



Mast Cells, Neuroinflammation and Pain in Fibromyalgia Syndrome

Theoharis C. Theoharides^{1,2,3,4*}, Irene Tsilioni¹ and Mona Bawazeer^{1,2,5}

¹ Laboratory of Molecular Immunopharmacology and Drug Discovery, Department of Immunology, Tufts University School of Medicine, Boston, MA, United States, ² Sackler School of Graduate Biomedical Sciences, Tufts University, Boston, MA, United States, ³ Department of Internal Medicine, Tufts Medical Center, Tufts University School of Medicine, Boston, MA, United States, ⁴ Department of Psychiatry, Tufts Medical Center, Tufts University School of Medicine, Boston, MA, United States, ⁵ Department of Basic Medical Sciences, College of Medicine, King Saud Bin Abdulaziz University for Health Sciences, Riyadh, Saudi Arabia

Fibromyalgia Syndrome (FMS) is a disorder of chronic, generalized muscular pain, accompanied by sleep disturbances, fatigue and cognitive dysfunction. There is no definitive pathogenesis except for altered central pain pathways. We previously reported increased serum levels of the neuropeptides substance P (SP) and its structural analogue hemokinin-1 (HK-1) together with the pro-inflammatory cytokines IL-6 and TNF in FMS patients as compared to sedentary controls. We hypothesize that thalamic mast cells contribute to inflammation and pain, by releasing neuro-sensitizing molecules that include histamine, IL-1 β , IL-6 and TNF, as well as calcitonin-gene related peptide (CGRP), HK-1 and SP. These molecules could either stimulate thalamic nociceptive neurons directly, or via stimulation of microglia in the diencephalon. As a result, inhibiting mast cell stimulation could be used as a novel approach for reducing pain and the symptoms of FMS.

OPEN ACCESS

Edited by:

Kempuraj Duraisamy,
University of Missouri, United States

Reviewed by:

Domenico De Berardis,
Azienda Usl Teramo, Italy

Mihir Gupta,

University of California, San Diego,
United States

*Correspondence:

Theoharis C. Theoharides
theoharis.theoharides@tufts.edu

Specialty section:

This article was submitted to
Cellular Neuropathology,
a section of the journal
Frontiers in Cellular Neuroscience

Received: 20 May 2019

Accepted: 16 July 2019

Published: 02 August 2019

Citation:

Theoharides TC, Tsilioni I and Bawazeer M (2019) Mast Cells, Neuroinflammation and Pain in Fibromyalgia Syndrome. *Front. Cell. Neurosci.* 13:353. doi: 10.3389/fncel.2019.00353

INTRODUCTION

Fibromyalgia Syndrome is a disorder of chronic generalized muscular pain, stiffness, generalized fatigue, sleep abnormalities, (Clauw et al., 2011; Schmidt-Wilcke and Clauw, 2011; Clauw, 2014) and cognitive problems (Theoharides et al., 2015b; Hauser et al., 2019) assessed by the FSQ (Ferrari and Russell, 2013), which has about 93% sensitivity and 92% specificity (Clauw, 2014). FMS affects about 5% of adults, primarily women 20–60 years of age (Branco et al., 2010) and belongs to a family of overlapping painful conditions (Table 1) known as CSS (Yunus, 2007; Theoharides, 2013). Central sensitization is recognized as the main mechanism involved (Woodman, 2013) and is characterized by allodynia, pain from an otherwise non-painful stimulus, (Russell and Larson, 2009) and hyperalgesia (Staud et al., 2001) due to an exaggerated response to a painful stimulus

Abbreviations: ACR, American College of Rheumatology; ASD, autism spectrum disorder; CGRP, calcitonin-gene related peptide; CIRS, chronic inflammatory response syndrome; CRH, corticotropin-releasing hormone; CSF, cerebrospinal fluid; CSS, central sensitivity syndromes; CXCL8, IL-8; Fc ϵ RI, high affinity surface receptors; FMS, fibromyalgia syndrome; FSQ, fibromyalgia survey questionnaire; IBS, irritable bowel syndrome; IC/BPS, interstitial cystitis/bladder pain syndrome; IgE, immunoglobulin E; MCP-1/CCL2, monocyte chemoattractant protein-1; ME/CFS, myalgic encephalomyelitis/chronic fatigue syndrome; MRGPRX2, mas-related G-protein coupled receptor member X2; (mt)DNA, mitochondrial; NGF, nerve growth factor; PAF, platelet activating factor; PTSD, post-traumatic stress disorder; SCF, stem cell factor; SP, substance P; TLRs, toll-like receptors; TMD, myogenic temporomandibular disorder.

(Woolf, 2011). The pathogenesis of FMS remains unknown and with no objective diagnostic criteria (McBeth and Mulvey, 2012; Wolfe and Walitt, 2013). FMS patients have reduced tolerance to pain, especially extremes of heat and cold (Desmeules et al., 2003). There is considerable evidence of altered circuitry of pain networks and (Jensen et al., 2012; Flodin et al., 2014) abnormal pain processing in FMS (Staud, 2011).

The PubMed database was searched between 1960 and 2018 using the terms fatigue, fibromyalgia, hypothalamus, inflammation, mast cells, pain and stress. Only articles in English were included.

Here we discuss how brain mast cell release of neurosensitizing mediators in the thalamus leads to focal inflammation and contribute to the pathogenesis of FMS.

Neurohormonal Triggers of Mast Cells Contribute to Focal Inflammation in the Diencephalon

It was recently proposed that FMS may involve localized inflammation in the hypothalamus (Theoharides et al., 2015c). Elevations in pro-inflammatory chemokines/cytokines could negatively impact symptoms (Bazzichi et al., 2007; Carvalho et al., 2008; Nugraha et al., 2013) leading to sensitization of peripheral and central nociceptors (Uceyler et al., 2011; Behm et al., 2012; Hornig et al., 2015). Increased levels of the pro-inflammatory chemokine IL-8 (CXCL8) have been reported in the serum and CSF in patients with FMS (Ross et al., 2010; Kadetoff et al., 2012; Rodriguez-Pinto et al., 2014). Chemokines facilitate nociception by directly acting on chemokine receptors present along the pain pathway (Abbadie, 2005; Charo and Ransohoff, 2006).

The cytokines TNF and IL-17 greatly contribute to the inflammatory response (Romero-Sanchez et al., 2011; Griffin et al., 2012). Plasma levels of IL-17 were increased and correlated with levels of TNF in patients with FMS (Pernambuco et al., 2013). CSF and serum IL-17 also positively correlated with pain (Meng et al., 2013) and anxiety (Liu et al., 2012). Mast cells, themselves, can secrete IL-17; moreover, IL-6 and TGF β from mast cells contribute to the development of Th-17 cells (Kenna and Brown, 2013).

TABLE 1 | Pain Syndromes Comorbid with FMS.

- Chronic inflammatory response syndrome (CIRS)
- Functional dyspepsia
- Gulf War Illness (GWI)
- Interstitial cystitis/bladder pain syndrome (IC/BPS)
- Irritable bowel syndrome (IBS)
- Mast cell mediator disorder (MCMD)
- Mastocytosis
- Migraines
- Myalgic Encephalomyelitis/Chronic Fatigue Syndrome (ME/CFS)
- Myogenic temporomandibular disorder (TMD)
- Myofacial pain syndrome
- Post-traumatic stress disorder (PTSD)
- Restless leg syndrome
- Temporomandibular pain syndrome (TMS)
- Tension headache

Fibromyalgia syndrome worsen by stress, (Geenen et al., 2002) which augments pain responses (Bote et al., 2012, 2013). Plasma concentrations of cortisol are increased in the evening, suggesting disruption of the circadian rhythm (Crofford et al., 2004). Serum levels of CRH, which is secreted under stress, were increased in patients with FMS (Tsiliioni et al., 2016). CRH was also increased in the CSF of such patients and correlated with severity of pain (McLean et al., 2006). Physiological stress was reported to be the most common trigger in patients with systemic mastocytosis (SM) (Jennings et al., 2014) who also commonly experience FMS (Theoharides et al., 2015d, 2019). We reported increased levels of CRH in the serum of one patient with indolent systemic mastocytosis (Theoharides et al., 2014). CRH can trigger human mast cells to release VEGF without histamine or tryptase (Cao et al., 2005). CRH also has synergistic action with NT stimulating VEGF release. As a result, there is increased vascular permeability in the skin and the blood-brain barrier (BBB) (Esposito et al., 2002; Donelan et al., 2006; Theoharides and Konstantinidou, 2007). Stress also disrupts the gut-blood barrier (Theoharides et al., 1999; Wallon et al., 2008) allowing for gut microbiome-associated molecules, such as propionate (Minerbi et al., 2019) to enter the brain and contribute to focal inflammation. These results have led to the conclusion that mast cells may serve as “immune gate to the brain” (Theoharides, 1990; Ribatti, 2015).

