Check for updates

OPEN ACCESS

EDITED BY Thuy Le, Duke University, United States

REVIEWED BY

Fatemeh Ahangarkani, Mazandaran University of Medical Sciences, Sari, Iran Eman A. Gouda M. Youssef, Lundquist Institute for Biomedical Innovation, United States

*CORRESPONDENCE Shima Aboutalebian Shima.aboutalebian@gmail.com

SPECIALTY SECTION

This article was submitted to Fungal Pathogenesis, a section of the journal Frontiers in Cellular and Infection Microbiology

RECEIVED 17 March 2022 ACCEPTED 23 August 2022 PUBLISHED 14 October 2022

CITATION

Erami M, Mirhendi H, Momen-Heravi M, Hezaveh SJH, Ahsaniarani AH, Sabet SS and Aboutalebian S (2022) A case of COVID-19-associated rhino-orbitocerebral mucormycosis caused by *Apophysomyces variabilis* with a review of the literature. *Front. Cell. Infect. Microbiol.* 12:898477.

doi: 10.3389/fcimb.2022.898477

COPYRIGHT

© 2022 Erami, Mirhendi, Momen-Heravi, Hezaveh, Ahsaniarani, Sabet and Aboutalebian. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

A case of COVID-19-associated rhino-orbito-cerebral mucormycosis caused by *Apophysomyces variabilis* with a review of the literature

Mahzad Erami^{1,2}, Hossein Mirhendi^{3,4}, Mansooreh Momen-Heravi², Seyed Jamal Hashemi Hezaveh¹, Amir Hossein Ahsaniarani⁵, Seddighe Sadat Sabet⁶ and Shima Aboutalebian^{3,4*}

¹Department of Medical Parasitology and Mycology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran, ²Infectious Disease Research Center, Kashan University of Medical Sciences, Kashan, Iran, ³Department of Medical Parasitology and Mycology, School of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran, ⁴Mycology Reference Laboratory, Research Core Facilities Laboratory, Isfahan University of Medical Sciences, Isfahan, Iran, ⁵Department Of Votriniolaryngology, School of Medicine, Matini Hospital, Kashan University of Medical Sciences, Kashan, Iran, ⁶Department of Pharmaceutical, Kashan University of Medical Sciences, Kashan, Iran,

A fatal case of COVID-19-associated mucormycosis (CAM) affected a 40-yearold woman who was initially admitted to our hospital due to a SARS-CoV-2 infection. Her clinical condition worsened, and she finally died because of respiratory failure, hemodynamic instability, and mucormycosis with invasion into the orbit and probably the brain. According to DNA sequence analysis of the fungus isolated from the patient, *Apophysomyces variabilis* was involved. This is the first published case of CAM and the third case of mucormycosis due to this mold.

KEYWORDS

mucormycosis, *apophysomyces variabilis*, COVID-19, immunocompetent patient, invasive fungal infection, antifungal susceptibility testing

Introduction

Mucormycosis (zygomycosis) is a disease caused by an opportunistic infection in humans and animals by members of the order Mucorales (Kwon-Chung, 2012; Binder et al., 2014). Mucormycosis is subdivided into the rhinocerebral, pulmonary, cutaneous, gastrointestinal, and disseminated types according to the distribution of lesions (Roden et al., 2005; Skiada et al., 2011) and mainly occurs in immunosuppressed hosts, including those with uncontrolled diabetes mellitus, hematological malignancies, transplant recipients, and patients receiving corticosteroid therapy (Farmakiotis and Kontoyiannis, 2016).

These infections have the capability of angio-invasion, inflicting vasculitis and thrombosis of vessels, which may result in massive areas of infarction and necrosis (Ibrahim et al., 2012). Regardless of its type, zygomycosis is an aggressive, life-threatening disease. As stated by the United States Centers for Disease Control and Prevention (CDC), this is an infrequent illness with an infection rate of <1% worldwide and with a 50% mortality (Dannaoui and Lackner, 2020); if left untreated, the mortality rate could be above 90% (Divakar, 2021).

