



Back to the Origin: *In Situ* Studies Are Needed to Understand Selection during Crop Diversification

Yolanda H. Chen¹, Lori R. Shapiro^{2,3}, Betty Benrey⁴ and Angélica Cibrián-Jaramillo^{5*}

¹ Department of Plant and Soil Science, University of Vermont, Burlington, VT, United States, ² Department of Microbiology and Immunology, Harvard Medical School, Boston, MA, United States, ³ Department of Applied Ecology, North Carolina State University, Raleigh, NC, United States, ⁴ Laboratory of Evolutionary Entomology, Institute of Biology, University of Neuchâtel, Neuchâtel, Switzerland, ⁵ Laboratorio Nacional de Genómica para la Biodiversidad, Centro de Investigación y de Estudios Avanzados, Irapuato, Mexico

OPEN ACCESS

Edited by:

Alejandro Casas, Instituto de Investigaciones en Ecosistemas y Sustentabilidad, Universidad Nacional Autónoma de México, Mexico

Reviewed by:

Shabir Hussain Wani, Michigan State University, United States Mukhtar Ahmed, Pir Mehr Ali Shah Arid Agriculture University, Pakistan

> *Correspondence: Angélica Cibrián-Jaramillo angelica.cibrian@gmail.com

Specialty section:

This article was submitted to Agroecology and Land Use Systems, a section of the journal Frontiers in Ecology and Evolution

> Received: 24 June 2017 Accepted: 25 September 2017 Published: 18 October 2017

Citation:

Chen YH, Shapiro LR, Benrey B and Cibrián-Jaramillo A (2017) Back to the Origin: In Situ Studies Are Needed to Understand Selection during Crop Diversification. Front. Ecol. Evol. 5:125. doi: 10.3389/fevo.2017.00125 Crop domestication has been embraced as a model system to study the genetics of plant evolution. Yet, the role of the environment, including biotic forces such as microbial and insect communities, in contributing to crop phenotypes under domestication and diversification has been poorly explored. In particular, there has been limited progress in understanding how human selection, agricultural cultivation (soil disturbance, fertilization, and irrigation), and biotic forces act as selective pressures on crop phenotypes. For example, geographically-structured pathogenic, pestiferous, and mutualistic interactions with crop plants have likely given rise to landraces that interact differently with local microbial and insect communities. In order to understand the adaptive role of crop traits, we argue that more studies should be conducted in the geographic centers of origin to test hypotheses on how abiotic, biotic, and human selective forces have shaped the phenotypes of domesticated plants during crop domestication and subsequent diversification into landraces. In these centers of origin, locally endemic species associated with wild ancestors have likely contributed to the selection on plant phenotypes. We address a range of questions that can only be studied in the geographic center of crop origin, placing emphasis on Mesoamerican polyculture systems, and highlight the significance of in situ studies for increasing the sustainability of modern agricultural systems.

Keywords: crop domestication, agroecology, evolutionary ecology, biogeography, epigenetics, human culture, insects, microbes

INTRODUCTION

The domestication of crop plants has fundamentally altered the relationship between humans and their environment (Larson et al., 2014). While the genetics of crops domestication has been widely studied for some common plant species (Darwin, 1868; Evans, 1993; Smartt and Simmonds, 1995; Ladizinsky, 1998; Hancock, 2012), the role of ecological interaction within centers of origin in contributing crop phenotypic diversity has been overlooked (Chen et al., 2015; Perez-Jaramillo et al., 2016). Prior to domestication, wild ancestors of crop plants evolved in association with a broad assemblage of microbes and insects, with which they engaged in a range of pathogenic,

1

predatory, commensal, and mutualistic interactions (Chen et al., 2013; Huang et al., 2016; Perez-Jaramillo et al., 2016). These ecological interactions were almost certainly altered by domestication and when early domesticates were introduced to new locations with unique climates, distinct local biodiversity, and different cultural methods of farming.

The majority of domestication events occurred in specific geographic regions, often within the native range of their wild ancestors (Vavilov, 1926, 1951; Meyer et al., 2012). Despite this, most studies rarely consider whether all interacting species are endemic in the center of origin. We found that only 1.6% of 1,532 studies comparing insect responses on wild and crop plants specifically accounted for biogeographical history (Chen et al., 2015). Geographically-explicit hypotheses are needed to understand in situ crop diversification for two reasons. First, human-mediated migration of crops to new regions within centers of origin influenced the genetic structuring of crop populations. Second, domesticated cultivars experienced novel selective pressures imposed by new environments and the cultural preferences of different indigenous peoples (Figure 1; Brush, 1995; Hugo et al., 2003).

In situ field studies documenting variation of ecological interactions are important to determine the extent to which landrace phenotypes respond to local adaptation and artificial selection. After initial crop domestication, early landraces were brought to new environments that were often different from the ecological conditions experienced by their wild progenitors (Hufford et al., 2012). In these new environments, landraces interacted with new species, different cropping system, and new abiotic conditions (Figure 1). Also, different groups of farmers may cultivate the same crop in different polyculture systems (Casas et al., 1996, 2007; Hugo et al., 2003), which is the simultaneous cultivation of multiple crops. As a result, landraces emerge over thousands of years due to natural selection exerted by local abiotic conditions, local insect, and microbial communities, as well as human selection on traits related to ease of cultivation, aesthetics, taste, and cultural preferences (Perales et al., 2005; Brush and Perales, 2007; Casas et al., 2007; Aguirre-Dugua et al., 2013).

Here, we discuss the factors that contribute to phenotypic variation in crops and landraces. We examine the role of human selection and niche construction activities on crop phenotypic plasticity, a major factor in local adaptation (Piperno, 2017). We conceptually address two questions that could deepen our understanding of crop evolution and local adaptation: (1) To what extent are crop plants locally adapted? and (2) What are the relative roles of human selection, human-mediated migration, the local abiotic environment, endemic biotic communities, and cultivation practices in the diversification of crops into landraces? We describe polyculture systems in Mesoamerica as a suitable model in which these questions could be pursued. Ultimately, whether plants can maintain historic beneficial interactions with associated species-or form new ones in their introduced regions-have important implications for the future sustainability of agriculture.

