



# Scaffold Searching of FDA and EMA-Approved Drugs Identifies Lead Candidates for Drug Repurposing in Alzheimer's Disease

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Shityakov S, Skorb EV, Förster CY and Dandekar T (2021) Scaffold Searching of FDA and EMA-Approved Drugs Identifies Lead Candidates for Drug Repurposing in Alzheimer's Disease. Front. Chem. 9:736509. doi: 10.3389/fchem.2021.736509 Clinical trials of novel therapeutics for Alzheimer's Disease (AD) have consumed a significant amount of time and resources with largely negative results. Repurposing drugs already approved by the Food and Drug Administration (FDA), European Medicines Agency (EMA), or Worldwide for another indication is a more rapid and less expensive option. Therefore, we apply the scaffold searching approach based on known amyloid-beta (AB) inhibitor tramiprosate to screen the DrugCentral database (n = 4,642) of clinically tested drugs. As a result, menadione bisulfite and camphotamide substances with protrombogenic and neurostimulation/ cardioprotection effects were identified as promising AB inhibitors with an improved binding affinity ( $\Delta G bind$ ) and blood-brain barrier permeation (logBB). Finally, the data was also confirmed by molecular dynamics simulations using implicit solvation, in particular as Molecular Mechanics Generalized Born Surface Area (MM-GBSA) model. Overall, the proposed in silico pipeline can be implemented through the early stage rational drug design to nominate some lead candidates for AD, which will be further validated in vitro and in vivo, and, finally, in a clinical trial.

Keywords: scaffold search, approved drugs, drug repurposing, alzheimer's disease, chemical similarity, molecular modeling

# INTRODUCTION

Alzheimer's Disease (AD) is a progressive neurodegenerative disorder, causing memory loss and in 60–70% of cases leading to dementia (Caltagirone et al., 1983). The AD pathology is widely believed to be associated with the production of  $\beta$ -amyloid peptide (A $\beta$ ), which is responsible for the plaque formations in the brain, disrupting normal neuronal functions (Takahashi et al., 2017). This pathological hallmark of AD might be considered in rational drug design and discovery as a promising therapeutic target to develop effective medication against this disorder. However, there is still a lack of efficient treatment for this disabling and ultimately fatal disease, e.g., donepezil and memantine usually provide at best only temporary and

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incomplete symptomatic relief (Nie et al., 2011). Moreover, various attempts had been made to use different drug binding sites (subregion-targets) in A $\beta$  that could stop its aggregation and formation of a senile plague (Nie et al., 2011). In particular, tramiprosate (3-Aminopropanesulfonic acid, TRA), a mimic of glycosaminoglycans, targets the HHQK subregion at the N-terminus of AB (Nie et al., 2011). The TRA treatment of TgCRND8 mice resulted in a 30% reduction in the brain plaque load and the same decrease in the cerebral levels of soluble and insoluble AB (Gervais et al., 2007). Additionally, a dose-dependent 60% reduction of plasma AB levels was also observed, suggesting that this influences the AB central pool, changing either its efflux or its metabolism in the brain (Gervais et al., 2007). Despite the structural simplicity, high specificity, and excellent in vivo AB inhibition properties of TRA, it subsequently failed in the late stages of phase III clinical trial (Rauk, 2008). However, the data obtained from in vitro/vivo experiments and clinical trials could provide valuable evidence that AB inhibitors and their scaffolds represent a viable drug designing methodology for AD treatment (Blazer and Neubig, 2009). Indeed, some scaffold-based techniques, such as scaffold hopping, have become a powerful tool to determine the most promising druglike candidates and already approved drugs in a drug repurposing protocol for AD (Kowal et al., 2019; Ballard et al., 2020). For example, some researchers had already performed in silico screening of a virtual library of a biaryl scaffold-containing compounds to inhibit the primary targets of AD therapeutics, such as acetylcholinesterase, β-secretase, monoamine oxidases, and N-methyl-D-aspartate receptor (Khalid et al., 2018). On the other hand, the scaffold searching of FDA and EMA-approved drug libraries was not previously performed to identify lead candidates for drug repurposing in AD. Moreover, the blood-brain Barrier (BBB) permeation of drug-like molecules is often considered in various pharmacokinetics (PK) studies as a pivotal PK-related descriptor (Shityakov et al., 2012; Carpenter et al., 2014). Therefore, in our study, we wanted to apply the molecular search based on the TRA

scaffold to examine a database of FDA and EMA-approved drugs using *in silico* computational approaches, such as virtual screening, molecular dynamics, and BBB-based descriptor analyses.