Most severe cases of COVID-19 suffer from low oxygen levels, high iron levels, a hyperglycemic state, acidosis, and lowered phagocytic activity. These conditions facilitate opportunistic fungal infections, such as mucormycosis (Singh et al., 2021). Recently, there has been a dramatic increase in COVID-19-associated mucormycosis (CAM) cases, driven especially by low- and middle-income countries and most extensively reported in India. The mortality rate varies between 14% and 70%, depending on the site of infection (Patel et al., 2020).

Early diagnosis and treatment are critical to reduce mortality in patients with zygomycosis (Binder et al., 2014). In addition, precise species identification of the fungus involved is critical, not only to ensure accurate diagnosis but also for developing speciesspecific antifungal drugs, as well as to improve knowledge of the epidemiology of the disease. Various organisms have been implicated as etiological agents of mucormycosis, most commonly *Rhizopus*. Other etiological agents include *Mucor*, *Rhizomucor*, *Lichtheimia* (previously known as *Absidia*), *Apophysomyces*, *Saksenaea*, *Cunninghamella*, *Syncephalastrum*, and *Cokeromyces* (Gomes et al., 2011).

Here, a fatal case of rhino-orbito-cerebral mucormycosis (ROCM) due to *Apophysomyces variabilis* in a 40-year-old woman with COVID-19 is reported. To the authors' knowledge, this is the first case of CAM caused by *A. variabilis* in Iran. To better understand the clinical characteristics of the cases of ROCM due to non-*Rhizopus* Mucorales, we reviewed all published cases of ROCM that were supported by data from molecular species identification.

Case report

On 11 September 2021, a 40-year-old Afghan woman was hospitalized in Shahid Beheshti Hospital, Kashan, Iran, due to COVID-19. She had a fever, dry cough, dyspnea, hemoptysis, myalgia, and severe weakness, which had been ongoing for 5 days. Physical examination revealed a body temperature of 38.7°C, a blood pressure of 100/80 mm Hg, a respiratory rate of 26 breaths per minute, and an oxygen saturation of 90% on ambient air. Laboratory examination revealed leukopenia, lymphopenia, and a significant increase in C-reactive protein (CRP), and the patient underwent oxygen supplementation therapy. Nasopharyngeal and oropharyngeal swabs were obtained and subject to a real-time

reverse transcriptase polymerase chain reaction test using the Light Cycler 96-well system (Roche, Germany) for SARS-CoV-2 targeting N and RdRp genes (Pishtaz Teb, Tehran, Iran) (Fakhim et al. 2021). The sample was positive with a cycle threshold value of 25, indicating ongoing viral infection. Cultures of urine and blood were unremarkable. A computed tomography (CT) scan of the chest demonstrated multifocal bilateral mixed ground-glass opacity and consolidation, including central and peripheral distribution, and bilateral pleural effusion (Figure 1A). The patient was started on remdesivir (100 mg), dexamethasone (8 mg intravenously/twice daily and prednisolone (125 mg/day), and anticoagulants (heparin (QZD)). Because of increased hypoxia and progressive reduction in oxygen saturation (≈45%), the patient became severely ill and underwent mechanical ventilation, and treatment with intravenous tocilizumab (8 mg/ kg) was started. After 7 days of hospitalization, the patient developed right-side periorbital edema, proptosis of the right globe, necrosis of the hard palate, and nasal bleeding (Figures 1B, C). A paranasal and orbital CT scan demonstrated opacification and an air-fluid level in the bilateral sphenoid and ethmoid sinuses and the right maxilla along with mucosal thickening in the left maxilla and fat haziness in the right extraand intraconal locations (Figures 1D, E). Samples from wounds, tissue necrosis, and bloody secretions of the eye were taken and subjected to direct examination by microscopy using a potassium hydroxide preparation (Figure 2A), fungal culture, and histopathology (Figure 2B). The condition was diagnosed as a probable invasive mucormycosis. Intravenous liposomal amphotericin B 5 mg/kg/day was initiated. However, the patient was in an unstable condition; therefore, surgery and endoscopy of the sinus tissues and debridement were not possible. Despite antifungal and antibacterial therapy, the patient's situation deteriorated, and the patient died on the evening of day 11 due to respiratory failure, hemodynamic instability, and mucormycosis with involvement of the orbit and probably the brain.

