



A Bioenergetic Framework for Assessing Soil Organic Matter Persistence

Elizabeth K. Williams* and Alain F. Plante

Department of Earth and Environmental Science, University of Pennsylvania, Philadelphia, PA, United States

OPEN ACCESS

Edited by:

Philippe C. Baveye,
AgroParisTech Institut des Sciences
et Industries du Vivant et
de L'environnement, France

Reviewed by:

Carsten W. Mueller,
Technische Universität München,
Germany
Federico Maggi,
University of Sydney, Australia

*Correspondence:

Elizabeth K. Williams
weliz@sas.upenn.edu

Specialty section:

This article was submitted to
Soil Processes,
a section of the journal
Frontiers in Earth Science

Received: 22 June 2018

Accepted: 10 September 2018

Published: 26 September 2018

Citation:

Williams EK and Plante AF (2018)
A Bioenergetic Framework
for Assessing Soil Organic Matter
Persistence. *Front. Earth Sci.* 6:143.
doi: 10.3389/feart.2018.00143

The emerging view of soil organic matter (SOM) persistence asserts that SOM exists as a continuum of organic material, continuously processed by the decomposer community from large biopolymers to small monomers and with increasing oxidation and solubility, protected from decomposition through mineral aggregation and adsorption. Microbial community and ecosystem dynamics regulate the exchange of both nutrients and carbon between the soil and the atmosphere through the mineralization of SOM. Because these ecosystem dynamics are driven by net energy flows, analysis of SOM bioenergetics can provide complementary constraints to SOM models as well as insight into the fundamental conundrum of why thermodynamically unstable organic matter persists in soil. Microbial substrate preference has been shown to depend on the energy status of the potential substrates in terms of energy required and energy returned. Here we propose a framework for assessing the persistence of SOM utilizing thermally determined activation energy (E_a) and energy density (ED), tested on a suite of soils that have undergone alteration in field or laboratory experiments designed to isolate persistent SOM. Comparison of these energetic parameters in this framework will determine whether a chemical or physical change during SOM decomposition resulted in a change in its environmental persistence. An expanded framework of bioenergetics changes during SOM formation, decomposition, and stabilization is proposed as persistent SOM is characterized by decreased ED and E_a , relative to the bulk SOM.

Keywords: soil organic matter, bioenergetics, thermal analysis, carbon cycling, activation energy

INTRODUCTION

Soil organic matter (SOM) contains more actively cycling carbon than the atmosphere and global terrestrial biomass combined (Jobbágy and Jackson, 2000; Stockmann et al., 2013), and for this reason, even small changes in the size of this carbon pool can have large impacts on the release and storage of atmospheric CO₂ and other greenhouse gases (CH₄, etc.; Davidson and Janssens, 2006; Bond-Lamberty and Thomson, 2010). Microbial community and ecosystem dynamics regulate the exchange of both nutrients and carbon between the soil and the atmosphere through the mineralization of SOM (Falkowski et al., 2008). These microbial community dynamics are driven by net energy flows, in which soil heterotrophs oxidize organic matter, acquiring energy that was fixed during photosynthesis, and channeling this energy into storage in the form of tissues (i.e., microbial biomass) or other organic materials, or expending this energy in metabolic processes (i.e., respiration; Odum et al., 1962; Currie, 2003). Despite its importance, the mechanisms controlling SOM stability and persistence are still debated (Schmidt et al., 2011). However, the growing

consensus is that environmental and biological controls (i.e., microbial community composition, organo-mineral associations, temperature, and moisture) exert a stronger control over SOM stability than molecular structure alone (Schmidt et al., 2011; Lehmann and Kleber, 2015).

Microbial substrate preference has been shown to depend on the energy status of the potential substrates in terms of energy required to metabolize the substrate and energy returned from metabolism of the substrate. In a study of thermophilic microbes, Amenabar et al. (2017) showed that the microbial energy demand (or required energy input from the microbial community) dictated the substrate choice rather than the potential energy supply of the substrates, with microbes preferring easier to decompose substrates over more energy rich substrates. Studying pyrogenic carbon, Harvey et al. (2016) demonstrated that the microbial community exhibited a greater preference for organic matter that returned the most energy for the smallest energy investment, which they expressed as the return-on-energy-investment (ROI) ratio. The energy status of SOM should therefore also be an important factor in its persistence. Barré et al. (2016) demonstrated that for a variety of climates and soil types, persistent SOM showed specific thermally determined energetic signatures, namely, that persistent SOM became less energy dense (yielding less energy during ramped combustion) with increasing duration of minimal C inputs via bare fallow treatment, suggesting that soil microorganisms preferentially mineralize high-energy SOM. In a similar study employing thermal decomposition of SOM, Williams et al. (2018) demonstrated distinct bioenergetic signatures between particulate (faster cycling) SOM vs mineral-associated (slower cycling) SOM. The particulate SOM fractions were characterized by larger energy densities and activation energies, whereas the mineral-associated SOM fractions were characterized by smaller energy densities and activation energies.

Here we propose a framework for assessing the persistence of SOM using thermally determined activation energy and energy density (ED). ED is a measure of the total net bond energy released during ramped combustion and can be used as a proxy for the molecular composition of SOM, such that more condensed or polymerized compounds would have higher ED. ED also represents the amount of available or potential chemical energy the microbial community might thus gain through the complete decomposition of the SOM for growth and respiration. Activation energy (E_a) is a measure of the input energy required to combust SOM and can be used as a proxy for the molecular composition of SOM and as a proxy for the energetic barrier to decomposition, and thus SOM persistence. We will test this framework using several soils that have undergone alteration in field or laboratory experiments designed to isolate persistent SOM: bare fallow and chemical fallow field trials, land-use/land-cover changes, long-term laboratory incubations, and acid hydrolysis. Many fractionation methods intended to isolate persistent SOM are still hampered by methodological problems (Bruun et al., 2008). In several persistent-SOM isolation method comparison studies (Jenkinson, 1971; Balesdent, 1996; Balabane and Plante, 2004; Plante et al., 2005; Krull et al., 2006; Paul et al., 2006; Bruun et al., 2008), the authors found

variable or no relationships between the isolated soil fractions and environmental persistence. Likewise, uncertainty in estimates of the size of labile and persistent SOM pools are a major source of error in modeling soil organic C turnover (Falloon and Smith, 2000) and response to climatic and environmental changes.