# **COMPUTATIONAL METHODS**

The 3D molecular structure (PBD ID: 2NAO) of a disease-relevant Aß fibril (1-42), containing chains A, B, and C, was downloaded from the Protein Data Bank (Walti et al., 2016) to be used in the study. The Ramachandran plot server (https://zlab.umassmed.edu/ bu/rama/index.pl) was implemented for the stereochemical validation of the receptor molecule to investigate the  $\phi - \psi$ dihedral angles in a Ramachandran plot. Altogether, observed statistics showed that 86.54% (90 residues) and 11.54% (12 residues) of all observed residues were in the core and allowed regions. Additionally, no steric clashes were detected in the peptide structure. The TRA molecule and its propanesulfonic scaffold were built by using the MarvinSketch software (ChemAxon, Hungary). A database, containing the FDA and EMA-approved drugs (n = 4,642), was obtained from the DrugCentral 2021 online drug compendium. The Molsoft ICM 3.8-3 scaffold search algorithm was used to filter the database to identify scaffold-containing drugs. All ligands were protonated at pH = 7.4 and T = 310 K using the MOE software. Prior to molecular docking, the CASTp (Computed Atlas of Surface Topology of proteins) algorithm (Naghibzadeh, 2001) was implemented to detect the location of the peptide-ligand binding site with Cartesian coordinates located at the grid center: x = 11.87 Å; y = 18.88 Å; z = -27.75 Å. The AutoDock molecular docking algorithm to calculate binding affinity ( $\Delta G_{bind}$ ) was implemented via the Raccoon v1.0 modeling suite to perform virtual screening. The receptor and ligand structure preparations for molecular docking included Gasteiger partial charges assignment and rotatable bonds definition according to the standard protocol published elsewhere (Shityakov et al., 2014). The inhibition

Compound	Structure	Indication			
1. Unithiol	HS SH 000	Mercury, arsenic, and lead poisoning			
2. Aurotioprol	H +AuS OH Na <sup>+</sup>	Moderate-severe rheumatoid arthritis and tuberculosis			
3. Acamprosate		Abstinence from alcohol			
4. Menadione	O O S O O O O H S S O O O H S S O O O H S S O	Hypoprothrombinemia			
5. Docusate sodium		Occasional constipation			
6. Camphotamide		Cardioprotection and neurostimulation			
7. Dermatan sulfate		Deep vein thrombosis			
8. Ecamsule	HO'O	Skin protection			
9. Sulfamazone		Sulfonamide antibiotic with antipyretic properties			

#### TABLE 1 Chemical structures and clinical indications of FDA and EMA-approved drugs (n = 13) containing propanesulfonic scaffold of TRA.

(Continued on following page)



TABLE 1 (Continued) Chemical structures and clinical indications of FDA and EMA-approved drugs (n = 13) containing propanesulfonic scaffold of TRA.

constants (*Ki*) and Ligand Efficiencies (*LE*) were calculated from the binding energy values as follows:

$$Ki = expigg[rac{\Delta G_{bind}}{RT}igg]$$
 $LE = rac{\Delta G_{bind}}{N_{atm}}$ 

where *R* (gas constant) is 1.98 cal(mol<sup>\*</sup>K) <sup>-1</sup>; *T* (room temperature) is 298.15 K;  $N_{atm}$  is the number of non-hydrogen atoms in a molecule. AutoDock v.4.2.5.1 was used in the study since its previous version incorrectly calculates part of the intermolecular desolvation energy term (Goodsell et al., 1996). The docking grid with dimension size of

