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*CORRESPONDENCE Yi-li Wang wangyili@cmc.edu.cn Xiao Zhang 954073462@qq.com

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Molecular docking and molecular dynamics study Lianhua Qingwen granules (LHQW) treats COVID-19 by inhibiting inflammatory response and regulating cell survival

Jun-Feng Cao^{1,2}, Yunli Gong¹, Mei Wu¹, Li Xiong¹, Shengyan Chen¹, Haonan Huang¹, Xinge Zhou¹, Ying-chun Peng^{1,3}, Xue-fang Shen^{1,3}, Jinyu Qu^{1,3}, Yi-li Wang^{1,3*} and Xiao Zhang^{1,2*}

¹Chengdu Medical College, Chengdu, China, ²Chengdu Medical College of Basic Medical Sciences, Chengdu, China, ³The First Affifiliated Hospital of Chengdu Medical College, Chengdu, China

Purpose: 2019 Coronavirus disease (COVID-19) is endangering health of populations worldwide. Latest research has proved that *Lianhua Qingwen granules* (LHQW) can reduce tissue damage caused by inflammatory reactions and relieve patients' clinical symptoms. However, the mechanism of LHQW treats COVID-19 is currently lacking. Therefore, we employed computer simulations to investigate the mechanism of LHQW treats COVID-19 by modulating inflammatory response.

Methods: We employed bioinformatics to screen active ingredients in LHQW and intersection gene targets. PPI, GO and KEGG was used to analyze relationship of intersection gene targets. Molecular dynamics simulations validated the binding stability of active ingredients and target proteins. Binding free energy, radius of gyration and the solvent accessible surface area were analyzed by supercomputer platform.

Results: COVID-19 had 4628 gene targets, LHQW had 1409 gene targets, intersection gene targets were 415. Bioinformatics analysis showed that intersection targets were closely related to inflammation and immunomodulatory. Molecular docking suggested that active ingredients (including: licopyranocoumarin, Glycyrol and 3-3-Oxopropanoic acid) in LHQW played a role in treating COVID-19 by acting on CSF2, CXCL8, CCR5, NLRP3, IFNG and TNF. Molecular dynamics was used to prove the binding stability of active ingredients and protein targets.

Conclusion: The mechanism of active ingredients in LHQW treats COVID-19 was investigated by computer simulations. We found that active ingredients in LHQW not only reduce cell damage and tissue destruction by inhibiting the inflammatory

response through CSF2, CXCL8, CCR5 and IFNG, but also regulate cell survival and growth through NLRP3 and TNF thereby reducing apoptosis.

KEYWORDS

COVID-19, lianhua qingwen granules (LHQW), molecular docking, bioinformatics analysis, molecular dynamics



Introduction

SARS-CoV-2 is a single-stranded RNA virus that belongs to the beta family of coronaviruses (Mahmoudinobar et al., 2021; Bui et al., 2021). The virus has a spherical lipid bilayer envelope with a spike (S) protein on its surface responsible for viral infection (Ilkhani et al., 2021; Qasem et al., 2021). The genome of SARS-CoV-2 is engaged by two open reading frames, they encode Nuclear Protein Phosphatase 1 α (PP1A) and Replicase polyprotein 1ab (pp1ab) (Citarella et al., 2021). The coronavirus protease M^{pro} (also known as 3C-like protease (3CL^{pro})) is a triple structural domain cysteine protease and M^{pro} is involved in the proteolytic processing of pp1a and pp1ab (Panikar et al., 2021). Numerous studies have suggested a critical role for the protease M^{pro} in coronavirus gene expression and replicase processing (Roe et al., 2021), and M^{pro} mediates viral replication and transcription (Sabbah et al., 2021). Study has been suggested that targeting and inhibiting M^{pro} activity is an effective way to prevent SARS-CoV-2 replication and transmission (Sultan et al., 2021). DasGupta et al., 2022 used mixed solvent molecular dynamics (MixMD) to simulate and

analyze the possible metastable sites of M^{pro}. Sztain et al., 2020 identified cryptic pocket structures within M^{pro} of SARS-CoV-2 by Gaussian accelerated molecular dynamics, and these pocket structures may serve as drug targets to develop protease inhibitors of COVID-19. Strömich et al., 2022 found by computer simulation that M^{pro} has multiple potentially acting binding sites capable of interfering with viral RNA transcription and protein translation. Günther et al., 2021 identified two metastable binding sites of M^{pro} as possible drug targets against SARS-CoV-2 by high-throughput X-ray crystallography analysis.

The clinical presentation of patients with COVID-19 ranges from mild flu-like symptoms to life-threatening multi-organ failure. (Sidiq et al., 2020). Studies have found that critically ill patients may develop acute myocardial injury, renal dysfunction and coagulation complications (Rello et al., 2020; Shi et al., 2021; Ciotti et al., 2021). Although relevant vaccines and drugs have been used clinically, clinical studies have found many adverse events (such as: fever, headache, muscle pain and joint pain) after vaccination in some populations (Vitiello et al., 2021; Li Z et al., 2020). The Ad26.COV2.S vaccine produced by Johnson had 34% untreated rate of vaccine efficacy against COVID-19 in a phase 3 clinical trial. (Hadj Hassine, 2022). Gam-COVID-Vac (Sputnik V) is a heterologous adenovirus vector vaccine based on SARS-CoV-2, its phase 3 clinical trial found mild adverse reactions in 94% of subjects, with a very small number of subjects experiencing serious adverse events. And there were four vaccine-related deaths in subjects (Fernandes et al., 2022). Both the Pfizer and Modena COVID-19 vaccines granted by the U.S. Food and Drug Administration (FDA) have shown some associated adverse reactions (Meo et al., 2021; Ng et al., 2021; Yamamoto, 2022). The antiviral drug raltegravir is the only drug approved for the treatment of COVID-19. Although there is little information on the adverse events of raltegravir, studies have reported that raltegravir use is associated with an increase in renal disease (Martinez-Lopez et al., 2020; Chouchana et al., 2021). Famipiravir is one of the anti-SARS-CoV-2 candidates, clinical trials have found adverse effects such as vomiting chest pain and elevated serum liver transaminase and uric acid levels in mild to moderate patients (Hassanipour et al., 2021).

In the treatment for COVID-19, China has always adhered to the principle of "treating both Chinese and Western medicine", and the Chinese herbal medicine Lianhua Qingwen granules (LHQW) has played an important role in the fight against the epidemic (Zeng et al., 2020). LHQW is a herbal formula consisting of 13 herbs, including forsythia, jinyinhua, ephedra, almonds, gypsum, panax quinquefolium, cotton horse kanji, fishy grass, patchouli, rhubarb, licorice, rhodiola, and menthol. LHQW has broad-spectrum antiviral, effective antibacterial, antipyretic and anti-inflammatory, anti-cough and cough, and immunomodulatory functions (Zhang X et al., 2020). Studies have shown that LHQW has good effects on various influenza virus-induced respiratory diseases (Zeng et al., 2020). Clinical studies have shown that Lotus Clear can improve clinical symptoms (such as: fever, fatigue and muscle aches) and shorten the duration of COVID-19 in patients with COVID-19. Experimental studies have also shown that lotus seeds can exert pharmacological effects in the treatment of COVID-19 by inhibiting viral replication and reducing cytokine release from host cells (Liu C et al., 2021; Runfeng et al., 2020; Hu et al., 2021). LHQW is commonly used as an adjuvant to COVID-19, and it is thought to provide symptomatic relief and shorten the course of the disease (Wang et al., 2020). Study found that LHQW reduced D-dimer (prognostic indicator of pneumonia) and erythrocyte sedimentation rate (marker of inflammation) in patients, thereby delaying disease progression (Shen et al., 2021). LHQW was found to improve not only the clinical symptoms but also the prognosis of patients with COVID-19 (Xiao et al., 2020; Li L. et al., 2020; Zeng et al., 2020; Zhuang et al., 2021). A pharmacodynamic study from 284 patients with neocrown pneumonia in China showed a cure rate of 91.5% in the LHQW treatment group, which was significantly higher than the 82.4% cure rate in the conventional group (Wu and Zhong, 2021). And clinical studies have shown that LHQW has improved the symptoms of fever, fatigue and cough in COVID-19 patients (Tong et al., 2020; Wang et al., 2022). Although LHQW has played a positive role in the treatment of COVID-19, the complex composition has hindered a more indepth study of the pharmacological mechanism of action of lotus clematis in the treatment of COVID-19.

Therefore, we further investigated the mechanism of LHQW treats COVID-19 by bioinformatics analysis and computer simulation. Molecular docking and molecular dynamics can allow for a comprehensive simulation of the interaction and binding stability among ligand active ingredients and receptor protein targets with the help of powerful computational capabilities.

In this study, bioinformatics was used to obtain active ingredients in LHQW and intersection gene targets. computer simulations were validated the relationship among active ingredients and protein targets by supercomputer platform. Molecular docking was used to validate affinity of ligand active ingredients and receptor proteins. Molecular dynamics was used to simulate the stability of binding complex.

Material and methods

Screening active ingredients and gene targets

Traditional Chinese Medicine Systems Pharmacology Database (TCMSP) was used to screen and analyse ingredients in *Lianhua Qingwen granules* (LHQW) (Mok et al., 2022). And active ingredients in LHQW were screened by the criterion of OB≥80% and DL≥0.2. Oral bioavailability (OB) is defined as the extent to which the active ingredient is utilized by the body, and OB can determine the impact of a compound on disease (Zhang Y. et al., 2020). In drug development, drug likeness (DL) is important to improve the success of drug discovery and development (Wu et al., 2022). GeneCards database was used to obtain disease gene targets and drug gene targets (Stelzer et al., 2016). Disease gene targets and drug gene targets were combined by Venny website to obtain intersection gene targets.

