#### Check for updates

#### **OPEN ACCESS**

EDITED BY Johnny Di Pierdomenico, University of Murcia, Spain

REVIEWED BY Raymond Scott Duncan, University of Missouri–Kansas City, United States

\*CORRESPONDENCE Harald Neumann ⊠ harald.neumann@uni-bonn.de

RECEIVED 06 May 2025 ACCEPTED 03 June 2025 PUBLISHED 27 June 2025

#### CITATION

Min Y, Cuevas-Rios G, Langmann T and Neumann H (2025) Sialylation as a checkpoint for inflammatory and complement-related retinal diseases. *Front. Cell. Neurosci.* 19:1623755.

doi: 10.3389/fncel.2025.1623755

#### COPYRIGHT

© 2025 Min, Cuevas-Rios, Langmann and Neumann. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

## Sialylation as a checkpoint for inflammatory and complement-related retinal diseases

## Yiduo Min<sup>1</sup>, German Cuevas-Rios<sup>1</sup>, Thomas Langmann<sup>2</sup> and Harald Neumann<sup>1\*</sup>

<sup>1</sup>Institute of Reconstructive Neurobiology, Medical Faculty & University Hospital Bonn, University of Bonn, Bonn, Germany, <sup>2</sup>Experimental Immunology of the Eye, Department of Ophthalmology, University Hospital Cologne, Cologne, Germany

Sialylation is a modification process involving the addition of sialic acid residues to the termini of glycoproteins and glycolipids in mammalian cells. Sialylation serves as a crucial checkpoint inhibitor of the complement and immune systems, particularly within the central nervous system (CNS), including the retina. Complement factor H (FH), complement factor properdin (FP), and sialic acid-binding immunoglobulinlike lectin (SIGLEC) receptors of retinal mononuclear phagocytes are key players in regulating the complement and innate immune systems in the retina by recognizing sialic acid (Sia) residues. Intact retinal sialylation prevents any long-lasting and excessive complement or immune activation in the retina. However, sialylated glycolipids are reduced in the CNS with aging, potentially contributing to chronic inflammatory processes in the retina. Particularly, genetically induced hyposialylation in mice leads to age-related, complement factor C3-mediated retinal inflammation and bipolar cell loss. Notably, most of the gene transcript pathways enriched in the mouse retina, following genetically induced hyposialylation, are also involved in age-related macular degeneration (AMD). Interestingly, intravitreal application of polysialic acid (polySia) controlled the innate immune responses in the mouse retina by blocking mononuclear phagocyte reactivity, inhibiting complement activation, and protecting against vascular damage in two different humanized SIGLEC-11 animal models. Accordingly, a polySia polymer conjugate has entered clinical phase II/III testing in patients with geographic atrophy secondary to AMD. Thus, hyposialylation or dysfunctional sialylation should be considered as an agerelated contributor to inflammatory retinal diseases, such as AMD. Consequently, sialic acid-based biologics could provide novel therapies for complement-related retinal diseases.

#### KEYWORDS

retina, sialylation, sialic acid, polysialic acid, complement, microglia, inflammation, age-related macular degeneration (AMD)

### **1** Introduction

The retina is an immune-privileged tissue with a blood–retinal barrier, composed of the microvascular endothelium and the retinal pigment epithelium (RPE) (Streilein et al., 2002; Chen et al., 2019), as well as an immune-suppressed microenvironment (Silverman and Wong, 2018; Murakami et al., 2020). However, genetic and environmental factors, such as diet and age, contribute to chronic immune responses that are associated with retinal degeneration (Amirul Islam et al., 2014; Thomas et al., 2021). In particular, age-related macular degeneration

(AMD), a leading cause of blindness in individuals above 60 years, is closely associated with retinal inflammation and oxidative stress. AMD is classified into two forms: dry and wet (Mitchell et al., 2018). Dry AMD is characterized by the accumulation of drusen and the inability of the RPE to clear them. Wet AMD is characterized by blood–retinal barrier leakage, parenchymal edema, and choroidal neovascularization (CNV) driven by the vascular endothelial growth factor (VEGF) (Thomas et al., 2021; Marchesi et al., 2024). Notably, transcriptome analyses of retinas from AMD patients revealed upregulation of genes involved in the chemokine, cytokine, and complement cascade signaling pathways (Saddala et al., 2019).

In recent years, it has been found that sialylation, a posttranslational modification of the glycocalyx termini by sialic acid residues (Schnaar et al., 2014), plays a key role in retinal complement and immune regulation. Sialic acid (Sia), the terminal carbohydrate of the glycocalyx, is found on mammalian cell surfaces in two types, N-acetylneuraminic acid (Neu5Ac) and N-glycolylneuraminic acid (Neu5Gc) (Varki et al., 2015; Pearce and Läubli, 2016). Only Neu5Ac is naturally present in humans due to a lineage-specific loss-offunction mutation in the gene encoding cytidine monophosphate-Nacetylneuraminic acid hydroxylase (CMAH) (Chou et al., 2002; Martin et al., 2005). Interestingly, several studies have shown that Neu5Gc from food sources, such as red meat, milk, and dairy products, can be incorporated into the glycocalyx of human tissue, potentially triggering immune responses against these non-human sialic acids, resulting in a condition termed xenosialitis (Oetke et al., 2001; Tangvoranuntakul et al., 2003; Hedlund et al., 2008). However, no evidence has been found of xenosialitis or Neu5Gc incorporation in the human retina. Moreover, Sias can form homopolymers with varying average degrees of polymerization (avDP) through glycosidic linkages. In mammals, polysialic acid (polySia) is a homopolymer composed of a2,8-linked Neu5Ac monomers, with a degree of polymerization ranging from 10 to approximately 100 (Schnaar et al., 2014). Sialylation can inhibit complement activation by targeting critical regulators of the complement cascade. In particular, complement factor H (FH) recognizes α2,3-linked Neu5Ac residues on the glycocalyx, thereby inhibiting the alternative complement pathway (Blaum et al., 2014). In addition, complement factor P (FP), a widely known positive regulator of the alternative complement pathway, has been shown to bind to low molecular weight a2,8-linked polySia with average degree of polymerization 20 (avDP20) and to reduce its ability to promote alternative complement activation. Sialylation also downregulates microglia activity by binding to sialic acid-binding immunoglobulin-like lectin (SIGLEC) receptors on immune cell membranes, serving as a checkpoint to maintain immune tissue homeostasis in the central nervous system (CNS; Klaus et al., 2021; Lünemann et al., 2021). However, intact sialylation is susceptible to aging and chronic inflammation. Thus, dysfunctional or reduced sialylation can disrupt retinal immune regulation, leading to excessive activation of the complement system, chronic inflammation, and tissue degeneration.

Given its essential role in retinal immune regulation, sialylation has become a promising therapeutic target for age-related and inflammatory retinal diseases (Nycholat et al., 2012; Rillahan et al., 2012; Zhong et al., 2022). Polysialylated ligands, such as polySia, have shown promising potential in preclinical studies to suppress complement activation, attenuate mononuclear phagocyte activity, and prevent vascular damage (Karlstetter et al., 2016; Krishnan et al., 2023). This review explores the role of sialylation as a checkpoint in complement-associated retinal diseases, focusing on the mechanisms underlying dysfunctional sialylation, its impact on retinal immune homeostasis, and its therapeutic potential.

