Check for updates

OPEN ACCESS

EDITED BY Ming Li, Osaka University, Japan

REVIEWED BY Elena Giusto, San Camillo Hospital (IRCCS), Italy Meiting Chen, Sun Yat-sen University Cancer Center (SYSUCC), China

*CORRESPONDENCE Alireza Afshar ⊠ Alireza.af2017@gmail.com Masoud Zare ⊠ md.zare77@gmail.com

[†]These authors share first authorship

RECEIVED 03 May 2024 ACCEPTED 24 September 2024 PUBLISHED 04 October 2024

CITATION

Salehpour A, Karimi Z, Ghasemi Zadeh M, Afshar M, Kameli A, Mooseli F, Zare M and Afshar A (2024) Therapeutic potential of mesenchymal stem cell-derived exosomes and miRNAs in neuronal regeneration and rejuvenation in neurological disorders: a mini review.

Front. Cell. Neurosci. 18:1427525. doi: 10.3389/fncel.2024.1427525

COPYRIGHT

© 2024 Salehpour, Karimi, Ghasemi Zadeh, Afshar, Kameli, Mooseli, Zare and Afshar. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Therapeutic potential of mesenchymal stem cell-derived exosomes and miRNAs in neuronal regeneration and rejuvenation in neurological disorders: a mini review

Aria Salehpour^{1†}, Zahra Karimi^{1†}, Mokhtar Ghasemi Zadeh^{1,2}, Mohammadreza Afshar^{1,2}, Ali Kameli¹, Fatemeh Mooseli^{1,2}, Masoud Zare^{1*} and Alireza Afshar^{1,2*}

¹The Persian Gulf Marine Biotechnology Research Center, The Persian Gulf Biomedical Sciences Research Institute, Bushehr University of Medical Sciences, Bushehr, Iran, ²Student Research Committee, Bushehr University of Medical Sciences, Bushehr, Iran

Mesenchymal stem cells (MSCs) have gained considerable attention in the field of regenerative medicine due to their ability to secrete small extracellular vesicles (EVs) known as exosomes. This review delves into the various biological activities of MSCs and the cell interactions enabled by these exosomes, with a focus on their potential for neuronal regeneration and the treatment of neurological disorders. We scrutinize findings from multiple studies that underscore the neuroprotective and neuro-regenerative effects of exosomes derived from MSCs, illuminating their mechanisms of action and therapeutic applications. This review thoroughly investigates all related pathways, miRNAs, and factors to suggest potential strategies for enhancing therapy for neurological disorders using exosomes and miRNAs, and for boosting neuronal regeneration.

KEYWORDS

neurological disorders, exosomes, mesenchymal stem cells, MSCs, regenerative medicine

1 Background

Neurological illnesses accounted for the second highest cause of mortality and the largest cause of disability in 2016 (Feigin and Vos, 2019). Globally, around 30% of individuals may have a neurological problem at some point in their lives (Feigin and Vos, 2019). Stroke, migraine, Alzheimer's disease, and other dementias are the most prevalent neurological reasons for disabilities and the number of fatalities has climbed by 39% and the number of life years lost due to disability has decreased by 15% in the last 30 years (Feigin and Vos, 2019; Feigin et al., 2020).

There has been significant progress in understanding the mechanisms of nerve regeneration (Wareham et al., 2022). MSCs have emerged as a promising treatment for nerve injury due to their ability to differentiate into various cell types and secrete a range of bioactive molecules (Xia et al., 2023). The colony-forming fibroblastic cells were

extracted from the bone marrow by Friedenstein and colleagues, who were the first to study their properties (Friedenstein et al., 1968).

These cells were distinguished by their preferential adhesion to tissue culture plastics. The groundbreaking research of Friedenstein et al. (1968) was expanded upon by a number of other groups, who demonstrated that these malleable adherent human cells produced from bone marrow could develop into several mesenchymal cell types, such as osteoblasts, chondrocytes, and adipocytes (Ashton et al., 1980; Bab et al., 1986; Castro-Malaspina et al., 1980). Because of their strong capacity for self-renewal and their capacity to generate bone and cartilage, these cells were dubbed "mesenchymal stem cells (MSCs)" and it was hypothesized that they were in charge of the regular turnover and upkeep of adult mesenchymal tissues (Caplan, 2005; Ma, 2010).

Most human cells excrete EVs, which transfer information from one cell to another, resulting in either positive or negative regulation (Thery et al., 2018). One subtype of EVs, exosomes, are released by eukaryotic cells into the extracellular environment and range in size from 30 to 150 nm (Li et al., 2023; Yanez-Mo et al., 2015). Exosomes are small membrane-bound vesicles that can transport and deliver proteins, lipids, nucleic acids, and other cargo to recipient cells (Gurung et al., 2021). Recent research has shown the presence of a variety of nucleic acids, including as mRNAs, miRNAs, and other non-coding RNAs (ncRNAs), in the exosomes. When exosomes circulate, these exosomal RNAs might be absorbed by nearby or distant cells, whereupon they modify recipient cells (Sato-Kuwabara et al., 2015). Exosomes also contain several cellular markers, including CD63, CD81, and CD9, which are often derived from endosomes (Kowal et al., 2016).

Exosomes produced by Mesenchymal Stem Cells (MSC-EXOs) play a crucial role in mediating cell interactions by transferring protein and DNA contents (Xin et al., 2013). Published research suggests that MSC-EXOs may promote neurogenesis, reduce neuronal cell apoptosis, stimulate the formation of new blood vessels, and alleviate local inflammation (Guo et al., 2019; Harrell et al., 2021; Qiu et al., 2015). As vital mediators of intracellular and intercellular communication, exosomes are immune-friendly and biocompatible, making them ideal native nanocarriers for stem cell (SC)-free therapy (He et al., 2018; Liu et al., 2022).

Many studies focus on methods that could support regeneration and lead to medical breakthroughs, such as enhancing neuron survival, increasing the rate of neurite outgrowth, and improving the extensive growth capacity of axons to overcome inhibitory signals. These studies aim to develop new therapies that support complete nerve regeneration (Scheib and Hoke, 2013; Uz et al., 2018). However, successful functional recovery remains insufficient and unsatisfactory (Liu et al., 2022). This situation has led to the exploration of SC-based treatments and their secretomes, such as exosomes, as alternative therapies with fewer drawbacks. This review compiles related studies to provide an overview of recent advancements and suggest potential directions for future research.

2 Materials and methods

The search query "("Exosomes" [Mesh]) AND ("Neurons" [Mesh]) AND ("Mesenchymal Stem Cells" [Mesh]) "was used to gather data for this review from the Pubmed database in January 2024. The following criteria were applied to include all relevant studies:

- 1 Studies that cite MSCs as the source of the exosomes being used.
- 2 Studies that provide methods for characterizing or isolating exosomes, rather than just stating EVs or media for culturing SCs.
- 3 Studies that define a neurological condition that MSC-EXOs aim to target.

This review excluded other publications that either did not mention the term MSCs or did not provide identification evidence for MSCs despite claiming a similar status (such as urine-derived SCs or adipose-derived SCs).

