T., Stringari, C., Sprowl-Tanio, S., Wang, K., TeSlaa, T., Hoverter, N. P., et al. (2014). Wnt signaling directs a metabolic program of glycolysis and angiogenesis in colon cancer. *EMBO J.* 33 (13), 1454–1473. doi:10.15252/embj. 201488598
- Penny, M. K., Lerario, A. M., Basham, K. J., Chukkapalli, S., Mohan, D. R., LaPensee, C., et al. (2023). Targeting oncogenic Wnt/ β -Catenin signaling in adrenocortical carcinoma disrupts ECM expression and impairs tumor growth. *Cancers (Basel)* 15 (14), 3559. doi:10.3390/cancers15143559

Pham, S. H., Pratt, K., Okolicsanyi, R. K., Oikari, L. E., Yu, C., Peall, I. W., et al. (2022). Syndecan-1 and -4 influence Wnt signaling and cell migration in human breast cancers. *Biochimie* 198, 60–75. doi:10.1016/j.biochi.2022.01.014

- Prendergast, G. C., Malachowski, W. P., DuHadaway, J. B., and Muller, A. J. (2017). Discovery of IDO1 inhibitors: from bench to bedside. *Cancer Res.* 77 (24), 6795–6811. doi:10.1158/0008-5472.CAN-17-2285
- Qian, Y., Wang, X., Liu, Y., Li, Y., Colvin, R. A., Tong, L., et al. (2014). Extracellular ATP is internalized by macropinocytosis and induces intracellular ATP increase and drug resistance in cancer cells. *Cancer Lett.* 351, 242–251. doi:10.1016/j.canlet.2014.06.008
- Qin, K., Yu, M., Fan, J., Wang, H., Zhao, P., et al. (2023). Canonical and noncanonical Wnt signaling: multilayered mediators, signaling mechanisms and major signaling crosstalk. *Genes Dis.* 11 (1), 103–134. doi:10.1016/j.gendis.2023.01.030
- Rainero, E. (2024). Macropinocytosis at the crossroad between nutrient scavenging and metabolism in cancer. *Curr. Opin. Cell Biol.* 88, 102359. doi:10.1016/j.ceb.2024.102359
- Raslan, A. A., and Yoon, J. K. (2020). WNT signaling in lung repair and regeneration. *Mol. Cells* 43 (9), 774–783. doi:10.14348/molcells.2020.0059
- Redelman-Sidi, G., Binyamin, A., Gaeta, I., Palm, W., Thompson, C. B., Romesser, P. B., et al. (2018). The canonical wnt pathway drives macropinocytosis in cancer. *Cancer Res.* 78 (16), 4658–4670. doi:10.1158/0008-5472.CAN-17-3199
- Reitzer, L. J., Wice, B. M., and Kennell, D. (1979). Evidence that glutamine, not sugar, is the major energy source for cultured HeLa cells. *J. Biol. Chem.* 254, 2669–2676. doi:10.1016/s0021-9258(17)30124-2
- Richards, T., Modarage, K., Dean, C., McCarthy-Boxer, A., Hilton, H., Esapa, C., et al. (2019). Atmin modulates Pkhd1 expression and may mediate Autosomal Recessive Polycystic Kidney Disease (ARPKD) through altered non-canonical Wnt/Planar Cell Polarity (PCP) signalling. *Biochim. Biophys. Acta Mol. Basis Dis.* 1865 (2), 378–390. doi:10.1016/j.bbadis.2018.11.003
- Rim, E. Y., Clevers, H., and Nusse, R. (2022). The wnt pathway: from signaling mechanisms to synthetic modulators. *Annu. Rev. Biochem.* 91, 571–598. doi:10.1146/annurev-biochem-040320-103615