Protein-protein interaction network and enrichment analysis

The STRING database was employed to analyze proteinprotein interaction (PPI) of *Lianhua Qingwen granules* (LHQW) treats COVID-19 (Szklarczyk et al., 2021). And core interaction protein targets were screened according to the criteria of nodal degree value and median centroid value greater than the mean. DAVID database was employed for gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analysis (Wu et al., 2020). This study obtained cellular component (CC), molecular function (MF) and biological process (BP) of the gene targets through GO enrichment. KEGG pathway enrichment analysis was performed on the relevant signaling pathways involved in interaction gene targets (Cao et al., 2022a).

Molecular docking

The Pubchem database was used to obtain 3D structures of small molecules, and the PDB database was used to obtain the structures of proteins (Jagdish and Marty Ytreberg, 2019; Zhou et al., 2020; Adegbola et al., 2021; Dela Cruz et al., 2022). Small molecules and proteins were subjected to energy minimization under MMFF94 force field. AutoDock Vina 1.1.2 software was used for molecular docking (Aprivanti et al., 2021). All protein seat receptors were pre-treated (including the removal of water molecules, salt ions, and other small molecules from the protein results) by PyMol 2.5 (Kashyap et al., 2021). The center of the docking box was defined using PyMol based on the location of the active site and the box side length was set to 22.5 Å. ADFRsuite 1.0 was used to convert all processed small molecules and receptor proteins into the PDBQT format required for molecular docking (Song et al., 2019; Kumar et al., 2020). The highest scoring docked conformations from the molecular docking output were used for subsequent molecular dynamics simulations (Cao et al., 2022b). We used the original crystal ligand of the target protein as a positive reference, and analyzed and compared the binding posture of the original crystal ligand and protein, the chemical bond length and the chemical bond angle by re-docking the original crystal ligand and protein. Finally, the consistency of the binding mode can indicate the correctness of the molecular docking protocol.

Molecule dynamics

The highest scoring conformations identified by molecular docking analysis were further validated by molecular dynamics simulation. Molecular dynamics (MD) simulation is based on Newtonian mechanics theory to model the motion of molecular systems (Jagdish et al., 2014). AMBER 18 software was used to perform molecular dynamics simulations (Mermelstein et al., 2018; Lee et al., 2018). GAFF2 small molecule force fields and ff14SB protein force fields were used to treat ligand small molecules and receptor proteins (Lee J. et al., 2020). Hydrogen atoms were added to all complex systems using the LEaP module and a truncated octahedral TIP3P solvent cassette was added at a distance of 10 Å from the system. Na+/Cl- was added to balance the charge of the complex system (Rieloff and Skepö, 2021). The steepest descent method with 2500 steps and the conjugate gradient method with 2500 steps were used for the energy optimization of the complex system (Lee, T. S. et al., 2020). After the energy optimization of the system was completed, the

system temperature was steadily increased from 0 K to 298.15 K at a fixed volume and constant ramp rate. The NVT (Isothermal isovolume) system simulation of 500 ps was performed to further uniformly distribute the solvent molecules in the solvent box at the system maintenance temperature of 298.15 K (Cao et al., 2022c). Finally, the composite system was subjected to a 100 ns NPT (isothermal isobaric) system simulation. The truncation distance of the non-bond was set to 10 Å during the molecular dynamics simulation. Particle mesh Ewald (PME) was used to calculate electrostatic interactions. SHAKE method was used to limit the bond length of hydrogen atoms. Langevin algorithm was used for temperature control of the simulation, and the collision frequency was set to 2 ps⁻¹. The simulated system pressure was 1 atm, the integration step was 2 fs, and the traces were saved at 10 ps intervals for subsequent analysis. The binding free energy can reflect the interaction between the ligand small molecule and the receptor protein (Wang et al., 2021). The interactions between ligand and receptor include both covalent and nonbonded interactions, but only non-bonded interactions (van der Waals interactions, electrostatic interactions and hydrogen bonding interactions) are generally present in complex systems of drug small molecules (ligands) and proteins (receptors) (Dasmahapatra et al., 2022). Therefore, AMBER 18 was used to calculate the binding free energy by MM/GBSA method in this study (Weng et al., 2019; Cao et al., 2022d).

Results

Active ingredientsand intersection gene targets

1391 ingredients in *Lianhua Qingwen granules* (LHQW) were obtained from TCMSP database. 10 active ingredients were screened by using DL \geq 0.2 and OB \geq 80% as the criteria (Table 1). GeneCards database was used to obtain 4628 COVID-19 gene targets and 1409 *Lianhua Qingwen granules* (LHQW) gene targets. And 415 intersection gene targets were processed by Venny (Figure 1).

Core intersection target screening and Protein-protein interaction network constructing

This study obtained 29 core intersection targets by relevance score (relevance score \geq 4). STRING database was used to analysis core intersection targets of LHQW and COVID-19, 13 core protein targets (including: NLRP3, TNF, CSF2, IFNG) were obtained by setting confidence degree (confidence degree>0.95) (Figures 2A, B).

TABLE 1 The active ingredients in Lianhua Qingwen granules (LHQW).

The active ingredients in Lianhua Qingwen granules (LHQW)

| Mol ID | Molecule Name | MW | AlogP | OB (%) | BBB | DL | HL |
|-----------|--|--------|-------|-----------|-------|------|-------|
| MOL003330 | Phillygenin | 372.45 | 2.38 | 95.04 | 0.07 | 0.57 | 1.97 |
| MOL002311 | Glycyrol | 366.39 | 4.85 | 90.78 | -0.2 | 0.67 | 9.85 |
| MOL003006 | (3R,8S,9R,9aS,10aS)-9-ethenyl-8-(beta-D-glucopyranosyloxy)-2,3,9,9a,10,10a-hexahydro-5-oxo-5H,8H-pyrano-pyridine-3-carboxylic acid | 281.29 | -0.96 | 87.47 | -0.89 | 0.23 | 5.5 |
| MOL012922 | I-SPD | 327.41 | 3.1 | 87.35 | 0.21 | 0.54 | 1.68 |
| MOL001734 | 3-3-oxopropanoic acid | 379.35 | 0.36 | 85.87 | -1.25 | 0.47 | 3.58 |
| MOL003306 | Pinoresinol monomethyl ether | 372.45 | 2.38 | 85.12 | 0 | 0.57 | 2.12 |
| MOL004990 | 7,2',4'-trihydroxy-5-methoxy-3-arylcoumarin | 300.28 | 2.56 | 83.71 | -0.59 | 0.27 | 0.99 |
| MOL000471 | Aloe-emodin | 270.25 | 1.67 | 83.38 | -1.07 | 0.24 | 31.49 |
| MOL003322 | Forsythinol | 372.45 | 2.38 | 81.25 | -0.08 | 0.57 | 2.72 |
| MOL004904 | Licopyranocoumarin | 384.41 | 3.04 | 80.36 | -0.62 | 0.65 | 0.08 |

MW, molecular weight.

AlogP, partition coefficient between octanol and water.

OB, oral bioavailability.

BBB, blood brain barrier.

DL, drug similarity.

HL, drug half-life.

GO and KEGG enrichment analysis

The 29 intersection gene targets were imported into DAVID database for GO and KEGG enrichment analysis. GO enrichment analysis yielded a total of 267 GO, including 224 biological processes (BP), 19 cell components (CC) and 24 molecular functions (MF). Biological processes were correlated with cellular response to lipopolysaccharide and inflammatory response. Among cell components, extracellular space and integral component of plasma membrane for a relatively large amount. In molecular functions, virus receptor activity and virus receptor activity were relatively high (Figures 3A–F). KEGG pathway yielded 71 pathways, and enriched pathways involved rheumatoid arthritis, coronavirus disease - COVID-19 and other signaling pathways (Figures 3G, H).

Molecular docking

The 10 active ingredients and 13 core intersection protein targets were used for molecular docking. The stability of protein receptor-small molecule ligand binding depends on the binding energy. The lower the binding energy of the complex, the more stable the receptor-ligand binding conformation (Figure 4). Molecular docking indicated that CCR5/Licopyranocoumarin was mainly maintained by hydrogen bonding and hydrophobic interactions. Licopyranocoumarin interacted with Arg-184, Tyr-232, Gln-41, Gly-44, Gly-46 and Ser-245 on the CCR5 protein by hydrogen bonding and with Val-230 and Pro-149 by hydrophobic interactions (Figure 5A). The binding of CSF2/3-Oxopropanoic acid was maintained mainly by hydrophobic

interactions. 3-3-Oxopropanoic acid interacted with His-15 on the CSF2 protein by hydrogen bonding and with Leu-55, Leu-59, Phe-47, Ile-117 and Ile-19 by hydrophobic interactions (Figure 5B). In the CXCL8/Glycyrol, Glycyrol interacted with Glu-29 and Val-27 on CXCL8 by hydrogen bonding and with Val-58, Ile-61, Leu-25 and Val-27 by hydrophobic interactions (Figure 5C). The binding of IFNG/Glycyrol indicated that Glycyrol hydrogen bonds with Ala-8 and Leu-28 on the protein, hydrophobic interaction with Leu-11, Phe-15, Phe-57, Leu-57 and Leu-30 (Figure 5D). Licopyranocoumarin could form hydrogen bonding with Ser-626 and Arg-578 on NLRP3 protein, and also with Val-353, Pro-352, Tyr-632, Ile-411, Phe-575 and Thr-439 formed a hydrophobic interaction (Figure 5E). The binding of TNF/Glycyrol was mainly through hydrophobic interaction, Glycyrol interacted with Phe-144, Ala-145, Asp-143, Gln-67 and Tyr-141 on TNF protein by hydrophobic interactions and with Glu-23 by hydrogen bonding (Figure 5F).

