

Impact of Neutrophil Extracellular **Traps on Thrombosis Formation: New Findings and Future Perspective**

Yilu Zhou, Zhendong Xu* and Zhigiang Liu*

Department of Anesthesiology, Shanghai First Maternity and Infant Hospital, School of Medicine, Tongii University, Shanghai, China

Thrombotic diseases seriously endanger human health, neutrophils and neutrophil extracellular traps (NETs) play an important role in abnormal thrombus formation. NETs are extracellular structures released by neutrophils upon stimulation by pathogens. NETs include neutrophil elastase (NE), myeloperoxidase (MPO), cathepsin G and other active substances. The network structure provided by NETs can prevent the spread of pathogens and effectively kill and eliminate pathogens. However, the components of NETs can also abnormally activate the coagulation pathway and participate in the formation of pathological thrombi. This review aims to summarize the mechanisms of NETs formation in detail; the research progress of NETs in venous thrombosis, arterial thrombosis, acquired disease-associated thrombosis, sepsis coagulation disorder; as well as the strategies to target NETs in thrombosis prevention and treatment.

Keywords: neutrophil, neutrophil extracellular traps, thrombosis, Innate immunity, thrombosis - immunology

INTRODUCTION

Neutrophil extracellular traps (NETs) are extracellular structures released by neutrophils under stimuli, such as pathogens, histones and bacteriostatic proteins (Laridan et al., 2019). NETs can confine pathogens in situ, prevent the systemic spread of pathogens and assist phagocytes in killing pathogens, which is considered to be another mechanism of action of neutrophils in innate immunity (Papayannopoulos, 2018). Because the network structure of NETs can provide a scaffold for the aggregation of red blood cells and platelets, and its components can also activate the coagulation pathway, NETs have recently been discovered to be involved in thrombosis formation (Noubouossie et al., 2019). This review elaborates the formation mechanism of NETs and their role in thrombosis formation and focuses on research progress in different kinds of thrombosis formation as well as related thrombosis prevention strategies.

NET FORMATION

NETs were first named by Brinkmann et al. in 2004, but the mechanism by which NETs undergo NETosis has not yet been clarified (Brinkmann et al., 2004). According to differences in the pathogenic stimulation of neutrophils, activation of signaling pathways, and cell membrane integrity, NETs formation can be classified into two types: suicidal NETosis and survival NETs

OPEN ACCESS

Edited by:

Meihong Deng, The Ohio State University, United States

Reviewed by:

Zhichao Fan. UCONN Health. United States Olga Ciepiela, Medical University of Warsaw, Poland

> *Correspondence: Zhigiang Liu drliuzhigiang@163.com

Zhendona Xu btxzd123@126.com

Specialty section:

This article was submitted to Microbes and Innate Immunity. a section of the journal Frontiers in Cellular and Infection Microbiology

> Received: 01 April 2022 Accepted: 04 May 2022 Published: 31 May 2022

Citation:

Zhou Y, Xu Z and Liu Z (2022) Impact of Neutrophil Extracellular Traps on Thrombosis Formation: New Findings and Future Perspective. Front, Cell, Infect, Microbiol, 12:910908. doi: 10.3389/fcimb.2022.910908 release (Thiam et al., 2020). In suicidal NETosis, neutrophils activate Fc receptors, Toll-like receptors (TLRs) and complement receptors under the stimulation of interleukin-8 and bacterial surface antigens. Activation of reduced coenzyme II oxidase leads to an increase in reactive oxygen species (ROS), and protein-arginine deiminase 4 (PAD4) further citrullinates chromatin after activation under the action of ROS, histone 3 and histone 4, causing DNA unwinding (Kapoor et al., 2018; Van Avondt and Hartl, 2018). ROS can also promote the release of myeloperoxidase (MPO) and neutrophil elastase (NE) in granules, and the NE can enter the nucleus to assist in the cleavage of histones and promote DNA unwinding (Jorch and Kubes, 2017). After the nucleus is ruptured, unwound DNA strands and histones enter the cytoplasm, and antibacterial proteins in the cytoplasm, such as MPO, CitH3, NE and cathepsin G, work together to form early NETs. At this time, neutrophils initiate the cell death program, and lysis of the cell membrane releases NETs (Kenny et al., 2017). In survival NETs release, neutrophils activate TLR-4 and TLR-2 receptors under stimuli such as bacterial lipopolysaccharide and gram-negative bacteria, resulting in PAD4 activation, and activated PAD4 enters the nucleus to citrullinate histone H3 and H4, causing DNA unwinding (Branzk et al., 2014; Thålin et al., 2019). Unlike suicidal NETosis, PAD4 is activated in a non-ROS-activated manner in surviving NETosis and is not accompanied by nuclear

or cell membrane ruptures. Neutrophils themselves do not die, and unwound DNA strands bud into the cytoplasm and form early NETs with bacteriostatic proteins. They are exocytosed and released to the outside of the cell in the form of vesicles. At this time, although neutrophils have no nuclear DNA, they still have the ability to phagocytose and kill bacteria (Adrover et al., 2020). In the regulation of NETosis, pathogen fragment size and particle content in neutrophils can influence the generation and release of NETs (Branzk et al., 2014; Adrover et al., 2020). In addition, specific subpopulations of macrophages can also participate in the degradation of NETs through phagocytosis (Haider et al., 2020). Recent studies have also found that the pore-forming protein gasdermin D (GSDMD) can promote the formation and release of NETs by participating in the lysis of the neutrophil granule membrane, nuclear membrane and cell membrane and regulating the permeability of neutrophils. The intranuclear transfer of NE further affects NETosis (Sollberger et al., 2018). The link between NETosis and various kinases, such as cyclindependent kinases is also under investigation (Amulic et al., 2017; Wolach et al., 2018). However, there is still considerable controversy about whether PAD4 activation is necessary in NETosis and whether the DNA in NETs can be of mitochondrial origin (Lood et al., 2016; Claushuis et al., 2018). In conclusion, the mechanism of NETs generation and release still needs to be further explored (Figures 1A-C).



NETS AND THROMBOSIS FORMATION

Thrombosis is an intravascular blood clot formed by substances such as platelets and coagulation factors in the case of vascular injury, abnormal blood flow or blood components and can play a role in repairing vascular injury and hemostasis under normal physiological conditions (Mackman et al., 2020). When the coagulation pathway is overactivated or fibrinolytic activity is decreased, the formation of thrombi will lead to vascular obstruction, resulting in a tissue blood supply disorder (Mackman et al., 2020). In addition, in situ thrombus shedding can also cause acute thromboembolism, such as acute myocardial infarction (AMI) (Wendelboe and Raskob, 2016), stroke (Farkas et al., 2019) with high fatality and disability rates. Several recent studies have found that neutrophils and NETs are ubiquitous in human arteriovenous thrombosis samples, mouse deep vein thrombosis (DVT) and other disease models (Wendelboe and Raskob, 2016; Noubouossie et al., 2017; Wang et al., 2018; Farkas et al., 2019). NETs have been further found to attract platelet activation and promote thrombosis by activating internal and external coagulation pathways, Generally, After vessel injury induced by neutrophils and NETs, TF under the endothelium is exposed to the vascular lumen and initiates extrinsic coagulation pathway. It serves as a cellular receptor for plasma factor VII/VIIa. Together, the factor VIIa : TF complex activates factor X and factor IX. Activated factor Xa in association with its cofactor Va, the prothrombinase complex, cleaves prothrombin to thrombin, which in turn cleaves fibrinogen to fibrin. Then, trans-glutaminase factor XIII cross-links fibrin to stabilize the thrombi (Noubouossie et al., 2017; Wang et al., 2018; Farkas et al., 2019). The network structure of NETs provides a scaffold for the deposition of platelets, erythrocytes, fibrinogen, von Willebrand factor (vWF) and other substances, such as platelet adhesion factors and extracellular bodies, which are conducive to thrombosis (Wang et al., 2018). Histones in NETs can attract platelet aggregation and activation through the interaction of fibrinogen, TLR2 and TLR4 and promote an increase in thrombin generation. In addition to the inhibition of activated protein C (APC), histones can also promote the expression of tissue factor on vascular endothelium and macrophages and promote coagulation (Yang et al., 2016; Noubouossie et al., 2017; Osada et al., 2017). NE and cathepsin G can promote the coagulation pathway induced by tissue factor and coagulation factor XII by hydrolyzing tissue factor pathway inhibitor (TFPI). Coagulation factor XII can also bind to surrounding platelets and DNA through its connection with NETs; it is directly activated by the coagulation pathway to expand coagulation (Vu et al., 2016; Döring et al., 2020). While cathepsin G promotes endothelial activation and prothrombin production, DNA inhibits fibrinolysis by forming complexes with fibrin and plasmin (Gould et al., 2015; Folco et al., 2018). In terms of the overall effect of NETs on thrombosis, NETs have been shown to promote the formation of erythrocyte-rich thrombi in vitro, and the erythrocytes in the thrombus are directly bound to NETs as observed by electron microscopy. In addition, NETs can interact with fibronectin and vWF to attract and promote platelet adhesion and activation (Gould et al., 2014; Martinod and

