



## OPEN ACCESS

## EDITED BY

Juan Pellico,  
King's College London, United Kingdom

## REVIEWED BY

Junjie Li,  
Kyushu University, Japan  
Fernando Herranz,  
Spanish National Research Council (CSIC),  
Spain

## \*CORRESPONDENCE

Jason J. Davis,  
✉ jason.davis@chem.ox.ac.uk

RECEIVED 03 April 2025

ACCEPTED 23 June 2025

PUBLISHED 14 August 2025

## CITATION

Duncan AM, Ellis CM, Smith JP, Leutloff L,  
Langton MJ and Davis JJ (2025) Organic  
functionality in responsive  
paramagnetic nanostructures.  
*Front. Chem.* 13:1605538.  
doi: 10.3389/fchem.2025.1605538

## COPYRIGHT

© 2025 Duncan, Ellis, Smith, Leutloff, Langton  
and Davis. This is an open-access article  
distributed under the terms of the [Creative  
Commons Attribution License \(CC BY\)](#). The use,  
distribution or reproduction in other forums is  
permitted, provided the original author(s) and  
the copyright owner(s) are credited and that the  
original publication in this journal is cited, in  
accordance with accepted academic practice.  
No use, distribution or reproduction is  
permitted which does not comply with these  
terms.

# Organic functionality in responsive paramagnetic nanostructures

Anna M. Duncan, Connor M. Ellis, James P. Smith, Lillian Leutloff,  
Matthew J. Langton and Jason J. Davis\*

Department of Chemistry, University of Oxford, Oxford, United Kingdom

Magnetic resonance imaging (MRI) has become an invaluable tool for diagnosing and monitoring a range of medical conditions, including cancer and cardiovascular disease, owing, in large part, to its high spatial resolution. Despite this, MRI suffers from an inherent low sensitivity, a drawback that can be mitigated through the use of exogenous contrast agents. Although molecular paramagnetic contrast agents are most commonly used, they suffer from significant limitations, including short circulation times, inadequate sensitivity, moderate (or no) tissue specificity, and potential toxicity. Recent advancements in nanomaterials research have paved the way for the development of paramagnetic nanoplateforms offering a promising alternative to these traditional chelates. Responsive contrast agents have gained attention due to their ability to generate local contrast in areas of particular interest, enabling the potential for disease-specific reporting where environmental factors including pH, ion concentration and biomolecule activity deviate from the norm. In addition to this, the generation of local or locality-specific contrast can help to overcome the intrinsic nonspecific nature of traditional contrast agents allowing for overall better treatment options. Purely organic nanoparticles, including those which are micellar, liposomal or dendritic and inorganic-polymer hybrids, can support step changes in MRI signal generation and its diagnostic potency by leveraging the specific and responsive characteristics of the organic components. This review seeks to illustrate how the integration of organic chemistry into magnetic nanostructures can enable responsive high-contrast generation.

## KEYWORDS

MRI, nanoparticles, polymers, responsive, organic

## 1 Introduction

### 1.1 Magnetic resonance imaging

The detection of disease and early or associated physiological irregularities is crucial in improving health outcomes (Deulkar et al., 2024). It aids prophylaxis, enhances surveillance capabilities, and facilitates a more effective and personalised treatment (Cormode et al., 2009; Mura and Couvreur, 2012; Crommelin and Florence, 2013). The early detection of diseases such as cancer is, for example, highly correlated with the probability of recovery; for example, prostate cancer detected in stages 1, 2, or three presents with nearly a 100% 1 year survival rate, this falling to a 87.6% 1 year survival rate when detected in stage 4 (Hawkes, 2019). Although ~50% of cancers are currently diagnosed at stages 3 and 4, a robust screening and improved imaging sensitivity can help physicians accurately determine the

presence of a lesion, and if it is likely to be malignant (Crosby et al., 2022). MRI scans of high detail and edge resolution, in particular, yield valuable information on tumour morphology, and thus inform on the likelihood of malignancy. Methodologies that support the non-invasive visualisation of pathology include magnetic resonance imaging (MRI), positron emission topography (PET), single photon emission tomography (SPECT), computed tomography (CT), ultrasound (US), optical imaging (OI) and photoacoustic imaging (PAI) (Smith and Gambhir, 2017; Liu Y. et al., 2019; Walter et al., 2020; Hsu et al., 2023). The choice of imaging method needs to be tailored to a specific resource availability and clinical diagnostic aim; MRI is a powerful imaging modality with an anatomical spatial resolution of 1 mm and unlimited depth penetration but with an associated low sensitivity ( $\mu\text{M}$ – $\text{mM}$ ) (Alvares et al., 2017). To address this and add functionality, contrast agents (CAs) can be used to enhance diagnostic value through either increased generic signal:noise and/or environmentally-specific signal generation (see Section 1.2).

Based on nuclear magnetic resonance (NMR) principles, MRI analyses probe the interaction of magnetically active (non-integer spin quantum number) nuclei with an externally applied magnetic field (Jackson et al., 2021). Clinical MRI scanners facilitate the spatial mapping of water  $^1\text{H}$  (proton) signal density within various soft tissue structures of the body, depending on their specific magnetochemical environment, the latter having a dependence on the local concentration of water (Bley et al., 2010; Kostevšek, 2020). Crucially, this allows for the detection of structural abnormalities and disease, including, but not limited to sites of tumour (as noted above), injury and infection (Brindle, 2008; Abramovitch et al., 1998; Modic et al., 1986). Different classes of MRI imaging employ various scanner design, pulse sequences and image weighting. The majority of MRI scanners for diagnostic purposes are closed-bore systems, where the magnetic field is generated by passing an electrical current through a superconducting niobium-titanium (Nb-Ti) solenoid ( $<9.3$  K) surrounded by copper (Busse et al., 2018; Warner, 2016; Zhang et al., 2019; Parizh et al., 2017). The generation of cross-sectional images necessitates the use of a strong (commonly 1.5–3 T) magnetic field to align the water proton spins (Berger, 2002); subsequently, longitudinal and transverse relaxation processes result in the restoration of equilibrium magnetisation as mapped by a Fourier transformation of the FID signals measured by the receiver (Edelstein et al., 1984). Subtle differences in the rates of these relaxation processes are reflective of local environment. The most commonly employed imaging sequences within MRI are  $T_1$ -weighted and  $T_2$ -weighted. The former utilise both short repetition times (TR) between successive pulse sequences and echo times (TE) between the delivery of the radiofrequency (RF) pulse and the reception of the echo signal, whilst the opposite is true for  $T_2$ -weighted scans (Kawahara et al., 2021; Jung and Weigel, 2013).

To elucidate the time constant of the longitudinal relaxation process ( $T_1$ ), a  $180^\circ$  ( $\pi$ ) RF pulse is used to re-orient the spins away from equilibrium, inverting their net magnetisation vector ( $M_z$ ) (Bain, 1990). Spin-lattice relaxation restores equilibrium magnetisation, with the rate of change along the z-axis ( $M_z$ ) described by a rate constant,  $R_1$ , where  $R_1 = 1/T_1$  (Williams et al., 2005; Spencer and Fishbein, 2000; Kingsley, 1999). To probe transverse relaxation, a  $90^\circ$  ( $\pi/2$ ) RF pulse is applied that

focuses protic spin into the  $xy$ -plane, specifically introducing a net magnetisation vector ( $M_{xy}$ ) (Xu and Chan, 1999). Loss of spin coherence results in  $M_{xy}$  progressively falling to zero and the associated detected free induction decay (FID) signal exponentially decaying with a time constant  $T_2$  (Le Botlan and Ouguerram, 1997). These concepts are summarised in Figure 1.

## 1.2 Contrast agents

MRI CAs allow an improved delineation (through differential image signal: noise) between different tissues, specific microenvironments, or anatomical structures within the body (De León-Rodríguez et al., 2015; Kim et al., 2000). The efficiency (effect per unit dose) of any given CA is defined by its relaxivity (Aime et al., 2009). Relaxivity,  $r_i$ , measured in  $\text{mM}^{-1} \text{s}^{-1}$ , is the linear gradient between the relaxation rate ( $R_i = 1/T_i$ , where  $i = 1, 2$ ) against CA concentration ( $\text{mM}$ ) (Werner et al., 2008). A shorter  $T_1$  corresponds to a brighter image, therefore, if specific tissue possesses relaxation rates that are either too slow (i.e., long  $T_1$ ), or too close to the relaxation times of water in neighbouring tissues CAs (commonly lanthanide-based, often containing chelated  $\text{Gd}^{3+}$ ) are introduced to improve the spatial resolution/clarity of the MR image (Li et al., 2019). A shorter  $T_2$  corresponds to a darkening of image contrast, with  $T_2$  CAs introducing local magnetic field inhomogeneities that lead to a decrease in signal intensity through promotion of the transversal relaxation pathway. Clinically, CAs that enhance longitudinal relaxivity are generally more useful than CAs that augment transversal relaxivity, as a darker scan area is also indicative of artefacts/clots (Na and Hyeon, 2009; Geraldès and Laurent, 2009). Notably, a high relaxivity also means that a lower dose of the CA can be administered, reducing off-target effects (Lancelot et al., 2020; Jacques et al., 2010). The rational design of CAs with improved relaxivity generally considers a programmed slowing of molecular tumbling (by virtue of the increased size of the probe) and/or enhanced inner/second/outer sphere effects through either increased hydration, conformational rigidification, or an increased viscosity of surrounding water molecules (Figure 2). (Botta and Tei, 2012)

As noted,  $T_1$  CAs typically contain paramagnetic ions such as  $\text{Gd}^{3+}$  (gadolinium-based CAs, GBCAs) and are commonly used to enhance the visualization of blood vessels, tumours, inflammation, and specific organs (Blomqvist et al., 2022; Iyad et al., 2023). They consist of kinetically stable Gd-chelates and include the use of ligands such as 2,2',2'',2'''-(1,4,7,10-tetraazacyclododecane-1,4,7,10-tetrayl)tetraacetic acid (DOTA) and  $N,N'$ -[(Carboxymethyl)azanediyl]di (ethane-2,1-diyl)bis [N-(carboxymethyl)glycine] (DTPA) (Bousquet et al., 1988). Low molecular weight high spin complexes of  $\text{Fe}^{3+}$  and  $\text{Mn}^{2+}$  can also be used as lower relaxivity substitutes but are, additionally, accompanied by not insignificant toxicity concerns (Wang et al., 2019; Manavalan et al., 2024; Devreux et al., 2021; Drahoš et al., 2012).

Many such Gd-based agents carry the risk of nephrogenic systemic fibrosis (NSF) resulting from the de-chelation of  $\text{Gd}^{3+}$  from its associated macrocycle, primarily affecting patients with compromised renal function (Ersoy and Rybicki, 2007; Lin and



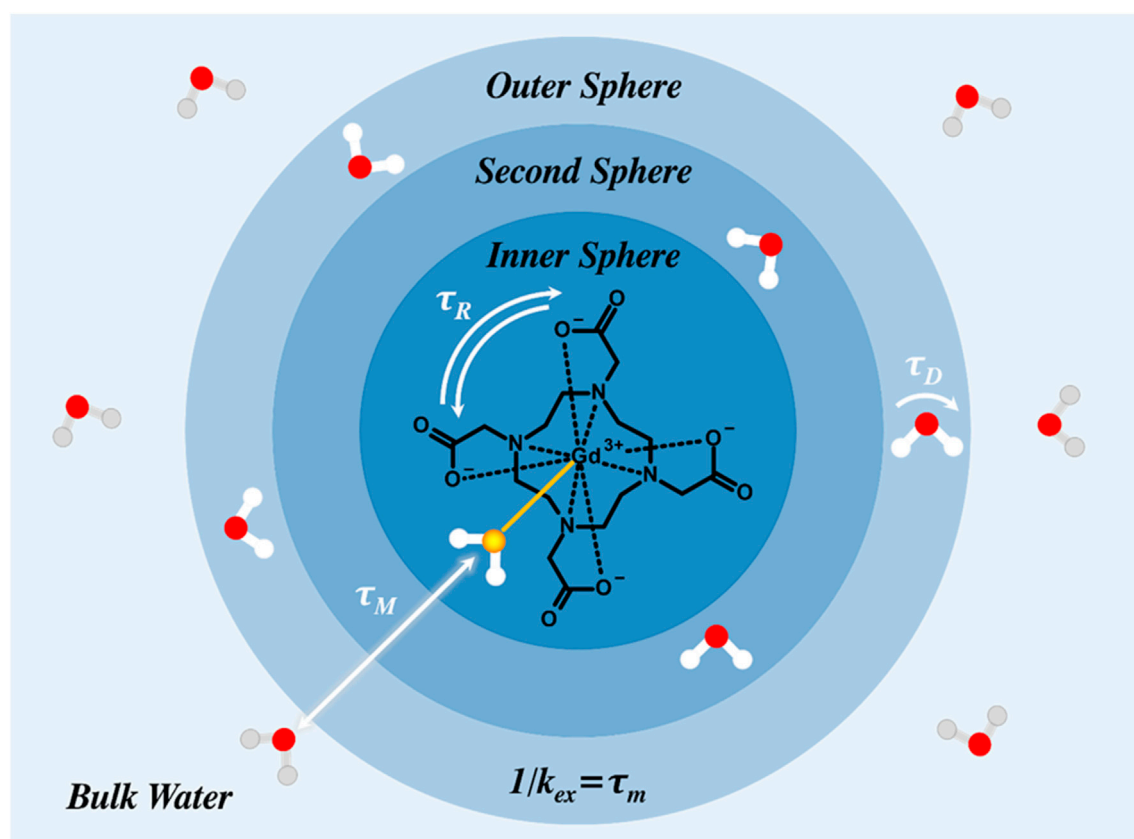


FIGURE 2

Water molecules can either coordinate directly with the paramagnetic centre (inner sphere), weakly interact with the chelating ligand (second sphere), or diffuse more freely (outer sphere). The rotational correlation time (or molecular tumbling rate) is denoted by  $\tau_R$ , the water exchange correlation time by  $\tau_M$  ( $k_{ex}$  = water exchange rate), and the diffusion correlation time by  $\tau_D$ .

(such as lipid vesicles (Langereis et al., 2013) and polymer micelles (Nasongkla et al., 2006)), and hybrid nanoparticles thereof (Arosio et al., 2013). The incorporation of paramagnetic chelates within a nanoparticle platform is an attractive way of modulating several Solomon-Bloembergen-Morgan (SBM)-governing parameters, such as those introduced in Figure 2, that are responsible for large baseline boosts in relaxivity (Wahsner et al., 2019). The anchoring of a Gd-chelate to a rigid or semi-rigid scaffold such as an inorganic nanoparticle or a polymer chain restricts the rotation of the chelate, stretching  $\tau_R$ , and typically moving the characteristic rotational frequency ( $1/\tau_R$ ) closer to alignment with the Larmor frequency of water (notably improving relaxivity) (Verwilt et al., 2015; Pellico et al., 2019a). Similarly, the confinement of a paramagnetic chelate within a nanoparticle scaffold, e.g., entrapping within a polymer matrix or porous inorganic nanoparticle, can lead to refined SBM parameters, such as restricted water exchange rate ( $\tau_M$ ), increased rotational correlation time ( $\tau_R$ ), and enhanced local water hydration (i.e., amplified outer sphere effects) with the central lanthanide, effects also able to support an improved relaxivity (Villaraza et al., 2010). These modified nanoparticulate scaffolds are generally metabolised in the liver, the main detoxification organ, with associated potential biosafety concerns (Wang et al., 2024). Inadequate elimination may result in the long-term accumulation of such nanomaterials in

hepatocytes, impairing their normal biological function. Harmful downstream effects may also result from the production of reactive oxygen species, associated destructive interactions with DNA/mitochondria, and effects on intracellular signalling pathways; a range of *in vivo* and *in vitro* hepatotoxicity studies have now taken place in an attempt to understand and mitigate these effects (Yao et al., 2019).

### 1.3 Responsive organic contrast

The pathophysiology accompanying many diseases can result in altered chemical environments in affected and surrounding tissues; malignant tumours possess, for example, acidic extracellular pH levels (by virtue of a high rate of aerobic glycolysis, increasing lactic acid production) and hypoxia (through overproduction of, e.g., glutathione (GSH) and cysteine) (Chen et al., 2020; Hao et al., 2018), with decreased tissue pH also found in conditions such as atherosclerosis (Naghavi et al., 2002) and renal disease (Kraut and Kurtz, 2005). Low extracellular pH is, thus, a common motivator in the rational design of bio-responsive particles (Kim et al., 2000; Wu and Zhao, 2013). The use of a stimulus-responsive imaging probe is, in general terms, a significant diagnostic asset in supporting disease-specific reporting (Neves and



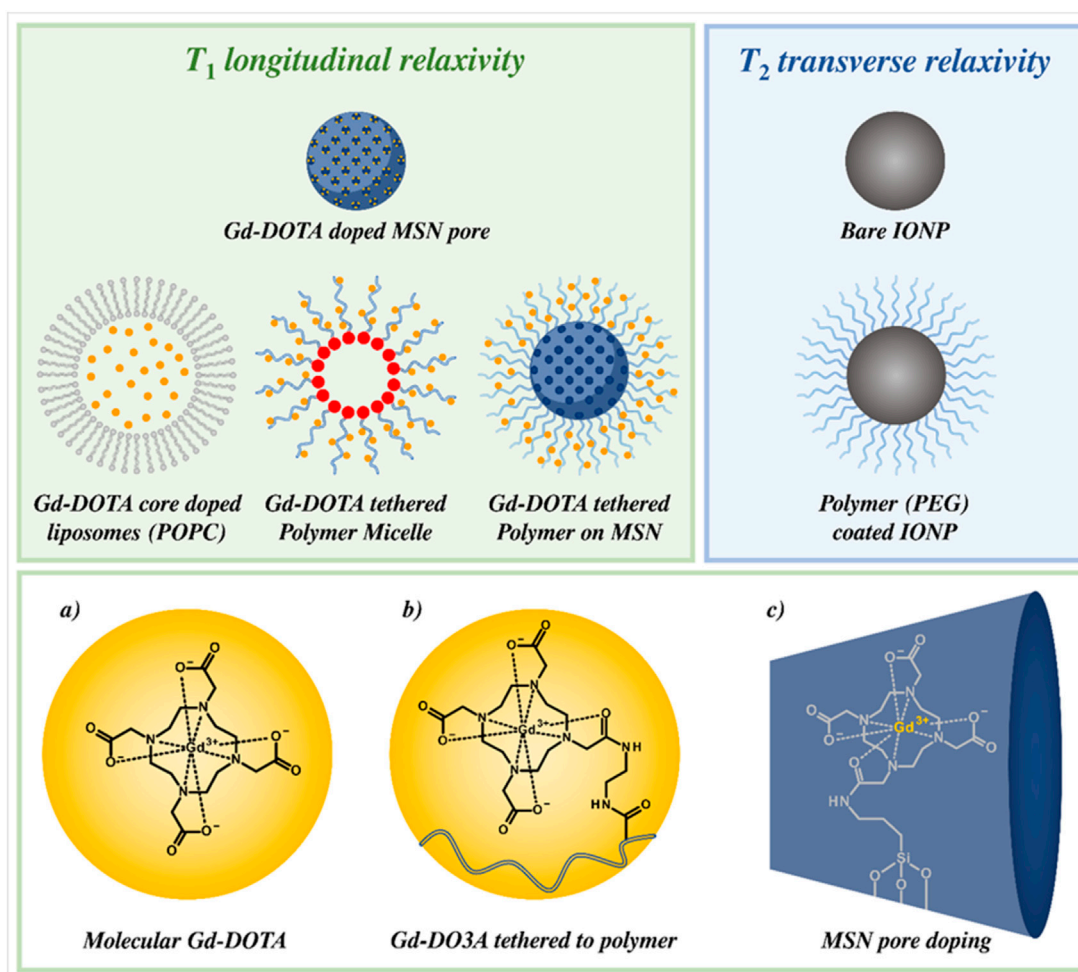


FIGURE 3

Overview of organically-tuneable IONPs, MSNs, artificial phospholipid vesicles (liposomes) and polymer micelles.  $T_1$  relaxivity (examples in green) is achieved through doping/tethering MR-active paramagnetic chelates that include Gd-complexes with a nanoparticle architecture. Displayed are: (a) molecular Gd-DOTA, (b) Gd-DO3A tethered to a polymeric strand, and (c) a doped MSN inner pore. The beneficial elongated rotational correlation characteristics associated with this integration enhance associated image contrast. Iron oxide-based particle platforms are used to promote  $T_2$  relaxation (blue), and can be wrapped in polymers (e.g., PEG) to improve probe biocompatibility and/or elongate blood circulation times.