- Roche, T. E., Baker, J. C., Yan, X., Hiromasa, Y., Gong, X., Peng, T., et al. (2001). Distinct regulatory properties of pyruvate dehydrogenase kinase and phosphatase isoforms. *Progr. Nucleic Acid. Res. Mol. Biol.* 70, 33–75. doi:10.1016/s0079-6603(01)70013-x
- Rong, Z., Xu, J., Yang, J., Wang, W., Tang, R., Zhang, Z., et al. (2024). CircRREB1 mediates metabolic reprogramming and stemness maintenance to facilitate pancreatic ductal adenocarcinoma progression. *Cancer Res.* 84 (24), 4246–4263. doi:10.1158/0008-5472.CAN-23-3596
- Saldanha, J., Singh, J., and Mahadevan, D. (1998). Identification of a frizzled like cysteine rich domain in the extracellular region of developmental receptor tyrosine kinases. *Protein Sci.* 7 (7), 1632–1635. doi:10.1002/pro.5560070718
- Sallusto, F., Cella, M., Danieli, C., and Lanzavecchia, A. (1995). Dendritic cells use macropinocytosis and the mannose receptor to concentrate macromolecules in the major histocompatibility complex class II compartment: downregulation by cytokines and bacterial products. *J. Exp. Med.* 182 (2), 389–400. doi:10.1084/jem.182.2.389
- Seliger, C., Leukel, P., Moeckel, S., Jachnik, B., Lottaz, C., Kreutz, M., et al. (2013). Lactate-modulated induction of THBS-1 activates transforming growth factor (TGF)-beta2 and migration of glioma cells *in vitro*. *PLoS One* 8 (11), e78935. doi:10.1371/journal.pone.0078935
- Seltzer, M. J., Bennett, B. D., Joshi, A. D., Gao, P., Thomas, A. G., Ferraris, D. V., et al. (2010). Inhibition of glutaminase preferentially slows growth of glioma cells with mutant IDH1. *Cancer Res.* 70 (22), 8981–8987. doi:10.1158/0008-5472.CAN-10-1666
- Shah, S., Dhawan, V., Holm, R., Nagarsenker, M. S., and Perrie, Y. (2020). Liposomes: advancements and innovation in the manufacturing process. *Adv. Drug Deliv. Rev.* 154–155, 102–122. doi:10.1016/j.addr.2020.07.002
- Shen, J., Cao, B., Wang, Y., Ma, C., Zeng, Z., Liu, L., et al. (2018). Hippo component YAP promotes focal adhesion and tumour aggressiveness via transcriptionally activating THBS1/FAK signalling in breast cancer. *J. Exp. Clin. Cancer Res.* 37 (1), 175. doi:10.1186/s13046-018-0850-z
- Shen, L., Sharma, D., Yu, Y., Long, F., and Karner, C. M. (2021). Biphasic regulation of glutamine consumption by WNT during osteoblast differentiation. *J. Cell Sci.* 134 (1), jcs251645. doi:10.1242/jcs.251645
- Shen, H., Wang, H., Mo, J., Zhang, J., Xu, C., Sun, F., et al. (2024). Unrestricted molecular motions enable mild photothermy for recurrence-resistant FLASH antitumor radiotherapy. *Bioact. Mater* 37, 299–312. doi:10.1016/j.bioactmat.2024.03.024
- Shi, D. L. (2022). Wnt/planar cell polarity signaling controls morphogenetic movements of gastrulation and neural tube closure. *Cell Mol. Life Sci.* 79 (12), 586. doi:10.1007/s00018-022-04620-8
- Shi, D. L. (2023). Planar cell polarity regulators in asymmetric organogenesis during development and disease. *J. Genet. Genomics* 50 (2), 63–76. doi:10.1016/j.jgg.2022.06.007