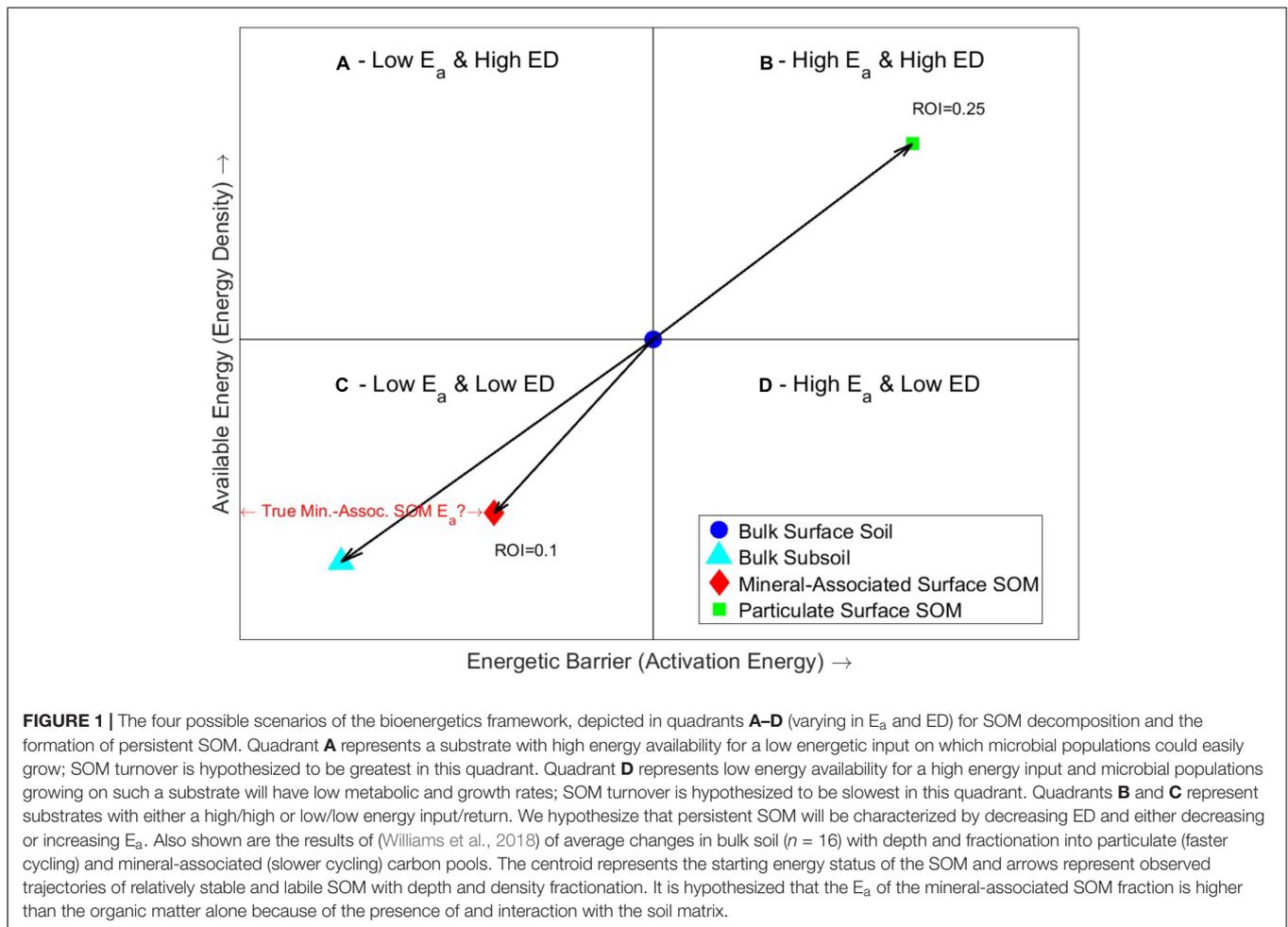
Because the energy status of SOM is a fundamental factor in its environmental persistence, we expect that comparison of changes in these two energetic parameters (ED and E_a) as discussed in the following bioenergetics framework will allow us to determine if a chemical or physical change during SOM decomposition has resulted in a change in its environmental persistence or biodegradability. We hypothesize that the ED will be lower in persistent SOM compared to the bulk soil as energy dense particulate organic matter (mostly comprised of plant residues) decompose into smaller particles and biomolecules. For E_a , we propose two competing hypotheses that offer the opportunity to test opposing theories of persistent SOM formation. If persistent SOM formation follows the humification theory, E_a is hypothesized to increase as persistent SOM would have been formed by the polymeric condensation of decomposition products into large and stable biopolymers (i.e., humus). If persistent SOM follows the progressive decomposition or selective preservation theories, E_a is hypothesized to decrease as the persistent SOM is comprised of smaller biomolecules.

MATERIALS AND METHODS

The Bioenergetic Framework

The stability and persistence of SOM is dependent upon the balance of the energetic barriers to decomposition (i.e., physical and chemical protection, availability of electron acceptors, or enzyme production) and the energy available or released to the microbial community upon decomposition for metabolic functions. Comparison of these energies (available energy and energy barrier) leads to four possible scenarios (**Figure 1**) varying in E_a and ED that have previously been postulated by Rovira et al. (2008) for litter decomposition and are now discussed in relation to SOM. Scenario 1, symbolized by the top-left quadrant (A), represents a substrate with a high energy availability for a low energetic investment. Microbial populations could easily grow on quadrant A substrates and thus SOM turnover would be fastest. Scenario 2, symbolized by the bottom-right quadrant (D), represents the opposite relationship in which the substrate has low energy availability but requires a high energy investment. Microbial populations growing on such a substrate will have low metabolic and growth rates and thus SOM would exhibit the slowest turnover and be considered persistent. Scenarios 3 and 4, represented by quadrants B and C, fall between the extremes of scenarios 1 and 2 (quadrants A and D) and in some cases may be equal in quality and biodegradability in terms of energy input and return. However, because of the greater available energy of quadrant B substrates, it is postulated that substrates in quadrant C are generally less biodegradable and more persistent than substrates in quadrant B.

In soils, the interpretation of E_a is complicated by the mineral matrix, which can bind with SOM, forming additional



barriers to decomposition reflected in both the thermal and natural decomposition of SOM (Leifeld and von Lützw, 2014). Counterintuitively, Williams et al. (2018) found that organic matter chemically protected by strong mineral bonds required less E_a than particulate organic matter (Figure 1). A similar difference was observed in respiration experiments of mineral and organic soils, in which E_a was lower in mineral soils (Leifeld and von Lützw, 2014). Williams et al. (2018) suggested that while the E_a of the bound-SOM was lower, it is still reflective of the additional barriers to decomposition provided by the mineral matrix, (as evidenced in experimental mixtures of kerogen and common soil minerals; Dembicki, 1994) as well as compositional differences between the particulate and mineral-associated SOM. Williams et al. (2018) also observed that with depth (and increased residence time) in the soil column, there was also a decrease in ED and E_a . Based on these energetic differences observed between mineral-associated and particulate organic matter, we hypothesize that there is a difference in environmental stability and persistence between quadrants B and C, such that organic matter trending toward quadrant C has a greater potential for environmental persistence likely because of increasing mineral association and the preferential loss of energy dense particulate organic matter.

Experimental Soil Sample Sets

Samples for this study were collected from archived soil samples from several previously published field-based and laboratory-based experiments designed to isolate or characterize persistent SOM through the loss or removal of labile SOM (Table 1). Additional details about these soils, experimental conditions, and additional analyses can be found in the following referenced works.

Field Experiments

We used soils from four long-term bare fallow (LTBF) field experiments from northwestern Europe, one chemical fallow field experiment from New Zealand, and five land-use conversion experiments located across a mean annual temperature gradient (Table 1). The LTBF soils used were from bare fallows located at Château de Versailles in Versailles, France, Rothamsted Research in Harpenden, United Kingdom, the Swedish University of Agricultural Sciences at Ultuna in Uppsala, Sweden, and at the Askov Experimental Station, Denmark (Barré et al., 2016). Surface soils (up to 25 cm depth) were collected from two field replicate plots at each location and from triplicate plots in Askov (see Table 1 for soil depths). Bare fallow treatments span 27 years at Askov to 79 years at Versailles (Table 1),

TABLE 1 | Location, treatment, depth, soil carbon contents (pre and post), and percent carbon change for the field and laboratory based experiments.

Experiment		Location	Treatment	Soil depth (cm)	Initial soil C content (mg/g)	Final soil C content (mg/g)	ΔC (%)
Field-based	Fallow ^{1,2}	Versailles, FR	79 years	0–25	17.5	6.2	–64.6
		Rothamsted, United Kingdom	49 years	0–23	30.3	9.7	–67.9
		Ultuna, SW	53 years	0–20	15.0	9.2	–38.7
		Askov, DK	27 years	0–20	16.5	11.9	–27.9
		Canterbury, NZ	13 years	0–7.5	31.1	20.8	–33.1
			13 years	7.5–15	23.8	19.4	–18.5
	Land-use change ³	Waggoner's Ranch, TX	Grassland to cropland	0–20	11.2	10.2	–8.9
			Mandan, ND	Grassland to cropland	0–20	32.4	28.0
		Akron, CO	Grassland to cropland	0–20	11.6	6.9	–40.5
		Nova Vida Ranch, BR	Forest to pasture	0–20	10.6	14.1	+ 33.0
		Alajuela, CR	Forest to pasture	0–20	200.2	141.6	–29.3
		Laboratory	Incubation ³ (588 days, 35°C)	TX, ND, CO	Grassland	0–20	18.4
BR, CR	Forest			0–20	105.4	94.7	–10.2
TX, ND, CO	Cropland			0–20	15.0	12.6	–16.0
BR, CR	Pasture			0–20	77.9	72.3	–7.2
Acid hydrolysis ⁴ (6 M HCl, 16 h, 95°C)	Breton, AB		Forest	0–5	67.2	48.5	–27.8
	Breton, AB		Conventional till	0–5	12.9	9.2	–28.7
	Breton, AB		Conventional till	0–5	17.9	12.7	–29.1
	Scott, SK		Conventional till	0–5	40.8	41.1	+ 0.7
	Scott, SK		Grassland	0–5	59.6	26.3	–55.9

Superscript numbers refer to the following referenced works, in which these data have been previously reported and other site or experimental information including detailed soil descriptions can be obtained: ¹Barré et al. (2016), ²Gregorich et al. (2015), ³Haddix et al. (2011), and ⁴Plante et al. (2006).

and samples were collected in the following years: (Versailles) 1929, 1934, 1939, 1950, 1960, 1968, 1980, 1991, 2002, 2008; (Rothamsted) 1959, 1963, 1971, 1987, 2000, 2008; (Ultuna) 1956, 1967, 1974, 1985, 1995, 2009; (Askov) 1956, 1962, 1968, 1976, 1983. During the duration of the bare fallows, soil organic C concentrations exhibited a relative decrease of 29–65% (Barré et al., 2016).

