



Tooth Microwear Texture in the Eastern Atlantic Harbour Seals (*Phoca vitulina vitulina*) of the German Wadden Sea and Its Implications for Long Term Dietary and Ecosystem Changes

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Marine mammals are increasingly threatened in their habitat by various anthropogenic impacts. This is particularly evident in prey abundance. Understanding the dietary strategies of marine mammal populations can help predict implications for their future health status and is essential for their conservation. In this study we provide a striking example of a new dietary proxy in pinnipeds to document marine mammal diets using a dental record. In this novel approach, we used a combination of 49 parameters to establish a dental microwear texture (DMTA) as a dietary proxy of feeding behaviour in harbour seals. This method is an established approach to assess diets in terrestrial mammals, but has not yet been applied to pinnipeds. Our aim was to establish a protocol, opening DMTA to pinnipeds by investigating inter- and intra-individual variations. We analysed the 244 upper teeth of 78 Atlantic harbour seals (Phoca vitulina vitulina). The specimens were collected in 1988 along the North Sea coast (Wadden Sea, Germany) and are curated by the Zoological Institute of Kiel University, Germany. An increasing surface texture roughness from frontal to distal teeth was found and related to different prey processing biomechanics. Ten and five year old individuals were similar in their texture roughness, whereas males and females were similar to each other with the exception of their frontal dentition. Fall and summer specimens also featured no difference in texture roughness. We established the second to fourth postcanine teeth as reference tooth positions, as those were unaffected by age, sex, season, or intraindividual variation. In summary, applying indirect dietary proxies, such as DMTA, will allow reconstructing dietary traits of pinnipeds using existing skeletal collection material.

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Combining DMTA with time series analyses is a very promising approach to track health status in pinniped populations over the last decades. This approach opens new research avenues and could help detect dietary shifts in marine environments in the past and the future.

Keywords: dental microwear texture analysis (DMTA), marine mammals, *Phoca vitulina* (Harbour seal), German Wadden Sea, pinnipeds, diet

INTRODUCTION

Marine mammals are increasingly threatened by cumulative anthropogenic activities, such as chemical pollution, marine litter, noise pollution, climate change, shipping, and offshoreconstruction (Lotze et al., 2005; Das et al., 2008; Gilles et al., 2009; Unger et al., 2017; Lehnert et al., 2018; Mikkelsen et al., 2019; Baltzer et al., 2020). Given their long life spans, their predatory feeding at high trophic levels as well as their proximity to coastal regions, marine mammals can be considered sentinels for ocean and human health (Bossart, 2011). The understanding of their population viability (aquatic food web, prey availability, distribution pattern) and the assessment of possible threats is an important basis for an informed management and conservation measures (Parrish et al., 2002; Kovacs et al., 2012).

The harbour seal (Phoca vitulina Linnaeus, 1758) is the most widespread pinniped confined to the Northern Hemisphere and a successful apex-predator in the Wadden Sea (Jensen et al., 2017). However, the status of the species is highly affected by human impact. In the first half of the 20th century, the harbour seal population in the Wadden Sea has been severely decimated by a heavy hunting pressure (Reijnders, 1981). After the hunting prohibition in the mid-1970s and the establishment of national parks in German waters, the harbour seal population recovered gradually (Brasseur et al., 2018). Since then, the growing seal population became increasingly threatened by a shift of prey availability as a result of fisheries interaction and climate change (Kovacs et al., 2012). Moreover, reoccurring virus epizootics have led to increased harbour seal mortality. In 2014, the Influenza A virus subtype H10N7 has caused the death of several thousand European harbour seals, with approximately 1,400 dead seals in the coastal waters of Schleswig Holstein, Germany, alone (Bodewes et al., 2015). In 1988 and 2002, the north-European population was struck by large scale epizootics caused by the phocine distemper virus (PDV), resulting in the largest mass mortality events in marine mammals recorded, with 23,000 and 30,000 dead harbour seals respectively (Härkönen et al., 2006). The PDV epidemics were associated with chronic exposure to contaminant loads that accumulate through aquatic food chains and weaken the immune function in harbour seals (Hall et al., 1992; De Swart et al., 1996; Ross et al., 1996). Many of the PDV infected harbour seal carcasses have been studied in an effort to better understand the risks and dangers to the population and to improve the conservation management (Härkönen and Heide-Jørgensen, 1990; Abt, 2002; Müller et al., 2004). As a side effect, the skeletal material has been accumulated in scientific collections. These large series of the harbour seal population are an exceptional well-documented treasure with the potential

to reconstruct harbour seal ecology and to engineer new tools to assess the health status of populations (Kahle et al., 2018; Ludolphy et al., 2018; Kierdorf et al., 2019).

