



Berberine Ameliorates Prenatal Dihydrotestosterone Exposure-Induced Autism-Like Behavior by Suppression of Androgen Receptor

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Many epidemiology studies have shown that maternal polycystic ovary syndrome (PCOS) results in a greater risk of autism spectrum disorders (ASD) development, although the detailed mechanism remains unclear. In this study, we aimed to investigate the potential mechanism and provide a possible treatment for PCOS-mediated ASD through three experiments: Experiment 1: real-time PCR and western blots were employed to measure gene expression in human neurons, and the luciferase reporter assay and chromatin immunoprecipitation (ChIP) was used to map the responsive elements on related gene promoters. Experiment 2: pregnant dams were prenatally exposed to dihydrotestosterone (DHT), androgen receptor (AR) knockdown (shAR) in the amygdala, or berberine (BBR), and the subsequent male offspring were used for autism-like behavior (ALB) assay followed by biomedical analysis, including gene expression, oxidative stress, and mitochondrial function. Experiment 3: the male offspring from prenatal DHT exposed dams were postnatally treated by either shAR or BBR, and the offspring were used for ALB assay followed by biomedical analysis. Our findings showed that DHT treatment suppresses the expression of estrogen receptor β (ER β) and superoxide dismutase 2 (SOD2) through AR-mediated hypermethylation on the ER β promoter, and BBR treatment suppresses AR expression through hypermethylation on the AR promoter. Prenatal DHT treatment induces ER β suppression, oxidative stress and mitochondria dysfunction in the amygdala with subsequent ALB behavior in male offspring, and AR knockdown partly diminishes this effect. Furthermore, both prenatal and postnatal treatment of BBR partly restores prenatal DHT exposure-mediated ALB. In conclusion, DHT suppresses ER β expression through the AR signaling pathway by hypermethylation on the ER β promoter, and BBR restores this effect through AR suppression. Prenatal DHT exposure induces ALB in offspring through AR-mediated

Abbreviations: ALB, autism-like behavior; AR, androgen receptor; ASD, autism spectrum disorders; BBR, berberine; ChIP, chromatin immunoprecipitation; DHT, dihydrotestosterone; ER β , estrogen receptor β ; O₂⁻, superoxide anions; PCOS, polycystic ovary syndrome; ROS, reactive oxygen species; SOD2, superoxide dismutase 2.

ER β suppression, and both prenatal and postnatal treatment of BBR ameliorates this effect. We conclude that BBR ameliorates prenatal DHT exposure-induced ALB through AR suppression, this study may help elucidate the potential mechanism and identify a potential treatment through using BBR for PCOS-mediated ASD.

Keywords: autism spectrum disorders, berberine, androgen, PCOS, autism-like behavior

BACKGROUND

Autism spectrum disorders (ASD) are a group of neurodevelopmental disorders characterized by impaired social communication and interaction, together with restrictive and repetitive behaviors (Rossignol and Frye, 2012; Bralten et al., 2018; Hoirisch-Clapauch and Nardi, 2019). It has been reported that prenatal androgen levels are associated with ASD, and fetal testosterone predicts sexually-differentiated childhood behavior (Tordjman et al., 1997; Auyeung et al., 2009; Ruta et al., 2011). Furthermore, our recent studies have shown that suppression of estrogen receptor β (ER β) and its target gene is associated with ASD development, while the expression of estrogen receptor α (ER α) has little correlation (Zou et al., 2017; Li et al., 2018; Xie et al., 2018), leading us to suppose that prenatal androgen exposure may induce ASD development through ER β suppression.

Polycystic ovary syndrome (PCOS) is a common endocrine disorder in women characterized by hyperandrogenism and ovarian dysfunction (Kosidou et al., 2017). Many epidemiology studies have shown that maternal PCOS is associated with a greater risk of ASD (Kosidou et al., 2016; Cherskov et al., 2018; Katsigianni et al., 2019); furthermore, prenatal androgen exposure may affect the neurodevelopment of offspring, resulting in ASD and attention-deficit/hyperactivity disorder (ADHD; Baron-Cohen et al., 2011; Berni et al., 2018; Cesta et al., 2020), while the detailed mechanism for PCOS-mediated ASD remains unclear.

Recently, the rodent PCOS model with chronic androgen treatment displays many reproductive and metabolic characteristics that are similar to that of human PCOS and provides many advantages for PCOS research, including its inexpensive cost, stable genetic background, shorter life span and generation time, et cetera (Ruta et al., 2011; Walters et al., 2012; Noroozadeh et al., 2017; Iwasa et al., 2020). Currently, three androgen-based PCOS models are typically used, including testosterone (T), dihydrotestosterone (DHT) and dehydroepiandrosterone (DHEA), and each of these models has its strengths and weaknesses (Walters et al., 2012), which we considered in the selection of the androgen for our model. DHT is a non-aromatizable androgen that cannot be converted into estradiol so that there is no interference from estrogen (Noroozadeh et al., 2017). In this study, we chose the DHT-induced rat as the PCOS research model.

Berberine (BBR) is a small isoquinoline alkaloid molecule that can be isolated from plants and holds many pharmacological activities, including antimicrobial, antitumoral, and metabolic modulation effects (Cai et al., 2016). It passes through the blood-brain barrier (BBB) and has many positive effects against some

neurodegenerative diseases, including Alzheimer's disease (AD) and Parkinson's disease (PD; Jiang et al., 2015; Cai et al., 2016). Also, BBR has been reported to suppress the androgen receptor (AR) signaling pathway (Li et al., 2011) and prostatic hyperplasia (Youn et al., 2018), and has many potential effects on PCOS (Li et al., 2013; An et al., 2014; Arentz et al., 2014; Wang et al., 2016), although the detailed mechanism remains unclear and there have been no reports on the effect of BBR on ASD development.

Here, we aim to investigate the mechanism and effect of BBR on PCOS-mediated ASD. The *in vitro* cell culture study shows that DHT (Berkel et al., 2018) suppresses expression of ER β and its target gene superoxide dismutase 2 (SOD2; Liu et al., 2014) in human neurons through AR expression, and BBR restores this effect through AR suppression. Further investigation shows that DHT/BBR-mediated gene suppression is due to hypermethylation on the related gene promoters. We further established an *in vivo* PCOS rat model by DHT administration (Mannerås et al., 2007; Paixão et al., 2017) and found that maternal PCOS induces autism-like behavior (ALB) in offspring through ER β /SOD2 suppression and subsequent oxidative stress and mitochondrial dysfunction. Prenatal treatment of AR knockdown restores this effect, indicating that prenatal DHT exposure induces ALB through the AR signaling pathway; interestingly, both prenatal and postnatal treatment of BBR partly restores this effect. We conclude that BBR ameliorates prenatal DHT exposure-induced ALB by AR suppression. Although it is important to note that the DHT-induced rat model may not completely mimic the PCOS situation in humans, these findings may indicate a potential mechanism and treatment for PCOS-mediated ASD, paving the way for a new avenue of research in this field.

MATERIALS AND METHODS

A detailed description can be found in **Supplementary Data S1**, and the primers used in this study are shown in **Supplementary Table S1**. The human neural progenitor cell ACS-5003 was obtained from ATCC and cultured in NPC medium with supplements and antibiotics. In some experiments, the cells were conditionally immortalized using an hTERT lentivirus vector with an extended life span to achieve higher transfection efficiency and experimental stability (Bodnar et al., 1998; Kong et al., 2016).

Construction of Human AR/ER β Reporter Plasmid

To construct AR/ER β reporter plasmids, the AR/ER β gene promoters (2 kb upstream of the transcription start site plus first exon) were amplified from Ensembl gene ID:

ARLNC1-205 ENST00000650661.1 (for AR) and ESR2-201 ENST00000267525.10 (for ER β) by PCR from human genomic DNA and subcloned into the pGL3-basic vector (#E1751, Promega) using restriction sites of Mlu I and Hind III with the following primers: AR forward: 5'-gctc-acgcgt- cga ggt tag gag ata aag acc -3' (Mlu I) and AR reverse: 5'- gtac- aagctt- ctt ttc tgt aca tct cta gat -3' (Hind III); ER β forward: 5'-gctc-acgcgt-attcaagacgagcctggcca -3' (Mlu I) and ER β reverse: 5'- gtac-aagctt- ctg tt tca ggt aag gtg tgt -3' (Hind III). To map AR or ER β promoter activity, the related deletion promoter constructs were generated by PCR methods and subcloned into the pGL3-basic vector (Zhang et al., 2017).