Chemical bare fallow soils were collected from three field replicate plots from Lincoln, New Zealand. Soils were sampled at three depths (0–7.5, 7.5–15, and 15–25 cm). These soils span 13 years (beginning in 2000) of chemical fallow conditions (Gregorich et al., 2015), resulting in a relative decrease of 33% in soil organic C concentrations in the surface soil and 18% in the two subsoils.

Soils from land-use conversion experiments were collected from five sites along a mean annual temperature gradient (2–25.6°C; Haddix et al., 2011). The temperate sites had paired native grassland and cultivated land uses (Mandan, ND, United States; Akron, CO, United States; and Waggoner Ranch, TX, United States) and the tropical sites had paired native forest and pasture land uses (Alajuela, Costa Rica, and Nova Vida Ranch, Brazil). Transitions from native grassland or forest to cultivated cropland or pasture occurred between 1957 and 1984. Surface soil samples (0–20 cm) were collected from each land-use, resulting in 10 land-use conversion samples (five native + five altered). The land-use conversion resulted in relative decreases in soil organic C concentration (9–40%) due to decades of intensive agriculture. Only the Brazil site

experienced a 33% increase in soil C, where an increase in the proportions of carbohydrates and peptides was noted that was not observed in the other locations (Haddix et al., 2011).

Laboratory Experiments

The 10 previously described land-use conversion soils, comprised of three grassland, three cropland, two forests, and two pasture soils, were subjected to a long-term laboratory incubation experiment. Samples were incubated for 588 days at 35°C and optimal, constant moisture conditions to maximize the microbial mineralization of labile SOM. Based on the total respired carbon (Haddix et al., 2011), the long-term incubations resulted in a relative decrease of 7–20% in soil organic C concentrations.

Five surface soil samples (0–5 cm) were collected from two long-term agricultural field experiments in Alberta, Canada and Saskatchewan, Canada (Plante et al., 2006). The two sites had paired either grassland or forest and cultivated (conventional till) land uses. Soils were hydrolyzed with 25 mL of 6 M HCl at 95°C for 16 h, resulting in relative decreases in soil organic C concentration decreases of 28–56%, although one site experienced a 0.7% increase in soil C.

Activation Energy and Energy Density Determinations

Thermal analyses were performed by ramped combustion using a Netzsch STA 449PC Jupiter simultaneous thermal

analyzer equipped with an automatic sample carrier (ASC) and a type-S platinum/ rhodium (Pt/PtRh) sample carrier (Netzsch-Gerätebau GmbH, Selb, Germany). Thermal analysis has been demonstrated to be highly reproducible on a variety of soil types (Plante et al., 2009), and thus thermal analyses analytical replicate aliquots were not performed. Samples were weighed to contain approximately 1 mg C and were heated from ambient temperature (25°C) to 105°C at 10°C min⁻¹ under an oxidizing atmosphere of 40 mL min⁻¹ of CO₂-free synthetic air (20% O₂ and N₂ balance). Samples were held at 105°C for 15 min to allow for drying/moisture equilibration, then heated at 10°C min⁻¹ to 800°C. Evolved CO₂ was analyzed using a coupled LICOR LI-820 infrared gas analyzer. Differential scanning calorimetry (DSC) heat flux (the exothermic or endothermic energy flux from the sample, referenced to an empty Pt/Rh crucible), thermogravimetric (TG) mass loss, and evolved CO₂ gas (CO₂-EGA) signals were recorded every second. DSC and CO₂ baselines were corrected *a posteriori* using the base R package and the smooth.spline() function using a spline and linear baseline, respectively (refer to Plante et al., 2011 for additional information on baseline corrections). DSC integration was also performed using the base R package and the sintergral() function.

Total exothermic energy content (in mJ) was determined by integrating the DSC heat flux (in mW) over the exothermic region 190–600°C, which represents the temperature range in which SOM is oxidized (Rovira et al., 2008). TG mass loss was determined for the same range. This mass loss is attributable largely to SOM combustion, with minor contributions from dehydration of minerals such as kaolinite. ED (in J mg⁻¹ OM) was thus determined by dividing energy content by TG mass loss (Rovira et al., 2008; Plante et al., 2011).

Activation energy (E_a , in kJ mol⁻¹ CO₂) was determined following the methods of Burnham and Braun (1999) and Williams et al. (2014), assuming first-order reaction kinetics during ramped combustion. The fraction of sample C remaining (x) for each time step was calculated from the evolved CO₂ data. Each data point over the decomposition temperature range (105–800°C) is then assumed to represent an instantaneous measurement of the rate constant of combustion (k) at each temperature, and E_a is determined from a modified Arrhenius plot of $\ln|\ln(x)|$ vs $1/T$, in which the slope of the regression is equal to $-E_a/R$ where $\ln|\ln(x)|$ is the natural log of the absolute value of the natural log of x and R is the universal gas constant. Thermally derived values of E_a are expected to be greater than what occurs naturally in soils or during laboratory soil incubations (Leifeld and von Lütow, 2014) because of the complete decomposition of each soil instead of a relatively biochemically labile portion and because of the lack of microbial and fungal enzymes to catalyze the decomposition reactions.

Additionally, the ROI ratio (Harvey et al., 2016) which has been demonstrated to be directly related to the biodegradability of specific types of organic matter and thus environmental persistence was calculated for each soil by dividing ED by E_a .

Statistical Analyses

Differences in ED, E_a , and ROI between pre- and post-treatment soils were analyzed using an independent one-way ANOVA. Statistical analyses were performed for each of the following paired pre- and post-treatment groupings: all soils ($n = 38$), soils separated by field ($n = 23$) or lab ($n = 15$) setting, and soils grouped by individual experiments [bare fallow; $n = 2$ ($\times 4$ experiments), chemical fallow; $n = 3$ ($\times 3$ soil depths), land-use; $n = 5$, incubation; $n = 10$, and hydrolysis; $n = 5$]. All analyses were performed using MATLAB and differences in E_a , ED, and ROI were considered statistically significant when $p \leq 0.05$.

RESULTS

The E_a and ED varied widely among the initial soils. E_a ranged from 57 to 69 kJ/mol CO₂ and ED ranged from 7 to 15 J/mg OM, reflecting the range of soil types, climates, land uses, vegetation types, and organic matter inputs. The ROI ratios varied between the initial and final soils, spanning 0.15–0.24 in the untreated soils to 0.07–0.33 in the final soils, implying changes in SOM biodegradability over the course of the experiments (Table 2). The changes in ROI were not strongly correlated to changes in carbon ($R^2 = 0.20$, $p = 0.04$; least squares linear regression). When analyzed collectively, only the E_a was statistically different (decreased) between pre- and post-treatment soils (ED $p = 0.20$; E_a $p = 0.005$; ROI $p = 0.48$).

Field-Based Experiments

In the field-based studies (i.e., bare/chemical fallow and land-use change), post-treatment soils displayed a decrease in E_a and ED (Figure 2) relative to the initial soils during the course of the experiments (ED $p < 0.001$; E_a $p = 0.003$), trending toward quadrant C. In the LTBF experiments, the evolution of and variation in ED and E_a with labile SOM loss over time is visible (Figures 2A–D). For the 79 year fallow at Versailles, only the decrease in ED over the course of the experiment was statistically significant (ED $p = 0.001$; E_a $p = 0.08$). For the other bare fallows, only the decrease in E_a was statistically significant: Askov (ED $p = 0.16$; E_a $p = 0.03$), Rothamsted (ED $p = 0.28$; E_a $p < 0.000$), Ultuna (ED $p = 0.23$; E_a $p = 0.003$).