# 2 Dysfunctional sialylation during aging and chronic inflammation: a trigger for retinal diseases

Sialic acid is a nine-carbon sugar that forms the terminal cap of the glycol residues of glycolipids and glycoproteins. Sias of the glycocalyx contribute to various biological processes, such as microdomain formation (Möckl et al., 2015), cell adhesion (Kelm et al., 1994), tissue homeostasis (Varki and Gagneux, 2012), immune cell modulation (Lübbers et al., 2018), cell migration (Bassagañas et al., 2014), chemokine sensing (Kiermaier et al., 2016), and growth factor retention (Sato and Kitajima, 2019).

Lower levels of sialylated gangliosides have been reported in the CNS of elderly individuals (Segler Stahl et al., 1983). In particular, the content of ganglioside-bound Sia in the whole brain of individuals aged 25 to 85 years was found to be reduced by approximately 65% at 85 years of age (Segler Stahl et al., 1983). Furthermore, an increased activity of the endogenous neuraminidases Neu1 and Neu4 was observed during inflammation, which can cleave Sias from the cell surface, suggesting that the Sia cap of the glycocalyx is compromised under pathological inflammatory conditions (Demina et al., 2018; Howlader et al., 2022). Oxidative damage has also been shown to desialylate cell surfaces (Eguchi et al., 2005; Cho et al., 2017). Furthermore, enzymatic desialylation can lead to complementdependent removal of neurons in vitro (Linnartz et al., 2012; Linnartz-Gerlach et al., 2015). Interestingly, oral supplementation with Sia ameliorated the oxidative damage in two different mouse models: a nephropathy model induced by aminoglycoside antibiotics and an oxidative stress model induced by a high-fat diet (Pawluczyk et al., 2015; Yida et al., 2015).

In mice that are heterozygous for the null mutant of UDP-Nacetylglucosamine 2-epimerase/N-acetylmannosamine kinase (Gne+/-), a crucial enzyme for Sia biosynthesis, a slight reduction in brain sialylation was noted (Klaus et al., 2020). This reduced sialylation not only led to non-inflammatory synapse and neuronal loss but was also linked to reduced microglial ramification and dependent on complement component 3 (C3) (Klaus et al., 2020). In a follow-up study focusing on the mouse retina, the number of bipolar cells in the retina of 9-month-old Gne+/- mice was reduced, which was again associated with inflammatory processes and dependent on C3 (Cuevas-Rios et al., 2024). Additionally, hyposialylation activated key biological pathways in the mouse retina at 9 months of age, including UV response, epithelialmesenchymal transition (EMT), p53 pathway, angiogenesis, apoptosis, KRAS signaling, IL6/JAK/STAT3 signaling, tumor necrosis factor-alpha (TNF-A) signaling via NFkB, cholesterol homeostasis, and complement (Cuevas-Rios et al., 2024). Interestingly, most of these enriched gene pathways have been directly or indirectly associated with AMD (Table 1).

Furthermore, several enzymes involved in sialylation and glycocalyx formation are associated with AMD (Emilsson et al., 2022). In a proteogenomic study, Emilsson et al. investigated associations between 4,782 human AMD candidate serum proteins and various disease stages of AMD and found that not

Retinal pathways enriched in mice with hyposialylation (Current for the second second	Retinal pathway contribution to AMD
UV exposure	Major cause of photoaging (Lueck et al., 2012; Sui et al., 2013)
Epithelial-mesenchymal transition	Pathogenesis of subretinal fibrosis (Kimura et al., 2015; Tuo et al., 2015; Shu et al., 2020; Shen et al., 2023)
Angiogenesis	Pathogenesis of choroidal neovascularization in wet AMD (Ferrara and Adamis, 2016; Wolf et al., 2022)
Apoptosis	Mediating RPE cell death, especially in dry AMD (Hanus et al., 2015; Lenin et al., 2023)
KRAS signaling	Promoting chronic inflammation, oxidative stress, RPE senescence, and impaired autophagy through downstream pathways such as JAK/STAT, PI3K/AKT, MAPK, and mTOR (Wang et al., 2019; Zhang et al., 2020; Ghosh et al., 2024)
IL6/JAK/STAT3 signaling	Driving chronic inflammation and establishing a pro-inflammatory feedback loop (Wiciński et al., 2021; Ahmed et al., 2024)
TNF-A signaling via NFkB	Promoting chronic inflammation and RPE cell apoptosis in dry AMD and inducing VEGF expression in wet AMD (Shi et al., 2006; Khan et al., 2021)
Cholesterol homeostasis	Accumulation of cholesterol and lipid deposits (drusen), impairing cholesterol clearance, and activation of inflammatory pathways (Pikuleva and Curcio, 2014; Ban et al., 2018)
Complement	Detailed discussion in Section 2.1.

TABLE 1 Associations between retinal hyposialylation and age-related macular degeneration (AMD) pathogenesis.

only complement and inflammation-related factors were enriched in the serum of patients at risk for early or late AMD, but also that the serum sialyltransferase (ST6 N-acetylgalactosaminide  $\alpha$ -2,6-sialyltransferase 1, ST6GALNAC1) and fucosyltransferase 5 (FUT5) were associated with early AMD (Emilsson et al., 2022). In a recent pilot study involving human retinal tissue, a reduction in Sia level at the Bruch's membrane/choroid interface in both the macular and peripheral regions of the retina in individuals with AMD (n = 3) was observed compared to controls (n = 5; Swan et al., 2025).

## 2.1 Sialylation as a protective mechanism against complement activation in the retina

The retina is particularly susceptible to complement-mediated damage due to its high metabolic activity and exposure to light-induced oxidative stress. The complement system consists of three activation pathways: classical, lectin, and alternative, all of which converge in C3 activation and lead to the formation of immunostimulatory factors such as C3a, C5a, and membrane attack complexes (MAC, C5b-9; Ricklin et al., 2010; Merle et al., 2015). As mentioned above, the loss of rod bipolar cells and the increase in inflammatory gene transcription were prevented after crossing *Gne+/–* mice that show hyposialylation with complement factor C3-deficient animals (Cuevas-Rios et al., 2024).

