



Editorial for “Regulatory RNAs in the nervous system”

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Until about a decade ago, the non-coding part of the genome was considered without function. The development of high-throughput RNA sequencing techniques (next-generation sequencing) revealed the existence of many transcripts that do not code for proteins in addition to the RNA components needed for mRNA translation: rRNAs and tRNAs. The aim of this issue was to put together reports on the role of non-coding RNAs in the nervous system, an emerging field not covered so far in a systematic manner.

Non-coding transcripts can be divided into three broad classes: (i) short RNAs (sRNAs), (ii) RNAs transcribed from the opposite strand of a protein-coding locus that contain sequences anti-sense with respect to the protein-coding transcript, (OS-RNAs) and (iii) long intergenic non-coding RNAs (lincRNAs). Many of these non-coding RNAs (nc-RNAs) can regulate the transcription or the translation of protein-coding genes. Almost on weekly basis, new findings reveal the regulatory role that nc-RNAs exert in many biological processes. Overall, these studies are making increasingly clear that, both in model organisms and in humans, complexity is not a function of the number of protein-coding genes, but results from the possibility of using combinations of genetic programs and controlling their spatial and temporal regulation during development, senescence and in disease by regulatory RNAs. This has generated a novel picture of gene regulatory networks where regulatory nc-RNAs represent novel layers of regulation. Publications reporting novel non-coding RNAs found using sequencing appears almost monthly, therefore dedicated bioinformatics techniques to analyze the result of this analysis are under development (Guffanti et al., 2014).

Particularly well-characterized is the role of microRNAs (miRNAs) in the post-transcriptional regulation of gene expression. MicroRNAs are short (~21 nt) nc-RNAs that arise from processing of a long primary transcript via a complex and well-described biosynthetic process. MicroRNAs bind to mRNAs (usually in the 3' untranslated region) and regulate gene expression by repressing mRNA translation and/or inducing degradation of the target mRNA. Up to now, several thousands of miRNAs have been predicted and identified in animals, plants and viruses (www.mirbase.org) and some microRNAs are highly conserved, facilitating the analysis of microRNA in non-model species. A feature of miRNAs is their combinatorial regulation: a given miRNA can target a multitude of different mRNAs and a given target might

similarly be targeted by multiple miRNAs; for this reason, they frequently represent the central nodes of several regulatory networks and may act as rheostat to provide stability and fine-tuning to gene expression networks (Osella et al., 2011; Siciliano et al., 2013). MicroRNAs are also relatively easy to study experimentally and novel methods to study their function are continually coming out (Chaudhuri et al., 2013; Knauss et al., 2013). They can be transfected in cells, microinjected in embryos or delivered *in vivo* to neurons and their function can be blocked, *in vitro* and *in vivo*, by modified antisense oligonucleotides (antagomiRs). For all these reasons, the majority of contributions to this e-book relate to miRNAs. In the nervous system, miRNAs have been involved in the regulation of cellular pathways controlling fundamental functions during development (Benchoua and Peschanski, 2013; Coolen et al., 2013; Cremisi, 2013; Hong et al., 2013; Iyengar et al., 2014; Iyer et al., 2014; Terzibasi Tozzini et al., 2014), synaptic plasticity (Tognini and Pizzorusso, 2012; Chiu et al., 2014), and in neurodegenerative disease. Intriguingly, miRNAs show a double-sided relationship with neuronal activity: electrical activity (Eacker et al., 2013; Pai et al., 2014) regulates miRNAs at the level of transcription, biogenesis, stability and specific targeting to dendrites and also axons and presynaptic terminals (Kaplan et al., 2013) on one side, but miRNAs are also able to regulate membrane conductances altering neuronal biophysical properties (Gavazzo et al., 2013). Synaptic localization is particularly relevant in the context of local translational control (Heise et al., 2014), thereby providing a molecular substrate for synaptic plasticity. Deregulation of expression of miRNAs is proposed not only as potential disease biomarker (Sheinerman and Umansky, 2013; Maffioletti et al., 2014), but it has been implicated directly in the pathogenesis of complex neurological and neuropsychiatric disease (Dogini et al., 2013; Goodall et al., 2013; Maciotta et al., 2013; Serafini et al., 2013; Barbato et al., 2014; Della Ragione et al., 2014; Elramah et al., 2014; Fragkouli and Doxakis, 2014; Kye and Goncalves Ido, 2014; Nieto-Diaz et al., 2014). This so-called RNA revolution also led to the exploitation of RNA interference and the development of related tools as potential treatment of a vast array of CNS disease that could benefit from regulation of disease-associated genes.

A second class of small RNAs are the piwi-interacting RNAs (piRNAs). These are slightly larger than miRNAs (24–32 nt) originate from intergenic repetitive sequences that are transcribed as a

long RNA and processed and play an important role in gametogenesis and transposon silencing. PiRNAs are expressed at low level (if at all) in somatic tissues and their role in the nervous system is still ill-characterized.