 $60 \times 60 \times 60$  Å and a grid spacing of 0.375 Å were used in the study. The Glide molecular docking algorithm with the Prime MM-GBSA (generalized Born solvent-accessible surface area) approach to calculate the free energy of binding ( $\Delta G_{PR}$ ) was implemented using the default settings, such as the OPLS3e force field, 0.25 of charge cutoff, and 0.8 scaling of the vdW radii of non-polar receptor and ligand atoms. The G-score value, as an empirical scoring function, was used to approximate the ligand binding free energy. All molecular dynamics (MD) simulations were performed using the AMBER 16 package with the FF99SB and GAFF force fields for the A $\beta$  peptide and its ligands (Case et al., 2005). The Antechamber module of AmberTools was employed to calculate the partial charges of the ligands using the semi-empirical AM1-BCC function according to the



FIGURE 1 | Relationship between cIRMS (A) as a root-mean-square difference between top two conformations in the lowest-energy cluster, cIRMSa (B) as an average of the root-mean-square difference between all conformations and the lowest energy conformation in the lowest-energy cluster and the number of torsions (N<sub>tor</sub>) in the ligand molecule.

TABLE 2 | Summary of AutoDock molecular docking results for drugs (n = 13), containing propanesulfonic scaffold of TRA.

Compound	∆G <sub>bind</sub> kcal/mol	cIRMSD Å	cIRMSD <sub>a</sub> Å	N <sub>atm</sub>	N <sub>tor</sub>	Κί μΜ	LE	т
1	-3.05	_	_	12	6	5.7 <sup>a</sup>	-0.25	0.57
2	-3.7	_	_	11	5	1.89 <sup>a</sup>	-0.34	0.67
3	-5.15	0.21	0.74	13	5	162.66	-0.39	0.71
4	-7.11	0.29	0.54	18	2	5.87	-0.39	0.56
5	-4.97	1.84	1.84	29	19	220.65	-0.17	0.63
6	-6.82	0.04	0.07	15	1	9.61	-0.46	0.5
7	-4.26	_	_	37	12	734.58	-0.12	0.56
8	-6.17	1.78	1.18	40	10	28.9	-0.15	0.45
9	-5.91	_	_	41	10	44.89	-0.14	0.33
10	-2.97	_	_	52	15	6.53 <sup>a</sup>	-0.06	0.27
11	0.66	_	_	61	30	3.06 <sup>b</sup>	0.01	0.44
12	-3.81	_	_	57	20	1.57 <sup>a</sup>	-0.07	0.44
13	-5.71	_	_	55	16	62.99	-0.1	0.42
14	-6.19	0.87	0.58	11	5	27.94	-0.56	0.67

<sup>a</sup>mM. <sup>b</sup>M

standard protocol (Marques et al., 2021). The systems were solvated with the TIP3P water models and neutralized by adding the Na + ions using the tLEap input script available from the AmberTools package. Long-range electrostatic interactions were modeled via the particlemesh Ewald method (Essmann et al., 1995). The SHAKE algorithm (Miyamoto and Kollman, 1992) was applied to constrain the length of covalent bonds, including the hydrogen atoms. Langevin thermostat was implemented to equilibrate the temperature of the system at 310 K. A 2.0-fs time step was used in all of the MD setups. For the minimization and equilibration (NVT and NPT ensembles) phases, 100,000 steps and a 1-ns period were used, respectively. Finally, 100-ns classical MD simulations, with no constraints as NPT ensemble, were performed for each of the peptide-ligand complexes using the molecular mechanics combined with the Poisson-Boltzmann (MM-PBSA) or generalized Born (MM-GBSA) augmented with the hydrophobic solvent-accessible surface area term (Kollman et al., 2000; Shityakov et al., 2017). The MM-PBSA/GBSA solvation models were applied as a postprocessing end-state method to calculate the free energies

 $(\Delta G_{PB} \text{ and } \Delta G_{GB})$  together with the entropies  $(T\Delta S)$  and enthalpies  $(\Delta H)$  for the analyzed molecules, namely:

#### $\Delta G_{PB} = \Delta H_{PB} - T\Delta S; \ \Delta G_{GB} = \Delta H_{GB} - T\Delta S$

To calculate the blood-brain barrier partitioning coefficients (logBB), the in-house python script, based on the Clark ( $logBB_{cl}$ ) and Rishton ( $logBB_{ri}$ ) linear regression models, was ran according to the following equations (Clark, 2003; Rishton et al., 2006):

$$logBB_{cl} = 0.152AlogP - 0.0148PSA + 0.139$$
  
$$logBB_{ri} = 0.155AlogP - 0.01PSA + 0.164$$

where *PSA* and *AlogP* are the polar surface area and atom-based octanol-water partitioning coefficient. The in-house PyMol script was applied to calculate the Buried Surface Area (*BSA*) of the peptide-ligand complexes according to the equation:

$$BSA = \frac{\left(ASA_{pep} + ASA_{lig}\right) - ASA_{comp}}{2}$$





where  $ASA_{pep}$  and  $ASA_{lig}$  and  $ASA_{comp}$  are the accessible surface areas of the peptide, ligand, and complex components. Molecular descriptors, such as molecular weight (*MW*), *AlogP*, H-bond acceptors (*HBA*), H-bond donors (*HBD*), *PSA*, quantitative estimate of drug-likeness (*QED*), and Tanimoto molecular similarity (*T*) indexes were calculated with the Biscu-it<sup>TM</sup> tools and Rcpi and Rcdk libraries within the *Python* and R environments (Voicu et al., 2020; Cao et al., 2015). Buried surface areas (*BSA*)

TABLE 3 Summary of molecular descriptors and partitioning coefficients (logBB) determined for lead compounds (drugs) with highest A<sup>β</sup> inhibition properties.

0	MW	AlogP	HBA	HBD	PSA	QED	logBB <sub>cl</sub>	logBB <sub>ri</sub>	logBB <sub>exp</sub>
Compound									
Menadione	253.25	0.98	4	0	88.18	0.76	-1.02	-0.57	_
Camphotamide	231.29	1.14	3	0	71.11	0.69	-0.74	-0.37	_
TRA	138.17	-0.91	3	1	80.06	0.58	-1.18	-0.78	_
Donepezil	379.5	4.36	4	0	38.77	0.72	0.23	0.45	0.89

**TABLE 4** Summary of binding affinities ( $\Delta G$  and G-score), entropy (T $\Delta S$ ), enthalpy ( $\Delta H_{PB}$  and  $\Delta H_{GB}$ ), and Buried Surface Area (BSA) values calculated for lead candidates and TRA. Entropy-enthalpy compensation data of the peptide-ligand complexes is obtained from the normal-mode analyses of 100 ns trajectories at the temperature of 298.15 K.

Compound	$\Delta G_{PB}$	$\Delta G_{GB}$	$\Delta G_{PR}$	G-score kcal/mol	T∆S	$\Delta H_{PB}$	$\Delta H_{GB}$	BSA, Å <sup>2</sup>
Menadione	-13.75	-19.71	-38.81	-6.51	-20.4	-34.15	-40.11	489.77
Camphotamide	-15.61	-14.86	-21.36	-5.08	-17.86	-33.47	-32.72	481.09
TRA	-2.28	-10.1	-16.31	-4.49	-17.91	-20.19	-28.01	373.56



FIGURE 4 | 2D Aβ-ligand interaction diagrams predicted from the AutoDock runs for TRA (A), menadione (B), and camphotamide (C) bound to Aβ peptide. The chain information is shown in parentheses. All hydrogen atoms are removed to enhance clarity.



have been calculated for 2D A $\beta$ -ligand interaction diagrams using the NACCESS program (Hubbard and Thornton, 1993; Wallace et al., 1995). Molecular graphics and visualization were performed with the LigPlot + program (EMBL-EBI, Wellcome Trust Genome Campus, Hinxton, United Kingdom) in order to build twodimensional interaction diagrams from three-dimensional coordinates. Linear regression analysis, followed by graphical representation was performed by PyMol and GraphPad Prism v.8 (GraphPad Software, San Diego, CA, United States). The differences were considered statistically significant at a *p*-value of <0.05.