Complement factor H is a key regulatory protein of the alternative pathway that prevents complement overactivation by inhibiting the formation and accelerating the decay of C3 convertase (Chen et al., 2007; Kopp et al., 2012). The human FH family consists of seven highly related members, namely, FH, FH-like 1 (FHL-1), and FH-related proteins 1 to 5 (FHR-1-5). All family members are composed of repetitive units, short consensus repeats (SCRs), of ~60 amino acids, which are also termed control complement protein domains. The regulatory function of FH depends on the binding to certain polyanions, such as Sia and glycosaminoglycans (Meri and Pangburn, 1990; Clark et al., 2013). The FH at position SCR20 interacts with  $\alpha$ 2,3-conjugated Sia moieties that are critical for the discrimination of self and non-self surfaces. The binding of FH to these  $\alpha$ 2,3-conjugated Sia residues on the cytosolic glycoconjugate complex can form a C3b-FH-Sia complex, which inhibits alternative complement pathway initiation and amplification (Figure 1; Blaum et al., 2014). However, studies have shown that the rs1061170 polymorphism (Tyr402His) in the FH gene, which is closely associated with an increased risk of AMD, does not directly affect the known Sia binding domain SCR20 but instead reduces the ability of FH to bind to retinal components, such as lipid peroxides and C-reactive protein (CRP; Molins et al., 2016; Romero-Vazquez et al., 2021; Giralt et al., 2024). Data indicate that intermediate AMD is associated with decreased serum FH levels and an increased serum monomeric C-reactive protein (mCRP)/FH ratio (Figure 1; Giralt et al., 2024). These changes lead to reduced FH levels and decreased protection of sialylated retinal cell surface membranes against complement attack, as well as increased mCRP-mediated activation of downstream complement pathways (Romero-Vázquez et al., 2020).

Pentraxin-3 (PTX3) has been recently proposed to act as an anchoring site for FH in Bruch's membrane and RPE, where it limits complement-dependent inflammatory response in a mouse model of oxidative stress-induced AMD (Wang et al., 2016). PTX3 is sialylated and is known to bind FH at SCR19-20, in addition to the side SCR7 (Deban et al., 2008; Inforzato et al., 2013). Interestingly, desialylation of PTX3 allows the binding of C1q and activation of the classical complement pathway (Inforzato et al., 2006).

After reduction or loss of Sia, the underlying glycan structures can trigger classical complement pathway activation (Figure 1). Thus, the remaining desialylated glycan cell surface structures are recognized by the complement complex C1 (C1qC1r<sub>2</sub>C1s), triggering a complement cascade response via opsonization and formation of a C4b2b complex, the classical pathway C3 convertase (Linnartz et al., 2012).

It is worth noting that FP plays an opposite function to FH. FP is the only positive regulator of the alternative pathway and prevents the rapid decay of C3/C5 convertases, thereby amplifying complement activity (Figure 1; Kouser et al., 2013; Chen et al., 2018). In an *in vitro* study, it was found that low molecular weight polySia reduced the binding of serum-derived FP to the cell surface of lesioned Hepa-1c1c7 and PC-12 neuroblastoma cells. In addition, polySia can lead to reduced cell lysis and reduced formation of membrane attack complexes (Karlstetter et al., 2016). Furthermore, the addition of polySia to human serum reduced the activity of the alternative complement pathway, which was triggered



#### FIGURE 1

Homeostatic control of the complement system and mononuclear phagocytes by sialylation. (A) Mononuclear phagocytes, such as microglia and invaded macrophages, are activated near drusen in inflammatory retinal diseases [e.g., in age-related macular degeneration (AMD)]. (B) Sialic acid/Nacetylneuraminic acid (Sia /NeuAc) or polysialic acid (polySia) displayed on glycoproteins [e.g., neural cell adhesion molecule (NCAM) and CD59] and glycolipids (e.g., gangliosides) of the intact glycocalyx engage SIGLEC receptors. Inhibitory retinal microglial SIGLECs—SIGLEC-7, –9, and –11 of humans-possess inhibitory signaling ITIM domains. Ligand binding triggers ITIM phosphorylation, enabling recruitment of phosphatases SHP1/2. This phosphatase activity counteracts signaling cascades initiated by ITAM-associated transmembrane molecules (e.g., TYROBP) that transmit inducing signals of the complement factor CR3. Complement factor H (FH) binds to Sia moieties on the intact host cell glycocalyx (e.g., PTX3), thereby inhibiting the initiation of the C3-mediated alternative complement pathway and blocking assembly of the C3 convertase (C3bBb). Thus, FH actively prevents aberrant activation of the alternative complement cascade on healthy sialylated cells. In contrast, desialylated glycomolecules [e.g., Pentraxin-3 (PTX3)] are opsonized by C1, initiating the classical complement cascade and generating the classical C3 convertase (C4b2b), as indicated by the dashed arrow. The classical C3 convertase then cleaves C3 into C3b and C3a. When C3b binds weakly to the altered cells in the absence of FH binding, it can initiate and amplify the alternative complement pathway via C3bBb. FH also binds to mCRP and thereby prevents its potential to recruit C1q to the surface of damaged cells. Binding of FP to desialylated surfaces drives C3b deposition. It stabilizes the formation and activation of the alternative C3bBb, together with the complement activator properdin (C3bBbP). However, soluble polySia has been shown to reduce the activation of the alternative complement pathway via binding to the FP. In summary, loss or reduction of sialylation-due to neuraminidase activity, oxidative damage, or aging-triggers both classical and alternative complement activation, enhancing C3b deposition and the formation of C3 convertases from both pathways. CR3, complement receptor 3; FH, complement factor H; FP, complement factor properdin; GCL, ganglion cell layer; INL, inner nuclear layer; IPL, inner plexiform layer; ITAM, immunoreceptor tyrosine-based activation motif; ITIM, immunoreceptor tyrosine-based inhibition motif; mCRP monomeric C-reactive protein; NFL, nerve fiber layer; ONL, outer nuclear layer; OPL, outer plexiform layer; RPE, retinal pigment epithelium; SIGLEC, sialic acid-binding immunoglobulin-like lectins. Figure was created using CorelDraw 2019 (Corel Corporation, Ottawa, Canada).

by plate-bound lipopolysaccharides (LPS) (Shahraz et al., 2022). Thus, polySia found on the glycocalyx attached to the retinal protein neural cell adhesion molecule (NCAM) could also prevent cell membrane damage by reducing the activity of the alternative complement pathway. Importantly, this inhibitory effect was chain length-dependent, since our group recently showed that soluble monosialic or oligosialic acids failed to bind to the complement regulatory protein FH and FP at physiological concentrations (Shahraz et al., 2022).

In summary, these findings highlight the critical role of multivariant recognition of Sia residues of the cell membrane glycocalyx in maintaining homeostasis of the complement system in the retina.

### 2.2 Sialylation regulates mononuclear phagocyte homeostasis via inhibitory SIGLEC receptors

Mononuclear phagocytes include resident microglia, parenchymal macrophages, and circulating monocytes (Wieghofer et al., 2021). Microglia are uniformly distributed on the plexiform layers in the mature retina and thoroughly cover the retinal environment through their dynamically moving branching processes (Damani et al., 2011; Silverman and Wong, 2018). In response to tissue damage, the microglia are activated, and their morphology changes from ramified to amoeboid, enabling them to rapidly migrate to the site of distress and phagocytose pathogens, dead neurons, and cellular debris (Sappington et al., 2022; Brown, 2023; Ronning et al., 2025). However, this activation of immune cells is also a double-edged sword. It has been shown that activated microglia and macrophages accumulate around drusen, leading to the death of photoreceptor cells and RPE cells (Killingsworth et al., 1990; Dietrich et al., 2020). In addition, mononuclear phagocytes accumulating in the subretinal space secrete VEGF, platelet-derived growth factor- $\beta$ , fibroblast growth factor-1, fibroblast growth factor-2, and transforming growth factor- $\beta$ 1 to promote CNV (Li et al., 2017).