Wagner, 2014). The connection between NETs and fibrinogen in thrombi can also promote fibrin deposition and improve thrombus stability; thus, thrombi containing NETs are more sensitive to tissue plasminogen activator (tPA) than thrombi with fibrin as the main component (Ducroux et al., 2018). The use of DNase I in mouse models can effectively prevent the formation of intravascular microthrombosis, which strongly suggests an effect of NETs on thrombosis (Jiménez-Alcázar et al., 2017). However, Noubouossie et al. (2017) pointed out that the promoting effect of NETs on thrombosis is achieved through its DNA and histone components rather than the NETs themselves. Therefore, evidence for a thrombopromoting effect of NETs as a whole remains to be further developed. Studies have shown that activated platelets can promote the formation and release of NETs through mechanisms such as P-selectin and high mobility group box protein 1 (Etulain et al., 2015; Kim and Lee, 2020). Therefore, the effects between NETosis and thrombosis may be reciprocal, and this mutually reinforcing cascade may play an important role in events such as thrombophilia in patients with thrombosis or an increased risk of thrombosis after infection (Figure 2).

NETS AND VENOUS THROMBOSIS

The relationship between NETs and venous thrombosis was first discovered in animal disease models (Wang et al., 2018). In the DVT disease model, staining of the thrombosed vein tissue indicated the presence of extracellular DNA; the DNA costained with vWF, and the serum DNA level was also increased, while the nonthrombotic vein tissue did not stain positive for extracellular DNA (Wang et al., 2018). A similar phenomenon was observed in a mouse DVT model, wherein activated neutrophils and NETs-specific markers, such as citrullinated histone H3 (CitH3) and extracellular DNA, were all present within the thrombus (Lim et al., 2018). Thus, the existence of NETs in venous thrombosis was preliminarily proven. In PAD4 knockout mice, NETosis is inhibited due to the inability of histones to be citrullinated under the action of PAD4 (Martinod et al., 2013). In addition, when using antibodies to inhibit neutrophil activation, DVT formation and thrombus diameter were significantly reduced in mice, while DNasetreated DVT mice had reduced thrombus recurrence and thrombus size (Mutua and Gershwin, 2021). Thus, the role of NETs in venous thrombosis has been clearly demonstrated. The relationship between NETs and venous thrombosis has also been demonstrated in human disease. First, the serum levels of DNA and activated neutrophil markers in DVT patients were higher than those in non-DVT patients, preliminarily verifying the association of NETs with venous thrombosis (Conteduca et al., 2022). The presence of extracellular DNA-histone complexes and neutrophils was further detected in thrombus samples obtained from DVT and PE patients by thrombectomy, and the different stages of thrombus formation were also detected by staining of thrombus cross-sections. The contents of neutrophils and CitH3 are different in the early stage of thrombosis; they are replaced by collagen fibers in the later stage, and the content of



NETs markers is greatly reduced (Savchenko et al., 2014). In vitro experiments showed that NETs promote the differentiation of fibroblasts into collagen-secreting myofibroblasts, suggesting that NETs may be mainly involved in early venous thrombosis. With the development of thrombi, NETs can stimulate the production of collagen fibers from fibroblasts to promote maturation of the thrombus structure (Chrysanthopoulou et al., 2014). NETs have also been used as serum markers in the differential diagnosis of DVT, and different grades of extracellular DNA and CitH3 were detected in the serum of hundreds of DVT and non-DVT patients (van Montfoort et al., 2013). However, due to the limited sample size and differences in experimental techniques, the experimental results of NETs and venous thrombosis have been inconsistent across studies (van Montfoort et al., 2013; Savchenko et al., 2014; Chrysanthopoulou et al., 2014). Currently, there remains a lack of experimental evidence with a large sample size, which affects the role of NETs in venous thrombosis. The application of NETs, how to improve the stability of the results and further exploration of their role in thrombus structure need to be explored.

NETS AND ARTERIAL THROMBOSIS

Unlike venous thrombosis, arterial thrombosis is more common in acute events resulting from thrombus shedding, such as AMI and ischemic stroke (Wendelboe and Raskob, 2016; Farkas et al., 2019). In this process, in addition to NETs *in situ* thrombi, the inflammatory response to tissue damage caused by ischemia can also recruit neutrophils (Farkas et al., 2019). Numerous neutrophils were found in tissue samples from acute myocardial infarction long before the discovery of NETs, and activated neutrophils were also found in thrombus samples obtained from patients with acute myocardial infarction (Bonaventura et al., 2020). Using NETs-specific markers and neutrophil antibodies, the widespread presence of NETs was further detected in thrombus samples and atherosclerotic plaques from patients with acute myocardial infarction, preliminarily demonstrating the existence of NETs in arterial thrombi (Riegger et al., 2016; Franck et al., 2018). Maugeri et al. (2014) also found the presence of NETs and platelet aggregation in coronary thrombi by immunostaining, verifying the attraction and aggregation of NETs to platelets. In addition, the content of NETs in recent coronary thrombi was higher than that in old coronary thrombi, and the content of NETs in coronary thrombi was positively correlated with the extent of myocardial infarction and the degree of ST-segment elevation (Mangold et al., 2015; Langseth et al., 2018). It was also positively correlated with the severity of prognostic outcomes in patients within two years, providing preliminary evidence for NETs as a disease prognostic indicator in myocardial infarction (Mangold et al., 2015; Langseth et al., 2018). In ischemic stroke, Laridan et al. (2017) detected the presence of NETs in thrombus samples. In a study of stroke thrombosis, NETs were found to be more abundant in cardiogenic and structurally mature old thrombi, and the content was positively correlated with the duration of thrombectomy and the number of surgical instruments used, which prompted the local microenvironment to affect NETosis (Boeckh-Behrens et al., 2016; De Meyer et al., 2017; Ducroux et al., 2018). In other arterial thrombi, such as intra-abdominal aortic aneurysm thrombi, DNA-histone complex-based assays have also suggested the presence of NETs (Fernández-Ruiz, 2018). It is worth noting that the content of NETs detected in thrombus samples from patients with acute myocardial infarction and ischemic stroke showed opposite results in thrombi of different maturities; by comparing arterial and venous thrombus samples, arterial thrombi with higher levels of DNA-histone complexes in NETs were found to have enhanced inflammatory responses and inflammatory cell recruitment within atheromatous plaques by promoting macrophage activation (Mangold et al., 2015; Warnatsch et al., 2015). NETs also affect the stability of atherosclerotic plaques by acting on smooth muscle cells through histones, resulting in reduced intraplaque smooth muscle cell content, reduced plaque interstitial collagen synthesis, and thinning of the fibrous cap, whereas PAD4 inhibition or anti-Ly6G can enhance the stability of plaques and the proportion of smooth muscle cells in plaques (Döring et al., 2020). Therefore, whether NETs may have different mechanisms of action in different etiologies and different origins of thrombi, whether the tissue microenvironment and vascular differences can modify NETs in thrombus formation, and whether NETs play a role in the early stage and development of arterial thrombosis need to be further explored.