Shi, D. L. (2024). Canonical and non-canonical wnt signaling generates molecular and cellular asymmetries to establish embryonic axes. *J. Dev. Biol.* 12 (3), 20. doi:10.3390/jdb12030020

- Shih, P. C., and Mei, K. C. (2021). Role of STAT3 signaling transduction pathways in cancer stem cell-associated chemoresistance. *Drug Discov. Today* 26 (6), 1450–1458. doi:10.1016/j.drudis.2020.11.032
- Slusarski, D. C., Yang-Snyder, J., Busa, W. B., and Moon, R. T. (1997). Modulation of embryonic intracellular Ca2+ signaling by Wnt-5A. *Dev. Biol.* 182, 114–120. doi:10.1006/dbio.1996.8463
- Smith, M., Newton, R., Owens, S., Gong, X., and Tian, C. (2020). 28 retrospective pooled analysis of epacadostat clinical studies identifies doses required for maximal pharmacodynamic effect in anti-PD-1 combination studies. *J. Immunother. Cancer* 8,
- Son, A., Kang, S., Choi, S., Shin, S. W., Kim, Y., et al. (2023). Targeting Na-H exchanger 1 overcomes nuclear factor kappa B-mediated tumor resistance to radiotherapy. *Neoplasia* 35, 100862. doi:10.1016/j.neo.2022.100862
- Sprowl-Tanio, S., Habowski, A. N., Pate, K. T., McQuade, M. M., Wang, K., Edwards, R. A., et al. (2016). Lactate/Pyruvate transporter MCT-1 is a direct Wnt target that confers sensitivity to 3-bromopyruvate in colon cancer. *Cancer Metab.* 4, 20. doi:10.1186/s40170-016-0159-3
- Strutt, H., and Strutt, D. (2009). Asymmetric localisation of planar polarity proteins: mechanisms and consequences. *Semin. Cell Dev. Biol.* 20 (8), 957–963. doi:10.1016/j.semcdb.2009.03.006
- Sun, G., Wu, L., Sun, G., Shi, X., Cao, H., and Tang, W. (2021a). WNT5a in colorectal cancer: research progress and challenges. *Cancer Manag. Res.* 13, 2483–2498. doi:10.2147/CMAR.S289819
- Sun, H., Wang, T., Atkinson, P. J., Billings, S. E., Dong, W., and Cheng, A. G. (2021b). Gpr125 marks distinct cochlear cell types and is dispensable for cochlear development and hearing. *Front. Cell Dev. Biol.* 28 (9), 690955. doi:10.3389/fcell.2021.690955
- Tanaka, K., Kitagawa, Y., and Kadowaki, T. (2002). Drosophila segment polarity gene product porcupine stimulates the posttranslational N-glycosylation of wingless in the endoplasmic reticulum. *J. Biol. Chem.* 277 (15), 12816–12823. doi:10.1074/jbc.M200187200
- Tejeda-Muñoz, N., and De Robertis, E. M. (2022). Wnt, GSK3, and macropinocytosis. Subcell. Biochem. 98, 169–187. doi:10.1007/978-3-030-94004-1_9
- Tejeda-Muñoz, N., and Mei, K. C. (2024). Wnt signaling in cell adhesion, development, and colon cancer. $IUBMB\ Life.\ 76\ (7),\ 383-396.\ doi:10.1002/iub.2806$
- Tejeda-Muñoz, N., and Robertis, E. M. D. (2022). Lysosomes are required for early dorsal signaling in the *Xenopus* embryo. *Proc. Natl. Acad. Sci. U. S. A.* 119, e2201008119. doi:10.1073/pnas.2201008119
- Tejeda-Muñoz, N., Albrecht, L. V., Bui, M. H., and De Robertis, E. M. (2019). Wnt canonical pathway activates macropinocytosis and lysosomal degradation of extracellular proteins. *Proc. Natl. Acad. Sci. U. S. A.* 116 (21), 10402–10411. doi:10.1073/pnas.1903506116
- Tejeda-Muñoz, N., Mei, K.-C., Sheladiya, P., and Monka, J. (2022a). Targeting membrane trafficking as a strategy for cancer treatment. *Vaccines.* 10, 790. doi:10.3390/vaccines10050790
- Tejeda-Muñoz, N., Monka, J., and De Robertis, E. M. (2022b). Protocol for culturing and imaging of ectodermal cells from Xenopus. *Star. Protoc.* 3, 101455. doi:10.1016/j.xpro.2022.101455
- Tejeda-Muñoz, N., Morselli, M., Moriyama, Y., Sheladiya, P., Pellegrini, M., and De Robertis, E. M. (2022c). Canonical wnt signaling induces focal adhesion and integrin Beta-1 endocytosis. *iScience* 25, 104123. doi:10.1016/j.isci.2022.104123
- Tejeda-Muñoz, N., Binder, G., and Mei, K. C. (2024). Emerging therapeutic strategies for wnt-dependent colon cancer targeting macropinocytosis. *Cells Dev.* 180, 203974. doi:10.1016/j.cdev.2024.203974
- Thaker, A. I., Rao, M. S., Bishnupuri, K. S., Kerr, T. A., Foster, L., Marinshaw, J. M., et al. (2013). IDO1 metabolites activate β -catenin signaling to promote cancer cell proliferation and colon tumorigenesis in mice. *Gastroenterology* 145 (2), 416–425. doi:10.1053/j.gastro.2013.05.002
- Theivanthiran, B., Evans, K. S., DeVito, N. C., Plebanek, M., Sturdivant, M., Wachsmuth, L. P., et al. (2020). A tumor-intrinsic PD-L1/NLRP3 inflammasome signaling pathway drives resistance to anti-PD-1 immunotherapy. *J. Clin. Invest* 130 (5), 2570–2586. doi:10.1172/JCI133055
- Thrasivoulou, C., Millar, M., and Ahmed, A. (2013). Activation of intracellular calcium by multiple Wnt ligands and translocation of β -catenin into the nucleus: a convergent model of Wnt/Ca2+ and Wnt/ β -catenin pathways. *J. Biol. Chem.* 288 (50), 35651–35659. doi:10.1074/jbc.M112.437913
- Tsujimura, N., Yamada, N. O., Kuranaga, Y., Kumazaki, M., Shinohara, H., Taniguchi, K., et al. (2016). A novel role of dickkopf-related protein 3 in macropinocytosis in human bladder cancer T24 cells. *Int. J. Mol. Sci.* 17 (11), 1846. doi:10.3390/ijms17111846
- Tufail, M., and Wu, C. (2023). WNT5A: a double-edged sword in colorectal cancer progression. *Mutat. Res. Rev. Mutat. Res.* 792, 108465. doi:10.1016/j.mrrev.2023.108465
- Valdivia, A., Goicoechea, S. M., Awadia, S., Zinn, A., and Garcia-Mata, R. (2017). Regulation of circular dorsal ruffles, macropinocytosis, and cell migration by RhoG and