Sialylation regulates the mononuclear phagocytes through signaling via their SIGLECs (Linnartz-Gerlach et al., 2014; Thiesler and Hildebrandt, 2024). When sialylation functions properly, microglia remain in a homeostatic state, enabling them to selectively and silently prune misconnected neuronal dendrites (Azevedo et al., 2020; Paolicelli et al., 2022). After transient transition of microglia into an activated state to eliminate damaged or apoptotic cells, SIGLEC receptors help to resolve the inflammatory reaction, thereby preventing ongoing damage to healthy tissues (Paolicelli et al., 2022). This regulatory capacity of SIGLECs is due to the presence of their immunoreceptor tyrosinebased inhibition motifs (ITIMs) and/or ITIM-like motifs in their intracellular signaling tails (Figure 1). Transmembrane proteins with this motif in the cytoplasmic domain could have inhibitory effects by recruiting serine/threonine phosphatase tyrosine-specific Src homology-2 domain-containing phosphatase-1 (SHP-1/ PTPN6) or Src homology-2 domain-containing phosphatase-2 (SHP-2/PTPN11; Linnartz-Gerlach et al., 2014; Lünemann et al., 2021). These proteins counter-regulate other immunoreceptors harboring immunoreceptor tyrosine activation motifs (ITAMs) so that immune cell-mediated activation processes, for example, phagocytosis, oxidative burst, and inflammation, are attenuated (Crocker et al., 2007). In vitro, Siglec-E of mice recognizes  $\alpha$ 2,3and  $\alpha$ 2,8-linked Sia residues of the glycocalyx. Upon stimulation with neural debris, this recognition suppresses the phagocytosis of neural debris, downregulates the transcription of pro-inflammatory cytokine genes, and reduces the production of reactive oxygen species (Claude et al., 2013). Moreover, human SIGLEC-11 binds to α2,8-conjugated Sias of the glycocalyx and inhibits LPS-induced gene transcription of pro-inflammatory mediators in cultured mouse microglia ectopically expressing the human SIGLEC-11 receptor, thus demonstrating a neuroprotective function of this human-specific microglial SIGLEC-11 receptor (Wang and Neumann, 2010). Expression of SIGLEC-7, -9, and -11 in retinal tissue from AMD patients was found to be significantly upregulated compared with healthy controls (Krishnan et al., 2023). Interestingly, SIGLEC-7 and -9 of mononuclear phagocytes recognized Sia residues on the sialylated complement inhibitory glycoprotein CD59, thereby suppressing their immune cell attack capacity (Wen et al., 2024).

Thus, ITIM-containing SIGLECs could be a novel therapeutic target for treating inflammatory retinal diseases involving activated mononuclear phagocytes.

## 2.3 Therapeutic potential of polysialylated ligands in age-related retinal diseases

Polysialic acid is an extended homopolymer of  $\alpha 2,8$ -conjugated Sia that is attached to several glycoproteins, such as neural cell adhesion molecule (NCAM), synCAM-1, neuropilin -2 (NRP2), and CD36, on the surface of mammalian neurons and immune cells (Sato and Kitajima, 2013). In a mouse model of laser-induced retinal injury, intravitreal application of polySia avDP20 reduced the mononuclear phagocyte activation, vascular leakage, and membrane attack complex deposition in humanized SIGLEC-11 transgenic mice (Karlstetter et al., 2016). In addition, *in vitro* studies showed that polySia avDP20 inhibited mononuclear phagocyte reactivity via the SIGLEC receptor and directly interfered with the activation of the alternative complement system (Karlstetter et al., 2016).

Polysialic acid-functionalized nanoparticles (polySia-NPs), in which polySia was conjugated to a core of polyethylene glycol and poly lactic-co-glycolic acid (PLGA) copolymers, were effective in reducing macrophage-induced inflammatory retinal responses after intravitreal application in humanized SIGLEC-11 transgenic mice (Krishnan et al., 2023). In a mouse model of bright light retinal damage, polySia-NPs bound to SIGLEC-E receptors on macrophages and elicited an anti-inflammatory effect with a reduction in IL-6, IL-1 $\beta$ , and TNF- $\alpha$  levels, as well as diminished loss of retinal outer nuclear layer thickness (Krishnan et al., 2023).

The proven *in vivo* safety and efficacy of polySia-NPs in treating animal models of AMD led to an investigational new drug (IND) application at the US Food and Drug Administration (FDA) and received approval for entry into human Phase II/III clinical trials for the treatment of geographic atrophy in AMD patients (Krishnan et al., 2024; SIGLEC study ClinicalTrials.gov Identifier NCT05839041).

## **3** Conclusion

Sialylation plays a critical role in maintaining retinal immune homeostasis and protecting against complement-mediated damage. Age-related hyposialylation disrupts this protective process, contributing to the pathogenesis of retinal diseases, such as AMD. The interplay between hyposialylation and complement activation creates a vicious cycle of chronic inflammation and degeneration, highlighting the need for therapies that target both processes simultaneously. Sia-based polymers, such as polySia, therefore represent a promising therapeutic approach for complement-related retinal diseases.

### Author contributions

YM: Visualization, Writing – review & editing, Writing – original draft. GC-R: Writing – review & editing, Writing – original draft. TL: Writing – review & editing, Writing – original draft. HN: Writing – review & editing, Writing – original draft.

## Funding

The author(s) declare that financial support was received for the research and/or publication of this article. HN is supported by the Deutsche Forschungsgemeinschaft (DFG; German Research Foundation) via FOR2953 (number 409784463) and SPP2395 (number 500260917). This publication was supported by the Open Access Publication Fund of the University of Bonn.

## **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## **Generative AI statement**

The author(s) declare that no Gen AI was used in the creation of this manuscript.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

### References

Ahmed, C. M., Johnson, H. M., and Lewin, A. S. (2024). Corneal application of SOCS1/3 peptides for the treatment of eye diseases mediated by inflammation and oxidative stress. *Front. Immunol.* 15:1416181. doi: 10.3389/FIMMU.2024.1416181

Amirul Islam, F. M., Chong, E. W., Hodge, A. M., Guymer, R. H., Aung, K. Z., Makeyeva, G. A., et al. (2014). Dietary patterns and their associations with age-related macular degeneration: the Melbourne collaborative cohort study. *Ophthalmology* 121, 1428–1434.e2. doi: 10.1016/J.OPHTHA.2014.01.002

Azevedo, M. D., Sander, S., and Tenenbaum, L. (2020). GDNF, a neuron-derived factor upregulated in glial cells during disease. J. Clin. Med. 9:456. doi: 10.3390/JCM9020456