3 Literature review

3.1 MSC-derived exosomes in various neurological disorders

MSC-EXOs have a wide range of therapeutic applications for the treatment of neurological disorders. Exosomes have been found to be beneficial against neurological disorders caused by oxidative stress, peripheral nerve injury, spinal cord injury (SCI), optic nerve damage, Parkinson's disease, neuroinflammation, status epilepticus (SE), and hearing loss (Guy and Offen, 2020; Riazifar et al., 2019). This section offers a comprehensive review of the currently available literature on the impact of MSC-EXOs on different neurological disorders.

3.1.1 Peripheral nerve injury

The crucial role of Schwann cells in healing peripheral nerve injury is underscored, as they can absorb and utilize exosomes from Umbilical Cord MSCs. A significant increase in the expression of endogenous Zeb2, c-JUN, and ERK1/2 in Schwann cells was shown after they absorbed exosomes from hypoxia-pretreated Umbilical Cord MSCs, suggesting a promising approach for nerve repair. The dosage of utilized hypoxic the exosomes was 2.76 mg/g of body weight. Local subcutaneous injection was employed to deliver the exosomes. This activation enhanced nerve regeneration and myelin repair, highlighting the potential therapeutic impact of exosomes on neural tissue (Zhu et al., 2022).

Other studies have investigated the interesting feature of fluorescent tracer uptake in MSC-EXOs and their role in supporting peripheral nerve injury (PNI) healing. MSC-EXOs have shown potential in promoting nerve regeneration and functional recovery following PNI as these exosomes mediate various biological activities and cellular interactions, including the transfer of proteins and DNA contents between donor and recipient cells (Sowa et al., 2016). Gelatin hydrogel tubes were used to implant Schwann cells and adiposederived SCs into the mice's artificially attenuated sciatic nerve injury model. Prior to transplantation, the gelatin hydrogel tube lumen was filled with Schwann cells and SCs generated from adipose tissue at a final density of around 1×10^4 cells/tube (Sowa et al., 2016). Notably, exosomes from different sources, such as adipose tissue and bone marrow, have shown effectiveness in supporting PNI healing (Kourembanas, 2015; Liu et al., 2020).

Reactive oxygen species (ROS) increase during intervertebral disc degeneration (IVDD) due to decreased Nrf2 expression, a key transcription factor that buffers against high ROS. It was found that BMSC-EXOs reduced ROS production while also reducing the apoptosis, inflammation, and degeneration of nucleus pulposus (NP) cells by inhibiting Keap1 and promoting Nrf2 expression (Xu et al., 2023). Nuclear translocation and Nrf2 were enhanced by BMSC-EXOs, while NF- κ B expression was suppressed. Antioxidative protein expression also increased after BMSC-EXOs therapy. For the first time, it was shown that BMSC-EXOs could reactivate the suppressed antioxidant response mechanisms in degenerating NP cells by modifying the Keap1/Nrf2 axis. BMSC-EXOs may be used as an immediate ROS modulator to treat intervertebral disc degeneration (Xu et al., 2023).

3.1.2 Spinal cord injury

Phosphatase and tensin homolog (PTEN) protein expression in the damaged spinal cord area might be reduced by MSC-EXOs loaded with PTEN small interfering RNA (ExoPTEN) after intranasal treatments. Intranasally administered ExoPTEN resulted in significant improvements in motor function, faster recovery of the urine reflex, and sensory recovery. Physiological changes such as reduced neuroinflammation and gliosis, enhanced angiogenesis and axonal regeneration, and improvements in structure and electrophysiology were observed along with functional recovery. The practical use of this fast, noninvasive, cell-free, lesion-specific, and efficient therapy in SCI and other conditions shows considerable potential (Guo et al., 2019).

MSC-EXOs may enhance functional recovery in SCI rats by reducing neuron loss. The miR-21/PTEN/PDCD4 signaling pathway emerged as a key regulator, promoting cell viability and suppressing cell death *in vivo* (Kang et al., 2019).

Bone marrow MSC-EXO could activate the Wnt/ β -catenin signaling pathway, which can be a potentially effective treatment for SCI. Bcl-2 protein expression increased while Bax, cleaved caspase-3, and cleaved caspase-9 protein expression decreased in response to MSC-EXOs intervention (Li et al., 2019).

MSC-EXOs modified with miR-133b may significantly aid in the recovery of neurological function in rats with spinal cord injuries by influencing the signaling pathway associated with axon recovery and the expressions of neurofilament, GAP43, GFAP, and myelin basic protein (Ren et al., 2019).

Administration of MSC-EXOs effectively can reduce inflammation after traumatic spinal cord injury (SCI) and inhibited the activation of A1 neurotoxic reactive astrocytes. Treatment with bone marrow MSC-EXOs reduced neuronal cell death, inhibited nitric oxide emission in microglia, and significantly increased human umbilical vein endothelial cell proliferation, migration, and angiogenic tubule development *in vivo* (Liu et al., 2019).

Moreover, the mechanical and thermal hypersensitivities of rats' right hindpaw caused by nerve ligation were reversed by a single intrathecal injection of exosomes. Treatment with MSC-EXOs inhibited the increase of c-Fos, CNPase, GFAP, and Iba1 induced by nerve ligation. Evidence suggests the potential involvement of exosomes' activities on glial and neuronal cells in their analgesic effects. Exosomes have also demonstrated anti-inflammatory and pro-neurotrophic properties in the ipsilateral L5/6 dorsal root ganglia of nerve-ligated rats by inhibiting the levels of TNF- α and interleukin-1 β (IL-1 β) and increasing the levels of IL-10, brain-derived neurotrophic factor, and glial cell line-derived neurotrophic factor (Shiue et al., 2019).

3.1.3 Hypoxic neuron injuries and exosome-mediated neuroprotection

Hypoxic conditions play a crucial role in determining the impact of exosomes, particularly in hypoxic-related inflammatory conditions. Differentiated MSCs and exosomes derived from immune cells show enhanced treatment efficacy in hypoxic-related disorders. MSC exosomes can enable miR-133b translocation to astrocytes and neurons, which controls gene expression and promotes neurite rebuilding and neurological rehabilitation after stroke *in vivo* (Xin et al., 2013).

Additionally, improved axonal elongation and myelination following a stroke may be facilitated by the miR-17-92 clusterenriched MSC-EXOs. This improved axon-myelin reconstruction could be partially mediated by the activation of the PI3K/Akt/mTOR pathway, which is initiated by PTEN inhibition (Xin et al., 2021). Also, Exosomes loaded with miR-29b-3p have demonstrated a reduction in ischemic brain damage by inhibiting neuronal death and promoting angiogenesis through the Akt signaling cascade (Liu et al., 2020).

Increased axonal elongation and myelination following a stroke may also be enhanced by miR-17-92 cluster-enriched MSC-EXOs. This enhanced axon-myelin reconstruction may be partially mediated by the triggering of the PI3K/Akt/mTOR pathway, which is initiated by a reduction of PTEN (Kourembanas, 2015). MSC-EXOs with increased miR-17-92 cluster improved axonal development via PTEN/mTOR signaling in the neurons. These findings suggest that exosomes produced by MSCs interact with cortical neurons to promote axonal development through the conveyance of biological materials, specifically miRNAs (Zhang et al., 2017).