Levels of the neuropeptide SP (Russell, 1998) and NGF (Giovengo et al., 1999) are elevated in the CSF of FMS patients. NGF has been reported to increase nociception and hyperalgesia (Maren, 2017). The SP receptor NK-1 has been involved in the pathophysiology of pain (Greenwood-Van et al., 2014). We reported increased serum levels of SP, its structural analogue Hemokinin-1 (HK-1) and TNF in patients with FMS (Tsiliioni et al., 2016). SP (Theoharides et al., 2010a,b) and NGF (Levi-Montalcini, 1987) can stimulate mast cells. Moreover, SP induced mast cell expression of CRHR-1 (Scholzen et al., 2001). Cerebrovascular mast cells were stimulated by CGRP, (Reynier-Rebuffel et al., 1994; Ottosson and Edvinsson, 1997) which is now well established to participate in the pathophysiology of headaches (Edvinsson, 2018). In addition to neuropeptides, sex hormones can also affect mast cell reactivity. For instance, estradiol augments immune (Kovats, 2015) and allergic (Hox et al., 2015) processes. In particular, we had reported expression of estrogen receptors on rodent mast cells (Pang et al., 1995). We also reported that 17 β -estradiol further increased stimulation of mast cells by SP (Theoharides et al., 1993). Such findings may help explain why FMS is more common in women.

In addition to allergic reactions, mast cells contribute to innate immunity, (Galli et al., 2011) autoimmunity (Rottem and Mekori, 2005) and inflammation (Theoharides et al., 2010a).

Thalamic Mast Cells Secrete Neurosensitizing Mediators

Increasing evidence supports the involvement of mast cells in FMS (Lucas et al., 2006; Pollack, 2014) and comorbid disorders (Theoharides, 2013) as well as other inflammatory (Galli et al., 2008; Theoharides et al., 2010a) and painful conditions, (Heron and Dubayle, 2013; Chatterjea and Martinov, 2014) as well

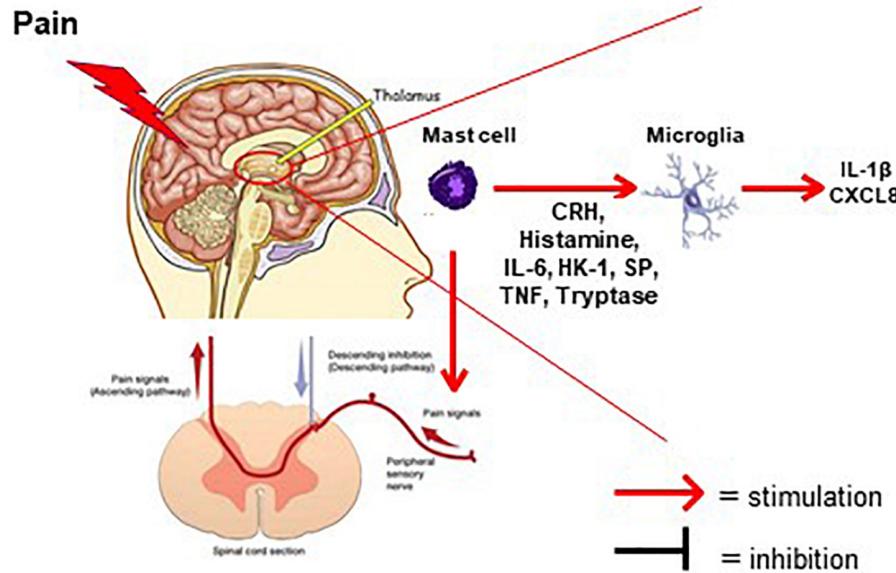


FIGURE 1 | Diagram depicting the involvement of mast cells in the generation of pain in FMS. Mast cells (violet color) in the thalamus secrete pro-inflammatory and neuro-sensitizing mediators (CRH, histamine, IL-6, HK-1, SP, TNF, Tryptase). These mediators can then activate either microglia in thalamic nuclei or ascending nociceptive tracks creating the sensation of pain. Possible natural molecules to inhibit stimulated mast cells and/or microglia are flavonoids such as luteolin or tetramethoxyluteolin (Methylut).

as neuroimmune interactions (Skaper et al., 2017) (Figure 1). Chronic urticaria, which involves stimulation of skin mast cells is more common in FMS (Torresani et al., 2009). Moreover, mast cells are significantly increased in the papillary dermis of FMS patients (Blanco et al., 2010). The chemokines monocyte chemoattractant protein-1 (MCP-1/CCL2) and eotaxin (CCL-11) are elevated in plasma of FMS patients (Zhang et al., 2008). MCP-1 is a strong mast cell chemoattractant (Conti et al., 1998) and also triggers mast cells in rodents (Conti and Theoharides, 1994). MCP-1 induced prolonged muscle hyperalgesia in rats via activation of its high-affinity receptor, CC Chemokine receptor 2 (CCR2), on the peripheral nerve terminals (Alvarez et al., 2014). Myoblasts treated with MCP-1 secreted significant amounts of the key pro-inflammatory cytokine IL-1 β (Zhang et al., 2008). C-reactive protein (CRP) is now considered a marker of chronic inflammation. CRP may be useful in the diagnostic of FMS (and depression/anxiety that often accompany FMS), even though there is no direct correlation reported (De Berardis et al., 2006, 2017; Orsolini et al., 2018).

Mast cells derive from the bone marrow and mature in response to SCF, which acts via the cell surface tyrosine kinase KIT receptor (Galli et al., 2011). Mast cell progenitors then migrate in all tissues. As a result, mast cell mediators can affect all organs and lead to multiple symptoms. Mast cells are found adjacent to blood vessels and nerve endings; in the brain, mast cells are located in the thalamus, hypothalamus and median eminence (Edvinsson et al., 1976; Lambracht-Hall et al., 1990; Theoharides et al., 2015d).

Mast cells are known to be stimulated by IgE, via activation of its unique surface receptors (Fc ϵ RI) (Rivera et al., 2008). Mast

cells can also be stimulated via TLRs, (Abraham and St John, 2010; Zhang et al., 2010). Stimulated mast cells secrete multiple vasoactive, pro-inflammatory and neuro-sensitizing molecules (Galli and Tsai, 2008; Theoharides et al., 2010a). Stimulation of mast cells can be augmented by the cytokine IL-33, (Fux et al., 2014) which synergizes with SP to induce release of impressive amounts of VEGF, (Theoharides et al., 2010b) TNF (Taracanova et al., 2017) or IL-1 β (Taracanova et al., 2018). As a result, mast cells can serve as “sensors of cell danger” (Theoharides, 1996; Enoksson et al., 2011; Theoharides et al., 2015a).

Mast cell secretory granules store many preformed pro-inflammatory and neuro-sensitizing mediators including bradykinin, histamine, TNF and tryptase (Nakae et al., 2005; Olszewski et al., 2007). Mast cells also release *de novo* synthesized molecules: (a) lipid mediators (leukotrienes, prostaglandins, and PAF), (b) cytokines (IL-6, IL-13, IL-33, TNF) and (c) chemokines (CXCL8, CCL2, CCL5), (Theoharides et al., 2015d; Mukai et al., 2018). Mast cell could often release mediators selectively without histamine or tryptase (Theoharides et al., 2007). Mast cells also release IL-31, which is important in the sensation of itching and pain, in response to IgE and SP, IL-33 and specifically their combination (Petric et al., 2018). We reported that mast cells can release mtDNA, which is mistaken as a pathogen and stimulates inflammatory responses (Zhang B. et al., 2012).

Finally, mast cells can release extracellular vesicles (exosomes) (Skokos et al., 2002, 2003) that could deliver regulatory molecules, including mtDNA and microRNAs (Kawikova and Askenase, 2014). Such microvesicles have been implicated in brain disorders (Tsiloni et al., 2014;

Kawikova and Askenase, 2014) and pain disorders (Rafiee et al., 2018; Silva-Freire et al., 2019). We recently reported that extracellular vesicles are increased in the serum of children with ASD, contained mtDNA and stimulated cultured human microglia to secrete the pro-inflammatory molecules IL-1 β and CXCL8 (Tsilioni and Theoharides, 2018).