Molecular dynamics and binding free energy

The root mean square deviation (RMSD) was used to reflect the fluctuating processes of the complexes. Higher RMSD values and fluctuations of the complexes indicate more intense motions of protein receptors and small molecule ligands. The MMGBSA method was used to calculate the binding free energy of the complexes, and the binding free energy can more accurately reflect the magnitude of the binding stability of the ligand small molecule to the receptor protein (Procacci, 2020). The binding free energy reflects the energy released by the binding of the





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processes (BP) were greatly enriched. (B) The subnetwork displayed the top 10 BP terms and related genes. (C) The top 10 terms in cellular components (CC) were greatly enriched. (D) The subnetwork displayed the first 10 CC terms and related genes. (E) The top 10 terms in molecular function (MF) were greatly enriched. (F) The subnetwork displayed the top 10 MF terms and related genes. (G) The top 20 KEGG pathways were showed. (H) The subnetworks displayed the top 20 KEGG pathways and related.

ligand and receptor into the complex (Al-Khafaji and Taskin Tok, 2020). The higher the value of the binding free energy, the more stable the structure of the complex. Except for CSF2/3-3-Oxopropanoic acid, the remaining five complexes gradually converged within the first 5 ns of the simulation and remained stable in the subsequent simulations. In particular, RMSD values of three complexes (CCR5/Licopyranocoumarin, NLRP3/Licopyranocoumarin and TNF/Glycyrol) were small (values below 3.5 Å) and fluctuated very stably. CSF2/3-3-Oxopropanoic acid and IFNG/Glycyrol both showed

significant fluctuations in the first 50 ns, but they rapidly stabilized after 50 ns. CXCL8/Glycyrol was never in a stable fluctuation throughout the simulation, indicating the relatively poor stability of CXCL8/Glycyrol (Figure 6). The binding free energies of CCR5/Licopyranocoumarin, CSF2/3-3-Oxopropanoic acid, CXCL8/Glycyrol, IFNG/Glycyrol, NLRP3/Licopyranocoumarin and TNF/Glycyrol were -25.52 \pm 2.86 kcal/mol, -6.98 \pm 5.02 kcal/mol, -39.15 \pm 1.96 kcal/mol, -34.16 \pm 4.59 kcal/mol, -38.17 \pm 1.51 kcal/mol and -16.84 \pm 1.92 kcal/mol (Table 2). The stability of CCR5/Licopyranocoumarin, CXCL8/





Binding conformation of active ingredients and protein targets. (A) CCR5/Licopyranocoumarin, (B) CSF2/3-3-Oxopropanoic acid, (C) CXCL8/ Glycyrol, (D) IFNG/Glycyrol, (E) NLRP3/Licopyranocoumarin, (F) TNF/Glycyrol.



Glycyrol and IFNG/Glycyrol was good and their values of binding free energy were greater than -30.0 kcal/mol. We found that the binding free energies of all complexes were mainly Van der Waals and electrostatic energies.

Analysis of root mean square fluctuations

In this study, root mean square fluctuations (RMSF) were used to model the vibrations of each residue after ligand and receptor binding to explore the fluctuating changes of macromolecular proteins at the residue level. Typically, the flexibility of a protein decreases upon binding of a ligand small molecule drug to a receptor protein, thereby modulating the active site of the protein. The simulation results show that all proteins except the ends of the protein have low RMSF after binding different small molecules, indicating that the core structure of the protein possesses good rigidity. Notably, the overall RMSFs of CCR5/Licopyranocoumarin, CSF2/3-3-Oxopropanoic acid, IFNG/Glycyrol, NLRP3/Licopyranocoumarin and TNF/Glycyrol were all less than 2.5 Å, indicating that the rigidity of the proteins increased significantly after the binding of proteins to small molecules (Figure 7).

Analysis of hydrogen bond

Hydrogen bonding is one of the strongest non-covalent binding interactions between receptor and ligand, and the higher the number of hydrogen bonds, the better the binding

TABLE 2 Binding free energies and energy components predicted by MM/GBSA (kcal/mol).

Results of binding free energies

| System name | CCR5/ Licopyranocoumarin | CSF2/3-3- Oxopropanoic acid | CXCL8/ Glycyrol | IFNG/ Glycyrol | NLRP3/ Licopyranocoumarin | TNF/ Glycyrol | |
|----------------------|-----------------------------|--------------------------------|--------------------|-------------------|------------------------------|-------------------|--|
| $\Delta E_{\rm vdw}$ | -31.41 ± 2.76 | -11.53 ± 6.29 | -46.22 ± 1.20 | -41.36 ± 1.57 | -43.33 ± 2.46 | -26.13 ± 2.16 | |
| ΔE_{elec} | -25.44 ± 5.08 | -12.81 ± 38.54 | -8.04 ± 2.37 | -18.88 ± 9.99 | -35.40 ± 2.72 | -13.98 ± 5.68 | |
| ΔG_{GB} | 5.62 ± 3.49 | 19.08 ± 40.98 | 20.90 ± 1.70 | 31.88 ± 5.76 | 46.01 ± 3.84 | 25.71 ± 5.40 | |
| ΔG_{SA} | -4.29 ± 0.24 | -1.72 ± 1.01 | -5.78 ± 0.16 | -5.80 ± 0.17 | -6.44 ± 0.10 | -2.44 ± 0.31 | |
| ΔG_{bind} | -25.52 ± 2.86 | -6.98 ± 5.02 | -39.15 ± 1.96 | -34.16 ± 4.59 | -38.17 ± 1.51 | -16.84 ± 1.92 | |

 ΔE_{vdW} , van der Waals energy.

 ΔE_{elec} , electrostatic energy.

 ΔG_{GB} , electrostatic contribution to solvation.

 ΔG_{SA} , non-polar contribution to solvation.

 ΔG_{bind} , binding free energy.



stability. CCR5/Licopyranocoumarin with a stable number of hydrogen bonds of about 3 throughout; followed by NLRP3/ Licopyranocoumarin and CXCL8/Glycyrol with the number of hydrogen bonds remaining at 2-3 at the late stage of the simulation. We found that CSF2/3-3-Oxopropanoic acid and CXCL8/Glycyrol showed a significant decrease in hydrogen bonding in the early stage of the simulation, and this result further explains the anomalous fluctuations of CSF2/3-3-Oxopropanoic acid and CXCL8/Glycyrol in the early stage of RSMD (Figure 8).

Analysis of the radius of gyration

The radius of rotation (Rog) can be used to reflect the tightness of the bonding of the complex system. The fluctuation size can very intuitively reflect the denseness of the composite or the system convergence. The values of Rog were TNF/Glycyrol, CCR5/Licopyranocoumarin, NLRP3/Licopyranocoumarin, CXCL8/Glycyrol, IFNG/Glycyrol and CSF2/3-3-Oxopropanoic acid from the largest to the smallest. The results of Rog were similar to the RMSD results, and the results indicated that TNF/Glycyrol, CCR5/Licopyranocoumarin could maintain very stable binding state (Figure 9).

Analysis of solvent accessible surface area

The solvent accessible surface area (SASA) is used to calculate the interface at which the complex is surrounded by the solvent. Because solvents behave differently under different conditions, SASA is a useful parameter for studying protein conformational dynamics in solvent environments. Based on the results of SASA fluctuation analysis, we could find that the fluctuation of CSF2/3-3-Oxopropanoic acid and NLRP3/Licopyranocoumarin were larger, which indicated that the exposure and burial area of the surface of this two proteins occur more changes (Figure 10).

Discussion

We explored the mechanism of *Lianhua Qingwen granules* (LHQW) treats COVID-19 by computer simulations. This study found that active ingredients in *Lianhua Qingwen granules* (LHQW) may reduce cell damage and tissue destruction composed of COVID-19 by inhibiting the inflammatory response and regulating cell survival. Firstly, Glycyrol may inhibit activation of the inflammatory response and chemotaxis of inflammatory cells through IFNG and CXCL8







thereby reducing the inflammatory response. Glycyrol may also regulate TNF to reduce cell damage and death thereby alleviating damage from the inflammatory response caused by infection. Secondly, Licopyranocoumarin may impede release of inflammatory factors and spread of inflammatory response through CCR5 and NLRP3 thereby controlling the extent of inflammatory effects and the degree of tissue damage. Finally, 3-3-Oxopropanoic acid may regulate the differentiation of inflammatory cells and cell survival through CSF2 thereby reducing apoptosis.

Therefore, these results demonstrate that active ingredients in *Lianhua Qingwen granules* (LHQW) treats COVID-19 by regulating cell survival and inhibiting inflammatory response from multiple targets.

Analysis of molecular docking and molecular dynamics

Molecular docking could be used to revealed strong affinity of active ingredients in *Lianhua Qingwen granules* (LHQW) (such as: Glycyrol Licopyranocoumarin and 3-3-Oxopropanoic acid) to protein targets (such as: IFNG, CXCL8, TNF, CCR5, NLRP3 and CSF2). And molecular dynamics results suggested that active ingredients in *Lianhua Qingwen granules* (LHQW) and protein could maintain a very stable binding state and thus exert effects in the treatment of COVID-19.

The binding of IFNG/Glycyrol indicated that the Glycyrol hydrogen bonds with Ala-8 and Leu-28 on IFNG, hydrophobic interaction with Leu-11, Phe-15, Phe-57, Leu-57 and Leu-30. In the CXCL8/Glycyrol, Glycyrol interacted with Glu-29 and Val-

27 on CXCL8 protein by hydrogen bonding and with Val-58, Ile-61, Leu-25 and Val-27 by hydrophobic interactions. The binding of TNF/Glycyrol was mainly through hydrophobic interaction, Glycyrol interacted with Phe-144, Ala-145, Asp-143, Gln-67 and Tyr-141 on the TNF protein by hydrophobic interactions and with Glu-23 by hydrogen bonding. The binding free energies of CXCL8/Glycyrol, IFNG/Glycyrol, TNF/Glycyrol were -39.15 \pm 1.96 kcal/mol, -34.16 \pm 4.59 kcal/mol and -16.84 \pm 1.92 kcal/mol. The number of CXCL8/Glycyrol, IFNG/Glycyrol and TNF/ Glycyrol hydrogen bonds remained around 2 in the late simulation period.