In the one field study considering soil depth (Figure 2E), there was a statistically significant decrease in the E_a with depth and no change in ED (initial ED $p = 0.14$; initial E_a $p = 0.03$; final ED $p = 0.12$; final E_a $p = 0.03$). At all depths, the 13 year chemical fallow soils demonstrated statistically significant decreases in E_a (0–7.5 cm $p = 0.03$; 7.5–15 cm $p = 0.001$; 15–25 cm $p = 0.001$) over time. However, only the two subsoil samples exhibited significant decreases in ED over the course of the fallow treatment (0–7.5 cm $p = 0.25$; 7.5–15 cm $p = 0.004$; 15–25 cm $p = 0.04$). Neither the changes in ED nor E_a were statistically significant with the changes in land-use (Figure 2F) from forest/grassland to pasture/cropland (ED $p = 0.44$; E_a $p = 0.22$).

Although ROIs for the field samples (collectively) decreased after treatment ($p = 0.04$), the decrease in E_a and ED of the field soils was not always reflected by a decrease in ROI. Only the Rothamsted ($p = 0.01$), Versailles ($p = 0.04$), and Ultuna

TABLE 2 | The changes in the ROI ratio with labile SOM loss or removal from the field and laboratory based experiments.

Experiment	Treatment	Soil depth (cm)	Initial ROI	Final ROI	Δ ROI (%)	
Field-based	Fallow	79 years	0–25	0.16	0.07	–56.3
		49 years	0–23	0.17	0.11	–35.3
		53 years	0–20	0.19	0.17	–10.5
		27 years	0–20	0.16	0.13	–18.8
		13 years	0–7.5	0.23	0.23	0.0
		13 years	7.5–15	0.22	0.22	0.0
		13 years	15–25	0.24	0.22	–8.3
	Land-use change	Grassland to cropland	0–20	0.17	0.13	–23.5
		Grassland to cropland	0–20	0.21	0.19	–9.5
		Grassland to cropland	0–20	0.19	0.12	–36.8
		Forest to pasture	0–20	0.15	0.16	+6.7
		Forest to pasture	0–20	0.22	0.22	0.0
		Grassland	0–20	0.20	0.19	–5.0
Laboratory	Incubation	Grassland	0–20	0.20	0.19	–5.0
		Forest	0–20	0.20	0.25	+25.0
		Cropland	0–20	0.16	0.18	+12.5
		Pasture	0–20	0.20	0.22	+10.0
	Acid hydrolysis	Forest	0–5	0.24	0.25	+4.2
		Conventional till	0–5	0.23	0.33	+43.5
		Conventional till	0–5	0.25	0.26	+4.0
		Conventional till	0–5	0.22	0.23	+4.5
		Grassland	0–5	0.20	0.23	+15.0

($p = 0.01$) bare fallows displayed decreased ROI (and decreased hypothesized biodegradability), while the remaining bare fallow (Askov, $p = 0.26$), chemical fallow soils (0–7.5 cm $p = 0.67$; 7.5–15 cm $p = 0.17$; 15–25 cm $p = 0.28$), and the land-use soils ($p = 0.20$) displayed no change in ROI possibly due to proportional changes in ED and E_a .

Laboratory-Based Experiments

In the laboratory-based experiments (i.e., incubation and acid hydrolysis), post-treatment soils displayed no statistically significant changes in E_a and ED (Figures 2G,H) relative to the initial soils (ED $p = 0.14$; E_a $p = 0.58$), when analyzed collectively. The long-term laboratory incubation (35°C, 588 days) resulted in highly variable alterations in ED and E_a (Figure 3A), seemingly dependent upon land-use (ED $p = 0.53$; E_a $p = 0.52$). Grassland soils displayed decreased ED and E_a (toward quadrant C) following incubation similar to the signature of mineral associated SOM (Williams et al., 2018) and the fallow soils, whereas forest and pasture soils displayed a decrease in E_a and increase in ED (toward quadrant A) similar to the decomposition of forest litter (Rovira et al., 2008), and croplands displayed increased ED and E_a (toward quadrant B) similar to the response of the hydrolyzed soils (Figure 2H) and the energetic signature of the particulate organic matter (Williams et al., 2018) in Figure 1. However, none of the changes in ED and E_a due to long-term incubation were statistically significant when separated by land-use (grassland ED $p = 0.22$; grassland E_a $p = 0.49$; forest ED $p = 0.84$; forest E_a $p = 0.42$; cropland ED $p = 0.63$; cropland E_a $p = 0.55$; pasture ED $p = 0.87$; pasture E_a $p = 0.68$). Labile SOM removal by acid hydrolysis (Figure 3B) resulted in statistically significant increases in E_a and

ED relative to the untreated soil, trending toward quadrant B (ED $p = 0.02$; E_a $p = 0.01$). The net changes in E_a and ED for all experiments are shown in Figure 3.

Return-on-energy-investments for the laboratory-based samples (collectively) did not change after treatment ($p = 0.14$). The ROI also did not change when the lab soils were separated by experiment (incubation; $p = 0.29$, hydrolysis; $p = 0.12$), possibly due to proportional changes in ED and E_a . Similar to the lack of pre- and post-treatment differences in ED and E_a with land-use, ROI also did not change (grassland $p = 0.64$; forest $p = 0.21$; cropland $p = 0.53$; pasture $p = 0.36$) with incubation.

DISCUSSION

The emerging view of SOM persistence asserts that SOM exists as a continuum of organic material, continuously processed by the decomposer community from large biopolymers to small monomers and with increasing oxidation and solubility, protected from decomposition through mineral aggregation and adsorption (Lehmann and Kleber, 2015) and that the persistence of SOM is thus a property of the ecosystem as a whole (Schmidt et al., 2011). Because these ecosystem dynamics are driven by net energy flows, analysis of SOM bioenergetics can provide complementary constraints to SOM models as well as insight into the fundamental conundrum of why thermodynamically unstable organic matter persists in soil.

Long-term bare fallow experiments present the best case for studying the bioenergetic status of persistent SOM (Ruhlmann, 1999). Soils are kept free of vegetation and additional organic inputs for extended periods of time (up to 79 years in the