Mast Cell Interactions With Microglia

Mast cells communicate with microglia (Skaper et al., 2012, 2014b). Mediators secreted from mast cells, (Zhang et al., 2016) such as histamine (Dong et al., 2014) and tryptase, (Zhang S. et al., 2012) can activate microglia leading to secretion of the pro-inflammatory cytokines IL-1 β , IL-6 and TNF. Microglia can also be activated by CRH secreted from mast cells (Wang et al., 2002; Kempuraj et al., 2004). Stimulation of brain mast cells in mice led to activation of microglia, which was decreased by administration of a mast cell inhibitor (Dong et al., 2017).

Microglia are involved in synapse plasticity, (Shemer et al., 2015; Wu et al., 2015; Reu et al., 2017) but are responsible for innate immunity of the brain (Ransohoff and Brown, 2012; Aguzzi et al., 2013). Microglia contribute to brain inflammation (Hagberg et al., 2012; Aguzzi et al., 2013; Nakagawa and Chiba, 2016) and the pathogenesis of different brain disorders, (Takeda et al., 2014; Reus et al., 2015; Faden et al., 2016; Garden and Campbell, 2016; Groh and Martini, 2017; Koutsouras et al., 2017; Pennisi et al., 2017; Jiang et al., 2018; Thonhoff et al., 2018) especially ASD (Vargas et al., 2005; Morgan et al., 2010; Suzuki et al., 2013; Edmonson et al., 2014; Gupta et al., 2014). Microglia in the thalamus have been discussed in the context of pain, especially maintaining the pain sensation even after the original painful stimulus is not present (Banati, 2002; Hansson, 2010; Saghaei et al., 2013; Blaszczyk et al., 2018).

CONCLUSION

Mast cells have been implicated in headaches (Theoharides, 1983; Theoharides et al., 2005) and pain (Xanthos et al., 2011; Aich et al., 2015; Gupta and Harvima, 2018). Activation of the mast cell-specific receptor, MRGPRX2, (McNeil et al., 2015) and its mouse analogue, Mrgprb2, mediated inflammatory mechanical and thermal hyperalgesia (Green et al., 2019). Hence, mast cells are key players of neuroendocrine (Theoharides, 2017) and painful disorders (Theoharides et al., 2019).

REFERENCES

- Abbadie, C. (2005). Chemokines, chemokine receptors and pain. *Trends Immunol.* 26, 529–534. doi: 10.1016/j.it.2005.08.001
- Abraham, S. N., and St John, A. L. (2010). Mast cell-orchestrated immunity to pathogens. *Nat. Rev. Immunol.* 10, 440–452. doi: 10.1038/nri2782
- Aguzzi, A., Barres, B. A., and Bennett, M. L. (2013). Microglia: scapegoat, saboteur, or something else? *Science* 339, 156–161. doi: 10.1126/science.1227901
- Aich, A., Afrin, L. B., and Gupta, K. (2015). Mast cell-mediated mechanisms of nociception. *Int. J. Mol. Sci.* 16, 29069–29092. doi: 10.3390/jims161226151
- Alvarez, P., Green, P. G., and Levine, J. D. (2014). Role for monocyte chemoattractant protein-1 in the induction of chronic muscle pain in the rat. *Pain* 155, 1161–1167. doi: 10.1016/j.pain.2014.03.004

In this context, inhibitors of mast cells (Harvima et al., 2014) would be useful in the treatment of FMS. Natural molecules could include the flavonoids, luteolin (Kempuraj et al., 2008; Theoharides et al., 2015c; Ashaari et al., 2018) and tetramethoxyluteolin, (Theoharides et al., 2017; Theoharides and Tsilioni, 2018) alone or in combination with other substances selected to reduce stress (Theoharides and Kavaloti, 2018). Other natural molecules could include palmitoylethanolamide, (Schweiger et al., 2019) which apparently inhibits neuro-inflammation (Skaper et al., 2013, 2015) and reduces pain (Skaper et al., 2014a; Impellizzeri et al., 2016).

FUTURE DIRECTIONS

Research should focus on identifying in serum of patients with FMS novel molecules that are involved in pain transmission such as bradykinin, CGRP and IL-31. Extracellular vesicles should also be isolated from the serum and CSF of FMS patients, their content identified, and their effect investigated on cultured human mast cells and microglia. Such possible interactions would serve as useful *in vitro* assays for the screening of potential novel treatment agents. Recent reports have also stressed the possible use of the cytokine IL-37, (Mastrangelo et al., 2018) which is known to have anti-inflammatory actions (Cavalli and Dinarello, 2018). It would be important to explore the possible use of IL-37 isoforms in the treatment of FMS.

AUTHOR CONTRIBUTIONS

TT, IT, and MB participated in searching the literature. TT and IT wrote or contributed to the writing of the manuscript. IT prepared the figure.

FUNDING

Some aspects of our work described were supported in part by the National Institutes of Health (NIH) (Grants NS38326 and AR47652), as well as the Michael and Katherine Johnson Family Fund to TT.

- Ashaari, Z., Hadjzadeh, M. A., Hassanzadeh, G., Alizamir, T., Yousefi, B., Keshavarzi, Z., et al. (2018). The flavone luteolin improves central nervous system disorders by different mechanisms: a review. *J. Mol. Neurosci.* 65, 491–506. doi: 10.1007/s12031-018-1094-2
- Banati, R. B. (2002). Brain plasticity and microglia: is transsynaptic glial activation in the thalamus after limb denervation linked to cortical plasticity and central sensitisation? *J. Physiol. Paris* 96, 289–299. doi: 10.1016/s0928-4257(02)0018-9
- Bazzichi, L., Rossi, A., Massimetti, G., Giannaccini, G., Giuliano, T., De Feo, F., et al. (2007). Cytokine patterns in fibromyalgia and their correlation with clinical manifestations. *Clin. Exp. Rheumatol.* 25, 225–230.