Licopyranocoumarin bound to CCR5 and NLRP3 could form very stable complex. Licopyranocoumarin interacted with Arg-184, Tyr-232, Gln-41, Gly-44, Gly-46 and Ser-245 on CCR5 by hydrogen bonding and with Val-230 and Pro-149 by hydrophobic interactions. Licopyranocoumarin could form hydrogen bonding with Ser-626 and Arg-578 on NLRP3 protein, and also with Val-353, Pro-352, Tyr-632, Ile-411, Phe-575 and Thr-439 formed a hydrophobic interaction. The binding free energy of CCR5/Licopyranocoumarin and NLRP3/ Licopyranocoumarin were -25.52 \pm 2.86 kcal/mol and -38.17 \pm 1.51 kcal/mol. The RMSD values of CCR5/Licopyranocoumarin and NLRP3/Licopyranocoumarin were small (values below 3.5 Å) and fluctuated very steadily.

The binding of the CSF2/3-3-Oxopropanoic acid was maintained mainly by hydrophobic interactions. The binding of the CSF2/3-3-Oxopropanoic acid was maintained by hydrophobic interactions. The small molecule 3-3-Oxopropanoic acid interacted with His-15 on the CSF2 protein by hydrogen bonding and with Leu-55, Leu-59, Phe-47, Ile-117 and Ile-19 by hydrophobic interactions. The binding

free energies of CSF2/3-3-Oxopropanoic acid was -6.98 \pm 5.02 kcal/mol. And the overall RMSF of CSF2/3-3-Oxopropanoic acid was less than 2.5 Å.

Glycyrol may inhibit activation of the inflammatory response and chemotaxis of inflammatory cells through IFNG and CXCL8

Interferon-gamma (IFNG) is essential for immune and tumor control of intracellular pathogens. However, abnormal IFNG expression is associated with many auto-inflammatory and autoimmune diseases. GO enrichment analysis includes interferon- γ receptor binding and cytokine activity. KEGG enrichment analysis includes IL27-mediated signaling pathways. C-X-C Motif Chemokine Ligand 8 (CXCL8) is a major mediator of the inflammatory response, CXCL8 clears pathogens and protects the host from infection by attracting neutrophils and T cells. GO enrichment includes interleukin 8 receptor binding and chemokine activity. KEGG enrichment analysis includes GPCR downstream signaling and bacterial infection in the airway.

IFNG is selectively produced by immune cells (such as: T lymphocytes and NKT cells). And the receptor for IFNG (IFNGR) is present in almost all cell types. IFNG can induce antiviral, antiproliferative and immunomodulatory effects in the direction of inflammation (Schoenborn and Wilson, 2007; Nöst et al., 2019; Ozger et al., 2021). IFNG encodes soluble cytokines secreted by the innate and adaptive immune systems. Elevated IFNG levels have been found in COVID-19 patients, and elevated IFNG levels are thought to exacerbate cytokine storms (Xu et al., 2022). IFNG is mainly produced by CD4 T cells. IFNG secretion enhances macrophage microbicidal mechanisms and regulates Th17 cell and tissue damage (Petrone et al., 2021). CD4 + T cells have two inflammatory subpopulations, Th1 and Th17 cells. Th1 and Th17 perform their effector functions by producing inflammatory cytokines (including IFNG and IL-17). Melgaço et al. showed that stinging viral proteins lead to activation of CD4 T helper cell and CD8 T to produce IFNG (Melgaço et al., 2021). Therefore, inhibition of IFNG or IL-17 seems to improve clinical status of critically ill patients. Shahbazi et al. showed an overall reduction in pro-inflammatory Th1 and Th17 lymphocytes in critically ill patients with COVID-19. The reduced frequency of these lymphocytes may be due to their migration to the lungs of critically ill patients and this increases lung inflammation (Shahbazi et al., 2021; Mele et al., 2021). Some studies have shown that the active ingredients of LHQW (such as: lonicerae and glycyrrhizin) can exert inhibitory effects on IFNG and reduce IFNG expression (Nöst et al., 2019; Liu P et al., 2021).

CXCL8 can be rapidly induced by pro-inflammatory cytokines (such as: TNF and IL-1b) (Liu et al., 2016). Kaiser et al. showed that neutrophils were recruited to express CXCL8 in the lung, which in turn activates and enhances CXCL8 release from peripheral neutrophils (Ha et al., 2017). Kaiser et al. found high concentrations of CXCL8 in bronchoalveolar lavage (BAL) fluid from patients with COVID-19 (Kaiser et al., 2021). Studies have shown that LHQW can reduce the gene level of interleukin 8 (CXCL8) (Dong et al., 2014).

Therefore, Glycyrol may inhibit activation of the inflammatory response and chemotaxis of inflammatory cells through IFNG and CXCL8 thereby reducing the inflammatory response.

Glycyrol may regulate TNF to reduce cell damage and death

Tumor necrosis factor alpha is a member of the TNF/TNFR cytokine superfamily (Balkwill, 2006). TNF- α can be released from the cell membrane by extracellular protein hydrolysis cleavage and functions as a cytokine. GO enrichment analysis includes cytokine activity and protein binding. KEGG enrichment analysis includes the regulation of dendritic cell developmental lineage pathways and cell survival.

TNF- α is an inflammatory cytokine produced by macrophages/monocytes during acute inflammation, and TNF- α is responsible for various intracellular signaling events (mediating the gene expression of growth factors, cytokines and transcription factors). TNF- α is also important in the fight against infection and cancer (Idriss and Naismith, 2000; Ramasamy and Subbian, 2021). And TNF-a affects different aspects of the immune system and regulates various pathological and physiological processes (Choudhary et al., 2021). TNF- α is also a major activator of IL-6 expression (Pons et al., 2020). Study found that TNF- α was a prominent feature of patient deterioration (Fara et al., 2020). The severity of COVID-19 is associated with damage produced by cytokine storm and type I IFN (IFN- α and IFN- β) (Ramasamy and Subbian, 2021; Krämer et al., 2021). Interestingly, the numbers of total T cells, CD4 T and CD8 T cells were negatively correlated with TNF-a, IL-6 levels, respectively. This leads to the conclusion that T cells are reduced and depleted in COVID-19 patients. Cytokines (such as IL-6 and TNF- α) may be involved in T-cell reduction (Diao et al., 2020). Study has been shown that most patients suffer from lymphocytopenia with elevated serum pro-inflammatory cytokine levels (such as: TNF-α, IL-6) (Khanmohammadi and Rezaei, 2021). In addition, Norooznezhad et al. found that NFκB, a very critical transcription factor in inflammation (especially cytokine storm) may lead to the expression of IL-6 and TNF-a (Pons et al., 2020; Norooznezhad and Mansouri, 2021; Attiq et al., 2021). Study has been shown that LHQW can inhibit virus-induced NF- κ B activation and alleviate virusinduced gene expression of IL-6, TNF- α (Shen and Yin, 2021).

Therefore, Glycyrol may also regulate TNF to reduce cell damage and death thereby alleviating damage from the inflammatory response caused by infection.

Licopyranocoumarin may impede the release of inflammatory factors and the spread of the inflammatory response through CCR5 and NLRP3

C Motif Chemokine Receptor 5 (CCR5, also known as CD195) is a G protein-coupled receptor (GPCR) (Zeng et al., 2022). CCR5 is expressed on the surface of leukocytes, especially T-CD4⁺ cells. CCR5 mediates the migration of macrophages to inflammatory areas facilitating the release of inflammatory cytokines and the amplification of immune responses (Iannaccone et al., 2020). GO enrichment analysis includes G protein-coupled receptor activity. KEGG enrichment analysis includes selective expression of chemokine receptors and autophagic pathways during T cell polarization. Activation of NLRP3 as a key component of the innate immune system plays a critical role in host defense against bacteria, fungi and viruses, and NLRP3 is also associated with metabolic and inflammatory conditions. GO enrichment includes peptidoglycan binding. KEGG enrichment analysis includes activation of NF-KappaB by PKR and inflammatory vesicles.

CCR5 may play a role in the inflammatory response to coronavirus infection. Study has been suggested that the CCR5 pathway is a suppressor of immune hyperactivation in severe COVID-19 (Cuesta-Llavona et al., 2021; Dieter et al., 2022). Lungs infected with SARS-COV-2 show upregulation of chemokines (including CCL4, CCL8, and CCL11), and all of these factors share CCR5 as their receptor (Hachim et al., 2020; Zeng et al., 2022). The active ingredient of LHQW can inhibit the secretion of CCL5 thereby inhibiting the interaction between CCL5 and CCR5, and suppressing the inflammatory response (Ko et al., 2006).

NLR Family Pyrin Domain Containing 3 (NLRP3) inflammatory vesicles are one of the most important components of the innate immune system and NLRP3 significantly enhances inflammation by increasing the production of IL-18 and gasdermin D (GSDMD) (López-Reyes et al., 2020; Freeman and Swartz, 2020; Saeedi-Boroujeni et al., 2021). The immune overreaction and cytokine storm in SARS-CoV-2 infection may be associated with NLRP3 inflammatory vesicle activation. In SARS-CoV-2 infection, NLRP3 inflammatory vesicle activation leads to the stimulation and synthesis of natural killer cells (NKs), NF- κ B and interferon γ (INF- γ) (Batiha et al., 2021; Rodrigues et al., 2021). The active ingredient of LHQW inhibits NLRP3 inflammatory vesicles (Liu et al., 2019), and LHQW blocks the onset of apoptosis and

inflammatory response by inhibiting the activation of NLRP3 (Chao et al., 2022).

Therefore, Licopyranocoumarin may impede release of inflammatory factors and spread of inflammatory response through CCR5 and NLRP3 thereby controlling the extent of inflammatory effects and the degree of tissue damage.