Preparation of Human AR Expression Lentivirus

The cDNA for human AR (obtained from Open Biosystems) was subcloned into the pLVX-Puro vector (from Clontech) with the restriction sites of XhoI and XbaI using the below primers: human AR forward primer: 5'- gtac-ctcag- atg gaa gtg cag tta ggg ctg -3' (XhoI) and human AR reverse primer: 5'- gtac-tctaga-tca ctg ggt gtg gaa ata gat -3' (XbaI). The lentivirus for either AR or empty control (CTL) was expressed by Lenti-XTM Lentiviral Expression Systems (from Clontech).

Preparation of shAR Knockdown Lentivirus

The shRNA lentivirus plasmid for human AR (sc-29204-SH), or non-target control (sc-108060) were purchased from Santa Cruz Biotechnology, and the shRNA lentivirus plasmid for rat AR was a kind gift from Dr. Haimou Zhang (from Hubei University). The related lentivirus for shAR or empty control (CTL) were expressed through Lenti-XTM Lentiviral Expression Systems (from Clontech) per manufacturers' instructions. The purified and condensed lentivirus was used for *in vivo* gene knockdown. The knockdown efficiency was confirmed by more than 65% of mRNA reduction compared to the control group in rat amygdala cells using real-time PCR (see **Supplementary Table S1**).

Luciferase Reporter Assay

1.0×10^5 of treated cells were seeded in a 6-well plate with complete medium to grow until they reached 80% confluence. Cells were then cotransfected by 3 μ g of reporter constructs, together with 0.2 μ g of pRL-CMV-Luc *Renilla* plasmid (from Promega). After treatment, the cells were harvested and the luciferase activity assays were carried out using the Dual-LuciferaseTM Assay System (Promega), and the transfection efficiencies were normalized using a cotransfected *Renilla* plasmid according to manufacturers' instructions. The reporter activities for AR and ER β were calculated (Zhang et al., 2017).

Chromatin Immunoprecipitation (ChIP)

Cells were washed and cross-linked using 1% formaldehyde for 20 min and terminated by 0.1 M glycine. Cell lysates were sonicated and centrifuged. Five-hundred micrograms of protein were pre-cleared by BSA/salmon sperm DNA with preimmune IgG and a slurry of Protein A Agarose beads. Immunoprecipitations were performed with the indicated antibodies, BSA/salmon sperm DNA and a 50% slurry of

Protein A agarose beads. Input and immunoprecipitates were washed and eluted, then incubated with 0.2 mg/ml Proteinase K for 2 h at 42°C, followed by 6 h at 65°C to reverse the formaldehyde crosslinking. DNA fragments were recovered through phenol/chloroform extraction and ethanol precipitation. A -150 bp fragment in the range of -300~0 from the transcription start site on the AR/ER β promoter was amplified by quantitative real-time PCR (qPCR) using the primers provided in **Supplementary Table S1** (Zhang et al., 2017; Zou et al., 2017).

In vivo Rat Experiments

Sprague-Dawley rats were obtained from Guangdong Medical Animal Center, and maintained under standard 12 h light/dark cycles and given *ad libitum* access to food and water.

Rat Protocol 1: Prenatal Treatment

The 2-month female rats were anesthetized and received treatments consisting of 60-day release pellets that were implanted on the dorsal neck. Hormone pellets contained 5 mg of either (DHT, #A-161), or vehicle pellets (CTL) containing the same matrix but with no hormone (Moran et al., 2007). Furthermore, the rats were implanted with a guide cannula targeting the amygdala (26 gauge; Plastics One; Neal-Perry et al., 2014; Hu et al., 2015). An osmotic minipump (Alzet model 2002; flow rate 0.5 μ l/h; Cupertino, CA, USA) connected to a 26-gauge internal cannula that extended 1 mm below the guide was implanted and used to deliver AR knockdown (shAR) or empty (EMP) lentivirus. A vehicle consisting of artificial cerebrospinal fluid (aCSF) was used for the infusion of the lentivirus. Infusion (flow rate 0.5 μ l/h) begun immediately after the placement of the minipump. 0.5 μ l of total 2×10^3 cfu of lentivirus was infused for 1 h. After 1 week of surgery recovery, the female rats were mated with proven male rats, and the successful pregnancy was confirmed by examining the vaginal plugs. The dams were then treated with vehicle (DMSO, 1 ml/kg) or BBR (10 mg/kg, dissolved in DMSO) by intraperitoneal (i.p.) injection every 2 days starting from day 1. The experimental rats were separated into 4 groups (10 per group). Group 1: CTL rats with empty control lentivirus infusion (CTL); Group 2: DHT rats with empty control lentivirus infusion (DHT); Group 3: DHT rats with shAR knockdown lentivirus infusion (DHT/shAR); Group 4: DHT rats with BBR injection (DHT/BBR). The amygdala neurons were isolated on embryonic day 18 (E18) as described below. After birth, the male offspring were separated from the dams on day 21 and fed until 7–8 weeks old for further experiments. The offspring were then used for ALB tests followed with biomedical analysis (Zou et al., 2017).

Rat Protocol 2: Postnatal Treatment

The 2-month female rats were prenatally exposed to either CTL or DHT during their 21-day pregnancy period as described by [Rat Protocol 1](#). Two-week old male offspring from the above dams were infused with either AR knockdown (shAR) or empty (EMP) lentivirus or treated with vehicle (VEH, 1 ml/kg of DMSO) or BBR (10 mg/kg, dissolved in DMSO) by intraperitoneal (i.p.) injection every 2 days as mentioned in [Rat Protocol 1](#). The rats were separated

into the below four groups (10 per group). Group 1: CTL rats with empty control lentivirus infusion (CTL); Group 2: DHT rats with empty control lentivirus infusion (DHT); Group 3: DHT rats with postnatal shAR knockdown lentivirus infusion (DHT/p-shAR); Group 4: DHT rats with postnatal BBR injection (DHT/p-BBR). After 2 weeks of lentivirus and BBR treatment, the offspring were then used for behavior tests and biomedical analysis, as discussed in [Rat Protocol 1](#) (Zou et al., 2017).

Animal Behavior Test

The animal behavior test of offspring was carried out at 7–8 weeks of age. ALB was evaluated using ultrasonic vocalizations (Silverman et al., 2010; Schaafsma et al., 2017), social recognition tests (Moy et al., 2004; Schaafsma et al., 2017) and a three-chambered social test (Moy et al., 2004; Silverman et al., 2010; Schaafsma et al., 2017; Choi et al., 2019; Wang et al., 2019).

Immunostaining

The isolated amygdala neurons were transferred to coverslips for incubation under growth conditions before being washed with PBS, fixed in 4% paraformaldehyde for 20 min, and incubated with 0.3% Triton X-100 in PBS for 15 min. After blocking with normal goat serum, sections with 8-oxo-dG anti-mouse antibody (1:100, #4354-MC-050, from Novus Biologicals) were incubated for 12 h at 4°C and subsequently with secondary antibody Alexa Fluor 488. The coverslips were then mounted by antifade Mountant with DAPI (staining nuclei, in blue). The photographs were taken using a Confocal Laser Microscope (Leica, 20× lens) and quantitated using ImageJ software.

RESULTS

BBR Restores DHT-Mediated ERβ/SOD2 Suppression by AR Suppression in Human Neurons

We evaluated the potential effect of DHT and BBR on the gene expression of AR, SOD2, and ERβ. The human ACS-5003 neurons were treated by either control (CTL), DHT (10 nM), or DHT with AR knockdown by lentivirus (DHT/shAR), or CTL with AR overexpression by lentivirus (CTL/↑AR), CTL with 10 μM BBR (CTL/BBR), or DHT with BBR (DHT/BBR) for 24 h, then cells were harvested for gene expression analysis. Our results showed that DHT treatment decreased expression of SOD2 and ERβ to 66% and 51%, respectively compared to the CTL group, while it did not have any effect on AR expression. We then evaluated the effect of shAR on DHT treatment (DHT/shAR). Our results showed that shAR decreased AR expression to 21%, indicating a successful AR gene manipulation by lentivirus knockdown; very interestingly, shAR completely restored DHT-mediated gene suppression of SOD2 and ERβ. We also measured the effect of AR overexpression (CTL/↑AR) and found that CTL/↑AR increased AR expression to 235% compared to CTL group, indicating a successful gene manipulation by lentivirus overexpression; furthermore, it decreased expression of SOD2 and ERβ to