## **RESULTS AND DISCUSSION**

Before molecular docking and MD simulations, the DrugCentral database (n = 4,642) was filtered to find the molecules, containing the propanesulfonic scaffold of TRA. The scaffold search protocol is based on the chemical similarity searching that can be used to screen a database of compounds for structural similarity to a query chemical structure. The small portion (n = 13) of FDA and EMA-approved drugs (**Table 1**), containing propanesulfonic scaffold of TRA, was identified as several hit molecules

suitable for further investigation using molecular docking to eventually locate some lead candidates. In fact, our study relies on the modified hit-to-lead protocol to identify promising lead compounds, inhibiting  $A\beta$  in order to highlight the possibility for their optimization.

The identified hit molecules belong to different chemical classes and can be prescribed to treat various pathological conditions, such as bacterial infections (leprosy and tuberculosis), lead or mercury poisoning, metabolic disorders, and neurological conditions. Subsequently, the rigid-flexible molecular docking was performed to calculate the affinity between hit substances and A $\beta$ , targeting the HHQK subregion at the N-terminus of the peptide. The lowest-energy A $\beta$  conformer of the NMR models (n = 10) was chosen as a receptor molecule for molecular docking with the potential energy of -1,106.70 kcal/mol, which makes it more stable in the solution. Additionally, the 3D alignment of the A $\beta$  peptide and HHQK region reviled the 2.0 Å structural deviation indicating a relatively "conserved" protein-receptor binding site in solution NMR models (**Supplementary Figures S1A,B**).

To accurately assess any correlations between the number of atoms in the ligand molecule and the conformational effects happening within the peptide-ligand binding site, we estimated



the root-mean-square (*RMS*) difference between the top two conformations and its average value calculated between all conformations and the lowest-energy conformation in the lowest-energy cluster (**Supplementary Figure S2**). A strong positive correlation ( $r^2 = 0.78$ ) with reliable statistics (*p*-value = 0.02) between the *clRMS*, as an *RMS* difference between top two conformations in the largest cluster, and the number of torsions ( $N_{tor}$ ) was observed by linear regression analysis (**Figure 1A**). Furthermore, a significant standard deviation of the *clRMSa* variable, as an average of the *RMS* difference between all conformations and the lowest energy conformation, was detected for some hit compounds (3, 4, 8, and 14), describing the elevated conformational change (**Figure 1B**), which greatly affects the correlation coefficient ( $r^2 = 0.63$ ).

Finally, menadione and camphotamide lead candidates were found among the top binders (Table 2 and Figures 2A,B) with the binding energy values (-7.11 and -6.82 kcal/mol) significantly lower than for the reference compound with a  $\Delta G_{bind}$  value of -6.19 kcal/mol (Table 2 and Figure 2C). It was previously published that menadione sodium bisulfite as a remedy against hemorrhagic disease caused by vitamin K deficiency could inhibit Aβ toxic formation and aggregation in C. elegans, extending its life span and reducing disruption of cellular membranes (Zhang et al., 2018). The co-authors also hypothesized that menadione might inhibit amyloid formation due to its backbone similarity to 1,4naphthoquinone, which shows strong anti-aggregation effects on amyloidogenic proteins, such as insulin and  $\alpha$ -synuclein. Similarly, the 6-hydroxy-nicotine intermediate from P. nicotinovorans as a precursor for camphotamide has been shown to have neuroprotective effects with putative applications in AD treatment (Wang et al., 2015; Hritcu et al., 2017). As a positive inotropic agent (cardiotonic), this substance could be potentially prescribed to treat AD-associated amyloid cardiomyopathy, inhibiting, directly and indirectly, AB accumulation in the heart.

Indeed, the inhibition of A $\beta$  and the formation of neurofibrillary tangles along the heart-brain axis could be reached through targeted supplementation of neurotrophic factors to the brain as it was hypothesized by Shityakov and coauthors Shityakov et al. (2021).

