



Does Physical Activity Age Wild Animals?

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There is considerable interest in why the process of aging varies between individuals, both in humans and other animals. However, in animals, aging in terms of survival (demographic senescence) is understood in considerably more detail than in terms of declines in the body's functional capacity (functional senescence). Oxidative damage is probably an important component of the aging process in many species. Physical activity typically increases levels of oxidative stress, and many animals exhibit long and intense periods of active behavior. These observations raise a question that has not yet been addressed: while in humans, at least, activity is considered beneficial to health, could high intensity activity play a part in the rate that wild animals age? Studies to date suggest that increased "effort" can lead to reduced survival in free-living animals, but "effort" refers to different processes in different studies, and is rarely clarified or quantified. To understand the role of activity in functional senescence, studies must measure the detailed activity of free-ranging animals, possibly describing it in terms of intensity, frequency and duration, coupled with records of resultant physiological and DNA damage.

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"Comparative biology teaches us that reproduction is life's solution to the inevitability of death in the hostile environments of Earth" (Carnes, 2007).

INTRODUCTION: AGING IN THE WILD

Time waits for no animal. Once mature, long-lived individuals of all species experience aging—a progressive decline in the functional capacity of their bodies, a loss of integration of homeostatic systems and an increased vulnerability to death (López-Otín et al., 2013; Li et al., 2015). For many years, wild animals were thought not to grow old, dying before senescence took hold (Medawar, 1952). More recently, however, focussed-studies on multiple taxa have shown otherwise (Nussey et al., 2013; Jones et al., 2014). In many species, individuals continue living well beyond their functional peaks (Froy et al., 2018), and often carry on reproducing albeit at a declining rate (Nussey et al., 2013; Jones et al., 2014). Aging in animals can be considered in terms of the decline in the body's performance (functional senescence) or the decline in fertility rate coupled with an increased risk of mortality (demographic senescence). Most work focusses on demographic senescence and we now know that there is a great diversity in trajectories of demographic senescence, not only between different species but also between individuals of a given species (Jones et al., 2008, 2014).

In contrast, functional senescence, at least in wild animals, is less well studied. Research to date has shown age-specific changes in the immunity of wild animals (Cichon et al., 2003) and in certain other components of their physiology such as haematocrit (blood oxygen stores), hormones and metabolism (Massot et al., 2011; Elliott et al., 2014). In humans we know that physiological aging starts at different times in different individuals, and that it can occur at different rates even within the same person, depending on the tissue or physiological system examined (Sehl and Yates, 2001; Belsky et al., 2015). For example, the autonomic nervous and reproductive-endocrine systems senesce nearly four times faster than the CNS or musculoskeletal systems (Sehl and Yates, 2001). Similar detail at the physiological level is missing in wild animals (Reichard, 2016), but we know that some phenotypic, physiological, and behavioral traits can age asynchronously (Elliott et al., 2014; Hayward et al., 2015; Kervinen et al., 2015), in a way similar to in ourselves. At the same time, we have a comparatively detailed picture of the interconnected nature of aging between humans and the environment (Dato et al., 2017), and while similar evidence is accruing in wild animals (Nussey et al., 2008; Marshall et al., 2015), our understanding is far from complete (Hammers et al., 2015).

There is a clear gap in our knowledge stemming from how the behavioral choices of animals influence their aging. Why should this be important? In this *Review*, we discuss whether the activities that animals carry out as part of their everyday lives could be a key component shaping their aging process. We discuss how activity could influence aging, what we are yet to understand about this process, and the key future directions that studies should take in order to drive forward the field of wild animal gerontology.

OXIDATIVE STRESS AS A COMPONENT OF AGING

Oxidative stress is one of the most important physiological costs of living and thus a key modulator of life-history trade-offs (Monaghan et al., 2009; Speakman and Garratt, 2013; Blount et al., 2015; Speakman et al., 2015). All aerobically respiring organisms produce reactive oxygen and nitrogen species (RS), which are highly reactive and can damage biomolecules including lipids, proteins, and DNA. The production of RS can be counterbalanced by various antioxidants (AO). Normally, cells are in a dynamic redox state whereby RS and AO are in so-called oxidative balance. When RS increases, there is a concomitant increase in the AO system to return the cells to oxidative equilibrium. An increase in RS production not matched by the AO response leads to an increase in oxidative damage and resulting oxidative stress (Costantini and Verhulst, 2009). There is considerable controversy as to the role of oxidative stress in aging, and research is needed to examine key mechanistic links between oxidative stress and senescence (Freitas and de Magalhães, 2011; Speakman et al., 2015). However, although we do not yet fully understand the functional links from production through to outcome for the whole organism (Speakman et al., 2015), oxidative stress is clearly a key underlying component of the aging process for many organisms.