Brindle, 2006). The integration of organic chemistry enables a tailored rational design (with characteristics engineered to respond to a physiological condition of interest), and improved relaxometric features within a facile chemical tunability (Davies et al., 2013; He et al., 2019; Matsumoto and Jasanoff, 2008; Priya James et al., 2014; Vithanarachchi and Allen, 2012). The so-generated environmentally-responsive CAs can enable localized contrast enhancement, activated at either a desired site or condition, and offer a spatial and temporal modulation of contrast (e.g., dynamic imaging as the responsive CA modulates upon exposure to an altered tissue environment) (Foster and Larsen, 2023). External stimuli such as those associated with pH, biomolecule presence, light, ion presence, specific temperature, or redox agent activity can be leveraged to interact with this CA functionality such that contrast is switched 'on' in the presence of a particular environmental condition *in vivo* (Ge and Liu, 2013; Zhang Z. et al., 2022). Early responsive, non-particulate, MRI contrast focused on the design of simple macrocycles that contained responsive side arm modifications which influenced  $q$

(hydration number) through differences in chelation state on exogenous ion addition (Viswanathan et al., 2010). Specifically, side arm ion-association can result in a conformational change that introduces a vacant coordination site at the paramagnetic centre. In recent work, responsive organic functionality has been leveraged to modulate more subtle SBM-governing parameters, such as the rotational correlation time ( $\tau_R$ ) through locally triggered changes in the size of the nanoparticle agent, e.g., by polymer swelling/deswelling or through altered colloidal properties with deviations in local pH (Cazares-Cortes et al., 2017). Similarly, water exchange rates ( $\tau_M$ ) can be manipulated by triggering changes in polymer hydration or chelate accessibility to bulk water within an organic agent (De Sarno et al., 2019).

Polymers are commonly employed in the design of responsive MRI CAs, either as wholly polymeric nanoparticles that include polymer micelles, or as coatings for inorganic/organic nanostructures such as mesoporous silica nanoparticles (MSNs) and IONPs (Elsabahy et al., 2015; Gao et al., 2011). The former micelles are organic nanoparticles composed of amphiphilic

polymer strands that undergo self-assembly above a critical micellar concentration (CMC) (Zhang and Zhao, 2016), whilst polymer coatings (such as PEG/PAA block copolymers) can be attached to nanoparticles such as MSNs, either through covalent functionalisation or electrostatic association (Pellico et al., 2019b). Responsive agents can also be generated from organic liposomes, phospholipid-based spherical NPs with a hollow core (and an ability to internally encapsulate MRI active cargo and/or drug); these have been explored as an (organic) responsive alternative to traditional MRI CAs, owing to their high chemical tuneability, versatility and good biocompatibility (Nsairat et al., 2022; Hingorani et al., 2015). A range of such polymer-inorganic hybrids and organic-based particles are summarised in Figure 3, with their operational mechanisms detailed in Sections 2,3.

The organic coating of MRI-active nanomaterials can have a profound effect on circulation time and practically-realistic contrast generation. The aggregation of NP CAs *in vivo* predictably correlates inversely with circulation lifetimes, strongly affects relaxivity, and is therefore a core consideration in design (Ta et al., 2017; Ahl et al., 1997). In the case of IONPs, aggregation takes place when interparticle distance is decreased and dipolar interactions increased, a process that may be facilitated by the desorption of capping ligands such as oleic acid (Roca et al., 2009). These aggregates, although possessing enhanced  $r_2$ , are more susceptible to uptake by macrophages. Similarly, aggregation of Ln-doped MSNs, for example, has an observed detrimental effect on their intrinsic relaxivity, due to the blocking/obstruction of the CA-containing pore (or surface) bound sites (Rizzi et al., 2021). In the case of MSNs, aggregation can also occur by protein-mediated neutralisation in serum, a process that is known to be circumvented by surface functionalisation with PEG or zwitterionic polymers (Liu et al., 2015; Lowe et al., 2015). In comparison, aggregation of liposomal structures (see Section 1.3 for discussion of stimuli-responsive liposomal CAs) can be induced by  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$  (Rahnfeld et al., 2018). Cholesterol or phosphatidylglycerol containing liposomes are more rapidly cleared by the reticuloendothelial system, although glycolipids or polymers can be incorporated with the phospholipid structure to mitigate this (Ahl et al., 1997). Suitable organic coatings, then, reduce the prevalence of aggregation and support favourable circulation times *in vivo*, and have accordingly play a critical role in MRI CA design, regardless of any additional responsive functionality.

## 2 Paramagnetic polymer-inorganic hybrid nanoparticles

### 2.1 Polymer functionalised iron oxide nanoparticles

Of the eight iron oxides known, magnetite ( $\text{Fe}_3\text{O}_4$ ) and its oxidised form maghemite ( $\gamma\text{-Fe}_2\text{O}_3$ ) are, in nanoparticulate form, common imaging probes which are routinely surface modified with biocompatible polymers such as PEG, sodium alginate, and poly (acrylic acid) (PAA) to improve colloidal stability under aqueous conditions (Dulińska-Litewka et al., 2019; Estelrich et al., 2015). There are many reported applications of IONPs, including the targeted delivery of therapeutics and for disease treatment

through a considered exploitation of their magnetic properties, e.g., in magnetic hyperthermia which promotes cell apoptosis in tumour tissue (Dadfar et al., 2019; Vallabani and Singh, 2018). Their use as imaging probes has been heavily analysed over the past few decades, and continues to receive interest due to their high magnetic moments, allowing for effective contrast generation at comparatively small dose. The saturation magnetisation of these is also known to be a function of morphology (size, structure and shape), and octahedral, tetrahedral, cuboid, plate and wire-like (*etc.*) IONPs have been successfully produced, building upon the traditional spherical structures (Xie et al., 2018). These geometric variants also have specific biocompatibility and bioclearance characteristics. Renal clearance, in general terms, has been shown to be ineffective for IONPs with diameters  $>50$  nm, resulting in accumulation *in vivo* over extended timeframes, a clear problem if repeat MRI investigations are required (Lapusan et al., 2024). Superparamagnetic iron oxide based nanoparticles have also been employed to enable  $T_1$  contrast generation, an ability ascribed to increased surface  $\text{Fe}^{3+}$  exposure, suppressed magnetisation values and surface effects that influence both magnetisation and water exchange (see Figure 4). (Wei et al., 2017; Bao et al., 2018) The reduced size of IONPs is associated with significantly improved blood half-lives (e.g., 10–14 h for ferumoxytol, used for vascular imaging) due, in large part, to their reduced rate of opsonisation compared to larger diameter equivalents (e.g., 8 min for ferumoxide) (Lapusan et al., 2024).

Despite their ease of fabrication and favourable magnetic characteristics, the fouling of IONPs by protein within the blood/extracellular fluid onto the NP surface typically results in facile aggregation of these particles, leading to their quick removal by macrophages in a process known as opsonisation (Suk et al., 2016). To mitigate this, IONPs are routinely surface functionalised at the point of synthesis by hydrophilic entities such as PEG/PDA/PAA (as shown in Figure 4), a process which also improves their shelf-life for *in vivo* MRI applications, again aiding their clinical translatability (Lapusan et al., 2024).

PEG modification is a particularly common means to improve blood circulation time, colloidal stability, and biocompatibility (Feng et al., 2018). These effects are commonly referred to as ‘stealth’ and reduce the rapid clearance of native particles by the reticuloendothelial system. There has been much work investigating the optimal thickness of the stabilising PEG coat on the surfaces of different sized iron oxide NPs, with molecular weights typically ranging from 2 kDa to 5 kDa. For example, Larsen et al. systematically varied the molecular weight of the PEG-coating of IONPs, ranging from 330–20,000 Da, to investigate the effect on blood circulation time and macrophage uptake (Larsen et al., 2012). In this work, the larger molecular weight (20 kDa) PEG-IONPs were observed to possess the longest blood circulation times (45 min) whereas the low molecular weight (330 Da) PEG-IONPs exhibited the fastest macrophage-based removal. PEG-IONPs with a 5 kDa polymer coat displayed the best contrast generation with an associated  $r_2 = 354 \text{ mM}^{-1} \text{ s}^{-1}$  at 3 T, in data that indicated that thick polymer coatings can detrimentally impact associated relaxivities by restricting the approach of water (and therefore its effective dephasing). In similar work, Leal et al. reported the synthesis of PEG-IONPs with a variety of molecular weights ranging from 600 Da to 8 kDa, with the optimal PEG thickness

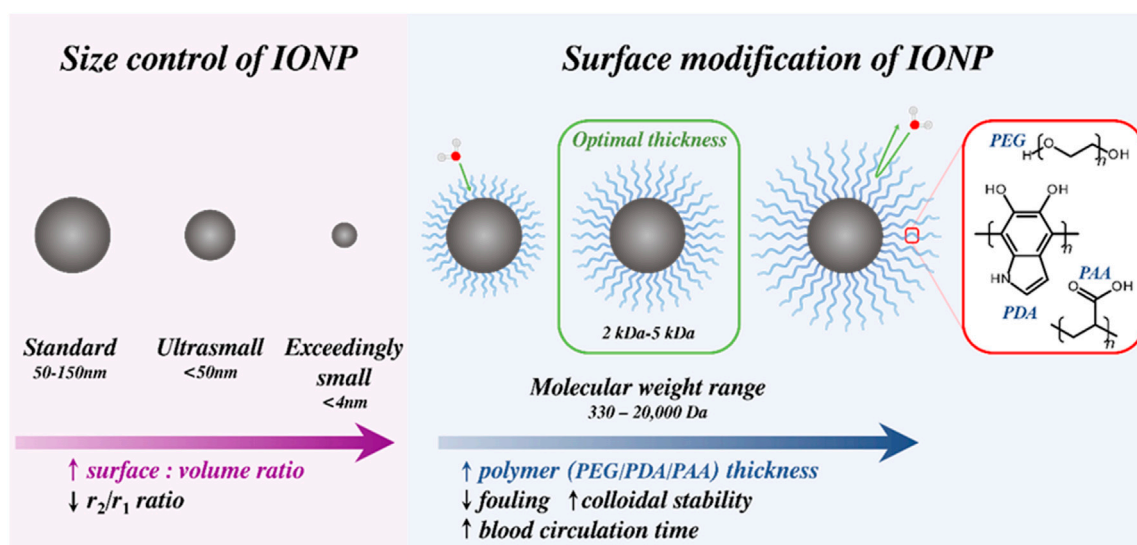


FIGURE 4

Overview of size effects and IONP surface modification. As diameter decreases, the increased surface effects mean that exceedingly small IONPs are suitable  $T_1$  CAs. Modification with hydrophilic polymers (e.g., polyethylene glycol, PEG; polydopamine, PDA; polyacrylic acid, PAA) improves biocompatibility, blood retention time and colloidal stability, assisting clinical translatability.

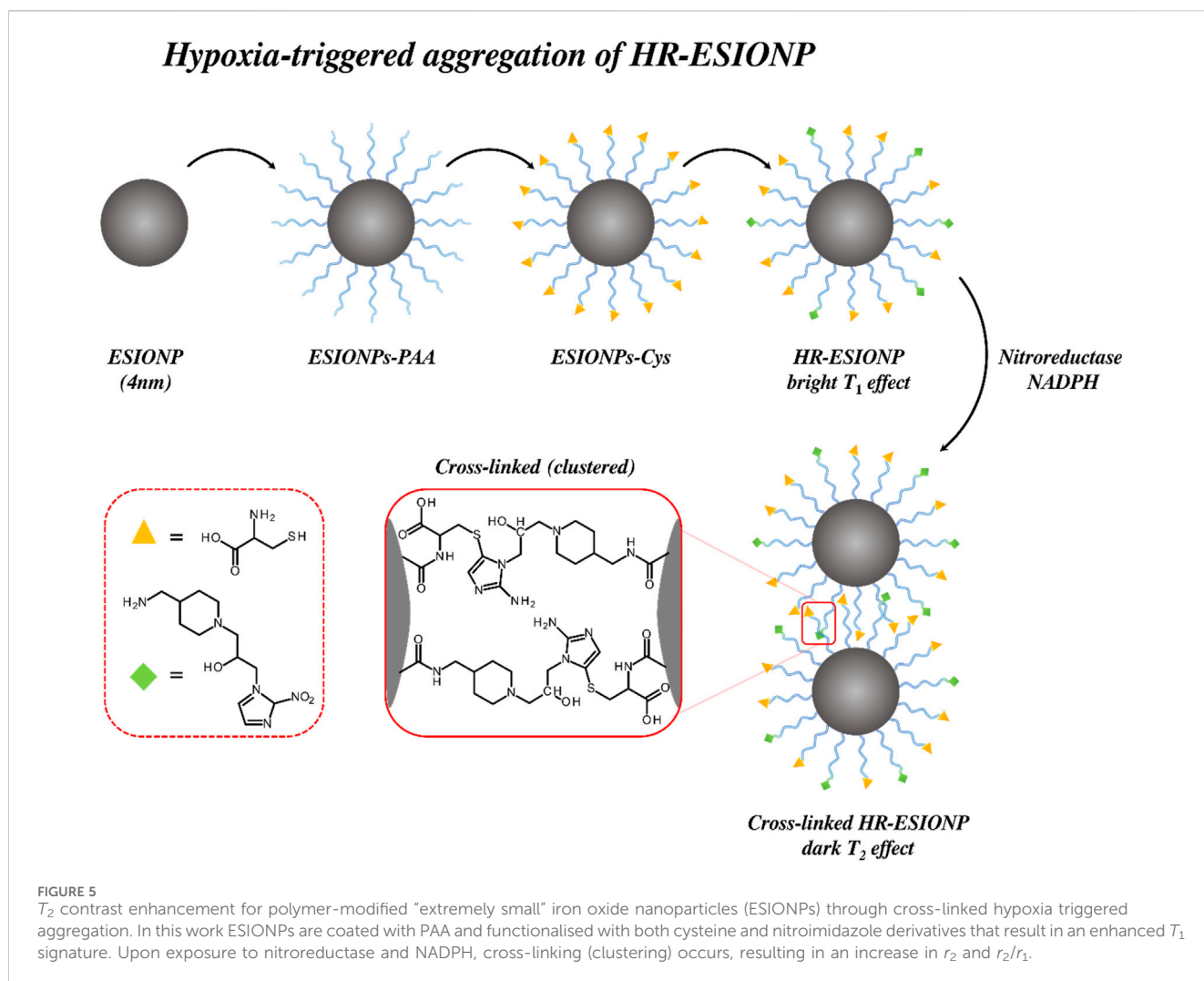
reported to be 3,000 Da (associated  $r_2 = 151.1 \pm 1.7 \text{ mM}^{-1} \text{ s}^{-1}$  at 9.4 T), conclusions mirrored in observations by Leal *et al.* (2015), Ramniceanu *et al.* (2016). These coating-optimised characteristics have also been observed with other polymers as illustrated, for example, by Cheah *et al.* where polydopamine (PDA), PAA, and PEG coated IONPs of tuneable sizes were investigated (PDA  $M_n = 189.6 \text{ Da}$ , PEG  $M_n = 5,000 \text{ Da}$ , and PAA  $M_n = 1800 \text{ Da}$ ) (Cheah *et al.*, 2021). All polymer-modified IONP formulations in this work exhibited good colloidal stabilities as resolved by dynamic light scattering (DLS), with the PAA-modified particles in particular displaying effective  $T_2$  contrast enhancement ( $r_2 = 75.3 \text{ mM}^{-1} \text{ s}^{-1}$  at 0.5 T).

While PEG and comparable polymers do impart a 'stealth' capacity for IONPs, a review of more than 5,000 publications on these particle systems by Wen *et al.* revealed that some 85% exhibited a rapid drop to half the original blood concentration within an hour of administration (Wen *et al.*, 2023). This finding suggests that more investigation needs to be done into PEG alternatives, such as modified lipids, or co-polymeric polymersomes (Carol *et al.*, 2015; Qi *et al.*, 2018). Such advancements would allow for the concentration of the CA to remain high through clinically-relevant periods, allowing for a lower initial dose. Current research aligns with this trend, with an increasing number of organic polymers now applied in the post-synthetic surface modifications of IONPs. Alternative organic functionalities, such as those provided by dendrimers or polymer micelles, are also now commonly employed as IONP protective coatings (Salimi *et al.*, 2018). While these studies show promise in addressing the systemic challenge of rapid clearance, further refinement is necessary to develop scalable, high contrast, and clinically approved agents with a true 'stealth effect' (Salimi *et al.*, 2018).

To achieve environmental-responsivity, an engineered triggered IONP aggregation can be employed to engender

significant switches in  $r_2$ . Lu *et al.* for example, have designed extremely small iron oxide nanoparticles (ESIONPs) with a diameter less than 4 nm, particles that typically exhibit  $T_1$  contrast in their monodispersed native state (Lu *et al.*, 2023). These particles were modified with polyacrylic acid (PAA) to improve dispersity in aqueous environments, and subsequently functionalised with nitroimidazole and cysteine derivatives, moieties capable of cross-linking in the presence of nitroreductase and NADPH, frequently present and overexpressed in hypoxic tumours as shown in Figure 5 (Yang *et al.*, 2017). The PAA coating on the ESIONPs enabled prolonged circulation and tumour site accumulation prior to this triggered aggregation.

Relaxivity measurements were taken at 0.5 T and 35 °C in PBS to examine the properties of the particles before and after exposure to a reducing environment. Prior to aggregation, the particles exhibited an  $r_2$  relaxivity of  $31.81 \text{ mM}^{-1} \text{ s}^{-1}$ , and an  $r_2/r_1$  ratio of 1.66 (ratiometric analyses facilitating concentration independent assessments) (Hagberg *et al.*, 2013). Upon treatment with NADPH and nitroreductase in a hypoxic environment,  $r_2$  was observed to increase to  $141.21 \text{ mM}^{-1} \text{ s}^{-1}$ , and the  $r_2/r_1$  ratio to 7.81 ( $\Delta r_2 = 109.4 \text{ mM}^{-1} \text{ s}^{-1}$ ). The shift in contrast-generating properties reflect the role of aggregation and clustering in dampening  $r_1$  relaxivity and increasing  $r_2$ . To examine *in vivo* performance, the particles were injected into mice bearing a 4T1 xenograft tumour, mimicking human breast cancer progression. The image intensity supported by the responsive particles was compared to that offered by the non-responsive particles, with the former darkening (and accumulating) in hypoxic tumour areas. The combination of  $T_1$ -to- $T_2$  contrast switching, and dark contrast enhancement meant these agents, then, acted as effective hypoxia-sensitive CAs. These studies, in combination, have exemplified the use of suitable organic



functionality, introduced onto the surface of IONPs through both grafting ‘to’ and ‘from’ approaches, in modulating IONP aggregation in a manner that is both tuneable and clinically pertinent.