Ban, N., Lee, T. J., Sene, A., Choudhary, M., Lekwuwa, M., Dong, Z., et al. (2018). Impaired monocyte cholesterol clearance initiates age-related retinal degeneration and vision loss. *JCI Insight* 3:120824. doi: 10.1172/JCI.INSIGHT.120824

Bassagañas, S., Pérez-Garay, M., and Peracaula, R. (2014). Cell surface sialic acid modulates extracellular matrix adhesion and migration in pancreatic adenocarcinoma cells. *Pancreas* 43, 109–117. doi: 10.1097/MPA.0B013E31829D9090

Blaum, B. S., Hannan, J. P., Herbert, A. P., Kavanagh, D., Uhrín, D., and Stehle, T. (2014). Structural basis for sialic acid-mediated self-recognition by complement factor H. *Nat. Chem. Biol.* 11, 77–82. doi: 10.1038/nchembio.1696

Brown, G. C. (2023). Cell death by phagocytosis. Nat. Rev. Immunol. 24, 91–102. doi: 10.1038/s41577-023-00921-6

Chen, J. Y., Cortes, C., and Ferreira, V. P. (2018). Properdin: a multifaceted molecule involved in inflammation and diseases. *Mol. Immunol.* 102, 58–72. doi: 10.1016/J.MOLIMM.2018.05.018

Chen, M., Forrester, J. V., and Xu, H. (2007). Synthesis of complement factor H by retinal pigment epithelial cells is down-regulated by oxidized photoreceptor outer segments. *Exp. Eye Res.* 84, 635–645. doi: 10.1016/J.EXER.2006.11.015

Chen, M., Luo, C., Zhao, J., Devarajan, G., and Xu, H. (2019). Immune regulation in the aging retina. *Prog. Retin. Eye Res.* 69, 159–172. doi: 10.1016/j.preteyeres.2018.10.003

Cho, A., Christine, M., Malicdan, V., Miyakawa, M., Nonaka, I., Nishino, I., et al. (2017). Sialic acid deficiency is associated with oxidative stress leading to muscle atrophy and weakness in GNE myopathy. *Hum. Mol. Genet.* 26, 3081–3093. doi: 10.1093/HMG/DDX192

Chou, H. H., Hayakawa, T., Diaz, S., Krings, M., Indriati, E., Leakey, M., et al. (2002). Inactivation of CMP-N-acetylneuraminic acid hydroxylase occurred prior to brain expansion during human evolution. *Proc. Natl. Acad. Sci. USA* 99, 11736–11741. doi: 10.1073/PNAS.182257399

Clark, S. J., Ridge, L. A., Herbert, A. P., Hakobyan, S., Mulloy, B., Lennon, R., et al. (2013). Tissue-specific host recognition by complement factor H is mediated by differential activities of its glycosaminoglycan-binding regions. *J. Immunol.* 190, 2049–2057. doi: 10.4049/JIMMUNOL.1201751

Claude, J., Linnartz-Gerlach, B., Kudin, A. P., Kunz, W. S., and Neumann, H. (2013). Microglial CD33-related Siglec-E inhibits neurotoxicity by preventing the phagocytosisassociated oxidative burst. *J. Neurosci.* 33, 18270–18276. doi: 10.1523/JNEUROSCI. 2211-13.2013

Crocker, P. R., Paulson, J. C., and Varki, A. (2007). Siglecs and their roles in the immune system. *Nat. Rev. Immunol.* 7, 255–266. doi: 10.1038/nri2056

Cuevas-Rios, G., Assale, T. A., Wissfeld, J., Bungartz, A., Hofmann, J., Langmann, T., et al. (2024). Decreased sialylation elicits complement-related microglia response and bipolar cell loss in the mouse retina. *Glia* 72, 2295–2312. doi: 10.1002/GLIA.24613

Damani, M. R., Zhao, L., Fontainhas, A. M., Amaral, J., Fariss, R. N., and Wong, W. T. (2011). Age-related alterations in the dynamic behavior of microglia. *Aging Cell* 10, 263–276. doi: 10.1111/J.1474-9726.2010.00660.X

Deban, L., Jarva, H., Lehtinen, M. J., Bottazzi, B., Bastone, A., Doni, A., et al. (2008). Binding of the long Pentraxin PTX3 to factor H: interacting domains and function in the regulation of complement activation. *J. Immunol.* 181, 8433–8440. doi: 10.4049/JIMMUNOL.181.12.8433

Demina, E. P., Pierre, W. C., Nguyen, A. L. A., Londono, I., Reiz, B., Zou, C., et al. (2018). Persistent reduction in sialylation of cerebral glycoproteins following postnatal inflammatory exposure. *J. Neuroinflammation* 15, 1–14. doi: 10.1186/S12974-018-1367-2

Dietrich, L., Lucius, R., Roider, J., and Klettner, A. (2020). Interaction of inflammatorily activated retinal pigment epithelium with retinal microglia and neuronal cells. *Exp. Eye Res.* 199:108167. doi: 10.1016/J.EXER.2020.108167

Eguchi, H., Ikeda, Y., Ookawara, T., Koyota, S., Fujiwara, N., Honke, K., et al. (2005). Modification of oligosaccharides by reactive oxygen species decreases sialyl Lewis x-mediated cell adhesion. *Glycobiology* 15, 1094–1101. doi: 10.1093/GLYCOB/CWJ003 organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Emilsson, V., Gudmundsson, E. F., Jonmundsson, T., Jonsson, B. G., Twarog, M., Gudmundsdottir, V., et al. (2022). A proteogenomic signature of age-related macular degeneration in blood. *Nat. Commun.* 13, 1–15. doi: 10.1038/s41467-022-31085-x

Ferrara, N., and Adamis, A. P. (2016). Ten years of anti-vascular endothelial growth factor therapy. *Nat. Rev. Drug Discov.* 15, 385–403. doi: 10.1038/nrd.2015.17

Ghosh, S., Hose, S., and Sinha, D. (2024). AKT2-mediated lysosomal dysfunction promotes secretory autophagy in retinal pigment epithelium (RPE) cells. *Autophagy* 20, 2841–2842. doi: 10.1080/15548627.2024.2413305

Giralt, L., Figueras-Roca, M., Eguileor, B. D. L., Romero, B., Zarranz-Ventura, J., Alforja, S., et al. (2024). C-reactive protein-complement factor H axis as a biomarker of activity in early and intermediate age-related macular degeneration. *Front. Immunol.* 15:1330913. doi: 10.3389/FIMMU.2024.1330913

Hanus, J., Anderson, C., and Wang, S. (2015). RPE necroptosis in response to oxidative stress and in AMD. Ageing Res. Rev. 24, 286–298. doi: 10.1016/J.ARR.2015.09.002

Hedlund, M., Padler-Karavani, V., Varki, N. M., and Varki, A. (2008). Evidence for a human-specific mechanism for diet and antibody-mediated inflammation in carcinoma progression. *Proc. Natl. Acad. Sci. USA* 105, 18936–18941. doi: 10.1073/PNAS. 0803943105