- Behm, F. G., Gavin, I. M., Karpenko, O., Lindgren, V., Gaitonde, S., Gashkoff, P. A., et al. (2012). Unique immunologic patterns in fibromyalgia. *BMC Clin. Pathol.* 12:25. doi: 10.1186/1472-6890-12-25
- Blanco, I., Beritez, N., Arguelles, M., Carcaba, V., Fernandez, F., Janciauskienė, S., et al. (2010). Abnormal overexpression of mastocytes in skin biopsies of fibromyalgia patients. *Clin. Rheumatol.* 29, 1403–1412. doi: 10.1007/s10067-010-1474-7
- Blaszczyk, L., Maitre, M., Leste-Lasserre, T., Clark, S., Cota, D., Oliet, S. H. R., et al. (2018). Sequential alteration of microglia and astrocytes in the rat thalamus following spinal nerve ligation. *J. Neuroinflammation* 15:349. doi: 10.1186/s12974-018-1378-z
- Bote, M. E., Garcia, J. J., Hinchado, M. D., and Ortega, E. (2012). Inflammatory/stress feedback dysregulation in women with fibromyalgia. *Neuroimmunomodulation* 19, 343–351. doi: 10.1159/000341664
- Bote, M. E., Garcia, J. J., Hinchado, M. D., and Ortega, E. (2013). Fibromyalgia: anti-inflammatory and stress responses after acute moderate exercise. *PLoS One* 8:e74524. doi: 10.1371/journal.pone.0074524
- Branco, J. C., Bannwarth, B., Failde, I., Abello, C. J., Blotman, F., Spaeth, M., et al. (2010). Prevalence of fibromyalgia: a survey in five European countries. *Semin. Arthritis Rheum.* 39, 448–453. doi: 10.1016/j.semarthrit.2008.12.003
- Cao, J., Papadopoulou, N., Kempuraj, D., Boucher, W. S., Sugimoto, K., Cetrulo, C. L., et al. (2005). Human mast cells express corticotropin-releasing hormone (CRH) receptors and CRH leads to selective secretion of vascular endothelial growth factor. *J. Immunol.* 174, 7665–7675. doi: 10.4049/jimmunol.174.12.7665
- Carvalho, L. S., Correa, H., Silva, G. C., Campos, F. S., Baiao, F. R., Ribeiro, L. S., et al. (2008). May genetic factors in fibromyalgia help to identify patients with differentially altered frequencies of immune cells? *Clin. Exp. Immunol.* 154, 346–352. doi: 10.1111/j.1365-2249.2008.03787.x
- Cavalli, G., and Dinarello, C. A. (2018). Suppression of inflammation and acquired immunity by IL-37. *Immunol. Rev.* 281, 179–190. doi: 10.1111/imr.12605
- Charo, I. F., and Ransohoff, R. M. (2006). The many roles of chemokines and chemokine receptors in inflammation. *N. Engl. J. Med.* 354, 610–621. doi: 10.1056/nejmra052723
- Chatterjea, D., and Martinov, T. (2014). Mast cells: versatile gatekeepers of pain. *Mol. Immunol.* 63, 38–44. doi: 10.1016/j.molimm.2014.03.001
- Clauw, D. J. (2014). Fibromyalgia: a clinical review. *JAMA* 311, 1547–1555. doi: 10.1001/jama.2014.3266
- Clauw, D. J., Arnold, L. M., and McCarberg, B. H. (2011). The science of fibromyalgia. *Mayo Clin. Proc.* 86, 907–911. doi: 10.4065/mcp.2011.0206
- Conti, P., Reale, M., Barbacane, R. C., Letourneau, R., and Theoharides, T. C. (1998). Intramuscular injection of hrRANTES causes mast cell recruitment and increased transcription of histidine decarboxylase: lack of effects in genetically mast cell-deficient W/Wv mice. *FASEB J.* 12, 1693–1700. doi: 10.1096/faseb.12.15.1693
- Conti, P., and Theoharides, T. C. (1994). Monocyte chemoattractant Protein-1 (MCP-1) is active on mast cells and causes clump formation. *Int. J. Immunopathol. Pharmacol.* 7, 149–151.
- Crofford, L. J., Young, E. A., Engleberg, N. C., Korszun, A., Brucksch, C. B., McClure, L. A., et al. (2004). Basal circadian and pulsatile ACTH and cortisol secretion in patients with fibromyalgia and/or chronic fatigue syndrome. *Brain Behav. Immun.* 18, 314–325. doi: 10.1016/s0889-1591(04)00021-2
- De Berardis, D., Campanella, D., Gambi, F., La, R. R., Carano, A., Conti, C. M., et al. (2006). The role of C-reactive protein in mood disorders. *Int. J. Immunopathol. Pharmacol.* 19, 721–725.
- De Berardis, D., Serroni, N., Campanella, D., Marini, S., Rapini, G., Valchera, A., et al. (2017). Alexithymia, suicide ideation, C-Reactive Protein, and serum lipid levels among outpatients with generalized anxiety disorder. *Arch. Suicide Res.* 21, 100–112. doi: 10.1080/13811118.2015.1004485
- Desmeules, J. A., Cedraschi, C., Rapiti, E., Baumgartner, E., Finckh, A., Cohen, P., et al. (2003). Neurophysiologic evidence for a central sensitization in patients with fibromyalgia. *Arthritis Rheum.* 48, 1420–1429. doi: 10.1002/art.10893
- Donelan, J., Boucher, W., Papadopoulou, N., Lytinas, M., Papaliodis, D., and Theoharides, T. C. (2006). Corticotropin-releasing hormone induces skin vascular permeability through a neurotensin-dependent process. *Proc. Natl. Acad. Sci. U.S.A.* 103, 7759–7764. doi: 10.1073/pnas.0602210103