The clinical samples obtained using sterile swabs and cultured on Sabouraud dextrose agar plates containing chloramphenicol (50 mg/L) (Biolife, Milan, Italy) yielded colonies of fluffy, cottony growth that filled the plate within 4 days. The colony surface was white when young, becoming cream-to-yellow or brownish-gray later on (Figure 3A). The reverse was white-to-pale yellow (Figure 3B). Microscopy of a slide prepared from the isolated colony revealed non-septate and branched hyphae, long and unbranched sporangiophores, and pear-shaped sporangia. Moreover, disintegration might have left a small collar at the foundation of the columella, and the sporangiospores were smooth and generally elongated and may have appeared as pale brown in mass (Figure 3C). Accurate morphologic identification at the species level was not possible; therefore, the isolate was subject to molecular identification. DNA was extracted using physical destruction of the cell wall by glass-bead manipulation followed by phenol-chloroform purification as described



(A) Chest computed tomography (CT) scan with multifocal bilateral mixed ground-glass opacity and consolidation, including central and peripheral distribution, and bilateral pleural effusion. (B, C) Swelling of the right eye and deep necrotic ulcer in the palate. (D, E) Paranasal and orbital CT scan demonstrated opacification and air-fluid level in the bilateral sphenoid and ethmoid sinuses and the right maxilla along with mucosal thickening in the left maxilla and fat haziness in the right extraconal and intraconal locations.

previously (Aboutalebian et al., 2021). The internal transcribed spacer (ITS) region was amplified using primer pairs ITS1 (TCCGTAGGTGAACCTGCGG) and ITS4 (TCCTCCGC TTATTGATATGC) (Aboutalebian et al., 2020). PCR products were purified and sequenced (Core Facilities Research Laboratory, Isfahan, Iran), and results were interpreted using NCBI BLAST (https://blast.ncbi.nlm.nih.gov/Blast.cgi) queries. The isolate was identified as *A. variabilis*, with a 99.86% sequence identity to the reference sequence. The sequence has been submitted to GenBank under the accession number OL804547.

Antifungal susceptibility testing (AFST) was performed based on the Clinical and Laboratory Standards Institute (CLSI) M38-A2 broth Dilution AFST (Wayne, 2008; Chakrabarti et al., 2010), and the minimum inhibitory concentrations were determined after 24 h of incubation at 35° C. *Candida parapsilosis* ATCC 22019 was used as quality control. AFST of the isolate against amphotericin B, fluconazole, itraconazole, voriconazole, posaconazole, and isavuconazole revealed minimum inhibitory concentration (MIC) values of 2, >64, 1, >16, 0.5, and 2 μ g/ml, respectively,



FIGURE 2

(A) Direct microscopic examination using 10% potassium hydroxide revealed broad, aseptate hyaline hyphae with wide-angle branching.
(B) Histopathologic examination with hematoxylin and eosin stain showing abundance in granulation tissue with foreign-body reaction and broad, aseptate hyphae in the intravascular space with wide-angle branching.



FIGURE 3

(A, B) Morphological characteristics of colonies on Sabouraud dextrose agar on the surface and the reverse after 4 days of incubation at 25°C. (C) Microscopic appearance of the isolate as observed on a slide prepared from the fungal culture.

and minimum effective concentration (MEC) for caspofungin was >8 μ g/ml. Written informed consent was obtained from the next of kin of the patient for the publication of any potentially identifiable images or data.

Discussion

We report the first case of CAM caused by *A. variabilis* occurring in an apparently immunocompetent patient with severe COVID-19, with fungal identification obtained using sequencing of the ITS region of ribosomal DNA.