Moreover, it was shown that both Normoxic Bone marrow MSC-EXOs and hypoxic Bone marrow MSC-EXOs have strong neuroprotective properties against pyroptosis mediated by the NLRP3 inflammasome *in vitro*. Patients may benefit from using hypoxic bone marrow MSC-EXOs to slow the progression of cerebral ischemia and hypoxia damage because they have a more noticeable protective impact than normoxic bone marrow MSC-EXOs (Kang et al., 2021).

3.1.4 Neuroinflammation

Exosomes derived from lipopolysaccharide (LPS)-preconditioned bone marrow MSCs could facilitate the polarization of macrophages toward an M2 phenotype by inhibiting TSG-6. This, in turn, shuts down the NF-KB/NLRP3 regulating axis and expedited functional recovery in a rat model. These exosomes transformed the pro-inflammation macrophage into a pro-regeneration macrophage. Significant amounts of TNF-stimulated gene-6 (TSG-6) were found in extracted exosomes, which inhibited NF-KB and NOD-like receptor protein 3 (NLRP3) and may provide a healing strategy for peripheral nerve damage (Li et al., 2022).

Human amniotic fluid MSC-EXOs could reduce an inflammatory injury induced by an LPS-conditioned medium from microglia *in vitro*. This may have been accomplished by reducing iNOS activity and releasing resolving factors. Additionally, these vesicles may have a neuroprotective function by preventing the detrimental effects of microglial activation (Zavatti et al., 2022).

3.1.5 Amyotrophic lateral sclerosis

Amyotrophic lateral sclerosis (ALS) is a rapidly progressing neurodegenerative disease characterized by the death of motor neurons in the brain and spinal cord. This eventually results in paralysis, muscular atrophy, and weakening. Despite considerable advancements in non-pharmacological therapies and pharmacological symptomatic treatments, effective disease-modifying medications are still elusive.

Exosomes derived from other SC types have the potential to mitigate oxidative damage, reinstate mitochondrial activities, restore the damaged cell endothelium, and enhance motor function in animals modeled with ALS (Wang et al., 2021). For instance, exosomes derived from adipose-derived SCs have been shown to exert a neuroprotective role on NSC-34 cells overexpressing ALS mutations *in vitro* (Bonafede et al., 2016). Recently, researchers discovered that exosomes formed from MSCs might promote tissue regeneration and lower inflammation *in vitro*; moreover, these cell-free products may protect degenerating motor neurons and provide a promising ALS treatment strategy (Gschwendtberger et al., 2023).

3.1.6 Optic nerve damage

MSC-EXOs, which rely on certain miRNAs to control mTORC1 signaling, are a feasible and promising regenerative treatment for optic nerve damage. The delivery of miR-222-3p and miR-22-3p was shown to explain the regenerative effect of MSC-EXOs on axons *in vivo* (Sang et al., 2023).

It was demonstrated that treating optic nerve crush (ONC) rats with intravitreal MSC-EXOs significantly increased the longevity of retinal ganglion cells (RGCs). TNF- α , IL-1 β , IL-6, IL-8, and MCP-1 were among the pro-inflammatory cytokines that showed a decrease in their levels, while IL-10, an anti-inflammatory factor, showed an increase. Furthermore, the injection of MSC-EXOs reduced the amount of apoptosis caused by ONC by upregulating the Bcl-2/Bax ratio and downregulating caspase-3 activity. Moreover, MSC-EXOs markedly increased AKT phosphorylation, while LY294002 reversed the effects of MSC-EXOs on apoptosis prevention. These findings showed that administering MSC-EXOs intravitreally can lessen the damage caused by ONC in a rat model (Cui et al., 2021).

3.1.7 Parkinson's disease

After Alzheimer's disease, Parkinson's disease (PD) is the second most prevalent neurological illness. The pathophysiologic abnormalities in the subsequent basal ganglia's circuitry are caused by the degradation of dopaminergic neurons in the substantia nigra of the midbrain, which is the hallmark of PD (Kalia and Lang, 2016).

In a progressive PD mouse model (α -synuclein A53T transgenic mice), extracted exosomes during the development of dopaminergic neurons from Bone Marrow Stem Cells (BMSCs) could significantly enhance motor, learning, and memory functions. The mechanism behind this effect may be linked to altered phospholipid composition and cholesterol metabolism in hippocampal neurons (Xu et al., 2022). Another study indicated that the injection of Bone Marrow MSC-EXOs into the striatum of PD model rats resulted in a downregulation of the protein levels of IL-6, IL-1 β , TNF- α , and ROS in the substantia nigra (Li et al., 2022).

3.1.8 Status epilepticus

Status Epilepticus (SE) is a subset of epilepsy, representing a debilitating neurological disorder often associated with significant mortality and morbidity numbers. It is a life-threatening neurologic condition that occurs when a person has a continuous seizure or multiple seizures without enough time to recover between them. The incidences of SE are around 50 patients per 100,000 population per year with a mortality rate of around 2.5% (Al-Sofyani, 2021).

IL-1 β -treated MSC-EXOs could prevent astrocytes and status epilepticus mice from experiencing LPS-induced inflammatory reactions, and its effects are mostly facilitated by the Nrf-2 signaling cascade. LPS-induced astrocyte inflammatory responses and astrogliosis could be strongly suppressed by the treatment of these exosomes as well (Liu et al., 2021).

3.1.9 Oxidative stress related neurological disorders and MSC-EXOs

Adipose-derived MSC-EXOs may activate the Nrf2-ARE signaling pathway in hippocampus neurons, therefore suppressing oxidative stress generated by methotrexate (MTX). Furthermore, the Nrf2-ARE signaling pathway has a direct role in mitigating the hippocampal neuronal damage caused by MTX. All things considered, adiposederived MSC-EXOs seem to have potential as medicines for treating neuronal damage caused by MTX while treating cancer (Huang et al., 2022). Additionally, exosomes derived from IL-1 MSCs conditioned may largely act through the Nrf-2 signaling pathway to suppress LPS-induced inflammatory responses in astrocytes and SE mice (Liu et al., 2021).

3.1.10 Hearing loss

Umbilical cord MSC-EXOs may reveal protective benefits in the treatment of ototoxicity-induced hearing loss. Umbilical cord MSC-EXOs could restore hearing loss caused by cisplatin-induced deafness. This was linked to increased expression of glia-derived nexin, mmu-miR-125a-5p, mmu-miR-125b-5p, and mmu-miR127-5p in inner ear tissues and increasing particular growth factors, galectin-3, and fibronectin. These findings may help decreasing inflammation or protecting cochlear hair cells from harm (Tsai et al., 2021).

4 Strategies for improving MSC-EXOs effects

To enhance the therapeutic effects of MSC-EXOs, specific strategies as suggested by available studies can be utilized:

4.1 Considering miRNA role in neural regeneration

One avenue involves investigating the miRNA cargo of exosomes, as certain miRNAs have been associated with angiogenesis, neuroprotection, and neuroimmune regulation. Differentially expressed miRNAs, such as miRNA-199b-5p and miRNA-132-3p, have been identified in exosomes and may play a crucial role in boosting the neural regeneration capacity of exosome therapy (Table 1) (Liu et al., 2022).