NETS AND ANTI-COAGULANT LIPID SYNDROME (APS) AND TUMOR-ASSOCIATED THROMBOSIS

Acquired thrombophilia refers to a group of clinical syndromes in which patients have a high risk of thromboembolism due to the presence of acquired thrombosis risk factors, and the embolism is mostly venous thromboembolism, including anticoagulant lipid syndrome (APS) and tumor-associated thrombosis (Stevens et al., 2016). APS is an autoimmune disease characterized by recurrent venous or arterial thrombosis and miscarriage and is accompanied by increased serum antiphospholipid antibody (aPL) titers (Wirestam et al., 2019). Studies have shown that the content of extracellular DNA in the serum of patients with primary APS is increased, and the ability of isolated neutrophils to release NETs is enhanced, preliminarily verifying the existence of NETs in APS (Yalavarthi et al., 2015). In addition, the serum antibody of APS patients is positively correlated with the NETs content, and the isolated aPL mAb can enhance the release of NETs from neutrophils, suggesting that aPL-mediated NETs release may be a mechanism of APS-related thrombosis (van der Linden et al., 2018). In animal experiments, by adding aPL monoclonal antibody isolated from the serum of APS patients to DVT mice, the risk and degree of thrombosis were increased, the content of CitH3 in the thrombus was higher than that of the control group, and the level of serum DNA was also increased, validating the mechanism by which aPL promotes thrombosis by stimulating NETs release (Meng et al., 2017). The analysis of serum antibodies in APS patients further revealed the presence of anti-NETs antibodies, which led to a decrease in the degradation and clearance capacity of NETs formed in blood vessels, revealing another possible mechanism related to NETs in APS (Zuo et al., 2020).

The risk of thrombosis in cancer patients is higher than that in healthy people, which seriously affects the prognosis of patients (Fernandes et al., 2019). In addition to tumor-derived cytokines and chemotherapy-mediated tissue damage, the causes of thrombosis have also been found to be related to neutrophil activation and the formation and release of NETs (Hisada et al., 2015). In tumor patients, the contents of CitH3 and histone-DNA complexes in the systemic circulation and thrombus are increased, and they can be used to predict the risk of arterial thromboembolism in patients with malignant tumors of the lung and pancreas, suggesting that NETs play a role in tumor-related thrombosis (Oklu et al., 2017; Mauracher et al., 2018; Grilz et al., 2019). In vitro experiments have shown that human-derived pancreatic tumor cells can stimulate normal neutrophils to increase the release of NETs, and the NETs release capacity of neutrophils from gastric cancer patients is also enhanced compared to that of neutrophils from non-tumor patients (Yang et al., 2015; Abdol Razak et al., 2017). In animal experiments, the formation and release of NETs and the incidence of thrombi in tumor mice were higher than those in non-tumor mice, and CitH3 could be directly detected in the thrombus, further illustrating the relationship between NETs and tumor thrombosis (Hisada et al., 2020). Leal et al. (2017) observed increased formation and release of NETs when using tumor-derived granulocyte colony-stimulating factor (G-CSF) to stimulate neutrophils, first suggesting that NETs are involved in tumor thrombus formation. However, there is currently a lack of evidence on thrombosis after NETosis inhibition in acquired thrombophilia, and the evidence related to the impact of NETs on tumor prognosis needs to be further developed.

NETS AND SEPSIS-ASSOCIATED COAGULATION DISORDER

Sepsis is a systemic inflammatory response syndrome caused by infection (Singer et al., 2016). Patients with sepsis are often characterized by activation of the coagulation system, a progressive prothrombotic state, and dysregulation of the anticoagulation system, which may lead to disseminated intravascular coagulation (DIC), microvascular thrombosis, hypoperfusion and eventually multiple organ dysfunction syndrome and death (Semeraro et al., 2012; Tsao et al., 2015). A large amount of lipopolysaccharide is shed by bacteria in patients with sepsis; this is an important substance for activating neutrophils to produce NETs (Clark et al., 2007), which may be one of the reasons for the formation of NETs in sepsis. Platelets participate in the coagulation process and also play a role in inducing inflammation and resisting infection in infectious diseases (Thomas and Storey, 2015). Activated platelets during sepsis can directly sequester or kill pathogens or promote pathogen clearance by activating macrophages and neutrophils, and the specific receptor TLR4 is activated to promote NETs formation, ultimately causing platelet aggregation and microthrombosis (McDonald et al., 2012). Neutrophils isolated from the blood of patients with sepsis can release tissue factor (TF) through NETs, and this form of TF can induce thrombin generation in vitro and play a key role in the activation of the coagulation system in sepsis (Kambas et al., 2012). Plasma and platelets isolated from the blood of patients with sepsis can induce neutrophils to release NETs in vitro, and NETs produced

by neutrophils from patients with sepsis have good procoagulant activity compared with healthy controls (Yang et al., 2017). NETs play an important role in thrombosis in septic patients through their interaction with platelets. Immune thrombosis refers to the innate immune response induced by thrombosis in blood vessels, especially in microvessels (Engelmann and Massberg, 2013). Immune thrombosis is supported by immune cells and specific thrombosis-related molecules and produces intravascular scaffolds that help identify and destroy pathogens, thereby protecting the body from pathogen-mediated damage (Loof et al., 2011). Neutrophils enhance thrombosis through a cellspecific mechanism of the immune response, and the NETs generated after activation have antibacterial and procoagulant activity (Kimball et al., 2016). NETs promote thrombosis in many ways and are a key factor in immune thrombosis (Goggs et al., 2020). Microvascular thrombotic diseases mainly include hemolytic uremic syndrome and thrombotic thrombocytopenic purpura (TTP), collectively referred to as thrombotic microangiopathy (TMA) (George and Nester, 2014). In hemolytic-uremic syndrome, in addition to promoting the inflammatory response and antibacterial effects, NETs also promote microvascular thrombosis, leading to renal failure in patients (Ramos et al., 2016; Leffler et al., 2017). In transplantation-related studies, elevated NETs levels significantly increased the risk of transplantation-related TMA (Arai et al., 2013), which may be due to the lack of DNase I, which can degrade NETs, in the plasma of these patients (Jiménez-Alcázar et al., 2015). ADAMTS13 is a plasma metalloproteinase that can cleave vWF and is a key enzyme in the diagnosis of TTP (Chauhan et al., 2006). In a recent study using exogenous recombinant human ADAMTS13 and DNase I-treated mice, the production of NETs in vivo was significantly reduced, indicating that ADAMTS13 may inhibit the formation of NETs and that DNase I can degrade NETs structures (Wong et al., 2020).