Mucormycosis is the third most common opportunistic invasive fungal infection and is characterized by high morbidity and mortality in immunocompromised and immunocompetent hosts (Bouza et al., 2006; Kontoyiannis et al., 2012). It manifests with a range of symptoms, such as rhino-orbito-cerebral, rhinoorbital, and pulmonary mucormycosis, which are caused by aspiration of the spores. The infection is potentially angioinvasive; therefore, the resulting disease is characterized by symptoms from vascular invasion and thrombosis, which lead to impaired blood flow and poor penetration of systemic antifungal treatment. The disease affects mainly immunocompromised individuals, including those with uncontrolled diabetes mellitus, hematologic malignancies, and high levels of iron and those on treatment with glucocorticoids. Mucormycosis can likewise appear as a cutaneous and subcutaneous illness in some trauma patients without known immunodeficiency.

Rhizopus is the genus most commonly involved in human mucormycosis, followed in frequency by *Mucor* and *Lichtheimia*, together representing 70%–80% of all cases of mucormycosis, whereas *Apophysomyces* spp. account for less than 3% of the infections (Gomes et al., 2011). Isolated from the environment of a

mango orchard in northern India, the genus Apophysomyces was first described in 1979 (Misra and Lata, 1979). Until 2010, Apophysomyces elegans was regarded as the only member of this genus; however, the phylogenetic studies over the following years have revealed that Apophysomyces contains at least five species: A. elegans, A. variabilis, A. trapeziformis, A. mexicanus and A. ossiformis. Differences in apophysis and sporangiospores have been observed among the species (Alvarez et al., 2010). The variety of infections with Apophysomyces spp. is underappreciated, since these fungi do not usually grow on the standard fungal culture media utilized in clinical laboratories. These fungi require a special nutrient-deficient growth medium (e.g., Czapek agar), a high temperature in comparison to other human pathogens (37°C-42°C), and prolonged incubation (7-10 days) (Padhye and Ajello, 1988). In spite of their low virulence, Apophysomyces spp. are presently viewed as emerging pathogens amongst mucoralean fungi. Unlike other members of Mucorales that affect immunocompromised hosts and those with uncontrolled diabetes, A. variabilis mainly affects immunocompetent persons in the absence of underlying comorbidities (Ribes et al., 2000).

Recent studies have shown that CAM may occur especially in immunocompromised hosts (Pal et al., 2021). A number of risk factors for CAM have been recognized, and some of these were present in the current case. Our patient had a significant increase in CRP (108 mg/L), and elevated CRP levels and extreme neutropenia are essentially associated with worse survival (Cho et al., 2015). Gode et al. claimed that a CRP level >4 mg/dl would be related to poor prognosis in patients with acute invasive fungal rhinosinusitis (Gode et al., 2015). Similarly, Cho et al. likewise found that patients with a CRP level >5.5 mg/dl had worse outcomes (Cho et al., 2015). Twu et al. found that a CRP \geq 1.025 mg/dl considerably accelerated the risk of developing orbital complications in patients with invasive fungal

rhinosinusitis (Twu et al., 2021). In addition, Mucorales require iron to grow, and enhanced iron availability, including high ferritin levels, is a risk factor for CAM (Jose et al., 2021). Our patient had high ferritin levels during the period of infection, consequently precipitating the rapid growth of the fungus. Hyperglycemia in patients with COVID-19 because of previous diabetes mellitus, damage to beta cells in the pancreas by COVID-19, corticosteroid therapy, and stress-related increased cortisol levels with glucocorticoid therapy can damage phagocytic processes, failing to halt spore germination and growth and leading to CAM. Further endothelial damage and increased expression of endothelial receptors have been observed in patients with COVID-19. Therefore, in our patient, glucocorticoid treatment, hyperglycemia and its complications, and dysfunction of innate immune cells due to high-dose corticosteroid therapy and tocilizumab are probable contributors to the pathogenesis of CAM (Prakash et al., 2021).