Howlader, M. A., Demina, E. P., Samarani, S., Guo, T., Caillon, A., Ahmad, A., et al. (2022). The Janus-like role of neuraminidase isoenzymes in inflammation. *FASEB J.* 36:e22285. doi: 10.1096/FJ.202101218R

Inforzato, A., Peri, G., Doni, A., Garlanda, C., Mantovani, A., Bastone, A., et al. (2006). Structure and function of the long pentraxin PTX3 glycosidic moiety: fine-tuning of the interaction with C1q and complement activation. *Biochemistry* 45, 11540–11551. doi: 10.1021/BI0607453

Inforzato, A., Reading, P. C., Barbati, E., Bottazzi, B., Garlanda, C., and Mantovani, A. (2013). The "sweet" side of a long pentraxin: how glycosylation affects PTX3 functions in innate immunity and inflammation. *Front. Immunol.* 3, 371–315. doi: 10.3389/FIMMU.2012.00407

Karlstetter, M., Kopatz, J., Aslanidis, A., Shahraz, A., Caramoy, A., Linnartz-Gerlach, B., et al. (2016). Polysialic acid blocks mononuclear phagocyte reactivity, inhibits complement activation, and protects from vascular damage in the retina. *EMBO Mol. Med.* 9:154. doi: 10.15252/EMMM.201606627

Kelm, S., Schauer, R., Manuguerra, J. C., Gross, H. J., and Crocker, P. R. (1994). Modifications of cell surface sialic acids modulate cell adhesion mediated by sialoadhesin and CD22. *Glycoconj. J.* 11, 576–585. doi: 10.1007/BF00731309

Khan, A. H., Pierce, C. O., De Salvo, G., Griffiths, H., Nelson, M., Cree, A. J., et al. (2021). The effect of systemic levels of TNF-alpha and complement pathway activity on outcomes of VEGF inhibition in neovascular AMD. *Eye* 36, 2192–2199. doi: 10.1038/\$41433-021-01824-3

Kiermaier, E., Moussion, C., Veldkamp, C. T., Gerardy-Schahn, R., De Vries, I., Williams, L. G., et al. (2016). Polysialylation controls dendritic cell trafficking by regulating chemokine recognition. *Science* 351, 186–190. doi: 10.1126/SCIENCE.AAD0512

Killingsworth, M. C., Sarks, J. P., and Sarks, S. H. (1990). Macrophages related to Bruch's membrane in age-related macular degeneration. *Eye* 4, 613–621. doi: 10.1038/eye.1990.86

Kimura, K., Orita, T., Liu, Y., Yang, Y., Tokuda, K., Kurakazu, T., et al. (2015). Attenuation of EMT in RPE cells and subretinal fibrosis by an RAR-γ agonist. *J. Mol. Med.* (*Berl*) 93, 749–758. doi: 10.1007/S00109-015-1289-8

Klaus, C., Hansen, J. N., Ginolhac, A., Gérard, D., Gnanapragassam, V. S., Horstkorte, R., et al. (2020). Reduced sialylation triggers homeostatic synapse and neuronal loss in middle-aged mice. *Neurobiol. Aging* 88, 91–107. doi: 10.1016/J.NEUROBIOLAGING.2020.01.008

Klaus, C., Liao, H., Allendorf, D. H., Brown, G. C., and Neumann, H. (2021). Sialylation acts as a checkpoint for innate immune responses in the central nervous system. *Glia* 69, 1619–1636. doi: 10.1002/GLIA.23945

Kopp, A., Hebecker, M., Svobodová, E., and Józsi, M. (2012). Factor H: a complement regulator in health and disease, and a mediator of cellular interactions. *Biomol. Ther.* 2, 46–75. doi: 10.3390/BIOM2010046

Kouser, L., Abdul-Aziz, M., Nayak, A., Stover, C. M., Sim, R. B., and Kishore, U. (2013). Properdin and factor H: opposing players on the alternative complement pathway "see-saw". *Front. Immunol.* 4:47548. doi: 10.3389/FIMMU.2013.00093

Krishnan, A., Callanan, D. G., Sendra, V. G., Lad, A., Christian, S., Earla, R., et al. (2024). Comprehensive ocular and systemic safety evaluation of polysialic acid-decorated immune modulating therapeutic nanoparticles (PolySia-NPs) to support entry into first-in-human clinical trials. *Pharmaceuticals* 17:481. doi: 10.3390/PH17040481

Krishnan, A., Sendra, V. G., Patel, D., Lad, A., Greene, M. K., Smyth, P., et al. (2023). Polysialic acid-nanoparticles inhibit macrophage mediated inflammation through Siglec agonism: a potential treatment for age related macular degeneration. *Front. Immunol.* 14:1237016. doi: 10.3389/FIMMU.2023.1237016

Lenin, R. R., Koh, Y. H., Zhang, Z., Yeo, Y. Z., Parikh, B. H., Seah, I., et al. (2023). Dysfunctional autophagy, Proteostasis, and mitochondria as a prelude to age-related macular degeneration. *Int. J. Mol. Sci.* 24:8763. doi: 10.3390/IJMS24108763

Li, L., Heiduschka, P., Alex, A. F., Niekämper, D., and Eter, N. (2017). Behaviour of CD11b-positive cells in an animal model of laser-induced choroidal neovascularisation. *Ophthalmologica* 237, 29–41. doi: 10.1159/000453550

Linnartz, B., Kopatz, J., Tenner, A. J., and Neumann, H. (2012). Sialic acid on the neuronal Glycocalyx prevents complement C1 binding and complement Receptor-3-mediated removal by microglia. *J. Neurosci.* 32, 946–952. doi: 10.1523/JNEUROSCI. 3830-11.2012

Linnartz-Gerlach, B., Mathews, M., and Neumann, H. (2014). Sensing the neuronal glycocalyx by glial sialic acid binding immunoglobulin-like lectins. *Neuroscience* 275, 113–124. doi: 10.1016/J.NEUROSCIENCE.2014.05.061

Linnartz-Gerlach, B., Schuy, C., Shahraz, A., Tenner, A. J., and Neumann, H. (2015). Sialylation of neurites inhibits complement-mediated macrophage removal in a human macrophage-neuron co-culture system. *Glia* 64:35. doi: 10.1002/GLIA. 22901

Lübbers, J., Rodríguez, E., and van Kooyk, Y. (2018). Modulation of immune tolerance via Siglec-sialic acid interactions. *Front. Immunol.* 9:422533. doi: 10.3389/FIMMU. 2018.02807

Lueck, K., Hennig, M., Lommatzsch, A., Pauleikhoff, D., and Wasmuth, S. (2012). Complement and UV-irradiated photoreceptor outer segments increase the cytokine secretion by retinal pigment epithelial cells. *Invest. Ophthalmol. Vis. Sci.* 53, 1406–1413. doi: 10.1167/IOVS.11-8889

Lünemann, J. D., von Gunten, S., and Neumann, H. (2021). Targeting sialylation to treat central nervous system diseases. *Trends Pharmacol. Sci.* 42, 998–1008. doi: 10.1016/J.TIPS.2021.09.002

Marchesi, N., Capierri, M., Pascale, A., and Barbieri, A. (2024). Different therapeutic approaches for dry and wet AMD. *Int. J. Mol. Sci.* 25:13053. doi: 10.3390/IJMS252313053

Martin, M. J., Rayner, J. C., Gagneux, P., Barnwell, J. W., and Varki, A. (2005). Evolution of human-chimpanzee differences in malaria susceptibility: relationship to human genetic loss of N-glycolylneuraminic acid. *Proc. Natl. Acad. Sci. USA* 102, 12819–12824. doi: 10.1073/PNAS.0503819102

Meri, S., and Pangburn, M. K. (1990). Discrimination between activators and nonactivators of the alternative pathway of complement: regulation via a sialic acid/ polyanion binding site on factor H. *Proc. Natl. Acad. Sci. USA* 87, 3982–3986.