NETS AND COMPLEMENT SYSTEM IN THROMBOSIS FORMATION

NETs formation is associated with systemic lupus erythematosus activity, and NETs can damage and kill endothelial cells and promote arterial inflammation of atherosclerotic plaques, thereby accelerating atherosclerosis in systemic lupus erythematosus (Barnado et al., 2016). Anti-phospholipid antibodies can stimulate neutrophils to produce NETs, which is closely related to thrombosis in patients with antiphospholipid antibody syndrome (Yalavarthi et al., 2015). The complement system is a part of the innate immune system composed of more than 30 proteins on the plasma and cell surfaces and is one of the main effector mechanisms of antibody-mediated immunity. Neutrophils are activated to release complement components, such as complement factor P, complement factor B, and C3, among which CFP is deposited on NETs and bacteria to induce formation of the membrane attack complex C5b (Yuen et al., 2016). In a mouse experiment with antiphospholipid antibodyinduced thrombus formation, thrombus formation was significantly inhibited in complement component C3 knockout (C3-/-) and C5 knockout (C5-/-) mice, indicating that complement activation plays an important role in antiphospholipid antibody-induced thrombosis (Pierangeli et al., 2005). In another mouse experiment, complement C5 promoted liver injury associated with histone-induced lethal thrombosis (Mizuno et al., 2017). There is also an interaction between complement and platelets, which is manifested in the binding of factor Va to the C5b-induced binding site, promotion of its binding to factor Xa, and then binding to the functional prothrombin binding site on the platelet surface. These studies provide a basis for the study of the role of complement in thrombosis. In a spontaneous small intestinal tumor model, tumor development and hypercoagulability were both associated with neutrophils, especially the appearance of low-density neutrophils. Comparing the effect of complement C3a receptor (C3aR) to that in the absence of C3aR, NETs are more likely to be generated under the effect of C3aR, leading to the activation of coagulation in small intestinal tumors, a hypercoagulable state and thrombosis (Wiedmer et al., 1986). During a variety of bacterial infections, complement plays an important role while promoting the release of NETs; complement CR1 plays a leading role in this process, while complement CR3 plays an assisting role (Guglietta et al., 2016). NETs can serve as direct scaffolds for thrombosis and complement activation, and the opsonization and lytic activity of the complement system enhance the antibacterial properties of NETs. Although NETs, complement proteins, and coagulation factors work differently, they can still function in a large complex to protect the host from hemorrhage and infection; this cooperation is not limited to the site of injury but rather also occurs in the bloodstream. When this balance is disturbed, serious complications may occur, such as sepsis, deep vein thrombosis, autoimmune diseases and even cancer (Palmer et al., 2016). There is a close relationship between NETs and complement, and both play a role in the process of thrombosis. However, the specific division of labor of these components and whether there is a feedback mechanism still need further research.

POTENTIAL THERAPEUTIC TARGETS FOR NETS

The research on NETs in arteriovenous thrombosis and acquired thrombosis has provided a potential target for the treatment of thrombotic diseases. At present, research on NETs associated thrombosis has mainly focused on the promotion of NETs degradation and the inhibition of NETosis. Studies have shown that the use of DNase I can achieve partial lysis of NETs, and the combined use of tPA can achieve complete thrombolysis, which makes up for the defects of incomplete thrombolysis and low efficiency with tPA alone (Mangold et al., 2015). The use of DNase I in mice with thrombosis can also effectively prevent the recurrence of stroke, myocardial infarction and DVT (Ducroux et al., 2018). However, whether the degradation of NETs by DNase I will lead to the dispersion of components such as histones with procoagulant activity and

increase the risk of thrombosis needs further study. Since APC can cleave NETs by cleaving histones, the strategy of using recombinant TMa to promote APC generation has also attracted researchers' attention (Osada et al., 2017). Recent studies have also shown that the NE inhibitor can effectively reduce the generation of NETs in mice with endotoxic shock, providing another potential method for NETs-related antithrombotic therapy (Okeke et al., 2020). In terms of NETosis inhibition, thrombus formation was reduced in PAD4 knockout DVT mice, the PAD4 selective inhibitor GSK484 effectively inhibited NETosis in vitro, and there was no significant decrease in immune function in PAD4-deficient animal models. NETs are powerful targets in thrombosis therapy (Lewis et al., 2015; Martinod et al., 2015). Antioxidants, such as vitamin C, can affect NETosis by reducing the production of ROS, and vitamin C supplementation in septic mice can reduce the production of NETs; however, the specific effect of vitamin C on thrombosis has not been further studied in animal models and clinical trials. The use of JAK signaling pathway inhibitors to observe NETosis inhibition and thrombosis reduction in thrombosis mice needs to be further explored, and the efficacy of the related drug ruxolitinib in thrombosis prevention and treatment has also been confirmed in clinical experiments (Mohammed et al., 2013). The immunosuppressant colchicine has been proven in clinical and basic experiments to improve the prognosis of patients with myocardial infarction and stroke by inhibiting neutrophils, but its specific inhibitory effect on NETosis remains to be explored in depth (Vannucchi et al., 2015; Tardif et al., 2019). In addition, inhibiting the interaction of NETs and coagulation pathways can also achieve thrombosis prevention. Using the rhADAMTS13 protease to specifically cleave vWF, thrombolysis was observed in animal thrombosis models, and the combination of rhADAMTS13 and DNase I effectively improved myocardial infarction by reducing the interaction of vWF with neutrophils and NETs in myocardial tissue of mice (Kelly et al., 2018). The platelet receptor inhibitor P2Y12 and aspirin can also inhibit the interaction between platelets and NETs to reduce thrombosis formation (Li et al., 2017; Thiam et al., 2020).

REFERENCES

- Abdol Razak, N., Elaskalani, O., and Metharom, P. (2017). Pancreatic Cancer-Induced Neutrophil Extracellular Traps: A Potential Contributor to Cancer-Associated Thrombosis. *Int. J. Mol. Sci.* 18 (3), 487. doi: 10.3390/ijms18030487
- Adrover, J. M., Aroca-Crevillén, A., Crainiciuc, G., Ostos, F., Rojas-Vega, Y., Rubio-Ponce, A., et al. (2020). Programmed 'Disarming' of the Neutrophil Proteome Reduces the Magnitude of Inflammation. *Nat. Immunol.* 21 (2), 135– 144. doi: 10.1038/s41590-019-0571-2
- Amulic, B., Knackstedt, S. L., Abu Abed, U., Deigendesch, N., Harbort, C. J., Caffrey, B. E., et al. (2017). Cell-Cycle Proteins Control Production of Neutrophil Extracellular Traps. *Dev. Cell.* 43 (4), 449–462.e5. doi: 10.1016/ j.devcel.2017.10.013
- Arai, Y., Yamashita, K., Mizugishi, K., Watanabe, T., Sakamoto, S., Kitano, T., et al. (2013). Serum Neutrophil Extracellular Trap Levels Predict Thrombotic

CONCLUSION AND FUTURE PERSPECTIVES

As a unique immune mechanism following the death of neutrophils, the role of NETs in thrombotic diseases has been verified in clinical samples and animal experiments, and research on NETs as antithrombotic therapy targets has been widely carried out. This study is expected to provide new insights into the treatment of existing thrombotic diseases and thrombotic complications. NETs-specific markers, such as CitH3 and the MPO-DNA complex, can also be used as biomarkers to provide the basis for the diagnosis or prognosis of thrombosis-related diseases and to explore the development and establishment of quantitative formulas for NETs-specific markers and NETs content in the circulation or local lesions. Methods such as improving the detection sensitivity of NETs can further provide new tools for precision medicine and outcome prediction. However, the results of existing studies have not been consistent, and there is still a lack of stronger experimental evidence for the actual effect of inhibiting NETs on promoting thrombolysis and preventing thrombosis. The issue of immunocompromised and increased bleeding risk also needs to be further explored. We look forward to future research on NETs to provide ideas and methods for solving thrombosisrelated diseases.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

FUNDING

This work was supported by the grants of Science and Technology Commission of Shanghai Municipality (20Y11901400) and Pudong Health Committee of Shanghai (PW2020D-13).

Microangiopathy After Allogeneic Stem Cell Transplantation. Biol. Blood Marrow Transpl. 19 (12), 1683–1689. doi: 10.1016/j.bbmt.2013.09.005

- Barnado, A., Crofford, L. J., and Oates, J. C. (2016). At the Bedside: Neutrophil Extracellular Traps (NETs) as Targets for Biomarkers and Therapies in Autoimmune Diseases. J. Leukoc. Biol. 99 (2), 265–278. doi: 10.1189/ jlb.5BT0615-234R
- Boeckh-Behrens, T., Kleine, J. F., Zimmer, C., Neff, F., Scheipl, F., Pelisek, J., et al. (2016). Thrombus Histology Suggests Cardioembolic Cause in Cryptogenic Stroke. *Stroke* 47 (7), 1864–1871. doi: 10.1161/STROKEAHA. 116.013105
- Bonaventura, A., Vecchié, A., Abbate, A., and Montecucco, F. (2020). Neutrophil Extracellular Traps and Cardiovascular Diseases: An Update. *Cells* 9 (1), 231. doi: 10.3390/cells9010231
- Branzk, N., Lubojemska, A., Hardison, S. E., Wang, Q., Gutierrez, M. G., Brown, G. D., et al. (2014). Neutrophils Sense Microbe Size and Selectively Release