A. variabilis has most often been isolated in cases of cutaneous mucormycosis (Chander et al., 2018). Nevertheless, a couple of cases of rhino-orbito-cerebral disease due to unusual Mucorales species have been reported; however, this is a condition that is probably underestimated due to difficulties in precise species identification in many laboratories. We reviewed 13 cases, including our current case, of ROCM published in the period 2000–2021, that were supported by data from molecular species identification. Eight of the 13 patients had been

otherwise healthy, and only five of 13 were diabetic or had other underlying systemic diseases. The average age was 44.5 years (range, 24-74 years). As seen in Table 1, all these patients have been treated with liposomal amphotericin B or amphotericin B. Of the reviewed patients with a rhino-orbitocerebral disease, six of 13 survived, and four died; for the remaining three patients, information was not available. Of the four who died, three were previously healthy, and one had diabetes (Table 1). Wolkow et al. reported a case of A. variabilis infection resulting in rhino-orbito-cerebral disease in an immunocompetent 74-year-old woman. This patient exhibited diffuse calvarial lytic lesions and overlying soft-tissue nodules, but without parenchymal intracranial involvement. There was radiographic and clinical evidence of infarction of the orbital contents and cavernous sinus thrombosis. That patient was treated with amphotericin B, isavuconazole, and terbinafine (Wolkow et al., 2017). Moreover, another case of A. variabilis infection has been reported, but no patient information is available (Bala et al., 2015).

Conclusions

This case alerts us to be more aware of opportunistic fungal infections caused by rare species of Mucorales in hospitals. Although it may be a difficult task, it is essential to be

TABLE 1 Clinical characteristics, treatment, and outcomes of patients with rhino-orbito-cerebral mucormycosis caused by rare or unusual species of Mucorales.

Case no.	Age/ sex	Geography	Underlying condition	Causative agent of	Antifungal	Outcome and comments	References
1	59/M	Australia	Immunocompetent	Apophysomyces elegans	L-AmB	Cured	(Fairley et al., 2000)
2	24/M	Columbia	Immunocompetent	A. elegans	L-AmB	Cured	(Garcia-Covarrubias et al., 2001)
3	50/M	NA	Diabetes mellitus, allergic rhinitis, and previous sinusitis	A. elegans	L-AmB	Cured	(Liang et al., 2006)
4	31/M	India	Immunocompetent	A. elegans	AmB	Died	(Schütz et al., 2006)
5	56/F	India	Immunocompetent	Saksenaea vasiformis	AmB	Died	(Baradkar et al., 2008)
6	40/M	Malaysia	Immunocompetent	S. vasiformis	AmB	Cured	(Shatriah et al., 2012)
7	25/M	India	Immunocompetent	A. elegans	AmB	Cured	(Parsi et al., 2013)
8	NA/M	India	Diabetic	Syncephalastrum racemosum	NA	NA	(Mathuram et al., 2013)
9	45/F	India	Diabetic	A. elegans	L-AmB	NA	(Biswas et al., 2015)
10	NA	India	NA	Apophysomyces variabilis	NA	NA	(Bala et al., 2015)
11	74/F	USA	Immunocompetent	A. variabilis	L-AmB, ISC, TRF	Cured	(Wolkow et al., 2017)
12	46/M	Mexico	Diabetes	A. ossiformis	L-AmB	Died	(Martínez-Herrera et al., 2020)
13	40/F	Iran	Immunocompetent	A. variabilis	L-AmB	Died	This study

M, male; F, female; NA, not available; L-AmB, liposomal amphotericin B; AmB, amphotericin B; ISC, isavuconazole; TRF, terbinafine.

conscious of the correct detection and identification of microorganisms to improve the prophylaxis and therapeutic decision and outcome and to reduce mortality.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

Ethical approval of the study was obtained from the ethics committee of Tehran University of Medical Sciences, Tehran, Iran (IR.TUMS.SPH.REC.1399.329). Written informed consent was obtained from the next of kin of the patient for the publication of any potentially identifiable images or data.

Author contributions

ME and SA performed all the experiments and participated in data collection. SA drafted the manuscript and assisted in data analysis and interpretation. ME, MH, JH, AA, and SS participated in collecting the clinical isolate and in data collection. HM was in charge of supervising the study and critical review of the manuscript.