BASAL METABOLIC RATE (BMR) AND OXIDATIVE STRESS

The primary biological process that results in the production of RS is an animal's basal metabolism. Two of the strongest covariates with between-species differences in lifespan are the amount of oxidative DNA damage (Adelman et al., 1988) and BMR (Hulbert et al., 2007; though see Munshi-South and Wilkinson, 2010). The pace of life theory postulates that species with a lower BMR mature at a later age and larger size, reproduce at slower rates, and have longer lifespans. In support of the pace of life theory, evolved changes in the BMR of a species occur in tandem with the evolution of different pace of life strategies (Auer et al., 2018), but lifespan is not simply a function of BMR, if it is a function of BMR at all (Speakman et al., 2002; Magalhães et al., 2007), and instead may be co-regulated or correlated with other components of an organism's life history (Glazier, 2014). Moreover, BMR is not an important driver of aging and lifespan at the within-species level (Rønning et al., 2014; Norin and Gamperl, 2017), and may even be beneficial (Speakman et al., 2002). For example, field voles Microtus agrestis kept at cooler temperatures and thus exhibiting a higher BMR did not have shorter lifespans (Selman et al., 2008), whilst between-individual natural variation in BMR had either no effect or a positive effect on longevity in zebra finches (Taeniopygia guttata; Rønning et al., 2014), lab mice (Mus musculus; Speakman et al., 2004) and Glanville fritillary butterflies (Melitaea cinxia; Niitepold and Hanski, 2012). BMR therefore is not the primary driver of individual variation in aging, probably because the body can largely remain in oxidative balance. Instead, variation in individual aging could be explained by sources of oxidative stress that overwhelm mechanisms of oxidative balance.

PHYSICAL ACTIVITY, AGING, AND "EFFORT"

Many organisms live active lives. During activity, metabolic rate increases to multiple times BMR (Piersma, 2010). A concomitant increase in oxidative stress and DNA damage under these conditions has been reported in a range of laboratory species (summarized by: Yap et al., 2017b) and in humans (Fisher-Wellman and Bloomer, 2009). Oxidative stress is a common consequence of physical activity. But does this mean that physical activity can shorten lifespan? After all, moderate physical activity is known to increase lifespan (e.g., lab rodents: Navarro et al., 2004; Boveris and Navarro, 2008; Vaanholt et al., 2010) and to offer protective benefits against oxidative damage (Radak et al., 2008). Potentially, the relationship between health and levels of physical activity is hormetic, whereby increases in physical activity lead to increases in physical or physiological benefits up to a threshold beyond which benefits then decline (Costantini et al., 2010; Pontzer, 2018; Figure 1A). Whilst evidence for hormetic responses are widespread in humans (Radak et al.,

2008; though see Arem et al., 2015) and lab animals (Goto et al., 2007), evidence in wild animals is rarer, quite possibly because so few studies carry out suitable experiments. To date, studies have mainly investigated the extremes of the theoretical curve only, such as low activity vs. high activity (e.g., Aniagu et al., 2006). Combining studies that have investigated activity at low, intermediate and high levels suggests that low levels of voluntary activity have no effect on oxidative stress (Selman et al., 2002), intermediate levels of activity are associated with lower levels of oxidative stress and/or higher antioxidant protection (Costantini et al., 2008), and higher levels of activity have higher oxidative stress and lower antioxidant protection (Costantini et al., 2008). This is consistent with the hormetic response. There is a critical need for more studies, and the testing of a fuller range of activity levels, to understand how commonly the health effects of activity are hormetic. The impact of increasing activity levels on health will no doubt vary between species and contexts. Knowing at what point exercise may start to negatively impact health is a challenge; for example, it is possible that organisms carrying out sustained exercise below their energetic ceiling (i.e., <7 x resting metabolic rate: Hammond and Diamond, 1997), may not suffer obvious costs (e.g., Welcker et al., 2010). Only when organisms exercise above this ceiling, may negative impacts on health be apparent (Pontzer, 2018) and it is not clear that dose-dependent studies of human mortality capture this (Arem et al., 2015). Furthermore, while in some cases the hormetic response may be apparent, in other species, increasing activity levels may have no effect on health until a threshold beyond which this increase becomes deleterious (Figure 1B). In other cases, perhaps simply the greater the activity level the greater the negative impact on health (Figure 1C). The shape of the response in humans can change both with physical training and antioxidant intake (Radak et al., 2017; Pontzer, 2018). For example, training can increase the level of exercise required before it becomes damaging or minimizes the oxidative stress damage (Radak et al., 2017). Similarly, experimental work in lab animals shows that training can decrease oxidative stress (Larcombe et al., 2010). In the wild, migrating birds may have higher intakes of dietary antioxidants, which could offer a protective benefit during long distance flight by countering increased RS production (Cooper-Mullin and McWilliams, 2016). The implications of these findings for the importance of animal exercise (Halsey, 2016), and exercise ecology (van Dijk and Matson, 2016), needs further exploration.

Despite reviews on humans (Powers et al., 2011) and examples from animal laboratory models (Yap et al., 2017b), as yet no studies have summarized broadly whether physical activity causes oxidative stress and DNA damage in wild animals either in the lab or wild. To date, such studies on wild animals have manipulated the "effort" they exhibit, an ill-defined term often used in the literature to allude to investment (physical and/or physiological) in some activity. To understand how manipulated changes in "effort" influence oxidative stress biomarkers and levels of DNA damage, we collated studies found in the literature (N = 15). We found that in about half of studies, physical activity such as long distance flight or raising offspring was found to cause increases in oxidative stress (7/13 studies) and DNA damage (3/6 studies: **Table 1**). Moreover, increases in DNA damage were most evident when physical activity was considered

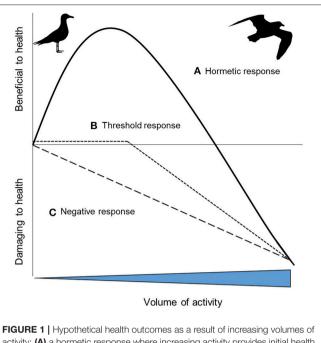


FIGURE 1 | Hypothetical health outcomes as a result of increasing volumes of activity: (A) a hormetic response where increasing activity provides initial health benefits, before then having an increasingly negative impact; (B) increasing activity has no or little impact on health up to a threshold, after which it becomes progressively more deleterious; (C) the impacts on health of increasing activity are progressively more deleterious [inspired by 43].

to be high; where activity levels were classified as "low" or "voluntary," there was no effect on oxidative stress or DNA damage (**Table 1**).