Neutrophil Extracellular Traps in Response to Large Pathogens. *Nat. Immunol.* 15 (11), 1017–1025. doi: 10.1038/ni.2987

- Brinkmann, V., Reichard, U., Goosmann, C., Fauler, B., Uhlemann, Y., DS, W., et al. (2004). Neutrophil Extracellular Traps Kill Bacteria. *Science* 303 (5663), 1532–1535. doi: 10.1126/science.1092385
- Chauhan, A. K., Motto, D. G., Lamb, C. B., Bergmeier, W., Dockal, M., Plaimauer, B., et al. (2006). Systemic Antithrombotic Effects of ADAMTS13. J. Exp. Med. 203 (3), 767–776. doi: 10.1084/jem.20051732
- Chrysanthopoulou, A., Mitroulis, I., Apostolidou, E., Arelaki, S., Mikroulis, D., Konstantinidis, T., et al. (2014). Neutrophil Extracellular Traps Promote Differentiation and Function of Fibroblasts. J. Pathol. 233 (3), 294–307. doi: 10.1002/path.4359
- Clark, S. R., Ma, A. C., Tavener, S. A., McDonald, B., Goodarzi, Z., Kelly, M. M., et al. (2007). Platelet TLR4 Activates Neutrophil Extracellular Traps to Ensnare Bacteria in Septic Blood. *Nat. Med.* 13 (4), 463–469. doi: 10.1038/nm1565
- Claushuis, T. A. M., van der Donk, L. E. H., Luitse, A. L., van Veen, H. A., van der Wel, N. N., van Vught, L. A., et al. (2018). Role of Peptidylarginine Deiminase 4 in Neutrophil Extracellular Trap Formation and Host Defense During *Klebsiella Pneumoniae*-Induced Pneumonia-Derived Sepsis. J. Immunol. 201 (4), 1241–1252. doi: 10.4049/jimmunol.1800314
- Conteduca, V., Scarpi, E., Wetterskog, D., Brighi, N., Ferroni, F., Rossi, A., et al. (2022). Plasma Tumor DNA is Associated With Increased Risk of Venous Thromboembolism in Metastatic Castration-Resistant Cancer Patients. *Int. J. Cancer* 150 (7), 1166–1173. doi: 10.1002/ijc.33834
- De Meyer, S. F., Andersson, T., Baxter, B., Bendszus, M., Brouwer, P., Brinjikji, W., et al. (2017). Analyses of Thrombi in Acute Ischemic Stroke: A Consensus Statement on Current Knowledge and Future Directions. *Int. J. Stroke* 12 (6), 606–614. doi: 10.1177/1747493017709671
- Döring, Y., Libby, P., and Soehnlein, O. (2020). Neutrophil Extracellular Traps Participate in Cardiovascular Diseases: Recent Experimental and Clinical Insights. Circ. Res. 126 (9), 1228–1241. doi: 10.1161/CIRCRESAHA.120.315931
- Ducroux, C., Di Meglio, L., Loyau, S., Delbosc, S., Boisseau, W., Deschildre, C., et al. (2018). Thrombus Neutrophil Extracellular Traps Content Impair tPA-Induced Thrombolysis in Acute Ischemic Stroke. *Stroke* 49 (3), 754–757. doi: 10.1161/STROKEAHA.117.019896
- El-Sayed, O. M., Dewyer, N. A., Luke, C. E., Elfline, M., Laser, A., Hogaboam, C., et al. (2016). Intact Toll-Like Receptor 9 Signaling in Neutrophils Modulates Normal Thrombogenesis in Mice. J. Vasc. Surg. 64 (5), 1450–1458.e1. doi: 10.1016/j.jvs.2015.08.070
- Engelmann, B., and Massberg, S. (2013). Thrombosis as an Intravascular Effector of Innate Immunity. *Nat. Rev. Immunol.* 13 (1), 34–45. doi: 10.1038/nri3345
- Etulain, J., Martinod, K., Wong, S. L., Cifuni, S. M., Schattner, M., and Wagner, D.
 D. (2015). P-Selectin Promotes Neutrophil Extracellular Trap Formation in Mice. *Blood* 126 (2), 242–246. doi: 10.1182/blood-2015-01-624023
- Farkas, Á. Z., Farkas, V. J., Gubucz, I., Szabó, L., Bálint, K., Tenekedjiev, K., et al. (2019). Neutrophil Extracellular Traps in Thrombi Retrieved During Interventional Treatment of Ischemic Arterial Diseases. *Thromb. Res.* 175, 46–52. doi: 10.1016/j.thromres.2019.01.006
- Fernandes, C. J., Morinaga, L. T. K., Alves, J. L. Jr, Castro, M. A., Calderaro, D., Jardim, C. V. P., et al. (2019). Cancer-Associated Thrombosis: The When, How and Why. *Eur. Respir. Rev.* 28 (151), 180119. doi: 10.1183/16000617.0119-2018
- Fernández-Ruiz, I. (2018). Inflammation: NETs Are Involved in AAA. Nat. Rev. Cardiol. 15 (5), 257. doi: 10.1038/ncardio.2018.28
- Folco, E. J., Mawson, T. L., Vromman, A., Bernardes-Souza, B., Franck, G., Persson, O., et al. (2018). Neutrophil Extracellular Traps Induce Endothelial Cell Activation and Tissue Factor Production Through Interleukin-1α and Cathepsin G. Arterioscler. Thromb. Vasc. Biol. 38 (8), 1901–1912. doi: 10.1161/ ATVBAHA.118.311150
- Franck, G., Mawson, T. L., Folco, E. J., Molinaro, R., Ruvkun, V., Engelbertsen, D., et al. (2018). Roles of PAD4 and NETosis in Experimental Atherosclerosis and Arterial Injury: Implications for Superficial Erosion. *Circ. Res.* 123 (1), 33–42. doi: 10.1161/CIRCRESAHA.117.312494
- George, J. N., and Nester, C. M. (2014). Syndromes of Thrombotic Microangiopathy. N Engl. J. Med. 371 (7), 654–666. doi: 10.1056/NEJMra1312353
- Goggs, R., Jeffery, U., LeVine, D. N., and Li, R. H. L. (2020). Neutrophil-Extracellular Traps, Cell-Free DNA, and Immunothrombosis in Companion Animals: A Review. Vet. Pathol. 57 (1), 6–23. doi: 10.1177/0300985819861721