GENETIC AND ENVIRONMENTAL CONTRIBUTIONS TO CROP PHENOTYPIC VARIATION

We focus on landraces as a natural experimental system to understand how natural and human selective forces have shaped the diversity of phenotypic traits of domesticated crop plants during local adaptation. In order to detect local adaptation, it is important to characterize the extent of variation and fitness of phenotypic traits in response to local selective pressures (Kawecki and Ebert, 2004). Variation in landrace phenotypes is affected by their genotype, local environmental variation, and plant phenotypic plasticity. Equation 1 describes how the components of phenotypic variation can be partitioned (Pigliucci, 2001):

$$V_P = V_G + V_E + V_{G x E} \tag{1}$$

Where V_p denotes the total phenotypic variation found for a population or subspecies, V_G is the total genetic variation, V_E is the total environmental variation, and V_{GxE} is the genotype \times environment interaction. We limit our treatment of V_p and V_G because they have been discussed elsewhere (Olsen and Wendel, 2013; Piperno, 2017). In contrast, there has been a minimal effort to understand the contributions of V_E and V_{GxE} to crop domestication and diversification. **Table 1** provides a set of examples illustrating the range of plant adaptations to abiotic, biotic, and human selection.

Total genetic variation (V_G) is influenced by evolutionary forces including selection, genetic drift, inbreeding, and gene flow (Hartl and Clark, 2006), which likely differ between wild progenitor and crop populations. Domesticated crops have reduced effective population sizes and are under intense selection for a suite of traits favored by humans (Doebley et al., 2006; Moreno et al., 2006). Cultural factors can also structure landrace populations and dispersal patterns, as people in different ethnolinguistic groups are less likely to trade seeds (Orozco-Ramírez et al., 2016).

The contribution of the environment to phenotypic variation (V_E) strongly differs between wild progenitors and domesticated crops grown in human constructed agricultural niches. In natural systems, V_E is affected by local abiotic conditions such as climate, precipitation, soil type, and nutrient availability. Domesticated crops are found in a range of cultivation systems, where the purpose of cultivation is to lessen the unpredictability of the local environment in order to favor uniform plant growth and yield. Cultivation has been shown to strongly affect plant-associated microbial and insect communities (Berg, 2009; Berendsen et al., 2012; Chen et al., 2013). Surprisingly, the majority of the insect and microbial species associated with a given crop in its center of origin remain undescribed (Chen et al., 2015; Coleman-Derr et al., 2016). Given that plant-biotic interactions can influence plant phenotypes (Henning et al., 2016), human niche activities can indirectly influence crop phenotypes by influencing the diversity and community structure of biotic assemblages.

Genotype by environment interactions (V_{GxE}) describe phenotypic plasticity, or how genotypes responds to a range of environmental conditions. Crops respond to agricultural



conditions differently from their wild ancestors in growth (**Table 1**). Also, plasticity itself may have been constrained by domestication. Wild progenitors may have retained a greater plasticity to unpredictable environments, whereas domesticated may have lost phenotypic plasticity. Growing plants in different environments can provide important insight on the origin of important traits associated with domestication. For instance, teosinte grown under historic Pleistocene conditions (atmospheric CO_2 and temperature) displayed maize-like phenotypes such as reduced tillering, uniform seed maturation, and bract-less seeds (Piperno et al., 2015). Therefore, hypotheses that account for environmental variation and the selective environment can provide important insight on how crop species adapted to divergent environmental conditions.

Crop responses to the environment can be heritable, independent of changes in the DNA sequence, which contributes another dimension to V_{GxE} . An emerging frontier is to understand the role of transgenerational epigenetics in crop adaptation to local environmental conditions (Piperno, 2017). Epigenetics studies can reveal how the environment can shape heritable gene expression (and thus the phenotype) without changing the underlying DNA sequence (Jablonka and Raz, 2009; Laland et al., 2014). Epigenetic changes in DNA methylation can regulate gene expression patterns during stress, which can influence the ability of crop varieties to respond to local environments (Ferreira et al., 2015). Therefore, *in situ* studies are critically needed to test the relative roles of genetics, epigenetics, and the environment in contributing to crop diversification.

MESOAMERICA AS A FIELD LABORATORY FOR *IN SITU* STUDIES

Centers of domestication offer field locations to understand how local environments and human cultural influences contributed to crop domestication and diversification into landraces. The legendary Russian botanist Nikolai Vavilov delineated eight geographic regions as "centers of domestication" where multiple crop species were domesticated (Vavilov, 1926, 1951). One such center is Mesoamerica, which is the region of origin for maize, beans, squash, peppers, avocado, vanilla, and thousands of non-commercialized plant species (Casas et al., 2007). Crop domestication in Mesoamerica has received far less attention than in other centers of origin (Casas et al., 2007). Mesoamerica is particularly well-suited for field studies on crop domestication and diversification for several reasons: (1) It hosts a dazzling array of landrace varieties that are often cultivated sympatrically with their wild ancestors, (2) It has an archeological record confirming **TABLE 1** | Phenotypic variation (V_p) in wild progenitors and landraces can be attributed to genetic variation (V_G), environmental variation (V_E), and variation attributed to genotype by environment interactions (V_{GxE}).