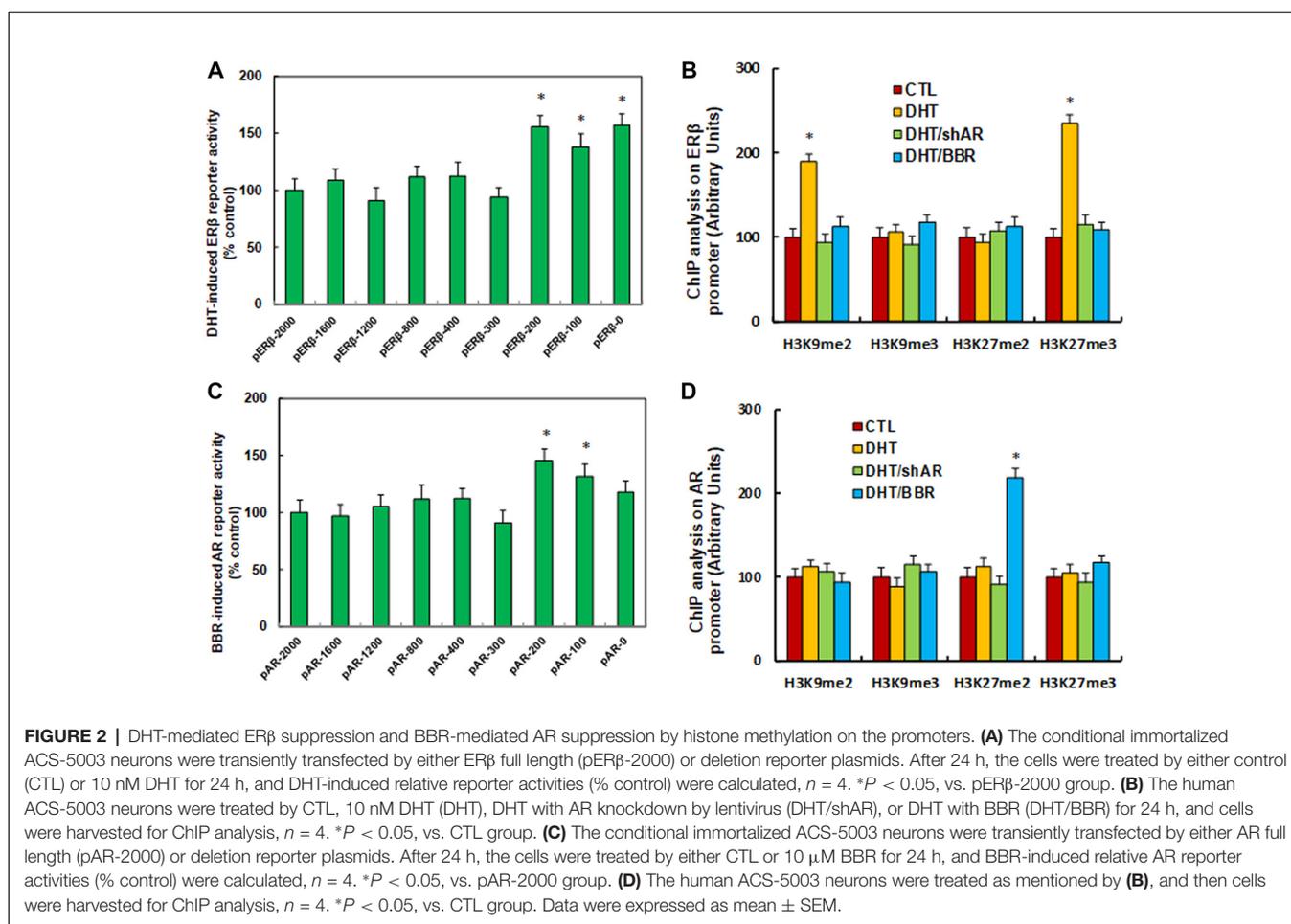
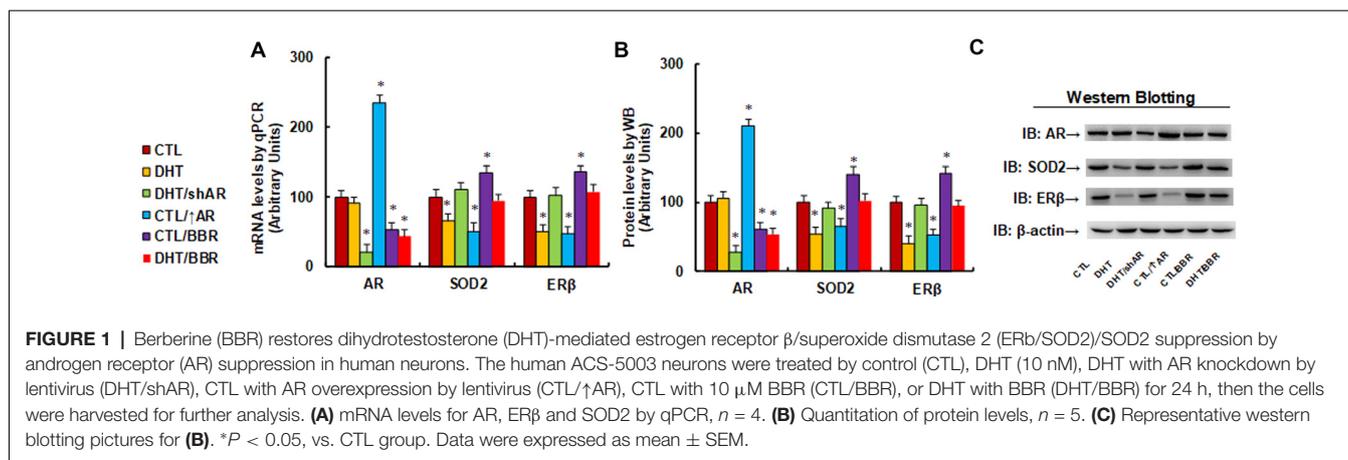
51% and 47%, respectively. We finally measured the effect of BBR (CTL/BBR), and the results showed that CTL/BBR and DHT/BBR decreased AR expression to 53% and 44%, respectively, and CTL/BBR increased expression of SOD2 and ERβ to 135% and 136%, respectively, compared to the CTL group. Our results indicate that DHT suppresses the expression of SOD2 and ERβ, and AR knockdown restores, while AR expression mimics, this effect; BBR restores this effect by AR suppression (see [Figure 1A](#)). We then measured the protein levels for these genes, and an expression pattern similar to that of the mRNA levels was observed (see [Figures 1B,C](#), [Supplementary Figure S1A](#)). We conclude that BBR restores DHT-mediated ERβ/SOD2 suppression by AR suppression.

DHT-Mediated ERβ Suppression Is Due to DHT-Induced Histone Methylation on the ERβ Promoter

We investigated the potential molecular mechanism for DHT-induced ERβ suppression. ERβ promoter deletion constructs were generated and transfected into conditional immortalized human neurons for ERβ reporter activity assay in the presence of either control (CTL) or 10 nM DHT for 24 h, and the DHT-induced relative reporter activities (% control) were calculated. We found that DHT-induced ERβ reporter suppression occurred among the −2,000, −1,600, −1,200, −800, −400, −300 deletion constructs according to Ensembl gene ID: ESR2-201 ENST00000267525.10; the suppression was restored in the −200, −100, and −0 deletion constructs, suggesting that DHT-responsive transcriptional element is located in the range of −300 to 0 on the ERβ promoter (see [Figure 2A](#)). We then measured the possible epigenetic changes in the area of −300 to 0 on ERβ promoter. The ChIP analysis showed that there was no significant difference in either histone H3 (K9, K14, K18, K23, K27) or H4 (K5, K8, K12, K16) acetylation (see [Supplementary Figure S2A](#)). Also, DHT does not affect histone H4 methylation (see [Supplementary Figure S2B](#)). Finally, DHT treatment showed no effect on the methylation of H3K9me3 and H3K27me2, while it increased H3K9me2 and H3K27me3 to 189% and 235%, respectively, compared to the CTL group; both DHT/shAR and DHT/BBR group completely restored this effect (see [Figure 2B](#)). This indicates that DHT-mediated ERβ suppression is due to DHT-induced histone methylation on the ERβ promoter through AR.