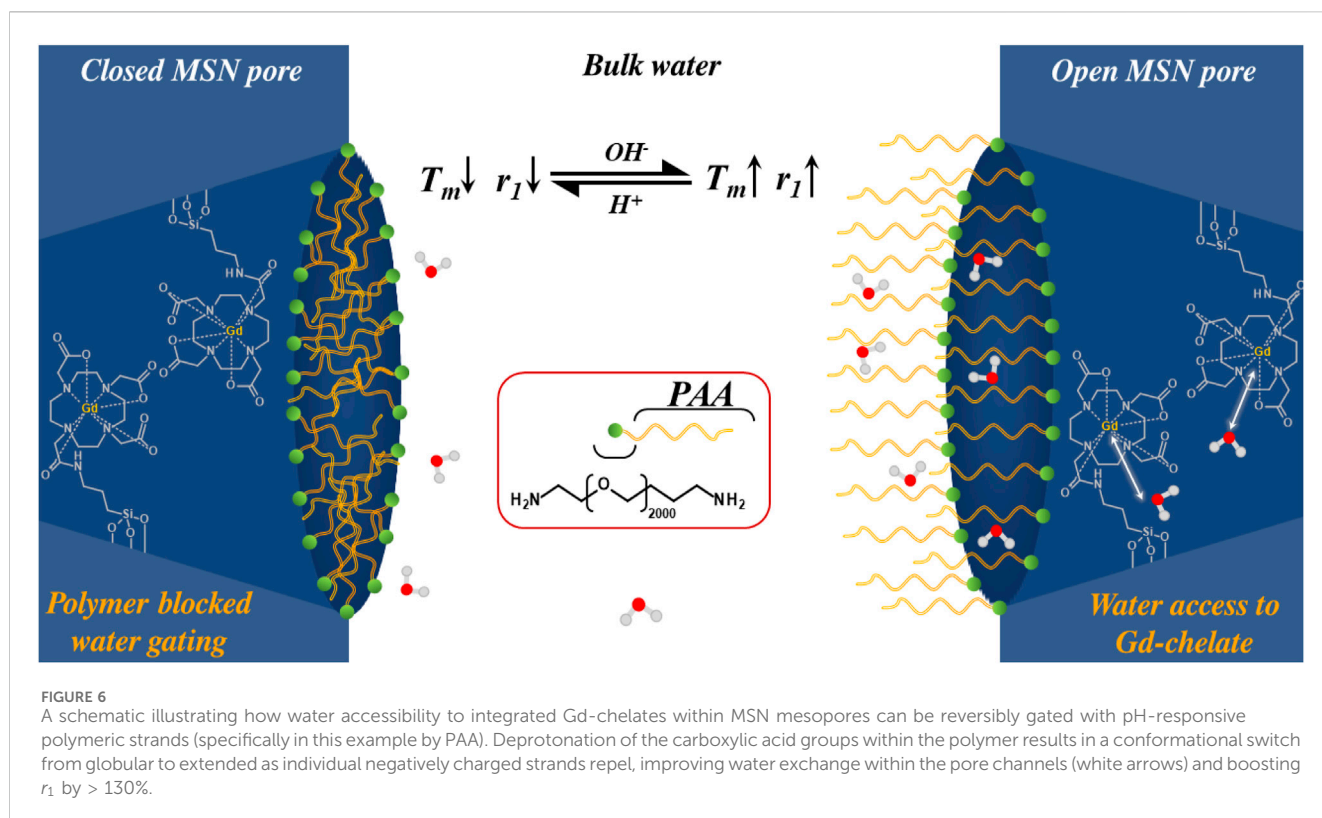
## 2.2 Polymer-supported MSNs

MSNs have gained significant popularity as a platform for the delivery of a variety of payloads, including drugs and contrast agents (Mohamed Isa et al., 2021), possessing several attractive physicochemical features including good colloidal stabilities and facile chemical surface tuneability (Wu et al., 2011). They can be readily functionalised with chemical moieties through post-synthetic silanol surface modification, including that where polymers are introduced to yield a passive solubilising coat, active targeting capability, or groups susceptible to protonation/deprotonation that support stimuli-responsivity (Yuan et al., 2020). In comparison to other nanoparticulate formulations, MSNs show particular clinical utility due to their good colloidal stability in the bloodstream, low toxicity and facile interfacial functionalisation; and characteristics that support complementary delivery and tumour-targeting capabilities (Lérida-Viso et al., 2023). Silica based

nanoparticles already have an established history in cancer imaging, with proven efficient clearance via hepatic and renal pathways, reinforcing their clinical translatability as a versatile platform (Benezra et al., 2011). From an MRI perspective, however, further studies are needed to comprehensively evaluate *in vivo*  $Gd^{3+}$  leaching kinetics from pore-doped MSNs (Carniato et al., 2018).

MSNs are typically coated with polymers in two distinct fashions, either through direct covalent binding or by electrostatic association. Covalent approaches most typically involve amide coupling or a surface-initiated polymerisation, with electrostatic assembly invoking supramolecular interactions between the charged portion of the polymer and the silica surface. Sodium alginate is, by way of example, a natural hydrophilic polysaccharide that has been widely employed for bio-responsive applications due to its negligible toxicity and high biodegradability (Sarkis et al., 2017). Li et al. have, for example, employed the aforementioned electrostatic association approach to yield dye cargo-entrapped gadolinium-doped MSNs (generated by an *in situ* doping of the MSN surface with  $Gd^{3+}$ ) coated with sodium alginate (Li Z. et al., 2022). Loss of electrostatic association between the MSNs and the alginate cap at a pH below the  $pK_a$  of the alginate carboxylate groups





( $pK_a < 4.5$ ) resulted in both triggered release of the fluorescent rhodamine B (RhB) dye and restored water access to the surface-doped paramagnetic  $Gd^{3+}$  ions. The latter displayed a corresponding five-fold switch in relaxivity, with  $\Delta r_1 = 40.57 \text{ mM}^{-1} \text{ s}^{-1}$  (at 0.5 T), of clinical note given that low pH is associated with endosome and lysosomal environments.

A very similar irreversible capping approach was adopted by He *et al.*, generating pH-responsive poly (*N,N*-dimethylacrylamide-co-THPA-functionalised *N*-(3-aminopropyl)methacrylamide) and poly (allylamine hydrochloride) surface coated MSNs loaded with  $Gd_2O_3$  nanoparticles (He *et al.*, 2019). As the pH decreases below pH 5.0, the poly (allylamine hydrochloride) protonates, reversing the previously favourable supramolecular association of the polymer to the charged particle surface and exposing the MSN pores. In doing so, the MR-active integrated  $Gd_2O_3$  particles were, again, released, resulting in a corresponding switch in  $r_1$  with  $\Delta r_1 = 5.71 \text{ mM}^{-1} \text{ s}^{-1}$  (at 3 T) as pH decreases from pH = 7.4 to pH = 5.0.

The above examples are indicative of potentially large ( $\Delta r_1 > 150\%$ ) triggered responses but are, in essence, irreversible. With covalently modified particles, the permanence of the polymer coating can support partially or fully reversible relaxivity modulations. When designing a particle that is covalently modified with a polymer, changes to the conformation and chemical character of the organic layer can be a potent means to generate reversible relaxivity switches. One approach was demonstrated in work by Pellico *et al.* who synthesised MSNs covalently functionalised with PEG/PAA block copolymers (Pellico *et al.*, 2019b). The use of a block copolymer enabled improved colloidal stability through the PEG block while also eliciting a pH-responsive, in this case a fully reversible, switch in

$T_1$  imaging capabilities through the PAA block. This assumes a globular conformation below pH 4.8, resulting in a tight capping of the MSN pores and reducing diffusive water access to integrated Gd-motifs, diminishing relaxivity. As illustrated in Figure 6, above pH 4.8, individual polymer strands extend into solution (Sarkar and Somasundaran, 2004), significantly improving water accessibility to the integrated paramagnetic complexes and inducing a significant, and reversible, >130% switch in  $r_1$  ( $\Delta r_1 = 11.4 \text{ mM}^{-1} \text{ s}^{-1}$ ) as pH is traversed from pH 3.0 to pH 10.0.

In a similar work by Yuan *et al.*, Gd-DOTA doped MSNs were surface modified with the pH-responsive poly (methacrylic acid) (pMAA), generated by use of a surface-initiated RAFT (SI-RAFT) polymerisation process (Figure 7). (Yuan *et al.*, 2023) Notably, introduction of the polymer coat enabled  $\tau_M$  to be modulated, directly influencing relaxivity. In this case, the reversibly charging polymer is not acting as a physical barrier to water access but as a potent hydrogen bond acceptor in the charged state; this is proposed to then modulate the relative viscosity of the local water pool and an SBM-theory supported promotion of relaxivity (Yuan *et al.*, 2023; Ju *et al.*, 2007; Sachar *et al.*, 2020). The  $r_1$  relaxivity was assessed at 1.4 T, across a pH range of 4.0–9.0, with a significant switch of >180% ( $\Delta r_1 = 30.3 \pm 3.2 \text{ mM}^{-1} \text{ s}^{-1}$ ) as the  $pK_a$  of pMAA was traversed. As these values exceed the theoretical maximum for the inner sphere contributions to total relaxivity, the shifts were assigned to increased outer sphere effects (De León-Rodríguez *et al.*, 2015).

It is clear from these that suitable polymers, either electrostatically or covalently associated with NPs within inorganic-organic hybrid architectures, are able to support significant (>130%) pH-dependent enhancements in relaxivity upon traversal of their  $pK_a$ , by virtue of conformational and/or

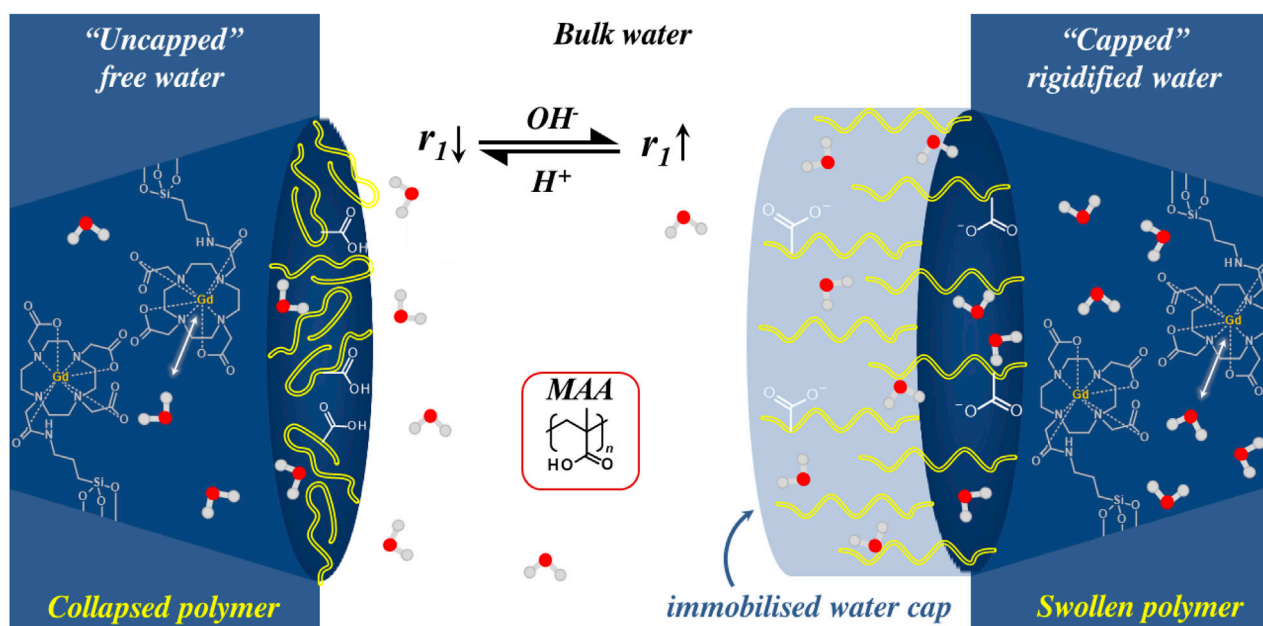


FIGURE 7

A schematic illustrating a pH-responsive  $T_1$  switch as observed with pMAA-Gd-MSNs. The pMAA shell in its uncharged/collapsed conformation allows limited hydration that results in a moderate  $r_1$ . When the pH traverses the  $pK_a$  of MAA, the polymer shell charges and swells, dramatically increasing local hydration. The so generated “water cap” (light blue shaded region) boosts  $r_1$  by > 180%.

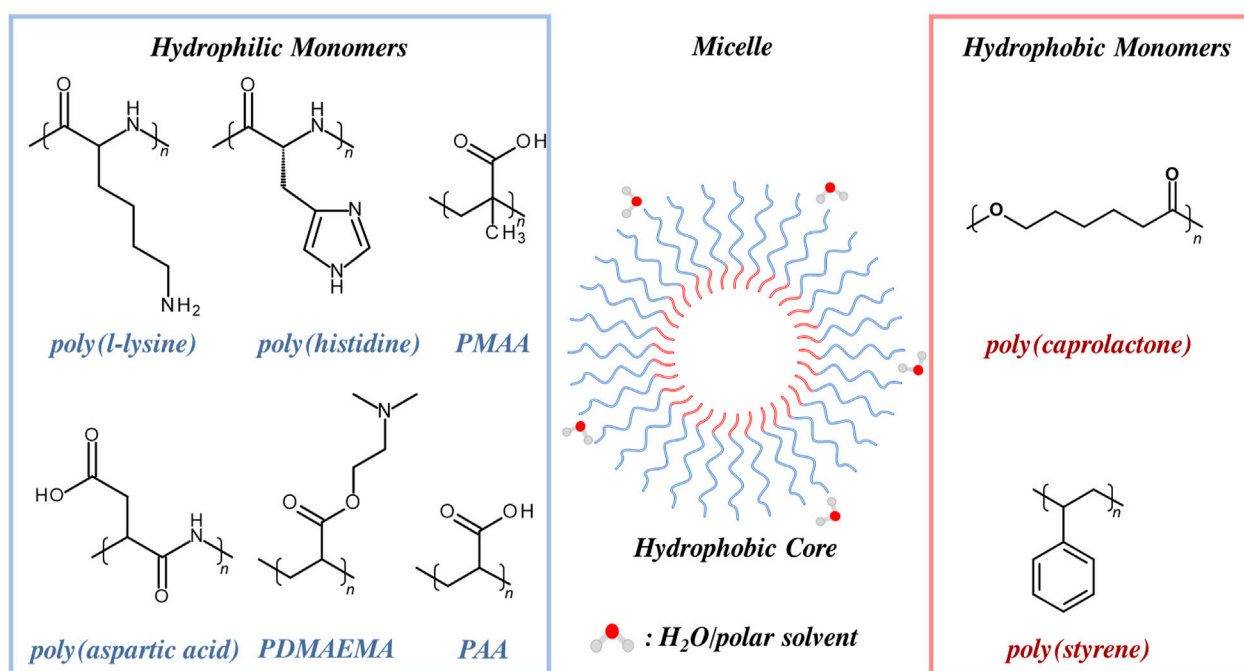


FIGURE 8

Examples of hydrophobic and hydrophilic polymers commonly utilised in micelle generation (hydrophobic core shown). Micelle surface functionality can be tuned through monomer selection. Carboxylic acid containing monomers (PAA, PMAA, PDMAEMA) and amino acid monomers (poly (aspartic acid), poly (L-lysine), poly (Histidine)) can be utilised to generate a physiologically-relevant pH-response. Hydrophobic monomers (PCL and PS) are also displayed.

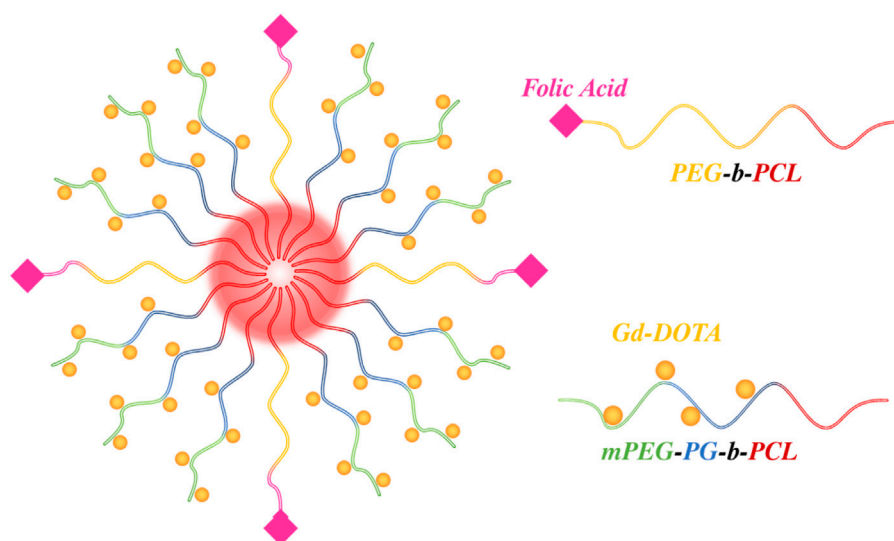


FIGURE 9

Schematic illustrating the micelle architecture employed by Cao *et al.* where MRI active Gd-DOTA moieties were tethered to individual mPEG-PG-*b*-PCL strands. Conjugation of folic acid to PEG-*b*-PCL polymeric strands enabled an active targeting capability (Yuan *et al.*, 2025).

electrostatic changes at individual monomer units. Only those approaches which employ covalent surface-bound polymers can support a reversible switch in image contrast upon cycling of bulk pH. The customisable nature of such polymer coatings, empowered through monomer choice, showcases the generation of inorganic-polymer hybrids as an attractive method by which a range of (multi) stimuli-responsive (*e.g.*, pH in combination with redox/light/enzyme) CAs could be considered. The so-generated responsive imaging probes, tailored to be responsive to a biological condition of interest, would enrich our understanding of disease pathology.

### 3 Paramagnetic organic nanoparticles

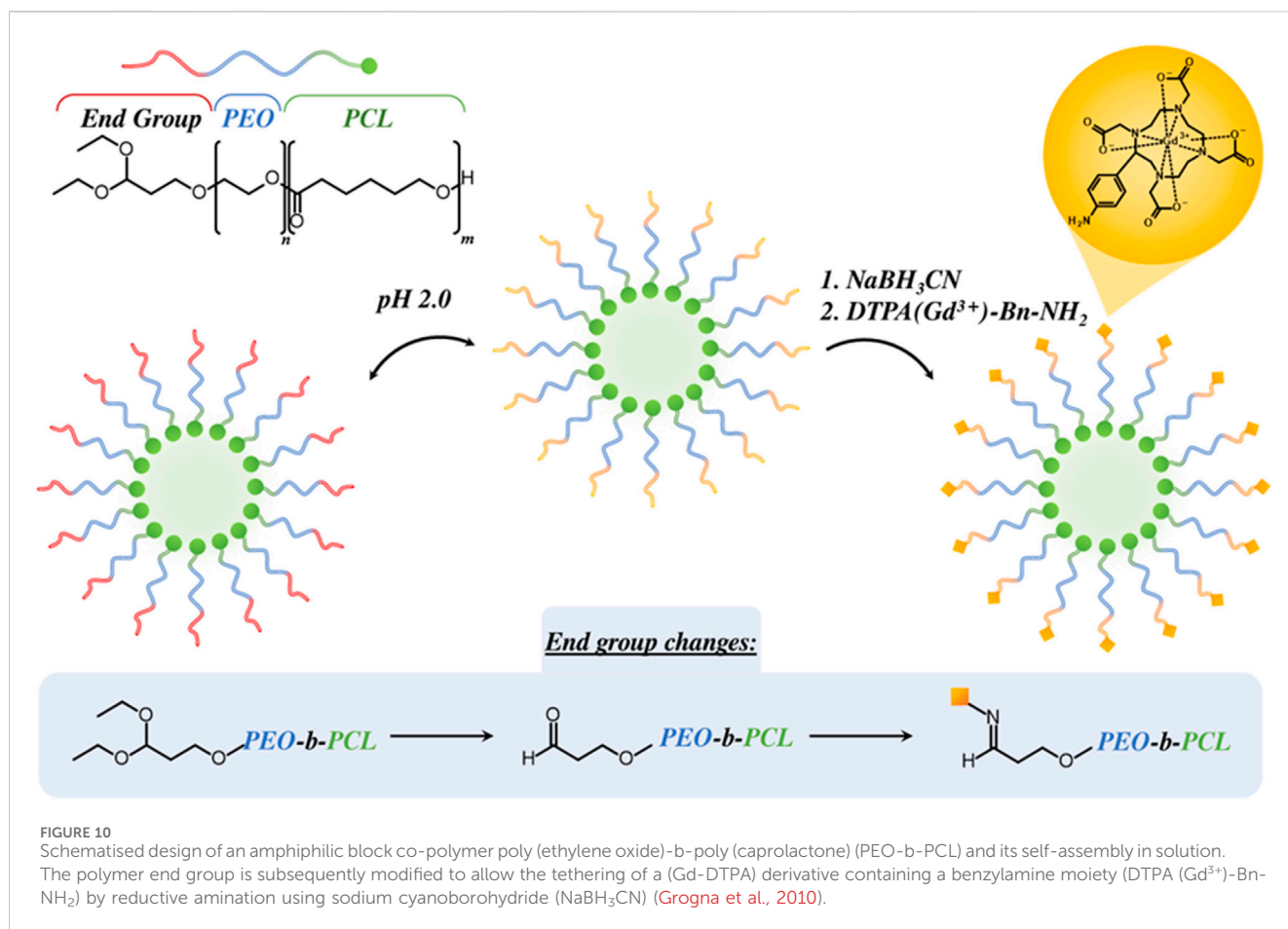
#### 3.1 Polymer micelles

Polymer micelles are typically composed of block co-polymers, containing both a hydrophobic and hydrophilic repeating unit, which self-assemble into micelles on suitable solvent exposure (Figure 8). (Yorulmaz Avsar *et al.*, 2019) Hydrophilic blocks will naturally form the solubilising and stabilising micellular shell in aqueous conditions, with the hydrophobic block associating with the micelle core. Amphiphilic block-copolymers have been widely investigated as platforms for contrast agents, drug delivery, and catalysis due to their high associated biocompatibility, improved colloidal stabilities over non-modified inorganic nanoparticle analogues (*e.g.*, MSNs and IONPs), customisability, and tissue penetration ability (Deng *et al.*, 2021). Polymer micelles are, for example, known to be able to capitalise on the tumour site accumulation characteristics of the EPR effect whilst avoiding glomerular filtration, promoting beneficially elongated blood retention times *in vivo* (Svenson, 2012). Physical properties, including the size and shape of the polymer micelle can be easily customised at the point of synthesis. They can be doped with a paramagnetic contrast agent such as Gd-DOTA through strand

covalent attachment, or through physical entrapment within the micellar core (Kazunori *et al.*, 1993). As prior discussed in Section 1.3., polymer micelles also offer improved relaxivities over their molecular analogues since the anchored or entrapped MR-active complexes have notably elongated rotational correlation times compared to the normal rotational characteristics of a molecular chelate (Lux and Sherry, 2018; Li *et al.*, 2012).

