



# Non-Targeted Metabolomic Analysis of Chicken Kidneys in Response to Coronavirus IBV Infection Under Stress Induced by Dexamethasone

Jun Dai<sup>1,2</sup>, Huan Wang<sup>2</sup>, Ying Liao<sup>2</sup>, Lei Tan<sup>2</sup>, Yingjie Sun<sup>2</sup>, Cuiping Song<sup>2</sup>, Weiwei Liu<sup>2</sup>, Chan Ding<sup>2,3</sup>, Tingrong Luo<sup>1,4\*</sup> and Xusheng Qiu<sup>2\*</sup>

## OPEN ACCESS

### Edited by:

Muhammad Imran Arshad,  
University of Agriculture, Pakistan

### Reviewed by:

Guijing Shen,  
Xiamen University, China  
Jinping Gu,  
Zhejiang University of Technology,  
China

### \*Correspondence:

Xusheng Qiu  
xsqiu1981@shvri.ac.cn  
Tingrong Luo  
tingrongluo@gxu.edu.cn

### Specialty section:

This article was submitted to  
Virus and Host,  
a section of the journal  
Frontiers in Cellular and  
Infection Microbiology

Received: 17 May 2022

Accepted: 20 June 2022

Published: 15 July 2022

### Citation:

Dai J, Wang H,  
Liao Y, Tan L, Sun Y, Song C,  
Liu W, Ding C, Luo T and Qiu X (2022)  
Non-Targeted Metabolomic Analysis  
of Chicken Kidneys in Response  
to Coronavirus IBV Infection Under  
Stress Induced by Dexamethasone.  
*Front. Cell. Infect. Microbiol.* 12:945865.  
doi: 10.3389/fcimb.2022.945865

<sup>1</sup> Laboratory of Veterinary Microbiology and Animal Infectious Diseases, College of Animal Sciences and Veterinary Medicine, Guangxi University, Nanning, China, <sup>2</sup> Shanghai Veterinary Research Institute, Chinese Academy of Agricultural Sciences, Shanghai, China, <sup>3</sup> Jiangsu Co-innovation Center for Prevention and Control of Important Animal Infectious Diseases and Zoonoses, Yangzhou University, Yangzhou, China, <sup>4</sup> State Key Laboratory for Conservation and Utilization of Subtropical Agro-Bioresources, Guangxi University, Nanning, China

Stress in poultry can lead to changes in body metabolism and immunity, which can increase susceptibility to infectious diseases. However, knowledge regarding chicken responses to viral infection under stress is limited. Dexamethasone (Dex) is a synthetic glucocorticoid similar to that secreted by animals under stress conditions, and has been widely used to induce stress in chickens. Herein, we established a stress model in 7-day-old chickens injected with Dex to elucidate the effects of stress on IBV replication in the kidneys. The metabolic changes, immune status and growth of the chickens under stress conditions were comprehensively evaluated. Furthermore, the metabolic profile, weight gain, viral load, serum cholesterol levels, cytokines and peripheral blood lymphocyte ratio were compared in chickens treated with Dex and infected with IBV. An LC-MS/MS-based metabolomics method was used to examine differentially enriched metabolites in the kidneys. A total of 113 metabolites whose abundance was altered after Dex treatment were identified, most of which were lipids and lipid-like molecules. The principal metabolic alterations in chicken kidneys caused by IBV infection included fatty acid, valine, leucine and isoleucine metabolism. Dex treatment before and after IBV infection mainly affected the host's tryptophan, phenylalanine, amino sugar and nucleotide sugar metabolism. In addition, Dex led to up-regulation of serum cholesterol levels and renal viral load in chickens, and to the inhibition of weight gain, peripheral blood lymphocytes and IL-6 production. We also confirmed that the exogenous cholesterol in DF-1 cells promoted the replication of IBV. However, whether the increase in viral load in kidney tissue is associated with the up-regulation of cholesterol levels induced by Dex must be demonstrated in future

experiments. In conclusion, chick growth and immune function were significantly inhibited by Dex. Host cholesterol metabolism and the response to IBV infection are regulated by Dex. This study provides valuable insights into the molecular regulatory mechanisms in poultry stress, and should support further research on the intrinsic link between cholesterol metabolism and IBV replication under stress conditions.

**Keywords:** dexamethasone, stress, non-targeted metabolomic, coronavirus, IBV

## INTRODUCTION

Animal stress is a complex, multidimensional and universal phenomenon that is difficult to evaluate and has important biological significance (Buwalda et al., 2012). Poultry stress is a global problem that causes large economic losses and threatens poultry health and welfare (Yang et al., 2014; Zaboli et al., 2019). Multiple stressors are present during the growth of chickens, including immune stress, oxidative stress, transport stress and cold or heat stress (Sun et al., 2018; Guo et al., 2020; Goel et al., 2021). Compared with mammals, chickens have a higher body temperature and are extremely sensitive to temperature changes during the breeding process (Lara and Rostagno, 2013; Collier and Gebremedhin, 2015), particularly in cold regions (Blahová et al., 2007). Cold or heat stress is the most common environmental stressor in chickens, and may cause pathophysiological stress (Castellani and Young, 2016; Lin et al., 2006; Wang et al., 2018; Hu and Cheng, 2021). For example, cold or heat stress not only affects tissue inflammatory factor levels (Zhao et al., 2013; Zhang et al., 2014) but also causes severe damage to the liver, heart and intestinal tissues in chickens (Zhang et al., 2011; Wei et al., 2018). Moreover, stress has been found to significantly increase susceptibility to necrotic enteritis and enhance bacterial translocation in a subclinical experimental model (Tsiouris et al., 2015; Borsoi et al., 2015; Zhou et al., 2021). Evidence has indicated that cold stress decreases immune function in animals, and temperature downshifts can make animals susceptible to viral infection. For example, outbreaks of H1N1 (Steel et al., 2010), RSV (Donaldson, 2006; Noyola and Mandeville, 2008; Tang, 2009; Zhang et al., 2013), PEDV (Kong et al., 2020), NDV (Pfitzer et al., 2000) and RV (Foxman et al., 2015) usually occur in cold winter seasons. Previous studies on stress in poultry have focused on changes in cerebellar development (Austdal et al., 2016), energy intake (Benson et al., 1993; Garriga et al., 2006; Raghebian et al., 2016; Hu et al., 2021), growth (Nawab et al., 2018; Chen et al., 2021; Kumar et al., 2021), inflammatory responses (Wein et al., 2017; Lan et al., 2020; Liu et al., 2021), oxidative responses (Gao et al., 2010; Jiao et al., 2018; Pan et al., 2019) and immunosuppression (Guo et al., 2020; Guo et al., 2021; Su et al., 2021). However, the detailed molecular mechanisms linking stress and viral replication remain unclear.

Studies have indicated that the hypothalamic-pituitary-adrenal (HPA) axis is an important system for integrating and regulating stress responses *in vivo* and *in vitro* (McEwen, 2000; Iyasere et al., 2017; Beckford et al., 2020). In general, various stressors can cause changes in certain HPA axes that are required for stress adaptation (Scheff et al., 2012). Activation of the HPA axis and sympathetic

nervous system when chickens are exposed to stress induces the rapid secretion of glucocorticoids (GCs) from the adrenal glands (Chrousos and Gold, 1992; Aguilera, 1994; Lin et al., 2006; Chrousos, 2009; Borsoi et al., 2015; Scanes, 2016). GCs are a class of steroid hormones that respond rapidly to environmental and physiological stimuli (Carsia et al., 1988; Revollo and Cidlowski, 2009; Alba et al., 2019; Häffelin et al., 2020). Excessive GCs are a sign of stress (Chrousos and Gold, 1992; Webster Marketon and Glaser, 2008; Quinteiro-Filho et al., 2017). Through the release of GCs, the HPA axis mobilizes energy reserves, thus ensuring that the organism has sufficient resources to respond to actual bodily harm or to prepare for anticipated harm and alleviate the adverse effects (Herman et al., 2016; Zheng et al., 2021). Additionally, GCs, the final product of the HPA axis, are cholesterol-derived molecules that exert pleiotropic and nongenomic effects through the glucocorticoid receptor (GR) (Chrousos and Kino, 2007). GCs also play an important role in maintaining resting and stress-associated homeostasis and affects the body's physiological adaptive response to stressors (Nicolaidis et al., 2015). The secretion of GC is a typical stress endocrine response (Sapolsky et al., 2000; Flinchum and Gwen, 2015) that is particularly important in poultry stress biology (Scanes, 2016). In poultry, two of the most common physiological parameters of stress are circulating concentrations of the adreno-cortical hormone corticosterone (CORT) and the heterophil:lymphocyte ratio (H:L) (Kalliecharan and Hall, 1974; Scanes, 2016). CORT is the main poultry glucocorticoid that regulates energy reserves to meet metabolic demands. Poultry stress causes elevated plasma CORT, stimulates glucocorticoid receptors, and may promote glycemia, lipolysis and proteolysis (Deviche et al., 2017). Therefore, proper control of the stress response is critical for the body, because inappropriate or prolonged activation of the HPA axis is energy-draining and has been implicated in many physiological disease states (Myers et al., 2012). Dex is a synthetic glucocorticoid similar to that secreted by animals under stress conditions (Lin et al., 2004; Gao et al., 2008), which has been used in many studies as a glucocorticoid mimic (Chang et al., 2015). In particular, Dex is widely used in the establishment of chicken stress models (Eid et al., 2006; Gao et al., 2010; El-Senousey et al., 2018; Pan et al., 2019; Zhou et al., 2019; Zhai et al., 2020; Osho and Adeola, 2020; Su et al., 2021). Therefore, Dex is a satisfactory stress-inducing drug that provides an effective way to study the relationship between stress and viral replication.

Herein, we established a chicken stress model by using Dex and found that Dex-induced chicken stress increased the IBV viral load in the kidneys. Untargeted metabolomics was used to detect differentially enriched metabolites in chicken kidney tissue

to evaluate the correlations between IBV replication and metabolic changes induced by Dex. Serum cholesterol levels significantly increased in chickens under stress. Many metabolic pathways and metabolites may be associated with the replication and pathogenicity of IBV. This finding provides insight into the intrinsic link between host metabolism and IBV replication under stress conditions.

## MATERIALS AND METHODS

### Cell Culture and Virus

DF-1 (ATCC, CRL-12203), Vero and H1299 cell lines were obtained from the American Type Culture Collection (ATCC) (Wang et al., 2019; Wang et al., 2021). The IBV QX and Beaudette strains (ATCC VR-22) were kept at the Shanghai Veterinary Research Institute, Chinese Academy of Agricultural Sciences, Shanghai (Wang et al., 2021; Gao et al., 2021). The IBV Beaudette strain used in this study was adapted to DF-1, H1299 and Vero cells, and was a gift from Prof. Dingxiang Liu's laboratory (South China Agricultural University) (Wang et al., 2019; Gao et al., 2021). The culture of DF-1, Vero and H1299 cell lines was performed according to previously described methods in our laboratory (Wang et al., 2019; Liu et al., 2019; Gong et al., 2021). In brief, DF-1 and Vero cells were grown in Dulbecco's modified Eagle's medium with 10% fetal calf serum. H1299 cells were maintained in Roswell Park Memorial Institute 1640 medium supplemented with 10% (v/v) fetal calf serum. The above cells were cultured at 37 °C under 5% CO<sub>2</sub>.

### Antibodies, Reagents and Chickens

Anti-IBV N antibody was obtained by immunization of a rabbit with IBV N antigen in our laboratory as previously described (Wang et al., 2021). CD4-FITC, CD8-PE and CD3-SPRD monoclonal antibodies were purchased from Southern Biotech (USA). The 25-hydroxycholesterol (HY-113134) and cholesterol (HY-N0322) were purchased from MedChemExpress. Dexamethasone sodium phosphate injection was purchased from Jiangxi Bolai Pharmacy Co., Ltd. Plasma total cholesterol (TC), high-density lipoprotein cholesterol (HDL-C) and low-density lipoprotein cholesterol (LDL-C) concentrations were assessed with an automatic biochemical analyzer (Rayto, chemray240). A lymphocyte separation kit (P8740) was purchased from Beijing Soleibao Technology Co., Ltd. ELISA kits for IFN- $\gamma$  (ZC-51624), IL-1 $\beta$  (ZC-51658), TNF- $\alpha$  (ZC-51975), IL-6 (ZC-51663) and IFN- $\beta$  (ZC-51619) were purchased from Shanghai Zhuo Cai Technology Co., Ltd. All specific pathogen free (SPF) embryonated eggs were purchased from Beijing Boehringer Ingelheim Vital Biotechnology Co., Ltd. (Beijing, China) and incubated as previously described (van de Ven et al., 2011; Maatjens et al., 2014; Mesquita et al., 2021). In summary, SPF embryonated eggs were incubated for 17 d in a multi-stage incubator (science incubator TM, Shandong, China) at a constant machine temperature of 37.6°C and relative humidity of 53%. Eggs were placed with the air cell up and turned hourly at an angle of 90°. From E17 to E21.5, the eggs

were not turned, the air temperature remained constant at 35.0° C, and the air speed was lower than 0.2 m/s, which is considered still air.

### Western Blot Analysis

DF-1 cells, Vero cells and H1299 cells were harvested at the indicated infection time points and lysed with 2 $\times$  SDS loading buffer in the presence of 100 mM dithiothreitol, then denatured at 100°C for 10 min. The obtained protein samples were separated with SDS-PAGE to detect intracellular IBV protein levels according to a previously described method (Wang et al., 2021).