its exchange factor, Trio. Trio. Mol. Biol. Cell 28 (13), 1768–1781. doi:10.1091/mbc.E16-06-0412

- Vallée, A., Lecarpentier, Y., and Vallée, J. N. (2021). The key role of the WNT/β-Catenin pathway in metabolic reprogramming in cancers under normoxic conditions. *Cancers (Basel)* 13 (21), 5557. doi:10.3390/cancers13215557
- VanderVorst, K., Dreyer, C. A., Hatakeyama, J., Bell, G. R. R., Learn, J. A., Berg, A. L., et al. (2023). Vangl-dependent Wnt/planar cell polarity signaling mediates collective breast carcinoma motility and distant metastasis. *Breast Cancer Res.* 25 (1), 52. doi:10.1186/s13058-023-01651-2
- Vaupel, P., and Multhoff, G. (2021). Revisiting the Warburg effect: historical dogma versus current understanding. J. Physiol. 599 (6), 1745–1757. doi:10.1113/JP278810
- Voloshanenko, O., Gmach, P., Winter, J., Kranz, D., and Boutros, M. (2017). Mapping of Wnt-Frizzled interactions by multiplex CRISPR targeting of receptor gene families. *FASEB J.* 31 (11), 4832–4844. doi:10.1096/fj.201700144R
- Wallingford, J. B. (2012). Planar cell polarity and the developmental control of cell behavior in vertebrate embryos. *Annu. Rev. Cell Dev. Biol.* 28, 627–653. doi:10.1146/annurev-cellbio-092910-154208
- Wang, Q., Symes, A. J., Kane, C. A., Freeman, A., Nariculam, J., Munson, P., et al. (2010). A novel role for Wnt/Ca2+ signaling in actin cytoskeleton remodeling and cell motility in prostate cancer. *PLoS One* 5, e10456. doi:10.1371/journal.pone.0010456
- Wang, G., Yu, Y., Wang, Y. Z., Yin, P. H., Xu, K., and Zhang, H. (2020). The effects and mechanisms of isoliquiritigenin loaded nanoliposomes regulated AMPK/mTOR mediated glycolysis in colorectal cancer. *Artif. Cells Nanomed Biotechnol.* 48 (1), 1231–1249. doi:10.1080/21691401.2020.1825092
- Wang, B., Chen, J., Caserto, J. S., Wang, X., and Ma, M. (2022a). An *in situ* hydrogel-mediated chemo-immunometabolic cancer therapy. *Nat. Commun.* 13 (1), 3821. doi:10.1038/s41467-022-31579-8
- Wang, B., Khan, S., Wang, P., Wang, X., Liu, Y., Chen, J., et al. (2022b). A highly selective GSK-3 β inhibitor CHIR99021 promotes osteogenesis by activating canonical and autophagy-mediated wnt signaling. *Front. Endocrinol. (Lausanne)* 13, 926622. doi:10.3389/fendo.2022.926622
- Wang, Z., Lan, H., Wang, Y., Zheng, Q., Li, C., Wang, K., et al. (2024a). Pyruvate carboxylase attenuates myocardial ischemia-reperfusion injury in heart transplantation via Wnt/ β -Catenin-Mediated glutamine metabolism. *Biomedicines* 12 (8), 1826. doi:10.3390/biomedicines12081826
- Wang, Q., Li, S., Xu, C., Wang, X., Yang, T., et al. (2024b). Glutaminolysis inhibition boosts photodynamic therapy to eliminate cancer stem cells. *Biomaterials* 306, 122497. doi:10.1016/j.biomaterials.2024.122497