BBR-Mediated AR Suppression Is Due to BBR-Induced Histone Methylation on the AR Promoter

We investigated the possible mechanism for BBR-induced AR suppression. AR promoter deletion constructs were generated and transfected into conditional immortalized human neurons for AR BBR reporter activity assay in the presence of either control (CTL) or 10 μM BBR for 24 h, and the BBR-induced relative reporter activities (% control) were calculated. We found that BBR-induced AR reporter suppression occurred among the −2,000, −1,600, −1,200, −800, −400, −300 deletion



constructs according to Ensembl gene ID: ARLNC1-205 ENST00000650661.1; the suppression was restored in the -200 , -100 , and -0 deletion reporter constructs, suggesting that BBR-responsive transcriptional element is located in the area of -300 to 0 on the AR promoter (see **Figure 2C**). We then measured epigenetic changes in this area. Our results showed that BBR did not affect either histone H3 or

H4 acetylation (see **Supplementary Figure S3A**), as well as histone H4 methylation (see **Supplementary Figure S3B**). We finally measured the effect of BBR on histone H3 methylation (see **Figure 2D**). We found that both DHT and DHT/shAR groups had no effect, and BBR treatment had no effect on the methylation of H3K9me2, H3K9me3, and H3K27me3, while it increased H3K27me2 to 219% compared to CTL

group. We conclude that BBR-mediated AR suppression is due to BBR-induced histone H3K27me2 methylation on the AR promoter.

Prenatal Treatment of Either shAR or BBR Ameliorates Prenatal DHT Exposure-Induced Oxidative Stress and Mitochondrial Dysfunction by AR Suppression

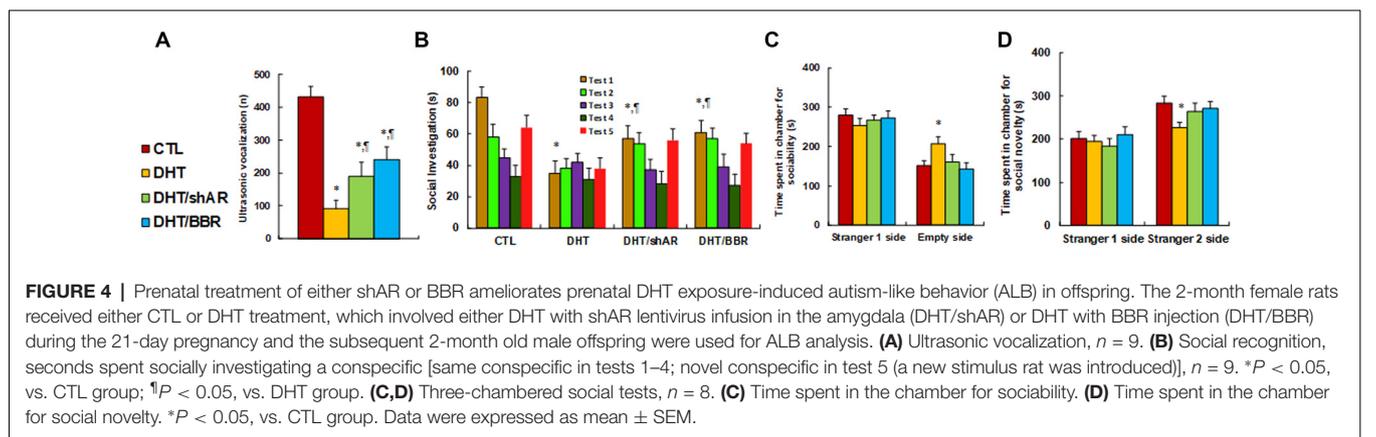
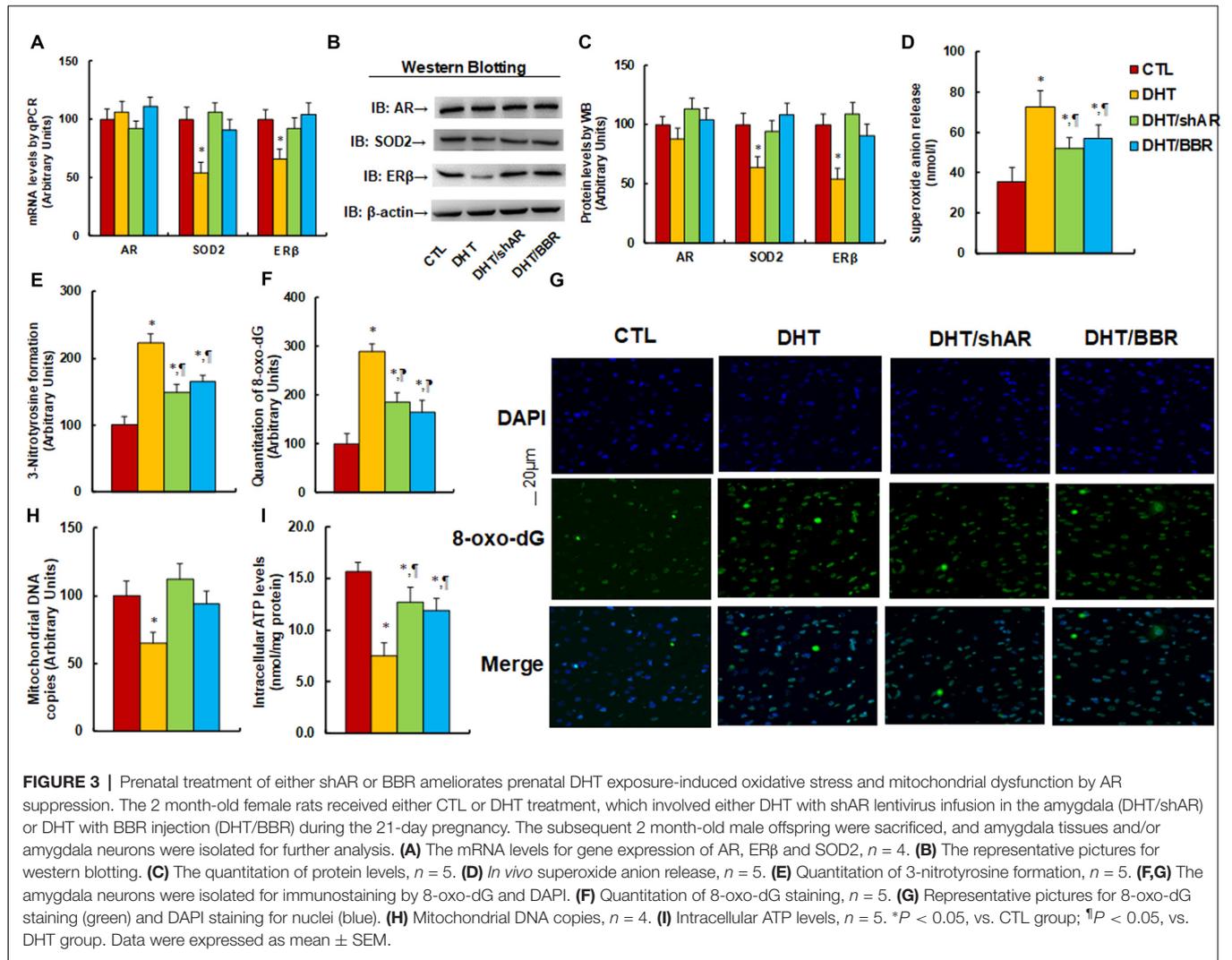
The 2 month-old female rats received either control (CTL) or DHT treatment, which involved either DHT with shAR lentivirus infusion in the amygdala (DHT/shAR) or DHT with BBR injection (DHT/BBR) during the 21-day pregnancy. The subsequent 2-month old male offspring were sacrificed and the brain tissues were isolated for further analysis. We first evaluated the mRNA expression of AR, SOD2, and ER β in the amygdala. The results showed that prenatal DHT exposure (DHT) decreased expression of SOD2 and ER β to 54% and 66%, respectively, compared to the CTL group; none of the prenatal treatments had any effect on AR expression, while both the DHT/shAR and DHT/BBR treatments completely restored the DHT-mediated suppression of SOD2 and ER β (see **Figure 3A**). We then measured the protein levels for those genes, and an expression pattern similar to that of the mRNA levels was observed (see **Figures 3B,C, Supplementary Figure S1B**). We then measured oxidative stress, and found that the DHT group showed increased O $_2^{\cdot-}$ release (see **Figure 3D**) and 3-nitrotyrosine (3-NT) formation (see **Figure 3E**) to 202% and 221%, respectively, compared to the CTL group. Furthermore, both the DHT/shAR and DHT/BBR groups partly restored DHT-mediated oxidative stress. We also measured 8-oxo-dG formation using immunohistochemistry (IHC) techniques as another *in vivo* oxidative stress biomarker (see **Figures 3E,G**). The results showed that the DHT group increased 8-oxo-dG formation to 289%, while DHT/shAR and DHT/BBR group increased 8-oxo-dG formation to 185% and 164%, respectively, partly restoring the DHT-mediated effect. Finally, we evaluated mitochondrial function. The results showed that DHT decreased mitochondrial DNA copies (see **Figure 3H**) and intracellular ATP levels (see **Figure 3I**) to 65% and 48%, respectively, compared to the CTL group, while the DHT/shAR and DHT/BBR groups completely restored DHT-induced suppressed mitochondrial replication and partly restored DHT-induced suppressed intracellular ATP levels. We also measured the potential effect of prenatal treatments on gene expression of the hypothalamus and hippocampus. Our results showed that in the DHT group, AR mRNA in the hypothalamus decreased to 73% (see **Supplementary Figure S4A**), while AR mRNA in the hippocampus decreased to 71% in the DHT/BBR group (see **Supplementary Figure S4B**) compared to CTL group. There was no effect on the expression of SOD2 and ER β . Our results suggest that prenatal BBR treatment ameliorates prenatal DHT exposure-induced oxidative stress and mitochondrial dysfunction by AR suppression.