Many wild-based studies manipulate "effort" by increasing brood or litter sizes and there is mixed evidence to support the contention that this leads to reduced survival-a measure of aging (Santos and Nakagawa, 2012). For example, Golet et al. (1998) reported that 29% (7/24) of studies manipulating clutch size negatively affected adult survival, whilst a study on jackdaws Corvus monedula showed that experimentally increased clutch sizes were associated with a decrease in long-term survival rates (Boonekamp et al., 2014). Unfortunately, few studies quantify the "effort" of the animals, for example in terms of their energy expenditure, and so are unable to show a link between effort and measures of senescence such as reduced survivability (though see: Daan et al., 1996). Moreover, there is only one study (and it is on captive animals-zebra finches), that includes measures of oxidative stress, DNA damage and survival (Sudyka et al., 2016). Hence, studies published so far investigating activity correlates of wild animal senescence have failed to combine all the following crucial elements: quantified physical activity, physiological and DNA damage, and measures of functional or demographic senescence.

PHYSICAL ACTIVITY: DURATION, FREQUENCY, OR INTENSITY?

Athletes can run for 100 m considerably faster than they can run a mile. The basic physiology underlying this phenomenon TABLE 1 | Published animal studies that assess "effort" and its impact on oxidative stress (OS), antioxidant biomarkers (AO), and DNA damage (telomere length, comet assays).

Species	Study type	Design	Measure of effort	Timing of sampling	Biomarkers	DNA damage	References
Zebra finch Taeniopygia guttata	C/E	Brood size increase		12 d	OS =	=	Sudyka et al., 2016
Zebra finch <i>Taeniopygia guttata</i>	C/E	Brood size increase		35 d 1 yr	$OS \uparrow AO =$ $OS \uparrow AO =$	↑ ↑	Reichert et al., 2014
Blue tit <i>Cyanistes caeruleus</i>	W/E	Brood size increase		14 d		\uparrow	Sudyka et al., 2014
Great tit <i>Parus major</i>	C/E	Wing feather removal		21 d	$OS=AO\uparrow$		Vaugoyeau et al., 2015
Pigeon <i>Columba livia</i>	W/E	Distance	Short distance (39 km) Long distance (179 km)	Post exercise	OS ↓ AO ↑ OS ↑ AO ↓		Costantini et al., 2008
Zebra finch <i>Taeniopygia guttata</i>	C/E	Flight activity	Low activity (~900 m) High activity(~2,700 m)	3 d	$\begin{array}{l} OS = AO = \uparrow \\ OS \uparrow AO = \downarrow \end{array}$		Costantini et al., 2013
Zebra finch <i>Taeniopygia guttata</i>	C/E	Flight activity	High activity (number of trips)	90 d	$OS \uparrow AO =$		Yap et al., 2017a
Great tit Parus major	W/E	Wing feather removal	Feeding rate (no change)	43 d	OS = AO =		Wegmann et al., 2015
Magellanic penguins Spheniscus magellanicus	W/N		Number offspring			=	Cerchiara et al., 2017
Chub <i>Squalius cephalus</i>	C/E	Swimming to exhaustion	Critical swimming speed	Post 2nd exercise bout	AO =	1	Aniagu et al., 2006
Snow bunting Plectrophenax nivalis	W/N	Feeding rate	Feeding rate	8–15 d	OS ↑		Guindre-Parker et al., 2013
European robin Erithacus rubecula	W/N	Migration flight		Post- nocturnal flight	OS ↑ AO ↑		Jenni-Eiermann et al., 2014
Bald ibis Geronticus eremite	W/N	Migration flight	Time spent flying	Pre vs. post flight	OS = AO =		Bairlein et al., 2015
Barn swallow <i>Hirundo rustica</i>	W/E	Brood size increase		13 d	$OS \uparrow AO =$		Pap et al., 2018
Field vole <i>Microtus agrestris</i>	C/E	Voluntary running	0 vs. 5.9 km/d	7 d	OS = AO =	=	Selman et al., 2002

Publications include both wild (W) and captive (C) studies, natural (N), and experimental (E) manipulations. Changes in measures relative to baseline or pre-exercise values are reported as increases (†), decreases (↓), no change (=), or a mixture (where multiple markers recorded) († =).

is that short-burst exercise employs ATP and phosphocreatine to generate energy anaerobically—instant energy stores that deplete over a matter of seconds—while longer-term exercise must also employ lactic acid and electron-transport phosphorylation aerobically (Thompson, 2017). There are analogies to short burst and long endurance activity in animal ecology (Killen et al., 2017). Barnacle geese *Branta leucopsis* make migratory flights lasting around 60 h in total during which their heart rates are ~60% maximum (Butler et al., 1998), representing moderate exercise of long duration. In contrast, activities such as flight take-off, predator avoidance, mating displays and active predation typically involve rates of energy expenditure at least several-fold that at rest (Vehrencamp et al., 1989; Green et al., 2009; Lane et al., 2010; Soulsbury, 2019), and can only be performed for short periods (Piersma, 2010).