	Phenotypic variation	Genetic variation	Environmental variation	Genotype × environmental variation
Abiotic	Wild emmer, bean, and lupine populations show adaptations to drought (Peleg et al., 2005; Cortés et al., 2013; Berger et al., 2017).	Drought-resistant wild emmer populations differ in gene expression in stress, transport, and metabolism genes (Huang et al., 2016).	Precipitation is a major factor explaining the wider geographic range of the domesticated Mesoamerican fruit tree, <i>Spondias</i> <i>purpurea</i> (Anacardiaceae), compared to its wild progenitor (Miller and Knouft, 2006).	Teosinte growth and a high affinity nitrate transport gene were more highly inducible under low nitrogen availability (Gaudin et al., 2011).
	The common bean, <i>Phaseolus vulgaris</i> , shows higher phosophorous efficiency than wild <i>P. vulgaris</i> populations (Araújo et al., 1997).	Geographic distribution of genetic diversity in historical barley landraces shows local adaptation to latitude (Aslan et al., 2015).	Temperature plays a major role in structuring the distribution of four early landraces of maize (Hufford et al., 2012).	Teosinte grown under Pleistocene-like climatic conditions produced maize-like phenotypes (Piperno et al., 2015).
Biotic	Wild emmer wheat (<i>Triticum</i> <i>turgidum</i> spp. <i>dicoccoides</i>) and chili show geographic variation in disease resistance (Tewksbury et al., 2008; Huang et al., 2016).	Wild watermelon roots are resistant to root-knot nematode damage while cultivated species are susceptible (Thies et al., 2016).	Irrigation increases susceptibility to fungal pathogens and increases the risk of fungal pathogen emergence (Stukenbrock and McDonald, 2008).	Wheat genotypes differ in their plasticity in terms of yield and suppression of aphid population growth (Tétard-Jones and Leifert, 2011).
	The microbiome of domesticated barley differs from wild barley (Bulgarelli et al., 2015).	The 6H chromosome of barley, <i>Hordeum vulgare</i> , is coevolving with the net form <i>Pyrenophora teres f.</i> <i>teres</i> but not the spot form of barley net blotch (Rau et al., 2015).	Cultivated chickpea is extremely susceptible to Ascochyta blight in the Near East in the fall, which selected for spring sowing of chickpea (Abbo et al., 2007).	Domesticated sunflowers respond to nitrogen inputs by developing larger flower heads, which leads to higher sunflower moth, <i>Homeosoma</i> <i>electellum</i> , herbivory but lower parasitism (Chen and Welter, 2005).
Human selection	Breadfruit, <i>Artocarpus altils</i> in Oceania are seedless and dependent upon humans for dispersal (Xing et al., 2012). Domesticated lupines, <i>Lupinus</i> spp., set seed earlier, produce larger seeds than wild lupines (Berger et al., 2017). In Cassava, <i>Manihot esculenta</i> , levels of toxic cyanogenic glucosides are dependent upon local preferences and knowledge on processing (McKey et al., 2010).	A SNP mutation in the promoter region of a P450 gene, CYP78A increases fruit size and reduces branching in tomato plants (Chakrabarti et al., 2013). Several species of domesticated cucurbits show signs of convergent evolution in the interruption of the cucurbitacin biosynthesis pathway (Zhou et al., 2016).	Insect communities are less diverse and show a shift in community composition in cultivated rice agroecosystems compared to wild rice (Chen et al., 2013). Agave grown in cultivated soils have lower diversity of soil prokaryotes in the rhizosphere than those in native soils (Coleman-Derr et al., 2016).	Domesticated chickpea, <i>Cicer</i> <i>arietinum</i> , have been selected to be insensitive to veralization compared to their wild ancestor, <i>Cicer reticulatum</i> (Pinhasi van-Oss et al., 2016). Maize shows lower tiller plasticity and lower nitrogen use efficiency compared to teosinte. Under low nitrogen, maize plants have lower leaf size (Gaudin et al., 2011). The common bean, <i>Phaseolus</i> <i>vulgaris</i> , shows higher phosophorous efficiency than wild <i>P. vulgaris</i> populations (Araújo et al., 1997).

Wild progenitors and domesticated crops display adaptive responses to abiotic, biotic, and human selection. Environmental variation (V_E) can be naturally occurring or arise from human niche construction activities. Wild progenitors and domesticated crops show different responses to the habitat conditions within which they evolved (V_{GxE}).

a long history of interactions between humans and many species of crop plants, and (3) Indigenous peoples continue oral and cultural traditions associated with the cultivation of these plants (Gepts, 2004; Staller et al., 2006; Piperno et al., 2007).

We focus on Mexico, the largest country within Mesoamerica, where much of the phenotypic and genotypic variation underlying local adaptation to environments has not been characterized. Many suspected centers of domestication and regions with wild relatives remain poorly explored for most crop plants. These underexplored regions that are historically relevant for the study of crop diversification include: the Balsas River Valley (Piperno et al., 2007), the Gulf Coast (Kraft et al., 2014), and the Tehuacan Valley for species other than maize (Vallejo et al., 2016). Needless to say, there are considerable opportunities to examine the selective forces that produced the extant array of landraces for native Mesoamerican crop plant species.

In Mesoamerica, traditional agroecosystems have been maintained cohesively for hundreds to thousands of years by indigenous peoples. Different cropping systems dominate in different climatic regions (**Figure 1**). In the Yucatan Peninsula, home gardens are highly diverse polyculture systems, and include crops such as avocado, annona, and papaya (Moreno-Calles et al., 2016). The inland and coastal regions are dominated by agroforestry systems paired with ornamental and woody species (Moreno-Calles et al., 2016). The oldest Mesoamerican polyculture systems continue to be maintained in Tlaxcala (Gonzalez-Jacome, 2016). In cold and dry highland environments, prehispanic terraces for water management still exist in Oaxaca and the Tehuacan Valley (Donkin, 1979). In

arid environments, drought-tolerant plants including cactus are cultivated in polyculture with chili pepper and other crops (Moreno-Calles et al., 2012, 2016). Slash and burn agriculture is the most widespread form of cultivation in the tropical deciduous and temperate forests, where crops are rotated after the plant cover is burned every few years. Slash and burn systems can be quite diverse, with at least 57 tree species from 33 plant families and many more herbaceous species (Moreno-Calles et al., 2016).

One of the most dominant cropping system in slash and burn agriculture is the milpa (Figure 1D), which is, at its most basic, the joint cultivation of maize, beans, squash (Zizumbo-Villarreal and Colunga-GarcíaMarín, 2010). Maize serves as a trellis for beans that fix nitrogen, while squash suppresses weeds. Although, the wild progenitors of maize, beans, and squash are native to separate regions in Mexico (Smith, 1997; Gepts, 2004; Piperno et al., 2007), these core milpa crops have been cultivated together for thousands of years. It is highly possible that associated microbes and insects have adapted to the milpa. Perhaps it should not be a surprise that the most devastating contemporary native insect pests of maize can all be collected from squash flowers (Metcalf and Lampman, 1989), and that several closely related species of leaf beetles in the genus Diabrotica damage maize, beans, and squash (Clark et al., 2001; Vidal et al., 2005; Eben and Espinosa de Los Monteros, 2013). There are many other unexplored questions on crops and biotic interactions in milpa systems such as: (1) To what extent have microbiomes, pathogens, mycorrhizae, herbivores, natural enemies, and pollinators adapted to polyculture and landrace traits? (2) Does intercropping select landraces to develop complementary use for light, nutrients, and water? and (3) How has the *milpa* shaped landrace phenotypes?