### Animal Experiments

A total of 260 healthy White Leghorns of similar weight were randomly and averagely divided into five groups: mock (N=52), Dex (N=52), IBV (N=52), IBV-P-Dex (pretreatment with Dex before infection, N=52) and IBV-A-Dex (treatment with Dex after infection, N=52). The feeding and care of chickens was performed in accordance with the Institutional Animal Care and Use Committee guidelines. The approval number was SHVRI-chicken-2022021802. According to previous studies (Zhou et al., 2019; Guo et al., 2020; Su et al., 2021), a chicken stress model was established through subcutaneous injection of Dex (2.0 mg/kg, once/day), and PBS was used as a negative control. Seven-day-old SPF chickens were challenged *via* an eye dropper with 200  $\mu$ l of 10<sup>5</sup> EID<sub>50</sub> of IBV-QX or PBS as the negative control. Chickens were observed daily, and their clinical symptoms and mortality were recorded. At 7 dpi (14 days old), the chickens' responses to IBV infection under stress induced by Dex were comprehensively assessed through measurement of viral load, histopathology, metabolomics, serum cholesterol concentration, peripheral blood lymphocyte ratio, survival rate and growth performance (**Supplementary Materials Figure S1** shows the experimental design).

### Histopathology

The trachea, heart, lung, liver, kidney and spleen tissues were isolated and soaked in 10% neutral formalin at room temperature for more than 48 h, then subjected to routine processing. H&E staining was performed, and the slices of organs for each chicken were observed under an optical microscope.

### Sample Preparation and Extraction

Samples were thawed on ice, and 50 mg tissue fragments were taken from each sample and mixed with 500  $\mu$ l ice-cold methanol/water (70%, v/v). After homogenization at 30 Hz for 2 min, the mixture was vibrated for 5 min and incubated on ice for 15 min. After centrifugation at 12,000 rpm at 4°C for 10 min, 400  $\mu$ l supernatant was transferred into another centrifuge tube; subsequently, 500  $\mu$ l of ethyl acetate/methanol (V, 1:3) was added into the original centrifuge tube. The mixture was oscillated for 5 min, incubated on ice for 15 min and centrifuged at 12,000 rpm at 4°C for 10 min, and 400  $\mu$ l of supernatant was collected. The two supernatants were combined and concentrated. Then 100  $\mu$ l of 70% methanol water was added

to the dried product, and ultrasonic treatment was performed for 3 min. Finally, samples were centrifuged at 12,000 rpm at 4°C for 3 min, and 60 µl of supernatant was aspirated for LC-MS/MS analysis.

## Metabolomics Analysis

Untargeted metabolomics profiling was performed using a UPLC-Q-TOF/MS (AB SCIEX, MA, USA). The chromatographic separation system was equipped with a Waters ACQUITY UPLC HSS T3 column (1.8 µm, 2.1 mm × 100 mm, Waters Co.). UPLC conditions were as follows: column temperature, 40°C; flow rate, 0.40 mL/min; injection volume, 1 µl. The mobile phase consisted of 0.1% formic acid in water (phase A), and 0.1% formic acid in acetonitrile (phase B). The linear gradient program was as follows: 95% to 10% phase A over 11 min, and holding 10% for 1 min, and 10% to 95% in 0.1 min, and holding 95% phase A for 1.9 min. The mass spectrometer was operated in positive/negative polarity mode with the following settings: Nitrogen was used as the drying gas, nebulizer gas and sheath gas, and the flow was maintained at 8 L/min, while sheath flow was at 11 L/min. The drying gas and sheath gas temperature was maintained at 325°C. The ESI+ and ESI- voltages were set at 2500 V and 1500 V respectively. Mass range was set at m/z 50-1700 and the resolution was 30,000 (FWHM). The mass spectrometer was calibrated daily in the mass range m/z 100-1700 before starting the sample analysis by using Agilent tune mix (Part no G1969-85000). The mass accuracy values were good in full scan range (mass error < 5 ppm).

## Quantitative Real-Time PCR Analysis

Total RNA from kidneys was used for quantitative real-time PCR (qPCR) analysis to confirm the propagation of the IBV in chicks, and the mock group was used as a negative control. The primer pairs used in the PCR assay for the IBV-N gene were IBV Fw-5'-CAG AAG AAG GGC TCT CGC ATT AC-3' and IBV Re-5'-AGG TTG AGC ATT GCC GTA ACA C-3'. Positive and negative strands of IBV genomic RNA were detected according to our previous report (Liao et al., 2011). The qPCR specific primers for ch-DHCR24 and ch-CH25H were designed with reference to previous literature (Liu et al., 2019). RNA was extracted with an RNeasy Mini Kit (QIAGEN, ID: 74104, Germany) according to the manufacturer's protocols. Validation of RNA-seq data by qPCR was performed as previously described (Liu et al., 2018).

## Statistical Analysis

The original data file obtained by LC-MS analysis was first converted into mzML format in Proteo Wizard software. Peak extraction, alignment and retention time correction were performed with the XCMS program. The "Support Vector Regression (SVR)" method was used to correct the peak area (Vapnik et al., 2008; Al-Zoubi et al., 2011; Sánchez-Illana et al., 2018). The peaks were filtered with a deletion rate > 50% in each group of samples. Metabolic identification information was then obtained by searching the laboratory's self-built database (Wuhan Metware Biotechnology Co., Ltd.) and integrating the public database and metDNA. Finally, statistical analysis was performed in the R program (**Supplementary Materials**

**Table S1**). Statistical analysis included univariate analysis and multivariate analysis, and univariate statistical analysis included Student's t-test and variance multiple analysis. Multivariate statistical analysis included principal component analysis (PCA), partial least squares discriminant analysis (PLS-DA) and orthogonal partial least squares discriminant analysis (OPLS-DA). The identification of differentially enriched metabolites and metabolic pathways was as previously described (Liu et al., 2019).

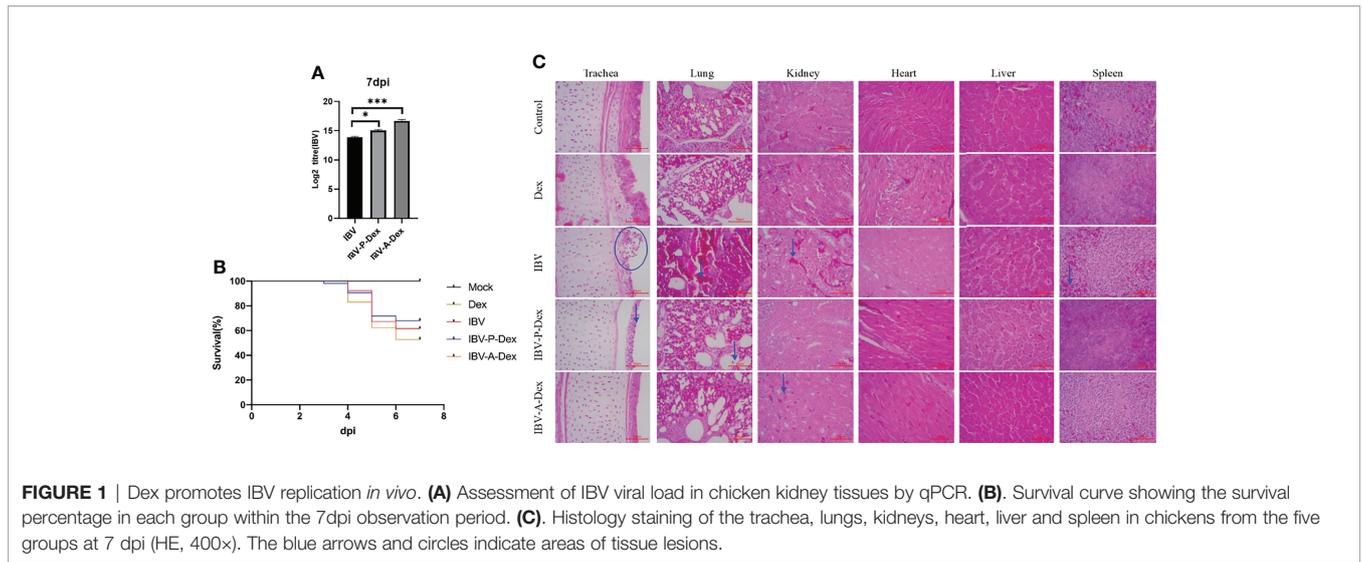
## RESULTS

### Dex Induces Weight Loss in Chickens and Promotes IBV Replication *In Vivo*

To evaluate the effects of Dex treatment and IBV infection on chicken growth, we weighed chickens at 1, 7 and 14 days of age. As shown in the **Supplementary Materials Table S2**, the body weights of the Dex and IBV groups were significantly lower than that of the control at 14 days of age. Thus, both Dex treatment and IBV infection inhibited chicken weight gain. To confirm whether Dex affected IBV replication *in vivo*, we assessed the effect of Dex on IBV replication, on the basis of chicken mortality and IBV viral load in kidney tissue (**Figures 1A, B**). The IBV viral loads in the IBV-P-Dex group and the IBV-A-Dex group were higher than that in the IBV group. Dex promoted the replication of IBV in chicken kidneys but did not cause a significant difference in mortality. In addition, we used pathological slides to assess whether Dex affected IBV pathogenicity. The trachea, lung, kidney, heart, liver and spleen tissues were subjected to histopathological observation with hematoxylin and eosin staining. All chickens infected with IBV-QX strains showed different degrees of pathological changes in the trachea, lung, kidneys and spleen, but not the heart and liver tissues (**Figure 1C**). No clear pathological changes were found in the control and Dex groups. Histological examination revealed substantial damage to the tracheal mucosal cortex: the mucosal cells were necrotic and shedding, and a small amount of inflammatory cells had infiltrated. Many mass hemorrhages were observed in the bronchial cavities and pulmonary chambers of the lungs, filling the cavities. Clear renal interstitial hyperemia was observed, and the renal tubular epithelial cells were swollen and degenerated. The splenic sinus was dilated and congested or bleeding in the red pulp area of the spleen, and a small amount of heterophilic granulocyte infiltration was observed. These results indicated that the IBV-QX strain successfully infected chickens and showed strong tropism for kidney and tracheal tissue, in agreement with findings from previous studies (Bouwman et al., 2019; Laconi et al., 2020).

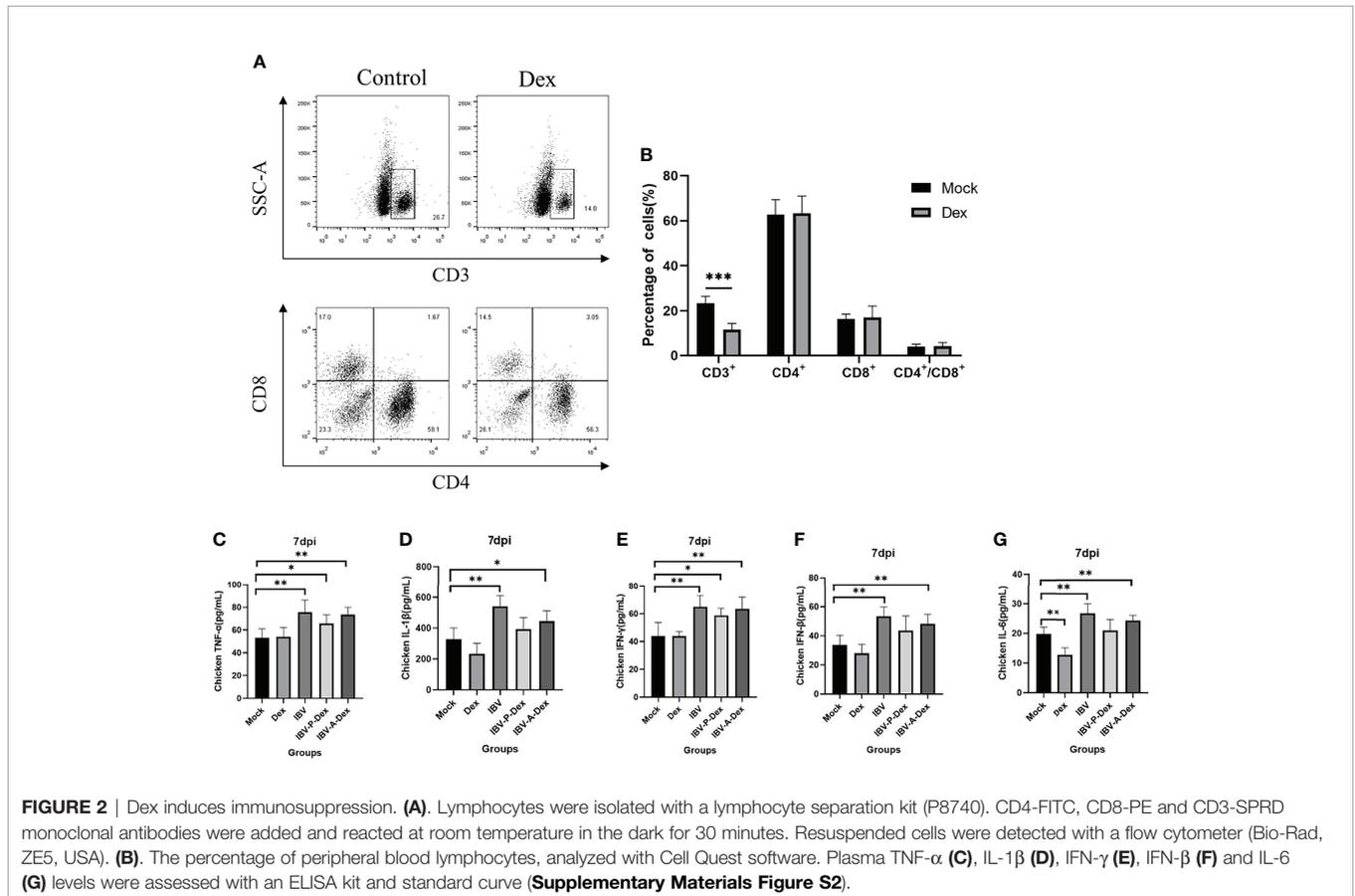
### Dex Induces Immunosuppression

Chicken stress can lead to immunosuppression and metabolic alterations, which can consequently increase susceptibility to infectious diseases (Wang et al., 2020; Kikusato et al., 2021). Therefore, we detected the percentage of CD3<sup>+</sup> T cell subsets and



cytokines in the peripheral blood to determine the immune status of the chickens. As shown in **Figures 2A, B**, the percentage of CD3+ T cell subsets in the Dex group was significantly lower than that in the control. ELISA indicated that the levels of the serum cytokines TNF- $\alpha$ , IL-1 $\beta$ , IFN- $\gamma$ , IFN-