References

Aboutalebian, S., Ahmadikia, K., Fakhim, H., Chabavizadeh, J., Okhovat, A., Nikaeen, M., et al. (2021). Direct detection and identification of the most common bacteria and fungi causing otitis externa by a stepwise multiplex PCR. *Front. Cell. infect. Microbiol.* 11, 210. doi: 10.3389/fcimb.2021.644060

Aboutalebian, S., Mahmoudi, S., Okhovat, A., Khodavaisy, S., and Mirhendi, H. (2020). Otomycosis due to the rare fungi talaromyces purpurogenus, naganishia albida and filobasidium magnum. *Mycopathologia* 185, 569–575. doi: 10.1007/s11046-020-00439-8

Alvarez, E., Stchigel, A. M., Cano, J., Sutton, D. A., Fothergill, A. W., Chander, J., et al. (2010). Molecular phylogenetic diversity of the emerging mucoralean fungus apophysomyces: proposal of three new species. *Rev. Iberoamericana Micol.* 27, 80– 89. doi: 10.1016/j.riam.2010.01.006

Bala, K., Chander, J., Handa, U., Punia, R. S., and Attri, A. K. (2015). A prospective study of mucormycosis in north India: experience from a tertiary care hospital. *Med. mycol.* 53, 248–257. doi: 10.1093/mmy/myu086

Baradkar, V., Mathur, M., Taklikar, S., Rathi, M., and Kumar, S. (2008). Fatal rhino-orbito-cerebral infection caused by saksenaea vasiformis in an immunocompetent individual: first case report from India. *Indian J. Med. Microbiol.* 26, 385–387. doi: 10.1016/S0255-0857(21)01823-5

Binder, U., Maurer, E., and Lass-Flörl, C. (2014). Mucormycosis-from the pathogens to the disease. *Clin. Microbiol. Infect* 20, 60–66. doi: 10.1111/1469-0691.12566

Biswas, D., Kotwal, A., Kakati, B., and Ahmad, S. (2015). Amphotericin b resistant apophysomyces elegans causing rhino-oculo-cerebral mucormycosis in an immunocompetent host. *J. Clin. Diagn. Res.* 9, DD01. doi: 10.7860/JCDR/2015/ 13929.6272

Bouza, E., Munoz, P., and Guinea, J. (2006). Mucormycosis: an emerging disease? Clin. Microbiol. Infect 12, 7–23. doi: 10.1111/j.1469-0691.2006.01604

All authors contributed to the article and approved the submitted version.

Funding

This work was supported by Isfahan University of Medical Sciences, Isfahan, Iran (grant number 1400180), which we gratefully acknowledge. Also, the authors thank the staff at Shahid Beheshti Hospital, Kashan, Iran.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Chakrabarti, A., Shivaprakash, M., Curfs-Breuker, I., Baghela, A., Klaassen, C., and Meis, J. (2010). Apophysomyces elegans: epidemiology, amplified fragment length polymorphism typing, and *in vitro* antifungal susceptibility pattern. *J. Clin. Microbiol.* 48, 4580–4585. doi: 10.1128/JCM.01420-10

Chander, J., Kaur, M., Singla, N., Punia, R., Singhal, S. K., Attri, A. K., et al. (2018). Mucormycosis: battle with the deadly enemy over a five-year period in India. *J. Fungi* 4, 46. doi: 10.3390/jof4020046

Cho, H.-J., Jang, M.-S., Hong, S. D., Chung, S.-K., Kim, H. Y., and Dhong, H.-J. (2015). Prognostic factors for survival in patients with acute invasive fungal rhinosinusitis. *Am. J. rhinol. Allergy* 29, 48–53. doi: 10.2500/ajra.2015.29.4115

Dannaoui, E., and Lackner, M. (2020). Mucorales and mucormycosis. J. Fungi 6 (1), 6. doi: 10.3390/jof6010006

Divakar, P. K. (2021). Fungal taxa responsible for Mucormycosis/"Black fungus" among COVID-19 patients in India. J. Fungi 7, 641. doi: 10.3390/jof7080641

Fairley, C., Sullivan, T. J., Bartley, P., Allworth, T., and Lewandowski, R. (2000). Survival after rhino-orbital-cerebral mucormycosis in an immunocompetent patient. *Ophthalmology* 107, 555–558. doi: 10.1016/S0161-6420(99)00142-6