The present Mesoamerican landscape is also a gradient of ecological contexts where one could study whether genes underlie phenotypes that are adaptive to local abiotic and biotic conditions. Reciprocal common garden studies with maize have found that highland landraces show higher fitness and seed quality in highland conditions, while lowland landraces have higher fitness in mid-altitude locations (Mercer et al., 2008). Traits such as pigmentation, stem hair, plant height, and flowering time have been shown to be adaptive to altitude, but completely different genes underlie local adaptation to highland conditions in Mesoamerica and South America (Mercer et al., 2008). Field sampling of Mexican teosinte populations helped to clarify that maize adaptation to the Mexican highlands resulted due to introgression from wild teosinte (Hufford et al., 2013). Understanding the genomic basis of local adaptation in crops relies on multiple in situ localities, where the ecological history can be reconstructed by testing for genomic regions under divergence (Pyhäjärvi et al., 2013), and the responses of candidate genes can be observed in local environments (Doust et al., 2014; Piperno, 2017). Traditional Mesoamerican agroecosystems are living biological and ethnographic systems that are suitable for studying how human-created niches in agroecosystems interact with local biotic and abiotic environments to shape landrace phenotypes. These *in situ* systems provide an important reference for examining how crop plants adapted as they have diversified within centers of origin.

IMPLICATIONS OF *IN SITU* STUDIES FOR SUSTAINABLE AGRICULTURE

Sustainable agriculture aims to reduce reliance on pesticides and fertilizers by utilizing biodiversity to provide ecological services that provision nutrients, protect crops, and enhance yields (Altieri, 1999). For the world's most important crops, the majority of production occurs outside their centers of origin (Khoury et al., 2016). Oftentimes, crops are grown in marginal environments, where they experience low nutrient availability, excess or limited water availability, temperature extremes, or pest outbreaks (Table 1). Under climate change, these pressures are predicted to intensify (Hatfield et al., 2010). Perhaps because centers of origin tend to be the geographic source for the major diseases (Leppik, 1970) and insect pests of crops (Chen, 2016), they are also the source for genes for resistance (Harlan, 1976; Hijmans et al., 2003; Zhang et al., 2017), insect natural enemies (van den Bosch, 1971; van Driesche et al., 2008), and microbes (Philippot et al., 2013; Perez-Jaramillo et al., 2016) that help plants to resist pests and tolerate abiotic stress.

In situ studies can also provide insight on whether human selection for crop yield is fundamentally at odds with traits that mediate beneficial plant-biotic interactions. First, crop domestication appears to have promoted pests more frequently than beneficial species, especially for economically-important traits such as fruit size and seed size (Chen et al., 2015). However, we do not know if direct trade-offs between yield and pest resistance exist, and whether this relationship may vary with different landraces and environments within centers of origin. The diversity and community structure of microbes and insects associated with wild ancestors and landraces have been inadequately described (Chen et al., 2013; Perez-Jaramillo et al., 2016), and geographic variation in patterns of biodiversity within centers of origin remain unexplored.

Second, plant genotypes vary in their ability to form positive relationships with beneficial species (Table 1; Smith and Goodman, 1999; Chen and Welter, 2005; Tamiru et al., 2011). Determining the relative roles of plant genetic diversity, microbial associates, and plant gene × environment interactions in conferring resistance to biotic and abiotic stresses (Philippot et al., 2013) would help elucidate whether breeding, microbial inoculation strategies, or natural enemy introductions would better support crop production in the diverse environments where crops are grown. Finally, in situ studies provide insight on the ecological function of crop genes and metabolites within their natural environment, which are oftentimes only explored in environments far from centers of origin. For examples, teosinte and some maize varieties emit the sesquiterpene (E)β-caryophyllene, which attracts entomopathogenic nematodes (Rasmann et al., 2005) and parasitoids (Kollner et al., 2008) in Europe. However, the role of this compound in landraces is not known, especially in Mesoamerica, where a diverse assemblage of species may be adapted to respond to plant signals. In situ studies can help resolve whether landrace varieties produce the compound, natural enemies are attracted to it, and whether breeding for (E)- β -caryophyllene would increase natural enemy attraction and enhance yield in the diverse worldwide locations where maize is now grown.

CONCLUSIONS

In situ ecological studies are an essential, but almost completely unexplored line of inquiry for evolutionary ecologists to understand the selective forces that contribute to local adaptation of landrace varieties. As one of the major centers of crop origin, Mesoamerica is an ideal location for in situ studies, because wild progenitors can be found growing sympatrically with domesticated landrace varieties cultivated in traditional polyculture systems. For many crops and cultivation systems, the unique combination of local abiotic, biotic, and cultural selective forces that shaped variation in crop phenotypes during domestication and diversification continue to coexist. We advocate that geographically-explicit studies will yield new insight into how selection from humans and the local environment contribute to landrace diversification and local adaptation. Such knowledge is immediately applicable toward understanding the capacity of crop plants to respond to the biotic