4.2 Macrophage polarization role and peripheral nerve injury

Exosomes derived from lipopolysaccharide (LPS) preconditioned MSCs (LPS pre-Exos) may facilitate the polarization

TABLE 1 Mesenchymal stem cells' exosomal miRNA targets and effects.

Type of cells	Disease	miRNA	Target/possible effect(s)	Reference(s)
Human umbilical cord mesenchymal/stromal stem cells (MSCs)	Retinitis pigmentosa	miRNA-21-5p miRNA-122-5p miRNA-146a-5p miRNA -let7a-5p miRNA -148a-3p miRNA -let7i-5p miRNA -222-3p miRNA -320a-3p miRNA -100-5p miRNA -26a-5p	Anti-inflammation	Zhang et al. (2022)
Bone marrow MSCs	Spinal cord injury	miR-124	Neuronal differentiation	Zou et al. (2014)
	Optic Nerve Crush Injury	miR-17-5p miR-30c-2 miR-92a miR-92a miR-292 miR-182	Neuroprotective	Mead et al. (2020)
	Нурохіа	miR-210	Increase migration of neural precursor cells	Wang et al. (2020)
	Spinal cord injury	miRNA-29b	Neuronal regeneration	Yu et al. (2019)
	Loss of retinal ganglion cells	SiRNA Argonaute 2	Neuroprotective and neuritogenic effects	Mead and Tomarev (2017)
	Recurrent laryngeal nerve injury	miR-21	Regeneration	Wu et al. (2017)
	Stroke	miR-1906	Neuroprotection	Haupt et al. (2021)
	Traumatic brain injury	miR-216a-5p	Promote neurogenesis and inhibit apoptosis	Xu et al. (2020)
	Spinal cord injury	miR-125a	Neuroprotective effect	Chang et al. (2021)
	Cognitive and behavioral distortion	miR-30c-5p miR-29b-3p miR-16-5p miR-221-3p miR-181a-5p miR-145-5p miR-31-5p miR-34a-5p miR-30c-5p miR-29b-3p miR-16-5p miR-221-3p miR-181a-5p miR-145-5p miR-145-5p miR-34a-5p miR-34a-5p miR-34a-5p miR-199a-3p	Anti-inflammatory Reduce apoptotic state, increase astrocyte, and microglia activation	El-Derany and Noureldein (2021)

(Continued)

10.3389/fncel.2024.1427525

Type of cells	Disease	miRNA	Target/possible effect(s)	Reference(s)
Wharton's Jelly MSCs	Hypoxic-ischemic	let-7b-5p let-7c-5p let-7c-5p let-7i-5p let-7i-5p let-7i-5p let-7i-5p let-7i-5p miR-9-3p miR-20a-5p miR-24-3p miR-26b-5p miR-29a-3p miR-29b-3p miR-29c-3p miR-92a-3p miR-93-5p miR-98-5p miR-106b-5p miR-133b miR-181a-5p miR-298 miR-320a miR-320a miR-320a miR-342-3p	Anti-apoptotic effect	Joerger-Messerli et al. (2018)
Adipose-derived MSCs	Acute sciatic nerve injury	miR-432-5p miR-195a-5p miR-184-3p miR-9-5p miR-137-3p miR-124-3p miR-132-3p	Neural differentiation	Roballo et al. (2019)
	Alzheimer's disease	miRNA-22	Inhibit apoptosis, Anti-inflammatory effect	Zhai et al. (2021)
human Adipose derived MSCs	Peripheral nerve injury	miRNA-199b-5p	Modulate peripheral nerve-related cellular functions	Liu et al. (2022)
MSCs	Intracerebral hemorrhage	miR-133b	Neuroprotective role for anti-apoptotic effect	Shen et al. (2018)
PC12 cells and MSCs	Spinal cord injury	miR-21 miR-19b	Suppresses the apoptosis of neuron, increase cell differentiation	n Xu et al. (2019)

of macrophages toward an M2 phenotype by inhibiting TNF-stimulated gene-6 (TSG-6). This subsequently modulates the NF-KB/NLRP3 axis (Li et al., 2022). This novel discovery presents a potential therapeutic strategy for peripheral nerve injury, demonstrating enhanced axon regrowth, remyelination, and improved M2 macrophage polarization upon local injection of LPS pre-Exos (Figure 1) (Li et al., 2022).

4.3 Utilizing IL-1 preconditioned MSC-EXOs

Treatment with IL-1 may greatly assist MSCs in the production of pro-trophic and anti-inflammatory compounds. IL-1 preconditioned MSC-EXOs can inhibit inflammatory responses in astrocytes and status epilepticus mice by acting via the Nrf-2 signaling pathway (Figure 1) (Liu et al., 2021).

4.4 Utilizing hypoxic preconditioned MSC-EXOs

Hypoxic conditions have been shown to play a critical role in enhancing the therapeutic impact of exosomes, particularly in hypoxia-related inflammatory conditions. Hypoxic conditioning of MSCs increases paracrine actions, altering key signaling pathways such as transforming growth factor β (TGF- β), NF- κ B, and PI3K/Akt (Kang et al., 2019; Tirpe et al., 2019). Hypoxic preconditioning of umbilical cord MSC-EXOs can activate crucial components, including Zeb2, c-JUN, and ERK1/2 in Schwann cells, promoting nerve regeneration and myelin repair (Figure 1) (Zhu et al., 2022).

4.5 Neuroprotective properties of normoxic and hypoxic bone marrow MSC-EXOs

Moreover, Bone marrow MSC-EXOs and hypoxic Bone marrow MSC-EXOs can reveal strong neuroprotective properties against pyroptosis mediated by the NLRP3 inflammasome. Patients may benefit from using hypoxic bone marrow MSC-EXOs to slow the course of cerebral ischemia and hypoxia damage, as they have a more noticeable protective impact than normoxic bone marrow MSC-EXOs (Figure 1) (Kang et al., 2021).

4.6 Autologous MSC-EXOs

Researchers created a scaffold based on autologous plasma exosomes (AP-EXO), which were loaded with peptides that promote proliferation and target neurons. After inducing SCI in rats, $30 \mu g$ of this scaffold was injected intravenously and induced strong axon regeneration across the lesion core, reaching levels over 30-fold larger than naïve therapy. This helped to restore intraspinal networks and increase motor functional recovery. The AP-EXO-based tailored therapy, which combines safety and effectiveness, allows for functional recovery after SCI and has shown great promise for use in biomedical applications. Expanding the use of combinatory peptides and autologous exosomes produced from human plasma is beneficial in fostering regeneration and recovery during SCI therapy (Ran et al., 2023).

4.7 Non-autologous MSC-EXOs

For targeted therapeutics including CRISPR/Cas9 gene editing, exosomes provide a potentially effective delivery system. Using a subcutaneous tumor model, the *in vivo* treatment effectiveness of CRISPR/Cas9 packed MSC-EXOs was assessed by insertion of cultivated KPC689 cells in B6-albino mouse.