$\beta$  and IL-6 in chickens were significantly higher at 7 dpi after infection with IBV than in the control (**Figures 2C-G**). In addition, with Dex intervention, the cytokine levels in the IBV-P-Dex and IBV-A-Dex groups were lower than those in the IBV group (**Figures 2C-G**). These results revealed that Dex

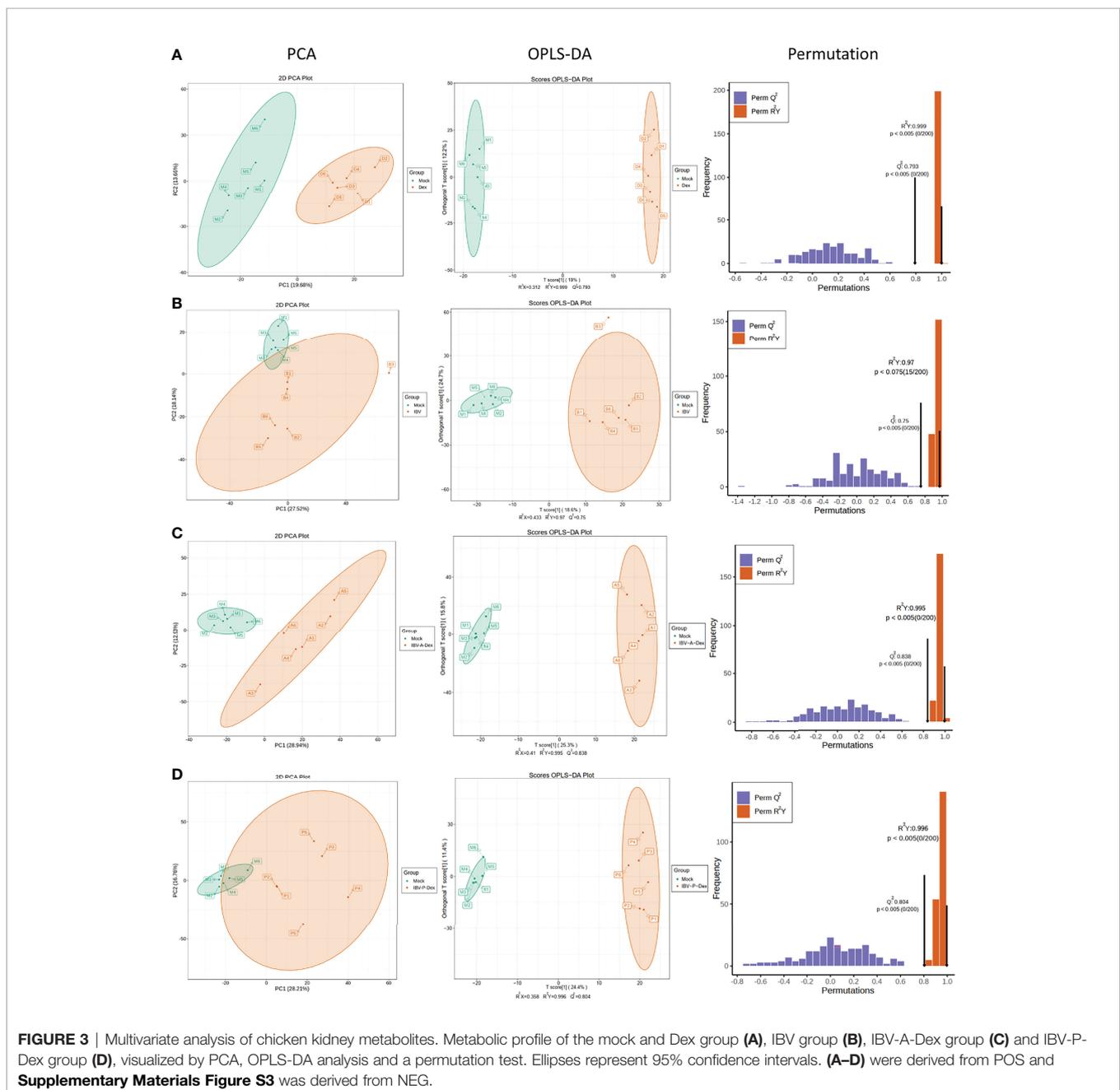


causes immunosuppression in chickens; however, whether immunosuppression is the main reason for the promotion of IBV replication remains unclear.

## Multivariate Analysis of Chicken Kidney Metabolites

To investigate the metabolic pathway changes associated with Dex treatment and IBV infection, we used an LC-MS/MS-based metabolomics method to examine differentially enriched metabolites in the chicken kidneys. Metabolite identification information was obtained by searching laboratory-built

databases, and integrating public libraries and met-DNA methods. To comprehensively collect reliable information on the mock and other groups (mock vs Dex, mock vs IBV, mock vs IBV-A-Dex, and mock vs IBV-P-Dex), we used the principle component analysis (PCA) to compare the metabolite composition (Figures 3A–D). OPLS-DA models were used to determine whether Dex affected metabolic patterns, and a permutation test was further applied to validate the accuracy and predictive ability of the OPLS-DA model (Figures 3A–D). The results of PCA and OPLS-DA analysis showed a clear separation between the content of the control and other groups, thus indicating that significant changes in the



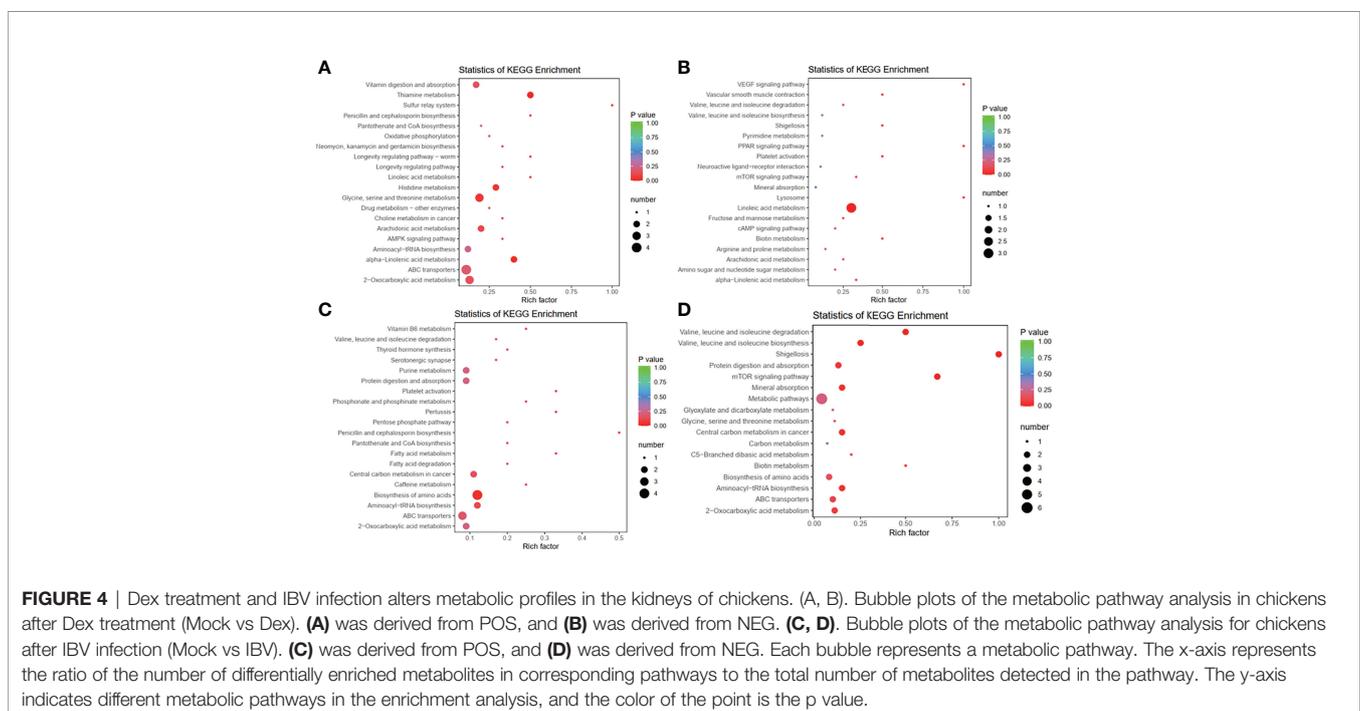
concentrations of metabolites in the kidneys were induced by Dex and IBV infection, which could be used for subsequent studies.

## Lipid Synthesis Is Regulated by Dex and IBV Infection in Chicken Kidneys

The differentially enriched metabolites between the control and Dex groups were key to explaining the occurrence of stress in chickens induced by Dex. The combination of fold change (fold change  $\geq 2$  and fold change  $\leq 0.5$ ), P value ( $<0.05$ ) and VIP ( $\geq 1$ ) in the OPLS-DA model was used to screen for differentially enriched metabolites, and a total of 147 differentially enriched metabolites were obtained in the Dex group. The lists of differentially enriched metabolites are shown in **Supplementary Materials Table S3** (POS) and **Table S4** (NEG), and were visualized *via* a volcano plot (**Supplementary Materials Figures S4A, B**) and hierarchical clustering (**Supplementary Materials Figures S5A, B**). A total of 113 renal metabolites (74 up-regulated, 39 down-regulated) derived from POS significantly changed after Dex treatment, most of which were involved in lipid metabolism pathways. Approximately 39.19% (29/74) of the 74 up-regulated metabolites were lipids and lipid-like molecules, according to the class I identification. According to the class II identification, fatty acyls (13/74), glycerophospholipids (6/74), carboxylic acids and derivatives (6/74), organonitrogen compounds (6/74), and steroids and steroid derivatives (5/76) were the top five metabolites. Of the 39 down-regulated metabolites, 43.59% (17/39) were lipids and lipid-like molecules, of which glycerophospholipids (7/39), glycerolipids (3/39), prenol lipids (3/39), and steroids and steroid derivatives (3/39) were the top three metabolites, according to class II (**Supplementary Materials Table S3**). In summary, Dex may

be involved in the regulation of host lipid metabolism, affecting the biosynthesis of fatty acyls, glycerolipids, prenol lipids, steroids and glycerophospholipids. In addition, the KEGG enrichment analysis results revealed that the differentially enriched metabolites participated in nine target pathways including vitamin digestion and absorption; linoleic acid and alpha-linolenic acid metabolism; thiamine, histidine and arachidonic acid metabolism; and glycine, serine and threonine metabolism (**Figures 4A, B**). These pathways are associated with amino acid metabolism and fatty acid metabolism, which are involved in protein and lipid synthesis, respectively.

We used PCA analysis, OPLS-DA model analysis, permutation tests (**Figure 3B** and **Figure S3B**) and KEGG pathway enrichment analysis (**Figures 4C, D**) to evaluate the changes in kidney metabolites in chicks after IBV infection. The lists of differentially enriched metabolites are shown in **Supplementary Materials Table S5** (POS) and **S6** (NEG), and are visualized *via* a volcano plot (**Supplementary Materials Figures S4C, D**) and hierarchical clustering (**Supplementary Materials Figures S5C, D**). The results showed that IBV infection caused a decrease in the levels of PE-NMe (14:0/20:1 (11Z)), eicosapentaenoyl PAF C-16, PA (20:0/a-15:0), PA (20:0/a-17:0), PC (18:1(11Z)/18:2(9Z,12Z)) and PA (21:0/14:0), which are involved in the biosynthesis of biofilms and bile. However, approximately 48% (24/50) of the 50 up-regulated metabolites were lipids and lipid-like molecules, according to the class I identification. An increase was observed in the levels of many nucleotides and derivatives, glycerophospholipids, and steroids and steroid derivatives, which are associated with cholesterol metabolism and nucleotide synthesis. The changes in these metabolites caused by IBV infection were found to contribute to viral replication.