- Gould, T. J., Vu, T. T., Stafford, A. R., Dwivedi, D. J., Kim, P. Y., Fox-Robichaud, A. E., et al. (2015). Cell-Free DNA Modulates Clot Structure and Impairs Fibrinolysis in Sepsis. Arterioscler. Thromb. Vasc. Biol. 35 (12), 2544–2553. doi: 10.1161/ATVBAHA.115.306035
- Gould, T. J., Vu, T. T., Swystun, L. L., Dwivedi, D. J., Mai, S. H., Weitz, J. I., et al. (2014). Neutrophil Extracellular Traps Promote Thrombin Generation Through Platelet-Dependent and Platelet-Independent Mechanisms. *Arterioscler. Thromb. Vasc. Biol.* 34 (9), 1977–1984. doi: 10.1161/ATVBAHA.114.304114
- Grilz, E., Mauracher, L. M., Posch, F., Königsbrügge, O., Zöchbauer-Müller, S., Marosi, C., et al. (2019). Citrullinated Histone H3, A Biomarker for Neutrophil Extracellular Trap Formation, Predicts the Risk of Mortality in Patients With Cancer. Br. J. Haematol. 186 (2), 311–320. doi: 10.1111/bjh.15906
- Guglietta, S., Chiavelli, A., Zagato, E., Krieg, C., Gandini, S., Ravenda, P. S., et al. (2016). Coagulation Induced by C3aR-Dependent NETosis Drives Protumorigenic Neutrophils During Small Intestinal Tumorigenesis. *Nat. Commun.* 7, 11037. doi: 10.1038/ncomms11037
- Haider, P., Kral-Pointner, J. B., Mayer, J., Richter, M., Kaun, C., Brostjan, C., et al. (2020). Neutrophil Extracellular Trap Degradation by Differently Polarized Macrophage Subsets. *Arterioscler. Thromb. Vasc. Biol.* 40 (9), 2265–2278. doi: 10.1161/ATVBAHA.120.314883
- Hisada, Y., Geddings, J. E., Ay, C., and Mackman, N. (2015). Venous Thrombosis and Cancer: From Mouse Models to Clinical Trials. *J. Thromb. Haemost.* 13 (8), 1372–1382. doi: 10.1111/jth.13009
- Hisada, Y., Grover, S. P., Maqsood, A., Houston, R., Ay, C., Noubouossie, D. F., et al. (2020). Neutrophils and Neutrophil Extracellular Traps Enhance Venous Thrombosis in Mice Bearing Human Pancreatic Tumors. *Haematologica* 105 (1), 218–225. doi: 10.3324/haematol.2019.217083
- Jiménez-Alcázar, M., Napirei, M., Panda, R., Köhler, E. C., Kremer Hovinga, J. A., Mannherz, H. G., et al. (2015). Impaired DNase1-Mediated Degradation of Neutrophil Extracellular Traps Is Associated With Acute Thrombotic Microangiopathies. J. Thromb. Haemost. 13 (5), 732–742. doi: 10.1111/ jth.12796
- Jiménez-Alcázar, M., Rangaswamy, C., Panda, R., Bitterling, J., Simsek, Y. J., Long, A. T., et al. (2017). Host DNases Prevent Vascular Occlusion by Neutrophil Extracellular Traps. *Science* 358 (6367), 1202–1206. doi: 10.1126/ science.aam8897
- Jorch, S. K., and Kubes, P. (2017). An Emerging Role for Neutrophil Extracellular Traps in Noninfectious Disease. *Nat. Med.* 23 (3), 279–287. doi: 10.1038/ nm.4294
- Kambas, K., Mitroulis, I., Apostolidou, E., Girod, A., Chrysanthopoulou, A., Pneumatikos, I., et al. (2012). Autophagy Mediates the Delivery of Thrombogenic Tissue Factor to Neutrophil Extracellular Traps in Human Sepsis. *PloS One* 7 (9), e45427. doi: 10.1371/journal.pone.0045427
- Kapoor, S., Opneja, A., and Nayak, L. (2018). The Role of Neutrophils in Thrombosis. *Thromb. Res.* 170, 87–96. doi: 10.1016/j.thromres.2018.08.005
- Kelly, P. J., Murphy, S., Coveney, S., Purroy, F., Lemmens, R., Tsivgoulis, G., et al. (2018). Anti-Inflammatory Approaches to Ischaemic Stroke Prevention. J. Neurol. Neurosurg. Psychiatry 89 (2), 211–218. doi: 10.1136/jnnp-2016-314817
- Kenny, E. F., Herzig, A., Krüger, R., Muth, A., Mondal, S., Thompson, P. R., et al. (2017). Diverse Stimuli Engage Different Neutrophil Extracellular Trap Pathways. *Elife* 6, e24437. doi: 10.7554/eLife.24437.032
- Kimball, A. S., Obi, A. T., Diaz, J. A., and Henke, P. K. (2016). The Emerging Role of NETs in Venous Thrombosis and Immunothrombosis. *Front. Immunol.* 7, 236. doi: 10.3389/fimmu.2016.00236
- Kim, S. W., and Lee, J. K. (2020). Role of HMGB1 in the Interplay Between NETosis and Thrombosis in Ischemic Stroke: A Review. *Cells* 9 (8), 1794. doi: 10.3390/cells9081794
- Langseth, M. S., Opstad, T. B., Bratseth, V., Solheim, S., Arnesen, H., Pettersen, A. Å., et al. (2018). Markers of Neutrophil Extracellular Traps are Associated With Adverse Clinical Outcome in Stable Coronary Artery Disease. *Eur. J. Prev. Cardiol.* 25 (7), 762–769. doi: 10.1177/2047487318760618
- Laridan, E., Denorme, F., Desender, L., François, O., Andersson, T., Deckmyn, H., et al. (2017). Neutrophil Extracellular Traps in Ischemic Stroke Thrombi. Ann. Neurol. 82 (2), 223–232. doi: 10.1002/ana.24993
- Laridan, E., Martinod, K., and De Meyer, S. F. (2019). Neutrophil Extracellular Traps in Arterial and Venous Thrombosis. Semin. Thromb. Hemost. 45 (1), 86– 93. doi: 10.1055/s-0038-1677040

- Leal, A. C., Mizurini, D. M., Gomes, T., Rochael, N. C., Saraiva, E. M., Dias, M. S., et al. (2017). Tumor-Derived Exosomes Induce the Formation of Neutrophil Extracellular Traps: Implications For The Establishment of Cancer-Associated Thrombosis. *Sci. Rep.* 7 (1), 6438. doi: 10.1038/s41598-017-06893-7
- Leffler, J., Prohászka, Z., Mikes, B., Sinkovits, G., Ciacma, K., Farkas, P., et al. (2017). Decreased Neutrophil Extracellular Trap Degradation in Shiga Toxin-Associated Haemolytic Uraemic Syndrome. *J. Innate Immun.* 9 (1), 12–21. doi: 10.1159/000450609
- Lewis, H. D., Liddle, J., Coote, J. E., Atkinson, S. J., Barker, M. D., Bax, B. D., et al. (2015). Inhibition of PAD4 Activity is Sufficient to Disrupt Mouse and Human NET Formation. *Nat. Chem. Biol.* 11 (3), 189–191. doi: 10.1038/ nchembio.1735
- Lim, W., Le Gal, G., Bates, S. M., Righini, M., Haramati, L. B., Lang, E., et al. (2018). American Society of Hematology 2018 Guidelines for Management of Venous Thromboembolism: Diagnosis of Venous Thromboembolism. *Blood Adv.* 2 (22), 3226–3256. doi: 10.1182/bloodadvances.2018024828
- Li, R. H. L., Ng, G., and Tablin, F. (2017). Lipopolysaccharide-Induced Neutrophil Extracellular Trap Formation in Canine Neutrophils is Dependent on Histone H3 Citrullination by Peptidylarginine Deiminase. Vet. Immunol. Immunopathol. 193-194, 29–37. doi: 10.1016/j.vetimm.2017.10.002
- Lood, C., Blanco, L. P., Purmalek, M. M., Carmona-Rivera, C., De Ravin, S. S., Smith, C. K., et al. (2016). Neutrophil Extracellular Traps Enriched in Oxidized Mitochondrial DNA are Interferogenic and Contribute to Lupus-Like Disease. *Nat. Med.* 22 (2), 146–153. doi: 10.1038/nm.4027
- Loof, T. G., Mörgelin, M., Johansson, L., Oehmcke, S., AI, O., Dickneite, G., et al. (2011). Coagulation, an Ancestral Serine Protease Cascade, Exerts a Novel Function in Early Immune Defense. *Blood* 118 (9), 2589–2598. doi: 10.1182/ blood-2011-02-337568
- Mackman, N., Bergmeier, W., Stouffer, G. A., and Weitz, J. I. (2020). Therapeutic Strategies for Thrombosis: New Targets and Approaches. *Nat. Rev. Drug Discovery* 19 (5), 333–352. doi: 10.1038/s41573-020-0061-0
- Mangold, A., Alias, S., Scherz, T., Hofbauer, M., Jakowitsch, J., Panzenböck, A., et al. (2015). Coronary Neutrophil Extracellular Trap Burden and Deoxyribonuclease Activity in ST-Elevation Acute Coronary Syndrome are Predictors of ST-Segment Resolution and Infarct Size. *Circ. Res.* 116 (7), 1182– 1192. doi: 10.1161/CIRCRESAHA.116.304944
- Martinod, K., Demers, M., Fuchs, T. A., Wong, S. L., Brill, A., Gallant, M., et al. (2013). Neutrophil Histone Modification by Peptidylarginine Deiminase 4 Is Critical for Deep Vein Thrombosis in Mice. *Proc. Natl. Acad. Sci. U. S. A.* 110 (21), 8674–8679. doi: 10.1073/pnas.1301059110
- Martinod, K., Fuchs, T. A., Zitomersky, N. L., Wong, S. L., Demers, M., Gallant, M., et al. (2015). PAD4-Deficiency Does Not Affect Bacteremia in Polymicrobial Sepsis and Ameliorates Endotoxemic Shock. *Blood* 125 (12), 1948–1956. doi: 10.1182/blood-2014-07-587709
- Martinod, K., and Wagner, D. D. (2014). Thrombosis: Tangled Up in NETs. *Blood* 123 (18), 2768–2776. doi: 10.1182/blood-2013-10-463646
- Maugeri, N., Campana, L., Gavina, M., Covino, C., De Metrio, M., Panciroli, C., et al. (2014). Activated Platelets Present High Mobility Group Box 1 to Neutrophils, Inducing Autophagy and Promoting the Extrusion of Neutrophil Extracellular Traps. J. Thromb. Haemost. 12 (12), 2074–2088. doi: 10.1111/jth.12710
- Mauracher, L. M., Posch, F., Martinod, K., Grilz, E., Däullary, T., Hell, L., et al. (2018). Citrullinated Histone H3, a Biomarker of Neutrophil Extracellular Trap Formation, Predicts the Risk of Venous Thromboembolism in Cancer Patients. *J. Thromb. Haemost.* 16 (3), 508–518. doi: 10.1111/jth.13951
- McDonald, B., Urrutia, R., Yipp, B. G., Jenne, C. N., and Kubes, P. (2012). Intravascular Neutrophil Extracellular Traps Capture Bacteria From the Bloodstream During Sepsis. *Cell Host Microbe* 12 (3), 324–333. doi: 10.1016/ i.chom.2012.06.011
- Meng, H., Yalavarthi, S., Kanthi, Y., Mazza, L. F., Elfline, M. A., Luke, C. E., et al. (2017). *In Vivo* Role of Neutrophil Extracellular Traps in Antiphospholipid Antibody-Mediated Venous Thrombosis. *Arthritis Rheumatol.* 69 (3), 655– 667. doi: 10.1002/art.39938
- Mizuno, T., Yoshioka, K., Mizuno, M., Shimizu, M., Nagano, F., Okuda, T., et al. (2017). Complement Component 5 Promotes Lethal Thrombosis. Sci. Rep. 7, 42714. doi: 10.1038/srep42714
- Mohammed, B. M., Fisher, B. J., Kraskauskas, D., Farkas, D., Brophy, D. F., Fowler, A. A.3rd, et al. (2013). Vitamin C: A Novel Regulator of Neutrophil Extracellular Trap Formation. *Nutrients* 5 (8), 3131–3151. doi: 10.3390/nu5083131