Mice were given a subcutaneous injection of one million KPC689 cells in the flank. 10⁹ CRISPR/Cas9-loaded exosomes (10⁹ exosomes, 10 µg plasmid DNA) were injected intratumorally. In syngeneic subcutaneous and orthotopic models of pancreatic cancer, exosomes equipped with CRISPR/Cas9 may target the mutant Kras^{G12D} oncogenic allele in pancreatic cancer cells to reduce proliferation and limit tumor development. This approach can be promising for neurological disorders as well (McAndrews et al., 2021).

5 Conclusion: future directions and implications for neurological therapeutics

5.1 Difficulties and solutions in using MSC-EXOs in therapeutic settings

There are several issues and difficulties with using MSC-EXOs in clinical settings. To comply with certain good manufacturing standards (GMP), translation of exosome-based clinical trials is necessary (Rezaie et al., 2022). Secondly, establishing uniformity in MSC culture and exosome separation techniques is required. A methodology for exosome separation, processing, identification, and functional dissection has been proposed earlier (Rai et al., 2021).

Thirdly, methods for effective delivery of exosomes to their intended locations are needed. To induce precise delivery, therapeutic exosomes can be genetically modified to express different targeting molecules. This can be done either directly, by adding covalent or non-covalent bonds to the exosomal surface, or indirectly, by genetically modifying the cells that produce exosomes (Choi et al., 2021).

Fourthly, recognizing the long-term safety and effectiveness of exosomes for nerve regeneration must be considered. Clinical research has shown that exosomes represent a promising therapeutic platform for several illnesses including Alzheimer's disease, PD, SCI, PNI, and ALS. Although this exciting field is developing swiftly, more multidisciplinary research is required to address these little vesicles since we still have little understanding of the underlying mechanisms governing the many roles that exosomes play (Namini et al., 2023).

The comprehensive findings across studies underscore the potential of MSC-EXOs as a cell-free biological therapy for various neurological disorders. The ability of exosomes to modulate inflammatory responses, activate specific signaling pathways, and



promote nerve regeneration positions them as promising candidates for future therapeutic interventions. Further research is warranted to elucidate the intricate mechanisms underlying exosome-mediated neuroprotection, offering new insights into neurological therapeutics and paving the way for innovative treatment strategies (Guo et al., 2019; Tirpe et al., 2019; Xu et al., 2022).

5.2 Future perspectives

As we move forward in the field of cellular regeneration, rejuvenation, and longevity of neuronal cells, further research is

needed to uncover the specific miRNA cargo within exosomes and clarify their role in neural regeneration and protection. Standardizing isolation and characterization techniques will enhance the reproducibility of results. Moreover, it will be crucial for translational medicine to explore the potential of MSC-EXOs in clinical trials for a variety of neurological conditions. In conclusion, MSC-EXOs are at the forefront of innovative therapeutic strategies for neurological disorders. Their multifaceted impact on neuronal regeneration, modulation of inflammatory responses, and diverse applications highlight their potential importance in clinical settings. Continued research and clinical trials will pave the way for harnessing the full therapeutic potential of MSC-EXOs in addressing the complex challenges of neurological disorders.

Author contributions

AS: Writing – review & editing, Writing – original draft. ZK: Writing – review & editing, Writing – original draft. MG: Writing – original draft. MA: Writing – original draft. AK: Writing – original draft. FM: Writing – original draft. MZ: Writing – review & editing, Writing – original draft, Investigation. AA: Writing – review & editing, Writing – original draft, Supervision, Methodology, Conceptualization.

Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

References

Al-Sofyani, K. A. (2021). An insight into the current understanding of status epilepticus: from concept to management. *Neurol. Res. Int.* 2021, 1–12. doi: 10.1155/2021/9976754

Ashton, B. A., Allen, T. D., Howlett, C. R., Eaglesom, C. C., Hattori, A., and Owen, M. (1980). Formation of bone and cartilage by marrow stromal cells in diffusion chambers in vivo. *Clin. Orthop. Relat. Res.* 151, 294–307.

Bab, I., Ashton, B. A., Gazit, D., Marx, G., Williamson, M. C., and Owen, M. E. (1986). Kinetics and differentiation of marrow stromal cells in diffusion chambers in vivo. *J. Cell Sci.* 84, 139–151. doi: 10.1242/jcs.84.1.139

Bonafede, R., Scambi, I., Peroni, D., Potrich, V., Boschi, F., Benati, D., et al. (2016). Exosome derived from murine adipose-derived stromal cells: neuroprotective effect on in vitro model of amyotrophic lateral sclerosis. *Exp. Cell Res.* 340, 150–158. doi: 10.1016/j.yexcr.2015.12.009

Caplan, A. I. (2005). Review: mesenchymal stem cells: cell-based reconstructive therapy in orthopedics. *Tissue Eng.* 11, 1198-1211. doi: 10.1089/ten.2005.11.1198

Castro-Malaspina, H., Gay, R. E., Resnick, G., Kapoor, N., Meyers, P., Chiarieri, D., et al. (1980). Characterization of human bone marrow fibroblast colony-forming cells (CFU-F) and their progeny. *Blood* 56, 289–301. doi: 10.1182/blood.V56.2.289.289

Chang, Q., Hao, Y., Wang, Y., Zhou, Y., Zhuo, H., and Zhao, G. (2021). Bone marrow mesenchymal stem cell-derived exosomal microRNA-125a promotes M2 macrophage polarization in spinal cord injury by downregulating IRF5. *Brain Res. Bull.* 170, 199–210. doi: 10.1016/j.brainresbull.2021.02.015

Choi, H., Choi, Y., Yim, H. Y., Mirzaaghasi, A., Yoo, J. K., and Choi, C. (2021). Biodistribution of exosomes and engineering strategies for targeted delivery of therapeutic exosomes. *Tissue Eng. Regen. Med.* 18, 499–511. doi: 10.1007/ s13770-021-00361-0

Cui, Y., Liu, C., Huang, L., Chen, J., and Xu, N. (2021). Protective effects of intravitreal administration of mesenchymal stem cell-derived exosomes in an experimental model of optic nerve injury. *Exp. Cell Res.* 407:112792. doi: 10.1016/j.yexcr.2021.112792

El-Derany, M. O., and Noureldein, M. H. (2021). Bone marrow mesenchymal stem cells and their derived exosomes resolve doxorubicin-induced chemobrain: critical role of their miRNA cargo. *Stem Cell Res Ther* 12, 1–23. doi: 10.1186/s13287-021-02384-9

Feigin, V. L., and Vos, T. (2019). Global burden of neurological disorders: from global burden of disease estimates to actions. *Neuroepidemiology* 52, 1–2. doi: 10.1159/000495197

Feigin, V. L., Vos, T., Nichols, E., Owolabi, M. O., Carroll, W. M., Dichgans, M., et al. (2020). The global burden of neurological disorders: translating evidence into policy. *Lancet Neurol.* 19, 255–265. doi: 10.1016/S1474-4422(19)30411-9

Friedenstein, A. J., Petrakova, K. V., Kurolesova, A. I., and Frolova, G. P. (1968). Heterotopic of bone marrow. Analysis of precursor cells for osteogenic and hematopoietic tissues. *Transplantation* 6, 230–247. doi: 10.1097/00007890-196803000-0009