Fakhim, H., Nasri, E., Aboutalebian, S., Gholipour, S., Nikaeen, M., Vaezi, A., et al. (2021). Asymptomatic carriers of coronavirus disease 2019 among healthcare workers in Isfahan, Iran. *Future Virol.* 16(2), 93–8. doi: 10.2217/fvl-2020-0224

Farmakiotis, D., and Kontoyiannis, D. P. (2016). Mucormycoses. Infect. Dis. Clinics 30, 143-163. doi: 10.1016/j.idc.2015.10.011

Garcia-Covarrubias, L., Bartlett, R., Barratt, D. M., and Wassermann, R. J. (2001). Rhino-orbitocerebral mucormycosis attributable to apophysomyces elegans in an immunocompetent individual: case report and review of the literature. *J. Trauma Acute Care Surg.* 50, 353–357. doi: 10.1097/00005373-200102000-00027

Gode, S., Turhal, G., Ozturk, K., Aysel, A., Midilli, R., and Karci, B. (2015). Acute invasive fungal rhinosinusitis: survival analysis and the prognostic indicators. *Am. J. rhinol. Allergy* 29, e164–e169. doi: 10.2500/ajra.2015.29.4245

Gomes, M. Z., Lewis, R. E., and Kontoyiannis, D. P. (2011). Mucormycosis caused by unusual mucormycetes, non-Rhizopus,-Mucor, and-lichtheimia species. *Clin. Microbiol. Rev.* 24, 411–445. doi: 10.1128/CMR.00056-10

Ibrahim, A. S., Spellberg, B., Walsh, T. J., and Kontoyiannis, D. P. (2012). Pathogenesis of mucormycosis. *Clin. Infect. Dis.* 54, S16–S22. doi: 10.1093/cid/ cir865

Jose, A., Singh, S., Roychoudhury, A., Kholakiya, Y., Arya, S., and Roychoudhury, S. (2021). Current understanding in the pathophysiology of SARS-CoV-2-Associated rhino-Orbito-Cerebral mucormycosis: A comprehensive review. J. Maxillofac. Oral. Surg. 20(3), 373–380. doi: 10.1007/s12663-021-01604-2

Kontoyiannis, D. P., Lewis, R. E., Lotholary, O., Spellberg, B., Petrikkos, G., Roillides, E., et al. (2012). Future directions in mucormycosis research. *Clin. Infect. Dis.* 54, S79–S85. doi: 10.1093/cid/cir886

Kwon-Chung, K. J. (2012). Taxonomy of fungi causing mucormycosis and entomophthoramycosis (zygomycosis) and nomenclature of the disease: molecular mycologic perspectives. *Clin. Infect. Dis.* 54, S8–S15. doi: 10.1093/cid/cir864

Liang, K. P., Tleyjeh, I. M., Wilson, W. R., Roberts, G. D., and Temesgen, Z. (2006). Rhino-orbitocerebral mucormycosis caused by apophysomyces elegans. J. Clin. Microbiol. 44, 892–898. doi: 10.1128/JCM.44.3.892-898.2006

Martínez-Herrera, E., Frías-De-León, M. G., Julián-Castrejón, A., Cruz-Benítez, L., Xicohtencatl-Cortes, J., and Hernández-Castro, R. (2020). Rhino-orbital mucormycosis due to apophysomyces ossiformis in a patient with diabetes mellitus: a case report. *BMC Infect. Dis.* 20, 1–4. doi: 10.1186/s12879-020-05337-4

Mathuram, A. J., Mohanraj, P., and Mathews, M. S. (2013). Rhino-orbital-cerebral infection by syncephalastrum racemosusm. J. Assoc. Phys. India 61, 339–340.

Misra, P., and Lata, K. (1979). Apophysomyces, a new genus of the mucorales. (USA: MYCOTAXON). 1979; 8(2): PP. 317–82.