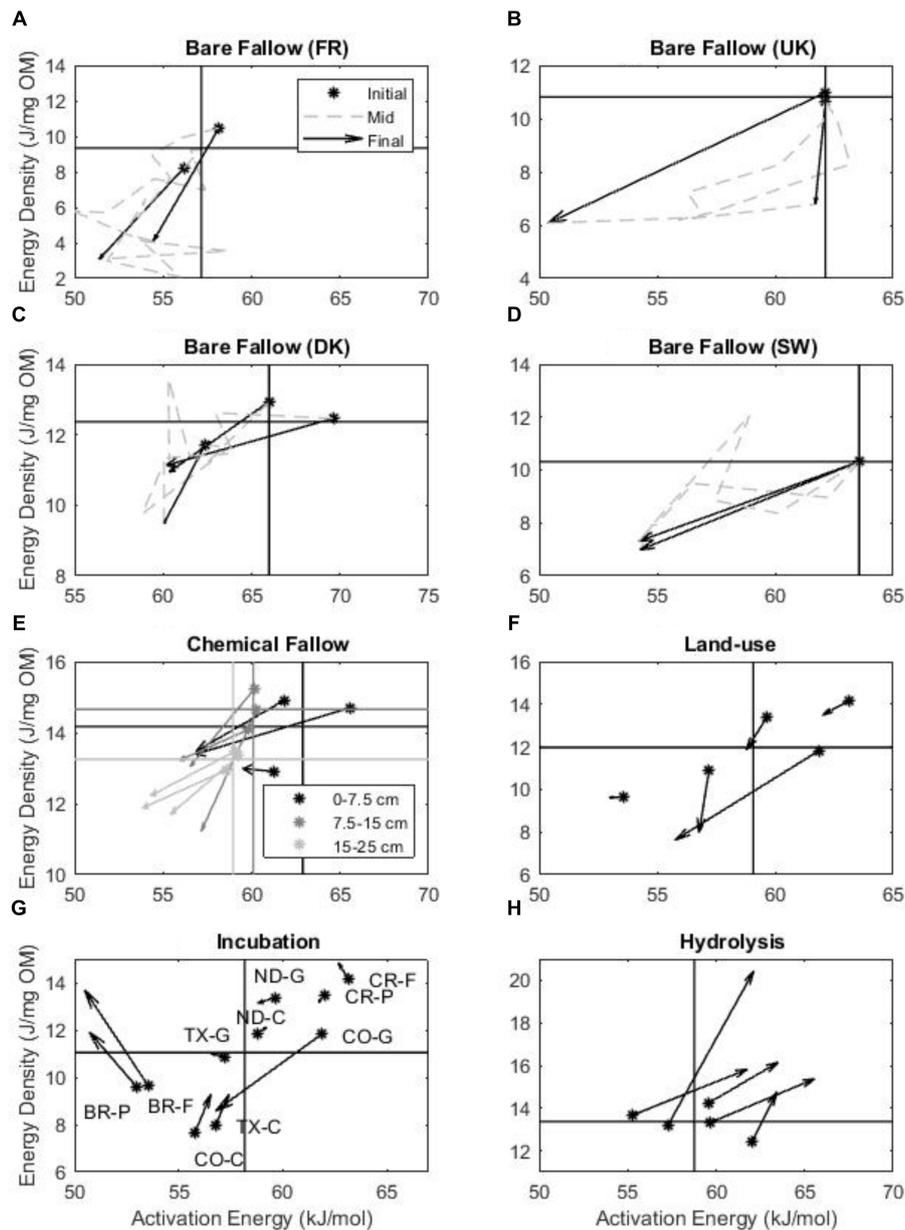


FIGURE 2 | The changes in E_a and ED with labile SOM loss or removal from the field and laboratory-based experiments presented in the bioenergetics framework. Changes are indicated by the circles (initial soils) and arrowheads (final soils). For each experiment, the axes cross at the average initial soil E_a and ED to demonstrate the trajectory into one of the four bioenergetics scenarios relative to the initial soil. **(A–D)** The change in ED and E_a over the course of the long-term bare fallow experiments (**A** Versailles, FR; **B** Rothamsted, United Kingdom; **C** Askov, DK; **D** Ultuna, SW). **(E)** The change in ED and E_a after 13 years of chemical fallow for surface soils and soils at depth. **(F)** The changes in ED and E_a after land-use change from grassland or forest to cultivated cropland or pasture. **(G)** The change in ED and E_a after laboratory incubation for 588 days at 35°C. Letters indicate the origin and land-use of each soil (G – grassland, P – pasture, C – cropland, F – forest) listed in **Table 1**. **(H)** Changes in the ED and E_a of SOM after acid hydrolysis. Please note the different axes scales.

experiments studied here), allowing initially present organic matter to decompose under field conditions and soils to become more enriched in persistent or stable SOM as the more labile SOM decomposes naturally (Barré et al., 2016). Although consisting of different durations and climatic conditions, all fallow experiments (**Figures 2A–E**) follow the first and third hypotheses of persistent SOM, trending into quadrant C

of the bioenergetics framework characterized by decreased ED and decreased E_a . This trajectory is comparable to the observed changes in ED and E_a for bulk soils with depth and fractionation into particulate and mineral-associated SOM (**Figure 1**). These reductions in ED over time were previously observed (Barré et al., 2016). This trend in the 79 year bare fallow is supported by the observed reduction in energy dense

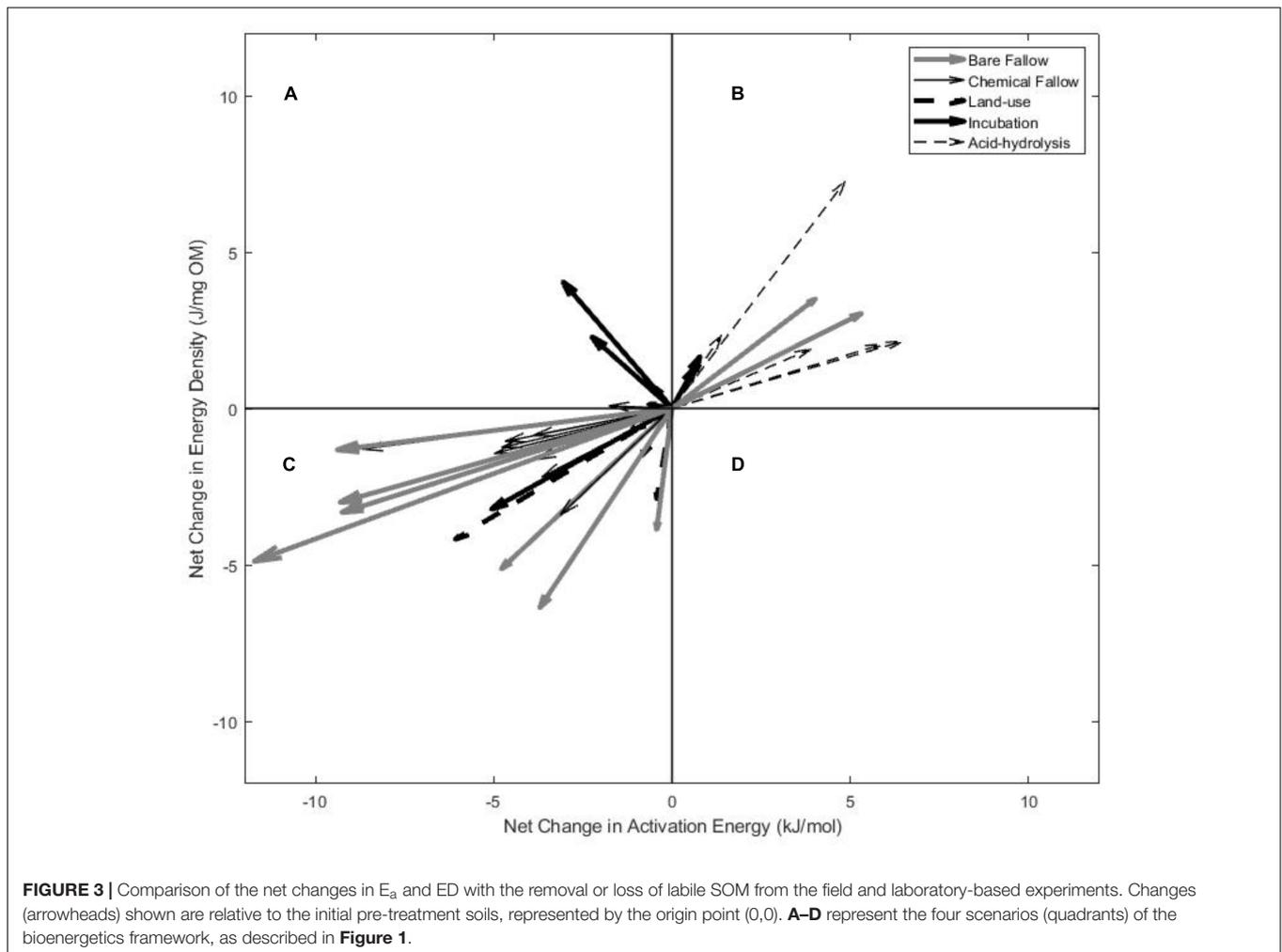


FIGURE 3 | Comparison of the net changes in E_a and ED with the removal or loss of labile SOM from the field and laboratory-based experiments. Changes (arrowheads) shown are relative to the initial pre-treatment soils, represented by the origin point (0,0). **A–D** represent the four scenarios (quadrants) of the bioenergetics framework, as described in **Figure 1**.

particulate organic matter (Balabane and Plante, 2004) over the course of the experiment, suggesting that organo-mineral complexes (similar to the mineral-associated SOM in Williams et al., 2018) dominate in the latter stages of the fallow and comprise the more persistent SOM. Mineral-associated soil fractions with the longest turnover are assumed to have larger proportions of easily metabolizable organic molecules with low thermodynamic stability (lowest abundance of aromatic groups, highest O-alkyl C/aromatic C ratio, highest proportion of thermally labile materials, and highest ratio of substituted fatty acids to lignin phenols), whereas the younger soil fractions are assumed to have higher proportions of stable organic matter considered more difficult to metabolize, as evidenced by Kleber et al. (2011). This decreased thermodynamic stability is evidenced in the decreased ED or E_a over time, simultaneously the decreased biodegradability is evidenced in the decreasing ROI ratios (**Table 2**).