In one such example, Cao *et al.* synthesised amphiphilic block copolymers containing methoxy poly (ethylene glycol)-polyglycerol (Gd-DOTA)-*b*-polycaprolactone (mPEG-PG (Gd-DOTA)-*b*-PCL) and folate terminated PEG-*b*-PCL which generated a micellular nanoparticle when mixed in a 4:1 ratio (Figure 9). (Cao *et al.*, 2017) Conjugation of Gd-DOTA to the mPEG-PG-*b*-PCL polymer strands supported a significant (rotationally-enabled) enhancement in MRI contrast, (corresponding  $r_1 = 14.01 \text{ mM}^{-1} \text{ s}^{-1}$  at 0.5 T, a >250% improvement over traditional clinically employed Magnevist). Folate was chosen for its tumour targeting capability, facilitating rapid internalisation of the micelles by receptor-mediated endocytosis, common for tissues that overexpress folate receptors, such as cancer (Kularatne and Low, 2010; Wang *et al.*, 2023).

While PEG is often chosen as the hydrophilic micellular component, poly (ethylene oxide) has also been reported. For example, Grogna *et al.* designed polymer micelles composed of poly (ethylene oxide)-*b*-poly (caprolactone) (PEO-*b*-PCL) as illustrated in Figure 10 (Grogna *et al.*, 2010). A Gd-diethylenetriaminepentaacetic acid (Gd-DTPA) derivative containing a benzylamine moiety was subsequently bound to the PEO block by a reductive amination process at the integrated amine groups. In this work, the effect of polymer strand flexibility on relaxivity was examined by investigating differences in  $r_1$  through polymer strand length ( $1,300 < M_n < 4,000$ ). It was observed that shorter polymer strands yielded the highest  $r_1$  ( $11.9 \text{ mM}^{-1} \text{ s}^{-1}$ ), ascribed to a more restricted free rotation of the Gd-DTPA probe and associated elongated  $\tau_R$  compared to the longer polymer strand micelles.

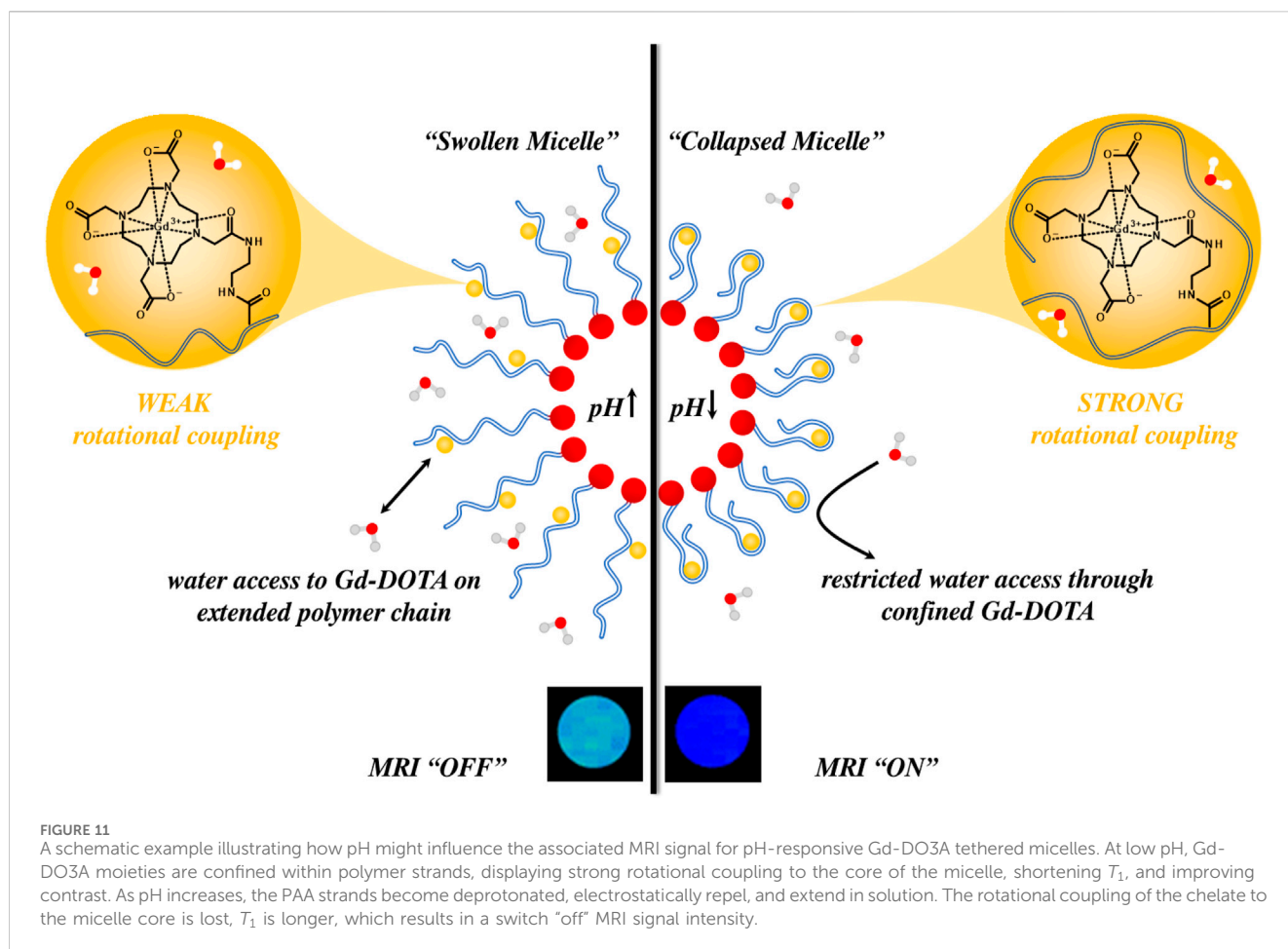


In terms of particle composition, monomers can be chosen to generate a micelle that is responsive to a specific environmental stimulus, such as deviations in pH or temperature. In the former, the selected monomer contains ionisable functional groups (e.g., carboxylic acid or amine) which have an associated  $pK_a$  that falls within a physiologically relevant range (pH 6.0–8.0). Upon traversal of the associated pH the native nanoparticulate structure can either conformational switch, or fragment entirely, as ionisable polymer blocks charge. Such changes can be used to modify parameters that control the (contrast-generating) relaxivity such as accessibility of water to the paramagnetic centres, and the rotational order parameter ( $F^2$ ), the latter describing the degree of independence of the paramagnetic complex rotation from the particulate scaffold. Examples of monomers utilised within assemblies include acrylic acid (AA), methacrylic acid (MAA), or dimethyl aminoethyl methacrylate (DMAEMA), all of which exhibit altered physicochemical properties at accessible, and relevant solution pH. Further examples can be generated from polymers comprised of amino acids, such as poly (aspartic acid), poly (L-lysine) and poly (histidine), synthesised from their respective N-carboxyanhydrides. These exhibit a pH-response through protonation/deprotonation of free carboxylic acid/amine groups, as well as through acid-regulated bond cleavage, and are additionally attractive as a result of their high associated biocompatibility (Yuan et al., 2025; Guo et al., 2020). Nitroimidazole derivatives and

disulfide bridges can be post-synthetically added to the polymer strands, enabling demicellisation in reduction-sensitive environments, e.g., tumour cells (Xu et al., 2022).

A more detailed theoretical consideration of the effects of modulating chelate rotational rigidity in these structures was explored further in work by Ellis *et al.* Here, poly (acrylic acid)-b-poly (styrene) (PAA-b-PS) block copolymers were synthesised, with individual PAA polymer strands subsequently functionalised with Gd-DO3A by amide coupling (Ellis et al., 2023). The PAA segment of the micelle remains largely deprotonated at normal physiological pH (7.1–7.4), above the  $pK_a$  of PAA, occupying an “extended” conformation since adjacent carboxylate groups electrostatically repel. As environmental pH falls below the  $pK_a$  of PAA, the carboxylate groups are protonated, PAA strands become much more mechanically coupled to the micelle core, with relaxivity significantly increasing as the rotational characteristics of Gd-DO3A became akin to the nanoparticle scaffold. This caused a notable switch in  $r_1$ , specifically  $\Delta r_1 = 9.70 \text{ mM}^{-1} \text{ s}^{-1}$  at 1.4 T as pH decreased from pH 7.5 to pH 4.0, an observation that was fully reversible. The order parameter was considered central to this. When  $F^2$  is equal to 0, the rotation of the paramagnetic complex is entirely independent of particle rotation, and when  $F^2$  is equal to one the complex is unable to rotate independently of the particle, *i.e.*, the mechanical coupling is “absolute” (Carniato et al., 2010). A  $F^2$  closer to one would be observed for rigid inorganic nanoparticles, such as small-pore MSNs or tightly contracted





polymer micelles, and is associated with an increase in the local rotational correlation time of the paramagnetic centre, and therefore, by extension,  $r_1$ . By designing a nanoparticle configuration that supports a locally triggered change in paramagnet-chelate rotation through polymer strand rigidity (see Figure 11), dramatic switches in  $r_1$  can, then, be achieved.

While the above examples demonstrate a generalised reversible swelling approach to modulate generated image contrast, as noted, a destructive (irreversible) fragmentation approach can also be employed to modulate relaxivity. In work by Kim *et al.*, for example, block copolymers comprised of PEG-*b*-poly (L-lactic acid), tethered to Gd-pentetic acid (DTPA) and PEG-*b*-poly (L-Histidine) (PEG-*b*-p (L-His)), for pH-sensitivity, were designed that self-assembled to form micelles in aqueous solution (Figure 12). (Kim *et al.*, 2014) At physiological pH ( $\approx 7.4$ ) the micelles retained a uniform and stable morphology, fragmenting on exposure to an acidic environment below the  $pK_a$  of p (L-His) ( $pK_a = 6.8$ ) as the imidazole groups protonate and undergo a hydrophobic to hydrophilic transition. The prior confined Gd-DTPA moieties, with initially highly-restricted water access, are released on fragmentation, improving  $\tau_M$ , and resulting in a productive switch in  $r_1$  ( $\Delta r_1 = 3.45 \text{ mM}^{-1} \text{ s}^{-1}$  at 4.7 T).

Physical and downstream relaxometric changes in purely organic nanoparticles may also be triggered by exposure to a reducing environment, specifically GSH levels present in cancer are typically 2–5 times higher than healthy tissue, with an intracellular concentration ranging from 1–10 mM (Ge *et al.*,

2024; Ding *et al.*, 2021). Polymeric nanoparticles with engineered disulphide motifs can, specifically, support a responsive character through bond reductive cleavage on exposure to, *e.g.*, elevated GSH, triggering disintegration of prior cross-linked polymeric strands and exposing encapsulated MR active agents such as Gd-chelates. This engineering has, for example, been demonstrated in work by Sigg *et al.*, with the reductive cleavage of amphiphilic heparin-poly (dimethylsiloxane) (hepPDMS) block-copolymers, complexed with  $\text{Gd}^{3+}$  (Sigg *et al.*, 2016). In this work, polymer hepPDMS strands were co-assembled with an integrated disulfide-linked peptide (peptide sequence  $\text{H}_2\text{N-H}_3\text{-X-[W-dL]}_3\text{-W-CONH}_2$ ) to form micelles, the latter containing the reducible motif  $\text{X} = (-\text{CH}_2)_2\text{-S-S-(CH}_2)_2\text{-NH-CO-(CH}_2)_2\text{-CO-}$  that connects oligohistidine (H3SSgT) and hydrophobic L-tryptophan-D-leucine units. This reducible linker restrained the oligohistidine from extending into solution and exposing the integrated paramagnetic chelates, limiting water accessibility. Exposure to the reducing agent dithiothreitol (DTT) was observed to cleave the cross-links, resulting in improved water flux to the  $\text{Gd}^{3+}$  motifs, and enhancing associated image contrast ( $\Delta r_1 = 10.20 \text{ mM}^{-1} \text{ s}^{-1}$  at 3 T).

### 3.2 Stimuli-responsive liposome CAs

Liposomes are inherently biomimetic as a result of their vesicular structure, comprising a synthetic amphiphilic bilayer

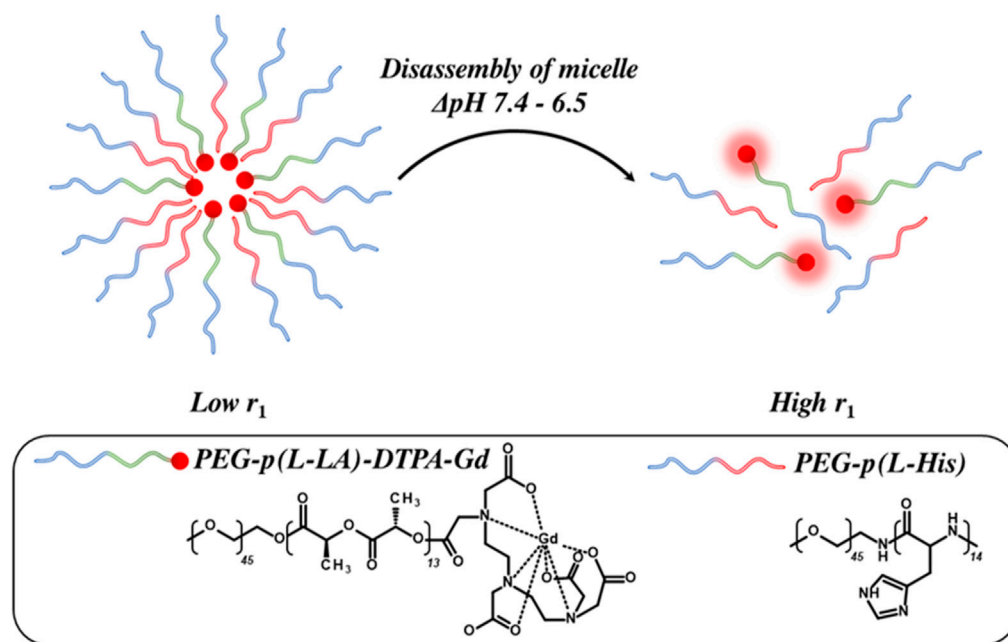


FIGURE 12

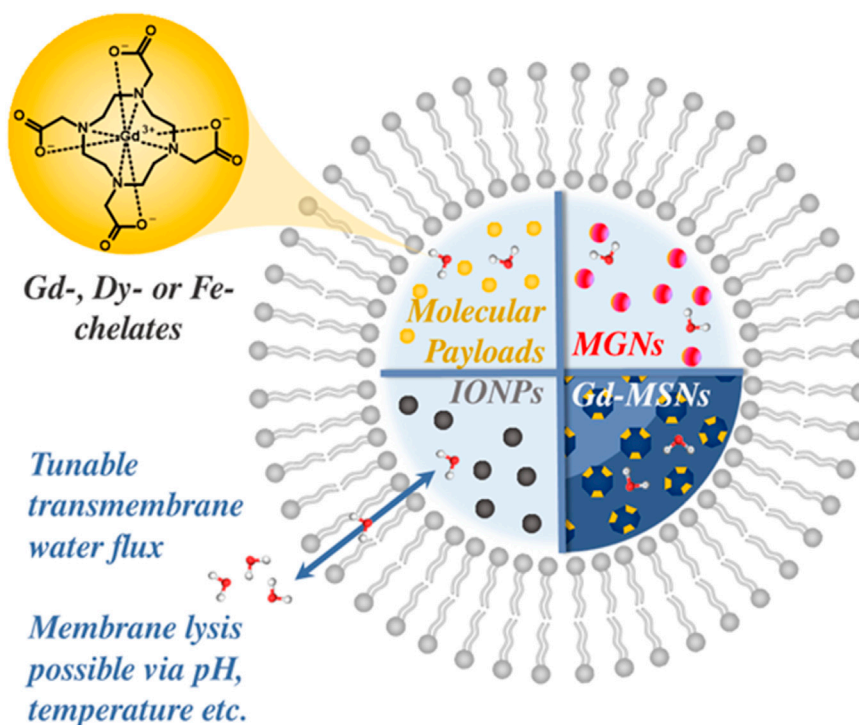
A schematic illustrating amphiphilic block copolymer methoxy poly (ethylene glycol)-*b*-poly (L-His) (PEG-*p* (L-His)), and methoxy poly (ethylene glycol)-*b*-poly (L-lactic acid)-diethylenetriaminopentaacetic acid dianhydride-gadolinium (PEG-*p* (L-LA)-DTPA-Gd) micelles. At physiological pH (7.4) the block copolymers self-assemble to generate colloiddally stable micelles that fragment below the  $\text{pK}_a$  of *p* (L-His), improving water accessibility to the core confined Gd-chelate.

membrane. They offer high levels of colloidal stability, advantageous circulation half-lives, and beneficial *in vivo* degradation (Mulder et al., 2006; Rideau et al., 2018). These simple models of a cell can be productively interfaced with contrast generation by the encapsulation of MRI-active payloads (typically Gd- (Ayyagari et al., 2006), Dy- (Castelli et al., 2009) or Fe- (Chowdhury et al., 2023) chelates). Nanoparticulate probes, such as IONPs (Cardellini et al., 2024; Li Q. et al., 2022; Zhang N. et al., 2022; Waqar et al., 2022), quantum dots (QDs) (Xu et al., 2018),  $\text{Mn}_3\text{O}_4$  nanoparticles (MGNs) (Thomas et al., 2023) or Gd-MSNs (Sun et al., 2018), can also be trapped with the hydrophilic core (Figure 13). (Soenen et al., 2011) Accommodation of these MRI active probes within liposomal scaffolds prevents non-specific interactions with serum proteins, offering protection against degradation during *in vivo* delivery, whilst additionally reducing opsonisation of the probe by RES (reticuloendothelial system) processes (Mulder et al., 2006; Zhang J. et al., 2022). In terms of clinical translatability, liposomes represent an established formulation for cancer nanomedicines, with Doxil (doxorubicin encapsulated within a PEGylated LUV) approved for oncology treatment since the 1990s (Cao et al., 2017; Wang et al., 2018).