3-3-Oxopropanoic acid may regulate the differentiation of inflammatory cells and cell survival through CSF2

Granulocyte macrophage colony-stimulating factor 2 (CSF2) is a key cytokine. CSF2 affects the survival, proliferation and differentiation of dendritic cells and macrophages by stimulating myeloid cells (Sielska et al., 2020; Saita et al., 2022). GO enrichment analysis includes inflammatory factor activity. KEGG enrichment analysis includes selective expression of chemokine receptors during T cell polarization and dendritic cell developmental lineage pathways.

Study has been shown that blockade of CSF2 signaling reduces airway inflammation and hyperresponsiveness in mouse models of environmentally induced lung injury and asthma (Burkhardt et al., 2012; Gilchrist et al., 2021). Lung tissue-resident memory-like TH17 cells are predominant immune cell type in bronchoalveolar lavage fluid (BAL) expressing the cytokine GM-CSF. GM-CSF were significantly elevated in the serum of patients with severe COVID-19 compared to patients with normal COVID19. Study has been shown that an enhanced frequency of GM-CSF/IFNG replicating T cells was found in the blood of COVID-19 patients and appeared to correlate with disease activity. Zhao et al. suggested that CSF2/GM-CSF expressing cells were present in the lung and co-express IL17A. Lung T+RM17 cells are a potential coordinating factor for excessive inflammation in severe COVID-19 (Schett et al., 2020; Zhao et al., 2021). Li et al. identified enhanced Th17 cell differentiation and cytokine responses in COVID-19 (Li et al., 2021). Some studies have analyzed and identified genes specifically expressed by SARS-CoV-2 infection, as well as genes altered due to coronavirus-2 and/or other respiratory viral infections. In particular, CSF2 expression appears to be associated with neo-coronavirus disease (Chandrashekar et al., 2021; Ferrarini et al., 2021). Cheng et al. determined that the cytokine gene CSF2 was upregulated after SARS-CoV-2 infection (Cheng et al., 2021). SARS-CoV-2 infection can activate T cells. Activated T cells rapidly proliferate and secrete granulocyte-macrophage colonystimulating factor (GM-CSF) and interleukin-16 (IL-16) (Shen and Yin, 2021).

Therefore, 3-3-Oxopropanoic acid may regulate the differentiation of inflammatory cells and cell survival through CSF2 thereby reducing apoptosis.

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Conclusion

We explored the mechanism of *Lianhua Qingwen granules* (LHQW) treats COVID-19 by molecular docking and molecular dynamics. We found that the active ingredients in LHQW not only reduce cell damage and tissue destruction by inhibiting the inflammatory response through CSF2, CXCL8, CCR5 and IFNG, but also regulate cell survival and growth through NLRP3 and TNF.

Therefore, the active ingredients in *Lianhua Qingwen* granules (LHQW) treats COVID-19 by regulating cell survival and inhibiting inflammatory response from multiple targets.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/supplementary material.

Author contributions

Author contributions: J-FC, YG, MW, LX, XZ (12th author) contributed to the conception of the study. J-FC, MW, SC, HH, XZ (7th author), Y-CP, X-FS, JQ contributed significantly to analysis and manuscript preparation. J-FC, YG, MW, LX

References

Adegbola, P. I., Semire, B., Fadahunsi, O. S., and Adegoke, A. E. (2021). Molecular docking and ADMET studies of allium cepa, azadirachta indica and xylopia aethiopica isolates as potential anti-viral drugs for covid-19. *Virusdisease* 32 (1), 85–97. doi: 10.1007/s13337-021-00682-7

Al-Khafaji, K., and Taskin Tok, T. (2020). Molecular dynamics simulation, free energy landscape and binding free energy computations in exploration the antiinvasive activity of amygdalin against metastasis. *Comput. Methods Programs. Biomed.* 195, 105660. doi: 10.1016/j.cmpb.2020.105660

Apriyanti, E., Satari, M. H., and Kurnia, D. (2021). Potential of MurA enzyme and GBAP in fsr quorum sensing system as antibacterial drugs target: *In vitro* and *in silico* study of antibacterial compounds from myrmecodia pendans. *Comb. Chem. High Throughput. Screen.* 24 (1), 109–118. doi: 10.2174/1386207323666200628111348

Attiq, A., Yao, L., Afzal, S., and Khan, M. A. (2021). The triumvirate of NF-κB, inflammation and cytokine storm in COVID-19. *Int. Immunopharmacol.* 101, 108255. doi: 10.1016/j.intimp.2021.108255

Balkwill, F. (2006). TNF-alpha in promotion and progression of cancer. *Cancer Metastasis. Rev.* 25, 409–416. doi: 10.1007/s10555-006-9005-3

Batiha, G. E., Al-Gareeb, D. A. I., Qusti, S., Alshammari, E. M., Rotimi, D., Adeyemi, O. S., et al. (2021). Common NLRP3 inflammasome inhibitors and covid-19: Divide and conquer. *Sci. Afr.*, e01084. doi: 10.1016/j.sciaf.2021.e01084

Bui, L. M., Phung, H.T. T., Ho Thi, T. T., Singh, V., Maurya, R., Khambhati, K., et al. (2021). Recent findings and applications of biomedical engineering for COVID-19 diagnosis: a critical review. *Bioengineered* 12, 8594–8613. doi: 10.1080/21655979.2021.1987821

Burkhardt, J., Kirsten, H., Wolfram, G., Quente, E., and Ahnert, P. (2012). Differential allelic expression of IL13 and CSF2 genes associated with asthma. *Genet. Mol. Biol.* 35, 567–574. doi: 10.1590/S1415-47572012005000055

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Cao, J.-F., Gong, Y., Wu, M., Yang, X., Xiong, L., Chen, S., et al. (2022c). Exploring the mechanism of action of licorice in the treatment of COVID-19 through bioinformatics analysis and molecular dynamics simulation. *Front. Pharmacol* 13, 1003310. doi: 10.3389/fphar.2022.1003310

Cao, J.-F., Hu, X., Xiong, L., Wu, M., Yang, X., Wang, C., et al. (2022a). Interference of interleukin-1 β mediated by lentivirus promotes functional recovery of spinal cord contusion injury in rats *via* the PI3K/AKT1 signaling pathway. *Mediators Inflammation* 2022, 6285099. doi: 10.1155/2022/6285099

Cao, J.-F., Yang, X., Xiong, L., Wu, M., Chen, S., Xiong, C., et al. (2022b). Mechanism of n-0385 blocking SARS-CoV-2 to treat COVID-19 based on molecular docking and molecular dynamics. *Front. Microbiol* 13, 1013911. doi: 10.3389/fmicb.2022.1013911

Cao, J.-F., Yang, X., Xiong, L., Wu, M., Chen, S., Xu, H., et al. (2022d). Exploring the mechanism of action of dapansutrile in the treatment of gouty arthritis based on molecular docking and molecular dynamics. *Front. Physiol* 13, 990469. doi: 10.3389/fphys.2022.990469

Chandrashekar, D. S., Athar, M., Manne, U., and Varambally, S. (2021). Comparative transcriptome analyses reveal genes associated with SARS-CoV-2 infection of human lung epithelial cells. *Sci. Rep.* 11, 16212. doi: 10.1038/s41598-021-95733-w

Chao, L., Lin, J., Zhou, J., Du, H., Chen, X., Liu, M., et al. (2022). Polyphenol rich forsythia suspensa extract alleviates DSS-induced ulcerative colitis in mice through the Nrf2-NLRP3 pathway. *Antioxid. (Basel).* 11 (3), 475. doi: 10.3390/antiox11030475

Cheng, L. C., Kao, T. J., Phan, N. N., Chiao, C. C., Yen, M. C., Chen, C. F., et al. (2021). Novel signaling pathways regulate SARS-CoV and SARS-CoV-2 infectious disease. *Med. (Baltimore).* 100, e24321. doi: 10.1097/MD.00000000024321

Chouchana, L., Preta, L. H., Tisseyre, M., Terrier, B., Treluyer, J. M., and Montastruc, F. (2021). Kidney disorders as serious adverse drug reactions of remdesivir in coronavirus disease 2019: a retrospective case-noncase study. *Kidney Int.* 99, 1235–1236. doi: 10.1016/j.kint.2021.02.015

Choudhary, S., Sharma, K., and Silakari, O. (2021). The interplay between inflammatory pathways and COVID-19: A critical review on pathogenesis and therapeutic options. *Microb. Pathog.* 150, 104673. doi: 10.1016/j.micpath.2020.104673

Ciotti, M., Benedetti, F., Zella, D., Angeletti, S., Ciccozzi, M., and Bernardini, S. (2021). SARS-CoV-2 infection and the COVID-19 pandemic emergency: The importance of diagnostic methods. *Chemotherapy* 66, 17–23. doi: 10.1159/000515343

Citarella, A., Scala, A., Piperno, A., and Micale., N. (2021). SARS-CoV-2 m(pro): A potential target for peptidomimetics and small-molecule inhibitors. *Biomolecules* 11 (4), 607. doi: 10.3390/biom11040607

Cuesta-Llavona, E., Gómez, J., Albaiceta, G. M., Amado-Rodríguez, L., García-Clemente, M., Gutiérrez-Rodríguez, J., et al. (2021). Variant-genetic and transcriptexpression analysis showed a role for the chemokine-receptor CCR5 in COVID-19 severity. *Int. Immunopharmacol.* 98, 107825. doi: 10.1016/j.intimp.2021.107825

DasGupta, D., Chan, W. K. B., and Carlson, H. A. (2022). Computational identification of possible allosteric sites and modulators of the SARS-CoV-2 main protease. *J. Chem. Inf. Model.* 62 (3), 618–626. doi: 10.1021/acs.jcim.1c01223