Organic matter in subsoils is characterized by longer turnover times compared to surface material (Rumpel and Kögel-Knabner, 2011) and also presents a useful opportunity to investigate SOM biogenetic changes at depth as microbial activity may be reduced by suboptimal environmental conditions, nutrient or energy

scarcity, and organic matter may be less accessible because of its association with mineral surfaces (Schmidt et al., 2011). At depth in the chemical fallow experiments (**Figure 2B**), soils also follow the third hypothesized trajectory of persistent SOM (**Figure 1**), with deeper soils trending into quadrant C with decreased E_a (as indicated in migrating origin points of the initial soils with depth). However, the ROI ratios (**Table 2**) do not suggest changes in SOM biodegradability with depth because of the lack of significant change in ED.

Changes in land-use are often disruptive to soil structure, alter natural C inputs, and often result in degradation of biogeochemically labile SOM (Guo and Gifford, 2002). However, when statistically analyzed collectively, changes in land-use in the soils were insufficient in addressing changes in the energy status of SOM, likely due to the continued input of labile organic matter inputs as well as the lack of field/site replicates, and thus there was no true isolation of persistent SOM with the land-use conversions.

Incubations offer a second-best option for studying the bioenergetic status of persistent SOM, as SOM is allowed to decompose in optimal conditions and positive correlations between ED and microbial respiration have been observed in

several studies (Plante et al., 2011; Peltre et al., 2013; Stone and Plante, 2015). Although conducted at an elevated temperature, compared to the native MATs, and for almost 2 years to speed the microbial degradation of labile SOM, these incubation experiments were also insufficient to observe differences in SOM E_a and ED. The variable responses in E_a and ED are likely a result of the different land-uses, soil types, climates, and vegetation and the interpretations are limited again by the lack of field/site replicates. Incubations are also limited by the fact that truly persistent SOM is not accessed by the microbes, but rather isolate a large intermediate pool of SOM (Haddix et al., 2011).

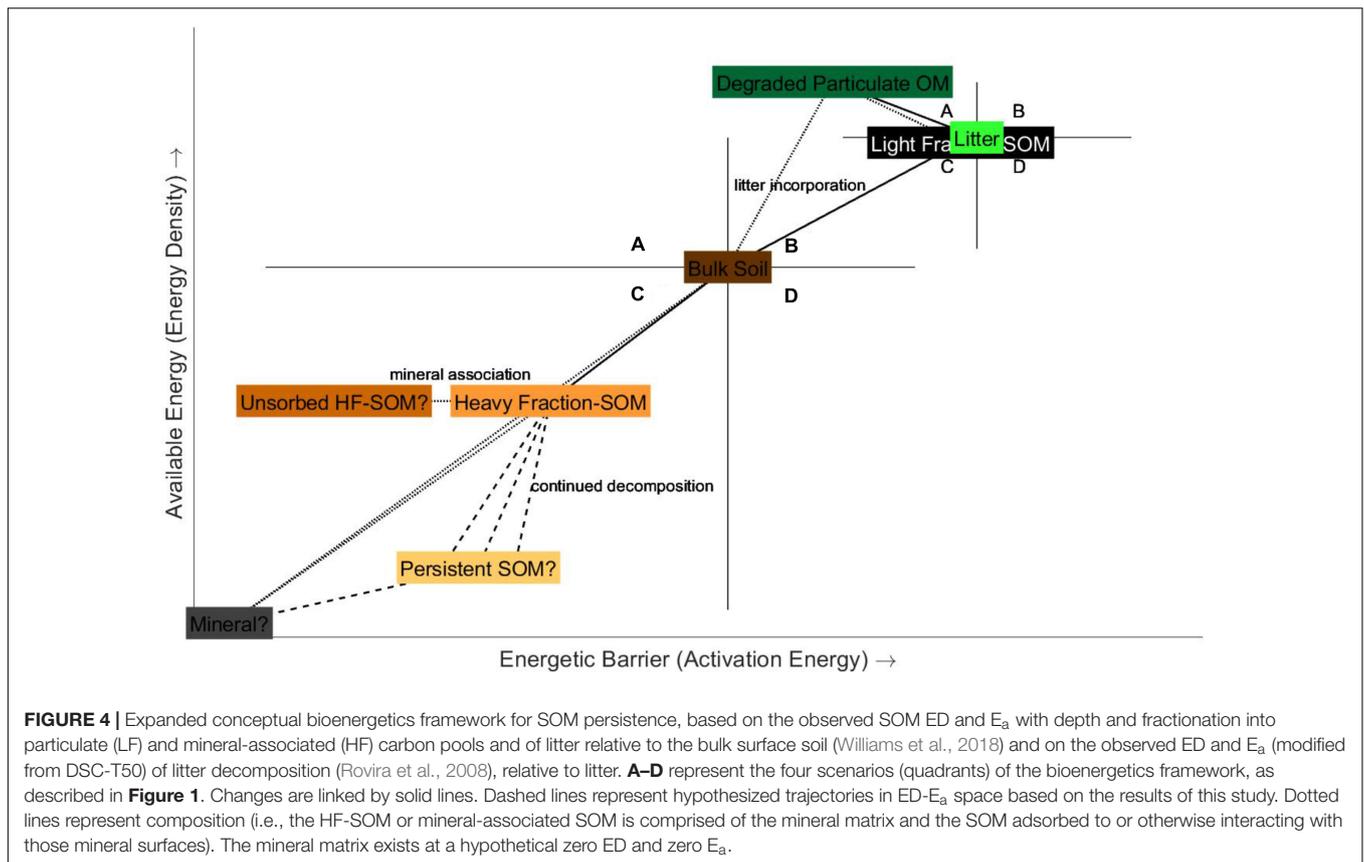
In direct contrast to the fallow experiments, the acid-hydrolyzed soils trend toward quadrant B (**Figure 3B**) following none of the hypothesized trajectories of persistent SOM but rather following the observed trajectories of labile SOM as presented in **Figure 1**. The use of acid hydrolysis (and other wet chemical oxidation methods such as with hydrogen peroxide) rests on the premise that organic molecules resistant to chemical hydrolysis are also resistant to soil enzymatic attack and are therefore also more biologically stable and persistent in the environment (Krull et al., 2006; Greenfield et al., 2013). While radiocarbon analysis of acid hydrolysis residues has been demonstrated to be older than the bulk soils (Trumbore et al., 1989; Paul et al., 2006), acid hydrolysis is known to preferentially hydrolyze nucleic acids, proteins, and carbohydrates with *O*-alkyl functional, while leaving alkyl and aromatic compounds (Kiem et al., 2000; Bruun et al., 2008) and has been shown to not equate with resistance (Greenfield et al., 2013). Additionally, in the study of young (40-days-old) and old (40-years-old) carbon mixtures, Bruun et al. (2008) found that acid hydrolysis consistently removed more old carbon, suggesting that the remaining material contained a large portion of faster cycling carbon. Similar to the results of Williams et al. (2018) in which unsaturated/aromatic soil (low H:C) carbon pools, trended toward scenario B, these results are also suggestive of the loss of non-aromatic organic material during acid hydrolysis or artifacts from the conversion of carbohydrates into molecules compositionally indistinguishable from non-hydrolyzable SOM (Greenfield et al., 2013).

The fact that none of the experimental soils trended into quadrant D (**Figures 2, 3**), or the most energetically unfavorable pool of organic substrates characterized by low ED and high E_a (Hypotheses 1 and 2), supports emerging concepts and models of SOM formation and stabilization away from classic humification theories. Decreased ED is expected with SOM decomposition as larger plant biopolymers are processed into smaller molecules; however, increased E_a is postulated to be the result of mineral association, condensation or polymerization reactions, or the loss of less molecularly complex OM. Mineral association is not expected to significantly affect the E_a of SOM in organic soils, but is expected to increase the E_a of SOM in mineral soils due to additional organic matter-mineral adsorption and complexation bonds which must be broken for SOM decomposition, mineralization or combustion (as conceptualized in **Figure 1**). However, it is expected that this increased E_a may be relatively small as evidenced in a study of the effects of various mineral matrices (quartz, calcite, dolomite,

kaolinite, and bentonite) on kerogen decomposition kinetics in which measured E_a was increased compared to kerogen alone, but only at low organic carbon concentrations (Dembicki, 1994) and by the observation that mineral-associated SOM displays lower E_a than particulate SOM (Williams et al., 2018). However, the presence of carbonates would increase the measured E_a as carbonates decompose into CO_2 at relatively high temperatures ($\sim 600\text{--}800^\circ\text{C}$) and would be indiscriminately incorporated into the E_a calculation. Condensation reactions, such as those occurring during the classic soil humification models, suggest the synthesis of large and stable biopolymers and molecules (humus) from decomposition products. However, the lack of evidence for the physical existence of humic substances *in situ* independent of the alkaline extraction procedure (Schmidt et al., 2011; Lehmann and Kleber, 2015) has led to the acceptance of other models of SOM formation that can be observed in the soil. Additionally, condensation or polymerization reactions occurring during SOM processing would result in an increase in ED as well as an increase in E_a and would likely not appear in quadrant D, but instead in quadrant B. Similarly, the loss of less molecularly complex OM would increase E_a but would also increase ED, as what is postulated to have happened during the acid hydrolysis experiments. Hypothesized substrates in quadrant D were expected to have the slowest turnover and thus be the most environmentally persistent. However, it does not seem plausible within the accepted theories of SOM formation and stabilization for SOM to exist in this hypothetical state, rather the most persistent SOM likely exists in the lower ED extremes of quadrant C (**Figure 4**) as suggested by the fallow and density fractionation experiments.