Exercise can be broadly characterized by three interacting components: (1) *Intensity* characterizes the percentage of oxygen consumption rate (VO_2) or heart rate relative to that at peak power output. In humans, high-intensity exercise is typically

characterized as including bouts at 64-90% VO_{2max} or 77-95% HR_{max} (Pollock et al., 1998). (2) Duration refers specifically to time spent carrying out the exercise, and (3) Frequency refers to the number of bouts of that exercise conducted. Thus the product of duration, frequency, and intensity in theory represents the overall volume of activity undertaken. We know that in isolation, increased duration (Bloomer et al., 2007), intensity (Wadley et al., 2016) and frequency (humans: Tanimura et al., 2010; dogs: Hinchcliff et al., 2000) of activity leads to increased oxidative stress. Furthermore, we know that in humans at least, oxidative stress is higher during anaerobic than aerobic exercise when conducted at similar intensities and duration (Bloomer et al., 2005). Therefore, activities that are of high intensity and anaerobic are quite possibly more immediately physiologically damaging than lower-intensity aerobic activities, perhaps even if the latter are of longer duration. For example, frequent and intense feeding activity by a passerine bird may be more costly to an individual than longer but less intense commuting flights between a foraging area and its nest, even if the volume of activity represented by the two activities is similar. With advances in data loggers to characterize animal activity, it is becoming progressively more feasible to quantify the composite parts of animal activities, in turn making it possible to relate the duration, intensity, and frequency of each part to both the energy costs and physiological costs of behavior.

CONCLUSIONS AND FUTURE STUDIES

There is a growing awareness that the active behaviors animals perform in their daily lives may come with short-term and sometimes also long-term physiological costs, increasing oxidative stress (Costantini et al., 2008), DNA damage (Aniagu et al., 2006) and in some case reducing survival (Daan et al., 1996). However, there is a gap in our knowledge about which components of these activities are most influential in driving senescence and under what circumstances this influence is greatest. To our knowledge, no studies of wild animals have yet combined a detailed quantification of their physical activity and measurements of the resultant physiological costs (immunity, OS, DNA damage, and physiological homeostasis), particularly during critical periods when physical activity may be heightened such as reproductive events or migration. An important aspect of attending to this knowledge gap will be the quantification of "effort". Furthermore, while most work to date investigating how activity affects aging has focussed on oxidative stress, physiological systems are highly interconnected and physical

REFERENCES

- Adelman, R., Saul, R. L., and Ames, B. N. (1988). Oxidative damage to DNA: relation to species metabolic rate and life span. PNAS 85, 2706–2708. doi: 10.1073/pnas.85.8.2706
- Aniagu, S. O., Day, N., Chipman, J. K., Taylor, E. W., Butler, P. J., and Winter, M. J. (2006). Does exhaustive exercise result in oxidative stress and associated DNA damage in the chub (*Leuciscus cephalus*)? *Envi. Mol. Mut.* 47, 616–623. doi: 10.1002/em.20247
- Arem, H., Moore, S. C., Patel, A., Hartge, P., De Gonzalez, A. B., Visvanathan, K., et al. (2015). Leisure time physical activity and mortality: a detailed pooled analysis of the dose-response relationship. *JAMA* 175, 959–967. doi: 10.1001/jamainternmed.2015.0533
- Auer, S. K., Dick, C. A., Metcalfe, N. B., and Reznick, D. N. (2018). Metabolic rate evolves rapidly and in parallel with the pace of life history. *Nature Comm.* 9:14. doi: 10.1038/s41467-017-02514-z
- Bairlein, F., Fritz, J., Scope, A., Schwendenwein, I., Stanclova, G., van Dijk, G., et al. (2015). Energy expenditure and metabolic changes of free-flying migrating northern bald ibis. *PLOS ONE* 10:e0134433. doi: 10.1371/journal.pone.0134433
- Belsky, D. W., Caspi, A., Houts, R., Cohen, H. J., Corcoran, D. L., Danese, A., et al. (2015). Quantification of biological aging in young adults. *PNAS* 112, E4104–E4110. doi: 10.1073/pnas.1506264112
- Bloomer, R., Davis, P., Consitt, L., and Wideman, L. (2007). Plasma protein carbonyl response to increasing exercise duration in aerobically trained men and women. *Int. J. Sport Med.* 28, 21–25. doi: 10.1055/s-2006-924140
- Bloomer, R. J., Goldfarb, A. H., Wideman, L., McKenzie, M. J., and Consitt, L. A. (2005). Effects of acute aerobic and anaerobic exercise on blood markers of oxidative stress. J. Str. Cond. Res. 19, 276–285. doi: 10.1519/14823.1
- Blount, J. D., Vitikainen, E. I. K., Stott, I., and Cant, M. A. (2015). Oxidative shielding and the cost of reproduction. *Biol. Rev.* 91, 483–497. doi: 10.1111/brv.12179

activity is likely to impact other biological processes such as the immune system, and hypothalamus-pituitary-adrenal axis (Eikenaar et al., 2018; Pontzer, 2018; *sensu* exercise immunology: van Dijk and Matson, 2016), which may also be associated with aging (Jurk et al., 2014).

Individual-based, long-term studies provide excellent opportunities to address a number of eco-evolutionary questions (Clutton-Brock and Sheldon, 2010), including how activity and exercise impact aging. A first step for example, could be to quantify exercise and physiological markers during the mating season of polygynous species such as red deer *Cervus elephaus* or black grouse *Lyrurus tetrix*, across multiple breeding seasons. This could prove insightful in describing differing rates of senescence between individuals, as indicated by, for instance, declining home range sizes (Froy et al., 2018). With the right techniques, there is huge potential in existing study systems to investigate the impacts of activity on animal senescence.