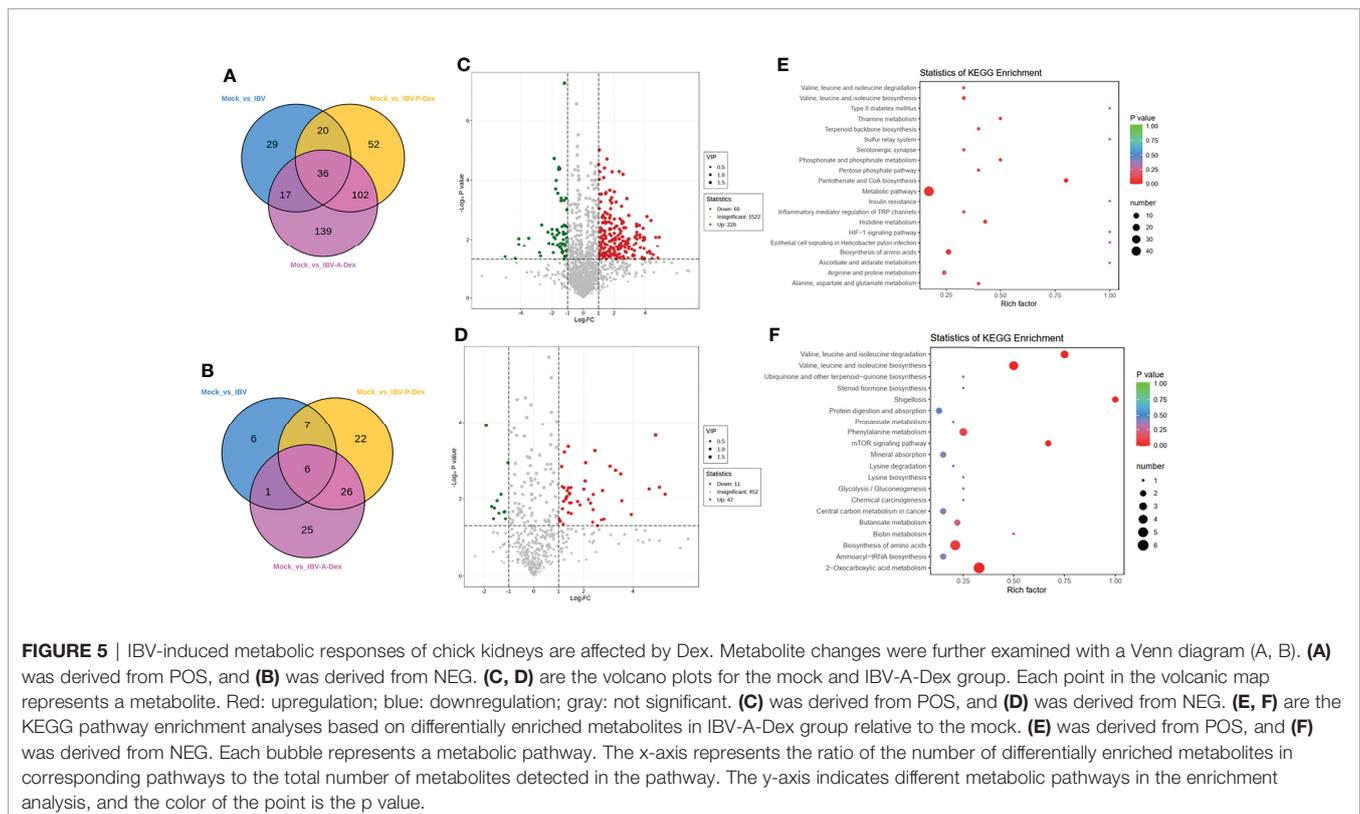
## IBV-Induced Metabolic Responses of Chicken Kidneys Are Affected by Dex

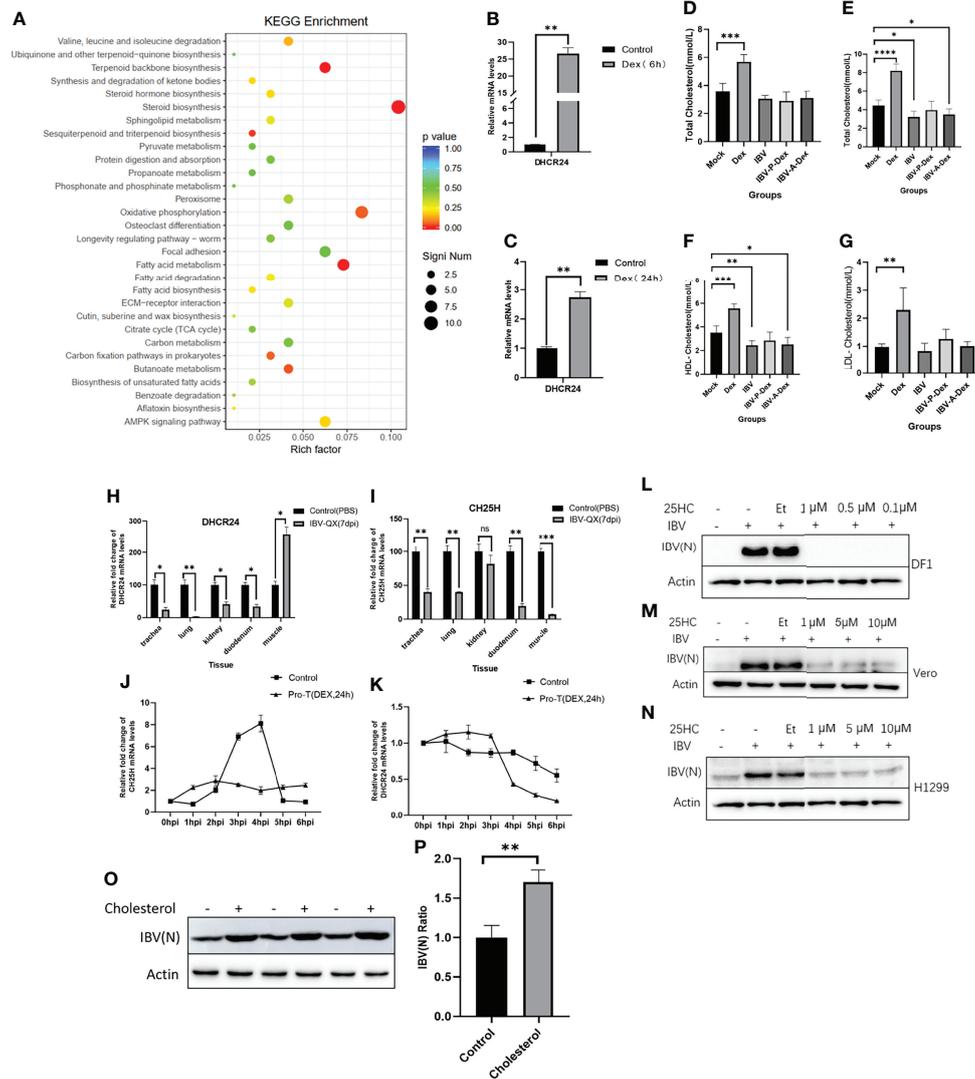
To confirm whether the IBV-induced metabolic responses of chicken kidneys were affected by Dex, we compared the metabolic profiles of the IBV group, IBV-P-Dex group and IBV-A-Dex group with the mock group. PCA and OPLS-DA analysis showed a clear separation between the mock and IBV-A-Dex or IBV-P-Dex groups (Figures 3C, D, Supplementary Materials Figures S3C, D), thus indicating that Dex induced significant changes in the concentrations of metabolites in the kidney responses to IBV infection. The global metabolite changes in terms of their similarity and uniqueness among the three groups were further examined through a Venn diagram (Figures 5A, B), in which 36 metabolites were common to the three groups (Figure 5A). The observed number of metabolites associated with IBV infection in the IBV-A-Dex group was approximately three-fold greater than that in the IBV-P-Dex group, and 24 metabolites were common to both groups (Supplementary Materials Figure S6A). These results indicated metabolic differences between Dex treatment before versus after IBV infection. The lists of differentially enriched metabolites are shown in Supplementary Materials Tables S7–S10, and the volcano plots are shown in Figures 5C, D and Supplementary Materials Figures S6C, D. These significantly differentially enriched metabolites included fatty acyls, amino acid derivatives, hormones and hormone related compounds, carboxylic acids and derivatives, small peptides, and steroids and steroid derivatives, according to the class II identification. In addition, the KEGG

enrichment analysis results are shown in Figures 5E, F and Supplementary Materials Figures S6E, F. The analysis revealed that the differentially enriched metabolites were involved in multiple metabolic pathways, including phenylalanine metabolism, 2-Oxocarboxylic acid metabolism, biosynthesis of amino acids, and synthesis and degradation of valine, leucine and isoleucine. These alterations in host metabolism are likely to be the underlying factors responsible for differences in the level of IBV replication in the kidneys.

## Correlation Between Cholesterol Metabolism and IBV Replication Under Stress

To evaluate the correlation between IBV replication and stress induced by Dex, we performed RNA-seq and untargeted metabolomics to obtain gene expression profiles from DF-1 cells and metabolic profiles from the chicken kidney, respectively. The RNA-seq data showed that Dex-driven changes in DF-1 cells were significantly enriched in cholesterol biosynthetic pathways. As shown in Figure 6A, Dex treatment drove the overexpression of numerous genes associated with steroid biosynthesis, terpenoid backbone biosynthesis and fatty acid metabolism. The mRNA level of DHCR24, a key rate-limiting enzyme for cholesterol biosynthesis, was significantly up-regulated in DF-1 cells after Dex treatment for 6 h or 24 h (Figures 6B, C). To verify whether Dex increased cholesterol levels in chickens, we used an automatic biochemical analyzer to measure plasma TC, HDL-C and LDL-C





**FIGURE 6** | Host cholesterol metabolism is affected by Dex treatment and IBV infection **(A)** KEGG analyses based on differentially expressed genes in DF-1 cells pretreated with Dex (10  $\mu\text{g}/\text{mL}$ ) for 24 h relative to controls. Circles indicate numbers of genes, and colors depict the enrichment. **(B, C)** qPCR detection of mRNA levels of DHCR24 in DF-1 cells treated with Dex (10  $\mu\text{g}/\text{mL}$ ) for 6 or 24 h, and the Dex-untreated group was used as a control. The gene expression was quantified relative to actin expression with the  $2^{-\Delta\Delta\text{CT}}$  method according to our previous research (Liu et al., 2018). **(D)** An automatic biochemical analyzer was used to detect the plasma TC level of chickens at 4 dpi (11 days old). Plasma TC **(E)**, HDL-C **(F)** and LDL-C **(G)** levels of chickens were detected with the automatic biochemical analyzer at 7 dpi (14 days of age). Transcript levels of DHCR24 **(H)** and CH25H **(I)** in chick kidney tissues were detected by qPCR at 7 dpi as previously described (Liu et al., 2018). Transcript levels of CH25H **(J)** and DHCR24 **(K)** in DF-1 cells were detected at the indicated time points after infection with 1 MOI of IBV. The cells were pretreated with Dex for 24 h before infection, and the Dex-untreated group was used as a control. L (DF-1), M (Vero) and N (H1299) were pretreated with 25HC at the indicated concentrations for 12 h, and the cells were then infected with IBV-Beaudette at an MOI of 1. Cells were collected at 12 hpi and subjected to western blot analysis. Cytotoxicity assays of 25HC in cells were based on previous studies (Zhang et al., 2019; Xie et al., 2019). **(O)** DF-1 cells were pretreated with cholesterol (10  $\mu\text{M}$ ) for 12 h, and cells were then infected with the IBV-Beaudette strain at an MOI of 1. Cells were collected at 12 hpi and subjected to western blot analysis; cholesterol-untreated cells were used as a control. **(P)** The signal of protein bands was determined in Image J software.

levels. Dex significantly up-regulated plasma TC, HDL-C and LDL-C levels in chickens (**Figures 6D–G**). The data also indicated that the chickens infected with IBV at 7 dpi, compared with those in the mock group, showed significantly lower plasma TC and HDL-C levels (**Figures 6E, F**). In addition, the transcript levels of DHCR24 and CH25H, two key rate-limiting enzymes in cholesterol metabolism, were significantly down-regulated in chickens at 7

dpi (**Figures 6H, I**). Simultaneously, when DF-1 cells were invaded by IBV, the mRNA levels of CH25H and CHCR24 also showed significant differences according to Dex pretreatment (**Figures 6J, K**). As shown in **Figures 6L–N**, pretreatment with 25HC for 12 h significantly inhibited IBV replication, particularly in DF-1 cells. Moreover, the addition of exogenous cholesterol in DF-1 cells promoted the replication of IBV (**Figures 6O, P**). Thus, Dex

treatment and IBV infection affected host cholesterol metabolism, and intrinsic link may exist between host cholesterol biosynthesis pathways and IBV infection.

## DISCUSSION

Metabolic changes are the end-result of adaptive and defensive biochemical reactions that occur during infection (Kuang et al., 2020). In recent years, metabolomics has been widely used in chicken research to understand various host responses during viral infection (Liu et al., 2019; Xu et al., 2019). Examination of the metabolomes of the kidneys (Xu et al., 2019), bursa of fabricius (Kuang et al., 2020), lungs (Liu et al., 2019), plasma (Chen et al., 2021) and chicken cell lines (DF-1 and LMH) (Lin et al., 2020; Xu et al., 2022) has provided a new method for evaluating the regulation of metabolic pathways in virus-host interactions involving viruses such as IBV, NDV, ALV, IBDV and ILTV (Liu et al., 2019; Xu et al., 2019; Kuang et al., 2020; Lin et al., 2020; Chen et al., 2021; Xu et al., 2022). IBV is a pathogenic chicken coronavirus, which is a highly infectious in domestic chickens of all ages and types, and affects the respiratory, renal and reproductive systems (Amarasinghe et al., 2017; de Wit and Cook, 2019). Although considerable attention has been paid to coronaviruses, the metabolic regulatory mechanisms underlying chicken responses to IBV infection under stress conditions remain unclear. In the present study, we established a poultry stress model in 7-day-old chickens injected with Dex to elucidate the effects of stress on metabolic changes and IBV replication in chicken kidneys. We used an LC-MS/MS-based metabolomics method to examine differentially enriched metabolites in the kidneys and assessed the effects of Dex on chicken body weight, T cell subsets, serum cholesterol levels, cytokines and viral load. This work provides new insights into the interaction between the IBV and host metabolism under stress.

This study further revealed the changes in metabolites and connected pathways in chickens after IBV infection or treatment by Dex. Particular attention was paid to whether host metabolism manipulated by Dex affected the infective response to IBV in chickens. The results of PCA, OPLS-DA and hierarchical clustering revealed significant differences in the global metabolite profiles of controls and chickens infected with the IBV-QX strain or treated with Dex (Figures 3A, B and Supplementary Figure S5). Compared with the control, Dex-treatment showed a significantly differences in amino acids, steroid biosynthesis, fatty acid metabolism, butanoate metabolism, thiamine metabolism, histidine metabolism, arachidonic acid metabolism and linoleic acid metabolism (Figures 4A, B). In summary, the observed changes in these metabolites increase understanding of the intracellular reactions through which Dex affects IBV infection by manipulating host metabolic pathways. Previous studies have reported that GCs are important regulators of lipid metabolism, and chronic exposure to GCs promotes lipogenesis (John et al., 2016; Hu et al., 2018). In addition, cholesterol is a critical GC regulatory serum component (Yang et al., 2014). GCs impair HDL-mediated cholesterol efflux beyond increasing HDL cholesterol

concentrations (Bouillet et al., 2020). This study supports previous observations in rats and chickens indicating that Dex treatment causes lipid accumulation (Cai et al., 2009; Wang et al., 2010; Cai et al., 2011; Wang et al., 2012), growth inhibition (Pan et al., 2019), changes in serum lipids (Malkawi et al., 2018) and hypercholesterolemia (Li et al., 2020), thus suggesting modulation of lipid metabolism disorders in chickens (Wang et al., 2010; Wang et al., 2012; Wang et al., 2012; Lv et al., 2018; Hu et al., 2020).