Liposomes, by their nature, offer a platform through which responsive MR contrast, in the presence of stimuli such as pH (Løkling et al., 2001; Torres et al., 2011), temperature (Zhang et al., 2015; Kono et al., 2011), ultrasound (Kim et al., 2016) and/or light (Reefing et al., 2019; Liu et al., 2022) can be achieved, as internally doped liposomes are initially in an 'MR silent' state due to the limited water diffusion through the hydrophobic membrane. Inherent in derived CAs is an ability for this natural water barrier to be modulated. The magnitude of

responsivity (relaxivity switch) may be increased by further improving the contrast 'off-state' to be more MRI 'silent' in nature, achieved by reducing the rate of water diffusion or bilayer permeability, e.g., through greater cholesterol loading (Saito and Shinoda, 2011). Central to this also is noting that inorganic imaging agents can either be readily encapsulated within the hydrophilic core of the vesicle or tethered to the bilayer itself (see Figure 14). In the latter, Gd-chelates are appended to the phospholipid hydrophilic heads and MR contrast generating. This provides a platform whereby relaxivity can be modulated by cleaving the pendant MR-responsive arm and modulating  $\tau_R$  (Accardo et al., 2009). This is, once again, a reflection of the dramatic change in tumbling rate of a  $\sim 100$  nm liposomal structure and a free Gd-chelate ( $\sim 1$  nm hydrodynamic size).

Liposomal MR responsivity may therefore be realised in two ways: destructively through lysis or fragmentation of the bilayer, triggering the controlled release of the  $T_1$  and/or  $T_2$  active payload, or non-destructively through selectively switching the permeability or transmembrane water flux (Mura et al., 2013; Nedyalkova et al., 2017; Fossheim et al., 1999). The latter is different to the mechanisms prior discussed for responsive micelle contrast generation in Section 3.1 where relaxivity is switched by either inducing a conformational change where local environment differs from the norm, or by inducing fragmentation entirely. The destructive approach in liposomes is generally used to achieve pH- or temperature-responsivity at regions of disease (generally acidic conditions and thermal stress), greatly switching water flux to initially integrated MR-active chelates (Yao et al., 2021). MR responsivity can also be achieved by chemically modifying



**FIGURE 13**  
Phospholipid bilayers allow for the encapsulation of  $T_1$  or  $T_2$  active payloads, such as molecular chelates (e.g., Gd-DOTA) or MR-active nanoparticles (Gd-MSNs, IONPs or MGNs). The rate of transmembrane water flux can be modulated in generating a responsive liposomal-based CA. Removal of the phospholipid membrane through lysis, or integration of transporter motifs that can retrospectively restore water flux, can enable a means by which relaxivity is modulated.

hydrophobic phospholipid tails within the bilayer to introduce either stimuli-switchable, *i.e.*, altered membrane permeability, or stimuli-cleavable, *i.e.*, directly impacting  $\tau_R$ , motifs (Figure 15).

In an example of a triggered manipulation of liposome membrane permeability, Simon *et al.* generated switchable vesicles by introducing azo functionalities within phospholipid tails to enable UV-responsive MRI contrast (Pitchaimani *et al.*, 2016). This tuneable membrane permeability specifically leveraged the structural impact of the azobenzene stereoisomerism with the *cis* (Z) isomeric form (produced through 370 nm irradiation) disrupting phospholipid packing, leading to an enhanced membrane permeability to water. This optical switching was demonstrated to be reversible, displaying UV-switchable MR contrast between MRI 'on' and 'off' states across three cycles.

Hyperthermia-responsive liposomes can be produced by using dipalmitoyl-phosphatidylcholine (DPPC), as demonstrated by Alawak *et al.* (2020). Here, MR active DPPC liposomes were shown to modulate diffusive water access to core-confined Gd-DTPA, with the thermo-responsive DPPC ultimately imparting a responsive  $r_1$  capacity ( $\Delta r_1 = 0.91 \text{ mM}^{-1} \text{ s}^{-1}$ ). As temperature was increased from 37.8°C, a DPPC gel-liquid phase transition (to a progressively more disordered state at  $T = 41.3^\circ \text{C}$ ), resulted in the lipid membrane 'melting', altering the inherent bilayer permeability, switching on contrast with concurrent doxorubicin release from the micelle core. Alternatively, the liposomal membrane can be fragmented through external deviations in pH. This was demonstrated in work by Li *et al.* who encapsulated Cu-doped

IONPs (size  $15 \pm 5 \text{ nm}$ ) within the core of hydrogenated soy phosphatidylcholine (HSPC) liposomes. Acid-triggered biodegradation ( $\text{pH} < 7.0$ ) enabled switchable  $T_2$  contrast as the lipid head group became protonated at low pH, destabilising the bilayer and resulting in membrane lysis and a darkening of  $T_2$ -weighted MR images (Thomas *et al.*, 2023; Paliwal *et al.*, 2015).

Irreversible rupture-based approaches to triggered MR contrast change can also be achieved through acid/UV induced bond cleavage from liposomal peripheries. In a switch "off" example of this, Liu *et al.* introduced a Gd-DTPA and *o*-nitrobenzyl (ONB) modified phospholipid within a tightly-packed vesicular membrane (Liu C. *et al.*, 2019). In this work, the liposomes were also internally doped with DOX for concurrent drug delivery. Initially, the covalent tethering of the Gd-chelates to the phospholipids significantly reduced (as we have noted previously) the tumbling rate of the  $\text{Gd}^{3+}$  moiety compared with a typical Gd-chelate (modulating  $\tau_R$ , boosting initial  $r_1$ ). This example, then, presents dual-trigger release/hydrolysis capabilities (pH and light), whereby subsequent UV cleavage of the ONB results in the release of Gd-DTPA and a concurrent decrease in rotational correlation time,  $\tau_R$  of the Gd moiety, and relaxometric switch.

The abnormal local concentrations of oxidising species (and reduced pH) at tumour tissue can be exploited in the generation of redox-responsive MRI contrast, which can report on, for example, the elevated reactive oxygen species (ROS) presence at such sites. In work by Thomas *et al.* a pH- and ROS-responsive probe was developed based on the encapsulation of  $\text{Mn}_3\text{O}_4$  nanoparticles

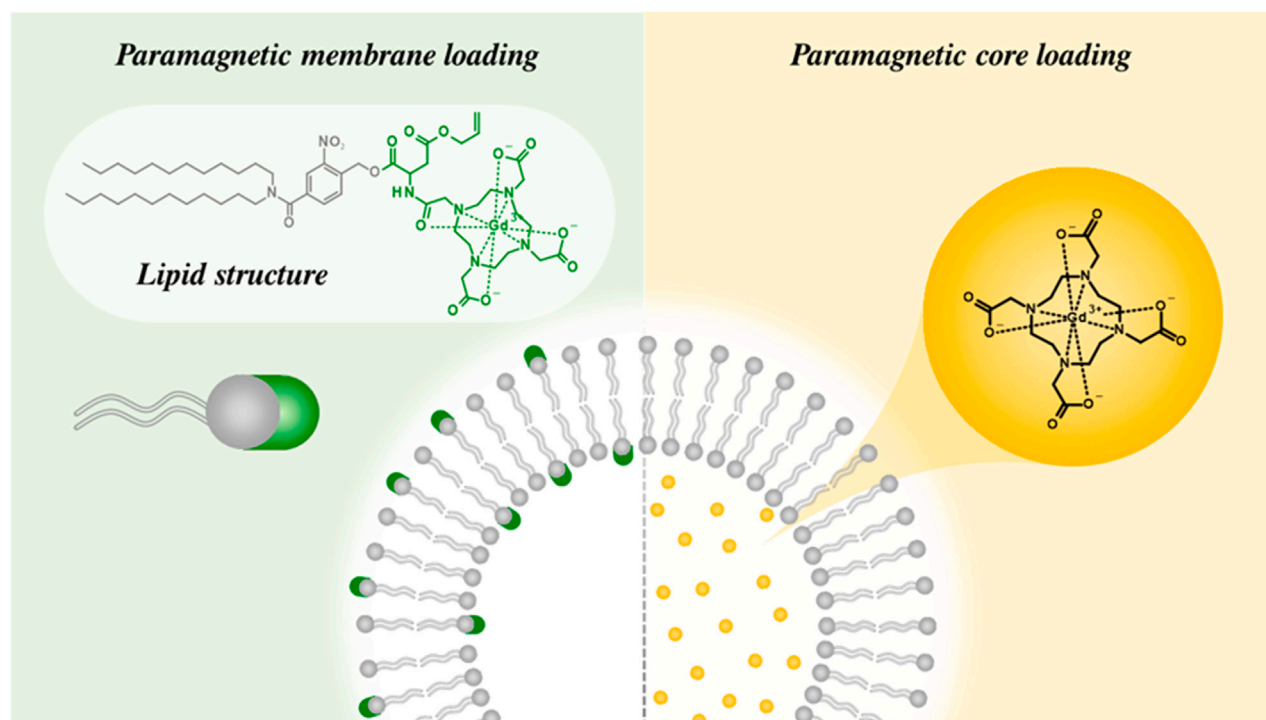


FIGURE 14

A schematic illustrating the means by which liposomes can be modified to be MR-active. Gd-chelates can either be encapsulated as a payload within the aqueous internal pool or covalently tethered to the phospholipid structure of the liposomal membrane (Liu C. et al., 2019).

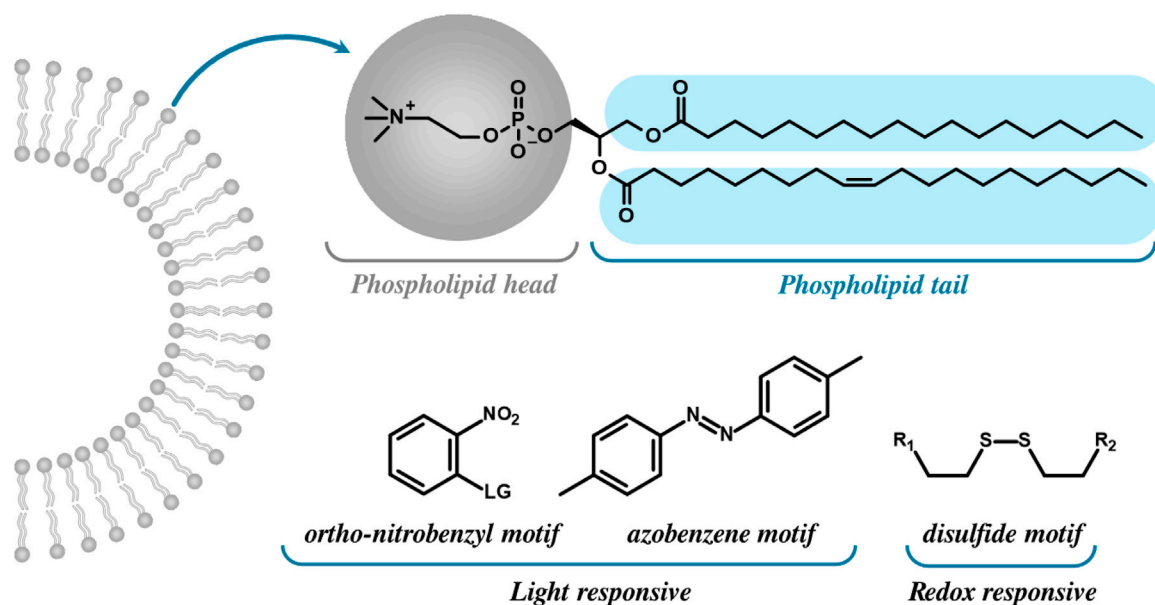
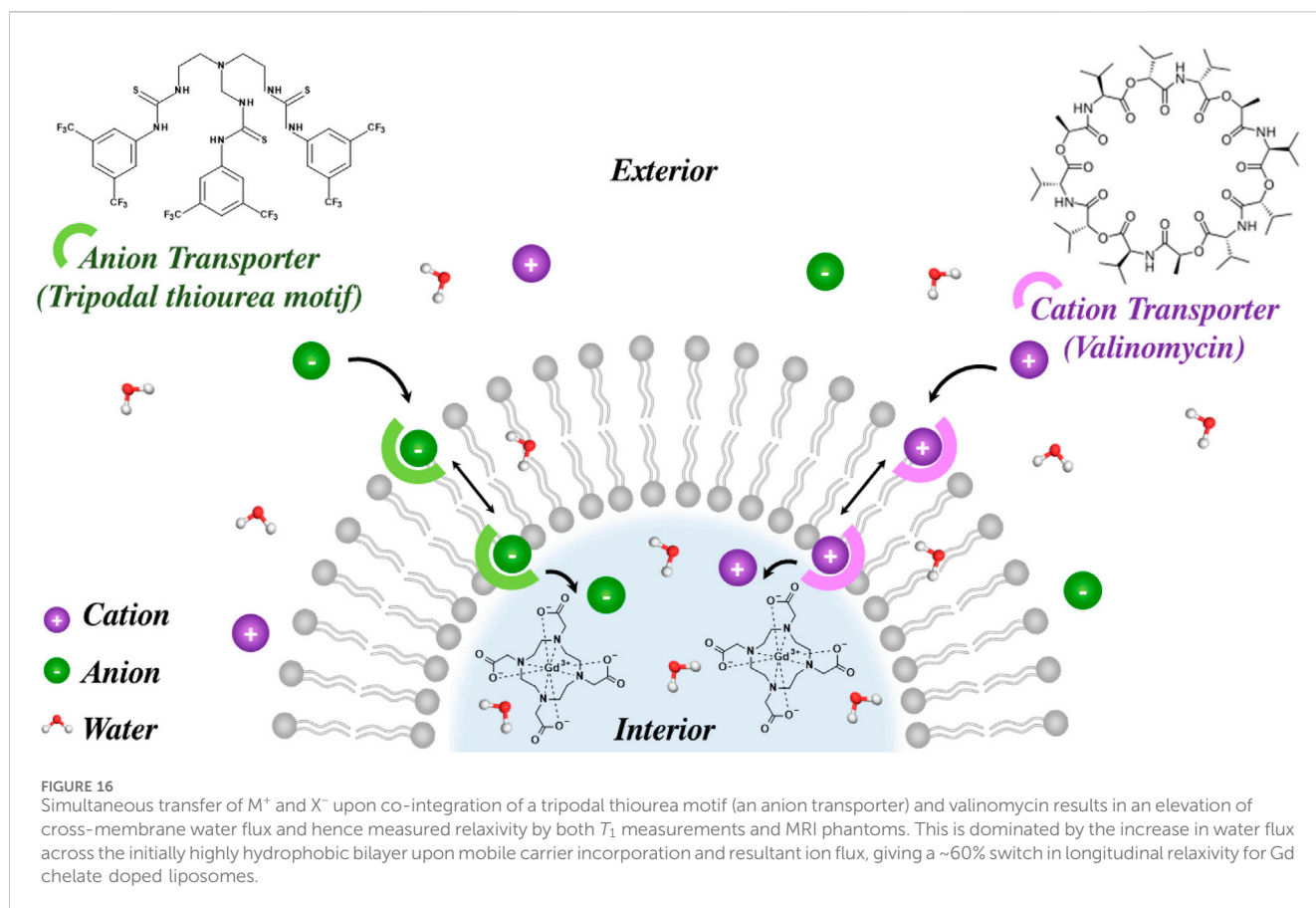


FIGURE 15

Bilayer structures can be chemically modified to become stimuli responsive in nature by incorporating light or redox responsive motifs into the phospholipid heads or tails. This is generally achieved by using a linker which can be cleaved upon exposure to certain stimuli, for example, the reduction of disulfides by TCEP (tris(2-carboxyethyl)phosphine hydrochloride) (Yao et al., 2021). Whilst integrated ortho-nitrobenzyl and azobenzene motifs can be reduced under certain conditions, as they are primarily light-responsive in nature. Azobenzenes integrated into phospholipid tails, for example, undergo E/Z photoswitching (Simon et al., 2022), altering the membrane permeability/diffusive water access. Phospholipid heads modified with ortho-nitrobenzyl groups, as an alternative, undergo photolysis and resultant cleavage of the leaving group such as a tethered paramagnet (Yao et al., 2021).





(size 20–30 nm) within PEGylated liposomes (composed of 1,2-distearoyl-sn-glycero-3-phosphoethanolamine-PEG, DPPC, and cholesterol) (Thomas et al., 2023). Switchable  $T_1$  contrast (enhancement) was generated here upon reduction of the  $Mn_3O_4$  nanoparticles, the production of highly paramagnetic  $Mn^{2+}$  and a  $\Delta r_1 = 3.84 \text{ mM}^{-1} \text{ s}^{-1}$  at 3 T.

These approaches so far generally follow one of two mechanistic approaches to modulate relaxivity in the presence of a particular external stimulus, depending on whether the liposome is either membrane modified or internally doped with the MRI signal-generator moiety. In the case of the former, a specific change in  $\tau_R$  results from covalent cleavage of the paramagnetic chelate. Alternatively, for the latter, the initially very limited diffusive water access across the lipid bilayer is reversed by altering membrane permeability, enhancing  $\tau_m$ , in a manner that may or may not be reversible (e.g., a destructive or geometric isomerism switch of bilayer motifs).

In work by Duncan et al., a more sophisticated triggered modulation in transmembrane water flux was used to generate ion-responsive contrast. Here cholesterol modified POPC liposomes internally doped with either Gd-DOTA or Gd-MSNs were generated (Duncan et al., 2024). This was followed by the dual integration of  $K^+$  and  $Cl^-$  selective ion carrier species, namely, valinomycin and a tripodal thiourea-based motifs, which could transport the desired ions across the bilayer; central here is noting that this ion transport is associated with significant associated water. Upon simultaneous transport of both cations and anions (ionic symport activation) the carrier-mediated

passage of both  $M^+$  and  $X^-$  across the bilayer was enabled, as displayed in Figure 16. Concurrently, ion associated water flux is greatly enhanced, elevating the cross-membrane exchange rate, allowing notable modulations in MRI contrast to be observed in  $T_1$  weighted MRI maps (with  $r_1$  increasing by up to 200% dependent on the nature of the integrated paramagnet). Such variations in  $r_1$  were found to correlate with the selectivity of the cationophore and anionophore present and support, of course, an engineered ion responsive contrast generation.

Stimuli responsive liposomal MRI CAs can be further developed to incorporate dual-modal ( $T_1$ - $T_2$ ) MRI probes. An example of this was demonstrated by Zhou et al., who investigated the NIR and  $H_2O_2$  activated oxidation of ferrocenyl (Fc-) compounds, specifically the oxidation of Fc (hydrophobic) to  $Fc^+$  (hydrophilic) within ferrocenylseleno-modified  $Gd^{3+}$ -doped liposomes through Fenton chemistry (Zhou et al., 2023). The associated transition from hydrophobic to hydrophilic (as  $Fc^+$ ) was proposed to improve local hydration and facilitate water exchange at both the  $Gd^{3+}$  and  $Fe^{3+}$  centres, switching both  $r_1$  and  $r_2$  with  $\Delta r_1 = 0.52 \text{ mM}^{-1} \text{ s}^{-1}$  and  $\Delta r_2 = 1.28 \text{ mM}^{-1} \text{ s}^{-1}$  recorded at 0.5 T.