Soil organic matter substrates characterized by high ED and low E_a (quadrant A) are expected to turnover relatively rapidly within the bioenergetics framework and thus would not become a dominant feature of the soils where persistent SOM dominates over labile SOM such as the post-treatment samples in this study. Only the coupled native forest/pasture incubation soils trended into quadrant A (**Figures 2, 3**). A trajectory into quadrant A, however, was observed by Rovira et al. (2008) during litter decomposition (**Figure 4**). Litter decay products were characterized by increased ED and decreased DSC-T50 (the temperature at which half of the total exothermic energy has been released) relative to fresh litter. For all of the soils included in this study as well as the soils, soil density fractions, and litter of Williams et al. (2018), DSC-T50 is positively correlated with E_a ($n = 254$; $R^2 = 0.6$, $p < 0.0001$). Thus, E_a is expected to decrease with decreasing DSC-T50 ($E_a = 0.25 \times \text{DSC-T50} - 27.55$) and this relationship also reinforces the usage of DSC-T50 as a measure of SOM decomposability as utilized by Rovira et al. (2008), Plante et al. (2011), Peltre et al. (2013), and Leifeld and von Lützow (2014). The native forest/pasture soils were characterized by the highest carbon contents (**Table 1**) and it is possible that greater litter inputs into these soils compared to the grassland/cultivated soils caused them to respond to the incubation in the same way as litter, with increasing ED.

In an expanded conceptual bioenergetics framework (**Figure 4**), we combine the results of this study, the litter decomposition study of Rovira et al. (2008), and the soil



density fractionation study of Williams et al. (2018) and summarize proposed bioenergetic changes during SOM formation, decomposition, and stabilization. Litter decomposes into relatively more energy-dense and biodegradable decay products (Rovira et al., 2008), plotting in quadrant A relative to the fresh litter. We expect that pyrogenic organic matter derived from litter would have a higher degree of molecular condensation and thus higher E_a (Harvey et al., 2012). The ED of pyrogenic organic matter would depend on the fire/charring intensity, as Harvey et al. (2016) demonstrated that chars produced at higher temperatures were characterized by lower net energies. However, some litter included in the study by Williams et al. (2018) contained visible pyrogenic organic matter and were from an energetics viewpoint indistinguishable from non-pyrogenic containing litter, thus pyrogenic organic matter is not explicitly depicted in **Figure 4**. When bulk soil is separated into particulate organic matter (represented by the LF from density fractionation) and mineral-associated organic matter (represented by the HF from density fractionation) pools, particulate SOM is more energy dense and requires greater energy input than bulk SOM or mineral-associated SOM. Particulate SOM is also likely composed to some extent of litter decay products or litter; hence, they are similar in ED and E_a in the framework. Mineral-associated SOM is comprised of the mineral matrix and the adsorbed or chemically complexed SOM, the former increasing the required energy input of the latter due to the additional bonds of SOM sorption onto mineral surfaces.

As the bulk soil decomposes, there is a net loss of the more labile particulate SOM, bringing the energetic signature of the bulk soil closer to that of the mineral-associated SOM. The mineral matrix exists at the hypothesized point of zero ED and zero E_a , as it offers no SOM for microbial decomposition and thus would yield no CO_2 . With continued decomposition, the energetic benefit to cost ratio continues to decrease, resulting in relatively thermodynamically unstable SOM being the most persistent.

CONCLUSION

By analyzing the energetics of a suite of soils from field and laboratory experiments designed to isolate persistent SOM within the proposed bioenergetics framework, this study has demonstrated that the signature of persistent SOM is that of decreasing available energy supply (ED) and of decreasing required energy input (E_a). This finding contradicts soil formation theories that propose that persistent SOM is formed by the polymeric condensation of decomposition products into large and stable biopolymers (humus), and instead supports theories that persistent SOM is comprised of smaller and smaller particulate SOM fragments and biomolecules, preserved due to interaction with soil minerals. SOM stabilization mechanisms, such as physical and chemical protection, can be viewed as energetic barriers to decomposition such that the stabilized SOM requires additional energies from the microbial community to

access. In this framework, we propose that this stabilized SOM is also less biodegradable than labile SOM because of its decreased energy contents. The conclusions of this framework should not be applied to carbonate soils as thermal decomposition of carbonate mineral may result in extraneous CO₂ incorporated into the calculation of E_a. The variable or increased ED and E_a exhibited by the laboratory experimental soils suggests that these methods do not always effectively isolate environmentally persistent SOM, but in some cases may target physically or chemically protected SOM.

AUTHOR CONTRIBUTIONS

EW conceived the conceptual framework, performed the thermal analyses, analyzed the data, and wrote the manuscript with input and contributions from AP.

REFERENCES

- Amenabar, M. J., Shock, E. L., Roden, E. E., Peters, J. W., and Boyd, E. S. (2017). Microbial substrate preference dictated by energy demand rather than supply. *Nat. Geosci.* 10, 577–581. doi: 10.1038/Ngeo2978
- Balabane, M., and Plante, A. F. (2004). Aggregation and carbon storage in silty soil using physical fractionation techniques. *Eur. J. Soil Sci.* 55, 415–427. doi: 10.1111/j.1351-0754.2004.0608.x
- Balesdent, J. (1996). The significance of organic separates to carbon dynamics and its modelling in some cultivated soils. *Eur. J. Soil Sci.* 47, 485–493. doi: 10.1111/j.1365-2389.1996.tb01848.x
- Barré, P., Plante, A. F., Cecillon, L., Lutfalla, S., Baudin, F., Bernard, S., et al. (2016). The energetic and chemical signatures of persistent soil organic matter. *Biogeochemistry* 130, 1–12. doi: 10.1007/s10533-016-0246-0
- Bond-Lamberty, B., and Thomson, A. (2010). Temperature-associated increases in the global soil respiration record. *Nature* 464, 579–582. doi: 10.1038/nature08930
- Bruun, S., Thomsen, I. K., Christensen, B. T., and Jensen, L. S. (2008). In search of stable soil organic carbon fractions: a comparison of methods applied to soils labelled with C-14 for 40 days or 40 years. *Eur. J. Soil Sci.* 59, 247–256. doi: 10.1111/j.1365-2389.2007.00985.x
- Burnham, A. K., and Braun, R. L. (1999). Global kinetic analysis of complex materials. *Energy Fuels* 13, 1–22. doi: 10.1021/ef9800765
- Currie, W. S. (2003). Relationships between carbon turnover and bioavailable energy fluxes in two temperate forest soils. *Glob. Change Biol.* 9, 919–929. doi: 10.1046/j.1365-2486.2003.00637.x
- Davidson, E. A., and Janssens, I. A. (2006). Temperature sensitivity of soil carbon decomposition and feedbacks to climate change. *Nature* 440, 165–173. doi: 10.1038/nature04514
- Dembecki, H. (1994). The effects of the mineral matrix on the determination of kinetic parameters using modified rock-aval pyrolysis - reply. *Organ. Geochem.* 21, 982–984.
- Falkowski, P. G., Fenchel, T., and Delong, E. F. (2008). The microbial engines that drive Earth's biogeochemical cycles. *Science* 320, 1034–1039. doi: 10.1126/science.1153213
- Falloon, P. D., and Smith, P. (2000). Modelling refractory soil organic matter. *Biol. Fertil. Soils* 30, 388–398.
- Greenfield, L. G., Gregorich, E. G., van Kessel, C., Baldock, J. A., Beare, M. H., Billings, S. A., et al. (2013). Acid hydrolysis to define a biologically-resistant pool is compromised by carbon loss and transformation. *Soil Biol. Biochem.* 64, 122–126. doi: 10.1016/j.soilbio.2013.04.009
- Gregorich, E. G., Gillespie, A. W., Beare, M. H., Curtin, D., Sanei, H., and Yanni, S. F. (2015). Evaluating biodegradability of soil organic matter by its thermal stability and chemical composition. *Soil Biol. Biochem.* 91, 182–191. doi: 10.1016/j.soilbio.2015.08.032

FUNDING

Funding to support this project was provided by the University of Pennsylvania Vice Provost for Research – Postdoctoral Fellowship for Academic Diversity and the National Science Foundation (EAR 1541588).