Merle, N. S., Noe, R., Halbwachs-Mecarelli, L., Fremeaux-Bacchi, V., and Roumenina, L. T. (2015). Complement system part II: role in immunity. *Front. Immunol.* 6:136998. doi: 10.3389/FIMMU.2015.00257

Mitchell, P., Liew, G., Gopinath, B., and Wong, T. Y. (2018). Age-related macular degeneration. *Lancet* 392, 1147–1159. doi: 10.1016/S0140-6736(18)31550-2

Möckl, L., Horst, A. K., Kolbe, K., Lindhorst, T. K., and Bräuchle, C. (2015). Microdomain formation controls spatiotemporal dynamics of cell-surface glycoproteins. *Chembiochem* 16, 2023–2028. doi: 10.1002/CBIC.201500361

Molins, B., Fuentes-Prior, P., Adán, A., Antón, R., Arostegui, J. I., Yagüe, J., et al. (2016). Complement factor H binding of monomeric C-reactive protein downregulates proinflammatory activity and is impaired with at risk polymorphic CFH variants. *Sci. Rep.* 6, 1–12. doi: 10.1038/srep22889

Murakami, Y., Ishikawa, K., Nakao, S., and Sonoda, K. H. (2020). Innate immune response in retinal homeostasis and inflammatory disorders. *Prog. Retin. Eye Res.* 74:100778. doi: 10.1016/J.PRETEYERES.2019.100778

Nycholat, C. M., Rademacher, C., Kawasaki, N., and Paulson, J. C. (2012). In silicoaided design of a glycan ligand of sialoadhesin for in vivo targeting of macrophages. *J. Am. Chem. Soc.* 134, 15696–15699. doi: 10.1021/JA307501E

Oetke, C., Hinderlich, S., Brossmer, R., Reutter, W., Pawlita, M., and Keppler, O. T. (2001). Evidence for efficient uptake and incorporation of sialic acid by eukaryotic cells. *Eur. J. Biochem.* 268, 4553–4561. doi: 10.1046/J.1432-1327.2001.02379.X

Paolicelli, R. C., Sierra, A., Stevens, B., Tremblay, M. E., Aguzzi, A., Ajami, B., et al. (2022). Microglia states and nomenclature: a field at its crossroads. *Neuron* 110, 3458–3483. doi: 10.1016/J.NEURON.2022.10.020

Pawluczyk, I. Z. A., Najafabadi, M. G., Brown, J. R., Bevington, A., and Topham, P. S. (2015). Sialic acid supplementation ameliorates puromycin aminonucleoside nephrosis in rats. *Lab. Investig.* 95, 1019–1028. doi: 10.1038/LABINVEST.2015.78

Pearce, O. M. T., and Läubli, H. (2016). Sialic acids in cancer biology and immunity. *Glycobiology* 26, 111–128. doi: 10.1093/GLYCOB/CWV097

Pikuleva, I. A., and Curcio, C. A. (2014). Cholesterol in the retina: the best is yet to come. *Prog. Retin. Eye Res.* 41, 64–89. doi: 10.1016/J.PRETEYERES.2014.03.002

Ricklin, D., Hajishengallis, G., Yang, K., and Lambris, J. D. (2010). Complement: a key system for immune surveillance and homeostasis. *Nat. Immunol.* 11, 785–797. doi: 10.1038/ni.1923

Rillahan, C. D., Schwartz, E., McBride, R., Fokin, V. V., and Paulson, J. C. (2012). 'Click and pick' identification of high affinity Sialoside ligands for Siglec-based targeting of leukocytes. *Angew. Chem. Int. Ed. Eng.* 51:11014. doi: 10.1002/ANIE.201205831

Romero-Vázquez, S., Adán, A., Figueras-Roca, M., Llorenç, V., Slevin, M., Vilahur, G., et al. (2020). Activation of C-reactive protein proinflammatory phenotype in the blood retinal barrier in vitro: implications for age-related macular degeneration. *Aging (Albany NY)* 12, 13905–13923. doi: 10.18632/AGING.103655

Romero-vazquez, S., Llorens, V., Soler-boronat, A., Figueras-roca, M., Adan, A., and Molins, B. (2021). Interlink between inflammation and oxidative stress in age-related macular degeneration: role of complement factor H. *Biomedicine* 9:763. doi: 10.3390/BIOMEDICINES9070763

Ronning, K. E., Burns, M. E., and Sennlaub, F. (2025). Monocytes in retinal degeneration: little cells with a big impact. *Adv. Exp. Med. Biol.* 1468, 133–137. doi: 10.1007/978-3-031-76550-6\_22

Saddala, M. S., Lennikov, A., Mukwaya, A., Fan, L., Hu, Z., and Huang, H. (2019). Transcriptome-wide analysis of differentially expressed chemokine receptors, SNPs, and SSRs in the age-related macular degeneration. *Hum. Genomics* 13:15. doi: 10.1186/S40246-019-0199-1

Sappington, R. M., Joachim, S. C., Freude, K., Guo, L., Choi, S., Bikkannavar, P., et al. (2022). Microglia: key players in retinal ageing and neurodegeneration. *Front. Cell. Neurosci.* 16:804782. doi: 10.3389/FNCEL.2022.804782

Sato, C., and Kitajima, K. (2013). Disialic, oligosialic and polysialic acids: distribution, functions and related disease. J. Biochem. 154, 115–136. doi: 10.1093/JB/MVT057

Sato, C., and Kitajima, K. (2019). Sialic acids in neurology. Adv. Carbohydr. Chem. Biochem. 76, 1–64. doi: 10.1016/BS.ACCB.2018.09.003

Schnaar, R. L., Gerardy-Schahn, R., and Hildebrandt, H. (2014). Sialic acids in the brain: gangliosides and polysialic acid in nervous system development, stability, disease, and regeneration. *Physiol. Rev.* 94, 461–518. doi: 10.1152/PHYSREV.00033.2013

Segler Stahl, K., Webster, J. C., and Brunngraber, E. G. (1983). Changes in the concentration and composition of human brain gangliosides with aging. *Gerontology* 29, 161–168.