REFERENCES

- Abbo, S., Frenkel, O., Sherman, A., and Shtienberg, D. (2007). The sympatric Ascochyta pathosystems of Near Eastern legumes, a key for better understanding of pathogen biology. *Eur. J. Plant Pathol.* 119, 111–118. doi: 10.1007/s10658-007-9116-x
- Aguirre-Dugua, X., Pérez-Negrón, E., and Casas, A. (2013). Phenotypic differentiation between wild and domesticated varieties of *Crescentia cujete L.* and culturally relevant uses of their fruits as bowls in the Yucatan Peninsula, Mexico. J. Ethnobiol. Ethnomed. 9:76. doi: 10.1186/1746-4269-9-76
- Altieri, M. A. (1999). The ecological role of biodiversity in agroecosystems. Agric. Ecosyst. Environ. 74, 19–31. doi: 10.1016/S0167-8809(99)00028-6
- Araújo, A. P., Teixeira, M. G., and Almeida, D. L. (1997). Phosphorus efficiency of wild and cultivated genotypes of common bean (*Phaseolus* vulgaris) under biological nitrogen fixation. Soil Biol. Biochem. 29, 951–957. doi: 10.1016/S0038-0717(96)00217-9
- Aslan, S., Forsberg, N. E. G., Hagenblad, J., and Leino, M. W. (2015). Molecular genotyping of historical barley landraces reveals novel candidate regions for local adaption. *Crop Sci.* 55, 2766–2776. doi: 10.2135/cropsci2015.02.0119
- Berendsen, R. L., Pieterse, C. M., and Bakker, P. A. (2012). The rhizosphere microbiome and plant health. *Trends Plant Sci.* 17, 478–486. doi: 10.1016/j.tplants.2012.04.001
- Berg, G. (2009). Plant-microbe interactions promoting plant growth and health: perspectives for controlled use of microorganisms in agriculture. *Appl. Microbiol. Biotechnol.* 84, 11–18. doi: 10.1007/s00253-009-2092-7
- Berger, J. D., Shrestha, D., and Ludwig, C. (2017). Reproductive strategies in Mediterranean legumes: trade-offs between phenology, seed size and vigor within and between wild and domesticated Lupinus species collected along aridity gradients. *Front. Plant Sci.* 8:548. doi: 10.3389/fpls.2017.00548
- Brush, S. B. (1995). *In situ* conservation of landraces in centers of crop diversity. *Crop Sci.* 35, 346–354. doi: 10.2135/cropsci1995.0011183X003500020009x
- Brush, S. B., and Perales, H. R. (2007). A maize landscape: ethnicity and agro-biodiversity in Chiapas Mexico. Agric. Ecosyst. Environ. 121, 211–221. doi: 10.1016/j.agee.2006.12.018
- Bulgarelli, D., Garrido-Oter, R., Munch, P. C., Weiman, A., Droge, J., Pan, Y., et al. (2015). Structure and function of the bacterial root microbiota in wild and domesticated barley. *Cell Host Microbe* 17, 392–403. doi: 10.1016/j.chom.2015.01.011

and abiotic conditions over the vast geographic ranges where they are now grown, and to identify sources of germplasm that might have adaptive traits for crops in their introduced ranges.

AUTHOR CONTRIBUTIONS

YC, LS, and AC conceived the Perspective. YC, AC, LS, and BB drafted the work and revised the content.

FUNDING

We thank our funders for their support: USDA National Institute of Food and Agriculture grant no. #VT-H02301MS to YC, NSF postdoctoral fellowship DBI-1202736 to LS, Swiss National Science Foundation, project No. 31003A_162860 to BB, and CONACyT Problemas Nacionales #247730 to AC.

ACKNOWLEDGMENTS

We thank Nicolas Marguler for the design of Figure 1.

- Casas, A., Otero-Arnaiz, A., Perez-Negron, E., and Valiente-Banuet, A. (2007). *In situ* management and domestication of plants in Mesoamerica. *Ann. Bot.* 100, 1101–1115. doi: 10.1093/aob/mcm126
- Casas, A., del Vázquez, M. C., Viveros, J. L., and Caballero, J. (1996). Plant management among the Nahua and the Mixtec in the Balsas River Basin, Mexico: an ethnobotanical approach to the study of plant domestication. *Hum. Ecol.* 24, 455–478. doi: 10.1007/BF02168862
- Chakrabarti, M., Zhang, N., Sauvage, C., Muños, S., Blanca, J., Cañizares, J., et al. (2013). A cytochrome P450 regulates a domestication trait in cultivated tomato. *Proc. Natl. Acad. Sci. U.S.A.* 110, 17125–17130. doi: 10.1073/pnas.1307313110
- Chen, Y. H. (2016). Crop domestication, global human-mediated migration, and the unresolved role of geography in pest control. *Elem. Sci. Anthr.* 4:106. doi: 10.12952/journal.elementa.000106
- Chen, Y. H., Gols, R., and Benrey, B. (2015). Crop domestication and naturally selected species interactions. *Annu. Rev. Entomol.* 60, 35–58. doi: 10.1146/annurev-ento-010814-020601
- Chen, Y. H., Langellotto, G. A., Barrion, A. T., and Cuong, N. L. (2013). Cultivation of domesticated rice alters arthropod biodiversity and community composition. *Ann. Entomol. Soc. Am.* 106, 100–110. doi: 10.1603/AN12082
- Chen, Y. H., and Welter, S. C. (2005). Crop domestication disrupts a native tritrophic interaction associated with the sunflower, *Helianthus annuus* (Asterales: Asteraceae). *Ecol. Entomol.* 30, 673–683. doi: 10.1111/j.0307-6946.2005.00737.x
- Clark, T. L., Meinke, L. J., and Foster, J. E. (2001). Molecular phylogeny of Diabrotica beetles (Coleoptera: Chrysomelidae) inferred from analysis of combined mitochondrial and nuclear DNA sequences. *Insect Mol. Biol.* 10, 303–314. doi: 10.1046/j.0962-1075.2001.00269.x
- Coleman-Derr, D., Desgarennes, D., Fonseca-Garcia, C., Gross, S., Clingenpeel, S., Woyke, T., et al. (2016). Biogeography and cultivation affect microbiome composition in the drought-adapted plant Subgenus Agave. *New Phytol.* 209, 798–811. doi: 10.1111/nph.13697
- Cortés, A. J., Monserrate, F. A., Ramírez-Villegas, J., Madri-án, S., and Blair, M. W. (2013). Drought tolerance in wild plant populations: the case of common beans (*Phaseolus vulgaris* L.). *PLoS ONE* 8:e62898. doi: 10.1371/journal.pone.0062898
- Darwin, C. (1868). *The Variation of Animals and Plants under Domestication*, Vol. 1, 2. London: John Murray.
- Doebley, J. F., Gaut, B. S., and Smith, B. D. (2006). The molecular genetics of crop domestication. *Cell* 127, 1309–1321. doi: 10.1016/j.cell.2006.12.006