Gschwendtberger, T., Thau-Habermann, N., von der Ohe, J., Luo, T., Hass, R., and Petri, S. (2023). Protective effects of EVs/exosomes derived from permanently growing human MSC on primary murine ALS motor neurons. *Neurosci. Lett.* 816:137493. doi: 10.1016/j.neulet.2023.137493

Guo, S., Perets, N., Betzer, O., Ben-Shaul, S., Sheinin, A., Michaelevski, I., et al. (2019). Intranasal delivery of mesenchymal stem cell derived exosomes loaded with phosphatase and Tensin homolog siRNA repairs complete spinal cord injury. *ACS Nano* 13, 10015–10028. doi: 10.1021/acsnano.9b01892

Gurung, S., Perocheau, D., Touramanidou, L., and Baruteau, J. (2021). The exosome journey: from biogenesis to uptake and intracellular signalling. *Cell Commun. Signal* 19:47. doi: 10.1186/s12964-021-00730-1

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Guy, R., and Offen, D. (2020). Promising opportunities for treating neurodegenerative diseases with mesenchymal stem cell-derived exosomes. *Biomol. Ther.* 10:1320. doi: 10.3390/biom10091320

Harrell, C. R., Volarevic, A., Djonov, V., and Volarevic, V. (2021). Mesenchymal stem cell-derived exosomes as new remedy for the treatment of neurocognitive disorders. *Int. J. Mol. Sci.* 22:1433. doi: 10.3390/ijms22031433

Haupt, M., Zheng, X., Kuang, Y., Lieschke, S., Janssen, L., Bosche, B., et al. (2021). Lithium modulates miR-1906 levels of mesenchymal stem cell-derived extracellular vesicles contributing to poststroke neuroprotection by toll-like receptor 4 regulation. *Stem Cells Transl. Med.* 10, 357–373. doi: 10.1002/sctm.20-0086

He, C., Zheng, S., Luo, Y., and Wang, B. (2018). Exosome Theranostics: biology and translational medicine. *Theranostics* 8, 237–255. doi: 10.7150/thno.21945

Huang, T., Tong, H., Zhou, H., Wang, J., Hu, L., Wang, Y., et al. (2022). ADSCexosomes alleviate MTX-induced rat neuronal damage by activating Nrf2-ARE pathway. *J. Mol. Neurosci.* 72, 1334–1344. doi: 10.1007/s12031-022-01996-x

Joerger-Messerli, M. S., Oppliger, B., Spinelli, M., Thomi, G., Di Salvo, I., Schneider, P., et al. (2018). Extracellular vesicles derived from Wharton's jelly mesenchymal stem cells prevent and resolve programmed cell death mediated by perinatal hypoxia-ischemia in neuronal cells. *Cell Transplant.* 27, 168–180. doi: 10.1177/0963689717738256

Kalia, L. V., and Lang, A. E. (2016). Parkinson disease in 2015: evolving basic, pathological and clinical concepts in PD. Nat. Rev. Neurol. 12, 65–66. doi: 10.1038/nrneurol.2015.249

Kang, X., Jiang, L., Chen, X., Wang, X., Gu, S., Wang, J., et al. (2021). Exosomes derived from hypoxic bone marrow mesenchymal stem cells rescue OGD-induced injury in neural cells by suppressing NLRP3 inflammasome-mediated pyroptosis. *Exp. Cell Res.* 405:112635. doi: 10.1016/j.yexcr.2021.112635

Kang, J., Li, Z., Zhi, Z., Wang, S., and Xu, G. (2019). MiR-21 derived from the exosomes of MSCs regulates the death and differentiation of neurons in patients with spinal cord injury. *Gene Ther.* 26, 491–503. doi: 10.1038/s41434-019-0101-8

Kourembanas, S. (2015). Exosomes: vehicles of intercellular signaling, biomarkers, and vectors of cell therapy. *Annu. Rev. Physiol.* 77, 13–27. doi: 10.1146/annurev-physiol-021014-071641

Kowal, J., Arras, G., Colombo, M., Jouve, M., Morath, J. P., Primdal-Bengtson, B., et al. (2016). Proteomic comparison defines novel markers to characterize heterogeneous populations of extracellular vesicle subtypes. *Proc. Natl. Acad. Sci. USA* 113, E968–E977. doi: 10.1073/pnas.1521230113

Li, R. Y., Hu, Q., Shi, X., Luo, Z. Y., and Shao, D. H. (2023). Crosstalk between exosomes and autophagy in spinal cord injury: fresh positive target for therapeutic application. *Cell Tissue Res.* 391, 1–17. doi: 10.1007/s00441-022-03699-6

Li, C., Jiao, G., Wu, W., Wang, H., Ren, S., Zhang, L., et al. (2019). Exosomes from bone marrow mesenchymal stem cells inhibit neuronal apoptosis and promote motor function recovery via the Wnt/beta-catenin signaling pathway. *Cell Transplant.* 28, 1373–1383. doi: 10.1177/0963689719870999

Li, Y., Li, Z., Gu, J., Xu, X., Chen, H., and Gui, Y. (2022). Exosomes isolated during dopaminergic neuron differentiation suppressed neuronal inflammation in a rodent model of Parkinson's disease. *Neurosci. Lett.* 771:136414. doi: 10.1016/j. neulet.2021.136414

Li, C., Li, X., Shi, Z., Wu, P., Fu, J., Tang, J., et al. (2022). Exosomes from LPSpreconditioned bone marrow MSCs accelerated peripheral nerve regeneration via M2 macrophage polarization: involvement of TSG-6/NF-kappaB/NLRP3 signaling pathway. *Exp. Neurol.* 356:114139. doi: 10.1016/j.expneurol.2022.114139

Liu, K., Cai, G. L., Zhuang, Z., Pei, S. Y., Xu, S. N., Wang, Y. N., et al. (2021). Interleukin-1beta-treated mesenchymal stem cells inhibit inflammation in hippocampal astrocytes through exosome-activated Nrf-2 signaling. Int. J. Nanomedicine 16, 1423–1434. doi: 10.2147/IJN.S289914

Liu, B., Kong, Y., Shi, W., Kuss, M., Liao, K., Hu, G., et al. (2022). Exosomes derived from differentiated human ADMSC with the Schwann cell phenotype modulate peripheral nerve-related cellular functions. *Bioact. Mater.* 14, 61–75. doi: 10.1016/j. bioactmat.2021.11.022

Liu, W., Wang, Y., Gong, F., Rong, Y., Luo, Y., Tang, P., et al. (2019). Exosomes derived from bone mesenchymal stem cells repair traumatic spinal cord injury by suppressing the activation of A1 neurotoxic reactive astrocytes. *J. Neurotrauma* 36, 469–484. doi: 10.1089/neu.2018.5835

Liu, C. Y., Yin, G., Sun, Y. D., Lin, Y. F., Xie, Z., English, A. W., et al. (2020). Effect of exosomes from adipose-derived stem cells on the apoptosis of Schwann cells in peripheral nerve injury. *CNS Neurosci. Ther.* 26, 189–196. doi: 10.1111/cns.13187