Dasmahapatra, U., Kumar, C. K., Das, S., Subramanian, P. T., Murali, P., Isaac, A. E., et al. (2022). *In-silico* molecular modelling, MM/GBSA binding free energy and molecular dynamics simulation study of novel pyrido fused imidazo[4,5-c] quinolines as potential anti-tumor agents. *Front. Chem.* 10, 991369. doi: 10.3389/ fchem.2022.991369

Dela Cruz, J. M. D., Dones, S. A. A., Villanueva, R. C., Labrador, A. M., and Santiago-Bautista, M. R. (2022). Molecular docking and *in silico* pharmacological screening of oleosin from Cocos nucifera complexed with tamoxifen in developing potential breast chemotherapeutic leads. *Asian Pac. J. Cancer Prev.* 23 (7), 2421– 2430. doi: 10.31557/APICP.2022.23.7.2421

Diao, B., Wang, C., Tan, Y., Chen, X., Liu, Y., Ning, L., et al. (2020). Reduction and functional exhaustion of T cells in patients with coronavirus disease 2019 (COVID-19). *Front. Immunol.* 11, 827. doi: 10.3389/fimmu.2020.00827

Dieter, C., Brondani, L. A., Leitão, C. B., Gerchman, F., Lemos, N. E., and Crispim, D. (2022). Genetic polymorphisms associated with susceptibility to COVID-19 disease and severity: A systematic review and meta-analysis. *PloS One* 17, e0270627. doi: 10.1371/journal.pone.0270627

Dong, L., Xia, J. W., Gong, Y., Chen, Z., Yang, H. H., Zhang, J., et al. (2014). Effect of lianhuaqingwen capsules on airway inflammation in patients with acute exacerbation of chronic obstructive pulmonary disease. *Evid. Based. Complement. Alternat. Med.* 2014, 637969. doi: 10.1155/2014/637969

Fara, A., Mitrev, Z., Rosalia, R. A., and Assas, B. M. (2020). Cytokine storm and COVID-19: a chronicle of pro-inflammatory cytokines. *Open Biol.* 10, 200160. doi: 10.1098/rsob.200160

Fernandes, Q., Inchakalody, V. P., Merhi, M., Mestiri, S., Taib, N., Moustafa Abo El-Ella, D., et al. (2022). Emerging COVID-19 variants and their impact on SARS-CoV-2 diagnosis, therapeutics and vaccines. *Ann. Med.* 54, 524–540. doi: 10.1080/ 07853890.2022.2031274

Ferrarini, M. G., Lal, A., Rebollo, R., Gruber, A. J., Guarracino, A., Gonzalez, I. M., et al. (2021). Genome-wide bioinformatic analyses predict key host and viral factors in SARS-CoV-2 pathogenesis. *Commun. Biol.* 4, 590. doi: 10.1038/s42003-021-02095-0

Freeman, T. L., and Swartz, T. H. (2020). Targeting the NLRP3 inflammasome in severe COVID-19. *Front. Immunol.* 11, 1518. doi: 10.3389/fimmu.2020.01518

Gilchrist, S. E., Pennelli, G. M., and Hafizi, S. (2021). Gas6/TAM signalling negatively regulates inflammatory induction of GM-CSF in mouse brain microglia. *Cells* 10 (12), 3281. doi: 10.3390/cells10123281

Günther, S., Reinke, P. Y. A., Fernández-García, Y., Lieske, J., Lane, T. J., and Ginn, H. M. (2021). X-Ray screening identifies active site and allosteric inhibitors of SARS-CoV-2 main protease. *Science* 372 (6542), 642–646. doi: 10.1126/science.abf7945

Hachim, M. Y., Hachim, I. Y., Naeem, K. B., Hannawi, H., Al Salmi, I., and Hannawi, S. (2020). C-c chemokine receptor type 5 links COVID-19, rheumatoid arthritis, and hydroxychloroquine: *in silico* analysis. *Transl. Med. Commun.* 5, 14. doi: 10.1186/s41231-020-00066-x

Ha, H., Debnath, B., and Neamati, N. (2017). Role of the CXCL8-CXCR1/2 axis in cancer and inflammatory diseases. *Theranostics* 7 (6), 1543–1588. doi: 10.7150/ thno.15625

Hadj Hassine, I. (2022). Covid-19 vaccines and variants of concern: A review. *Rev. Med. Virol.* 32, e2313. doi: 10.1002/rmv.2313

Hassanipour, S., Arab-Zozani, M., Amani, B., Heidarzad, F., Fathalipour, M., and Martinez-de-Hoyo, R. (2021). The efficacy and safety of favipiravir in treatment of COVID-19: a systematic review and meta-analysis of clinical trials. *Sci. Rep.* 11, 11022. doi: 10.1038/s41598-021-90551-6

Hu, K., Guan, W. J., Bi, Y., Zhang, W., Li, L., Zhang, B., et al. (2021). Efficacy and safety of lianhuaqingwen capsules, a repurposed Chinese herb, in patients with coronavirus disease 2019: A multicenter, prospective, randomized controlled trial. *Phytomedicine* 85, 153242. doi: 10.1016/j.phymed.2020.153242

Iannaccone, G., Scacciavillani, R., Del Buono, M. G., Camilli, M., Ronco, C., Lavie, C. J., et al. (2020). Weathering the cytokine storm in COVID-19: Therapeutic implications. *Cardiorenal. Med.* 10, 277–287. doi: 10.1159/000509483

Idriss, H. T., and Naismith, J.H. (2000). TNF alpha and the TNF receptor superfamily: structure-function relationship(s). *Microsc. Res. Tech.* 50, 184–195. doi: 10.1002/1097-0029(20000801)50:3<184::AID-JEMT2>3.0.CO;2-H

Ilkhani, H., Hedayat, N., and Farhad, S. (2021). Novel approaches for rapid detection of COVID-19 during the pandemic: A review. *Anal. Biochem.* 634, 114362. doi: 10.1016/j.ab.2021.114362

Jagdish, S. P., Berteotti, A., Ronsisvalle, S., Rocchia, W., and Cavalli, A. (2014). Steered molecular dynamics simulations for studying protein–ligand interaction in cyclin-dependent kinase 5. *J. Chem. Inf. Modeling.* 54 (2), 470–480. doi: 10.1021/ci4003574

Jagdish, S. P., and Marty Ytreberg, F. (2019). Calculation of protein-protein binding free energies using umbrella sampling with dual resolution water models. *Biophys. J.* 116 (3), 291a. doi: 10.1016/j.bpj.2018.11.1571

Kaiser, R., Leunig, A., Pekayvaz, K., Popp, O., Joppich, M., Polewka, V., et al. (2021). Self-sustaining IL-8 loops drive a prothrombotic neutrophil phenotype in severe COVID-19. *JCI Insight* 6 (18), e150862. doi: 10.1172/jci.insight.150862

Kashyap, D., Jakhmola, S., Tiwari, D., Kumar, R., Moorthy, N. S. H. N., Elangovan, M., et al. (2021). Plant derived active compounds as potential anti SARS-CoV-2 agents: an *in-silico* study. *J. Biomol. Struct. Dyn.* 6, 1–22. doi: 10.1080/ 07391102.2021.1947384

Khanmohammadi, S., and Rezaei, N. (2021). Role of toll-like receptors in the pathogenesis of COVID-19. J. Med. Virol. 93, 2735-2739. doi: 10.1002/jmv.26826

Ko, H. C., Wei, B. L., and Chiou, W. F. (2006). The effect of medicinal plants used in Chinese folk medicine on RANTES secretion by virus-infected human epithelial cells. *J. Ethnopharmacol.* 107, 205–210. doi: 10.1016/j.jep.2006.03.004

Krämer, B., Knoll, R., Bonaguro, L., ToVinh, M., Raabe, J., Astaburuaga-García, R., et al. (2021). Early IFN- α signatures and persistent dysfunction are distinguishing features of NK cells in severe COVID-19. *Immunity* 54, 2650–69.e14. doi: 10.1016/j.immuni.2021.09.002

Kumar, D., Meena, M. K., Kumari, K., Patel, R., Jayaraj, A., and Singh, P. (2020). *In-silico* prediction of novel drug-target complex of nsp3 of CHIKV through molecular dynamic simulation. *Heliyon* 6 (8), e04720. doi: 10.1016/ j.heliyon.2020.e04720

Lee, T. S., Allen, B. K., Giese, T. J., Guo, Z., Li, P., Lin, C., et al. (2020). Alchemical binding free energy calculations in AMBER20: Advances and best practices for drug discovery. *J. Chem. Inf. Model.* 60, 5595–5623. doi: 10.1021/ acs.jcim.0c00613

Lee, T. S., Cerutti, D. S., Mermelstein, D., Lin, C., LeGrand, S., Giese, T. J., et al. (2018). GPU-Accelerated molecular dynamics and free energy methods in Amber18: Performance enhancements and new features. *J. Chem. Inf. Model.* 58 (10), 2043–2050. doi: 10.1021/acs.jcim.8b00462

Lee, J., Hitzenberger, M., Rieger, M., Kern, N. R., Zacharias, M., and Im, W. (2020). CHARMM-GUI supports the amber force fields. *J. Chem. Phys.* 153 (3), 035103. doi: 10.1063/5.0012280

Li, Z., Chen, H., Zhang, H., Li, Y., Wang, C., Bai, L., et al. (2020). Similarity and specificity of traditional Chinese medicine formulas for management of coronavirus disease 2019 and rheumatoid arthritis. *ACS Omega*. 5, 30519–30530. doi: 10.1021/acsomega.0c04377

Li, Z., Huang, Z., Li, X., Huang, C., Shen, J., Li, S., et al. (2021). Bioinformatic analyses hinted at augmented T helper 17 cell differentiation and cytokine response as the central mechanism of COVID-19-associated Guillain-Barré syndrome. *Cell Prolif.* 54, e13024. doi: 10.1111/cpr.13024