- Donkin, R. A. (1979). Agricultural Terracing in the Aboriginal New World. Flagstaff, AZ: The Wenner-Gren Foundation for Anthropological Research, The University of Arizona.
- Doust, A. N., Lukens, L., Olsen, K. M., Mauro-Herrera, M., Meyer, A., and Rogers, K. (2014). Beyond the single gene: how epistasis and gene-byenvironment effects influence crop domestication. *Proc. Natl. Acad. Sci. U.S.A.* 111, 6178–6183. doi: 10.1073/pnas.1308940110
- Eben, A., and Espinosa de Los Monteros, A. (2013). Tempo and mode of evolutionary radiation in Diabroticina beetles (genera Acalymma, Cerotoma, and Diabrotica). *Zookeys* 207–321. doi: 10.3897/zookeys.332.5220
- Evans, L. T. (1993). Crop Evolution, Adaptation, and Yield. Cambridge: Cambridge University Press.
- Ferreira, L. J., Azevedo, V., Maroco, J., Oliveira, M. M., and Santo, A. P. (2015). Salt tolerant and sensitive rice varieties display differential methylome flexibility under salt stress. *PLoS ONE* 10:e0124060. doi: 10.1371/journal.pone.0124060
- Gaudin, A. C. M., McClymont, S. A., and Raizada, M. N. (2011). The nitrogen adaptation strategy of the wild teosinte ancestor of modern maize, *Zea mays* subsp. parviglumis. *Crop Sci.* 51, 2780–2795. doi: 10.2135/cropsci2010.12.0686
- Gepts, P. (2004). Crop domestication as a long-term selection experiment. *Plant Breed. Rev.* 24, 1–44. doi: 10.1002/9780470650288.ch1
- Gonzalez-Jacome, A. (2016). "Sistemas agrícolas en orografías complejas: las terrazas de Tlaxcala," in *Etnoagroforestería en México*, eds A. I. Moreno-Calles, A. Casas, V. M. Toledo, and M. Vallejo-Ramos (Mexico: UNAM), 111–146.
- Hancock, J. F. (2012). *Plant Evolution and the Origin of Crop Species*. Wallingford, CT; Oxfordshire: CABI.
- Harlan, J. R. (1976). Diseases as a factor in plant evolution. *Annu. Rev. Phytopathol.* 14, 31–51. doi: 10.1146/annurev.py.14.090176.000335
- Hartl, D. L., and Clark, A. G. (2006). *Principles of Population Genetics, 4th Edn.* Sunderland, MA: Sinauer Associates.
- Hatfield, J. L., Boote, K. J., Kimball, B. A., Ziska, L. H., Izaurralde, R. C., Ortf, D., et al. (2010). Climate impacts on agriculture: implications for crop production. *Agron. J.* 103, 351–370. doi: 10.2134/agronj2010.0303
- Henning, J. A., Weston, D. J., Pelletier, D. A., Timm, C. M., Jawdy, S. S., and Classen, A. T. (2016). Root bacterial endophytes alter plant phenotype, but not physiology. *PeerJ* 4:e2606. doi: 10.7717/peerj.2606
- Hijmans, R. J., Jacobs, M., Bamberg, J. B., and Spooner, D. M. (2003). Frost tolerance in wild potato species: assessing the predictivity of taxonomic, geographic, and ecological factors. *Euphytica* 130, 47–59. doi: 10.1023/A:1022344327669
- Huang, L., Raats, D., Sela, H., Klymiuk, V., Lidzbarsky, G., Feng, L., et al. (2016). Evolution and adaptation of wild emmer wheat populations to biotic and abiotic stresses. *Annu. Rev. Phytopathol.* 54, 279–301. doi: 10.1146/annurev-phyto-080614-120254
- Hufford, M. B., Lubinksy, P., Pyhäjärvi, T., Devengenzo, M. T., Ellstrand, N. C., and Ross-Ibarra, J. (2013). The genomic signature of crop-wild introgression in maize. *PLoS Genet*. 9:e1003477. doi: 10.1371/journal.pgen.1003477
- Hufford, M. B., Martínez-Meyer, E., Gaut, B. S., Eguiarte, L. E., Tenaillon, M. I., Cienfuegos, E., et al. (2012). Inferences from the historical distribution of wild and domesticated maize provide ecological and evolutionary insight. *PLoS ONE* 7:e47659. doi: 10.1371/journal.pone.0047659
- Hugo, P. R., Brush, S. B., and Qualset, C. O. (2003). Dynamic management of maize landraces in Central Mexico. *Econ. Bot.* 57, 21–34. doi: 10.1663/0013-0001(2003)057[0021:DMOMLI]2.0.CO;2
- Jablonka, E., and Raz, G. (2009). Transgenerational epigenetic inheritance: prevalence, mechanisms, and implications for the study of heredity and evolution. Q. Rev. Biol. 84, 131–176. doi: 10.1086/598822
- Kawecki, T. J., and Ebert, D. (2004). Conceptual issues in local adaptation. *Ecol. Lett.* 7, 1225–1241. doi: 10.1111/j.1461-0248.2004.00684.x
- Khoury, C. K., Achicanoy, H. A., Bjorkman, A. D., Navarro-Racines, C., Guarino, L., Flores-Palacios, X., et al. (2016). Origins of food crops connect countries worldwide. *Proc. R. Soc. B* 283, 468–474. doi: 10.1098/rspb.201 6.0792
- Kollner, T. G., Held, M., Lenk, C., Hiltpold, I., Turlings, T. C. J., Gershenzon, J., et al. (2008). A maize (*E*)-β-caryophyllene synthase implicated in indirect defense responses against herbivores is not expressed in most American maize varieties. *Plant Cell* 20, 482–494. doi: 10.1105/tpc.107.051672
- Kraft, K. H., Brown, C. H., Nabhan, G. P., Luedeling, E., Luna Ruiz, J. D. J., Coppens d'Eeckenbrugge, G., et al. (2014). Multiple lines of evidence for the origin of

domesticated chili pepper, *Capsicum annuum*, in Mexico. *Proc. Natl. Acad. Sci.* U.S.A. 111, 6165–6170. doi: 10.1073/pnas.1308933111