Padhye, A. A., and Ajello, L. (1988). Simple method of inducing sporulation by apophysomyces elegans and saksenaea vasiformis. *J. Clin. Microbiol.* 26, 1861–1863. doi: 10.1128/jcm.26.9.1861-1863.1988

Pal, R., Singh, B., Bhadada, S. K., Banerjee, M., Bhogal, R. S., Hage, N., et al. (2021). COVID-19-associated mucormycosis: An updated systematic review of literature. *Mycoses* 64 (12), 1452–1459 doi: 10.1111/myc.13338

Parsi, K., Itgampalli, R. K., Vittal, R., and Kumar, A. (2013). Perineural spread of rhino-orbitocerebral mucormycosis caused by apophysomyces elegans. *Ann. Indian Acad. Neurol.* 16, 414. doi: 10.4103/0972-2327.116921

Patel, A., Kaur, H., Xess, I., Michael, J., Savio, J., Rudramurthy, S., et al. (2020). A multicentre observational study on the epidemiology, risk factors, management and outcomes of mucormycosis in India. *Clin. Microbiol. Infect* 26, 944. e9–944. e15. doi: 10.1016/j.cmi.2019.11.021

Prakash, H., Skiada, A., Paul, R. A., Chakrabarti, A., and Rudramurthy, S. M. (2021). Connecting the dots: interplay of pathogenic mechanisms between COVID-19 disease and mucormycosis. *J. Fungi* 7, 616. doi: 10.3390/jof7080616

Ribes, J. A., Vanover-Sams, C. L., and Baker, D. J. (2000). Zygomycetes in human disease. Clin. Microbiol. Rev. 13, 236–301. doi: 10.1128/CMR.13.2.236

Roden, M. M., Zaoutis, T. E., Buchanan, W. L., Knudsen, T. A., Sarkisova, T. A., Schaufele, R. L., et al. (2005). Epidemiology and outcome of zygomycosis: a review of 929 reported cases. *Clin. Infect. Dis.* 41, 634–653. doi: 10.1086/432579

Schütz, P., Behbehani, J. H., Khan, Z. U., Ahmad, S., Kazem, M. A., Dhar, R., et al. (2006). Fatal rhino-orbito-cerebral zygomycosis caused by apophysomyces elegans in a healthy patient. *J. Oral. Maxillofac. Surg.* 64, 1795–1802. doi: 10.1016/j.joms.2006.05.010

Shatriah, I., Mohd-Amin, N., Tuan-Jaafar, T. N., Khanna, R. K., Yunus, R., and Madhavan, M. (2012). Rhino-orbito-cerebral mucormycosis in an immunocompetent patient: case report and review of literature. *Middle East Afr. J. Ophthalmol.* 19, 258. doi: 10.4103/0974-9233.95269

Singh, A., Singh, R., Joshi, S., and Misra, A. (2021). Mucormycosis in COVID-19: A systematic review of cases reported worldwide and in India, diabetes & metabolic syndrome. clinical research & reviews. J. Pre-proof. (2021)15 (4),102146 doi: 10.1016/j.dsx.2021.05.019

Skiada, A., Pagano, L., Groll, A., Zimmerli, S., Dupont, B., Lagrou, K., et al. (2011). Zygomycosis in Europe: analysis of 230 cases accrued by the registry of the European confederation of medical mycology (ECMM) working group on zygomycosis between 2005 and 2007. *Clin. Microbiol. Infect* 17, 1859–1867. doi: 10.1111/j.1469-0691.2010.03456.x

Twu, K.-H., Kuo, Y.-J., Ho, C.-Y., Kuan, E. C., Wang, W.-H., and Lan, M.-Y. (2021). Invasive fungal rhinosinusitis with and without orbital complications: Clinical and laboratory differences. *J. Fungi* 7, 573. doi: 10.3390/jof7070573

Wayne, P. (2008). Clinical and laboratory standards institute (CLSI): Reference method for broth dilution antifungal susceptibility testing of filamentous fungi (Carol Stream, IL: Allured Publishing Corporation).

Wolkow, N., Jakobiec, F. A., Stagner, A. M., Cunnane, M. E., Piantadosi, A. L., Basgoz, N., et al. (2017). Chronic orbital and calvarial fungal infection with apophysomyces variabilis in an immunocompetent patient. *Survey Ophthalmol.* 62, 70–82. doi: 10.1016/j.survophthal.2016.05.006