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All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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- Boonekamp, J. J., Salomons, M., Bouwhuis, S., Dijkstra, C., and Verhulst, S. (2014). Reproductive effort accelerates actuarial senescence in wild birds: an experimental study. *Ecol. Lett.* 17, 599–605. doi: 10.1111/ele.12263
- Boveris, A., and Navarro, A. (2008). Systemic and mitochondrial adaptive responses to moderate exercise in rodents. *Free Rad. Biol. Med.* 44, 224–229. doi: 10.1016/j.freeradbiomed.2007.08.015
- Butler, P. J., Woakes, A. J., and Bishop, C. M. (1998). Behaviour and physiology of Svalbard barnacle geese *Branta leucopsis* during their autumn migration. *J. Av. Biol.* 29:536. doi: 10.2307/3677173
- Carnes, B. A. (2007). Senescence viewed through the lens of comparative biology. Ann. New York Acad. Sci. 1114:14–22. doi: 10.1196/annals. 1396.045
- Cerchiara, J. A., Risques, R. A., Prunkard, D., Smith, J. R., Kane, O. J., and Boersma, P. D. (2017). Magellanic penguin telomeres do not shorten with age with increased reproductive effort, investment, and basal corticosterone. *Ecol. Evol.* 7, 5682–5691. doi: 10.1002/ece3.3128
- Cichon, M., Sendecka, J., and Gustafsson, L. (2003). Age-related decline in humoral immune function in collared flycatchers. J. Evol. Biol. 16, 1205–1210. doi: 10.1046/j.1420-9101.2003.00611.x
- Clutton-Brock, T., and Sheldon, B. C. (2010). Individuals and populations: the role of long-term, individual-based studies of animals in ecology and evolutionary biology. *Trends Ecol. Evol.* 25, 562–573. doi: 10.1016/j.tree.2010.08.002
- Cooper-Mullin, C., and McWilliams, S. R. (2016). The role of the antioxidant system during intense endurance exercise: lessons from migrating birds. J. Exp. Biol. 219, 3684–3695. doi: 10.1242/jeb.123992
- Costantini, D., Dell'Ariccia, G., and Lipp, H.-P. (2008). Long flights and age affect oxidative status of homing pigeons (*Columba livia*). J. Exp. Biol. 211, 377–381. doi: 10.1242/jeb.012856
- Costantini, D., Metcalfe, N. B., and Monaghan, P. (2010). Ecological processes in a hormetic framework. *Ecol. Lett.* 13, 1435–1447. doi: 10.1111/j.1461-0248.2010.01531.x