- Ladizinsky, G. (1998). Plant Evolution under Domestication. Dordrecht; New York, NY: Kluwer Academic Publishers.
- Laland, K., Uller, T., Feldman, M., Sterelny, K., Müller, G. B., Moczek, A., et al. (2014). Does evolutionary theory need a rethink? *Nature* 514, 161–164. doi: 10.1038/514161a
- Larson, G., Piperno, D. R., Allaby, R. G., Purugganan, M. D., Andersson, L., Arroyo-Kalin, M., et al. (2014). Current perspectives and the future of domestication studies. *Proc. Natl. Acad. Sci. U.S.A.* 111, 6139–6146. doi: 10.1073/pnas.1323964111
- Leppik, E. E. (1970). Gene centers of plants as sources of disease resistance. Annu. Rev. Phytopathol. 8, 323–344. doi: 10.1146/annurev.py.08.090170.0 01543
- McKey, D., Cavagnaro, T. R., Cliff, J., and Gleadow, R. (2010). Chemical ecology in coupled human and natural systems: people, manioc, multitrophic interactions and global change. *Chemoecology* 20, 109–133. doi: 10.1007/s00049-010-0047-1
- Mercer, K., Martínez-Vásquez, Á., and Perales, H. R. (2008). Asymmetrical local adaptation of maize landraces along an altitudinal gradient. *Evol. Appl.* 1, 489–500. doi: 10.1111/j.1752-4571.2008.00038.x
- Metcalf, R. L., and Lampman, R. L. (1989). The chemical ecology of Diabroticites and Cucurbitaceae. *Experientia* 45, 240–247. doi: 10.1007/BF01951810
- Meyer, R. S., DuVal, A. E., and Jensen, H. R. (2012). Patterns and processes in crop domestication: an historical review and quantitative analysis of 203 global food crops. *New Phytol.* 196, 29–48. doi: 10.1111/j.1469-8137.2012.04253.x
- Miller, A. J., and Knouft, J. H. (2006). GIS-based characterization of the geographic distributions of wild and cultivated populations of the Mesoamerican fruit tree *Spondias purpurea* (Anacardiaceae). Am. J. Bot. 93, 1757–1767. doi: 10.3732/ajb.93.12.1757
- Moreno, L. L., Tuxill, J., Moo, E. Y., Reyes, L. A., Alejo, J. C., and Jarvis, D. I. (2006). Traditional maize storage methods of Mayan farmers in Yucatan, Mexico: implications for seed selection and crop diversity. *Biodivers. Conserv.* 15, 1771–1795. doi: 10.1007/s10531-004-6679-0
- Moreno-Calles, A., Casas, A., García-Frapolli, E., and Torres-García, I. (2012). Traditional agroforestry systems of multi-crop "milpa" and "chichipera" cactus forest in the arid Tehuacan Valley, Mexico: their management and role in people's subsistence. *Agrofor. Syst.* 84, 207–226. doi: 10.1007/s10457-011-9460-x
- Moreno-Calles, A. I., Casas, A., Rivero-Romero, A. D., Romero-Bautista, Y. A., Rangel-Landa, S., Fisher-Ortíz, R. A., et al. (2016). Ethnoagroforestry: integration of biocultural diversity for food sovereignty in Mexico. J. Ethnobiol. Ethnomed. 12, 54. doi: 10.1186/s13002-016-0127-6
- Olsen, K. M., and Wendel, J. F. (2013). A bountiful harvest: genomic insights into crop domestication phenotypes. Annu. Rev. Plant Biol. 64, 47–70. doi: 10.1146/annurev-arplant-050312-120048
- Orozco-Ramírez, Q., Ross-Ibarra, J., Santacruz-Varela, A., and Brush, S. (2016). Maize diversity associated with social origin and environmental variation in Southern Mexico. *Heredity* 116, 477–484. doi: 10.1038/hdy.2016.10
- Peleg, Z., Fahima, T., Abbo, S., Krugman, T., Nevo, E., Yakir, D., et al. (2005). Genetic diversity for drought resistance in wild emmer wheat and its ecogeographical associations. *Plant Cell Environ.* 28, 176–191. doi: 10.1111/j.1365-3040.2005.01259.x
- Perales, H. R., Benz, B. F., and Brush, S. B. (2005). Maize diversity and ethnolinguistic diversity in Chiapas, Mexico. Proc. Natl. Acad. Sci. U.S.A. 102, 949–954. doi: 10.1073/pnas.0408701102
- Perez-Jaramillo, J. E., Mendes, R., and Raaijmakers, J. M. (2016). Impact of plant domestication on rhizosphere microbiome assembly and functions. *Plant Mol. Biol.* 90, 635–644. doi: 10.1007/s11103-015-0337-7
- Philippot, L., Raaijmakers, J. M., Lemanceau, P., and van der Putten, W. H. (2013). Going back to the roots: the microbial ecology of the rhizosphere. *Nat. Rev. Microbiol.* 11, 789–799. doi: 10.1038/nrmicro3109
- Pigliucci, M. (2001). Phenotypic Plasticity: Beyond Nature and Nurture. Baltimore, MD: Johns Hopkins University Press.
- Pinhasi van-Oss, R., Sherman, A., Zhang, H.-B., Vandemark, G., Coyne, C., and Abbo, S. (2016). Vernalization response of domesticated × wild chickpea progeny is subject to strong genotype by environment interaction. *Plant Breed*. 135, 102–110. doi: 10.1111/pbr.12325