Prenatal Treatment of Either shAR or BBR Ameliorates Prenatal DHT Exposure-Induced Autism-Like Behavior (ALB) in Offspring

We measured the effect of prenatal DHT exposure on ALBs with the prenatal treatment of either shAR or BBR. The results showed that ultrasonic vocalization frequency decreased to 21% in the DHT group compared to the CTL group, while both DHT/shAR and DHT/BBR group partly restored this effect (see **Figure 4A**). Also, the social recognition showed significant differences for those 4 treatments ($F_{(3,32)} = 2.741$, $P = 0.041$). The *Post hoc* analysis showed that habituation to the same stimulus conspecific (tests 1–4) was significant in CTL group ($F_{(3,32)} = 4.105$, $P < 0.01$), DHT/shAR group ($F_{(3,32)} = 3.067$, $P = 0.044$), and DHT/BBR group ($F_{(3,32)} = 3.279$, $P = 0.036$), but not in DHT group. Dishabituation was significant in CTL group ($F_{(1,8)} = 4.191$, $P < 0.01$), DHT/shAR group ($F_{(1,8)} = 3.611$, $P = 0.024$), and DHT/BBR group ($F_{(1,8)} = 3.816$, $P = 0.019$), but not significant in DHT group (**Figure 4B**). Also, the three-chambered social tests showed that DHT group increased time spent in the empty side of the chamber for sociability (see **Figure 4C**) to 137%, and decreased time spent in the Stranger 2 side of the chamber for social novelty (see **Figure 4D**) to 80%, respectively; while both DHT/shAR and DHT/BBR group completely restored this effect. Additionally, there was no difference in the Stranger 1 side in terms of either sociability or social novelty. The test animal spent most of its time on the Stranger side, with around 300 s for all different treatments (see **Figure 4C**). A statistically significant difference between the CTL and DHT groups could not be distinguished in evaluating time spent in the Stranger side, while there was a significant difference in time spent on the Empty side, indicating that DHT treatment induces social impairment. Furthermore, we measured the patterns of social behavior in male offspring, finding that DHT treatment decreased time spent Sniffing to 59%, Mounting to 57%, and 60% in total social interaction, with no difference in Grooming partners (see **Supplementary Figure S5**), and both treatment of the DHT/shAR and DHT/BBR groups completely restored this effect. Our results suggest that prenatal treatment of either shAR or BBR ameliorates prenatal DHT exposure-induced social deficits related to the autistic phenotype in offspring.

Postnatal BBR Treatment Ameliorates Prenatal DHT Exposure-Induced ALB in Offspring, While Postnatal shAR Treatment Does Not Affect

We measured the possible effect of postnatal BBR treatment on prenatal DHT exposure-mediated ALB in offspring. The 2-month female rats received either control (CTL) or DHT treatment during their pregnancy, and the subsequent 2-month-old male offspring from the DHT group received either shAR lentivirus infusion in the amygdala (DHT/p-shAR) or DHT with BBR injection (DHT/p-BBR) for 1 week, and then those male offspring were used for ALB analysis followed by a biomedical analysis of the isolated amygdala. We first evaluated the mRNA



expression of AR, SOD2, and ERβ in the amygdala. The results showed that prenatal DHT exposure (DHT) decreased expression of SOD2 and ERβ to 51% and 54%, respectively, but had no effect on AR expression. Additionally, postnatal shAR

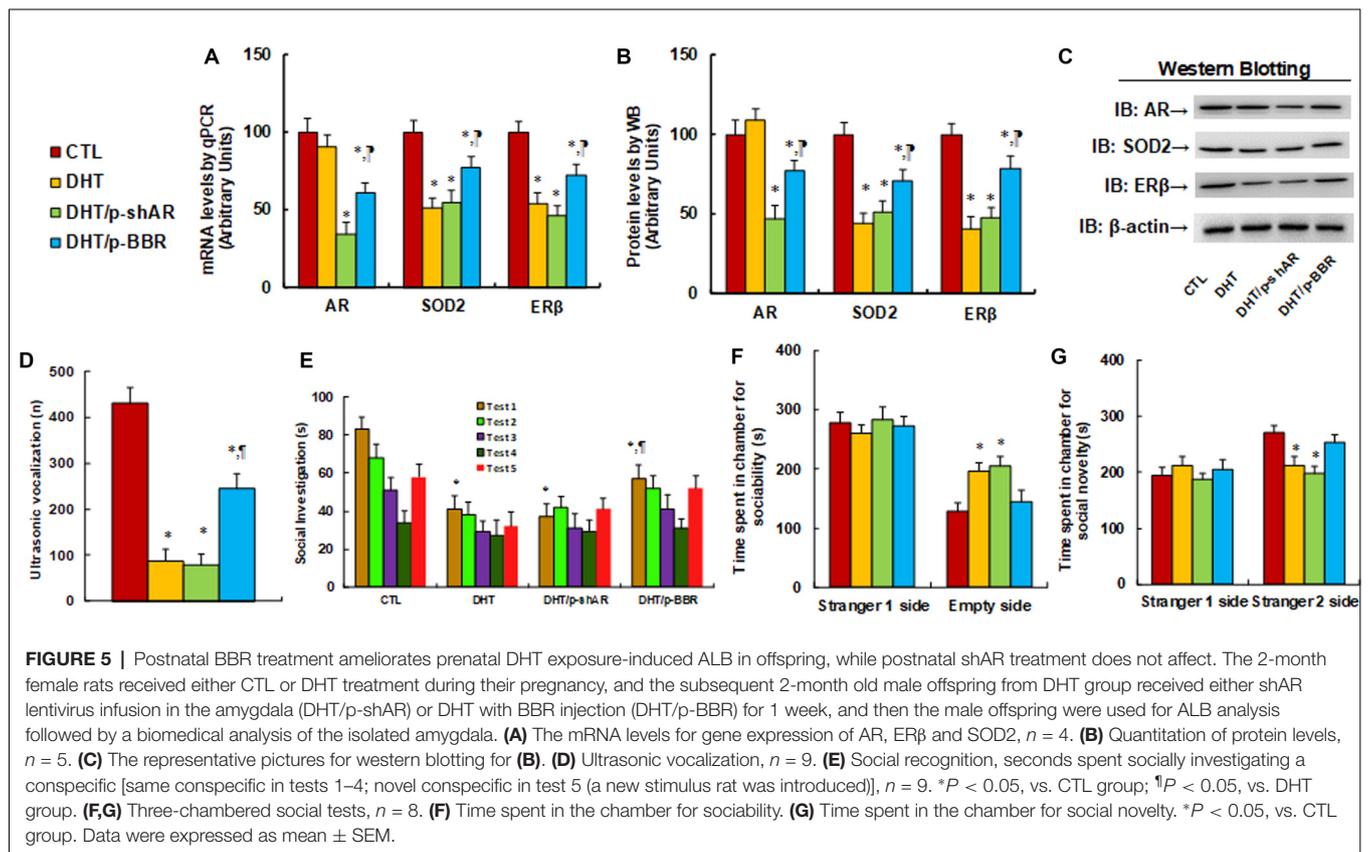
treatment (DHT/p-shAR) decreased AR expression to 34%, indicating a successful gene manipulation by shAR lentivirus knockdown, but did not affect prenatal DHT exposure-mediated suppression of SOD2 and ERβ. On the other hand, postnatal BBR