To investigate how molecular similarity contributes to the A $\beta$  binding, the Tanimoto coefficients as an appropriate choice for a fingerprint-based similarity and ligand efficiency as the affinity normalized by the number of non-hydrogen atoms were implemented. A moderate negative correlation ( $r^2 \approx 0.5$ ) with reliable statistics (*p*-value = 0.008) between the *T* and *LE* parameters was observed by linear regression analysis (**Figure 3**).

In other words, if chemical compounds match more to the TRA scaffold, they also have very similar binding modes calculated per number of heavy atoms in the molecule. In addition, all hit molecules were subdivided into 4 big groups or clusters by the hierarchical clustering according to their E-state molecular fingerprints and pairwise similarity matrix, where the lead molecules (compounds 4 and 6) ended up in the same cluster (**Figure 3B**).

Next, the *AlogP*, *QED*, *PSA*, and *logBB* values for menadione, camphotamide, and TRA as a reference, and donepezil as standard control were calculated to evaluate the drug ability to possess the optimal BBB permeation properties and druglikness (**Table 3**).

The AlogP parameter for lead candidates suggested their mild lipophilic properties, which were significantly higher than for the reference substance but not as good as for the standard control. On the other hand, the QED parameters for lead candidates were very similar to those determined for donepezil, reflecting the distribution of molecular properties, such as the number of hydrogen bond donors and acceptors, the number of aromatic rings, and the presence of unwanted chemical functionalities. However, for a CNS-active compound to permeate the BBB, a surface area less than 60-70 Å<sup>2</sup> is usually required to achieve the desired brain bioavailability for the administered drug (Shityakov et al., 2014; Kelder et al., 1999). Judging by relatively low AlogP and high PSA, there is a risk that the BBB permeation of the lead candidates might not be entirely sufficient as confirmed by the logBB, which optimally should be logBB > 0 (Shityakov et al., 2015). Therefore, further lead optimization by in situ enumeration or fragment replacement might be a plausible choice to make chemical modifications, improving brain bioavailability (logBB) together with selectivity, pharmacokinetic and pharmacodynamic parameters, and decreasing toxicity. Reaction-based in situ enumeration can be achieved by proposing synthetically feasible candidates from a reagent library in particular, which reagents are most likely to produce potential active compounds for the binding site (Mok et al., 2017). On the contrary, the fragment replacement protocol performs scaffold hopping by replacing part of the scaffold structure while maintaining the favorable binding between the receptor and the ligand (Vainio et al., 2013). The elevated experimental logBB index of 0.89 for donepezil was mainly due to its carrier-mediated transport to the brain (Kim et al., 2010). It could probably be accomplished by organic cation or choline transporters (OCT1-3 and CHT1) and not just crossing the BBB via passive diffusion through endothelial cells (Kim et al., 2010).

To validate further molecular docking results, the free energy of binding based on implicit solvation models was calculated for amyloiddrug complexes. The MM-PBSA/GBSA calculations (**Table 4**), using 100 ns MD trajectories, confirmed the previous data completely (GBSA) and partially (PBSA), revealing much higher binding affinities of lead compounds to  $A\beta$  in comparison to the reference.

The former protocol provided the best affinity for menadione  $(\Delta G_{GB} = -19.71 \text{ kcal/mol})$ , probably because in some extensive studies the MM-GBSA approach was computationally more efficient, achieving better accuracy but being less rigorous (Hou et al., 2011). Therefore, the second validation was done to clarify the MM-PBSA/ GBSA discrepancy by utilizing the Prime algorithm with the MM-GBSA-based scoring function. As a result, the G-score data and the entropy-enthalpy compensation analysis confirmed the previous findings, describing the exothermic nature ( $\Delta H < 0$ ) of the binding process with the decreased disorder ( $T\Delta S < 0$ ), which could occur spontaneously depending on temperature. Additionally, the BSA values (Table 4 and Figures 4A-C), which measures the size of the Aβ-drug interface, also confirmed the previous binding affinity pattern, where the BSA elevation leads to an increase in binding as it was already published for the peptide-ligand and host-guest systems (Shityakov et al., 2020; Esmaeilpour et al., 2021).