Ma, T. (2010). Mesenchymal stem cells: from bench to bedside. World J. Stem Cells 2, 13–17. doi: 10.4252/wjsc.v2.i2.13

McAndrews, K. M., Xiao, F., Chronopoulos, A., LeBleu, V. S., Kugeratski, F. G., and Kalluri, R. (2021). Exosome-mediated delivery of CRISPR/Cas9 for targeting of oncogenic Kras(G12D) in pancreatic cancer. *Life Sci. Alliance* 4:e202000875. doi: 10.26508/lsa.202000875

Mead, B., Cullather, E., Nakaya, N., Niu, Y., Kole, C., Ahmed, Z., et al. (2020). Viral delivery of multiple miRNAs promotes retinal ganglion cell survival and functional preservation after optic nerve crush injury. *Exp. Eye Res.* 197:108071. doi: 10.1016/j.exer.2020.108071

Mead, B., and Tomarev, S. (2017). BMSC-derived exosomes promote survival of retinal ganglion cells through miRNA-dependent mechanisms. *Stem Cells Transl. Med.* 6, 1273–1285. doi: 10.1002/sctm.16-0428

Namini, M. S., Daneshimehr, F., Beheshtizadeh, N., Mansouri, V., Ai, J., Jahromi, H. K., et al. (2023). Cell-free therapy based on extracellular vesicles: a promising therapeutic strategy for peripheral nerve injury. *Stem Cell Res Ther* 14:254. doi: 10.1186/s13287-023-03467-5

Qiu, X. C., Jin, H., Zhang, R. Y., Ding, Y., Zeng, X., Lai, B. Q., et al. (2015). Donor mesenchymal stem cell-derived neural-like cells transdifferentiate into myelin-forming cells and promote axon regeneration in rat spinal cord transection. *Stem Cell Res Ther* 6:105. doi: 10.1186/s13287-015-0100-7

Rai, A., Fang, H., Fatmous, M., Claridge, B., Poh, Q. H., Simpson, R. J., et al. (2021). A protocol for isolation, purification, characterization, and functional dissection of exosomes. *Methods Mol. Biol.* 2261, 105–149. doi: 10.1007/978-1-0716-1186-9_9

Ran, N., Li, W., Zhang, R., Lin, C., Zhang, J., Wei, Z., et al. (2023). Autologous exosome facilitates load and target delivery of bioactive peptides to repair spinal cord injury. *Bioact. Mater.* 25, 766–782. doi: 10.1016/j.bioactmat.2022.07.002

Ren, Z. W., Zhou, J. G., Xiong, Z. K., Zhu, F. Z., and Guo, X. D. (2019). Effect of exosomes derived from MiR-133b-modified ADSCs on the recovery of neurological function after SCI. *Eur. Rev. Med. Pharmacol. Sci.* 23, 52–60. doi: 10.26355/eurrev_201901_16747

Rezaie, J., Feghhi, M., and Etemadi, T. (2022). A review on exosomes application in clinical trials: perspective, questions, and challenges. *Cell Commun. Signal* 20:145. doi: 10.1186/s12964-022-00959-4

Riazifar, M., Mohammadi, M. R., Pone, E. J., Yeri, A., Lasser, C., Segaliny, A. I., et al. (2019). Stem cell-derived exosomes as nanotherapeutics for autoimmune and neurodegenerative disorders. *ACS Nano* 13, 6670–6688. doi: 10.1021/acsnano.9b01004

Roballo, K. C. S., da Silveira, J. C., Bressan, F. F., de Souza, A. F., Pereira, V. M., Porras, J. E. P., et al. (2019). Neurons-derived extracellular vesicles promote neural differentiation of ADSCs: a model to prevent peripheral nerve degeneration. *Sci. Rep.* 9:11213. doi: 10.1038/s41598-019-47229-x

Sang, X., Tang, L., Zhao, L., Xu, N., Liu, F., Shen, Y., et al. (2023). Umbilical cord mesenchymal stem cell-derived exosomes promote axon regeneration during optic nerve injury through microRNA-dependent mTORC1 signalling. *Clin. Transl. Med.* 13:e1319. doi: 10.1002/ctm2.1319

Sato-Kuwabara, Y., Melo, S. A., Soares, F. A., and Calin, G. A. (2015). The fusion of two worlds: non-coding RNAs and extracellular vesicles – diagnostic and therapeutic implications (review). *Int. J. Oncol.* 46, 17–27. doi: 10.3892/ijo.2014.2712

Scheib, J., and Hoke, A. (2013). Advances in peripheral nerve regeneration. *Nat. Rev. Neurol.* 9, 668–676. doi: 10.1038/nrneurol.2013.227

Shen, H., Yao, X., Li, H., Li, X., Zhang, T., Sun, Q., et al. (2018). Role of exosomes derived from miR-133b modified MSCs in an experimental rat model of intracerebral hemorrhage. *J. Mol. Neurosci.* 64, 421–430. doi: 10.1007/s12031-018-1041-2

Shiue, S. J., Rau, R. H., Shiue, H. S., Hung, Y. W., Li, Z. X., Yang, K. D., et al. (2019). Mesenchymal stem cell exosomes as a cell-free therapy for nerve injury-induced pain in rats. *Pain* 160, 210–223. doi: 10.1097/j.pain.000000000001395

Sowa, Y., Kishida, T., Imura, T., Numajiri, T., Nishino, K., Tabata, Y., et al. (2016). Adipose-derived stem cells promote peripheral nerve regeneration in vivo without differentiation into Schwann-like lineage. *Plast. Reconstr. Surg.* 137, 318e–330e. doi: 10.1097/01.prs.0000475762.86580.36

Thery, C., Witwer, K. W., Aikawa, E., Alcaraz, M. J., Anderson, J. D., Andriantsitohaina, R., et al. (2018). Minimal information for studies of extracellular vesicles 2018 (MISEV2018): a position statement of the International Society for Extracellular Vesicles and update of the MISEV2014 guidelines. *J. Extracell. Vesicles* 7:1535750. doi: 10.1080/20013078.2018.1535750 Tirpe, A. A., Gulei, D., Ciortea, S. M., Crivii, C., and Berindan-Neagoe, I. (2019). Hypoxia: overview on hypoxia-mediated mechanisms with a focus on the role of HIF genes. *Int. J. Mol. Sci.* 20:6140. doi: 10.3390/ijms20246140

Tsai, S. C., Yang, K. D., Chang, K. H., Lin, F. C., Chou, R. H., Li, M. C., et al. (2021). Umbilical cord mesenchymal stromal cell-derived exosomes rescue the loss of outer hair cells and repair Cochlear damage in cisplatin-injected mice. *Int. J. Mol. Sci.* 22:6664. doi: 10.3390/ijms22136664

Uz, M., Das, S. R., Ding, S., Sakaguchi, D. S., Claussen, J. C., and Mallapragada, S. K. (2018). Advances in controlling differentiation of adult stem cells for peripheral nerve regeneration. *Adv. Healthc. Mater.* 7:e1701046. doi: 10.1002/adhm.201701046

Wang, F., Jie, Z., Jian, Z., Wei, D., and Zhujuan, Z. (2020). miR-210 enhances mesenchymal stem cell-modulated neural precursor cell migration. *Mol. Med. Reports.* 21, 2405–14.