treatment (DHT/p-BBR) decreased AR expression to 61% and partly restored prenatal DHT exposure-mediated suppression of SOD2 and ER β (see **Figure 5A**). We then measured the protein levels for those genes, and an expression pattern similar to that of the mRNA levels was observed (see **Figures 5B,C**, **Supplementary Figure S1C**). We continued to evaluate the effect of postnatal BBR treatment on ALBs. The ultrasonic vocalization results showed that ultrasonic vocalization frequency decreased to 20% in the DHT group compared to the CTL group, and DHT/p-shAR treatment had no effect, while this effect was partly restored in the DHT/p-BBR group (see **Figure 5D**). Furthermore, the social recognition tests showed a significant difference between 4 treatments ($F_{(3,32)} = 2.947$, $P = 0.046$), and the *Post hoc* analysis showed that habituation to the same stimulus conspecific (tests 1–4) was significant in CTL group ($F_{(3,32)} = 4.264$, $P < 0.01$) and DHT/p-BBR group ($F_{(3,32)} = 3.410$, $P = 0.034$), but not in DHT and DHT/p-shAR groups. Dishabituation was significant in CTL group ($F_{(1,8)} = 4.361$, $P < 0.01$), and borderline significant in DHT/p-BBR group ($F_{(1,8)} = 3.017$, $P = 0.037$), but was not significant in DHT and DHT/p-shAR groups (see **Figure 5E**). Also, three-chambered social tests showed that in DHT group, time spent in the empty side of the chamber for sociability (see **Figure 5F**) increased to 153%, and time spent in the empty side of the chamber for social novelty (see **Figure 5G**) decreased to 79%, respectively; there was no noticeable effect in DHT/p-shAR group, while this effect was completely restored in DHT/p-BBR group. Also, there was no difference in the Stranger 1 side in

terms of either sociability or social novelty. Besides, we measured the repetitive patterns of social behavior in male offspring and found that DHT treatment decreased time spent Sniffing to 57%, Mounting to 45%, and total social interaction to 56%, with no difference in Grooming partners (see **Supplementary Figure S6**). DHT/p-shAR treatment showed no effect, while DHT/p-BBR treatment partly restored this effect. Our results suggest that postnatal BBR treatment ameliorates prenatal DHT exposure-induced ALB in offspring, but postnatal shAR treatment shows no effect. The ineffectiveness of p-shAR treatment can be explained by the hypothesis that even though the treatment can decrease AR expression in offspring, it cannot restore the epigenetic changes on the ER β promoter, while p-BBR treatment can potentially penetrate the BBB and modify DHT-induced epigenetic changes.

DISCUSSION

In this study, the neural progenitor cell ACS-5003 was used to mimic the differentiation of stem cells into neurons during embryo development (Wang et al., 2019). The *in vitro* experiments demonstrate that DHT suppresses ER β expression through AR-mediated hypermethylation on ER β promoter, and BBR restores this effect through AR suppression. Furthermore, the *in vivo* experiments show that prenatal DHT exposure induces ALB in offspring, and both prenatal and postnatal BBR treatment ameliorates this effect.



Prenatal DHT Exposure-Mediated Autism-Like Behavior

We have previously found that male offspring are more sensitive than female offspring in ER β suppression-mediated ALB (Zou et al., 2017). In this study, we chose male offspring as our experimental target. Here, we report that prenatal androgen exposure may also be a potential risk for ASD development. PCOS is characterized by hyperandrogenism (Kosidou et al., 2017), and many epidemiological studies demonstrated that PCOS is associated with ASD development (Kosidou et al., 2016; Cherskov et al., 2018; Katsigianni et al., 2019). Thus, we hypothesize that prenatal androgen exposure may trigger ASD development. The DHT treatment was used to mimic PCOS conditions during the *in vivo* rat experiments (Mannerås et al., 2007; Noroozadeh et al., 2017; Paixão et al., 2017). Our results show that prenatal DHT exposure induces ALB in offspring with suppressed expression of ER β and SOD2 in the amygdala (Zou et al., 2017; Li et al., 2018; Xie et al., 2018). AR knockdown partly diminishes this effect, indicating that DHT induces ALB through the AR signaling pathway. Also, it has been reported that the sex hormone regulates the expression of FOXP1 (Takayama et al., 2014; Fröhlich et al., 2017), SHANK (Berkel et al., 2018), and RORA (Sarachana et al., 2011), which is essential for ASD, further confirming the potential role of the AR signaling pathway on ASD development. This can partly explain why the androgen can be related to male/female bias to ASD (Becker, 2012; Schaafsma and Pfaff, 2014; Mogi et al., 2015). This is the first time that the potential mechanism for PCOS-mediated ASD development has been discovered through AR-mediated ER β suppression, providing a potential target for PCOS-mediated ASD through AR suppression (Baron-Cohen et al., 2011; Lee et al., 2017; Cherskov et al., 2018; Shay et al., 2018).

Prenatal-DHT Exposure-Mediated Gene Expression in Autistic Offspring

We have previously found that prenatal progesterin exposure suppresses ER β expression in the amygdala instead of in other brain areas and subsequently contributes to ASD development in offspring, with male offspring being more susceptible than females. Additionally, ER β overexpression in the amygdala significantly restores prenatal progesterin exposure-induced ALB (Zou et al., 2017; Xie et al., 2018), and maternal diabetes-induced autistic offspring show significant SOD2 suppression in the amygdala as well (Wang et al., 2019). Our results indicate that gene expression in the amygdala plays an important role in ASD development. In this study, we supposed that the amygdala should be the most sensitive organ in prenatal DHT exposure-induced autistic offspring; thus, we conducted the related manipulation and analysis of the amygdala in male offspring. Furthermore, our data showed that prenatal DHT treatment affects gene expression of AR, SOD2, and ER β in the amygdala (see **Figure 5A**) and also suppresses AR expression in the hypothalamus and hippocampus, but has no effect on ER β and SOD2 (see **Supplementary Figure S4**), although the possible reason for this still requires further investigation. Besides, we found that prenatal DHT exposure induces SOD2 suppression

in offspring, which is consistent with previous findings that either maternal diabetes (Wang et al., 2019) or prenatal exposure of progesterin (Zou et al., 2017; Li et al., 2018) induces SOD2 suppression and oxidative stress in neurons. Treatment of antioxidants such as resveratrol (Xie et al., 2018) partly restores this effect, indicating that oxidative stress is associated with the ASD phenotype, which is consistent with findings from recent literature (Mandic-Maravic et al., 2019; Bjørklund et al., 2020).

Role of BBR on PCOS-Mediated ASD

BBR is one of the herbal medicine that has been employed for the management of neurodegenerative diseases such as AD and PD due to its multi-faceted defensive capabilities, such as its protective role in atherosclerosis-related to lipid- and glucose-lowering properties (Jiang et al., 2015; Cai et al., 2016). Also, BBR has been reported to inhibit the AR signaling pathway (Li et al., 2011; Tian et al., 2016; Youn et al., 2018), while the related mechanism remains unclear. Besides, BBR has been used for the treatment of PCOS to improve pregnancy outcomes through its effect on insulin resistance (Li et al., 2013; Chen et al., 2017) and normalizing the endocrine and metabolic parameters in PCOS women (An et al., 2014; Arentz et al., 2014; Wang et al., 2016), while there have been no reports on the effect of BBR on PCOS-mediated ASD development. In this study, we report that BBR restores DHT-mediated ER β suppression by AR suppression and subsequently ameliorates prenatal DHT exposure-mediated ALB in offspring. Both prenatal and postnatal treatment of BBR protects from prenatal DHT exposure-induced ALB in offspring, indicating that BBR may be a potential candidate for the treatment of PCOS-induced ASD. We have previously found that prenatal progesterin exposure and maternal diabetes contribute to ASD development through ER β suppression, and ER β overexpression in the brain restores this effect, indicating that ER β suppression may contribute to ASD development (Zou et al., 2017; Li et al., 2018; Xie et al., 2018; Wang et al., 2019). On the other hand, many potential prenatal factors may suppress ER β expression, and DHT is only one of them. In this case, berberine treatment can only be active for androgen or PCOS-mediated ASD, and may not work efficiently on treating ASD mediated by other factors.

CONCLUSION

Prenatal DHT exposure induces ALB in offspring through the AR signaling pathway; both prenatal and postnatal treatment of BBR partly restore this effect by suppression of AR expression. We conclude that BBR ameliorates prenatal DHT exposure-induced ALB by AR suppression. This is the first time that the potential mechanism for PCOS-mediated ASD development has been discovered, and BBR may be a potential clinical treatment for PCOS-mediated ASD patients.

DATA AVAILABILITY STATEMENT

All datasets generated for this study are included in the article/**Supplementary Material**.