- Wang, J., Dong, X., Lei, J., Zhang, Y., Chen, S., and He, Y. (2024c). β -catenin orchestrates Gli1+ cell fate in condylar development and TMJOA. *J. Dent. Res.* 103 (12), 1291–1301. doi:10.1177/00220345241274354
- Warburg, O. (1956). On the origin of cancer cells. *Science* 123, 309–314. doi:10.1126/science.123.3191.309
- Wei, P. S., Thota, N., John, G., Chang, E., Lee, S., Wang, Y., et al. (2024). Enhancing RNA-lipid nanoparticle delivery: organ- and cell-specificity and barcoding strategies. *J. Control Release* 375, 366–388. doi:10.1016/j.jconrel.2024.08.030
- Welliver, T. P., Chang, S. L., Linderman, J. J., and Swanson, J. A. (2011). Ruffles limit diffusion in the plasma membrane during macropinosome formation. *J. Cell Sci.* 124 (23), 4106–4114. doi:10.1242/jcs.091538
- Williams, M. L. K., and Solnica-Krezel, L. (2020). Cellular and molecular mechanisms of convergence and extension in zebrafish. *Curr. Top. Dev. Biol.* 136, 377–407. doi:10.1016/bs.ctdb.2019.08.001
- Wise, D. R., and Thompson, C. B. (2010). Glutamine addiction: a new therapeutic target in cancer. *Trends Biochem. Sci.* 35, 427–433. doi:10.1016/j.tibs.2010.05.003
- Wise, D. R., DeBerardinis, R. J., Mancuso, A., Sayed, N., Zhang, X.-Y., Pfeiffer, H. K., et al. (2008). Myc regulates a Transcriptional Program that stimulates mitochondrial glutaminolysis and leads to glutamine addiction. *Proc. Natl. Acad. Sci. U. S. A.* 105 (48), 18782–18787. doi:10.1073/pnas.0810199105
- Wong, C. C., Xu, J., Bian, X., Wu, J. L., Kang, W., Qian, Y., et al. (2020). In colorectal cancer cells with mutant KRAS, SLC25A22-Mediated glutaminolysis reduces DNA demethylation to increase WNT signaling, stemness, and drug resistance. *Gastroenterology* 159 (6), 2163–2180. doi:10.1053/j.gastro.2020.08.016
- Xu, Y. K., and Nusse, R. (1998). The frizzled CRD domain is conserved in diverse proteins including several receptor tyrosine kinases. *Curr. Biol.* 8, R405–R406. doi:10.1016/s0960-9822(98)70262-3
- Xu, G., Zhang, Q., Cheng, R., Qu, J., and Li, W. (2025). Survival strategies of cancer cells: the role of macropinocytosis in nutrient acquisition, metabolic reprogramming, and therapeutic targeting. *Autophagy* 21 (4), 693–718. doi:10.1080/15548627.2025.2452149
- Yang, Y., and Mlodzik, M. (2015). Wnt-Frizzled/planar cell polarity signaling: cellular orientation by facing the wind (Wnt). *Annu. Rev. Cell Dev. Biol.* 31, 623–646. doi:10.1146/annurev-cellbio-100814-125315
- Yang, K., Wang, X., Zhang, H., Wang, Z., Nan, G., Li, Y., et al. (2016). The evolving roles of canonical WNT signaling in stem cells and tumorigenesis: implications in targeted cancer therapies. *Lab. Invest* 96 (2), 116–136. doi:10.1038/labinvest.2015.144

Yoo, H. C., and Han, J. M. (2022). Amino acid metabolism in cancer drug resistance. Cells $11\ (1)$, 140. doi:10.3390/cells11010140