Liu, P., Bai, X., Zhang, T., Zhou, L., Li, J., and Zhang, L. (2019). The protective effect of lonicera japonica polysaccharide on mice with depression by inhibiting NLRP3 inflammasome. *Ann. Transl. Med.* 7, 811. doi: 10.21037/atm.2019.12.64

Liu, M., Gao, Y., Yuan, Y., Yang, K., Shi, S., Tian, J., et al. (2021). Efficacy and safety of herbal medicine (Lianhuaqingwen) for treating COVID-19: A systematic review and meta-analysis. *Integr. Med. Res.* 10, 100644. doi: 10.1016/j.imr.2020.100644

Liu, Q., Li, A., Tian, Y., Wu, J. D., Liu, Y., Li, T., et al. (2016). The CXCL8-CXCR1/2 pathways in cancer. *Cytokine Growth Factor. Rev.* 31, 61–71. doi: 10.1016/j.cytogfr.2016.08.002

Liu, C., Yin, Z., Feng, T., Zhang, M., Zhou, Z., and Zhou, Y. (2021). An integrated network pharmacology and RNA-seq approach for exploring the preventive effect of lonicerae japonicae flos on LPS-induced acute lung injury. *J. Ethnopharmacol.* 264, 113364. doi: 10.1016/j.jep.2020.113364

Li, L., Zhang, Z., Zhou, W., Chen, J., Jin, H., Fang, H., et al. (2020). Lianhua qingwen prescription for coronavirus disease 2019 (COVID-19) treatment: Advances and prospects. *BioMed. Pharmacother.* 130, 110641. doi: 10.1016/j.biopha.2020.110641

López-Reyes, A., Martinez-Armenta, C., Espinosa-Velázquez, R., Vázquez-Cárdenas, P., Cruz-Ramos, M., Palacios-Gonzalez, B., et al. (2020). NLRP3 inflammasome: The stormy link between obesity and COVID-19. *Front. Immunol.* 11, 570251. doi: 10.3389/fimmu.2020.570251

Mahmoudinobar, F., Britton, D., and Montclare, J. K. (2021). Protein-based lateral flow assays for COVID-19 detection. *Protein Eng. Des. Sel.* 34, gzab010. doi: 10.1093/protein/gzab010

Martinez-Lopez, A., Cuenca-Barrales, C., Montero-Vilchez, T., Molina-Leyva, A., and Arias-Santiago, S. (2020). Review of adverse cutaneous reactions of pharmacologic interventions for COVID-19: A guide for the dermatologist. J. Am. Acad. Dermatol. 83, 1738–1748. doi: 10.1016/j.jaad.2020.08.006

Mele, D., Calastri, A., Maiorano, E., Cerino, A., Sachs, M., Oliviero, B., et al. (2021). High frequencies of functional virus-specific CD4(+) T cells in SARS-CoV-2 subjects with olfactory and taste disorders. *Front. Immunol.* 12, 748881. doi: 10.3389/fimmu.2021.748881

Melgaço, J. G., Azamor, T., Silva, A. M. V., Linhares, J. H. R., Dos Santos, T. P., Mendes, Y. S., et al. (2021). Two-step *In vitro* model to evaluate the cellular immune response to SARS-CoV-2. *Cells* 10 (9), 2206. doi: 10.3390/cells10092206

Meo, S. A., Bukhari, I. A., Akram, J. A., Meo, S., and Klonoff, D. C. (2021). COVID-19 vaccines: comparison of biological, pharmacological characteristics and adverse effects of Pfizer/BioNTech and moderna vaccines. *Eur. Rev. Med. Pharmacol. Sci.* 25, 1663–1669. doi: 10.26355/eurrev_202102_24877

Mermelstein, D. J., Lin, C., Nelson, G., Kretsch, R., McCammon, J. A., and Walker, R. C. (2018). Fast and flexible gpu accelerated binding free energy calculations within the amber molecular dynamics package. *J. Comput. Chem.* 39, 1354–1358. doi: 10.1002/jcc.25187

Mok, T. N., He, Q., Zhang, X., Sin, T. H., Wang, H., Hou, H., et al. (2022). Effects of 6-hydroxykaempferol: A potential natural product for amelioration of tendon impairment. *Front. Pharmacol.* 13, 919104. doi: 10.3389/fphar.2022.919104

Ng, X. L., Betzler, B. K., Testi, I., Ho, S. L., Tien, M., Ngo, W. K., et al. (2021). Ocular adverse events after COVID-19 vaccination. *Ocul. Immunol. Inflamm.* 29, 1216–1224. doi: 10.1080/09273948.2021.1976221

Norooznezhad, A. H., and Mansouri, K. (2021). Endothelial cell dysfunction, coagulation, and angiogenesis in coronavirus disease 2019 (COVID-19). *Microvasc. Res.* 137, 104188. doi: 10.1016/j.mvr.2021.104188

Nöst, X., Pferschy-Wenzig, E. M., Nikles, S., He, X., Fan, D., Lu, A., et al. (2019). Identification of constituents affecting the secretion of pro-inflammatory cytokines in LPS-induced U937 cells by UHPLC-HRMS-Based metabolic profiling of the traditional Chinese medicine formulation huangqi jianzhong tang. *Molecules* 24 (17), 3116. doi: 10.3390/molecules24173116

Ozger, H. S., Dizbay, M., Corbacioglu, S. K., Aysert, P., Demirbas, Z., Tunccan, O. G., et al. (2021). The prognostic role of neopterin in COVID-19 patients. *J. Med. Virol.* 93, 1520–1525. doi: 10.1002/jmv.26472

Panikar, S., Shoba, G., Arun, M., Sahayarayan, J. J., Usha Raja Nanthini, A., Chinnathambi, A., et al. (2021). 'Essential oils as an effective alternative for the treatment of COVID-19: Molecular interaction analysis of protease (M(pro)) with pharmacokinetics and toxicological properties'. *J. Infect. Public Health* 14, 601–610. doi: 10.1016/j.jiph.2020.12.037

Petrone, L., Petruccioli, E., Vanini, V., Cuzzi, G., Gualano, G., Vittozzi, P., et al. (2021). Coinfection of tuberculosis and COVID-19 limits the ability to *in vitro* respond to SARS-CoV-2. *Int. J. Infect. Dis.* 113 Suppl 1, S82–s87. doi: 10.1016/j.ijid.2021.02.090

Pons, S., Fodil, S., Azoulay, E., and Zafrani, L. (2020). The vascular endothelium: the cornerstone of organ dysfunction in severe SARS-CoV-2 infection. *Crit. Care* 24, 353. doi: 10.1186/s13054-020-03062-7

Procacci, P. (2020). Methodological uncertainties in drug-receptor binding free energy predictions based on classical molecular dynamics. *Curr. Opin. Struct. Biol.* 67, 127–134. doi: 10.1016/j.sbi.2020.08.001

Qasem, A., Shaw, A. M., Elkamel, E., and Naser, S. A. (2021). Coronavirus disease 2019 (COVID-19) diagnostic tools: A focus on detection technologies and limitations. *Curr. Issues Mol. Biol.* 43, 728–748. doi: 10.3390/cimb43020053

Ramasamy, S., and Subbian, S. (2021). Critical determinants of cytokine storm and type I interferon response in COVID-19 pathogenesis. *Clin. Microbiol. Rev.* 34 (4), e0016321. doi: 10.1128/CMR.00299-20

Rello, J., Belliato, M., Dimopoulos, M. A., Giamarellos-Bourboulis, E. J., Jaksic, V., Martin-Loeches, I., et al. (2020). Update in COVID-19 in the intensive care unit from the 2020 HELLENIC Athens international symposium. *Anaesth. Crit. Care Pain Med.* 39, 723–730. doi: 10.1016/j.accpm.2020.10.008

Rieloff, E., and Skepö, M. (2021). Molecular dynamics simulations of phosphorylated intrinsically disordered proteins: A force field comparison. *Int. J. Mol. Sci.* 22 (18), 10174. doi: 10.3390/ijms221810174

Rodrigues, T. S., de Sá, K. S. G., Ishimoto, A. Y., Becerra, A., Oliveira, S., Almeida, L., et al. (2021). Inflammasomes are activated in response to SARS-CoV-2 infection and are associated with COVID-19 severity in patients. *J. Exp. Med.* 218 (3), e20201707. doi: 10.1084/jem.20201707

Roe, M. K., Junod, N. A., Young, A. R., Beachboard, D. C., and Stobart, C. C. (2021). Targeting novel structural and functional features of coronavirus protease nsp5 (3CL(pro), m(pro)) in the age of COVID-19. *J. Gen. Virol.* 102 (3), 001558. doi: 10.1099/jgv.0.001558

Runfeng, L., Yunlong, H., Jicheng, H., Weiqi, P., Qinhai, M., Yongxia, S., et al. (2020). Lianhuaqingwen exerts anti-viral and anti-inflammatory activity against novel coronavirus (SARS-CoV-2). *Pharmacol. Res.* 156, 104761. doi: 10.1016/j.phrs.2020.104761

Sabbah, D. A., Hajjo, R., Bardaweel, S. K., and Zhong, H. A. (2021). An updated review on SARS-CoV-2 main proteinase (M(Pro)): Protein structure and small-molecule inhibitors. *Curr. Top. Med. Chem.* 21, 442–460. doi: 10.2174/1568026620666201207095117

Saeedi-Boroujeni, A., Mahmoudian-Sani, M. R., Nashibi, R., Houshmandfar, S., Tahmaseby Gandomkari, S., and Khodadadi, A. (2021). Tranilast: a potential antiinflammatory and NLRP3 inflammasome inhibitor drug for COVID-19. *Immunopharmacol. Immunotoxicol.* 43, 247-258. doi: 10.1080/ 08923973.2021.1925293

Saita, K., Moriuchi, Y., Iwagawa, T., Aihara, M., Takai, Y., Uchida, K., et al. (2022). Roles of CSF2 as a modulator of inflammation during retinal degeneration. *Cytokine* 158, 155996. doi: 10.1016/j.cyto.2022.155996