Our findings are broadly consistent with those of other comprehensive studies of human coronaviruses. A strong relationship between cholesterol and coronavirus replication has been widely documented in the literature (Thorp and Gallagher, 2004; Syed et al., 2010; Heaton and Randall, 2011; Blanc et al., 2013; Del Campo and Romero-Gómez, 2015; Guo et al., 2017; Meher et al., 2019; Lange et al., 2019; Cao et al., 2020; Radenkovic et al., 2020; Balgoma et al., 2020; Luquain-Costaz et al., 2020; Sanders et al., 2021; Cheng et al., 2021). Our previous study has also shown that cholesterol has an important role in coronavirus entry, membrane fusion and pathological syncytia formation; therefore cholesterol metabolic mechanisms may be promising drug targets for coronavirus infections (Dai et al., 2022). Herein, we obtained the gene expression profiles of DF-1 cells exposed to Dex for 24 h, and the RNA-seq data indicated that many genes associated with cholesterol biosynthesis were up-regulated by Dex. In view of this finding, we examined serum cholesterol levels in Dex-treated chickens and found that Dex treatment significantly upregulated serum cholesterol concentrations, including TC, HDL-C and LDL-C (Figures 6D–G). Previous research has shown that viral infections may cause host cells to alter the expression of cholesterol metabolizing enzymes and metabolites; similarly, cholesterol metabolism can also regulate host antiviral responses (Osuna-Ramos et al., 2018; Xiao et al., 2020). This notion was also supported by our data demonstrating that IBV infection significantly downregulated CH25H and DHCR24 transcript levels in chicks (Figures 6H, I). To support viral entry and increase virion production, IBV must consume host cholesterol. Similar results were also confirmed in chickens (Figures 6E, F). Moreover, the addition of exogenous cholesterol to DF-1 cells promoted IBV replication, but whether Dex promotion of IBV replication levels *in vivo* is associated with cholesterol biosynthesis remains to be further verified in subsequent experiments. In addition, most of the renal differentially enriched metabolites induced by Dex were involved in lipid metabolism pathways— notably cholesterol metabolism—which are very important for the replication of coronavirus IBV. The promotion of IBV replication in chicken kidney tissue may be associated with the up-regulation of cholesterol levels induced by Dex. The identification of these differentially enriched metabolites and metabolic pathways may lead to the development of drugs to combat coronavirus infections, thus further underscoring the importance of cholesterol in coronavirus infection. Therefore, weaponizing host cholesterol metabolism dysregulation against coronavirus infections may serve as an effective antiviral strategy (Sturley et al., 2020; Proto et al., 2021). Understanding the relationship between cholesterol biosynthesis and coronavirus infection will be an important direction for future research (Daniloski et al., 2021).

Dex has been investigated before in animal models and small clinical trials for infections with different coronaviruses, but the results have been mixed. Differences in timing and dose underlie many of the inconsistent and sometimes conflicting results across studies of GC therapy (Cain and Cidlowski, 2020). GCs are often used in combination with antiviral drugs to counteract painful inflammation and are known to inhibit the replication of some viruses (Lancz et al., 1990; Kim et al., 2017). However, this approach is controversial, because GC treatment has been suggested to increase the viral yield and susceptibility, thereby increasing lung lesions and increasing or prolonging shedding of viruses, such as SARS-CoV (Li et al., 2003), SIVs (Ali et al., 2013), HSV-1 (Erlandsson et al., 2002; Hara et al., 2009; Du et al., 2012; Pechan et al., 2014), MMTV (Parks et al., 1974; Indik et al., 2007), retroviruses (Solodushko et al., 2009), PRRSV-1 (Singleton et al., 2018) and FFV (Lee and Shin, 2018). Consequently, GC use is often hampered by the onset of adverse effects or resistance (Petta et al., 2016).

In summary, the metabolome profiles of chickens under stress induced by Dex and infected with the IBV-QX strain were analyzed to establish the characteristics by LC-MS/MS. The identification of these differentially enriched metabolites and metabolic pathways revealed that Dex treatment targets the cholesterol biosynthesis pathway in chicks and DF-1 cells. Host responses to IBV infection are also regulated by Dex. In addition, chicken growth and immune function are significantly inhibited by Dex. Although this study has some limitations, such as the use of a single tissue type and single time point for metabolite detection, it nonetheless provides a comprehensive analysis of host metabolic profile changes that occur during stress, and supplies new information for exploring the molecular regulatory mechanisms of avian stress. This article provides a comprehensive and in-depth understanding of poultry stress and should serve as a basis for further research to clarify the interaction between the virus and the host under stress.

## DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**. Further inquiries can be directed to the corresponding authors.

## ETHICS STATEMENT

The animal study was reviewed and approved by Shanghai Veterinary Research Institute, Chinese Academy of Agricultural Sciences.

## REFERENCES

- Aguilera, G. (1994). Regulation of Pituitary ACTH Secretion During Chronic Stress. *Front. Neuroendocrinol.* 15 (4), 321–350. doi: 10.1006/frne.1994.1013
- Alba, A. C., Strauch, T. A., Keisler, D. H., Wells, K. D., and Kesler, D. C. (2019). Using a Keratinase to Degrade Chicken Feathers for Improved Extraction of Glucocorticoids. *Gen. Comp. Endocrinol.* 270, 35–40. doi: 10.1016/j.ygcen.2018.10.002

## AUTHOR CONTRIBUTIONS

JD and XQ designed the experiments, analyzed the data, and wrote the manuscript. JD and HW performed the experiments. XQ, TL, CD, WL, LT, CS, and YS gave suggestions during the experiments. JD, XQ and CD revised the manuscript. All authors read and approved the final manuscript.

## FUNDING

This work was supported by the Shanghai Agriculture Applied Technology Development Program, China (G20180207), National Natural Science Foundation of China (grant No. 32030108), Shanghai Agriculture Applied Technology Development Program, China (2022-02-08-00-12-F01154) and Natural Science Foundation of Shanghai (grant Nos. 21ZR1476800 and 20ZR1469400).

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fcimb.2022.945865/full#supplementary-material>

**Supplementary Table S1** | Specific R package from this article.

**Supplementary Table S2** | Changes in body weight in chickens

**Supplementary Table S3** | The lists of differentially enriched metabolites in mock vs Dex (POS)

**Supplementary Table S4** | The lists of differentially enriched metabolites in mock vs Dex (NEG)

**Supplementary Table S5** | The lists of differentially enriched metabolites in mock vs IBV (POS)

**Supplementary Table S6** | The lists of differentially enriched metabolites in mock vs IBV (NEG)

**Supplementary Table S7** | The lists of differentially enriched metabolites in mock vs IBV-A-Dex (POS)

**Supplementary Table S8** | The lists of differentially enriched metabolites in mock vs IBV-A-Dex (NEG)

**Supplementary Table S9** | The lists of differentially enriched metabolites in mock vs IBV-P-Dex (POS)

**Supplementary Table S10** | The lists of differentially enriched metabolites in mock vs IBV-P-Dex (NEG)

Ali, A., Ibrahim, M., Eladl, A. H., Saif, Y. M., and Lee, C.-W. (2013). Enhanced Replication of Swine Influenza Viruses in Dexamethasone-Treated Juvenile and Layer Turkeys. *Vet. Microbiol.* 162 (2-4), 353–359. doi: 10.1016/j.vetmic.2012.10.007

Al-Zoubi, N., Kachrimanis, K., Younis, K., and Malamataris, S. (2011). Optimization of Extended-Release Hydrophilic Matrix Tablets by Support Vector Regression. *Drug Dev. Ind. Pharm.* 37 (1), 80–87. doi: 10.3109/03639045.2010.492396