Long non-coding RNAs are a heterogeneous population and are much less studied (see Ernst and Morton, 2013). They can be associated to chromatin and either interfere with transcription of the target gene(s) or induce epigenetic modifications. Long ncRNAs can indeed interact with chromatin remodellers such as Polycomb and target these to specific genomic regions. Opposite-strand RNAs can hybridize with their protein-coding complementary transcript and modulate splicing or induce RNA degradation. Finally, long ncRNAs derived from pseudogenes can act as competitive inhibitors for miRNAs thereby increasing the expression of their protein-coding paralog. Examples of these mechanisms relate to transcription of repetitive elements (Pascarella et al., 2014) or fine tuning of developmental patterning and positional information in the central nervous system mediated by regulation of the spatial pattern of expression of Hox genes in *Drosophila* (Gummalla et al., 2014).

REFERENCES

- Barbato, C., Pezzola, S., Caggiano, C., Antonelli, M., Frisone, P., Ciotti, M. T., et al. (2014). A lentiviral sponge for miR-101 regulates RanBP9 expression and amyloid precursor protein metabolism in hippocampal neurons. *Front. Cell. Neurosci.* 8:37. doi: 10.3389/fncel.2014.00037
- Benchoua, A., and Peschanski, M. (2013). Pluripotent stem cells as a model to study non-coding RNAs function in human neurogenesis. *Front. Cell. Neurosci.* 7:140. doi: 10.3389/fncel.2013.00140
- Chaudhuri, A. D., Yelamanchili, S. V., and Fox, H. S. (2013). Combined fluorescent *in situ* hybridization for detection of microRNAs and immunofluorescent labeling for cell-type markers. *Front. Cell. Neurosci.* 7:160. doi: 10.3389/fncel.2013.00160
- Chiu, H., Alqadah, A., and Chang, C. (2014). The role of microRNAs in regulating neuronal connectivity. *Front. Cell. Neurosci.* 7:283. doi: 10.3389/fncel.2013.00283
- Coolen, M., Katz, S., and Bally-Cuif, L. (2013). miR-9: a versatile regulator of neurogenesis. *Front. Cell. Neurosci.* 7:220. doi: 10.3389/fncel.2013.00220
- Cremisi, F. (2013). MicroRNAs and cell fate in cortical and retinal development. *Front. Cell. Neurosci.* 7:141. doi: 10.3389/fncel.2013.00141
- Della Ragione, F., Gagliardi, M., D'Esposito, M., and Matarazzo, M. R. (2014). Non-coding RNAs in chromatin disease involving neurological defects. *Front. Cell. Neurosci.* 8:54. doi: 10.3389/fncel.2014.00054
- Dogini, D. B., Avansini, S. H., Vieira, A. S., and Lopes-Cendes, I. (2013). MicroRNA regulation and dysregulation in epilepsy. *Front. Cell. Neurosci.* 7:172. doi: 10.3389/fncel.2013.00172
- Eacker, S. M., Dawson, T. M., and Dawson, V. L. (2013). The interplay of microRNA and neuronal activity in health and disease. *Front. Cell. Neurosci.* 7:136. doi: 10.3389/fncel.2013.00136
- Elramah, S., Landry, M., and Favereaux, A. (2014). MicroRNAs regulate neuronal plasticity and are involved in pain mechanisms. *Front. Cell. Neurosci.* 8:31. doi: 10.3389/fncel.2014.00031
- Ernst, C., and Morton, C. C. (2013). Identification and function of long non-coding RNA. *Front. Cell. Neurosci.* 7:168. doi: 10.3389/fncel.2013.00168
- Fragkouli, A., and Doxakis, E. (2014). miR-7 and miR-153 protect neurons against MPP(+)-induced cell death via upregulation of mTOR pathway. *Front. Cell. Neurosci.* 8:182. doi: 10.3389/fncel.2014.00182
- Gavazzo, P., Vassalli, M., Costa, D., and Pagano, A. (2013). Novel ncRNAs transcribed by Pol III and elucidation of their functional relevance by biophysical approaches. *Front. Cell. Neurosci.* 7:203. doi: 10.3389/fncel.2013.00203
- Goodall, E. F., Heath, P. R., Bandmann, O., Kirby, J., and Shaw, P. J. (2013). Neuronal dark matter: the emerging role of microRNAs in neurodegeneration. *Front. Cell. Neurosci.* 7:178. doi: 10.3389/fncel.2013.00178
- Guffanti, A., Simchovitz, A., and Soreq, H. (2014). Emerging bioinformatics approaches for analysis of NGS-derived coding and non-coding RNAs in neurodegenerative diseases. *Front. Cell. Neurosci.* 8:89. doi: 10.3389/fncel.2014.00089
- Gummalla, M., Galetti, S., Maeda, R. K., and Karch, F. (2014). Hox gene regulation in the central nervous system of *Drosophila*. *Front. Cell. Neurosci.* 8:96. doi: 10.3389/fncel.2014.00096