- Dong, H., Zhang, W., Zeng, X., Hu, G., Zhang, H., He, S., et al. (2014). Histamine induces upregulated expression of histamine receptors and increases release of inflammatory mediators from microglia1. *Mol. Neurobiol.* 49, 1487–1500. doi: 10.1007/s12035-014-8697-6
- Dong, H., Zhang, X., Wang, Y., Zhou, X., Qian, Y., and Zhang, S. (2017). Suppression of brain mast cells degranulation inhibits microglial activation and central nervous system inflammation. *Mol. Neurobiol.* 54, 997–1007. doi: 10.1007/s12035-016-9720-x
- Edmonson, C., Ziats, M. N., and Rennert, O. M. (2014). Altered glial marker expression in autistic post-mortem prefrontal cortex and cerebellum. *Mol. Autism* 5:3. doi: 10.1186/2040-2392-5-3
- Edvinsson, L. (2018). The CGRP pathway in migraine as a viable target for therapies. *Headache* 58(Suppl. 1), 33–47. doi: 10.1111/head.13305
- Edvinsson, L., Owman, C., and Sjöberg, N. O. (1976). Autonomic nerves, mast cells and amine receptors in human brain vessels. A histochemical and pharmacological study. *Brain Res.* 115, 377–393. doi: 10.1016/0006-8993(76)90356-5
- Enoksson, M., Lyberg, K., Moller-Westerberg, C., Fallon, P. G., Nilsson, G., and Lunderius-Andersson, C. (2011). Mast cells as sensors of cell injury through IL-33 recognition. *J. Immunol.* 186, 2523–2528. doi: 10.4049/jimmunol.1003383
- Esposito, P., Chandler, N., Kandere-Grzybowska, K., Basu, S., Jacobson, S., Connolly, R., et al. (2002). Corticotropin-releasing hormone (CRH) and brain mast cells regulate blood-brain-barrier permeability induced by acute stress. *J. Pharmacol. Exp. Ther.* 303, 1061–1066. doi: 10.1124/jpet.102.038497
- Faden, A. I., Wu, J., Stoica, B. A., and Loane, D. J. (2016). Progressive inflammation-mediated neurodegeneration after traumatic brain or spinal cord injury. *Br. J. Pharmacol.* 173, 681–691. doi: 10.1111/bph.13179
- Ferrari, R., and Russell, A. S. (2013). A questionnaire using the modified 2010 American College of rheumatology criteria for fibromyalgia: specificity and sensitivity in clinical practice. *J. Rheumatol.* 40, 1590–1595. doi: 10.3899/jrheum.130367
- Flodin, P. D., Martinsen, S., Lofgren, M., Bileviciute-Ljungar, I., Kosek, E., and Fransson, P. (2014). Fibromyalgia is associated with decreased connectivity between pain- and sensorimotor brain areas. *Brain Connect.* 4, 587–594. doi: 10.1089/brain.2014.0274
- Fux, M., Pecaric-Petkovic, T., Odermatt, A., Hausmann, O. V., Lorentz, A., Bischoff, S. C., et al. (2014). IL-33 is a mediator rather than a trigger of the acute allergic response in humans. *Allergy* 69, 216–222. doi: 10.1111/all.12309
- Galli, S. J., Borregaard, N., and Wynn, T. A. (2011). Phenotypic and functional plasticity of cells of innate immunity: macrophages, mast cells and neutrophils. *Nat. Immunol.* 12, 1035–1044. doi: 10.1038/ni.2109
- Galli, S. J., and Tsai, M. (2008). Mast cells: versatile regulators of inflammation, tissue remodeling, host defense and homeostasis. *J. Dermatol. Sci.* 49, 7–19. doi: 10.1016/j.jdermsci.2007.09.009
- Galli, S. J., Tsai, M., and Piliponsky, A. M. (2008). The development of allergic inflammation. *Nature* 454, 445–454. doi: 10.1038/nature07204
- Garden, G. A., and Campbell, B. M. (2016). Glial biomarkers in human central nervous system disease. *Glia* 64, 1755–1771. doi: 10.1002/glia.22998
- Geenen, R., Jacobs, J. W., and Bijlsma, J. W. (2002). Evaluation and management of endocrine dysfunction in fibromyalgia. *Rheum. Dis. Clin. North Am.* 28, 389–404. doi: 10.1016/s0889-857x(01)00009-6
- Giovengo, S. L., Russell, I. J., and Larson, A. A. (1999). Increased concentrations of nerve growth factor in cerebrospinal fluid of patients with fibromyalgia. *J. Rheumatol.* 26, 1564–1569.
- Green, D. P., Limjyunawong, N., Gour, N., Pundir, P., and Dong, X. (2019). A Mast-Cell-Specific receptor mediates neurogenic inflammation and pain. *Neuron* 101, 412–420. doi: 10.1016/j.neuron.2019.01.012
- Greenwood-Van, M. B., Mohammadi, E., Tyler, K., Pietra, C., Bee, L. A., and Dickenson, A. (2014). Synergistic effect of 5-hydroxytryptamine 3 and neurokinin 1 receptor antagonism in rodent models of somatic and visceral pain. *J. Pharmacol. Exp. Ther.* 351, 146–152. doi: 10.1124/jpet.114.216028
- Griffin, G. K., Newton, G., Tarrio, M. L., Bu, D. X., Maganto-Garcia, E., Azcutia, V., et al. (2012). IL-17 and TNF-alpha sustain neutrophil recruitment during inflammation through synergistic effects on endothelial activation. *J. Immunol.* 188, 6287–6299. doi: 10.4049/jimmunol.1200385
- Groh, J., and Martini, R. (2017). Neuroinflammation as modifier of genetically caused neurological disorders of the central nervous system: understanding pathogenesis and chances for treatment. *Glia* 65, 1407–1422. doi: 10.1002/glia.23162