- Amarasinghe, A., Abdul-Cader, M. S., Nazir, S., De Silva Senapathi, U., van der Meer, F., Cork, S. C., et al. (2017). Infectious Bronchitis Corona Virus Establishes Productive Infection in Avian Macrophages Interfering With Selected Antimicrobial Functions. *PLoS One* 12 (8), e0181801. doi: 10.1371/journal.pone.0181801
- Austdal, L. P. E., Bjørnstad, S., Mathisen, G. H., Aden, P. K., Mikkola, I., Paulsen, R. E., et al. (2016). Glucocorticoid Effects on Cerebellar Development in a Chicken Embryo Model: Exploring Changes in PAX6 and Metalloproteinase-9 After Exposure to Dexamethasone. *J. Neuroendocrinol.* 28 (12), 1–16. doi: 10.1111/jne.12438
- Balgoma, D., Gil-de-Gómez, L., and Montero, O. (2020). Lipidomics Issues on Human Positive ssRNA Virus Infection: An Update. *Metabolites* 10 (9), 356. doi: 10.3390/metabo10090356
- Beckford, R. C., Ellestad, L. E., Proszkowiec-Weglarz, M., Farley, L., Brady, K., Angel, R., et al. (2020). Effects of Heat Stress on Performance, Blood Chemistry, and Hypothalamic and Pituitary mRNA Expression in Broiler Chickens. *Poult. Sci.* 99 (12), 6317–6325. doi: 10.1016/j.psj.2020.09.052
- Benson, B. N., Calvert, C. C., Roura, E., and Klasing, K. C. (1993). Dietary Energy Source and Density Modulate the Expression of Immunologic Stress in Chicks. *J. Nutr.* 123 (10), 1714–1723. doi: 10.1093/jn/123.10.1714
- Blahová, J., Dobšíková, R., Straková, E., and Suchý, P. (2007). Effect of Low Environmental Temperature on Performance and Blood System in Broiler Chickens (*Gallus Domesticus*). *Acta. Veterinaria Brno.* 76 (8), S17–S23. doi: 10.2754/avb200776S8S017
- Blanc, M., Hsieh, W. Y., Robertson, K. A., Kropp, K. A., Forster, T., Shui, G., et al. (2013). The Transcription Factor STAT-1 Couples Macrophage Synthesis of 25-Hydroxycholesterol to the Interferon Antiviral Response. *Immunity* 38 (1), 106–118. doi: 10.1016/j.immuni.2012.11.004
- Borsoi, A., Quinteiro-Filho, W. M., Calefi, A. S., Ferreira, A. J. P., Astolfi-Ferreira, C. S., Florio, J. C., et al. (2015). Effects of Cold Stress and Salmonella Heidelberg Infection on Bacterial Load and Immunity of Chickens. *Avian Pathol.* 44 (6), 490–497. doi: 10.1080/03079457.2015.1086976
- Bouillet, B., Gautier, T., Denimal, D., Samson, M., Masson, D., Pais de Barros, J. P., et al. (2020). Glucocorticoids Impair HDL-Mediated Cholesterol Efflux Besides Increased HDL Cholesterol Concentration: A Proof of Concept. *Eur. J. Endocrinol.* 183 (3), 297–306. doi: 10.1530/EJE-20-0477
- Bouwman, K. M., Parsons, L. M., Berends, A. J., Vries, R., and Verheije, M. H. (2019). Three Amino Acid Changes in Avian Coronavirus Spike Protein Allow Binding to Kidney Tissue. *J. Virol.* 94 (2), e01363–e01319. doi: 10.1128/JVI.01363-19
- Buwalda, B., Scholte, J., de Boer, S. F., Coppens, C. M., and Koolhaas, J. M. (2012). The Acute Glucocorticoid Stress Response Does Not Differentiate Between Rewarding and Aversive Social Stimuli in Rats. *Horm. Behav.* 61 (2), 218–226. doi: 10.1016/j.yhbeh.2011.12.012
- Cain, D. W., and Cidlowski, J. A. (2020). After 62 Years of Regulating Immunity, Dexamethasone Meets COVID-19. *Nat. Rev. Immunol.* 20 (10), 587–588. doi: 10.1038/s41577-020-00421-x
- Cai, Y., Song, Z., Wang, X., Jiao, H., and Lin, H. (2011). Dexamethasone-Induced Hepatic Lipogenesis is Insulin Dependent in Chickens (*Gallus Gallus Domesticus*). *Stress* 14 (3), 273–281. doi: 10.3109/10253890.2010.543444
- Cai, Y., Song, Z., Zhang, X., Wang, X., Jiao, H., and Lin, H. (2009). Increased *De Novo* Lipogenesis in Liver Contributes to the Augmented Fat Deposition in Dexamethasone Exposed Broiler Chickens (*Gallus Gallus Domesticus*). *Comp. Biochem. Physiol. C. Toxicol. Pharmacol.* 150 (2), 164–169. doi: 10.1016/j.cbpc.2009.04.005
- Cao, X., Yin, R., Albrecht, H., Fan, D., and Tan, W. (2020). Cholesterol: A New Game Player Accelerating Vasculopathy Caused by SARS-CoV-2? *Am. J. Physiol. Endocrinol. Metab.* 319 (1), E197–E202. doi: 10.1152/ajpendo.00255.2020
- Carsia, R. V., Weber, H., and Lauterio, T. J. (1988). Protein Malnutrition in the Domestic Fowl Induces Alterations in Adrenocortical Function. *Endocrinology* 122 (2), 673–680. doi: 10.1210/endo-122-2-673
- Castellani, J. W., and Young, A. J. (2016). Human Physiological Responses to Cold Exposure: Acute Responses and Acclimatization to Prolonged Exposure. *Autonomic Neurosci.* 2016, 63–74. doi: 10.1016/j.autneu.2016.02.009
- Chang, W. H., Li, J. J., Zhang, S., Zheng, A. J., Yuan, J. L., Cai, H. Y., et al. (2015). Effects of Glucocorticoid-Induced Stress on Absorption of Glycylsarcosine in Jejunum of Broilers. *Poult. Sci.* 94 (4), 700–705. doi: 10.3382/ps/pev041
- Cheng, K., Riva, L., Sinha, S., Pal, L. R., Nair, N. U., Martin-Sancho, L., et al. (2021). Genome-Scale Metabolic Modeling Reveals SARS-CoV-2-Induced Host Metabolic Reprogramming and Identifies Metabolic Antiviral Targets. *bioRxiv*. doi: 10.1101/2021.01.27.428543
- Chen, Y., Li, H.-W., Cong, F., and Lian, Y.-X. (2021). Metabolomics Profiling for Identification of Potential Biomarkers in Chickens Infected With Avian Leukosis Virus Subgroup J (ALV-J). *Res. Vet. Sci.* 139, 166–171. doi: 10.1016/j.rvsc.2021.07.026
- Chen, S., Yong, Y., and Ju, X. (2021). Effect of Heat Stress on Growth and Production Performance of Livestock and Poultry: Mechanism to Prevention. *J. Therm. Biol.* 99, 103019. doi: 10.1016/j.jtherbio.2021.103019
- Chrousos, G. P. (2009). Stress and Disorders of the Stress System. *Nat. Rev. Endocrinol.* 5 (7), 374–381. doi: 10.1038/nrendo.2009.106
- Chrousos, G. P., and Gold, P. W. (1992). The Concepts of Stress and Stress System Disorders. Overview of Physical and Behavioral Homeostasis. *JAMA* 267 (9), 1244–1252. doi: 10.1001/jama.1992.03480090092034
- Chrousos, G. P., and Kino, T. (2007). Glucocorticoid Action Networks and Complex Psychiatric and/or Somatic Disorders. *Stress* 10 (2), 213–219. doi: 10.1080/10253890701292119
- Collier, R. J., and Gebremedhin, K. G. (2015). Thermal Biology of Domestic Animals. *Annu. Rev. Anim. Biosci.* 3, 513–532. doi: 10.1146/annurev-animal-022114-110659
- Dai, J., Wang, H., Liao, Y., Tan, L., Sun, Y., Song, C., et al. (2022). Coronavirus Infection and Cholesterol Metabolism. *Front. Immunol.* 13. doi: 10.3389/fimmu.2022.791267
- Danilowski, Z., Jordan, T. X., Wessels, H.-H., Hoagland, D. A., Kasela, S., Legut, M., et al. (2021). Identification of Required Host Factors for SARS-CoV-2 Infection in Human Cells. *Cell* 184 (1), 92–105.e16. doi: 10.1016/j.cell.2020.10.030
- Del Campo, J. A., and Romero-Gómez, M. (2015). Modulation of Host Lipid Metabolism by Hepatitis C Virus: Role of New Therapies. *World J. Gastroenterol.* 21 (38), 10776–10782. doi: 10.3748/wjg.v21.i38.10776
- Deviche, P., Bittner, S., Gao, S., and Valle, S. (2017). Roles and Mechanistic Bases of Glucocorticoid Regulation of Avian Reproduction. *Integr. Comp. Biol.* 57 (6), 1184–1193. doi: 10.1093/icb/ixc112
- de Wit, J. J. S., and Cook, J. K. A. (2019). Spotlight on Avian Pathology: Infectious Bronchitis Virus. *Avian Pathol.* 48 (5), 393–395. doi: 10.1080/03079457.2019.1617400
- Donaldson, G. C. (2006). Climate Change and the End of the Respiratory Syncytial Virus Season. *Clin Infect Dis.* 42 (5), 677–679. doi: 10.1086/500208
- Du, T., Zhou, G., and Roizman, B. (2012). Induction of Apoptosis Accelerates Reactivation of Latent HSV-1 in Ganglionic Organ Cultures and Replication in Cell Cultures. *Proc. Natl. Acad. Sci. U.S.A.* 109 (36), 14616–14621. doi: 10.1073/pnas.1212661109
- Eid, Y., Ebeid, T., and Younis, H. (2006). Vitamin E Supplementation Reduces Dexamethasone-Induced Oxidative Stress in Chicken Semen. *Br. Poult. Sci.* 47 (3), 350–356. doi: 10.1080/00071660600753912
- El-Senousey, H. K., Chen, B., Wang, J. Y., Atta, A. M., Mohamed, F. R., and Nie, Q. H. (2018). Effects of Dietary Vitamin C, Vitamin E, and Alpha-Lipoic Acid Supplementation on the Antioxidant Defense System and Immune-Related Gene Expression in Broilers Exposed to Oxidative Stress by Dexamethasone. *Poult. Sci.* 97 (1), 30–38. doi: 10.3382/ps/pex298
- Erlandsson, A. C., Bladh, L. G., Stierna, P., Yucel-Lindberg, T., Hammarsten, O., Modéer, T., et al. (2002). Herpes Simplex Virus Type 1 Infection and Glucocorticoid Treatment Regulate Viral Yield, Glucocorticoid Receptor and NF-kappaB Levels. *J. Endocrinol.* 175 (1), 165–176. doi: 10.1677/joe.0.1750165
- Flinchum, G., and Gwen, B. (2015). *Sturkie's Avian Physiology* (6th edition). *Am. J. Veterinary Res.* 76 (4), 291–292.
- Foxman, E. F., Storer, J. A., Fitzgerald, M. E., Wasik, B. R., Hou, L., Zhao, H., et al. (2015). Temperature-Dependent Innate Defense Against the Common Cold Virus Limits Viral Replication at Warm Temperature in Mouse Airway Cells. *Proc. Natl. Acad. Sci. U.S.A.* 112 (3), 827–832. doi: 10.1073/pnas.1411030112
- Gao, B., Gong, X., Fang, S., Weng, W., Wang, H., Chu, H., et al. (2021). Inhibition of Anti-Viral Stress Granule Formation by Coronavirus Endoribonuclease Nsp15 Ensures Efficient Virus Replication. *PLoS Pathog.* 17 (2), e1008690. doi: 10.1371/journal.ppat.1008690
- Gao, J., Lin, H., Song, Z. G., and Jiao, H. C. (2008). Corticosterone Alters Meat Quality by Changing Pre- and Postslaughter Muscle Metabolism. *Poult. Sci.* 87 (8), 1609–1617. doi: 10.3382/ps.2007-00007

- Gao, J., Lin, H., Wang, X. J., Song, Z. G., and Jiao, H. C. (2010). Vitamin E Supplementation Alleviates the Oxidative Stress Induced by Dexamethasone Treatment and Improves Meat Quality in Broiler Chickens. *Poult. Sci.* 89 (2), 318–327. doi: 10.3382/ps.2009-00216
- Garriga, C., Hunter, R. R., Amat, C., Planas, J. M., Mitchell, M. A., and Moretó, M. (2006). Heat Stress Increases Apical Glucose Transport in the Chicken Jejunum. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 290 (1), R195–R201. doi: 10.1152/ajpregu.00393.2005
- Goel, A., Ncho, C. M., and Choi, Y. H. (2021). Regulation of Gene Expression in Chickens by Heat Stress. *J. Anim. Sci. Biotechnol.* 12 (1), 11. doi: 10.1186/s40104-020-00523-5
- Gong, Y., Tang, N., Liu, P., Sun, Y., Lu, S., Liu, W., et al. (2021). Newcastle Disease Virus Degrades SIRT3 via PINK1-PRKN-Dependent Mitophagy to Reprogram Energy Metabolism in Infected Cells. *Autophagy* 1–19. doi: 10.1080/15548627.2021.1990515
- Guo, H., Huang, M., Yuan, Q., Wei, Y., Gao, Y., Mao, L., et al. (2017). The Important Role of Lipid Raft-Mediated Attachment in the Infection of Cultured Cells by Coronavirus Infectious Bronchitis Virus Beaudette Strain. *PLoS One* 12 (1), 513. doi: 10.1371/journal.pone.0170123
- Guo, Y., Jiang, R., Su, A., Tian, H., Zhang, Y., Li, W., et al. (2020). Identification of Genes Related to Effects of Stress on Immune Function in the Spleen in a Chicken Stress Model Using Transcriptome Analysis. *Mol. Immunol.* 124, 180–189. doi: 10.1016/j.molimm.2020.06.004
- Guo, Y., Su, A., Tian, H., Ding, M., Wang, Y., Tian, Y., et al. (2021). TMT-Based Quantitative Proteomic Analysis Reveals the Spleen Regulatory Network of Dexamethasone-Induced Immune Suppression in Chicks. *J. Proteomics* 248, 104353. doi: 10.1016/j.jprot.2021.104353
- Guo, Y., Su, A., Tian, H., Zhai, M., Li, W., Tian, Y., et al. (2020). Transcriptomic Analysis of Spleen Revealed Mechanism of Dexamethasone-Induced Immune Suppression in Chicks. *Genes (Basel)* 11 (5). doi: 10.3390/genes11050513
- Häffelin, K. E., Lindenwald, R., Kaufmann, F., Döhrring, S., Spindler, B., Preisinger, R., et al. (2020). Corticosterone in Feathers of Laying Hens: An Assay Validation for Evidence-Based Assessment of Animal Welfare. *Poult. Sci.* 99 (10), 4685–4694. doi: 10.1016/j.psj.2020.06.065
- Hara, Y., Shiraiishi, A., Kobayashi, T., Kadota, Y., Shirakata, Y., Hashimoto, K., et al. (2009). Alteration of TLR3 Pathways by Glucocorticoids may be Responsible for Immunosusceptibility of Human Corneal Epithelial Cells to Viral Infections. *Mol. Vis.* 15, 937–948. doi: 10.1016/j.visres.2008.04.032
- Heaton, N. S., and Randall, G. (2011). Multifaceted Roles for Lipids in Viral Infection. *Trends Microbiol.* 19 (7), 368–375. doi: 10.1016/j.tim.2011.03.007
- Herman, J. P., McKlveen, J. M., Ghosal, S., Kopp, B., Wulsin, A., Makinson, R., et al. (2016). Regulation of the Hypothalamic-Pituitary-Adrenocortical Stress Response. *Compr. Physiol.* 6 (2), 603–621. doi: 10.1002/cphy.c150015
- Hu, J. Y., and Cheng, H. W. (2021). Warm Perches: A Novel Approach for Reducing Cold Stress Effect on Production, Plasma Hormones, and Immunity in Laying Hens. *Poult. Sci.* 100 (8), 101294. doi: 10.1016/j.psj.2021.101294
- Hu, Y., Feng, Y., Zhang, L., Jia, Y., Cai, D., Qian, S.-B., et al. (2020). GR-Mediated FTO Transactivation Induces Lipid Accumulation in Hepatocytes via Demethylation of mA on Lipogenic mRNAs. *RNA Biol.* 17 (7), 930–942. doi: 10.1080/15476286.2020.1736868
- Hu, X., Li, X., Xiao, C., Kong, L., Zhu, Q., and Song, Z. (2021). Effects of Dietary Energy Level on Performance, Plasma Parameters, and Central AMPK Levels in Stressed Broilers. *Front. Vet. Sci.* 8, 681858. doi: 10.3389/fvets.2021.681858
- Hu, Y., Sun, Q., Hu, Y., Hou, Z., Zong, Y., Omer, N. A., et al. (2018). Corticosterone-Induced Lipogenesis Activation and Lipophagy Inhibition in Chicken Liver Are Alleviated by Maternal Betaine Supplementation. *J. Nutr.* 148 (3), 316–325. doi: 10.1093/jn/nxx073
- Indik, S., Günzburg, W. H., Kulich, P., Salmons, B., and Rouault, F. (2007). Rapid Spread of Mouse Mammary Tumor Virus in Cultured Human Breast Cells. *Retrovirology* 4, 73. doi: 10.1186/1742-4690-4-73
- Iyasere, O. S., Beard, A. P., Guy, J. H., and Bateson, M. (2017). Elevated Levels of the Stress Hormone, Corticosterone, Cause 'Pessimistic' Judgment Bias in Broiler Chickens. *Sci. Rep.* 7 (1), 6860. doi: 10.1038/s41598-017-07040-y
- Jiao, H., Zhou, K., Zhao, J., Wang, X., and Lin, H. (2018). A High-Calorie Diet Rich in Soy Oil Alleviates Oxidative Damage of Skeletal Muscles Induced by Dexamethasone in Chickens. *Redox Rep.* 23 (1), 68–82. doi: 10.1080/13510002.2017.1405494
- John, K., Marino, J. S., Sanchez, E. R., and Hinds, T. D. (2016). The Glucocorticoid Receptor: Cause of or Cure for Obesity? *Am. J. Physiol. Endocrinol. Metab.* 310 (4), E249–E257. doi: 10.1152/ajpendo.00478.2015
- Kallicharan, R., and Hall, B. K. (1974). A Developmental Study of the Levels of Progesterone, Corticosterone, Cortisol, and Cortisone Circulating in Plasma of Chick Embryos. *Gen. Comp. Endocrinol.* 24 (4), 364–372. doi: 10.1016/0016-6480(74)90149-X
- Kikusato, M., Xue, G., Pastor, A., Niewold, T. A., and Toyomizu, M. (2021). Effects of Plant-Derived Isoquinoline Alkaloids on Growth Performance and Intestinal Function of Broiler Chickens Under Heat Stress. *Poult. Sci.* 100 (2), 957–963. doi: 10.1016/j.psj.2020.11.050
- Kim, M. S., Lee, S. J., Choi, S. H., Kang, Y. J., and Kim, K. H. (2017). Dexamethasone Treatment Decreases Replication of Viral Hemorrhagic Septicemia Virus in Epithelioma Papulosum Cyprini Cells. *Arch. Virol.* 162 (5), 1387–1392. doi: 10.1007/s00705-017-3248-x
- Kong, F., Xu, Y., Ran, W., Yin, B., Feng, L., and Sun, D. (2020). Cold Exposure-Induced Up-Regulation of Hsp70 Positively Regulates PEDV mRNA Synthesis and Protein Expression In Vitro. *Pathogens* 9 (4), 246. doi: 10.3390/pathogens9040246
- Kuang, J., Xu, P., Shi, Y., Yang, Y., Liu, P., Chen, S., et al. (2020). Nephropathogenic Infectious Bronchitis Virus Infection Altered the Metabolome Profile and Immune Function of the Bursa of Fabricius in Chicken. *Front. Vet. Sci.* 7, 628270. doi: 10.3389/fvets.2020.628270
- Kumar, M., Ratwan, P., Dahiya, S. P., and Nehra, A. K. (2021). Climate Change and Heat Stress: Impact on Production, Reproduction and Growth Performance of Poultry and its Mitigation Using Genetic Strategies. *J. Therm. Biol.* 97, 102867. doi: 10.1016/j.jtherbio.2021.102867
- Lacóni, A., Weerts, E. A. W. S., Bloodgood, J. C. G., Deniz Marrero, J. P., Berends, A. J., Coccio, G., et al. (2020). Attenuated Live Infectious Bronchitis Virus QX Vaccine Disseminates Slowly to Target Organs Distant From the Site of Inoculation. *Vaccine* 38 (6), 1486–1493. doi: 10.1016/j.vaccine.2019.11.064
- Lancz, G., Whitaker-Dowling, P., Marsh, Y. V., Bradley, G., Eppstein, D. A., Hackney, J. F., et al. (1990). Inhibition of Vesicular Stomatitis Virus Replication in Dexamethasone-Treated L929 Cells. *Proc. Soc. Exp. Biol. Med.* 193 (3), 190–196. doi: 10.3181/00379727-193-43024
- Lange, P. T., Lagunoff, M., and Tarakanova, V. L. (2019). Chewing the Fat: The Conserved Ability of DNA Viruses to Hijack Cellular Lipid Metabolism. *Viruses* 11 (2), 119. doi: 10.3390/v11020119
- Lan, R., Li, Y., Chang, Q., and Zhao, Z. (2020). Dietary Chitosan Oligosaccharides Alleviate Heat Stress-Induced Intestinal Oxidative Stress and Inflammatory Response in Yellow-Feather Broilers. *Poult. Sci.* 99 (12), 6745–6752. doi: 10.1016/j.psj.2020.09.050
- Lara, L. J., and Rostagno, M. H. (2013). Impact of Heat Stress on Poultry Production. *Anim. (Basel)* 3 (2), 356–369. doi: 10.3390/ani3020356
- Lee, G.-E., and Shin, C.-G. (2018). Influence of Pretreatment With Immunosuppressive Drugs on Viral Proliferation. *J. Microbiol. Biotechnol.* 28 (10), 1716–1722. doi: 10.4014/jmb.1807.06054
- Liao, Y., Wang, X., Huang, M., Tam, J. P., and Liu, D. X. (2011). Regulation of the P38 Mitogen-Activated Protein Kinase and Dual-Specificity Phosphatase 1 Feedback Loop Modulates the Induction of Interleukin 6 and 8 in Cells Infected With Coronavirus Infectious Bronchitis Virus. *Virology* 420 (2), 106–116. doi: 10.1016/j.virol.2011.09.003
- Li, L., Hu, W., Liu, K., Zhang, D., Liu, M., Li, X., et al. (2020). miR-148a/LDLR Mediates Hypercholesterolemia Induced by Prenatal Dexamethasone Exposure in Male Offspring Rats. *Toxicol. Appl. Pharmacol.* 395, 114979. doi: 10.1016/j.taap.2020.114979
- Li, X.-w., Jiang, R.-m., and Guo, J.-z. (2003). [Glucocorticoid in the Treatment of Severe Acute Respiratory Syndrome Patients: A Preliminary Report]. *Zhonghua Nei. Ke. Za. Zhi.* 42 (6), 378–381.
- Lin, H., Decuyper, E., and Buyse, J. (2004). Oxidative Stress Induced by Corticosterone Administration in Broiler Chickens (*Gallus Gallus Domesticus*) 1. Chronic Exposure. *Comp. Biochem. Physiol. B. Biochem. Mol. Biol.* 139 (4), 737–744. doi: 10.1016/j.cbpc.2004.09.013
- Lin, H., Jiao, H. C., Buyse, J., and Decuyper, E. (2006). Strategies for Preventing Heat Stress in Poultry. *World's Poultry Sci. J.* 62 (1), 71–85. doi: 10.1079/WPS200585
- Lin, J., Yi, X., and Zhuang, Y. (2020). Coupling Metabolomics Analysis and DOE Optimization Strategy Towards Enhanced IBDV Production by Chicken Embryo Fibroblast DF-1 Cells. *J. Biotechnol.* 307, 114–124. doi: 10.1016/j.jbiotec.2019.10.018
- Liu, L., Liu, X., Cui, H., Liu, R., Zhao, G., and Wen, J. (2019). Transcriptional Insights Into Key Genes and Pathways Controlling Muscle Lipid Metabolism in Broiler Chickens. *BMC Genomics* 20 (1), 863. doi: 10.1186/s12864-019-6221-0