Liposomal MRI CAs represent, then, an attractive organic scaffold where generated MR contrast can be specifically modulated by a programmable and wide variety of stimuli. Responsivity in these frameworks may be achieved both by non-reversible (i.e., membrane lysis or peripheral cleavages of MRI active moieties) or reversible (water gating) means. Furthermore, such responsive probes have highly developed theranostic applications

(e.g., integration of the anti-cancer agent doxorubicin) in addition to a diagnostic approach ( $r_1$  and/or  $r_2$  active payloads) (Pitchaimani et al., 2016; Karimi et al., 2016). Given their biomimetic nature, and therefore high associated biocompatibilities, one can foresee these as emerging and potent theranostic agents.

## 4 Outlook and future work

Recent developments in nanoparticulate organic chemistry can be leveraged in the generation of contrast supporting agents that are highly responsive in specific physiologically-relevant environments.

Through all of these approaches, which employ organic/organically-coated nanoparticles, including those which are liposomal, micellar, dendritic and inorganic-polymer hybrid in nature, the importance in considering water access, exchange rate, and rotational correlation when designing a particular CA is emphasised. These platforms maintain the typical benefits of nanoparticulate CAs such as high tunability, morphological control, and the possibility of internal cargo loading. Additionally, as is generally true for nanoparticle-based MRI CAs, baseline relaxivity/generated contrast is greatly increased due to beneficially elongated rotational correlation characteristics that result in the tumbling of the paramagnetic-chelate becoming slowed and closer to the optimal Larmor frequency (as demonstrated by SBM theory). Inorganic-polymer hybrids include polymer amended IONPs and MSNs that can control associated image contrast through a considered manipulation of these same governing SBM parameters. Strategies include the irreversible aggregation/disaggregation of IONPs through control of the organic coating chemistry that can dramatically influence the magnetic susceptibility of the imaging probe, resulting in a significant switch in  $r_2$ . Alternatively, the use of surface-polymer bound MSNs can be employed to controllably gate the release of an integrated MR-active cargo, restoring initially highly restricted (necessary) water access. This can be either through irreversible loss of electrostatic association to the particle surface as polymer  $pK_a$  is traversed, or by reversible extension of individual polymers into solution through repulsion of adjacent charged strands (beneficially elongating  $\tau_D/\tau_m$ ).

Similar approaches are also used for the generation of responsive “purely organic” agents; the irreversible fragmentation of micellular and liposomal contrast agents under local stimulus is a common method to release core confined integrated MRI-active agents and restore water access, for example,. Here, the fragmentation is often caused by either a change in the ionisation state of the organic component, mediated through solution pH and  $pK_a$ , or through reduction of cleavable bridges that include disulfide links. Alternatively, as is the case for polymer micelles, rotational characteristics of polymer-tethered paramagnetic chelates can be controllably, and often reversibly, altered through a modulation of polymer conformation. Specifically, extension of individual strands into solution, as  $pK_a$  is traversed or disulfide bonds are cleaved, can result in a switch in relaxivity as mechanical coupling of the chelate to the nanoparticle core is lost and  $\tau_R$  becomes dictated by rotation

of the chelate. Liposomal agents have a long and potent history as therapeutic delivery systems but can also be powerfully integrated into environmentally-responsive contrast agents. With these water flux from bulk can be influenced by virtue of the (programmable) lipid bilayer which can provide a barrier to water exchange on rigidification (e.g., by cholesterol doping), restorable by melting, distortion or through the integration of appropriate transporters (e.g., solvated ion transporters).

To summarise, the marriage of inorganic magnetochemistry with organic functionality supports a colourful range of responsive CAs. These already integrate a diverse array of mechanistic approaches to switch  $T_1$  or  $T_2$  image contrast on exposure to a range of external environmental stimuli. Continued developments in this field look to generate highly biocompatible configurations supporting a significant switch ‘on’ in relaxivity under conditions which align with physiologically relevant microenvironments and associated diagnostic need. Going forwards, one can foresee that this combination of broad chemical know-how represents a potent means of enabling the high contrast imaging of specific pathology, potentially with integrated therapy.

## Author contributions

AD: Writing – review and editing. CE: Writing – review and editing. JS: Writing – review and editing. LL: Writing – review and editing. MJL: Writing – review and editing. JD: Conceptualization, Writing – original draft, Writing – review and editing.

## Funding

The author(s) declare that financial support was received for the research and/or publication of this article. MJL is a Royal Society University Research Fellow.

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

## References

- Abramovitch, R., Marikovsky, M., Meir, G., and Neeman, M. (1998). Stimulation of tumour angiogenesis by proximal wounds: spatial and temporal analysis by MRI. *Br. J. Cancer* 77 (3), 440–447. doi:10.1038/bjc.1998.70
- Accardo, A., Tesauro, D., Aloj, L., Pedone, C., and Morelli, G. (2009). Supramolecular aggregates containing lipophilic Gd(III) complexes as contrast agents in MRI. *Coord. Chem. Rev.* 253 (17–18), 2193–2213. doi:10.1016/j.ccr.2009.01.015
- Ahl, P. L., Bhatia, S. K., Meers, P., Roberts, P., Stevens, R., Dause, R., et al. (1997). Enhancement of the *in vivo* circulation lifetime of 1- $\alpha$ -distearoylphosphatidylcholine liposomes: importance of liposomal aggregation *versus* complement opsonization. *Biochimica Biophysica Acta (BBA) - Biomembr.* 1329 (2), 370–382. doi:10.1016/s0005-2736(97)00129-6
- Aime, S., Castelli, D. D., Cich, S. G., Gianolio, E., and Terreno, E. (2009). Pushing the sensitivity envelope of lanthanide-based magnetic resonance imaging (MRI) contrast agents for molecular imaging applications. *Accounts Chem. Res.* 42 (7), 822–831. doi:10.1021/ar800192p
- Alawak, M., Mahmoud, G., Dayyih, A. A., Duse, L., Pinnapireddy, S. R., Engelhardt, K., et al. (2020). Magnetic resonance activatable thermosensitive liposomes for controlled doxorubicin delivery. *Mater. Sci. Eng. C* 115, 111116. doi:10.1016/j.msec.2020.111116
- Alghamdi, S. A. (2023). Gadolinium-based contrast agents in pregnant women: a literature review of MRI safety. *Cureus* 15, e38493. doi:10.7759/cureus.38493
- Alvares, R. D. A., Szulc, D. A., and Cheng, H.-L. M. (2017). A scale to measure MRI contrast agent sensitivity. *Sci. Rep.* 7 (1), 15493. doi:10.1038/s41598-017-15732-8
- Arosio, P., Thévenot, J., Orlando, T., Orsini, F., Corti, M., Mariani, M., et al. (2013). Hybrid iron oxide-copolymer micelles and vesicles as contrast agents for MRI: impact of the nanostructure on the relaxometric properties. *J. Mater. Chem. B* 1 (39), 5317. doi:10.1039/c3tb00429e
- Ayyagari, A. L., Zhang, X., Ghaghada, K. B., Annapragada, A., Hu, X., and Bellamkonda, R. V. (2006). Long-circulating liposomal contrast agents for magnetic resonance imaging. *Magnetic Reson. Med.* 55 (5), 1023–1029. doi:10.1002/mrm.20846
- Bain, A. D. (1990). The choice of parameters in an NMR experiment. Application to the inversion-recovery T1 method. *J. Magnetic Reson.* 89 (1), 153–160. doi:10.1016/0022-2364(90)90170-E
- Bao, Y., Sherwood, J. A., and Sun, Z. (2018). Magnetic iron oxide nanoparticles as T1 contrast agents for magnetic resonance imaging. *J. Mater. Chem. C* 6 (6), 1280–1290. doi:10.1039/C7TC05854C
- Benezra, M., Penate-Medina, O., Zanzonico, P. B., Schaer, D., Ow, H., Burns, A., et al. (2011). Multimodal silica nanoparticles are effective cancer-targeted probes in a model of human melanoma. *J. Clin. Investigation* 121 (7), 2768–2780. doi:10.1172/jci45600
- Berger, A. (2002). Magnetic resonance imaging. *BMJ Clin. Res. ed.* 324 (7328), 35. doi:10.1136/bmj.324.7328.35
- Bley, T. A., Wieben, O., François, C. J., Brittain, J. H., and Reeder, S. B. (2010). Fat and water magnetic resonance imaging. *J. Magnetic Reson. Imaging* 31 (1), 4–18. doi:10.1002/jmri.21895
- Blomqvist, L., Nordberg, G. F., Nurchi, V. M., and Aaseth, J. O. (2022). Gadolinium in medical imaging—usefulness, toxic reactions and possible countermeasures—A review. *Biomolecules* 12 (6), 742. doi:10.3390/biom12060742
- Botta, M., and Tei, L. (2012). Relaxivity enhancement in macromolecular and nanosized Gd<sup>III</sup>-based MRI contrast agents. *Eur. J. Inorg. Chem.* 2012 (12), 1945–1960. doi:10.1002/ejic.201101305
- Bousquet, J. C., Saini, S., Stark, D. D., Hahn, P. F., Nigam, M., Wittenberg, J., et al. (1988). Gd-DOTA: characterization of a new paramagnetic complex. *Radiology* 166 (3), 693–698. doi:10.1148/radiology.166.3.3340763
- Brindle, K. (2008). New approaches for imaging tumour responses to treatment. *Nat. Rev. Cancer* 8 (2), 94–107. doi:10.1038/nrc2289
- Bunzen, H., and Jiráček, D. (2022). Recent advances in metal-organic frameworks for applications in magnetic resonance imaging. *ACS Appl. Mater. & Interfaces* 14 (45), 50445–50462. doi:10.1021/acsami.2c10272
- Busse, H., Kahn, T., and Moche, M. (2018). Techniques for interventional MRI guidance in closed-bore systems. *Top. Magnetic Reson. Imaging* 27 (1), 9–18. doi:10.1097/rmr.0000000000000150
- Cao, Y., Liu, M., Zhang, K., Zu, G., Kuang, Y., Tong, X., et al. (2017). Poly(glycerol) used for constructing mixed polymeric micelles as T1 MRI contrast agent for tumor-targeted imaging. *Biomacromolecules* 18 (1), 150–158. doi:10.1021/acs.biomac.6b01437
- Cardellini, J., Surpi, A., Muzzi, B., Pacciani, V., Innocenti, C., Sangregorio, C., et al. (2024). Magnetic-plasmonic nanoscale liposomes with tunable optical and magnetic properties for combined multimodal imaging and drug delivery. *ACS Appl. Nano Mater.* 7 (4), 3668–3678. doi:10.1021/acsanm.3c05100
- Carniato, F., Tei, L., and Botta, M. (2018). Gd-based mesoporous silica nanoparticles as MRI probes. *Eur. J. Inorg. Chem.* 2018 (46), 4936–4954. doi:10.1002/ejic.201801039
- Carniato, F., Tei, L., Cossi, M., Marchese, L., and Botta, M. (2010). A chemical strategy for the relaxivity enhancement of Gd(III) chelates anchored on mesoporous silica nanoparticles. *Chem. - A Eur. J.* 16 (35), 10727–10734. doi:10.1002/chem.201000499
- Carol, H., Fan, M. M. Y., Harasym, T. O., Boehm, I., Mayer, L. D., Houghton, P., et al. (2015). Efficacy of CPX-351, (cytarabine:daunorubicin) liposome injection, against acute lymphoblastic leukemia (ALL) xenograft models of the pediatric preclinical testing program. *Pediatr. Blood & Cancer* 62 (1), 65–71. doi:10.1002/pbc.25133
- Castelli, D. D., Terreno, E., Cabella, C., Chaabane, L., Lanzardo, S., Tei, L., et al. (2009). Evidence for *in vivo* macrophage mediated tumor uptake of paramagnetic/fluorescent liposomes. *NMR Biomed.* 22 (10), 1084–1092. doi:10.1002/nbm.1416
- Cazares-Cortes, E., Espinosa, A., Guigner, J.-M., Michel, A., Griffete, N., Wilhelm, C., et al. (2017). Doxorubicin intracellular remote release from biocompatible oligo(ethylene glycol) methyl ether methacrylate-based magnetic nanogels triggered by magnetic hyperthermia. *ACS Appl. Mater. & Interfaces* 9 (31), 25775–25788. doi:10.1021/acsami.7b06553
- Cheah, P., Brown, P., Qu, J., Tian, B., Patton, D. L., and Zhao, Y. (2021). Versatile surface functionalization of water-dispersible iron oxide nanoparticles with precisely controlled sizes. *Langmuir* 37 (3), 1279–1287. doi:10.1021/acs.langmuir.0c03314
- Chen, P.-S., Chiu, W.-T., Hsu, P.-L., Lin, S.-C., Peng, I. C., Wang, C.-Y., et al. (2020). Pathophysiological implications of hypoxia in human diseases. *J. Biomed. Sci.* 27 (1), 63. doi:10.1186/s12929-020-00658-7
- Chowdhury, M. S. I., Kras, E. A., Turowski, S. G., Spornyak, J. A., and Morrow, J. R. (2023). Liposomal MRI probes containing encapsulated or amphiphilic Fe(III) coordination complexes. *Biomaterials Sci.* 11 (17), 5942–5954. doi:10.1039/D3BM00029J
- Cormode, D. P., Skajaa, T., Fayad, Z. A., and Mulder, W. J. (2009). Nanotechnology in medical imaging: probe design and applications. *Arterioscler. Thromb. Vasc. Biol.* 29 (7), 992–1000. doi:10.1161/atvbaha.108.165506
- Crommelin, D. J., and Florence, A. T. (2013). Towards more effective advanced drug delivery systems. *Int. J. Pharm.* 454 (1), 496–511. doi:10.1016/j.ijpharm.2013.02.020
- Crosby, D., Bhatia, S., Brindle, K. M., Coussens, L. M., Dive, C., Emberton, M., et al. (2022). Early detection of cancer. *Science* 375 (6586), eaay9040. doi:10.1126/science.aay9040
- Dadfar, S. M., Roemhild, K., Drude, N. I., von Stillfried, S., Knüchel, R., Kiessling, F., et al. (2019). Iron oxide nanoparticles: diagnostic, therapeutic and theranostic applications. *Adv. Drug Deliv. Rev.* 138, 302–325. doi:10.1016/j.addr.2019.01.005
- Davies, G.-L., Kramberger, I., and Davis, J. J. (2013). Environmentally responsive MRI contrast agents. *Chem. Commun.* 49 (84), 9704. doi:10.1039/c3cc44268c
- De León-Rodríguez, L. M., Martins, A. F., Pinho, M. C., Rofsky, N. M., and Sherry, A. D. (2015). Basic MR relaxation mechanisms and contrast agent design. *J. Magn. Reson. Imaging* 42 (3), 545–565. doi:10.1002/jmri.24787
- Deng, T., Zhang, L., Li, X., Zink, J. I., and Wu, H. H. (2021). Responsive nanoparticles to enable a focused ultrasound-stimulated magnetic resonance imaging spotlight. *ACS Nano* 15 (9), 14618–14630. doi:10.1021/acsnano.1c04339
- De Sarno, F., Ponsiglione, A. M., Russo, M., Grimaldi, A. M., Forte, E., Netti, P. A., et al. (2019). Water-mediated nanostructures for enhanced MRI: impact of water dynamics on relaxometric properties of Gd-DTPA. *Theranostics* 9 (6), 1809–1824. doi:10.7150/thno.27313
- Deulkar, P., Singam, A., and Jain, A. (2024). A comprehensive review of the role of biomarkers in the early detection of endocrine disorders in critical illnesses. *Cureus* 16 (5), e61409. doi:10.7759/cureus.61409
- Devreux, M., Henoumont, C., Dioury, F., Boutry, S., Vacher, O., Elst, L. V., et al. (2021). Mn<sup>2+</sup> complexes with Pyclyen-Based derivatives as contrast agents for magnetic resonance imaging: synthesis and relaxometry characterization. *Inorg. Chem.* 60 (6), 3604–3619. doi:10.1021/acs.inorgchem.0c03120
- Ding, Y., Dai, Y., Wu, M., and Li, L. (2021). Glutathione-mediated nanomedicines for cancer diagnosis and therapy. *Chem. Eng. J.* 426, 128880. doi:10.1016/j.cej.2021.128880
- Dražoš, B., Lukeš, I., and Tóth, É. (2012). Manganese(II) complexes as potential contrast agents for MRI. *Eur. J. Inorg. Chem.* 2012 (12), 1975–1986. doi:10.1002/ejic.201101336
- Dulińska-Litewka, J., Łazarczyk, A., Hałubiec, P., Szafranski, O., Karnas, K., and Karczewicz, A. (2019). Superparamagnetic iron oxide nanoparticles-current and prospective medical applications. *Mater. (Basel)* 12 (4), 617. doi:10.3390/ma12040617
- Duncan, A. M., Ellis, C. M., Levingston, H., Kerckhoffs, A., Mózes, F. E., Langton, M. J., et al. (2024). Ion carrier modulated MRI contrast. *Chem. Sci.* 15 (34), 13937–13941. doi:10.1039/D3SC03466F
- Edelstein, W. A., Bottomley, P. A., and Pfeifer, L. M. (1984). A signal-to-noise calibration procedure for NMR imaging systems: signal-To-Noise calibration for NMR imaging systems. *Med. Phys.* 11 (2), 180–185. doi:10.1118/1.595484
- Ellis, C. M., Yuan, D., Mózes, F. E., Miller, J. J., and Davis, J. J. (2023). Reversible pH-responsive MRI contrast with paramagnetic polymer micelles. *Chem. Commun.* 59 (12), 1605–1608. doi:10.1039/D2CC06255K
- Elsabahy, M., Heo, G. S., Lim, S.-M., Sun, G., and Wooley, K. L. (2015). Polymeric nanostructures for imaging and therapy. *Chem. Rev.* 115 (19), 10967–11011. doi:10.1021/acs.chemrev.5b00135