Yu, J., Chen, L., Cui, B., Widhopf, G. F., Shen, Z., Wu, R., et al. (2016). Wnt5a induces ROR1/ROR2 heterooligomerization to enhance leukemia chemotaxis and proliferation. *J. Clin. Investig.* 126, 585–598. doi:10.1172/JCI83535

Zeng, L., Fagotto, F., Zhang, T., Hsu, W., Vasicek, T. J., Perry, W. L., et al. (1997). The mouse fused locus encodes axin, an inhibitor of the wnt signaling pathway that regulates embryonic axis formation. *Cell* 90 (1), 181–192. doi:10.1016/s0092-8674(00)80324-4

Zhai, L., Bell, A., Ladomersky, E., Lauing, K. L., Bollu, L., Sosman, J. A., et al. (2020). Immunosuppressive IDO in cancer: mechanisms of action, animal models, and targeting strategies. *Front. Immunol.* 11, 1185. doi:10.3389/fimmu.2020.01185

Zhang, B., Shi, Y., Gong, A., Pan, Z., Shi, H., Yang, H., et al. (2016). HucMSC exosome-delivered 14-3-3 ζ orchestrates self-control of the wnt response via modulation of YAP during cutaneous regeneration. *Stem Cells* 34 (10), 2485–2500. doi:10.1002/stem.2432

Zhang, L., Zeng, S., Yu, Z., Zhang, G., Xiong, Z., Xie, F., et al. (2022). Overexpression of activating transcription Factor-2 (ATF-2) activates Wnt/Ca2+ signaling pathways and promotes proliferation and invasion in non-small-cell lung cancer. *Dis. Markers* 2022, 5772089. doi:10.1155/2022/5772089

Zhang, N., Yuan, M., and Wang, J. (2023). LGR4: a new receptor member in endocrine and metabolic diseases. $\it Endocr. Rev. 44 (4), 647-667. doi:10.1210/endrev/bnad003$

Zhao, F., Xiao, C., Evans, K. S., Theivanthiran, T., DeVito, N., Holtzhausen, A., et al. (2018). Paracrine Wnt5a- β -Catenin signaling triggers a metabolic program that drives dendritic cell tolerization. *Immunity* 48 (1), 147–160. doi:10.1016/j.immuni.2017.12.004

Zheng, S., and Sheng, R. (2024). The emerging understanding of frizzled receptors. FEBS Lett. 598 (16), 1939-1954. doi:10.1002/1873-3468.14903

Zhou, B., Magana, L., Hong, Z., Huang, L. S., Chakraborty, S., Tsukasaki, Y., et al. (2020). The angiocrine Rspondin3 instructs interstitial macrophage transition via metabolic-epigenetic reprogramming and resolves inflammatory injury. *Nat. Immunol.* 21 (11), 1430–1443. doi:10.1038/s41590-020-0764-8

Zhu, X., Xu, M., Leu, N. A., Morrisey, E. E., and Millar, S. E. (2023). FZD2 regulates limb development by mediating β -catenin-dependent and -independent Wnt signaling pathways. *Dis. Model Mech.* 16 (3), dmm049876. doi:10.1242/dmm.049876

Zhuang, Q., Zhao, B., Lin, Z., Liang, Y., Zhao, Q., Wang, Y., et al. (2023). Navoximod modulates local HSV-1 replication to reshape tumor immune microenvironment for enhanced immunotherapy via an injectable hydrogel. *Commun. Biol.* 6 (1), 621. doi:10.1038/s42003-023-04983-z

Zou, G., and Park, J. I. (2023). Wnt signaling in liver regeneration, disease, and cancer. Clin. Mol. Hepatol. 29(1), 33-50. doi:10.3350/cmh.2022.0058

Zuo, Q., He, J., Zhang, S., Wang, H., Jin, G., et al. (2021). PPAR γ Coactivator-1 α suppresses metastasis of hepatocellular carcinoma by inhibiting warburg effect by PPAR γ -Dependent WNT/ β -Catenin/Pyruvate dehydrogenase kinase isozyme 1 axis. Hepatology 73 (2), 644–660. doi:10.1002/hep.31280

Zylkiewicz, E., Sokol, S. Y., and Hoppler, S. (2014). "Wnt signaling in early vertebrate development: from fertilization to gastrulation," in *Wnt signaling in development and disease*. Editors S. Hoppler, and R. T. Moon (Hoboken, NJ: John Wiley and Sons), 251–266.