- Liu, W., Qiu, X., Song, C., Sun, Y., Meng, C., Liao, Y., et al. (2018). Deep Sequencing-Based Transcriptome Profiling Reveals Avian Interferon-Stimulated Genes and Provides Comprehensive Insight Into Newcastle Disease Virus-Induced Host Responses. *Viruses* 10 (4), 162. doi: 10.3390/v10040162
- Liu, P., Yin, Y., Gong, Y., Qiu, X., Sun, Y., Tan, L., et al. (2019). *In Vitro* and *In Vivo* Metabolomic Profiling After Infection With Virulent Newcastle Disease Virus. *Viruses* 11 (10), 962. doi: 10.3390/v11100962
- Liu, Y., Yu, M., Cui, J., Du, Y., Teng, X., and Zhang, Z. (2021). Heat Shock Proteins Took Part in Oxidative Stress-Mediated Inflammatory Injury via NF- $\kappa$ B Pathway in Excess Manganese-Treated Chicken Livers. *Ecotoxicol. Environ. Saf.* 226, 112833. doi: 10.1016/j.ecoenv.2021.112833
- Luquin-Costaz, C., Rabia, M., Hullin-Matsuda, F., and Delton, I. (2020). Bis (monoacylglycerol)phosphate, an Important Actor in the Host Endocytic Machinery Hijacked by SARS-CoV-2 and Related Viruses. *Biochimie* 179, 247–256. doi: 10.1016/j.biochi.2020.10.018
- Lv, Z. P., Peng, Y. Z., Zhang, B. B., Fan, H., Liu, D., and Guo, Y. M. (2018). Glucose and Lipid Metabolism Disorders in the Chickens With Dexamethasone-Induced Oxidative Stress. *J. Anim. Physiol. Anim. Nutr. (Berl)* 102 (2), e706–e717. doi: 10.1111/jpn.12823
- Maatjens, C. M., Reijrink, I. A. M., Molenaar, R., van der Pol, C. W., Kemp, B., and van den Brand, H. (2014). Temperature and CO<sub>2</sub> During the Hatching Phase. I. Effects on Chick Quality and Organ Development. *Poult. Sci.* 93 (3), 645–654. doi: 10.3382/ps.2013-03491
- Malkawi, A. K., Alzoubi, K. H., Jacob, M., Matic, G., Ali, A., Al Faraj, A., et al. (2018). Metabolomics Based Profiling of Dexamethasone Side Effects in Rats. *Front. Pharmacol.* 9, 46. doi: 10.3389/fphar.2018.00046
- McEwen, B. S. (2000). Allostasis and Allostatic Load: Implications for Neuropsychopharmacology. *Neuropsychopharmacology* 22 (2), 108–124. doi: 10.1016/S0893-133X(99)00129-3
- Meher, G., Bhattacharjya, S., and Chakraborty, H. (2019). Membrane Cholesterol Modulates Oligomeric Status and Peptide-Membrane Interaction of Severe Acute Respiratory Syndrome Coronavirus Fusion Peptide. *J. Phys. Chem. B.* 123 (50), 10654–10662. doi: 10.1021/acs.jpcc.9b08455
- Mesquita, M. A., Araújo, I. C. S., Café, M. B., Arnhold, E., Mascarenhas, A. G., Carvalho, F. B., et al. (2021). Results of Hatching and Rearing Broiler Chickens in Different Incubation Systems. *Poult. Sci.* 100 (1), 94–102. doi: 10.1016/j.psj.2020.09.028
- Myers, B., McKlveen, J. M., and Herman, J. P. (2012). Neural Regulation of the Stress Response: The Many Faces of Feedback. *Cell Mol. Neurobiol* 1, 10.1007/s10571-012-9801-y. doi: 10.1007/s10571-012-9801-y
- Nawab, A., Ibtisham, F., Li, G., Kieser, B., Wu, J., Liu, W., et al. (2018). Heat Stress in Poultry Production: Mitigation Strategies to Overcome the Future Challenges Facing the Global Poultry Industry. *J. Therm. Biol.* 78, 131–139. doi: 10.1016/j.jtherbio.2018.08.010
- Nicolaides, N. C., Kyrtzi, E., Lamprokostopoulou, A., Chrousos, G. P., and Charmandari, E. (2015). Stress, the Stress System and the Role of Glucocorticoids. *Neuroimmunomodulation* 22 (1–2), 6–19. doi: 10.1159/000362736
- Noyola, D. E., and Mandeville, P. B. (2008). Effect of Climatological Factors on Respiratory Syncytial Virus Epidemics. *Epidemiol. Infect.* 136 (10), 1328–1332. doi: 10.1017/S0950268807000143
- Osho, S. O., and Adeola, O. (2020). Chitosan Oligosaccharide Supplementation Alleviates Stress Stimulated by in-Feed Dexamethasone in Broiler Chickens. *Poult. Sci.* 99 (4), 2061–2067. doi: 10.1016/j.psj.2019.11.047
- Osuna-Ramos, J. F., Reyes-Ruiz, J. M., and Del Ángel, R. M. (2018). The Role of Host Cholesterol During Flavivirus Infection. *Front. Cell Infect. Microbiol.* 8, 388. doi: 10.3389/fcimb.2018.00388
- Pan, L., Ma, X. K., Zhao, P. F., and Piao, X. S. (2019). Weeping Forsythia Extract Alleviates Dexamethasone-Induced Oxidative Injury of Breast Muscles in Broilers. *Animal* 13 (11), 2660–2668. doi: 10.1017/S175173111900096X
- Parks, W. P., Scolnick, E. M., and Kozikowski, E. H. (1974). Dexamethasone Stimulation of Murine Mammary Tumor Virus Expression: A Tissue Culture Source of Virus. *Science* 184 (4133), 158–160. doi: 10.1126/science.184.4133.158
- Pechan, P., Ardinger, J., Ketavarapu, J., Rubin, H., Wadsworth, S. C., and Scaria, A. (2014). Aurintricarboxylic Acid Increases Yield of HSV-1 Vectors. *Mol. Ther. Methods Clin. Dev.* 1, 6. doi: 10.1038/mtm.2013.6
- Petta, I., Dejager, L., Ballegeer, M., Lievens, S., Tavernier, J., De Bosscher, K., et al. (2016). The Interactome of the Glucocorticoid Receptor and Its Influence on the Actions of Glucocorticoids in Combatting Inflammatory and Infectious Diseases. *Microbiol. Mol. Biol. Rev.* 80 (2), 495–522. doi: 10.1128/MMBR.00064-15
- Pfitzer, S., Verwoerd, D. J., Gerdes, G. H., Labuschagne, A. E., Erasmus, A., Manvell, R. J., et al. (2000). Newcastle Disease and Avian Influenza A Virus in Wild Waterfowl in South Africa. *Avian Dis.* 44 (3), 655–660. doi: 10.2307/1593107
- Proto, M. C., Fiore, D., Piscopo, C., Pagano, C., Galgani, M., Bruzzaniti, S., et al. (2021). Lipid Homeostasis and Mevalonate Pathway in COVID-19: Basic Concepts and Potential Therapeutic Targets. *Prog. Lipid Res.* 82, 101099. doi: 10.1016/j.plipres.2021.101099
- Quinteiro-Filho, W. M., Calefi, A. S., Cruz, D. S. G., Aloia, T. P. A., Zager, A., Astolfi-Ferreira, C. S., et al. (2017). Heat Stress Decreases Expression of the Cytokines, Avian  $\beta$ -Defensins 4 and 6 and Toll-Like Receptor 2 in Broiler Chickens Infected With *Salmonella* Enteritidis. *Vet. Immunol. Immunopathol.* 186, 19–28. doi: 10.1016/j.vetimm.2017.02.006
- Radenkovic, D., Chawla, S., Pirro, M., Sahebkar, A., and Banach, M. (2020). Cholesterol in Relation to COVID-19: Should We Care About it? *J. Clin. Med.* 9 (6), 1909. doi: 10.3390/jcm9061909
- Ragheb, M., Sadeghi, A. A., and Aminafshar, M. (2016). Energy Sources and Levels Influenced on Performance Parameters, Thyroid Hormones, and HSP70 Gene Expression of Broiler Chickens Under Heat Stress. *Trop. Anim. Health Prod.* 48 (8), 1697–1702. doi: 10.1007/s11250-016-1146-y
- Revollo, J. R., and Cidlowski, J. A. (2009). Mechanisms Generating Diversity in Glucocorticoid Receptor Signaling. *Ann. N. Y. Acad. Sci.* 1179, 167–178. doi: 10.1111/j.1749-6632.2009.04986.x
- Sánchez-Illana, Á., Pérez-Guaita, D., Cuesta-García, D., Sanjuan-Herráez, J. D., Vento, M., Ruiz-Cerdá, J. L., et al. (2018). Model Selection for Within-Batch Effect Correction in UPLC-MS Metabolomics Using Quality Control - Support Vector Regression. *Anal. Chim. Acta* 1026, 62–68. doi: 10.1016/j.aca.2018.04.055
- Sanders, D. W., Jumper, C. C., Ackerman, P. J., Bracha, D., Donick, A., Kim, H., et al. (2021). *Et Al*: SARS-CoV-2 Requires Cholesterol for Viral Entry and Pathological Syncytia Formation. *Elife* 10, e65962. doi: 10.7554/eLife.65962.sa2
- Sapolsky, R. M., Romero, L. M., and Munck, A. U. (2000). How do Glucocorticoids Influence Stress Responses? Integrating Permissive, Suppressive, Stimulatory, and Preparative Actions. *Endocr. Rev.* 21 (1), 55–89. doi: 10.1210/edrv.21.1.0389
- Scanes, C. G. (2016). Biology of Stress in Poultry With Emphasis on Glucocorticoids and the Heterophil to Lymphocyte Ratio. *Poult. Sci.* 95 (9), 2208–2215. doi: 10.3382/ps/pew137
- Scheff, J. D., Calvano, S. E., Lowry, S. F., and Androulakis, I. P. (2012). Transcriptional Implications of Ultradian Glucocorticoid Secretion in Homeostasis and in the Acute Stress Response. *Physiol. Genomics* 44 (2), 121–129. doi: 10.1152/physiolgenomics.00128.2011
- Singleton, H., Graham, S. P., Frossard, J.-P., Bodman-Smith, K. B., and Steinbach, F. (2018). Infection of Monocytes With European Porcine Reproductive and Respiratory Syndrome Virus (PRRSV-1) Strain Lena is Significantly Enhanced by Dexamethasone and IL-10. *Virology* 517, 199–207. doi: 10.1016/j.virol.2018.02.017
- Solodushko, V., Bitko, V., and Fouty, B. (2009). Dexamethasone and Mifepristone Increase Retroviral Infectivity Through Different Mechanisms. *Am. J. Physiol. Lung Cell Mol. Physiol.* 297 (3), L538–L545. doi: 10.1152/ajplung.00162.2009
- Steel, J., Staeheli, P., Mubareka, S., García-Sastre, A., Palese, P., and Lowen, A. C. (2010). Transmission of Pandemic H1N1 Influenza Virus and Impact of Prior Exposure to Seasonal Strains or Interferon Treatment. *J. Virol.* 84 (1), 21–26. doi: 10.1128/JVI.01732-09
- Sturley, S. L., Rajakumar, T., Hammond, N., Higaki, K., Márka, Z., Márka, S., et al. (2020). Potential COVID-19 Therapeutics From a Rare Disease: Weaponizing Lipid Dysregulation to Combat Viral Infectivity. *J. Lipid Res.* 61 (7), 972–982. doi: 10.1194/jlr.R120000851
- Su, A., Guo, Y., Tian, H., Zhou, Y., Li, W., Tian, Y., et al. (2021). Analysis of miRNA and mRNA Reveals Core Interaction Networks and Pathways of Dexamethasone-Induced Immunosuppression in Chicken Bursa of Fabricius. *Mol. Immunol.* 134, 34–47. doi: 10.1016/j.molimm.2021.02.022
- Sun, F., Zuo, Y. Z., Ge, J., Xia, J., Li, X. N., Lin, J., et al. (2018). Transport Stress Induces Heart Damage in Newly Hatched Chicks via Blocking the Cytoprotective Heat Shock Response and Augmenting Nitric Oxide Production. *Poult. Sci.* 97 (8), 2638–2646. doi: 10.3382/ps/pey146