- Costantini, D., Monaghan, P., and Metcalfe, N. B. (2013). Loss of integration is associated with reduced resistance to oxidative stress. J. Exp. Biol. 216, 2213–2220. doi: 10.1242/jeb.083154
- Costantini, D., and Verhulst, S. (2009). Does high antioxidant capacity indicate low oxidative stress? *Funct. Ecol.* 23, 506–509. doi: 10.1111/j.1365-2435.2009.01546.x
- Daan, S., Deerenberg, C., and Dijkstra, C. (1996). Increased daily work precipitates natural death in the kestrel. J. Anim. Ecol. 65, 539-544. doi: 10.2307/5734
- Dato, S., Rose, G., Crocco, P., Monti, D., Garagnani, P., Franceschi, C., et al. (2017). The genetics of human longevity: an intricacy of genes, environment, culture and microbiome. *Mech. Age. Dev.* 165, 147–155. doi: 10.1016/j.mad.2017.03.011
- Eikenaar, C., Isaksson, C., and Hegemann, A. (2018). A hidden cost of migration? Innate immune function versus antioxidant defense. *Ecol. Evol.* 8, 2721–2728. doi: 10.1002/ece3.3756
- Elliott, K. H., Hare, J. F., Le Vaillant, M., Gaston, A. J., Ropert-Coudert, Y., and Anderson, W. G. (2014). Ageing gracefully: physiology but not behaviour declines with age in a diving seabird. *Funct. Ecol.* 29, 219–228. doi: 10.1111/1365-2435.12316
- Fisher-Wellman, K., and Bloomer, R. J. (2009). Acute exercise and oxidative stress: a 30 year history. *Dyn. Med.* 8:1. doi: 10.1186/1476-5918-8-1
- Freitas, A. A., and de Magalhães, J. P. (2011). A review and appraisal of the DNA damage theory of ageing. *Mut. Res.* 728, 12–22. doi: 10.1016/j.mrrev.2011.05.001
- Froy, H., Börger, L., Regan, C. E., Morris, A., Morris, S., Pilkington, J. G., et al. (2018). Declining home range area predicts reduced late-life survival in two wild ungulate populations. *Ecol. Lett.* 21, 1001–1009. doi: 10.1111/ele.12965
- Glazier, D. S. (2014). Is metabolic rate a universal "pacemaker" for biological processes? *Biol. Rev.* 90, 377–407. doi: 10.1111/brv.12115
- Golet, G. H., Irons, D. B., and Estes, J. A. (1998). Survival costs of chick rearing in black-legged kittiwakes. J. Anim. Ecol. 67, 827–841. doi: 10.1046/j.1365-2656.1998.00233.x
- Goto, S., Naito, H., Kaneko, T., Chung, H. Y., and Radák, Z. (2007). Hormetic effects of regular exercise in aging: correlation with oxidative stress. *Appl. Phys. Nutr. Met.* 32, 948–953. doi: 10.1139/h07-092
- Green, J., White, C., Bunce, A., Frappell, P., and Butler, P. (2009). Energetic consequences of plunge diving in gannets. *End. Spec. Res.* 10, 269–279. doi: 10.3354/esr00223
- Guindre-Parker, S., Baldo, S., Gilchrist, H. G., Macdonald, C. A., Harris, C. M., and Love, O. P. (2013). The oxidative costs of territory quality and offspring provisioning. J. Evol. Biol. 26, 2558–2565. doi: 10.1111/jeb.12256
- Halsey, L. G. (2016). Do animals exercise to keep fit? J. Anim. Ecol. 85, 614–620. doi: 10.1111/1365-2656.12488
- Hammers, M., Kingma, S. A., Bebbington, K., van de Crommenacker, J., Spurgin, L. G., Richardson, D. S., et al. (2015). Senescence in the wild: Insights from a long-term study on Seychelles warblers. *Exp. Geront.* 71, 69–79. doi: 10.1016/j.exger.2015.08.019
- Hammond, K. A., and Diamond, J. (1997). Maximal sustained energy budgets in humans and animals. *Nature* 386:457.
- Hayward, A. D., Moorad, J., Regan, C. E., Berenos, C., Pilkington, J. G., Pemberton, J. M., et al. (2015). Asynchrony of senescence among phenotypic traits in a wild mammal population. *Exp. Geront.* 71, 56–68. doi: 10.1016/j.exger.2015. 08.003
- Hinchcliff, K. W., Reinhart, G. A., DiSilvestro, R., Reynolds, A., Blostein-Fujii, A., and Swenson, R. A. (2000). Oxidant stress in sled dogs subjected to repetitive endurance exercise. *Am. J. Vet Res.* 61, 512–517. doi: 10.2460/ajvr.2000. 61.512
- Hulbert, A. J., Pamplona, R., Buffenstein, R., and Buttemer, W. A. (2007). Life and death: metabolic rate, membrane composition, and life span of animals. *Phys. Rev.* 87, 1175–1213. doi: 10.1152/physrev.00047.2006
- Jenni-Eiermann, S., Jenni, L., Smith, S., and Costantini, D. (2014). Oxidative stress in endurance flight: an unconsidered factor in bird migration. *PLoS ONE* 9:e97650. doi: 10.1371/journal.pone.0097650
- Jones, O. R., Gaillard, J. M., Tuljapurkar, S., Alho, J. S., Armitage, K. B., Becker, P. H., et al. (2008). Senescence rates are determined by ranking on the fast-slow life-history continuum. *Ecol. Lett.* 11, 664–673. doi: 10.1111/j.1461-0248.2008.01187.x
- Jones, O. R., Scheuerlein, A., Salguero-Gómez, R., Camarda, C. G., Schaible, R., Casper, B. B., et al. (2014). Diversity of ageing across the tree of life. *Nature* 505, 169–173. doi: 10.1038/nature12789