- Piperno, D. R. (2017). Assessing elements of an extended evolutionary synthesis for plant domestication and agricultural origin research. *Proc. Natl. Acad. Sci.* U.S.A. 114, 6429–6437. doi: 10.1073/pnas.1703658114
- Piperno, D. R., Holst, I., Winter, K., and McMillan, O. (2015). Teosinte before domestication: experimental study of growth and phenotypic variability in Late Pleistocene and early Holocene environments. *Quat. Int.* 363, 65–77. doi: 10.1016/j.quaint.2013.12.049
- Piperno, D. R., Moreno, J. E., Iriarte, J., Holst, I., Lachniet, M., Jones, J. G., et al. (2007). Late Pleistocene and Holocene environmental history of the Iguala Valley, central Balsas Watershed of Mexico. *Proc. Natl. Acad. Sci. U.S.A.* 104, 11874–11881. doi: 10.1073/pnas.0703442104
- Pyhäjärvi, T., Hufford, M. B., Mezmouk, S., and Ross-Ibarra, J. (2013). Complex patterns of local adaptation in teosinte. *Genome Biol. Evol.* 5, 1594–1609. doi: 10.1093/gbe/evt109
- Rasmann, S., Köllner, T. G., Degenhardt, J., Hiltpold, I., Toepfer, S., Kuhlmann, U., et al. (2005). Recruitment of entomopathogenic nematodes by insect-damaged maize roots. *Nature* 434, 732–737. doi: 10.1038/nature03451
- Rau, D., Rodriguez, M., Leonarda Murgia, M., Balmas, V., Bitocchi, E., Bellucci, E., et al. (2015). Co-evolution in a landrace meta-population: two closely related pathogens interacting with the same host can lead to different adaptive outcomes. *Sci. Rep.* 5:12834. doi: 10.1038/srep12834
- Smartt, J., and Simmonds, N. W. (1995). *Evolution of Crop Plants*. New York, NY: John Wiley & Sons, Inc.
- Smith, B. D. (1997). The initial domestication of *Cucurbita pepo* in the Americas 10,000 years ago. *Science* 276, 932–934. doi: 10.1126/science.276. 5314.932
- Smith, K. P., and Goodman, R. M. (1999). Host variation for interactions with beneficial plant-associated microbes. Annu. Rev. Phytopathol. 37, 473–491. doi: 10.1146/annurev.phyto.37.1.473
- Staller, J., Tykot, R., and Benz, B. (2006). *Histories of Maize Multidisciplinary Approaches to the Prehistory, Linguistics, Biogeography, Domestication, and Evolution of Maize.* New York, NY: Elsevier.
- Stukenbrock, E. H., and McDonald, B. A. (2008). The origins of plant pathogens in agro-ecosystems. Annu. Rev. Phytopathol. 46, 75–100. doi: 10.1146/annurev.phyto.010708.154114
- Tamiru, A., Bruce, T. J., Woodcock, C. M., Caulfield, J. C., Midega, C. A., Ogol, C. K., et al. (2011). Maize landraces recruit egg and larval parasitoids in response to egg deposition by a herbivore. *Ecol. Lett.* 14, 1075–1083. doi: 10.1111/j.1461-0248.2011.01674.x
- Tétard-Jones, C., and Leifert, C. (2011). Plasticity of yield components of winter wheat in response to cereal aphids. NJAS Wageningen J. Life Sci. 58, 139–143. doi: 10.1016/j.njas.2011.01.003
- Tewksbury, J. J., Reagan, K. M., Machnicki, N. J., Carlo, T. A., Haak, D. C., Pe-aloza, A. L. C., et al. (2008). Evolutionary ecology of pungency in wild

chilies. Proc. Natl. Acad. Sci. U.S.A. 105, 11808–11811. doi: 10.1073/pnas.08026 91105

- Thies, J. A., Ariss, J. J., Kousik, C. S., Hassell, R. L., and Levi, A. (2016). Resistance to southern root-knot nematode (*Meloidogyne incognita*) in wild watermelon (*Citrullus lanatus* var. *citroides*). J. Nematol. 48, 14–19. doi: 10.21307/jofnem-2017-004
- Vallejo, M., Casas, A., Moreno-Calles, A. I., and Blancas, J. (2016). "Los sistemas agroforestales del Valle de Tehuacán: una perspectiva regional," in *Etnoagroforestería en México*, eds A. I. Moreno-Calles, A. Casas, V. M. Toledo, and M. Vallejo-Ramos (Mexico, DF: UNAM), 193–218.
- van den Bosch, R. (1971). Biological control of insects. Annu. Rev. Ecol. Syst. 2, 45–66. doi: 10.1146/annurev.es.02.110171.000401
- van Driesche, R., Hoddle, M., and Center, T. (2008). Control of Pests and Weeds by Natural Enemies. Hoboken, NJ: Wiley-Blackwell.
- Vavilov, N. I. (1926). Studies on the origin of cultivated plants. Bull. Appl. Bot. Plant Breed. 16, 139–245.
- Vavilov, N. I. (1951). The origin, variation, immunity and breeding of cultivated plants. Chron. Bot. 13, 1–366. doi: 10.1097/00010694-195112000-00018
- Vidal, S., Kuhlmann, U., and Edwards, C. R. (eds). (2005). Western Corn Rootworm: Ecology and Management. Wallingford, CT: CABI.
- Xing, X., Koch, A. M., Jones, A. M. P., Ragone, D., Murch, S., and Hart, M. M. (2012). Mutualism breakdown in breadfruit domestication. Proc. R. Soc. London B Biol. Sci. 279, 1122–1130. doi: 10.1098/rspb.2011.1550
- Zhang, H., Mittal, N., Leamy, L. J., Barazani, O., and Song, B. H. (2017). Back into the wild? Apply untapped genetic diversity of wild relatives for crop improvement. *Evol. Appl.* 10, 5–24. doi: 10.1111/eva.12434
- Zhou, Y., Ma, Y., Zeng, J., Duan, L., Xue, X., Wang, H., et al. (2016). Convergence and divergence of bitterness biosynthesis and regulation in Cucurbitaceae. *Nat. Plants* 2:16183. doi: 10.1038/nplants.2016.183
- Zizumbo-Villarreal, D., and Colunga-GarcíaMarín, P. (2010). Origin of agriculture and plant domestication in West Mesoamerica. *Genet. Resour. Crop Evol.* 57, 813–825. doi: 10.1007/s10722-009-9521-4

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Chen, Shapiro, Benrey and Cibrián-Jaramillo. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.