- Ersoy, H., and Rybicki, F. J. (2007). Biochemical safety profiles of gadolinium-based extracellular contrast agents and nephrogenic systemic fibrosis. *J. Magnetic Reson. Imaging* 26 (5), 1190–1197. doi:10.1002/jmri.21135
- Estelrich, J., Sánchez-Martín, M. J., and Busquets, M. A. (2015). Nanoparticles in magnetic resonance imaging: from simple to dual contrast agents. *Int. J. Nanomedicine* 10, 1727–1741. doi:10.2147/ijn.S76501
- Feng, Q., Liu, Y., Huang, J., Chen, K., Huang, J., and Xiao, K. (2018). Uptake, distribution, clearance, and toxicity of iron oxide nanoparticles with different sizes and coatings. *Sci. Rep.* 8 (1), 2082. doi:10.1038/s41598-018-19628-z
- Fossheim, S. L., Fahlvik, A. K., Klaveness, J., and Muller, R. N. (1999). Paramagnetic liposomes as MRI contrast agents: influence of liposomal physicochemical properties on the *in vitro* relaxivity. *Magn. Reson. Imaging* 17 (1), 83–89. doi:10.1016/S0730-725X(98)00141-6
- Foster, D., and Larsen, J. (2023). Polymeric metal contrast agents for T(1)-Weighted magnetic resonance imaging of the brain. *ACS Biomater. Sci. Eng.* 9 (3), 1224–1242. doi:10.1021/acsbomaterials.2c01386
- Gaharwar, U. S., Meena, R., and Rajamani, P. (2019). Clearance and morphological alterations of intravenously administered iron oxide nanoparticles in Male wistar rats. *Int. J. Nanomedicine* 14, 9677–9692. doi:10.2147/IJN.S223142
- Gao, G. H., Lee, J. W., Nguyen, M. K., Im, G. H., Yang, J., Heo, H., et al. (2011). pH-responsive polymeric micelle based on PEG-poly( $\beta$ -amino ester)/(amido amine) as intelligent vehicle for magnetic resonance imaging in detection of cerebral ischemic area. *J. Control. Release* 155 (1), 11–17. doi:10.1016/j.jconrel.2010.09.012
- Ge, M., Papagiannakopoulos, T., and Bar-Peled, L. (2024). Reductive stress in cancer: coming out of the shadows. *Trends Cancer* 10 (2), 103–112. doi:10.1016/j.trecan.2023.10.002
- Ge, Z., and Liu, S. (2013). Functional block copolymer assemblies responsive to tumor and intracellular microenvironments for site-specific drug delivery and enhanced imaging performance. *Chem. Soc. Rev.* 42 (17), 7289. doi:10.1039/c3cs60048c
- Geraldes, C. F. G. C., and Laurent, S. (2009). Classification and basic properties of contrast agents for magnetic resonance imaging. *Contrast Media & Mol. Imaging* 4 (1), 1–23. doi:10.1002/cmmi.265
- Grogna, M., Cloots, R., Luxen, A., Jérôme, C., Passirani, C., Lautram, N., et al. (2010). Polymer micelles decorated by gadolinium complexes as MRI blood contrast agents: design, synthesis and properties. *Polym. Chem.* 1 (9), 1485–1490. doi:10.1039/C0PY00117A
- Guo, Z., Sui, J., Ma, M., Hu, J., Sun, Y., Yang, L., et al. (2020). pH-Responsive charge switchable PEGylated  $\epsilon$ -poly-L-lysine polymeric nanoparticles-assisted combination therapy for improving breast cancer treatment. *J. Control Release* 326, 350–364. doi:10.1016/j.jconrel.2020.07.030
- Hagberg, G. E., and Scheffler, K. (2013). Effect of  $r_1$  and  $r_2$  relaxivity of gadolinium-based contrast agents on the T<sub>1</sub>-weighted MR signal at increasing magnetic field strengths. *Contrast Media Mol. Imaging* 8 (6), 456–465. doi:10.1002/cmmi.1565
- Hao, D., Ai, T., Goerner, F., Hu, X., Runge, V. M., and Tweedle, M. (2012). MRI contrast agents: basic chemistry and safety. *J. Magnetic Reson. Imaging* 36 (5), 1060–1071. doi:10.1002/jmri.23725
- Hao, G., Xu, Z. P., and Li, L. (2018). Manipulating extracellular tumour pH: an effective target for cancer therapy. *RSC Adv.* 8 (39), 22182–22192. doi:10.1039/C8RA02095G
- Hawkes, N. (2019). Cancer survival data emphasise importance of early diagnosis. *BMJ* 364, 1408. doi:10.1136/bmj.1408
- He, K., Li, J., Shen, Y., and Yu, Y. (2019). pH-Responsive polyelectrolyte coated gadolinium oxide-doped mesoporous silica nanoparticles (Gd<sub>2</sub>O<sub>3</sub>@MSNs) for synergistic drug delivery and magnetic resonance imaging enhancement. *J. Mater. Chem. B* 7 (43), 6840–6854. doi:10.1039/C9TB01654F
- Hingorani, D. V., Bernstein, A. S., and Pagel, M. D. (2015). A review of responsive MRI contrast agents: 2005–2014. *Contrast Media & Mol. Imaging* 10 (4), 245–265. doi:10.1002/cmmi.1629
- Hsu, J. C., Tang, Z., Eremina, O. E., Sofias, A. M., Lammers, T., Lovell, J. F., et al. (2023). Nanomaterial-based contrast agents. *Nat. Rev. Methods Prim.* 3 (1), 30. doi:10.1038/s43586-023-00211-4
- Ilyad, N., Ahmad, M. S., Alkhatib, S. G., and Hjouj, M. (2023). Gadolinium contrast agents-challenges and opportunities of a multidisciplinary approach: literature review. *Eur. J. Radiology Open* 11, 100503. doi:10.1016/j.ejro.2023.100503
- Jackson, C. E., Moseley, I. P., Martinez, R., Sung, S., and Zadrozny, J. M. (2021). A reaction-coordinate perspective of magnetic relaxation. *Chem. Soc. Rev.* 50 (12), 6684–6699. doi:10.1039/D1CS00001B
- Jacques, V., Dumas, S., Sun, W.-C., Troughton, J. S., Greenfield, M. T., and Caravan, P. (2010). High-relaxivity magnetic resonance imaging contrast agents part 2: optimization of inner- and second-sphere relaxivity. *Investig. Radiol.* 45 (10), 613–624. doi:10.1097/RLI.0b013e3181ee6a49
- Jeon, M., Halbert, M. V., Stephen, Z. R., and Zhang, M. (2021). Iron oxide nanoparticles as T<sub>1</sub> contrast agents for magnetic resonance imaging: fundamentals, challenges, applications, and perspectives. *Adv. Mater.* 33 (23), 1906539. doi:10.1002/adma.201906539
- Ju, S.-P., Lee, W.-J., Huang, C.-I., Cheng, W.-Z., and Chung, Y.-T. (2007). Structure and dynamics of water surrounding the poly(methacrylic acid): a molecular dynamics study. *J. Chem. Phys.* 126 (22), 224901. doi:10.1063/1.2743963
- Jung, B. A., and Weigel, M. (2013). Spin echo magnetic resonance imaging. *J. Magnetic Reson. Imaging* 37 (4), 805–817. doi:10.1002/jmri.24068
- Karimi, M., Ghasemi, A., Sahandi Zangabad, P., Rahighi, R., Moosavi Basri, S. M., Mirshekari, H., et al. (2016). Smart micro/nanoparticles in stimulus-responsive drug/gene delivery systems. *Chem. Soc. Rev.* 45 (5), 1457–1501. doi:10.1039/C5CS00798D
- Kawahara, D., and Nagata, Y. (2021). T1-weighted and T2-weighted MRI image synthesis with convolutional generative adversarial networks. *Rep. Pract. Oncol. Radiotherapy J. Gt. Cancer Cent. Poznan Pol. Soc. Radiat. Oncol.* 26 (1), 35–42. doi:10.5603/RPOR.a2021.0005
- Kazunori, K., Glenn S. K., Masayuki, Y., Teruo, O., and Yasuhisa, S. (1993). Block copolymer micelles as vehicles for drug delivery. *J. Control. Release* 24 (1), 119–132. doi:10.1016/0168-3659(93)90172-2
- Kim, H. R., You, D. G., Park, S.-J., Choi, K.-S., Um, W., Kim, J.-H., et al. (2016). MRI monitoring of tumor-selective anticancer drug delivery with stable thermosensitive liposomes triggered by high-intensity focused ultrasound. *Mol. Pharm.* 13 (5), 1528–1539. doi:10.1021/acs.molpharmaceut.6b00013
- Kim, K. S., Park, W., Hu, J., Bae, Y. H., and Na, K. (2014). A cancer-recognizable MRI contrast agents using pH-responsive polymeric micelle. *Biomaterials* 35 (1), 337–343. doi:10.1016/j.biomaterials.2013.10.004
- Kim, R. J., Wu, E., Rafael, A., Chen, E.-L., Parker, M. A., Simonetti, O., et al. (2000). The use of contrast-enhanced magnetic resonance imaging to identify reversible myocardial dysfunction. *N. Engl. J. Med.* 343 (20), 1445–1453. doi:10.1056/NEJM200011163432003
- Kingsley, P. B. (1999). Methods of measuring spin-lattice (T<sub>1</sub>) relaxation times: an annotated bibliography. *Concepts Magnetic Reson.* 11 (4), 243–276. doi:10.1002/(sici)1099-0534(1999)11:4<243::aid-cmr5>3.3.co;2-3
- Kirchin, M. A., and Runge, V. M. (2003). Contrast agents for magnetic resonance imaging: safety update. *Top. magnetic Reson. imaging TMRI* 14 (5), 426–435. doi:10.1097/00002142-200310000-00007
- Kono, K., Nakashima, S., Kokuryo, D., Aoki, I., Shimomoto, H., Aoshima, S., et al. (2011). Multi-functional liposomes having temperature-triggered release and magnetic resonance imaging for tumor-specific chemotherapy. *Biomaterials* 32 (5), 1387–1395. doi:10.1016/j.biomaterials.2010.10.050
- Kostevšek, N. (2020). A review on the optimal design of magnetic nanoparticle-based T<sub>2</sub> MRI contrast agents. *Magnetochemistry* 6 (1), 11. doi:10.3390/magnetochemistry6010011
- Kraut, J. A., and Kurtz, I. (2005). Metabolic acidosis of CKD: diagnosis, clinical characteristics, and treatment. *Am. J. Kidney Dis.* 45 (6), 978–993. doi:10.1053/j.ajkd.2005.03.003
- Kularatne, S. A., and Low, P. S. (2010). Targeting of nanoparticles: folate receptor. *Methods Mol. Biol.* 624, 249–265. doi:10.1007/978-1-60761-609-2\_17
- Lancelot, E., Raynaud, J.-S., and Desché, P. (2020). Current and future MR contrast agents: seeking a better chemical stability and relaxivity for optimal safety and efficacy. *Investig. Radiol.* 55 (9), 578–588. doi:10.1097/RLI.0000000000000684
- Langereis, S., Geelen, T., Grüll, H., Strijkers, G. J., and Nicolay, K. (2013). Paramagnetic liposomes for molecular MRI and MRI-guided drug delivery. *NMR Biomed.* 26 (7), 728–744. doi:10.1002/nbm.2971
- Lapusan, R., Borlan, R., and Focsan, M. (2024). Advancing MRI with magnetic nanoparticles: a comprehensive review of translational research and clinical trials. *Nanoscale Adv.* 6 (9), 2234–2259. doi:10.1039/d3na01064c
- Larsen, E. K. U., Nielsen, T., Wittenborn, T., Rydtoft, L. M., Lokanathan, A. R., Hansen, L., et al. (2012). Accumulation of magnetic iron oxide nanoparticles coated with variably sized polyethylene glycol in murine tumors. *Nanoscale* 4 (7), 2352. doi:10.1039/c2nr11554a
- Lazaro-Carrillo, A., Filice, M., Guillén, M. J., Amaro, R., Viñambres, M., Tabero, A., et al. (2020). Tailor-made PEG coated iron oxide nanoparticles as contrast agents for long lasting magnetic resonance molecular imaging of solid cancers. *Mater. Sci. Eng. C* 107, 110262. doi:10.1016/j.msec.2019.110262
- Leal, M. P., Muñoz-Hernández, C., Berry, C. C., and García-Martín, M. L. (2015). *In vivo* pharmacokinetics of T<sub>2</sub> contrast agents based on iron oxide nanoparticles: optimization of blood circulation times. *RSC Adv.* 5 (94), 76883–76891. doi:10.1039/C5RA15680G
- Le Botlan, D. J., and Ouguerram, L. (1997). Spin-spin relaxation time determination of intermediate states in heterogeneous products from free induction decay NMR signals. *Anal. Chim. Acta* 349 (1–3), 339–347. doi:10.1016/S0003-2670(97)00274-2
- Lérida-Viso, A., Estepa-Fernández, A., García-Fernández, A., Martí-Centelles, V., and Martínez-Máñez, R. (2023). Biosafety of mesoporous silica nanoparticles: towards clinical translation. *Adv. Drug Deliv. Rev.* 201, 115049. doi:10.1016/j.addr.2023.115049
- Li, H., and Meade, T. J. (2019). Molecular magnetic resonance imaging with Gd(III)-Based contrast agents: challenges and key advances. *J. Am. Chem. Soc.* 141 (43), 17025–17041. doi:10.1021/jacs.9b09149



- Li, Q., Gao, W., Zhang, C., Wang, P., Wang, X., Yan, M., et al. (2022b). A biodegradable high-efficiency magnetic nanoliposome promotes tumor microenvironment-responsive multimodal tumor therapy along with switchable T<sub>2</sub> magnetic resonance imaging. *ACS Appl. Mater. & Interfaces* 14 (21), 24160–24173. doi:10.1021/acsami.2c04158
- Li, Y., Beija, M., Laurent, S., Elst, L. v., Muller, R. N., Duong, H. T. T., et al. (2012). Macromolecular ligands for gadolinium MRI contrast agents. *Macromolecules* 45 (10), 4196–4204. doi:10.1021/ma300521c
- Li, Z., Guo, J., Qi, G., Zhang, M., and Hao, L. (2022a). pH-Responsive drug delivery and imaging study of hybrid mesoporous silica nanoparticles. *Molecules* 27, 6519. doi:10.3390/molecules27196519
- Lin, S. P., and Brown, J. J. (2007). MR contrast agents: physical and pharmacologic basics. *J. Magnetic Reson. Imaging* 25 (5), 884–899. doi:10.1002/jmri.20955
- Liu, C., Chen, H., Zhou, H., Yu, S., Zhao, Y., Wang, N., et al. (2022). MRI-FI-guided superimposed stimulus-responsive co-assembled liposomes for optimizing transmembrane drug delivery pathways and improving cancer efficacy. *Appl. Mater. Today* 26, 101368. doi:10.1016/j.apmt.2022.101368
- Liu, C., Ewert, K. K., Wang, N., Li, Y., Safinya, C. R., and Qiao, W. (2019b). A multifunctional lipid that forms contrast-agent liposomes with dual-control release capabilities for precise MRI-Guided drug delivery. *Biomaterials* 221, 119412. doi:10.1016/j.biomaterials.2019.119412
- Liu, T.-P., Wu, S.-H., Chen, Y.-P., Chou, C.-M., and Chen, C.-T. (2015). Biosafety evaluations of well-dispersed mesoporous silica nanoparticles: towards in vivo-relevant conditions. *Nanoscale* 7 (15), 6471–6480. doi:10.1039/c4nr07421a
- Liu, Y., Bhattarai, P., Dai, Z., and Chen, X. (2019a). Photothermal therapy and photoacoustic imaging via nanotheranostics in fighting cancer. *Chem. Soc. Rev.* 48 (7), 2053–2108. doi:10.1039/C8CS00618K
- Løkling, K.-E., Fossheim, S. L., Skurtveit, R., Bjørnerud, A., and Klaveness, J. (2001). pH-sensitive paramagnetic liposomes as MRI contrast agents: *in vitro* feasibility studies. *Magn. Reson. Imaging* 19 (5), 731–738. doi:10.1016/S0730-725X(01)00380-0
- Lowe, S., O'Brien-Simpson, N. M., and Connal, L. A. (2015). Antibiofouling polymer interfaces: poly(ethylene glycol) and other promising candidates. *Polym. Chem.* 6 (2), 198–212. doi:10.1039/c4py01356e
- Lu, Z., Yan, J., Xu, M., Sun, L., Liu, J., Zhang, Y., et al. (2023). Hypoxia-responsive aggregation of iron oxide nanoparticles for T1-to-T2 switchable magnetic resonance imaging of tumors. *ACS Appl. Nano Mater.* 6 (1), 119–130. doi:10.1021/acsanm.2c03850
- Lux, J., and Sherry, A. D. (2018). Advances in gadolinium-based MRI contrast agent designs for monitoring biological processes *in vivo*. *Curr. Opin. Chem. Biol.* 45, 121–130. doi:10.1016/j.cbpa.2018.04.006
- Manavalan, R. K., Enoch, K., Volegov, A. S., Angusamy, G., and Nallasivam, S. (2024). Review on medical applications of manganese oxide (mn<sup>2+</sup>, Mn<sup>3+</sup>, and Mn<sup>4+</sup>) magnetic nanoparticles. *J. Nanomater.* 2024, 1–29. doi:10.1155/2024/1073915
- Matsumoto, Y., and Jasanoff, A. (2008). T2 relaxation induced by clusters of superparamagnetic nanoparticles: Monte carlo simulations. *Magn. Reson. Imaging* 26 (7), 994–998. doi:10.1016/j.mri.2008.01.039
- Modic, M. T., Pflanze, W., Feiglin, D. H. I., and Belhobek, G. (1986). Magnetic resonance imaging of musculoskeletal infections. *Radiologic Clin. N. Am.* 24 (2), 247–258. doi:10.1016/S0033-8389(22)01074-0
- Mohamed Isa, E. D., Ahmad, H., Abdul Rahman, M. B., and Gill, M. R. (2021). Progress in mesoporous silica nanoparticles as drug delivery agents for cancer treatment. *Pharmaceutics* 13 (2), 152. doi:10.3390/pharmaceutics13020152
- Morcós, S. K. (2008). Extracellular gadolinium contrast agents: differences in stability. *Eur. J. Radiology* 66 (2), 175–179. doi:10.1016/j.ejrad.2008.01.025
- Mulder, W. J. M., Strijkers, G. J., Van Tilborg, G. A. F., Griffioen, A. W., and Nicolay, K. (2006). Lipid-based nanoparticles for contrast-enhanced MRI and molecular imaging. *NMR Biomed.* 19 (1), 142–164. doi:10.1002/nbm.1011
- Mura, S., and Couvreur, P. (2012). Nanotheranostics for personalized medicine. *Adv. Drug Deliv. Rev.* 64 (13), 1394–1416. doi:10.1016/j.addr.2012.06.006
- Mura, S., Nicolas, J., and Couvreur, P. (2013). Stimuli-responsive nanocarriers for drug delivery. *Nat. Mater.* 12 (11), 991–1003. doi:10.1038/nmat3776
- Na, H. B., and Hyeon, T. (2009). Nanostructured T1 MRI contrast agents. *J. Mater. Chem.* 19 (35), 6267. doi:10.1039/b902685a
- Naghavi, M., John, R., Naguib, S., Siadat, M. S., Grasu, R., Kurian, K. C., et al. (2002). pH heterogeneity of human and rabbit atherosclerotic plaques; a new insight into detection of vulnerable plaque. *Atherosclerosis* 164 (1), 27–35. doi:10.1016/S0021-9150(02)00018-7
- Nasongkla, N., Bey, E., Ren, J., Ai, H., Khemtong, C., Guthi, J. S., et al. (2006). Multifunctional polymeric micelles as cancer-targeted, MRI-ultrasensitive drug delivery systems. *Nano Lett.* 6 (11), 2427–2430. doi:10.1021/nl061412u
- Nedyalkova, M., Donkova, B., Romanova, J., Tzvetkov, G., Madurga, S., and Simeonov, V. (2017). Iron oxide nanoparticles – *in Vivo/In vitro* biomedical applications and *in silico* studies. *Adv. Colloid Interface Sci.* 249, 192–212. doi:10.1016/j.cis.2017.05.003
- Neves, A. A., and Brindle, K. M. (2006). Assessing responses to cancer therapy using molecular imaging. *Biochimica Biophysica Acta (BBA) - Rev. Cancer* 1766 (2), 242–261. doi:10.1016/j.bbcan.2006.10.002
- Normann, P. T., Frøysa, A., and Svaland, M. (1995). Interference of gadodiamide injection (OMNISCAN®) on the colorimetric determination of serum calcium. *Scand. J. Clin. Laboratory Investigation* 55 (5), 421–426. doi:10.3109/00365519509104981
- Nsairat, H., Khater, D., Sayed, U., Odeh, F., Al Bawab, A., and Alshaer, W. (2022). Liposomes: structure, composition, types, and clinical applications. *Heliyon* 8 (5), e09394. doi:10.1016/j.heliyon.2022.e09394
- Paliwal, S. R., Paliwal, R., and Vyas, S. P. (2015). A review of mechanistic insight and application of pH-sensitive liposomes in drug delivery. *Drug Deliv.* 22 (3), 231–242. doi:10.3109/10717544.2014.882469
- Parizh, M., Lvovsky, Y., and Sumption, M. (2017). Conductors for commercial MRI magnets beyond NbTi: requirements and challenges. *Supercond. Sci. Technol.* 30 (1), 014007. doi:10.1088/0953-2048/30/1/014007
- Pellico, J., Ellis, C. M., and Davis, J. J. (2019a). Nanoparticle-based paramagnetic contrast agents for magnetic resonance imaging. *Contrast Media & Mol. Imaging* 2019, 1–13. doi:10.1155/2019/1845637
- Pellico, J., Ellis, C. M., Miller, J., and Davis, J. J. (2019b). Water gated contrast switching with polymer-silica hybrid nanoparticles. *Chem. Commun.* 55 (59), 8540–8543. doi:10.1039/C9CC03312B
- Pellico, J., Ruiz-Cabello, J., and Herranz, F. (2023). Radiolabeled iron oxide nanomaterials for multimodal nuclear imaging and positive contrast magnetic resonance imaging (MRI): a review. *ACS Appl. Nano Mater.* 6 (22), 20523–20538. doi:10.1021/acsanm.3c04269
- Pitchaimani, A., Thanh Nguyen, T. D., Wang, H., Bossmann, S. H., and Aryal, S. (2016). Design and characterization of gadolinium infused theranostic liposomes. *RSC Adv.* 6 (43), 36898–36905. doi:10.1039/C6RA00552G
- Priya James, H., John, R., Alex, A., and Anoop, K. R. (2014). Smart polymers for the controlled delivery of drugs - a concise overview. *Acta Pharm. Sin. B* 4 (2), 120–127. doi:10.1016/j.apsb.2014.02.005
- Qi, H., Zhou, H., Tang, Q., Lee, J. Y., Fan, Z., Kim, S., et al. (2018). Block copolymer crystallites with an ultrathin shell to extend blood circulation time. *Nat. Commun.* 9 (1), 3005. doi:10.1038/s41467-018-05396-x
- Rahnfeld, L., Thamm, J., Steiniger, F., Van Hoogevest, P., and Luciani, P. (2018). Study on the *in situ* aggregation of liposomes with negatively charged phospholipids for use as injectable depot formulation. *Colloids Surfaces B Biointerfaces* 168, 10–17. doi:10.1016/j.colsurfb.2018.02.023
- Ramnicanu, G., Doan, B.-T., Vezignol, C., Graillot, A., Loubat, C., Mignet, N., et al. (2016). Delayed hepatic uptake of multi-phosphonic acid poly(ethylene glycol) coated iron oxide measured by real-time magnetic resonance imaging. *RSC Adv.* 6 (68), 63788–63800. doi:10.1039/C6RA09896G
- Reefing, F., Stuart, M. C. A., Samplonius, D. F., Dierckx, R. A. J. O., Feringa, B. L., Helfrich, W., et al. (2019). A light-responsive liposomal agent for MRI contrast enhancement and monitoring of cargo delivery. *Chem. Commun.* 55 (72), 10784–10787. doi:10.1039/C9CC05516A
- Rezaei, B., Harun, A., Wu, X., Iyer, P. R., Mostafa, S., Ciannella, S., et al. (2024). Effect of polymer and cell membrane coatings on theranostic applications of nanoparticles: a review. *Adv. Healthc. Mater.* 13, 2401213. doi:10.1002/adhm.202401213
- Rideau, E., Dimova, R., Schwill, P., Wurm, F. R., and Landfester, K. (2018). Liposomes and polymersomes: a comparative review towards cell mimicking. *Chem. Soc. Rev.* 47 (23), 8572–8610. doi:10.1039/C8CS00162F
- Rizzi, F., Castaldo, R., Latronico, T., Lasala, P., Gentile, G., Lavorgna, M., et al. (2021). High surface area mesoporous silica nanoparticles with tunable size in the sub-micrometer regime: insights on the size and porosity control mechanisms. *Molecules* 26 (14), 4247. doi:10.3390/molecules26144247
- Roca, A. G., Veintemillas-Verdaguer, S., Port, M., Robic, C., Serna, C. J., and Morales, M. P. (2009). Effect of nanoparticle and aggregate size on the relaxometric properties of MR contrast agents based on high quality magnetite nanoparticles. *J. Phys. Chem. B* 113 (19), 7033–7039. doi:10.1021/jp807820s
- Sachar, H. S., Pial, T. H., Desai, P. R., Etha, S. A., Wang, Y., Chung, P. W., et al. (2020). Densely grafted polyelectrolyte brushes trigger “water-In-salt”-Like scenarios and ultraconfinement effect. *Matter* 2 (6), 1509–1521. doi:10.1016/j.matt.2020.02.022
- Saito, H., and Shinoda, W. (2011). Cholesterol effect on water permeability through DPPC and PSM lipid bilayers: a molecular dynamics study. *J. Phys. Chem. B* 115 (51), 15241–15250. doi:10.1021/jp201611p
- Salimi, M., Sarkar, S., Saber, R., Delavari, H., Alizadeh, A. M., and Mulder, H. T. (2018). Magnetic hyperthermia of breast cancer cells and MRI relaxometry with dendrimer-coated iron-oxide nanoparticles. *Cancer Nanotechnol.* 9 (1), 7. doi:10.1186/s12645-018-0042-8
- Sarkar, D., and Somasundaran, P. (2004). Conformational dynamics of poly(acrylic acid). A study using surface plasmon resonance spectroscopy. *Langmuir* 20 (11), 4657–4664. doi:10.1021/la035727q