Mutua, V., and Gershwin, L. J. (2021). A Review of Neutrophil Extracellular Traps (NETs) in Disease: Potential Anti-NETs Therapeutics. *Clin. Rev. Allergy Immunol.* 61 (2), 194–211. doi: 10.1007/s12016-020-08804-7

NETs in Thrombosis Formation

- Noubouossie, D. F., Reeves, B. N., Strahl, B. D., and Key, N. S. (2019). Neutrophils: Back in the Thrombosis Spotlight. *Blood* 133 (20), 2186–2197. doi: 10.1182/ blood-2018-10-862243
- Noubouossie, D. F., Whelihan, M. F., Yu, Y. B., Sparkenbaugh, E., Pawlinski, R., Monroe, D. M., et al. (2017). *In Vitro* Activation of Coagulation by Human Neutrophil DNA and Histone Proteins But Not Neutrophil Extracellular Traps. *Blood* 129 (8), 1021–1029. doi: 10.1182/blood-2016-06-722298
- Okeke, E. B., Louttit, C., Fry, C., Najafabadi, A. H., Han, K., Nemzek, J., et al. (2020). Inhibition of Neutrophil Elastase Prevents Neutrophil Extracellular Trap Formation and Rescues Mice From Endotoxic Shock. *Biomaterials* 238, 119836. doi: 10.1016/j.biomaterials.2020.119836
- Oklu, R., Sheth, R. A., Wong, K. H. K., Jahromi, A. H., and Albadawi, H. (2017). Neutrophil Extracellular Traps are Increased in Cancer Patients But Does Not Associate With Venous Thrombosis. *Cardiovasc. Diagn. Ther.* 7 (Suppl 3), S140–S149. doi: 10.21037/cdt.2017.08.01
- Osada, K., Minami, T., Arioka, T., Sakai, T., Tawara, S., Kawasaki, K., et al. (2017). Thrombomodulin Alfa Attenuates the Procoagulant Effect and Cytotoxicity of Extracellular Histones Through the Promotion of Protein C Activation. *Thromb. Res.* 160, 51–57. doi: 10.1016/j.thromres.2017.10.019
- Palmer, L. J., Damgaard, C., Holmstrup, P., and Nielsen, C. H. (2016). Influence of Complement on Neutrophil Extracellular Trap Release Induced by Bacteria. *J. Periodontal. Res.* 51 (1), 70–76. doi: 10.1111/jre.12284
- Papayannopoulos, V. (2018). Neutrophil Extracellular Traps in Immunity and Disease. Nat. Rev. Immunol. 18 (2), 134–147. doi: 10.1038/nri.2017.105
- Pierangeli, S. S., Girardi, G., Vega-Ostertag, M., Liu, X., Espinola, R. G., and Salmon, J. (2005). Requirement of Activation of Complement C3 and C5 for Antiphospholipid Antibody-Mediated Thrombophilia. *Arthritis Rheumatol.* 52 (7), 2120–2124. doi: 10.1002/art.21157
- Ramos, M. V., Mejias, M. P., Sabbione, F., Fernandez-Brando, R. J., Santiago, A. P., Amaral, M. M., et al. (2016). Induction of Neutrophil Extracellular Traps in Shiga Toxin-Associated Hemolytic Uremic Syndrome. *J. Innate Immun.* 8 (4), 400–411. doi: 10.1159/000445770
- Riegger, J., Byrne, R. A., Joner, M., Chandraratne, S., Gershlick, A. H., Ten Berg, J. M., et al. (2016). Prevention of Late Stent Thrombosis by an Interdisciplinary Global European Effort (PRESTIGE) Investigators. Histopathological Evaluation of Thrombus in Patients Presenting With Stent Thrombosis. A Multicenter European Study: A Report of the Prevention of Late Stent Thrombosis by an Interdisciplinary Global European Effort Consortium. *Eur. Heart J.* 37 (19), 1538–1549. doi: 10.1093/erheartj/ehv419
- Savchenko, A. S., Martinod, K., Seidman, M. A., Wong, S. L., Borissoff, J. I., Piazza, G., et al. (2014). Neutrophil Extracellular Traps Form Predominantly During the Organizing Stage of Human Venous Thromboembolism Development. J. Thromb. Haemost. 12 (6), 860–870. doi: 10.1111/jth.12571
- Semeraro, N., Ammollo, C. T., Semeraro, F., and Colucci, M. (2012). Sepsis, Thrombosis and Organ Dysfunction. *Thromb. Res.* 129 (3), 290–295. doi: 10.1016/j.thromres.2011.10.013
- Singer, M., Deutschman, C. S., Seymour, C. W., Shankar-Hari, M., Annane, D., Bauer, M., et al. (2016). The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). JAMA 315 (8), 801–810. doi: 10.1001/ jama.2016.0287
- Sollberger, G., Choidas, A., Burn, G. L., Habenberger, P., Di Lucrezia, R., Kordes, S., et al. (2018). Gasdermin D Plays a Vital Role in the Generation of Neutrophil Extracellular Traps. *Sci. Immunol.* 3 (26), eaar6689. doi: 10.1126/ sciimunol.aar6689
- Stevens, S. M., Woller, S. C., Bauer, K. A., Kasthuri, R., Cushman, M., Streiff, M., et al. (2016). Guidance for the Evaluation and Treatment of Hereditary and Acquired Thrombophilia. J. Thromb. Thrombol 41 (1), 154–164. doi: 10.1007/ s11239-015-1316-1
- Tardif, J. C., Kouz, S., Waters, D. D., Bertrand, O. F., Diaz, R., Maggioni, A. P., et al. (2019). Efficacy and Safety of Low-Dose Colchicine After Myocardial Infarction. N. Engl. J. Med. 381 (26), 2497–2505. doi: 10.1056/ NEJMoa1912388
- Thålin, C., Hisada, Y., Lundström, S., Mackman, N., and Wallén, H. (2019). Neutrophil Extracellular Traps: Villains and Targets in Arterial, Venous, and Cancer-Associated Thrombosis. *Arterioscler. Thromb. Vasc. Biol.* 39 (9), 1724– 1738. doi: 10.1161/ATVBAHA.119.312463