Wang, K., Li, Y., Ren, C., Wang, Y., He, W., and Jiang, Y. (2021). Extracellular vesicles as innovative treatment strategy for amyotrophic lateral sclerosis. *Front. Cell Dev. Biol.* 9:754630. doi: 10.3389/fcell.2021.754630

Wareham, L. K., Liddelow, S. A., Temple, S., Benowitz, L. I., Di Polo, A., Wellington, C., et al. (2022). Solving neurodegeneration: common mechanisms and strategies for new treatments. *Mol. Neurodegener*, 17:23. doi: 10.1186/s13024-022-00524-0

Wu, W., Zhang, S., Chen, Y., and Liu, H. (2017). Biological function and mechanism of bone marrow mesenchymal stem cells-packed poly (3, 4-ethylenedioxythiophene) (PEDOT) scaffolds for peripheral nerve injury: the involvement of miR-21-notch signaling pathway. *Curr. Neurovasc. Res.* 14, 19–25. doi: 10.217 4/1567202614666161123112832

Xia, Y., Zhu, J., Yang, R., Wang, H., Li, Y., and Fu, C. (2023). Mesenchymal stem cells in the treatment of spinal cord injury: mechanisms, current advances and future challenges. *Front. Immunol.* 14:1141601. doi: 10.3389/fimmu.2023.1141601

Xin, H., Li, Y., Liu, Z., Wang, X., Shang, X., Cui, Y., et al. (2013). MiR-133b promotes neural plasticity and functional recovery after treatment of stroke with multipotent mesenchymal stromal cells in rats via transfer of exosome-enriched extracellular particles. *Stem Cells* 31, 2737–2746. doi: 10.1002/stem.1409

Xin, H., Liu, Z., Buller, B., Li, Y., Golembieski, W., Gan, X., et al. (2021). MiR-17-92 enriched exosomes derived from multipotent mesenchymal stromal cells enhance axonmyelin remodeling and motor electrophysiological recovery after stroke. *J. Cereb. Blood Flow Metab.* 41, 1131–1144. doi: 10.1177/0271678X20950489

Xu, G., Ao, R., Zhi, Z., Jia, J., and Yu, B. (2019). miR-21 and miR-19b delivered by hMSC-derived EVs regulate the apoptosis and differentiation of neurons in patients with spinal cord injury. *J. Cell. Physiol.* 234, 10205–10217. doi: 10.1002/jcp.27690

Xu, H., Jia, Z., Ma, K., Zhang, J., Dai, C., Yao, Z., et al. (2020). Protective effect of BMSCs-derived exosomes mediated by BDNF on TBI via miR-216a-5p. *Med. Sci. Monit.* 26:e920855. doi: 10.12659/MSM.920855

Xu, X., Li, Z., Zuo, H., Chen, H., and Gui, Y. (2022). Mesenchymal stem cell-derived exosomes altered neuron cholesterol metabolism via Wnt5a-LRP1 axis and alleviated cognitive impairment in a progressive Parkinson's disease model. *Neurosci. Lett.* 787:136810. doi: 10.1016/j.neulet.2022.136810

Xu, G., Lu, X., Liu, S., Zhang, Y., Xu, S., Ma, X., et al. (2023). MSC-derived exosomes ameliorate intervertebral disc degeneration by regulating the Keap1/Nrf2 Axis. *Stem Cell Rev. Rep.* 19, 2465–2480. doi: 10.1007/s12015-023-10570-w

Yanez-Mo, M., Siljander, P. R., Andreu, Z., Zavec, A. B., Borras, F. E., Buzas, E. I., et al. (2015). Biological properties of extracellular vesicles and their physiological functions. *J. Extracell. Vesicles* 4:27066. doi: 10.3402/jev.v4.27066

Yu, T., Zhao, C., Hou, S., Zhou, W., Wang, B., and Chen, Y. (2019). Exosomes secreted from miRNA-29b-modified mesenchymal stem cells repaired spinal cord injury in rats. *Braz. J. Med. Biol. Res.* 52:e8735. doi: 10.1590/1414-431x20198735

Zavatti, M., Gatti, M., Beretti, F., Palumbo, C., and Maraldi, T. (2022). Exosomes derived from human amniotic fluid mesenchymal stem cells preserve microglia and neuron cells from Abeta. *Int. J. Mol. Sci.* 23:4967. doi: 10.3390/ijms23094967

Zhai, L., Shen, H., Sheng, Y., and Guan, Q. (2021). ADMSC Exo-MicroRNA-22 improve neurological function and neuroinflammation in mice with Alzheimer's disease. J. Cell. Mol. Med. 25, 7513–7523. doi: 10.1111/jcmm.16787

Zhang, Y., Chopp, M., Liu, X. S., Katakowski, M., Wang, X., Tian, X., et al. (2017). Exosomes derived from mesenchymal stromal cells promote axonal growth of cortical neurons. *Mol. Neurobiol.* 54, 2659–2673. doi: 10.1007/s12035-016-9851-0

Zhang, J., Li, P., Zhao, G., He, S., Xu, D., Jiang, W., et al. (2022). Mesenchymal stem cell-derived extracellular vesicles protect retina in a mouse model of retinitis pigmentosa by anti-inflammation through miR-146a-Nr4a3 axis. *Stem Cell Res Ther* 13, 1–22. doi: 10.1186/s13287-022-03100-x

Zhu, Z., Zhang, Y., Huang, Z., Hao, H., and Yan, M. (2022). Hypoxic culture of umbilical cord mesenchymal stem cell-derived sEVs prompts peripheral nerve injury repair. *Front. Cell. Neurosci.* 16:897224. doi: 10.3389/fncel.2022.897224

Zou, D., Chen, Y., Han, Y., Lv, C., and Tu, G. (2014). Overexpression of microRNA-124 promotes the neuronal differentiation of bone marrow-derived mesenchymal stem cells. *Neural Regen. Res.* 9, 1241–1248. doi: 10.4103/1673-5374.135333

Glossary

MSCs	Mesenchymal stem cells		
MSC-EXOs	Mesenchymal stem cells derived exosomes		
EVs	extracellular vesicles		
TGF-β	transforming growth factor β		
TSG-6	TNF-stimulated gene-6		
miRNA	micro RNA		
Akt	protein kinase B		
РІЗК	phosphatidylinositol 3-kinase		
ROS	reactive oxygen species		
IVDD	intervertebral disc		
Map2	microtubule-associated protein		
CNPase	2',3'-cyclic-nucleotide 39-phosphodiesterase		
Iba1	ionized calcium-binding adaptor molecule 1		
GFAP	glial fibrillary acidic protein		
GAP43	growth associated protein 43		
NP	nucleus pulposus		
mTOR	mammalian target of rapamycin		
NLRP3	NOD-like receptor protein 3		
MTX	methotrexate		
IL-1	Interleukin-1beta		
ALS	amyotrophic lateral sclerosis		
SE	Status Epilepticus		
PD	Parkinson's disease		
ONC	optic nerve crush		
RGCs	retinal ganglion cells		
AP-EXO	autologous plasma exosomes		