Schett, G., Manger, B., Simon, D., and Caporali, R. (2020). COVID-19 revisiting inflammatory pathways of arthritis. *Nat. Rev. Rheumatol.* 16, 465–470. doi: 10.1038/s41584-020-0451-z

Schoenborn, J. R., and Wilson, C. B. (2007). Regulation of interferon-gamma during innate and adaptive immune responses. *Adv. Immunol.* 96, 41–101. doi: 10.1016/S0065-2776(07)96002-2

Shahbazi, M., Jafari, M., Moulana, Z., Sepidarkish, M., Bagherzadeh, M., Rezanejad, M., et al. (2021). Reduced frequency of T helper 17 and T helper 1 cells and their association with critical coronavirus disease 2019. *Apmis* 129, 271–279. doi: 10.1111/apm.13129

Shen, P., Li, J., Tu, S., Wu, Y., Peng, Y., Chen, G., et al. (2021). Positive effects of lianhuaqingwen granules in COVID-19 patients: A retrospective study of 248 cases. *J. Ethnopharmacol.* 278, 114220. doi: 10.1016/j.jep.2021.114220

Shen, X., and Yin, F. (2021). The mechanisms and clinical application of traditional Chinese medicine lianhua-qingwen capsule. *BioMed. Pharmacother.* 142, 111998. doi: 10.1016/j.biopha.2021.111998

Shi, M., Peng, B., Li, A., Li, Z., Song, P., Li, J., et al. (2021). Broad anti-viral capacities of lian-Hua-Qing-Wen capsule and jin-Hua-Qing-Gan granule and rational use against COVID-19 based on literature mining. *Front. Pharmacol.* 12, 640782. doi: 10.3389/fphar.2021.640782

Sidiq, Z., Hanif, M., Dwivedi, K. K., and Chopra, K. K. (2020). Benefits and limitations of serological assays in COVID-19 infection. *Indian J. Tuberc.* 67, S163–Ss66. doi: 10.1016/j.ijtb.2020.07.034

Sielska, M., Przanowski, P., Pasierbińska, M., Wojnicki, K., Poleszak, K., Wojtas, B., et al. (2020). Tumour-derived CSF2/granulocyte macrophage colony stimulating factor controls myeloid cell accumulation and progression of gliomas. *Br. J. Cancer.* 123, 438–448. doi: 10.1038/s41416-020-0862-2

Song, L. F., Lee, T. S., Zhu, C., York, D. M., and Merz, K. M.Jr. (2019). Using AMBER18 for relative free energy calculations. *J. Chem. Inf. Model.* 59 (7), 3128–3135. doi: 10.1021/acs.jcim.9b00105

Stelzer, G., Rosen, N., Plaschkes, I., Zimmerman, S., Twik, M., Fishilevich, S., et al. (2016). The GeneCards suite: From gene data mining to disease genome sequence analyses. *Curr. Protoc. Bioinf.* 54, 1.30.1–1.30.33. doi: 10.1002/cpbi.5

Strömich, L., Wu, N., Barahona, M., and Yaliraki, S. N. (2022). Allosteric hotspots in the main protease of SARS-CoV-2. *J. Mol. Biol.* 434 (17), 167748. doi: 10.1016/j.jmb.2022.167748

Sultan, A., Ali, R., Sultan, T., Ali, S., Khan, N. J., and Parganiha., A. (2021). Circadian clock modulating small molecules repurposing as inhibitors of SARS-CoV-2 m(pro) for pharmacological interventions in COVID-19 pandemic. *Chronobiol. Int.* 38, 971–985. doi: 10.1080/07420528.2021.1903027

Szklarczyk, D., Gable, A. L., Nastou, K. C., Lyon, D., Kirsch, R., Pyysalo, S., et al. (2021). The STRING database in 2021: customizable protein-protein networks, and functional characterization of user-uploaded gene/measurement sets. *Nucleic Acids Res.* 49, D605–Dd12. doi: 10.1093/nar/gkab835

Sztain, T., Amaro, R., and McCammon, J. A. (2020). Elucidation of cryptic and allosteric pockets within the SARS-CoV-2 protease. bioRxiv [Preprint]. 2020.07.23.218784. J. Chem. Inf. Model 61 (7), 3495–3501. doi: 10.1101/2020.07.23.218784

Tong, T., Wu, Y., Ni, W., Shen, A., and Liu, S. (2020). The potential insights of traditional Chinese medicine on treatment of COVID-19. *Chin. Med.* 15, 51. doi: 10.1186/s13020-020-00326-w

Vitiello, A., Ferrara, F., Troiano, V., and La Porta, R. (2021). COVID-19 vaccines and decreased transmission of SARS-CoV-2. *Inflammopharmacology* 29, 1357– 1360. doi: 10.1007/s10787-021-00847-2

Wang, E., Fu, W., Jiang, D., Sun, H., Wang, J., Zhang, X., et al. (2021). VAD-MM/GBSA: A variable atomic dielectric MM/GBSA model for improved accuracy in protein-ligand binding free energy calculations. *J. Chem. Inf. Model.* 61 (6), 2844–2856. doi: 10.1021/acs.jcim.1c00091

Wang, D., Li, Z., and Liu, Y. (2020). An overview of the safety, clinical application and antiviral research of the COVID-19 therapeutics. J. Infect. Public Health 13, 1405–1414. doi: 10.1016/j.jiph.2020.07.004

Wang, D., Yu, M., Xie, W., Huang, L., Wei, J., and Lei, Y. (2022). Metaanalysis on the effect of combining lianhua qingwen with Western medicine to treat coronavirus disease 2019. *J. Integr. Med.* 20, 26–33. doi: 10.1016/ j.joim.2021.10.005

Weng, G., Wang, E., Chen, F., Sun, H., Wang, Z., and Hou, T. (2019). Assessing the performance of MM/PBSA and MM/GBSA methods. 9. prediction reliability of binding affinities and binding poses for protein-peptide complexes. *Phys. Chem. Chem. Phys.* 21, 10135–10145. doi: 10.1039/C9CP01674K

Wu, X., Sui, Z., Zhang, H., Wang, Y., and Yu, Z. (2020). Integrated analysis of IncRNA-mediated ceRNA network in lung adenocarcinoma. *Front. Oncol.* 10, 554759. doi: 10.3389/fonc.2020.554759

Wu, Y., You, X., Lin, Q., Xiong, W., Guo, Y., Huang, Z., et al. (2022). Exploring the pharmacological mechanisms of xihuang pills against prostate cancer via integrating network pharmacology and experimental validation *In vitro* and *In vivo*. *Front. Pharmacol.* 12, 791269. doi: 10.3389/fphar.2021.791269

Wu, Y., and Zhong, P. (2021). Clinical progress on management of pneumonia due to COVID-19 with Chinese traditional patent medicines. *Front. Pharmacol.* 12, 655063. doi: 10.3389/fphar.2021.655063

Xiao, M., Tian, J., Zhou, Y., Xu, X., Min, X., Lv, Y., et al. (2020). Efficacy of huoxiang zhengqi dropping pills and lianhua qingwen granules in treatment of COVID-19: A randomized controlled trial. *Pharmacol. Res.* 161, 105126. doi: 10.1016/j.phrs.2020.105126

Xu, X. R., Zhang, W., Wu, X. X., Yang, H. Q., Sun, Y. T., Pu, Y. T., et al. (2022). Analysis of mechanisms of shenhuang granule in treating severe COVID-19 based on network pharmacology and molecular docking. *J. Integr. Med* 20 (6), 561–74. doi: 10.1016/j.joim.2022.07.005

Yamamoto, K. (2022). Adverse effects of COVID-19 vaccines and measures to prevent them. *Virol. J.* 19, 100. doi: 10.1186/s12985-022-01831-0

Zeng, Z., Lan, T., Wei, Y., and Wei, X. (2022). CCL5/CCR5 axis in human diseases and related treatments. *Genes Dis.* 9, 12-27. doi: 10.1016/j.gendis.2021.08.004

Zeng, M., Li, L., and Wu, Z. (2020). Traditional Chinese medicine lianhua qingwen treating corona virus disease 2019(COVID-19): Meta-analysis of randomized controlled trials. *PloS One* 15, e0238828. doi: 10.1371/journal.pone.0238828

Zhang, X., Cao, D., Liu, J., Zhang, Q., and Liu, M. (2020). Efficacy and safety of lianhua qingwen combined with conventional antiviral Western medicine in the treatment of coronavirus disease (covid-19) in 2019: Protocol for a systematic review and meta-analysis. *Med. (Baltimore).* 99, e21404. doi: 10.1097/MD.00000000021404

Zhang, W., Chen, Y., Jiang, H., Yang, J., Wang, Q., Du, Y., et al. (2020). Integrated strategy for accurately screening biomarkers based on metabolomics coupled with network pharmacology. *Talanta* 211, 120710. doi: 10.1016/ j.talanta.2020.120710

Zhao, Y., Kilian, C., Turner, J. E., Bosurgi, L., Roedl, K., Bartsch, P., et al. (2021). Clonal expansion and activation of tissue-resident memory-like Th17 cells expressing GM-CSF in the lungs of severe COVID-19 patients. *Sci. Immunol.* 6 (56), eabf6692. doi: 10.1126/sciimmunol.abf6692

Zhou, M., Wang, Y., Lin, X., Wan, J., and Wen, C. (2020). Specific TLR4 blocking effect of a novel 3,4-dihydropyrimidinone derivative. *Front. Pharmacol.* 11, 624059. doi: 10.3389/fphar.2020.624059

Zhuang, J., Dai, X., Wu, Q., Cai, H., Fu, X., Zhang, W., et al. (2021). A metaanalysis for lianhua qingwen on the treatment of coronavirus disease 2019 (COVID-19). Complement. Ther. Med. 60, 102754. doi: 10.1016/j.ctim.2021.102754