- Thiam, H. R., Wong, S. L., Qiu, R., Kittisopikul, M., Vahabikashi, A., Goldman, A. E., et al. (2020). NETosis Proceeds by Cytoskeleton and Endomembrane Disassembly and PAD4-Mediated Chromatin Decondensation and Nuclear Envelope Rupture. *Proc. Natl. Acad. Sci. U. S. A.* 117 (13), 7326–7337. doi: 10.1073/pnas.1909546117
- Thiam, H. R., Wong, S. L., Wagner, D. D., and Waterman, C. M. (2020). Cellular Mechanisms of NETosis. Annu. Rev. Cell Dev. Biol. 36, 191–218. doi: 10.1146/ annurev-cellbio-020520-111016
- Thomas, M. R., and Storey, R. F. (2015). The Role of Platelets in Inflammation. *Thromb. Haemost.* 114 (3), 449–458. doi: 10.1160/TH15-12-1067
- Tsao, C. M., Ho, S. T., and Wu, C. C. (2015). Coagulation Abnormalities in Sepsis. Acta Anaesthesiol. Taiwan 53 (1), 16–22. doi: 10.1016/j.aat.2014.11.002
- Van Avondt, K., and Hartl, D. (2018). Mechanisms and Disease Relevance of Neutrophil Extracellular Trap Formation. *Eur. J. Clin. Invest* 48 Suppl 2, e12919. doi: 10.1111/eci.12919
- van der Linden, M., van den Hoogen, L. L., Westerlaken, G. H. A., Fritsch-Stork, R. D. E., van Roon, J. A. G., Radstake, T. R. D. J., et al. (2018). Neutrophil Extracellular Trap Release Is Associated With Antinuclear Antibodies in Systemic Lupus Erythematosus and Anti-Phospholipid Syndrome. *Rheumatol.* (*Oxf*) 57 (7), 1228–1234. doi: 10.1093/rheumatology/key067
- van Montfoort, M. L., Stephan, F., Lauw, M. N., Hutten, B. A., Van Mierlo, G. J., Solati, S., et al. (2013). Circulating Nucleosomes and Neutrophil Activation as Risk Factors for Deep Vein Thrombosis. *Arterioscler. Thromb. Vasc. Biol.* 33 (1), 147–151. doi: 10.1161/ATVBAHA.112.300498
- Vannucchi, A. M., Kiladjian, J. J., Griesshammer, M., Masszi, T., Durrant, S., Passamonti, F., et al. (2015). Ruxolitinib Versus Standard Therapy for the Treatment of Polycythemia Vera. N. Engl. J. Med. 372 (5), 426–435. doi: 10.1056/NEJMoa1409002
- Vu, T. T., Leslie, B. A., Stafford, A. R., Zhou, J., Fredenburgh, J. C., and Weitz, J. I. (2016). Histidine-Rich Glycoprotein Binds DNA and RNA and Attenuates Their Capacity to Activate the Intrinsic Coagulation Pathway. *Thromb. Haemost.* 115 (1), 89–98. doi: 10.1160/TH15-04-0336
- Wang, Y., Luo, L., Braun, O. Ö., Westman, J., Madhi, R., Herwald, H., et al. (2018). Neutrophil Extracellular Trap-Microparticle Complexes Enhance Thrombin Generation via the Intrinsic Pathway of Coagulation in Mice. Sci. Rep. 8 (1), 4020. doi: 10.1038/s41598-018-22156-5
- Warnatsch, A., Ioannou, M., Wang, Q., and Papayannopoulos, V. (2015). Inflammation. Neutrophil Extracellular Traps License Macrophages for Cytokine Production in Atherosclerosis. *Science* 349 (6245), 316–320.
- Wendelboe, A. M., and Raskob, G. E. (2016). Global Burden of Thrombosis: Epidemiologic Aspects. *Circ. Res.* 118 (9), 1340–1347. doi: 10.1161/ CIRCRESAHA.115.306841
- Wiedmer, T., Esmon, C. T., and Sims, P. J. (1986). On the Mechanism by Which Complement Proteins C5b-9 Increase Platelet Prothrombinase Activity. J. Biol. Chem. 261 (31), 14587–14592. doi: 10.1016/S0021-9258(18)66911-X
- Wirestam, L., Arve, S., Linge, P., and Bengtsson, A. A. (2019). Neutrophils-Important Communicators in Systemic Lupus Erythematosus and Antiphospholipid Syndrome. *Front. Immunol.* 10, 2734. doi: 10.3389/ fmmu.2019.02734

- Wolach, O., Sellar, R. S., Martinod, K., Cherpokova, D., McConkey, M., Chappell,
 R. J., et al. (2018). Increased Neutrophil Extracellular Trap Formation
 Promotes Thrombosis in Myeloproliferative Neoplasms. *Sci. Transl. Med.* 10 (436), eaan8292. doi: 10.1126/scitranslmed.aan8292
- Wong, S. L., Goverman, J., Staudinger, C., and Wagner, D. D. (2020). Recombinant Human ADAMTS13 Treatment and Anti-NET Strategies Enhance Skin Allograft Survival in Mice. Am. J. Transpl. 20 (4), 1162–1169. doi: 10.1111/ajt.15703
- Yalavarthi, S., Gould, T. J., Rao, A. N., Mazza, L. F., Morris, A. E., Núñez-Álvarez, C., et al. (2015). Release of Neutrophil Extracellular Traps by Neutrophils Stimulated With Antiphospholipid Antibodies: A Newly Identified Mechanism of Thrombosis in the Antiphospholipid Syndrome. *Arthritis Rheumatol.* 67 (11), 2990–3003. doi: 10.1002/art.39247
- Yang, X., Li, L., Liu, J., Lv, B., and Chen, F. (2016). Extracellular Histones Induce Tissue Factor Expression in Vascular Endothelial Cells *via* TLR and Activation of NF-κb and AP-1. *Thromb. Res.* 137, 211–218. doi: 10.1016/ j.thromres.2015.10.012
- Yang, S., Qi, H., Kan, K., Chen, J., Xie, H., Guo, X., et al. (2017). Neutrophil Extracellular Traps Promote Hypercoagulability in Patients With Sepsis. *Shock* 47 (2), 132–139. doi: 10.1097/SHK.00000000000741
- Yang, C., Sun, W., Cui, W., Li, X., Yao, J., Jia, X., et al. (2015). Procoagulant Role of Neutrophil Extracellular Traps in Patients With Gastric Cancer. Int. J. Clin. Exp. Pathol. 8 (11), 14075–14086.
- Yuen, J., Pluthero, F. G., Douda, D. N., Riedl, M., Cherry, A., Ulanova, M., et al. (2016). NETosing Neutrophils Activate Complement Both on Their Own NETs and Bacteria via Alternative and Non-Alternative Pathways. Front. Immunol. 7, 137. doi: 10.3389/fimmu.2016.00137
- Zuo, Y., Yalavarthi, S., Gockman, K., Madison, J. A., Gudjonsson, J. E., Kahlenberg, J. M., et al. (2020). Anti-Neutrophil Extracellular Trap Antibodies and Impaired Neutrophil Extracellular Trap Degradation in Antiphospholipid Syndrome. *Arthritis Rheumatol.* 72 (12), 2130–2135. doi: 10.1002/art.41460

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Zhou, Xu and Liu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.