- Gupta, K., and Harvima, I. T. (2018). Mast cell-neural interactions contribute to pain and itch. *Immunol. Rev.* 282, 168–187. doi: 10.1111/imr.12622
- Gupta, S., Ellis, S. E., Ashar, F. N., Moes, A., Bader, J. S., Zhan, J., et al. (2014). Transcriptome analysis reveals dysregulation of innate immune response genes and neuronal activity-dependent genes in autism. *Nat. Commun.* 5:5748. doi: 10.1038/ncomms6748
- Hagberg, H., Gressens, P., and Mallard, C. (2012). Inflammation during fetal and neonatal life: implications for neurologic and neuropsychiatric disease in children and adults. *Ann. Neurol.* 71, 444–457. doi: 10.1002/ana.22620
- Hansson, E. (2010). Long-term pain, neuroinflammation and glial activation. *Scand. J. Pain* 1, 67–72. doi: 10.1016/j.sjpain.2010.01.002
- Harvima, I. T., Levi-Schaffer, F., Draber, P., Friedman, S., Polakovicova, I., Gibbs, B. F., et al. (2014). Molecular targets on mast cells and basophils for novel therapies. *J. Allergy Clin. Immunol.* 134, 530–544. doi: 10.1016/j.jaci.2014.03.007
- Hauser, W., Sarzi-Puttini, P., and Fitzcharles, M. A. (2019). Fibromyalgia syndrome: under-, over- and misdiagnosis. *Clin. Exp. Rheumatol.* 37(Suppl. 116), 90–97.
- Heron, A., and Dubayle, D. (2013). A focus on mast cells and pain. *J. Neuroimmunol.* 264, 1–7. doi: 10.1016/j.jneuroim.2013.09.018
- Hornig, M., Montoya, J. G., Klimas, N. G., Levine, S., Felsenstein, D., Bateman, L., et al. (2015). Distinct plasma immune signatures in ME/CFS are present early in the course of illness. *Sci. Adv.* 1:e1400121. doi: 10.1126/sciadv.1400121
- Hox, V., Desai, A., Bandara, G., Gilfillan, A. M., Metcalfe, D. D., and Olivera, A. (2015). Estrogen increases the severity of anaphylaxis in female mice through enhanced endothelial nitric oxide synthase expression and nitric oxide production. *J. Allergy Clin. Immunol.* 135, 729–736. doi: 10.1016/j.jaci.2014.11.003
- Impellizzeri, D., Di, P. R., Cordaro, M., Gugliandolo, E., Casili, G., Morittu, V. M., et al. (2016). Adelmidrol, a palmitoylethanolamide analogue, as a new pharmacological treatment for the management of acute and chronic inflammation. *Biochem. Pharmacol.* 119, 27–41. doi: 10.1016/j.bcp.2016.09.001
- Jennings, S., Russell, N., Jennings, B., Slee, V., Sterling, L., Castells, M., et al. (2014). The mastocytosis society survey on mast cell disorders: patient experiences and perceptions. *J. Allergy Clin. Immunol. Pract.* 2, 70–76. doi: 10.1016/j.jaip.2013.09.004
- Jensen, K. B., Loitoile, R., Kosek, E., Petzke, F., Carville, S., Fransson, P., et al. (2012). Patients with fibromyalgia display less functional connectivity in the brain's pain inhibitory network. *Mol. Pain* 8:32. doi: 10.1186/1744-8069-8-32
- Jiang, N. M., Cowan, M., Moonah, S. N., and Petri, W. A. Jr. (2018). The impact of systemic inflammation on neurodevelopment. *Trends Mol. Med.* 24, 794–804. doi: 10.1016/j.molmed.2018.06.008
- Kadetoff, D., Lampa, J., Westman, M., Andersson, M., and Kosek, E. (2012). Evidence of central inflammation in fibromyalgia-increased cerebrospinal fluid interleukin-8 levels. *J. Neuroimmunol.* 242, 33–38. doi: 10.1016/j.jneuroim.2011.10.013
- Kawikova, I., and Askenase, P. W. (2014). Diagnostic and therapeutic potentials of exosomes in CNS diseases. *Brain Res.* 1617, 63–71. doi: 10.1016/j.brainres.2014.09.070
- Kempuraj, D., Papadopoulou, N. G., Lytinas, M., Huang, M., Kandere-Grzybowska, K., Madhappan, B., et al. (2004). Corticotropin-releasing hormone and its structurally related urocortin are synthesized and secreted by human mast cells. *Endocrinology* 145, 43–48. doi: 10.1210/en.2003-0805
- Kempuraj, D., Tagen, M., Iliopoulos, B. P., Clemons, A., Vasiadi, M., Boucher, W., et al. (2008). Luteolin inhibits myelin basic protein-induced human mast cell activation and mast cell dependent stimulation of Jurkat T cells. *Br. J. Pharmacol.* 155, 1076–1084. doi: 10.1038/bjp.2008.356
- Kenna, T. J., and Brown, M. A. (2013). The role of IL-17-secreting mast cells in inflammatory joint disease. *Nat. Rev. Rheumatol.* 9, 375–379. doi: 10.1038/nrrheum.2012.205
- Koutsouras, G. W., Ramos, R. L., and Martinez, L. R. (2017). Role of microglia in fungal infections of the central nervous system. *Virulence* 8, 705–718. doi: 10.1080/21505594.2016.1261789
- Kovats, S. (2015). Estrogen receptors regulate innate immune cells and signaling pathways. *Cell Immunol.* 294, 63–69. doi: 10.1016/j.cellimm.2015.01.018
- Lambracht-Hall, M., Dimitriadou, V., and Theoharides, T. C. (1990). Migration of mast cells in the developing rat brain. *Dev. Brain Res.* 56, 151–159. doi: 10.1016/0165-3806(90)90077-c
- Levi-Montalcini, R. (1987). The nerve growth factor 35 years later. *Science* 237, 1154–1162. doi: 10.1126/science.3306916
- Liu, Y., Ho, R. C., and Mak, A. (2012). The role of interleukin (IL)-17 in anxiety and depression of patients with rheumatoid arthritis. *Int. J. Rheum. Dis.* 15, 183–187. doi: 10.1111/j.1756-185X.2011.01673.x
- Lucas, H. J., Brauch, C. M., Settas, L., and Theoharides, T. C. (2006). Fibromyalgia-new concepts of pathogenesis and treatment. *Int. J. Immunopathol. Pharmacol.* 19, 5–10.
- Maren, S. (2017). Synapse-Specific encoding of fear memory in the amygdala. *Neuron* 95, 988–990. doi: 10.1016/j.neuron.2017.08.020
- Mastrangelo, F., Frydas, I., Ronconi, G., Kritis, S. K., Tettamanti, L., Caraffa, A., et al. (2018). Low-grade chronic inflammation mediated by mast cells in fibromyalgia: role of IL-37. *J. Biol. Regul. Homeost Agents* 32, 195–198.
- McBeth, J., and Mulvey, M. R. (2012). Fibromyalgia: mechanisms and potential impact of the ACR 2010 classification criteria. *Nat. Rev. Rheumatol.* 8, 108–116. doi: 10.1038/nrrheum.2011.216
- McLean, S. A., Williams, D. A., Stein, P. K., Harris, R. E., Lyden, A. K., Whalen, G., et al. (2006). Cerebrospinal fluid corticotropin-releasing factor concentration is associated with pain but not fatigue symptoms in patients with fibromyalgia. *Neuropsychopharmacology* 31, 2776–2782. doi: 10.1038/sj.npp.1301200
- McNeil, B. D., Pundir, P., Meeker, S., Han, L., Undem, B. J., Kulka, M., et al. (2015). Identification of a mast-cell-specific receptor crucial for pseudo-allergic drug reactions. *Nature* 519, 237–241. doi: 10.1038/nature14022
- Meng, X., Zhang, Y., Lao, L., Saito, R., Li, A., Backman, C. M., et al. (2013). Spinal interleukin-17 promotes thermal hyperalgesia and NMDA NR1 phosphorylation in an inflammatory pain rat model. *Pain* 154, 294–305. doi: 10.1016/j.pain.2012.10.022
- Minerbi, A., Gonzalez, E., Brereton, N. J. B., Anjarkouchian, A., Dewar, K., Fitzcharles, M. A., et al. (2019). Altered microbiome composition in individuals with fibromyalgia. *Pain* doi: 10.1097/j.pain.0000000000001640 [Epub ahead of print].
- Morgan, J. T., Chana, G., Pardo, C. A., Achim, C., Semendeferi, K., Buckwalter, J., et al. (2010). Microglial activation and increased microglial density observed in the dorsolateral prefrontal cortex in autism. *Biol. Psychiatry* 68, 368–376. doi: 10.1016/j.biopsych.2010.05.024
- Mukai, K., Tsai, M., Saito, H., and Galli, S. J. (2018). Mast cells as sources of cytokines, chemokines, and growth factors. *Immunol. Rev.* 282, 121–150. doi: 10.1111/imr.12634
- Nakae, S., Suto, H., Kakurai, M., Sedgwick, J. D., Tsai, M., and Galli, S. J. (2005). Mast cells enhance T cell activation: importance of mast cell-derived TNF. *Proc. Natl. Acad. Sci. U.S.A.* 102, 6467–6472. doi: 10.1073/pnas.0501912102
- Nakagawa, Y., and Chiba, K. (2016). Involvement of neuroinflammation during brain development in social cognitive deficits in autism spectrum disorder and schizophrenia. *J. Pharmacol. Exp. Ther.* 358, 504–515. doi: 10.1124/jpet.116.234476
- Nugraha, B., Korallus, C., Kielstein, H., and Gutenbrunner, C. (2013). CD3+CD56+natural killer T cells in fibromyalgia syndrome patients: association with the intensity of depression. *Clin. Exp. Rheumatol.* 31, S9–S15.
- Olszewski, M. B., Groot, A. J., Dastych, J., and Knol, E. F. (2007). TNF trafficking to human mast cell granules: mature chain-dependent endocytosis. *J. Immunol.* 178, 5701–5709. doi: 10.4049/jimmunol.178.9.5701
- Orsolini, L., Sarchione, F., Vellante, F., Fornaro, M., Matarazzo, L., Martinotti, G., et al. (2018). Protein-C reactive as biomarker predictor of schizophrenia phases of illness? A systematic review. *Curr. Neuropharmacol.* 16, 583–606. doi: 10.2174/1570159X16666180119144538
- Ottosson, A., and Edvinsson, L. (1997). Release of histamine from dural mast cells by substance P and calcitonin gene-related peptide. *Cephalgia* 17, 166–174. doi: 10.1046/j.1468-2982.1997.1703166.x
- Pang, X., Cotreau-Bibbo, M. M., Sant, G. R., and Theoharides, T. C. (1995). Bladder mast cell expression of high affinity estrogen receptors in patients with interstitial cystitis. *Br. J. Urol.* 75, 154–161. doi: 10.1111/j.1464-410x.1995.tb07303.x
- Pennisi, M., Crupi, R., Di, P. R., Ontario, M. L., Bella, R., Calabrese, E. J., et al. (2017). Inflammasomes, hormesis, and antioxidants in neuroinflammation: role