To analyse the movements of the studied complexes, the rootmean-square deviation (RMSD) and fluctuation (RMSF) values together with the radius of gyration  $(R_{e})$  with respect to the initial conformation were plotted versus time (Supplementary Figures S3A-E). The receptor RMSD values were relatively high (RMSD =10 Å) stabilized after about 20 ns for the compound 4 and 6/AB complexes and after 40 ns for TRA/A $\beta$  (Supplementary Figure S3A). The ligand RMSD remained low within 1.0 Å and stabilized almost instantly (Supplementary Figure S3B). The receptor RMSF values produced some picks associated with the high flexibility of the AB termini (Supplementary Figure S3C). The atomic fluctuations of ligands showed more stable profiles especially for compounds 4 and 6 (Supplementary Figure S3D). The decreased  $R_{g}$  values were associated with the receptor compactness, which was elevated during the simulation (Supplementary Figure S3E). Finally, the number of H-bonds between the receptor and its ligands and the fraction of residues involved in H-bonding were assessed to find the H-binding contribution to the affinity. These parameters were the highest for compound 4 with more H-bonds formed, (Supplementary Figure S3F) significantly increasing the residue fraction (Supplementary Figure S3G).

Finally, per-residue and pairwise energy decomposition analyses were employed to evaluate the energetic contribution of drugs and the binding site residues. Some unique residues, such as Tyr10 (B) for TRA, Lys16 (A, B) for menadione, and Lys16 (B, C) for camphotamide were identified at the energetic threshold ( $\Delta G = -3.0$  kcal/mol) and below by one or both implicit solvation protocols (**Figures 5, 6**). In particular, all amino acid residues involved in the interaction with the lead candidates had exceeded the energetic threshold (**Figure 5A,B**), which was not observed for the reference molecule (**Figure 5C**). Meanwhile, the Lys16 targeting in A $\beta$  by the oxidation of a catechol structure to the *o*-quinone forming the *o*-quinone-A $\beta$ adduct is believed to be responsible for its anti-aggregation activity (Liu et al., 2017). In the pairwise interaction, only Tyr10 (B) was detected slightly above the adjusted energetic threshold ( $\Delta G = -6.0 \text{ kcal/mol}$ ), and Lys16 (B) had the lowest energy during the MD simulation of the A $\beta$ -menadione complex (**Figure 6**). Highlighting the contribution of these residues for the inhibitor of this site, curcumin and resveratrol were found to interact with Arg5, Ser8, Tyr10, Gln15, Lys16, Leu17, and Phe20 (Fu et al., 2014). Besides, solvent-accessible residues, such as Phe20 at the N-terminal.

KLVFF stretch was reported as crucial in the interactions of these two inhibitors (Fu et al., 2014). Some studies have also documented the pairwise interactions for some amyloid variants (A $\beta_{10-35}$ ) to illustrate residue energetic contributions in a process of A $\beta$  reorganization driven basically by inter-chain hydrophobic and hydrophilic interactions and also solvation/desolvation effects (Campanera and Pouplana 2010).

## CONCLUSION

In this study, the scaffold searching approach based on known AB inhibitor tramiprosate to screen the DrugCentral database (n =4,642) was employed to identify hit compounds (n = 13) and lead candidates (n = 2) for AD drug repurposing. Two lead candidates, namely menadione bisulfite and camphotamide, out of 13 hit compounds were identified as promising AB inhibitors with the improved  $\Delta G_{bind}$  and logBB parameters. The binding affinity modes were also confirmed by molecular dynamics simulations using implicit solvation models, in particular MM-GBSA. We assume that this scaffold searching methodology in conjunction with pharmaceutical profiles (logBB) can be applied as a starting routine for drug repurposing in AD. Overall, the proposed computational pipeline can be implemented through the early stage rational drug design to nominate drugs for AD that, after additional in vitro and in vivo validation, could be readily evaluated in a clinical trial.

## DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding authors.

## **AUTHOR CONTRIBUTIONS**

SS, ES, CF, and TD conceptualized the topic. SS performed the experiments and wrote the manuscript. All co-authors read the manuscript. All authors contributed to the article and approved the submitted version.

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### SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fchem.2021.736509/full#supplementary-material

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