Shahraz, A., Lin, Y., Mbroh, J., Winkler, J., Liao, H., Lackmann, M., et al. (2022). Low molecular weight polysialic acid binds to properdin and reduces the activity of the alternative complement pathway. *Sci. Rep.* 12, 1–12. doi: 10.1038/s41598-022-09407-2

Shen, G., Li, Y., Zeng, Y., Hong, F., Zhang, J., Wang, Y., et al. (2023). Kallistatin deficiency induces the oxidative stress-related epithelial-mesenchymal transition of retinal pigment epithelial cells: a novel protagonist in age-related macular degeneration. *Invest. Ophthalmol. Vis. Sci.* 64:15. doi: 10.1167/IOVS.64.12.15

Shi, X., Semkova, I., Müther, P. S., Dell, S., Kociok, N., and Joussen, A. M. (2006). Inhibition of TNF-alpha reduces laser-induced choroidal neovascularization. *Exp. Eye Res.* 83, 1325–1334. doi: 10.1016/J.EXER.2006.07.007

Shu, D. Y., Butcher, E., and Saint-Geniez, M. (2020). EMT and EndMT: emerging roles in age-related macular degeneration. *Int. J. Mol. Sci.* 21, 1–26. doi: 10.3390/IJMS21124271

Silverman, S. M., and Wong, W. T. (2018). Microglia in the retina: roles in development, maturity, and disease. *Annu Rev Vis Sci* 4, 45–77. doi: 10.1146/ANNU REV-VISION-091517-034425

Streilein, J. W., Ma, N., Wenkel, H., Fong Ng, T., and Zamiri, P. (2002). Immunobiology and privilege of neuronal retina and pigment epithelium transplants. *Vis. Res.* 42, 487–495. doi: 10.1016/S0042-6989(01)00185-7

Sui, G. Y., Liu, G. C., Liu, G. Y., Gao, Y. Y., Deng, Y., Wang, W. Y., et al. (2013). Is sunlight exposure a risk factor for age-related macular degeneration? A systematic review and meta-analysis. *Br. J. Ophthalmol.* 97, 389–394. doi: 10.1136/BJOPHT HALMOL-2012-302281

Swan, J., Toomey, C. B., Bergstrand, M., Cuello, H. A., Robie, J., Yu, H., et al. (2025). The sialome of the retina, alteration in age-related macular degeneration (AMD) pathology and potential impacts on complement factor H. *bioRxiv*. doi: 10.1101/2025.03.09.642149

Tangvoranuntakul, P., Gagneux, P., Diaz, S., Bardor, M., Varki, N., Varki, A., et al. (2003). Human uptake and incorporation of an immunogenic nonhuman dietary sialic acid. *Proc. Natl. Acad. Sci. USA* 100, 12045–12050. doi: 10.1073/PNAS.2131556100

Thiesler, H., and Hildebrandt, H. (2024). Polysialic acid-Siglec immune checkpoints of microglia and macrophages: perspectives for therapeutic intervention. *Neural Regen. Res.* 21, 661–662. doi: 10.4103/NRR.NRR-D-24-01195

Thomas, C. J., Mirza, R. G., and Gill, M. K. (2021). Age-related macular degeneration. *Med. Clin. North Am.* 105, 473–491. doi: 10.1016/J.MCNA.2021.01.003

Tuo, J., Wang, Y., Cheng, R., Li, Y., Chen, M., Qiu, F., et al. (2015). Wnt signaling in age-related macular degeneration: human macular tissue and mouse model. *J. Transl. Med.* 13:330. doi: 10.1186/S12967-015-0683-X

Varki, A., and Gagneux, P. (2012). Multifarious roles of sialic acids in immunity. *Ann.* N. Y. Acad. Sci. 1253, 16–36. doi: 10.1111/J.1749-6632.2012.06517.X

Varki, A., Schnaar, R. L., and Schauer, R. (2015). Sialic acids and other nonulosonic acids. essentials of glycobiology (3rd ed.). Cold Spring Harbor (NY): Cold Spring Harbor Laboratory Press. 179–195.

Wang, L., Cano, M., Datta, S., Wei, H., Ebrahimi, K. B., Gorashi, Y., et al. (2016). Pentraxin 3 recruits complement factor H to protect against oxidative stress-induced complement and inflammasome overactivation. *J. Pathol.* 240, 495–506. doi: 10.1002/PATH.4811

Wang, Y., and Neumann, H. (2010). Alleviation of neurotoxicity by microglial human Siglec-11. J. Neurosci. 30, 3482–3488. doi: 10.1523/JNEUROSCI.3940-09.2010

Wang, S., Wang, X., Cheng, Y., Ouyang, W., Sang, X., Liu, J., et al. (2019). Autophagy dysfunction, cellular senescence, and abnormal immune-inflammatory responses in AMD: from mechanisms to therapeutic potential. *Oxidative Med. Cell. Longev.* 2019;3632169. doi: 10.1155/2019/3632169

Wen, R. M., Stark, J. C., Marti, G. E. W., Fan, Z., Lyu, A., Marques, F. J. G., et al. (2024). Sialylated glycoproteins suppress immune cell killing by binding to Siglec-7 and Siglec-9 in prostate cancer. *J. Clin. Invest.* 134:180282. doi: 10.1172/JCI180282

Wiciński, M., Seredyka-Burduk, M., Liberski, S., Marczak, D., Pol, M., Malinowski, B., et al. (2021). Evaluation of blood coagulation parameters and ADMA, NO, IL-6, and IL-18 serum levels in patients with Neovascular AMD before, during, and after the initial loading phase of intravitreal Aflibercept. *Life (Basel)* 11:441. doi: 10.3390/LIFE11050441

Wieghofer, P., Hagemeyer, N., Sankowski, R., Schlecht, A., Staszewski, O., Amann, L., et al. (2021). Mapping the origin and fate of myeloid cells in distinct compartments of the eye by single-cell profiling. *EMBO J.* 40:e105123. doi: 10.15252/EMBJ.2020105123

Wolf, A. T., Harris, A., Oddone, F., Siesky, B., Verticchio Vercellin, A., and Ciulla, T. A. (2022). Disease progression pathways of wet AMD: opportunities for new target discovery. *Expert Opin. Ther. Targets* 26, 5–12. doi: 10.1080/14728222.2022.2030706

Yida, Z., Imam, M. U., Ismail, M., Ismail, N., Ideris, A., and Abdullah, M. A. (2015). High fat diet-induced inflammation and oxidative stress are attenuated by N-acetylneuraminic acid in rats. *J. Biomed. Sci.* 22, 1–10. doi: 10.1186/S12929-015-0211-6

Zhang, M., Jiang, N., Chu, Y., Postnikova, O., Varghese, R., Horvath, A., et al. (2020). Dysregulated metabolic pathways in age-related macular degeneration. *Sci. Rep.* 10:2464. doi: 10.1038/S41598-020-59244-4

Zhong, X., D'antona, A. M., Scarcelli, J. J., and Rouse, J. C. (2022). New opportunities in glycan engineering for therapeutic proteins. *Antibodies* 11:5. doi: 10.3390/ANTIB11010005