- of NLRP3 in Alzheimer disease. *J. Neurosci. Res.* 95, 1360–1372. doi: 10.1002/jnr.23986
- Pernambuco, A. P., Schetino, L. P., Alvim, C. C., Murad, C. M., Viana, R. S., Carvalho, L. S., et al. (2013). Increased levels of IL-17A in patients with fibromyalgia. *Clin. Exp. Rheumatol.* 31, S60–S63.
- Petra, A. I., Tsilioni, I., Taracanova, A., Katsarou-Katsari, A., and Theoharides, T. C. (2018). Interleukin 33 and interleukin 4 regulate interleukin 31 gene expression and secretion from human laboratory of allergic diseases 2 mast cells stimulated by substance P and/or immunoglobulin E. *Allergy Asthma Proc.* 39, 153–160. doi: 10.2500/aap.2018.38.4105
- Pollack, S. (2014). Mast cells in fibromyalgia. *Clin. Exp. Rheumatol.* 33(1 Suppl. 88):S140.
- Rafiee, Z. A., Falahatian, M., and Alsahebfosoul, F. (2018). Serum levels of histamine and diamine oxidase in multiple sclerosis. *Am. J. Clin. Exp. Immunol.* 7, 100–105.
- Ransohoff, R. M., and Brown, M. A. (2012). Innate immunity in the central nervous system. *J. Clin. Invest.* 122, 1164–1171. doi: 10.1172/JCI58644
- Reu, P., Khosravi, A., Bernard, S., Mold, J. E., Salehpour, M., Alkass, K., et al. (2017). The lifespan and turnover of microglia in the human brain. *Cell Rep.* 20, 779–784. doi: 10.1016/j.celrep.2017.07.004
- Reus, G. Z., Fries, G. R., Stertz, L., Badawy, M., Passos, I. C., Barichello, T., et al. (2015). The role of inflammation and microglial activation in the pathophysiology of psychiatric disorders. *Neuroscience* 300, 141–154. doi: 10.1016/j.neuroscience.2015.05.018
- Reynier-Rebuffel, A.-M., Mathieu, P., Callebert, J., Dimitriadou, V., Farjaudon, N., Kacem, K., et al. (1994). Substance P, calcitonin gene-related peptide, and capsaicin release serotonin from cerebrovascular mast cells. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 267, R1421–R1429.
- Ribatti, D. (2015). The crucial role of mast cells in blood-brain barrier alterations. *Exp. Cell Res.* 338, 119–125. doi: 10.1016/j.yexcr.2015.05.013
- Rivera, J., Fierro, N. A., Olivera, A., and Suzuki, R. (2008). New insights on mast cell activation via the high affinity receptor for IgE. *Adv. Immunol.* 98, 85–120. doi: 10.1016/S0065-2776(08)00403-3
- Rodriguez-Pinto, I., Agmon-Levin, N., Howard, A., and Shoenfeld, Y. (2014). Fibromyalgia and cytokines. *Immunol. Lett.* 161, 200–203. doi: 10.1016/j.imlet.2014.01.009
- Romero-Sánchez, C., Jaimes, D. A., Londono, J., De Avila, J., Castellanos, J. E., Bello, J. M., et al. (2011). Association between Th-17 cytokine profile and clinical features in patients with spondyloarthritis. *Clin. Exp. Rheumatol.* 29, 828–834.
- Ross, R. L., Jones, K. D., Bennett, R. M., Ward, R. L., Druker, B. J., and Wood, L. J. (2010). Preliminary evidence of increased pain and elevated cytokines in fibromyalgia patients with defective growth hormone response to exercise. *Open Immunol. J.* 3, 9–18. doi: 10.2174/1874226201003010009
- Rottem, M., and Mekori, Y. A. (2005). Mast cells and autoimmunity. *Autoimmun. Rev.* 4, 21–27. doi: 10.1016/j.autrev.2004.05.001
- Russell, I. J. (1998). Advances in fibromyalgia: possible role for central neurochemicals. *Am. J. Med. Sci.* 315, 377–384. doi: 10.1016/s0002-9629(15)40355-6
- Russell, I. J., and Larson, A. A. (2009). Neurophysiopathogenesis of fibromyalgia syndrome: a unified hypothesis. *Rheum. Dis. Clin. North Am.* 35, 421–435. doi: 10.1016/j.rdc.2009.06.005
- Saghaei, E., Abbaszadeh, F., Naseri, K., Ghorbanpoor, S., Afhami, M., Haeri, A., et al. (2013). Estradiol attenuates spinal cord injury-induced pain by suppressing microglial activation in thalamic VPL nuclei of rats. *Neurosci. Res.* 75, 316–323. doi: 10.1016/j.neures.2013.01.010
- Schmidt-Wilcke, T., and Clauw, D. J. (2011). Fibromyalgia: from pathophysiology to therapy. *Nat. Rev. Rheumatol.* 7, 518–527. doi: 10.1038/nrrheum.2011.98
- Scholzen, T. E., Steinhoff, M., Bonaccorsi, P., Klein, R., Amadesi, S., Geppetti, P., et al. (2001). Neutral endopeptidase terminates substance P-induced inflammation in allergic contact dermatitis. *J. Immunol.* 166, 1285–1291. doi: 10.4049/jimmunol.166.2.1285
- Schweiger, V., Martini, A., Bellamoli, P., Donadello, K., Schievano, C., Del, B. G., et al. (2019). Ultramicronized palmitoylethanolamide (um-PEA) as add-on treatment in fibromyalgia syndrome (FMS): retrospective observational study on 407 patients. *CNS Neurol. Disord. Drug Targets* doi: 10.2174/1874226201003010009 [Epub ahead of print].
- Shemer, A., Erny, D., Jung, S., and Prinz, M. (2015). Microglia plasticity during health and disease: an immunological perspective. *Trends Immunol.* 36, 614–624. doi: 10.1016/j.it.2015.08.003
- Silva-Freire, N., Mayado, A., Teodosio, C., Jara-Acevedo, M., Varez-Twose, I., Matito, A., et al. (2019). Bone marrow mast cell antibody-targetable cell surface protein expression profiles in systemic mastocytosis. *Int. J. Mol. Sci.* 20:E552. doi: 10.3390/ijms20030552
- Skaper, S. D., Facci, L., Barbierato, M., Zusso, M., Bruschetta, G., Impellizzeri, D., et al. (2015). N-Palmitoylethanolamine and neuroinflammation: a novel therapeutic strategy of resolution. *Mol. Neurobiol.* 52, 1034–1042. doi: 10.1007/s12035-015-9253-8
- Skaper, S. D., Facci, L., Fusco, M., la Valle, M. F., Zusso, M., Costa, B., et al. (2014a). Palmitoylethanolamide, a naturally occurring disease-modifying agent in neuropathic pain. *Inflammopharmacology* 22, 79–94. doi: 10.1007/s10787-013-0191-7
- Skaper, S. D., Facci, L., and Giusti, P. (2014b). Neuroinflammation, microglia and mast cells in the pathophysiology of neurocognitive disorders: a review. *CNS Neurol. Disord. Drug Targets* 13, 1654–1666. doi: 10.2174/18742262010030224206
- Skaper, S. D., Facci, L., and Giusti, P. (2013). Glia and mast cells as targets for palmitoylethanolamide, an anti-inflammatory and neuroprotective lipid mediator. *Mol. Neurobiol.* 48, 340–352. doi: 10.1007/s12035-013-8487-6
- Skaper, S. D., Facci, L., Zusso, M., and Giusti, P. (2017). Neuroinflammation, mast cells, and glia: dangerous liaisons. *Neuroscientist* 23, 478–498. doi: 10.1177/1073858416687249
- Skaper, S. D., Giusti, P., and Facci, L. (2012). Microglia and mast cells: two tracks on the road to neuroinflammation. *FASEB J.* 26, 3103–3117. doi: 10.1096/fj.11-197194
- Skokos, D., Botros, H. G., Demeure, C., Morin, J., Peronet, R., Birkenmeier, G., et al. (2003). Mast cell-derived exosomes induce phenotypic and functional maturation of dendritic cells and elicit specific immune responses in vivo. *J. Immunol.* 170, 3037–3045. doi: 10.4049/jimmunol.170.6.3037
- Skokos, D., Goubran-Botros, H., Roa, M., and Mecheli, S. (2002). Immunoregulatory properties of mast cell-derived exosomes. *Mol. Immunol.* 38, 1359–1362. doi: 10.1016/s0161-5890(02)00088-3
- Staud, R. (2011). Brain imaging in fibromyalgia syndrome. *Clin. Exp. Rheumatol.* 29, S109–S117.
- Staud, R., Vierck, C. J., Cannon, R. L., Mauderli, A. P., and Price, D. D. (2001). Abnormal sensitization and temporal summation of second pain (wind-up) in patients with fibromyalgia syndrome. *Pain* 91, 165–175. doi: 10.1016/s0304-3959(00)00432-2
- Suzuki, K., Sugihara, G., Ouchi, Y., Nakamura, K., Futatsubashi, M., Takebayashi, K., et al. (2013). Microglial activation in young adults with autism spectrum disorder. *JAMA Psychiatry* 70, 49–58.
- Takeda, S., Sato, N., and Morishita, R. (2014). Systemic inflammation, blood-brain barrier vulnerability and cognitive/non-cognitive symptoms in Alzheimer disease: relevance to pathogenesis and therapy. *Front. Aging Neurosci.* 6:171. doi: 10.3389/fnagi.2014.00171
- Taracanova, A., Alevizos, M., Karagkouni, A., Weng, Z., Norwitz, E., Conti, P., et al. (2017). SP and IL-33 together markedly enhance TNF synthesis and secretion from human mast cells mediated by the interaction of their receptors. *Proc. Natl. Acad. Sci. U.S.A.* 114, E4002–E4009. doi: 10.1073/pnas.1524845114
- Taracanova, A., Tsilioni, I., Conti, P., Norwitz, E. R., Leeman, S. E., and Theoharides, T. C. (2018). Substance P and IL-33 administered together stimulate a marked secretion of IL-1beta from human mast cells, inhibited by methoxyluteolin. *Proc. Natl. Acad. Sci. U.S.A.* 115, E9381–E9390. doi: 10.1073/pnas.1810133115
- Theoharides, T. C. (1983). Mast cells and migraines. *Perspect. Biol. Med.* 26, 672–675. doi: 10.1353/pbm.1983.0028
- Theoharides, T. C. (1990). Mast cells: the immune gate to the brain. *Life Sci.* 46, 607–617. doi: 10.1016/0024-3205(90)90129-f
- Theoharides, T. C. (1996). Mast cell: a neuroimmunoendocrine master player. *Int. J. Tissue React.* 18, 1–21.
- Theoharides, T. C. (2013). Atopic conditions in search of pathogenesis and therapy. *Clin. Ther.* 35, 544–547. doi: 10.1016/j.clinthera.2013.04.002