- Syed, G. H., Amako, Y., and Siddiqui, A. (2010). Hepatitis C Virus Hijacks Host Lipid Metabolism. *Trends Endocrinol. Metab.* 21 (1), 33–40. doi: 10.1016/j.tem.2009.07.005
- Tang, J. W. (2009). The Effect of Environmental Parameters on the Survival of Airborne Infectious Agents. *J. R. Soc. Interface* 6 Suppl 6, S737–S746. doi: 10.1098/rsif.2009.0227.focus
- Thorpe, E. B., and Gallagher, T. M. (2004). Requirements for CEACAMs and Cholesterol During Murine Coronavirus Cell Entry. *J. Virol.* 78 (6), 2682–2692. doi: 10.1128/JVI.78.6.2682-2692.2004
- Tsiouris, V., Georgopoulou, I., Batzios, C., Pappaioannou, N., Ducatelle, R., and Fortomaris, P. (2015). The Effect of Cold Stress on the Pathogenesis of Necrotic Enteritis in Broiler Chicks. *Avian. Pathol.* 44 (6), 430–435. doi: 10.1080/03079457.2015.1083094
- van de Ven, L. J. F., Baller, L., van Wagenberg, A. V., Kemp, B., and van den Brand, H. (2011). Effects of Egg Position During Late Incubation on Hatching Parameters and Chick Quality. *Poult. Sci.* 90 (10), 2342–2347. doi: 10.3382/ps.2011-01467
- Vapnik, V., Golowich, S. E., and Smola, A. (2008). Support Vector Method for Function Approximation, Regression Estimation, and Signal Processing. *Adv. Neural Inf. Process. Syst.* 9, 281–287. <http://dx.doi.org/>
- Wang, X., Lin, H., Song, Z., and Jiao, H. (2010). Dexamethasone Facilitates Lipid Accumulation and Mild Feed Restriction Improves Fatty Acids Oxidation in Skeletal Muscle of Broiler Chicks (*Gallus Gallus Domesticus*). *Comp. Biochem. Physiol. C. Toxicol. Pharmacol.* 151 (4), 447–454. doi: 10.1016/j.cbpc.2010.01.010
- Wang, H., Liu, D., Sun, Y., Meng, C., Tan, L., Song, C., et al. (2021). Upregulation of DUSP6 Impairs Infectious Bronchitis Virus Replication by Negatively Regulating ERK Pathway and Promoting Apoptosis. *Vet. Res.* 52 (1), 7. doi: 10.1186/s13567-020-00866-x
- Wang, Y., Saelao, P., Chanthavixay, K., Gallardo, R., Bunn, D., Lamont, S. J., et al. (2018). Physiological Responses to Heat Stress in Two Genetically Distinct Chicken Inbred Lines. *Poult. Sci.* 97 (3), 770–780. doi: 10.3382/ps/pex363
- Wang, Y., Saelao, P., Kern, C., Jin, S., Gallardo, R. A., Kelly, T., et al. (2020). Liver Transcriptome Responses to Heat Stress and Newcastle Disease Virus Infection in Genetically Distinct Chicken Inbred Lines. *Genes (Basel)* 11 (9), 1067. doi: 10.3390/genes11091067
- Wang, X. J., Song, Z. G., Jiao, H. C., and Lin, H. (2012). Dexamethasone Facilitates Lipid Accumulation in Chicken Skeletal Muscle. *Stress* 15 (4), 443–456. doi: 10.3109/10253890.2011.639413
- Wang, X. J., Wei, D. L., Song, Z. G., Jiao, H. C., and Lin, H. (2012). Effects of Fatty Acid Treatments on the Dexamethasone-Induced Intramuscular Lipid Accumulation in Chickens. *PLoS One* 7 (5), e36663. doi: 10.1371/journal.pone.0036663
- Wang, H., Yuan, X., Sun, Y., Mao, X., Meng, C., Tan, L., et al. (2019). Infectious Bronchitis Virus Entry Mainly Depends on Clathrin Mediated Endocytosis and Requires Classical Endosomal/Lysosomal System. *Virology* 528, 118–136. doi: 10.1016/j.virol.2018.12.012
- Webster Marketon, J. I., and Glaser, R. (2008). Stress Hormones and Immune Function. *Cell Immunol.* 252 (1-2), 16–26. doi: 10.1016/j.cellimm.2007.09.006
- Wein, Y., Geva, Z., Bar-Shira, E., and Friedman, A. (2017). Transport-Related Stress and its Resolution in Turkey Pullets: Activation of a Pro-Inflammatory Response in Peripheral Blood Leukocytes. *Poult. Sci.* 96 (8), 2601–2613. doi: 10.3382/ps/pex076
- Wei, H., Zhang, R., Su, Y., Bi, Y., Li, X., Zhang, X., et al. (2018). Effects of Acute Cold Stress After Long-Term Cold Stimulation on Antioxidant Status, Heat Shock Proteins, Inflammation and Immune Cytokines in Broiler Heart. *Front. Physiol.* 9, 1589. doi: 10.3389/fphys.2018.01589
- Xiao, J., Li, W., Zheng, X., Qi, L., Wang, H., Zhang, C., et al. (2020). Targeting 7-Dehydrocholesterol Reductase Integrates Cholesterol Metabolism and IRF3 Activation to Eliminate Infection. *Immunity* 52 (1), 109–122.e6. doi: 10.1016/j.immuni.2019.11.015
- Xie, T., Feng, M., Dai, M., Mo, G., Ruan, Z., Wang, G., et al. (2019). Cholesterol-25-Hydroxylase Is a Chicken ISG That Restricts ALV-J Infection by Producing 25-Hydroxycholesterol. *Viruses* 11 (6), 498. doi: 10.3390/v11060498
- Xu, P., Liu, P., Zhou, C., Shi, Y., Wu, Q., Yang, Y., et al. (2019). A Multi-Omics Study of Chicken Infected by Nephropathogenic Infectious Bronchitis Virus. *Viruses* 11 (11), 1070. doi: 10.3390/v11111070
- Xu, L., Wang, Z., Chen, Z., Cui, L., Liu, Z., Liang, Y., et al. (2022). PFT- $\alpha$  Inhibits Gallid Alpha Herpesvirus 1 Replication by Repressing Host Nucleotide Metabolism and ATP Synthesis. *Vet. Microbiol.* 269, 109435. doi: 10.1016/j.vetmic.2022.109435
- Yang, N., Caratti, G., Ince, L. M., Poolman, T. M., Trebble, P. J., Holt, C. M., et al. (2014). Serum Cholesterol Selectively Regulates Glucocorticoid Sensitivity Through Activation of JNK. *J. Endocrinol.* 223 (2), 155–166. doi: 10.1530/JOE-14-0456
- Yang, X., Luo, Y. H., Zeng, Q. F., Zhang, K. Y., and Wang, J. P. (2014). Effects of Low Ambient Temperatures and Dietary Vitamin C Supplement on Growth Performance, Blood Parameters, and Antioxidant Capacity of 21-Day-Old Broilers. *Poult. Sci.* 93 (4), 898–905. doi: 10.3382/ps.2013-03438
- Zaboli, G., Huang, X., Feng, X., and Ahn, D. U. (2019). How can Heat Stress Affect Chicken Meat Quality? - a Review. *Poult. Sci.* 98 (3), 1551–1556. doi: 10.3382/ps/pey399
- Zhai, M., Guo, Y., Su, A., Tian, H., Sun, G., Kang, X., et al. (2020). Identification of Genes Related to Dexamethasone-Induced Immunosuppression in Chicken Thymus Using Transcriptome Analysis. *Res. Vet. Sci.* 132, 318–327. doi: 10.1016/j.rvsc.2020.07.002
- Zhang, Z.-W., Bi, M.-Y., Yao, H.-D., Fu, J., Li, S., and Xu, S.-W. (2014). Effect of Cold Stress on Expression of AMPK $\alpha$ -PPAR $\alpha$  Pathway and Inflammation Genes. *Avian Dis.* 58 (3), 415–426. doi: 10.1637/10763-010814-Reg.1
- Zhang, Z. W., Lv, Z. H., Li, J. L., Li, S., Xu, S. W., and Wang, X. L. (2011). Effects of Cold Stress on Nitric Oxide in Duodenum of Chicks. *Poult. Sci.* 90 (7), 1555–1561. doi: 10.3382/ps.2010-01333
- Zhang, X. L., Shao, X. J., Wang, J., and Guo, W. L. (2013). Temporal Characteristics of Respiratory Syncytial Virus Infection in Children and its Correlation With Climatic Factors at a Public Pediatric Hospital in Suzhou. *J Clin Virol* 58, 4, 666–670. doi: 10.1016/j.jcv.2013.09.027
- Zhang, Y., Song, Z., Wang, M., Lan, M., Zhang, K., Jiang, P., et al. (2019). Cholesterol 25-Hydroxylase Negatively Regulates Porcine Intestinal Coronavirus Replication by the Production of 25-Hydroxycholesterol. *Vet. Microbiol.* 231, 129–138. doi: 10.1016/j.vetmic.2019.03.004
- Zhao, F.-Q., Zhang, Z.-W., Wang, C., Zhang, B., Yao, H.-D., Li, S., et al. (2013). The Role of Heat Shock Proteins in Inflammatory Injury Induced by Cold Stress in Chicken Hearts. *Cell Stress Chaperones* 18 (6), 773–783. doi: 10.1007/s12192-013-0429-8
- Zheng, H.-T., Zhuang, Z.-X., Chen, C.-J., Liao, H.-Y., Chen, H.-L., Hsueh, H.-C., et al. (2021). Effects of Acute Heat Stress on Protein Expression and Histone Modification in the Adrenal Gland of Male Layer-Type Country Chickens. *Sci. Rep.* 11 (1), 6499. doi: 10.1038/s41598-021-85868-1
- Zhou, H. J., Kong, L. L., Zhu, L. X., Hu, X. Y., Busye, J., and Song, Z. G. (2021). Effects of Cold Stress on Growth Performance, Serum Biochemistry, Intestinal Barrier Molecules, and Adenosine Monophosphate-Activated Protein Kinase in Broilers. *Animal* 15 (3), 100138. doi: 10.1016/j.animal.2020.100138
- Zhou, Y., Tian, W., Zhang, M., Ren, T., Sun, G., Jiang, R., et al. (2019). Transcriptome Analysis Revealed Regulation of Dexamethasone Induced microRNAs in Chicken Thymus. *J. Cell Biochem.* 120 (4), 6570–6579. doi: 10.1002/jcb.27950

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Dai, Wang, Liao, Tan, Sun, Song, Liu, Ding, Luo and Qiu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.