- Sarkis, S., Silencieux, F., Markwick, K. E., Fortin, M.-A., and Hoesli, C. A. (2017). Magnetic resonance imaging of alginate beads containing pancreatic beta cells and paramagnetic nanoparticles. *ACS Biomaterials Sci. & Eng.* 3 (12), 3576–3587. doi:10.1021/acsbomaterials.7b00404
- Sigg, S. J., Santini, F., Najer, A., Richard, P. U., Meier, W. P., and Palivan, C. G. (2016). Nanoparticle-based highly sensitive MRI contrast agents with enhanced relaxivity in reductive milieu. *Chem. Commun.* 52 (64), 9937–9940. doi:10.1039/C6CC03396B
- Simon, J., Schwalm, M., Morstein, J., Trauner, D., and Jasanoff, A. (2022). Mapping light distribution in tissue by using MRI-Detectable photosensitive liposomes. *Nat. Biomed. Eng.* 7 (3), 313–322. doi:10.1038/s41551-022-00982-3
- Smith, B. R., and Gambhir, S. S. (2017). Nanomaterials for *in vivo* imaging. *Chem. Rev.* 117 (3), 901–986. doi:10.1021/acs.chemrev.6b00073
- Soenen, S. J., Velde, G. V., Ketkar-Atre, A., Himmelreich, U., and De Cuyper, M. (2011). Magnetoliposomes as magnetic resonance imaging contrast agents. *WIREs Nanomedicine Nanobiotechnology* 3 (2), 197–211. doi:10.1002/wnan.122
- Spencer, R. G. S., and Fishbein, K. W. (2000). Measurement of spin-lattice relaxation times and concentrations in systems with chemical exchange using the one-pulse sequence: breakdown of the Ernst model for partial saturation in nuclear magnetic resonance spectroscopy. *J. Magnetic Reson.* 142 (1), 120–135. doi:10.1006/jmre.1999.1925
- Suk, J. S., Xu, Q., Kim, N., Hanes, J., and Ensign, L. M. (2016). PEGylation as a strategy for improving nanoparticle-based drug and gene delivery. *Adv. Drug Deliv. Rev.* 99 (Pt A), 28–51. doi:10.1016/j.addr.2015.09.012
- Sun, Q., You, Q., Wang, J., Liu, L., Wang, Y., Song, Y., et al. (2018). Theranostic nanoplatfrom: triple-modal imaging-guided synergistic cancer therapy based on liposome-conjugated mesoporous silica nanoparticles. *ACS Appl. Mater. & Interfaces* 10 (2), 1963–1975. doi:10.1021/acsami.7b13651
- Svenson, S. (2012). Clinical translation of nanomedicines. *Curr. Opin. Solid State Mater. Sci.* 16 (6), 287–294. doi:10.1016/j.cossms.2012.10.001
- Ta, H. T., Li, Z., Wu, Y., Cowin, G., Zhang, S., Yago, A., et al. (2017). Effects of magnetic field strength and particle aggregation on relaxivity of ultra-small dual contrast iron oxide nanoparticles. *Mater. Res. Express* 4 (11), 116105. doi:10.1088/2053-1591/aa96c3
- Taylor, K. M. L., Kim, J. S., Rieter, W. J., An, H., Lin, W., and Lin, W. (2008). Mesoporous silica nanospheres as highly efficient MRI contrast agents. *J. Am. Chem. Soc.* 130 (7), 2154–2155. doi:10.1021/ja710193c
- Thomas, R. G., Kim, S., Nagareddy, R., Vijayan, V., Pullickal, A. M., Yoon, M. S., et al. (2023). Lipo-MGN nanoparticle hypoxia attenuation-mediated single-dose radiotherapy- and pH/ROS-responsive T1 contrast magnetic resonance imaging in hepatocellular carcinoma. *Cancer Nanotechnol.* 14 (1), 53. doi:10.1186/s12645-023-00182-x
- Torres, E., Mainini, F., Napolitano, R., Fedeli, F., Cavalli, R., Aime, S., et al. (2011). Improved paramagnetic liposomes for MRI visualization of pH triggered release. *J. Control. Release* 154 (2), 196–202. doi:10.1016/j.jconrel.2011.05.017
- Vallabani, N. V. S., and Singh, S. (2018). Recent advances and future prospects of iron oxide nanoparticles in biomedicine and diagnostics. *3 Biotech.* 8 (6), 279. doi:10.1007/s13205-018-1286-z
- Verwilt, P., Park, S., Yoon, B., and Kim, J. S. (2015). Recent advances in Gd-chelate based bimodal optical/MRI contrast agents. *Chem. Soc. Rev.* 44 (7), 1791–1806. doi:10.1039/C4CS00336E
- Villaraza, A. J. L., Bumb, A., and Brechbiel, M. W. (2010). Macromolecules, dendrimers, and nanomaterials in magnetic resonance imaging: the interplay between size, function, and pharmacokinetics. *Chem. Rev.* 110 (5), 2921–2959. doi:10.1021/cr900232t
- Viswanathan, S., Kovacs, Z., Green, K. N., Ratnakar, S. J., and Sherry, A. D. (2010). Alternatives to gadolinium-based metal chelates for magnetic resonance imaging. *Chem. Rev.* 110 (5), 2960–3018. doi:10.1021/cr900284a
- Vithanarachchi, S. M., and Allen, M. J. (2012). Strategies for target-specific contrast agents for magnetic resonance imaging. *Curr. Mol. Imaging* 1 (1), 12–25. doi:10.2174/2211555211201010012
- Wahsner, J., Gale, E. M., Rodríguez-Rodríguez, A., and Caravan, P. (2019). Chemistry of MRI contrast agents: current challenges and new frontiers. *Chem. Rev.* 119 (2), 957–1057. doi:10.1021/acs.chemrev.8b00363
- Walter, A., Paul-Gilloteaux, P., Plochberger, B., Sefc, L., Verkade, P., Mannheim, J. G., et al. (2020). Correlated multimodal imaging in life sciences: expanding the biomedical horizon. *Front. Phys.* 8, 47. doi:10.3389/fphy.2020.00047
- Wang, B., Hu, S., Teng, Y., Chen, J., Wang, H., Xu, Y., et al. (2024). Current advance of nanotechnology in diagnosis and treatment for malignant tumors. *Signal Transduct. Target. Ther.* 9 (1), 200. doi:10.1038/s41392-024-01889-y
- Wang, D., Sun, Y., Liu, Y., Meng, F., and Lee, R. J. (2018). Clinical translation of immunoliposomes for cancer therapy: recent perspectives. *Expert Opin. Drug Deliv.* 15 (9), 893–903. doi:10.1080/17425247.2018.1517747
- Wang, H., Jordan, V. C., Ramsay, I. A., Sojoodi, M., Fuchs, B. C., Tanabe, K. K., et al. (2019). Molecular magnetic resonance imaging using a redox-active iron complex. *J. Am. Chem. Soc.* 141 (14), 5916–5925. doi:10.1021/jacs.9b00603
- Wang, L., Evans, J. C., Ahmed, L., and Allen, C. (2023). Folate receptor targeted nanoparticles containing niraparib and doxorubicin as a potential candidate for the treatment of high grade serous ovarian cancer. *Sci. Rep.* 13 (1), 3226. doi:10.1038/s41598-023-28424-3
- Waqar, H., Riaz, R., Ahmed, N. M., Majeed, A. I., and Abbas, S. R. (2022). Monodisperse magnetic lecithin-PFP submicron bubbles as dual imaging contrast agents for ultrasound (US) and MRI. *RSC Adv.* 12 (17), 10504–10513. doi:10.1039/D2RA01542K
- Warner, R. (2016). Ultra-high field magnets for whole-body MRI. *Supercond. Sci. Technol.* 29 (9), 094006. doi:10.1088/0953-2048/29/9/094006
- Wei, H., Bruns, O. T., Kaul, M. G., Hansen, E. C., Barch, M., Wiśniowska, A., et al. (2017). Exceedingly small iron oxide nanoparticles as positive MRI contrast agents. *Proc. Natl. Acad. Sci.* 114 (9), 2325–2330. doi:10.1073/pnas.1620145114
- Wen, P., Ke, W., Dirisala, A., Toh, K., Tanaka, M., and Li, J. (2023). Stealth and pseudo-stealth nanocarriers. *Adv. Drug Deliv. Rev.* 198, 114895. doi:10.1016/j.addr.2023.114895
- Werner, E. J., Datta, A., Jocher, C. J., and Raymond, K. N. (2008). High-relaxivity MRI contrast agents: where coordination chemistry meets medical imaging. *Angew. Chem. Int. Ed.* 47 (45), 8568–8580. doi:10.1002/anie.200800212
- Williams, L.-A., Gelman, N., Picot, P. A., Lee, D. S., Ewing, J. R., Han, V. K., et al. (2005). Neonatal brain: regional variability of *in vivo* MR imaging relaxation rates at 3.0 T—initial experience. *Radiology* 235 (2), 595–603. doi:10.1148/radiol.2352031769
- Wu, S.-H., Hung, Y., and Mou, C.-Y. (2011). Mesoporous silica nanoparticles as nanocarriers. *Chem. Commun.* 47 (36), 9972–9985. doi:10.1039/C1CC11760B
- Wu, W., and Zhao, S. (2013). Metabolic changes in cancer: beyond the warburg effect. *Acta Biochim. Biophys. Sin. (Shanghai)* 45 (1), 18–26. doi:10.1093/abbs/gms104
- Xie, W., Guo, Z., Gao, F., Gao, Q., Wang, D., Liaw, B.-s., et al. (2018). Shape-size- and structure-controlled synthesis and biocompatibility of iron oxide nanoparticles for magnetic theranostics. *Theranostics* 8 (12), 3284–3307. doi:10.7150/thno.25220
- Xu, H. L., Yang, J. J., ZhuGe, D. L., Lin, M. T., Zhu, Q. Y., Jin, B. H., et al. (2018). Glioma-targeted delivery of a Theranostic liposome integrated with quantum dots, superparamagnetic iron oxide, and cilengitide for dual-imaging guiding cancer surgery. *Adv. Healthc. Mater.* 7 (9), 1701130. doi:10.1002/adhm.201701130
- Xu, Y., Chen, P., Tang, L., Zhang, X., Shi, F., Ning, X., et al. (2022). Hypoxia responsive and tumor-targeted mixed micelles for enhanced cancer therapy and real-time imaging. *Colloids Surf. B Biointerfaces* 215, 112526. doi:10.1016/j.colsurfb.2022.112526
- Xu, Z., and Chan, A. K. (1999). A near-resonance solution to the bloch equations and its application to RF pulse design. *J. Magnetic Reson.* 138 (2), 225–231. doi:10.1006/jmre.1999.1732
- Yan, Y., Liu, Y., Li, T., Liang, Q., Thakur, A., Zhang, K., et al. (2023). Functional roles of magnetic nanoparticles for the identification of metastatic lymph nodes in cancer patients. *J. Nanobiotechnology* 21 (1), 337. doi:10.1186/s12951-023-02100-0
- Yang, D., Tian, H. Y., Zang, T. N., Li, M., Zhou, Y., and Zhang, J. F. (2017). Hypoxia imaging in cells and tumor tissues using a highly selective fluorescent nitroreductase probe. *Sci. Rep.* 7 (1), 9174. doi:10.1038/s41598-017-09525-2
- Yao, W., Liu, C., Wang, N., Zhou, H., Chen, H., and Qiao, W. (2021). Triple-responsive targeted hybrid liposomes with high MRI performance for tumor diagnosis and therapy. *Mater. Chem. Front.* 5 (16), 6226–6243. doi:10.1039/D1QM00788B
- Yao, Y., Zang, Y., Qu, J., Tang, M., and Zhang, T. (2019). The toxicity of metallic nanoparticles on liver: the subcellular damages, mechanisms, and outcomes. *Int. J. Nanomedicine* 14, 8787–8804. doi:10.2147/ijn.s212907
- Yorulmaz Avsar, S., Kyropoulou, M., Di Leone, S., Schoenenberger, C.-A., Meier, W. P., and Palivan, C. G. (2019). Biomolecules turn self-assembling amphiphilic block Copolymer platforms into biomimetic interfaces. *Front. Chem.* 6, 645. doi:10.3389/fchem.2018.00645
- Yuan, D., Ellis, C. M., and Davis, J. J. (2020). Mesoporous silica nanoparticles in bioimaging. *Materials* 13 (17), 3795. doi:10.3390/ma13173795
- Yuan, D., Ellis, C. M., Mózes, F. E., and Davis, J. J. (2023). Ultrahigh magnetic resonance contrast switching with water gated polymer-silica nanoparticles. *Chem. Commun.* 59 (40), 6008–6011. doi:10.1039/D3CC01205K
- Yuan, H., Jiang, M., Fang, H., and Tian, H. (2025). Recent advances in poly(amino acids), polypeptides, and their derivatives in drug delivery. *Nanoscale* 17 (7), 3549–3584. doi:10.1039/D4NR04481A
- Zeng, Y., Zhang, D., Wu, M., Liu, Y., Zhang, X., Li, L., et al. (2014). Lipid-AuNPs@PDA nanohybrid for MRI/CT imaging and photothermal therapy of hepatocellular carcinoma. *ACS Appl. Mater. & Interfaces* 6 (16), 14266–14277. doi:10.1021/am503583s
- Zhang, D., Zhang, J., Bian, X., Zhang, P., Wu, W., and Zuo, X. (2024). Iron oxide nanoparticle-based T1 contrast agents for magnetic resonance imaging: a review. *Nanomaterials* 15 (1), 33. doi:10.3390/nano15010033
- Zhang, J., Liu, W., Zhang, P., Song, Y., Ye, Z., Fu, H., et al. (2022c). Polymers for improved delivery of iodinated contrast agents. *ACS Biomater. Sci. Eng.* 8 (1), 32–53. doi:10.1021/acsbomaterials.1c01082

Zhang, K., Liu, M., Tong, X., Sun, N., Zhou, L., Cao, Y., et al. (2015). Aptamer-modified temperature-sensitive liposomal contrast agent for magnetic resonance imaging. *Biomacromolecules* 16 (9), 2618–2623. doi:10.1021/acs.biomac.5b00250

Zhang, N., Shu, G., Qiao, E., Xu, X., Shen, L., Lu, C., et al. (2022b). DNA-functionalized liposomes *in vivo* fusion for NIR-II/MRI guided pretargeted ferroptosis therapy of metastatic breast cancer. *ACS Appl. Mater. & Interfaces* 14 (18), 20603–20615. doi:10.1021/acsami.2c01105

Zhang, P., Li, J., Guo, Q., Zhu, Y., Yan, K., Wang, R., et al. (2019). “Chapter 15 - NbTi superconducting wires and applications,” in *Titanium for consumer applications* (Elsevier), 279–296.

Zhang, Y., and Zhao, H. (2016). Surfactant behavior of amphiphilic polymer-tethered nanoparticles. *Langmuir* 32 (15), 3567–3579. doi:10.1021/acs.langmuir.6b00267

Zhang, Z., Wang, Y., Rizk, M. M. I., Liang, R., Wells, C. J. R., Gurnani, P., et al. (2022a). Thermo-responsive nano-in-micro particles for MRI-Guided chemotherapy. *Biomater. Adv.* 134, 112716. doi:10.1016/j.msec.2022.112716

Zhou, T., Zhang, S., Zhang, L., Jiang, T., Wang, H., Huang, L., et al. (2023). Redox ferrocenylseleno compounds modulate longitudinal and transverse relaxation times of FNPs-Gd MRI contrast agents for multimodal imaging and photo-Fenton therapy. *Acta Biomater.* 164, 496–510. doi:10.1016/j.actbio.2023.04.006