- Theoharides, T. C. (2017). Neuroendocrinology of mast cells: challenges and controversies. *Exp. Dermatol.* 26, 751–759. doi: 10.1111/exd.13288
- Theoharides, T. C., Alysandratos, K. D., Angelidou, A., Delivanis, D. A., Sismanopoulos, N., Zhang, B., et al. (2010a). Mast cells and inflammation. *Biochim. Biophys. Acta* 1822, 21–33.
- Theoharides, T. C., Zhang, B., Kempuraj, D., Tagen, M., Vasiadi, M., Angelidou, A., et al. (2010b). IL-33 augments substance P-induced VEGF secretion from human mast cells and is increased in psoriatic skin. *Proc. Natl. Acad. Sci. U.S.A.* 107, 4448–4453. doi: 10.1073/pnas.1000803107
- Theoharides, T. C., Dimitriadou, V., Letourneau, R. J., Rozniecki, J. J., Vliagoftis, H., and Boucher, W. S. (1993). Synergistic action of estradiol and myelin basic protein on mast cell secretion and brain demyelination: changes resembling early stages of demyelination. *Neuroscience* 57, 861–871. doi: 10.1016/0306-4522(93)90030-j
- Theoharides, T. C., Donelan, J., Kandere-Grzybowska, K., and Konstantinidou, A. (2005). The role of mast cells in migraine pathophysiology. *Brain Res. Brain Res. Rev.* 49, 65–76. doi: 10.1016/j.brainresrev.2004.11.006
- Theoharides, T. C., and Kavalaliot, M. (2018). Stress, inflammation and natural treatments. *J. Biol. Regul. Homeost Agents* 32, 1345–1347.
- Theoharides, T. C., Kempuraj, D., Tagen, M., Conti, P., and Kalogeromitros, D. (2007). Differential release of mast cell mediators and the pathogenesis of inflammation. *Immunol. Rev.* 217, 65–78. doi: 10.1111/j.1600-065x.2007.00519.x
- Theoharides, T. C., and Konstantinidou, A. (2007). Corticotropin-releasing hormone and the blood-brain-barrier. *Front. Biosci.* 12:1615–1628. doi: 10.2741/2174
- Theoharides, T. C., Letourneau, R., Patra, P., Hesse, L., Pang, X., Boucher, W., et al. (1999). Stress-induced rat intestinal mast cell intragranular activation and inhibitory effect of sulfated proteoglycans. *Dig. Dis. Sci.* 44, 87S–93S.
- Theoharides, T. C., Petra, A. I., Stewart, J. M., Tsilioni, I., Panagiotidou, S., and Akin, C. (2014). High serum corticotropin-releasing hormone (CRH) and bone marrow mast cell CRH receptor expression in a mastocytosis patient. *J. Allergy Clin. Immunol.* 134, 1197–1199. doi: 10.1016/j.jaci.2014.05.023
- Theoharides, T. C., Petra, A. I., Taracanova, A., Panagiotidou, S., and Conti, P. (2015a). Targeting IL-33 in autoimmunity and inflammation. *J. Pharmacol. Exp. Ther.* 354, 24–31. doi: 10.1124/jpet.114.222505
- Theoharides, T. C., Stewart, J. M., Hatzigelaki, E., and Kolaitis, G. (2015b). Brain "fog," inflammation and obesity: key aspects of neuropsychiatric disorders improved by luteolin. *Front. Neurosci.* 9:225. doi: 10.3389/fnins.2015.00225
- Theoharides, T. C., Tsilioni, I., Arbetman, L., Panagiotidou, S., Stewart, J. M., Gleason, R. M., et al. (2015c). Fibromyalgia, a syndrome in search of pathogenesis and therapy. *J. Pharmacol. Exp. Ther.* 355, 255–263.
- Theoharides, T. C., Valent, P., and Akin, C. (2015d). Mast cells, mastocytosis, and related disorders. *N. Engl. J. Med.* 373, 163–172. doi: 10.1056/nejmra1409760
- Theoharides, T. C., Stewart, J. M., and Tsilioni, I. (2017). Tolerability and benefit of a tetramethoxyluteolin-containing skin lotion. *Int. J. Immunopathol. Pharmacol.* 30, 146–151. doi: 10.1177/0394632017707610
- Theoharides, T. C., and Tsilioni, I. (2018). Tetramethoxyluteolin for the treatment of neurodegenerative diseases. *Curr. Top. Med. Chem.* 18, 1872–1882. doi: 10.2174/156802661766181119154247
- Theoharides, T. C., Tsilioni, I., and Ren, H. (2019). Recent advances in our understanding of mast cell activation - or should it be mast cell mediator disorders? *Expert. Rev. Clin. Immunol.* 15, 639–656. doi: 10.1080/1744666X.2019.1596800
- Thonhoff, J. R., Simpson, E. P., and Appel, S. H. (2018). Neuroinflammatory mechanisms in amyotrophic lateral sclerosis pathogenesis. *Curr. Opin. Neurol.* 31, 635–639. doi: 10.1097/WCO.0000000000000599
- Torresani, C., Bellafiore, S., and De Panfilis, G. (2009). Chronic urticaria is usually associated with fibromyalgia syndrome. *Acta Derm. Venereol.* 89, 389–392. doi: 10.2340/00015555-0653
- Tsilioni, I., Panagiotidou, S., and Theoharides, T. C. (2014). Exosomes in neurologic and psychiatric disorders. *Clin. Ther.* 36, 882–888. doi: 10.1016/j.clinthera.2014.05.005
- Tsilioni, I., Russell, I. J., Stewart, J. M., Gleason, R. M., and Theoharides, T. C. (2016). Neuropeptides CRH, SP, HK-1, and inflammatory cytokines IL-6 and TNF are increased in serum of patients with fibromyalgia syndrome, implicating mast cells. *J. Pharmacol. Exp. Ther.* 356, 664–672. doi: 10.1124/jpet.115.230060
- Tsilioni, I., and Theoharides, T. C. (2018). Extracellular vesicles are increased in the serum of children with autism spectrum disorder, contain mitochondrial DNA, and stimulate human microglia to secrete IL-1beta. *J. Neuroinflammation* 15:239. doi: 10.1186/s12974-018-1275-5
- Uceyler, N., Hauser, W., and Sommer, C. (2011). Systematic review with meta-analysis: cytokines in fibromyalgia syndrome. *BMC Musculoskelet Disord.* 12:245. doi: 10.1186/1471-2474-12-245
- Vargas, D. L., Nascimbene, C., Krishnan, C., Zimmerman, A. W., and Pardo, C. A. (2005). Neuroglial activation and neuroinflammation in the brain of patients with autism. *Ann. Neurol.* 57, 67–81. doi: 10.1002/ana.20315
- Wallon, C., Yang, P., Keita, A. V., Ericson, A. C., McKay, D. M., Sherman, P. M., et al. (2008). Corticotropin releasing hormone (CRH) regulates macromolecular permeability via mast cells in normal human colonic biopsies in vitro. *Gut* 57, 50–58. doi: 10.1136/gut.2006.117549
- Wang, W., Ji, P., Riopelle, R. J., and Dow, K. E. (2002). Functional expression of corticotropin-releasing hormone (CRH) receptor 1 in cultured rat microglia. *J. Neurochem.* 80, 287–294. doi: 10.1046/j.jnc.2001.00687.x
- Wolfe, F., and Walitt, B. (2013). Culture, science and the changing nature of fibromyalgia. *Nat. Rev. Rheumatol.* 9, 751–755. doi: 10.1038/nrrheum.2013.96
- Woodman, I. (2013). Fibromyalgia: fibromyalgia-all in the brain? *Nat. Rev. Rheumatol.* 9:565. doi: 10.1038/nrrheum.2013.137
- Woolf, C. J. (2011). Central sensitization: implications for the diagnosis and treatment of pain. *Pain* 152, S2–S15. doi: 10.1016/j.pain.2010.09.030
- Wu, Y., Dissing-Olesen, L., MacVicar, B. A., and Stevens, B. (2015). Microglia: dynamic mediators of synapse development and plasticity. *Trends Immunol.* 36, 605–613. doi: 10.1016/j.it.2015.08.008
- Xanthos, D. N., Gaderer, S., Drdla, R., Nuro, E., Abramova, A., Ellmeier, W., et al. (2011). Central nervous system mast cells in peripheral inflammatory nociception. *Mol. Pain* 7:42. doi: 10.1186/1744-8069-7-42
- Yunus, M. B. (2007). Fibromyalgia and overlapping disorders: the unifying concept of central sensitivity syndromes. *Semin. Arthritis Rheum.* 36, 339–356. doi: 10.1016/j.semarthrit.2006.12.009
- Zhang, B., Asadi, S., Weng, Z., Sismanopoulos, N., and Theoharides, T. C. (2012). Stimulated human mast cells secrete mitochondrial components that have autocrine and paracrine inflammatory actions. *PLoS One* 7:e49767. doi: 10.1371/journal.pone.0049767
- Zhang, S., Zeng, X., Yang, H., Hu, G., and He, S. (2012). Mast cell tryptase induces microglia activation via protease-activated receptor 2 signaling. *Cell Physiol. Biochem.* 29, 931–940. doi: 10.1159/000171029
- Zhang, Q., Raof, M., Chen, Y., Sumi, Y., Sursal, T., Junger, W., et al. (2010). Circulating mitochondrial DAMPs cause inflammatory responses to injury. *Nature* 464, 104–107. doi: 10.1038/nature08780
- Zhang, X., Wang, Y., Dong, H., Xu, Y., and Zhang, S. (2016). Induction of microglial activation by mediators released from mast cells. *Cell Physiol. Biochem.* 38, 1520–1531. doi: 10.1159/000443093
- Zhang, Z., Cherryholmes, G., Mao, A., Marek, C., Longmate, J., Kalos, M., et al. (2008). High plasma levels of MCP-1 and eotaxin provide evidence for an immunological basis of fibromyalgia. *Exp. Biol. Med.* 233, 1171–1180. doi: 10.3181/0712-RM-328
- Conflict of Interest Statement:** TT is the inventor of US patents No. 7,906,153 and No. 8,268,365 for the treatment of neuroinflammatory conditions.
- The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.
- Copyright © 2019 Theoharides, Tsilioni and Bawazeer. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.