ACKNOWLEDGMENTS

We thank Pierre Barré and Folk van Oort for supplying samples from the Long-Term Bare Fallow network, Mike Beare and Denis Curtin from Landcare New Zealand for supplying samples from the chemical fallow experiments, and Michelle Haddix and Rich Conant for supplying samples from the land-use conversion/MAT transect.

- Guo, L. B., and Gifford, R. M. (2002). Soil carbon stocks and land use change: a meta analysis. *Glob. Change Biol.* 8, 345–360. doi: 10.1046/j.1354-1013.2002.00486.x
- Haddix, M. L., Plante, A. F., Conant, R. T., Six, J., Steinweg, J. M., Magrini-Bair, K., et al. (2011). The role of soil characteristics on temperature sensitivity of soil organic matter. *Soil Sci. Soc. Am. J.* 75, 56–68. doi: 10.2136/sssaj2010.0118
- Harvey, O. R., Kuo, L. J., Zimmerman, A. R., Louchouart, P., Amonette, J. E., and Herbert, B. E. (2012). An index-based approach to assessing recalcitrance and soil carbon sequestration potential of engineered black carbons (Biochars). *Environ. Sci. Technol.* 46, 1415–1421. doi: 10.1021/es2040398
- Harvey, O. R., Myers-Pigg, A. N., Kuo, L. J., Singh, B. P., Kuehn, K. A., and Louchouart, P. (2016). Discrimination in degradability of soil pyrogenic organic matter follows a return-on-energy-investment principle. *Environ. Sci. Technol.* 50, 8578–8585. doi: 10.1021/acs.est.6b01010
- Jenkinson, D. S. (1971). Studies on decomposition of C14 labelled organic matter in soil. *Soil Sci.* 111, 64–70. doi: 10.1097/00010694-197101000-00008
- Jobbágy, E. G., and Jackson, R. B. (2000). The vertical distribution of soil organic carbon and its relation to climate and vegetation. *Ecol. Appl.* 10, 423–436.
- Kiem, R., Knicker, H., Korschens, M., and Kogel-Knabner, I. (2000). Refractory organic carbon in C-depleted arable soils, as studied by C-13 NMR spectroscopy and carbohydrate analysis. *Organ. Geochem.* 31, 655–668. doi: 10.1016/S0146-6380(00)00047-4
- Kleber, M., Nico, P. S., Plante, A. F., Filley, T., Kramer, M., Swanston, C., et al. (2011). Old and stable soil organic matter is not necessarily chemically recalcitrant: implications for modeling concepts and temperature sensitivity. *Glob. Change Biol.* 17, 1097–1107. doi: 10.1111/j.1365-2486.2010.02278.x
- Krull, E. S., Swanston, C. W., Skjemstad, J. O., and McGowan, J. A. (2006). Importance of charcoal in determining the age and chemistry of organic carbon in surface soils. *J. Geophys. Res. Biogeosci.* 111:G04001.
- Lehmann, J., and Kleber, M. (2015). The contentious nature of soil organic matter. *Nature* 528, 60–68. doi: 10.1038/nature16069
- Leifeld, J., and von Lütow, M. (2014). Chemical and microbial activation energies of soil organic matter decomposition. *Biol. Fertil. Soils* 50, 147–153. doi: 10.1007/s00374-013-0822-6
- Odum, E. P., Connell, C. E., and Davenport, L. B. (1962). Population energy-flow of 3 primary consumer components of old-field ecosystems. *Ecology* 43, 88–96. doi: 10.2307/1932043
- Paul, E. A., Morris, S. J., Conant, R. T., and Plante, A. F. (2006). Does the acid hydrolysis-incubation method measure meaningful soil organic carbon pools? *Soil Sci. Soc. Am. J.* 70, 1023–1035. doi: 10.2136/sssaj2005.0103
- Peltre, C., Fernandez, J. M., Craine, J. M., and Plante, A. F. (2013). Relationships between biological and thermal indices of soil organic matter stability differ with soil organic carbon level. *Soil Sci. Soc. Am. J.* 77, 2020–2028. doi: 10.2136/sssaj2013.02.0081

- Plante, A. F., Conant, R. T., Paul, E. A., Paustian, K., and Six, J. (2006). Acid hydrolysis of easily dispersed and microaggregate-derived silt- and clay-sized fractions to isolate resistant soil organic matter. *Eur. J. Soil Sci.* 57, 456–467. doi: 10.1111/j.1365-2389.2006.00792.x
- Plante, A. F., Fernandez, J. M., Haddix, M. L., Steinweg, J. M., and Conant, R. T. (2011). Biological, chemical and thermal indices of soil organic matter stability in four grassland soils. *Soil Biol. Biochem.* 43, 1051–1058. doi: 10.1016/j.soilbio.2011.01.024
- Plante, A. F., Fernandez, J. M., and Leifeld, J. (2009). Application of thermal analysis techniques in soil science. *Geoderma* 153, 1–10. doi: 10.1016/j.geoderma.2009.08.016
- Plante, A. F., Pernes, M., and Chenu, C. (2005). Changes in clay-associated organic matter quality in a C depletion sequence as measured by differential thermal analyses. *Geoderma* 129, 186–199. doi: 10.1016/j.geoderma.2004.12.043
- Rovira, P., Kurz-Besson, C., Couteaux, M. M., and Vallejo, V. R. (2008). Changes in litter properties during decomposition: a study by differential thermogravimetry and scanning calorimetry. *Soil Biol. Biochem.* 40, 172–185. doi: 10.1016/j.soilbio.2007.07.021
- Ruhlmann, J. (1999). A new approach to estimating the pool of stable organic matter in soil using data from long-term field experiments. *Plant Soil* 213, 149–160. doi: 10.1023/A:1004552016182
- Rumpel, C., and Kögel-Knabner, I. (2011). Deep soil organic matter—a key but poorly understood component of terrestrial C cycle. *Plant Soil* 338, 143–158. doi: 10.1007/s11104-010-0391-5
- Schmidt, M. W., Torn, M. S., Abiven, S., Dittmar, T., Guggenberger, G., Janssens, I. A., et al. (2011). Persistence of soil organic matter as an ecosystem property. *Nature* 478, 49–56. doi: 10.1038/nature10386
- Stockmann, U., Adams, M. A., Crawford, J. W., Field, D. J., Henakaarchchi, N., Jenkins, M., et al. (2013). The knowns, known unknowns and unknowns of sequestration of soil organic carbon. *Agric. Ecosyst. Environ.* 164, 80–99.
- Stone, M. M., and Plante, A. F. (2015). Relating the biological stability of soil organic matter to energy availability in deep tropical soil profiles. *Soil Biol. Biochem.* 89, 162–171. doi: 10.1016/j.soilbio.2015.07.008
- Trumbore, S. E., Vogel, J. S., and Southon, J. R. (1989). Ams C-14 measurements of fractionated soil organic-matter - an approach to deciphering the soil carbon-cycle. *Radiocarbon* 31, 644–654.
- Williams, E. K., Fogel, M. L., Berhe, A. A., and Plante, A. F. (2018). Distinct bioenergetic signatures in particulate versus mineral-associated soil organic matter. *Geoderma* 330, 107–116. doi: 10.1016/j.geoderma.2018.05.024
- Williams, E. K., Rosenheim, B. E., McNichol, A. P., and Masiello, C. A. (2014). Charring and non-additive chemical reactions during ramped pyrolysis: applications to the characterization of sedimentary and soil organic material. *Organ. Geochem.* 77, 106–114. doi: 10.1016/j.orggeochem.2014.10.006

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Williams and Plante. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.