ETHICS STATEMENT

The animal protocol conformed to US NIH guidelines (Guide for the Care and Use of Laboratory Animals, No. 85-23, revised 1996), and was reviewed and approved by the Institutional Animal Care and Use Committee from Guangzhou University of Chinese Medicine.

AUTHOR CONTRIBUTIONS

PY wrote the article. PY, LL, and XL designed, analyzed the data and interpreted the experiments. XL, YL, and ZW performed the vector constructions and gene expression analysis. XC, YL, ZW, and MW performed the statistical analysis and part of the rat experiments. YW, MX, and ML performed gene analysis and part of the mapping analysis. DX, JL, and CW performed

the remaining experiments. All authors read and approved the final manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fncel.2020.00087/full#supplementary-material>.

REFERENCES

- An, Y., Sun, Z., Zhang, Y., Liu, B., Guan, Y., and Lu, M. (2014). The use of berberine for women with polycystic ovary syndrome undergoing IVF treatment. *Clin. Endocrinol.* 80, 425–431. doi: 10.1111/cen.12294
- Arentz, S., Abbott, J. A., Smith, C. A., and Bensoussan, A. (2014). Herbal medicine for the management of polycystic ovary syndrome (PCOS) and associated oligo/amenorrhoea and hyperandrogenism; a review of the laboratory evidence for effects with corroborative clinical findings. *BMC Complement. Altern. Med.* 14:511. doi: 10.1186/1472-6882-14-511
- Auyeung, B., Baron-Cohen, S., Ashwin, E., Knickmeyer, R., Taylor, K., Hackett, G., et al. (2009). Fetal testosterone predicts sexually differentiated childhood behavior in girls and in boys. *Psychol. Sci.* 20, 144–148. doi: 10.1111/j.1467-9280.2009.02279.x
- Baron-Cohen, S., Lombardo, M. V., Auyeung, B., Ashwin, E., Chakrabarti, B., and Knickmeyer, R. (2011). Why are autism spectrum conditions more prevalent in males? *PLoS Biol.* 9:e1001081. doi: 10.1371/journal.pbio.1001081
- Becker, K. G. (2012). Male gender bias in autism and pediatric autoimmunity. *Autism Res.* 5, 77–83. doi: 10.1002/aur.1227
- Berkel, S., Eltokhi, A., Fröhlich, H., Porras-Gonzalez, D., Rafiullah, R., Sprengel, R., et al. (2018). Sex hormones regulate *SHANK* expression. *Front. Mol. Neurosci.* 11:337. doi: 10.3389/fnmol.2018.00337
- Berni, T. R., Morgan, C. L., Berni, E. R., and Rees, D. A. (2018). Polycystic ovary syndrome is associated with adverse mental health and neurodevelopmental outcomes. *J. Clin. Endocrinol. Metab.* 103, 2116–2125. doi: 10.1210/jc.2017-02667
- Björklund, G., Meguid, N. A., El-Bana, M. A., Tinkov, A. A., Saad, K., Dadar, M., et al. (2020). Oxidative stress in autism spectrum disorder. *Mol. Neurobiol.* doi: 10.1007/s12035-019-01742-2 [Epub ahead of print].
- Bodnar, A. G., Ouellette, M., Frolkis, M., Holt, S. E., Chiu, C. P., Morin, G. B., et al. (1998). Extension of life-span by introduction of telomerase into normal human cells. *Science* 279, 349–352. doi: 10.1126/science.279.5349.349
- Bralten, J., van Hulzen, K. J., Martens, M. B., Galesloot, T. E., Arias Vasquez, A., Kiemeny, L. A., et al. (2018). Autism spectrum disorders and autistic traits share genetics and biology. *Mol. Psychiatry* 23, 1205–1212. doi: 10.1038/mp.2017.98
- Cai, Z., Wang, C., and Yang, W. (2016). Role of berberine in Alzheimer's disease. *Neuropsychiatr. Dis. Treat.* 12, 2509–2520. doi: 10.2147/ndt.s114846
- Cesta, C. E., Öberg, A. S., Ibrahimson, A., Yusuf, I., Larsson, H., Almqvist, C., et al. (2020). Maternal polycystic ovary syndrome and risk of neuropsychiatric disorders in offspring: prenatal androgen exposure or genetic confounding? *Psychol. Med.* 50, 616–624. doi: 10.1017/S0033291719000424
- Chen, Q., Mo, R., Wu, N., Zou, X., Shi, C., Gong, J., et al. (2017). Berberine ameliorates diabetes-associated cognitive decline through modulation of aberrant inflammation response and insulin signaling pathway in DM rats. *Front. Pharmacol.* 8:334. doi: 10.3389/fphar.2017.00334
- Cherskov, A., Pohl, A., Allison, C., Zhang, H., Payne, R. A., and Baron-Cohen, S. (2018). Polycystic ovary syndrome and autism: a test of the prenatal sex steroid theory. *Transl. Psychiatry* 8:136. doi: 10.1038/s41398-018-0186-7
- Choi, Y., Park, H., Kang, S., Jung, H., Kweon, H., Kim, S., et al. (2019). NGL-1/LRRC4C-mutant mice display hyperactivity and anxiolytic-like behavior associated with widespread suppression of neuronal activity. *Front. Mol. Neurosci.* 12:250. doi: 10.3389/fnmol.2019.00250
- Fröhlich, H., Rafiullah, R., Schmitt, N., Abele, S., and Rappold, G. A. (2017). Foxp1 expression is essential for sex-specific murine neonatal ultrasonic vocalization. *Hum. Mol. Genet.* 26, 1511–1521. doi: 10.1093/hmg/ddx055
- Hoirisch-Clapauch, S., and Nardi, A. E. (2019). Autism spectrum disorders: let's talk about glucose? *Transl. Psychiatry* 9:51. doi: 10.1038/s41398-019-0370-4
- Hu, M., Richard, J. E., Maliqueo, M., Kokosar, M., Fornes, R., Benrick, A., et al. (2015). Maternal testosterone exposure increases anxiety-like behavior and impacts the limbic system in the offspring. *Proc. Natl. Acad. Sci. U S A* 112, 14348–14353. doi: 10.1073/pnas.1507514112
- Iwasa, T., Matsuzaki, T., Mayila, Y., Kawakita, T., Yanagihara, R., and Irahara, M. (2020). The effects of chronic oxytocin administration on body weight and food intake in DHT-induced PCOS model rats. *Gynecol. Endocrinol.* 36, 55–60. doi: 10.1080/09513590.2019.1631276
- Jiang, W., Li, S., and Li, X. (2015). Therapeutic potential of berberine against neurodegenerative diseases. *Sci. China Life Sci.* 58, 564–569. doi: 10.1007/s11427-015-4829-0
- Katsigianni, M., Karageorgiou, V., Lambrinou, I., and Siristatidis, C. (2019). Maternal polycystic ovarian syndrome in autism spectrum disorder: a systematic review and meta-analysis. *Mol. Psychiatry* 24, 1787–1797. doi: 10.1038/s41380-019-0398-0
- Kong, D., Zhan, Y., Liu, Z., Ding, T., Li, M., Yu, H., et al. (2016). SIRT1-mediated ER β suppression in the endothelium contributes to vascular aging. *Aging Cell* 15, 1092–1102. doi: 10.1111/ace1.12515
- Kosidou, K., Dalman, C., Widman, L., Arver, S., Lee, B. K., Magnusson, C., et al. (2016). Maternal polycystic ovary syndrome and the risk of autism spectrum disorders in the offspring: a population-based nationwide study in Sweden. *Mol. Psychiatry* 21, 1441–1448. doi: 10.1038/mp.2015.183
- Kosidou, K., Dalman, C., Widman, L., Arver, S., Lee, B. K., Magnusson, C., et al. (2017). Maternal polycystic ovary syndrome and risk for attention-deficit/hyperactivity disorder in the offspring. *Biol. Psychiatry* 82, 651–659. doi: 10.1016/j.biopsych.2016.09.022
- Lee, B. K., Arver, S., Widman, L., Gardner, R. M., Magnusson, C., Dalman, C., et al. (2017). Maternal hirsutism and autism spectrum disorders in offspring. *Autism Res.* 10, 1544–1546. doi: 10.1002/aur.1797
- Li, J., Cao, B., Liu, X., Fu, X., Xiong, Z., Chen, L., et al. (2011). Berberine suppresses androgen receptor signaling in prostate cancer. *Mol. Cancer Ther.* 10, 1346–1356. doi: 10.1158/1535-7163.MCT-10-0985
- Li, L., Li, M., Lu, J., Ge, X., Xie, W., Wang, Z., et al. (2018). Prenatal progesterone exposure is associated with autism spectrum disorders. *Front. Psychiatry* 9:611. doi: 10.3389/fpsy.2018.00611