- Jurk, D., Wilson, C., Passos, J. F., Oakley, F., Correia-Melo, C., Saretzki, G., et al. (2014). Chronic inflammation induces telomere dysfunction and accelerates ageing in mice. *Nat. Comm.* 2:5172. doi: 10.1038/ncomms5172
- Kervinen, M., Lebigre, C., Alatalo, R. V., Siitari, H., and Soulsbury, C. D. (2015). Life-history differences in age-dependent expressions of multiple ornaments and behaviors in a lekking bird. *Am. Nat.* 185, 13–27. doi: 10.1086/679012
- Killen, S. S., Calsbeek, R., and Williams, T. D. (2017). The ecology of exercise: mechanisms underlying individual variation in behavior, activity, and performance: an introduction to symposium. *Int. Comp. Biol.* 57, 185–194. doi: 10.1093/icb/icx083
- Lane, J. E., Boutin, S., Speakman, J. R., and Humphries, M. M. (2010). Energetic costs of male reproduction in a scramble competition mating system. J. Anim. Ecol. 79, 27–34. doi: 10.1111/j.1365-2656.2009.01592.x
- Larcombe, S. D., Coffey, J. S., Bann, D., Alexander, L., and Arnold, K. E. (2010). Impacts of dietary antioxidants and flight training on post-exercise oxidative damage in adult parrots. *Com. Biochem. Phys. B: Biochem. Mol. Biol.* 155, 49–53. doi: 10.1016/j.cbpb.2009.09.009
- Li, Q., Wang, S., Milot, E., Bergeron, P., Ferrucci, L., Fried, L. P., et al. (2015). Homeostatic dysregulation proceeds in parallel in multiple physiological systems. Age. Cell 14, 1103–1112. doi: 10.1111/acel.12402
- López-Otín, C., Blasco, M. A., Partridge, L., Serrano, M., and Kroemer, G. (2013). The hallmarks of aging. *Cell* 153, 1194–1217. doi: 10.1016/j.cell.2013.05.039
- Magalhães, J. P. D., Costa, J., and Church, G. M. (2007). An analysis of the relationship between metabolism, developmental schedules, and longevity using phylogenetic independent contrasts. J. Geront. A Biol. Sci. Med. Sci.62, 149–160. doi: 10.1093/gerona/62.2.149
- Marshall, H. H., Vitikainen, E. I., Mwanguhya, F., Businge, R., Kyabulima, S., Hares, M. C., et al. (2015). Lifetime fitness consequences of early-life ecological hardship in a wild mammal population. *Ecol. Evol.* 7, 1712–1724. doi: 10.1002/ece3.2747
- Massot, M., Clobert, J., Montes-Poloni, L., Haussy, C., Cubo, J., and Meylan, S. (2011). An integrative study of ageing in a wild population of common lizards. *Funct. Ecol.* 25, 848–858. doi: 10.1111/j.1365-2435.2011.01837.x
- Medawar, P. B. (1952). An Unsolved Problem of Biology. London: H.K. Lewis and Co.
- Monaghan, P., Metcalfe, N. B., and Torres, R. (2009). Oxidative stress as a mediator of life history trade-offs: mechanisms, measurements and interpretation. *Ecol. Lett.* 12, 75–92. doi: 10.1111/j.1461-0248.2008.01258.x
- Munshi-South, J., and Wilkinson, G. S. (2010). Bats and birds: Exceptional longevity despite high metabolic rates. Age. Res. Rev. 9, 12–19. doi: 10.1016/j.arr.2009.07.006
- Navarro, A., Gomez, C., López-Cepero, J. M., and Boveris, A. (2004). Beneficial effects of moderate exercise on mice aging: survival, behavior, oxidative stress, and mitochondrial electron transfer. Am. J. Phys. Reg. Inter. Comp. Physiol. 286, R505–R511. doi: 10.1152/ajpregu.00208.2003
- Niitepold, K., and Hanski, I. (2012). A long life in the fast lane: positive association between peak metabolic rate and lifespan in a butterfly. J. Exp. Biol. 216, 1388–1397. doi: 10.1242/jeb.080739
- Norin, T., and Gamperl, A. K. (2017). Metabolic scaling of individuals vs. populations: Evidence for variation in scaling exponents at different hierarchical levels. *Funct. Ecol.* 32, 379–388. doi: 10.1111/1365-2435.12996
- Nussey, D. H., Coulson, T., Festa-Bianchet, M., and Gaillard, J.-M. (2008). Measuring senescence in wild animal populations: towards a longitudinal approach. *Funct. Ecol.* 22, 393–406. doi: 10.1111/j.1365-2435.2008.01408.x
- Nussey, D. H., Froy, H., Lemaitre, J.,-F., Gaillard, J.-M., and Austad, S.N. (2013). Senescence in natural populations of animals: Widespread evidence and its implications for bio-gerontology. *Ageing. Res. Rev.* 12, 214–225. doi: 10.1016/j.arr.2012.07.004
- Pap, P. L., Vincze, O., Fülöp, A., Székely-Béres, O., Pătras, L., Pénzes, J., et al. (2018). Oxidative physiology of reproduction in a passerine bird: a field experiment. *Behav. Ecol. Sociobiol.* 72:34. doi: 10.1007/s00265-017-2434-x
- Piersma, T. (2010). Why marathon migrants get away with high metabolic ceilings: towards an ecology of physiological restraint. J. Exp. Biol. 214, 295–302. doi: 10.1242/jeb.046748
- Pollock, M. L., Gaesser, G. A., Butcher, J. D., Després, J. P., Dishman, R. K., Franklin, B. A., et al. (1998). ACSM position stand: the recommended quantity and quality of exercise for developing and maintaining cardiorespiratory and muscular fitness, and flexibility in healthy adults. *Med Sci Sports Exerc*, 30, 975–991.