- Heise, C., Gardoni, F., Culotta, L., di Luca, M., Verpelli, C., and Sala, C. (2014). Elongation factor-2 phosphorylation in dendrites and the regulation of dendritic mRNA translation in neurons. *Front. Cell. Neurosci.* 8:35. doi: 10.3389/fncel.2014.00035
- Hong, J., Zhang, H., Kawase-Koga, Y., and Sun, T. (2013). MicroRNA function is required for neurite outgrowth of mature neurons in the mouse postnatal cerebral cortex. *Front. Cell. Neurosci.* 7:151. doi: 10.3389/fncel.2013.00151
- Iyengar, B. R., Choudhary, A., Sarangdhar, M. A., Venkatesh, K. V., Gadgil, C. J., and Pillai, B. (2014). Non-coding RNA interact to regulate neuronal development and function. *Front. Cell. Neurosci.* 8:47. doi: 10.3389/fncel.2014.00047
- Iyer, A. N., Bellon, A., and Baudet, M. L. (2014). microRNAs in axon guidance. *Front. Cell. Neurosci.* 8:78. doi: 10.3389/fncel.2014.00078
- Kaplan, B. B., Kar, A. N., Gioio, A. E., and Aschrafi, A. (2013). MicroRNAs in the axon and presynaptic nerve terminal. *Front. Cell. Neurosci.* 7:126. doi: 10.3389/fncel.2013.00126
- Knauss, J. L., Bian, S., and Sun, T. (2013). Plasmid-based target protectors allow specific blockade of miRNA silencing activity in mammalian developmental systems. *Front. Cell. Neurosci.* 7:163. doi: 10.3389/fncel.2013.00163
- Kye, M. J., and Goncalves Ido, C. (2014). The role of miRNA in motor neuron disease. *Front. Cell. Neurosci.* 8:15. doi: 10.3389/fncel.2014.00015
- Maciotta, S., Meregalli, M., and Torrente, Y. (2013). The involvement of microRNAs in neurodegenerative diseases. *Front. Cell. Neurosci.* 7:265. doi: 10.3389/fncel.2013.00265
- Maffioletti, E., Tardito, D., Gennarelli, M., and Bocchio-Chiavetto, L. (2014). Micro spies from the brain to the periphery: new clues from studies on microRNAs in neuropsychiatric disorders. *Front. Cell. Neurosci.* 8:75. doi: 10.3389/fncel.2014.00075
- Nieto-Diaz, M., Esteban, F. J., Reigada, D., Munoz-Galdeano, T., Yunta, M., Caballero-Lopez, M., et al. (2014). MicroRNA dysregulation in spinal cord injury: causes, consequences and therapeutics. *Front. Cell. Neurosci.* 8:53. doi: 10.3389/fncel.2014.00053
- Osella, M., Bosia, C., Cora, D., and Caselle, M. (2011). The role of incoherent microRNA-mediated feedforward loops in noise buffering. *PLoS Comput. Biol.* 7:e1001101. doi: 10.1371/journal.pcbi.1001101
- Pai, B., Siripornmongkolchai, T., Berentsen, B., Pakzad, A., Vieuille, C., Pallesen, S., et al. (2014). NMDA receptor-dependent regulation of miRNA expression and association with Argonaute during LTP *in vivo*. *Front. Cell. Neurosci.* 7:285. doi: 10.3389/fncel.2013.00285
- Pascarella, G., Lazarevic, D., Plessy, C., Bertin, N., Akalin, A., Vlachouli, C., et al. (2014). NanoCAGE analysis of the mouse olfactory epithelium identifies the expression of vomeronasal receptors and of proximal LINE elements. *Front. Cell. Neurosci.* 8:41. doi: 10.3389/fncel.2014.00041
- Serafini, G., Pompili, M., Hansen, K. F., Obrietan, K., Dwivedi, Y., Amore, M., et al. (2013). MicroRNAs: fundamental regulators of gene expression in major affective disorders and suicidal behavior? *Front. Cell. Neurosci.* 7:208. doi: 10.3389/fncel.2013.00208
- Sheinerman, K. S., and Umansky, S. R. (2013). Circulating cell-free microRNA as biomarkers for screening, diagnosis and monitoring of neurodegenerative diseases and other neurologic pathologies. *Front. Cell. Neurosci.* 7:150. doi: 10.3389/fncel.2013.00150
- Siciliano, V., Garzilli, I., Fracassi, C., Criscuolo, S., Ventre, S., and di Bernardo, D. (2013). MiRNAs confer phenotypic robustness to gene networks by suppressing biological noise. *Nat. Commun.* 4:2364. doi: 10.1038/ncomms3364
- Terzibasi Tozzini, E., Savino, A., Ripa, R., Battistoni, G., Baumgart, M., and Cellerino, A. (2014). Regulation of microRNA expression in the neuronal stem cell niches during aging of the short-lived annual fish *Nothobranchius furzeri*. *Front. Cell. Neurosci.* 8:51. doi: 10.3389/fncel.2014.00051

Tognini, P., and Pizzorusso, T. (2012). MicroRNA212/132 family: molecular transducer of neuronal function and plasticity. *Int. J. Biochem. Cell Biol.* 44, 6–10. doi: 10.1016/j.biocel.2011.10.015

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