- Li, Y., Ma, H., Zhang, Y., Kuang, H., Ng, E. H., Hou, L., et al. (2013). Effect of berberine on insulin resistance in women with polycystic ovary syndrome: study protocol for a randomized multicenter controlled trial. *Trials* 14:226. doi: 10.1186/1745-6215-14-226
- Liu, Z., Gou, Y., Zhang, H., Zuo, H., Zhang, H., Liu, Z., et al. (2014). Estradiol improves cardiovascular function through up-regulation of SOD2 on vascular wall. *Redox Biol.* 3, 88–99. doi: 10.1016/j.redox.2014.11.001
- Mandic-Maravic, V., Mitkovic-Voncina, M., Pljesa-Ercegovac, M., Savic-Radojevic, A., Djordjevic, M., Pekmezovic, T., et al. (2019). Autism spectrum disorders and perinatal complications-is oxidative stress the connection? *Front. Psychiatry* 10:675. doi: 10.3389/fpsy.2019.00675
- Mannerås, L., Cajander, S., Holmang, A., Seleskovic, Z., Lystig, T., Lonn, M., et al. (2007). A new rat model exhibiting both ovarian and metabolic characteristics of polycystic ovary syndrome. *Endocrinology* 148, 3781–3791. doi: 10.1210/en.2007-0168
- Mogi, K., Takanashi, H., Nagasawa, M., and Kikusui, T. (2015). Sex differences in spatiotemporal expression of AR, ER α and ER β mRNA in the perinatal mouse brain. *Neurosci. Lett.* 584, 88–92. doi: 10.1016/j.neulet.2014.10.028
- Moran, A. L., Nelson, S. A., Landisch, R. M., Warren, G. L., and Lowe, D. A. (2007). Estradiol replacement reverses ovariectomy-induced muscle contractile and myosin dysfunction in mature female mice. *J. Appl. Physiol.* 102, 1387–1393. doi: 10.1152/jappphysiol.01305.2006
- Moy, S. S., Nadler, J. J., Perez, A., Barbaro, R. P., Johns, J. M., Magnuson, T. R., et al. (2004). Sociability and preference for social novelty in five inbred strains: an approach to assess autistic-like behavior in mice. *Genes Brain Behav.* 3, 287–302. doi: 10.1111/j.1601-1848.2004.00076.x
- Neal-Perry, G., Yao, D., Shu, J., Sun, Y., and Etgen, A. M. (2014). Insulin-like growth factor-I regulates LH release by modulation of kisspeptin and NMDA-mediated neurotransmission in young and middle-aged female rats. *Endocrinology* 155, 1827–1837. doi: 10.1210/en.2013-1682
- Noroozzadeh, M., Behboudi-Gandevani, S., Zadeh-Vakili, A., and Ramezani Tehrani, F. (2017). Hormone-induced rat model of polycystic ovary syndrome: a systematic review. *Life Sci.* 191, 259–272. doi: 10.1016/j.lfs.2017.10.020
- Paixão, L., Ramos, R. B., Lavarda, A., Morsh, D. M., and Spritzer, P. M. (2017). Animal models of hyperandrogenism and ovarian morphology changes as features of polycystic ovary syndrome: a systematic review. *Reprod. Biol. Endocrinol.* 15:12. doi: 10.1186/s12958-017-0231-z
- Rosignol, D. A., and Frye, R. E. (2012). A review of research trends in physiological abnormalities in autism spectrum disorders: immune dysregulation, inflammation, oxidative stress, mitochondrial dysfunction and environmental toxicant exposures. *Mol. Psychiatry* 17, 389–401. doi: 10.1038/mp.2011.165
- Ruta, L., Ingudomnukul, E., Taylor, K., Chakrabarti, B., and Baron-Cohen, S. (2011). Increased serum androstenedione in adults with autism spectrum conditions. *Psychoneuroendocrinology* 36, 1154–1163. doi: 10.1016/j.psyneuen.2011.02.007
- Sarachana, T., Xu, M., Wu, R. C., and Hu, V. W. (2011). Sex hormones in autism: androgens and estrogens differentially and reciprocally regulate RORA, a novel candidate gene for autism. *PLoS One* 6:e17116. doi: 10.1371/journal.pone.0017116
- Schaafsma, S. M., Gagnidze, K., Reyes, A., Norstedt, N., Mansson, K., Francis, K., et al. (2017). Sex-specific gene-environment interactions underlying ASD-like behaviors. *Proc. Natl. Acad. Sci. U S A* 114, 1383–1388. doi: 10.1073/pnas.1619312114
- Schaafsma, S. M., and Pfaff, D. W. (2014). Etiologies underlying sex differences in autism spectrum disorders. *Front. Neuroendocrinol.* 35, 255–271. doi: 10.1016/j.yfrne.2014.03.006
- Shay, D. A., Vieira-Potter, V. J., and Rosenfeld, C. S. (2018). Sexually dimorphic effects of aromatase on neurobehavioral responses. *Front. Mol. Neurosci.* 11:374. doi: 10.3389/fnmol.2018.00374
- Silverman, J. L., Yang, M., Lord, C., and Crawley, J. N. (2010). Behavioural phenotyping assays for mouse models of autism. *Nat. Rev. Neurosci.* 11, 490–502. doi: 10.1038/nrn2851
- Takayama, K., Suzuki, T., Tsutsumi, S., Fujimura, T., Takahashi, S., Homma, Y., et al. (2014). Integrative analysis of FOXP1 function reveals a tumor-suppressive effect in prostate cancer. *Mol. Endocrinol.* 28, 2012–2024. doi: 10.1210/me.2014-1171
- Tian, Y., Zhao, L., Wang, Y., Zhang, H., Xu, D., Zhao, X., et al. (2016). Berberine inhibits androgen synthesis by interaction with aldo-keto reductase 1C3 in 22Rv1 prostate cancer cells. *Asian J. Androl.* 18, 607–612. doi: 10.4103/1008-682x.169997
- Tordjman, S., Ferrari, P., Sulmont, V., Duyme, M., and Roubertoux, P. (1997). Androgenic activity in autism. *Am. J. Psychiatry* 154, 1626–1627. doi: 10.1176/ajp.154.11.1626-a
- Walters, K. A., Allan, C. M., and Handelsman, D. J. (2012). Rodent models for human polycystic ovary syndrome. *Biol. Reprod.* 86:149. doi: 10.1095/biolreprod.111.097808
- Wang, Y., Fu, X., Xu, J., Wang, Q., and Kuang, H. (2016). Systems pharmacology to investigate the interaction of berberine and other drugs in treating polycystic ovary syndrome. *Sci. Rep.* 6:28089. doi: 10.1038/srep28089
- Wang, X., Lu, J., Xie, W., Lu, X., Liang, Y., Li, M., et al. (2019). Maternal diabetes induces autism-like behavior by hyperglycemia-mediated persistent oxidative stress and suppression of superoxide dismutase 2. *Proc. Natl. Acad. Sci. U S A* 116, 23743–23752. doi: 10.1073/pnas.1912625116
- Xie, W., Ge, X., Li, L., Yao, A., Wang, X., Li, M., et al. (2018). Resveratrol ameliorates prenatal progesterone exposure-induced autism-like behavior through ER β activation. *Mol. Autism* 9:43. doi: 10.1186/s13229-018-0225-5
- Youn, D. H., Park, J., Kim, H. L., Jung, Y., Kang, J., Lim, S., et al. (2018). Berberine improves benign prostatic hyperplasia via suppression of 5 α reductase and extracellular signal-regulated kinase *in vivo* and *in vitro*. *Front. Pharmacol.* 9:773. doi: 10.3389/fphar.2018.00773
- Zhang, H., Li, L., Li, M., Huang, X., Xie, W., Xiang, W., et al. (2017). Combination of betulinic acid and chidamide inhibits acute myeloid leukemia by suppression of the HIF1 α pathway and generation of reactive oxygen species. *Oncotarget* 8, 94743–94758. doi: 10.18632/oncotarget.21889
- Zou, Y., Lu, Q., Zheng, D., Chu, Z., Liu, Z., Chen, H., et al. (2017). Prenatal levonorgestrel exposure induces autism-like behavior in offspring through ER β suppression in the amygdala. *Mol. Autism* 8:46. doi: 10.1186/s13229-017-0159-3

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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