- Pontzer, H. (2018). Energy constraint as a novel mechanism linking exercise and health. *Phys.* 33, 384–393. doi: 10.1152/physiol.00027.2018
- Powers, S. K., Nelson, W. B., and Hudson, M. B. (2011). Exercise-induced oxidative stress in humans: Cause and consequences. *Free Rad. Biol. Med.* 51, 942–950. doi: 10.1016/j.freeradbiomed.2010.12.009
- Radak, Z., Chung, H. Y., and Goto, S. (2008). Systemic adaptation to oxidative challenge induced by regular exercise. *Free Rad. Biol. Med.* 44, 153–159. doi: 10.1016/j.freeradbiomed.2007.01.029
- Radak, Z., Ishihara, K., Tekus, E., Varga, C., Posa, A., Balogh, L., et al. (2017). Exercise, oxidants, and antioxidants change the shape of the bell-shaped hormesis curve. *Redox Biol.* 12, 285–290. doi: 10.1016/j.redox.2017.02.015
- Reichard, M. (2016). Evolutionary ecology of aging: time to reconcile field and laboratory research. *Ecol. Evol.* 6, 2988–3000. doi: 10.1002/ece3.2093
- Reichert, S., Stier, A., Zahn, S., Arrivé, M., Bize, P., Massemin, S., et al. (2014). Increased brood size leads to persistent eroded telomeres. *Front. Ecol. Evol.* 2:9. doi: 10.3389/fevo.2014.00009
- Rønning, B., Moe, B., Berntsen, H. H., Noreen, E., and Bech, C. (2014). Is the rate of metabolic ageing and survival determined by basal metabolic rate in the zebra finch? *PLoS ONE* 9:e108675. doi: 10.1371/journal.pone.0108675
- Santos, E. S. A., and Nakagawa, S. (2012). The costs of parental care: a meta-analysis of the trade-off between parental effort and survival in birds. J. Evol. Biol. 25, 1911–1917. doi: 10.1111/j.1420-9101.2012.02569.x
- Sehl, M. E., and Yates, F. E. (2001). Kinetics of human aging: I. rates of senescence between ages 30 and 70 years in healthy people. J. Geront, Series A: Biol. Sc. and Med. Sci. 56, B198–B208. doi: 10.1093/gerona/56.5.b198
- Selman, C., McLaren, J. S., Collins, A. R., Duthie, G. G., and Speakman, J. R. (2002). Antioxidant enzyme activities, lipid peroxidation, and DNA oxidative damage: the effects of short-term voluntary wheel running. *Arch. Bioch. Biophys.* 401, 255–261. doi: 10.1016/s0003-9861(02)00050-4
- Selman, C., McLaren, J. S., Collins, A. R., Duthie, G. G., and Speakman, J. R. (2008). The impact of experimentally elevated energy expenditure on oxidative stress and lifespan in the short-tailed field vole *Microtus agrestis. Proc. Roy. Soc. B: Biol. Sci.* 275, 1907–1916. doi: 10.1098/rspb.2008.0355
- Soulsbury, C. D. (2019). Income and capital breeding in males: energetic and physiological limitations on male mating strategies. J. Exp. Biol. doi: 10.1242/jeb.184895
- Speakman, J. R., Blount, J. D., Bronikowski, A. M., Buffenstein, R., Isaksson, C., Kirkwood, T. B., et al. (2015). Oxidative stress and life histories: unresolved issues and current needs. *Ecol. Evol.* 5, 5745–5757. doi: 10.1002/ece3.1790
- Speakman, J. R., and Garratt, M. (2013). Oxidative stress as a cost of reproduction: Beyond the simplistic trade-off model. *BioEssays* 36, 93–106. doi: 10.1002/bies.201300108
- Speakman, J. R., Selman, C., McLaren, J. S., and Harper, E. J. (2002). Living fast, dying when? The link between aging and energetics. J. Nutr. 132, 1583S-1597S. doi: 10.1093/jn/132.6.1583S
- Speakman, J. R., Talbot, D. A., Selman, C., Snart, S., McLaren, J. S., Redman, P., et al. (2004). Uncoupled and surviving: individual mice with high metabolism have greater mitochondrial uncoupling and live longer. *Age. Cell* 3, 87–95. doi: 10.1111/j.1474-9728.2004.00097.x
- Sudyka, J., Arct, A., Drobniak, S., Dubiec, A., Gustafsson, L., and Cichon, M. (2014). Experimentally increased reproductive effort alters telomere

length in the blue tit (Cyanistes caeruleus). J. Evol. Biol. 27, 2258-2264. doi: 10.1111/jeb.12479

- Sudyka, J., Casasole, G., Rutkowska, J., and Cichon, M. (2016). Elevated reproduction does not affect telomere dynamics and oxidative stress. *Behav. Ecol. Sociobiol.* 70, 2223–2233. doi: 10.1007/s00265-016-2226-8
- Tanimura, Y., Shimizu, K., Tanabe, K., Kono, I., and Ajisaka, R. (2010). Effects of three consecutive days exercise on lymphocyte DNA damage in young men. *Eur. J. Appl. Phys.* 110, 307–314. doi: 10.1007/s00421-010-1499-2
- Thompson, M. A. (2017). Physiological and biomechanical mechanisms of distance specific human running performance. *Int. Comp. Biol.* 57, 293–300. doi: 10.1093/icb/icx069
- Vaanholt, L. M., Daan, S., Garland, J.r., T., and Visser, G.H. (2010). Exercising for life? energy metabolism, body composition, and longevity in mice exercising at different intensities. *Phys. Biochem. Zool.* 83, 239–251. doi: 10.1086/648434
- van Dijk, J. G. B., and Matson, K. D. (2016). Ecological immunology through the lens of exercise immunology: new perspective on the links between physical activity and immune function and disease susceptibility in wild animals. *Int. Comp. Biol* 56, 290–303. doi: 10.1093/icb/icw045
- Vaugoyeau, M., Decenciere, B., Perret, S., Karadas, F., Meylan, S., and Biard, C. (2015). Is oxidative status influenced by dietary carotenoid and physical activity after moult in the great tit (*Parus major*)? *J. Exp. Biol.* 218, 2106–2115. doi: 10.1242/jeb.111039
- Vehrencamp, S. L., Bradbury, J. W., and Gibson, R. M. (1989). The energetic cost of display in male sage grouse. *Anim. Behav.* 38, 885–896.
- Wadley, A. J., Turner, J. E., and Aldred, S. (2016). Factors influencing postexercise plasma protein carbonyl concentration. *Free Rad. Res.* 50, 375–384. doi: 10.3109/10715762.2015.1131824
- Wegmann, M., Voegeli, B., and Richner, H. (2015). Oxidative status and reproductive effort of great tits in a handicapping experiment. *Behav. Ecol.* 26, 747–754. doi: 10.1093/beheco/arv006
- Welcker, J., Moe, B., Bech, C., Fyhn, M., Schultner, J., Speakman, J. R., et al. (2010). Evidence for an intrinsic energetic ceiling in free-ranging kittiwakes *Rissa tridactyla. J. Anim. Ecol.* 79, 205–213. doi: 10.1111/j.1365-2656.2009. 01626.x
- Yap, K. N., Kim, O. R., Harris, K. C., and Williams, T. D. (2017a). Physiological effects of increased foraging effort in a small passerine. *J. Exp. Biol.* 220, 4282–4291. doi: 10.1242/jeb.160812
- Yap, K. N., Serota, M. W., and Williams, T. D. (2017b). The physiology of exercise in free-living vertebrates: what can we learn from current model systems? *Int. Comp. Biol.* 57, 195–206. doi: 10.1093/icb/icx016

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