The dental wear in curated tooth material can be a powerful tool for dietary discrimination. This dietary proxy has been studied on skeletal material from scientific collections for more than 40 years (Walker et al., 1978; Ungar, 2004). Dental wear occurs whenever a tooth comes into contact with another tooth (attrition) or ingesta (abrasion) (Kaiser et al., 2015). During the last decade, these microscopic scars have increasingly been interpreted by three-dimensional non-contact automated methods (for review see Schulz-Kornas et al., 2021) called dental microwear texture analysis (DMTA). These scars are continuously recorded and erased with each subsequent feeding event (Müller et al., 2014; Winkler et al., 2019a), thus the enamel surface merely contains a record of an individual's last few meals (Teaford and Oyen, 1989). Winkler et al. (2020a) showed in feeding experiments with rats (Rattus norvegicus forma *domestica*), that the overwriting of wear features on a micrometer scale takes at least two weeks.

Regarding terrestrial carnivorans, most DMTA measurements were taken on shearing and crushing facets of the maxillary fourth premolar and the mandibular first molar (Schubert et al., 2010; Ungar et al., 2010; De Santis et al., 2013; De Santis, 2016), which are differentiated into a pair of carnassial (Van Valen, 1969). These teeth are most probably used to efficiently process prey by shearing flesh and breaking bones. However, the dentition of marine mammals at large is specialised for capturing, rather than processing (Werth, 2000; Armfield et al., 2013; Loch and Simoes-Lopes, 2013). In contrast to terrestrial carnivorans, marine carnivores are not able to pin down struggling prey in their aqueous environment (Kienle and Berta, 2016). Furthermore, the mastication is reduced in marine mammal species, since oral processing of prey would be difficult and most probably inefficient, as prey items may drift away during consumption (Werth, 2000; Churchill and Clementz, 2015). To overcome those difficulties, the majority of marine mammals use a piercing bite to capture rapidly moving prey items, then swallow them whole (Adam and Berta, 2002; Armfield et al., 2013; Jones et al., 2013; Kienle and Berta, 2016). Regarding its skeletal and dental anatomy, the harbour seal is categorised as a pierce feeder (Adam and Berta, 2002; Churchill and Clementz, 2015), using its frontal teeth to seize agile prey, then swallowing it whole (Adam and Berta, 2002; Armfield et al., 2013; Jones et al., 2013; Kienle and Berta, 2016). The harbour seal uses raptorial biting to capture large prey, whereas smaller prey is captured using suction feeding (Kienle, 2014; Marshall et al., 2014; Ydesen et al., 2014). Similar to other phocids, the harbour seal displays an increased homodonty, simple cusp morphology, reduced carnassials (typical of carnivoran dentition) and a serial eruption sequence of the postcanine teeth (Loughlin, 1982; Adam and Berta, 2002; Churchill and Clementz, 2015). The deciduous teeth shed already *in utero* or shortly after birth, in order for the permanent dentition to be fully functional at the end of a short suckling period (4 – 6 weeks) (Meyer and Matzke, 2004).

In the North Sea, the diet composition of the harbour seal consists mostly of gadoid and flat fishes or clupeids and sand eels (Pierce et al., 1991; Gilles et al., 2008; de la Vega et al., 2016). Stomach content analyses conducted in the 70s and 80s revealed benthic fish, such as flounders, plaice, and gobies to be the most important prey species in the Wadden Sea (Behrends, 1985; Sievers, 1989), reflecting that harbour seals are opportunistic hunters. Additionally, harbour seals' food composition changed with increasing age, depending on size of both hunter and prey and availability of prey species (Behrends, 1985). Between 1981 and 1984 juvenile harbour seal stomach contents consisted of flatfish, gobies, and shrimp, while a shift towards more plaice, flounders and gadoids was observed with increasing age in sub-adults and adults (Behrends, 1985). Behavioural differences in harbour seals exist not only between age classes, but also between the sexes (Härkönen et al., 1999). Temporal and spatial behavioural differences between male and female harbour seals have been reported for water depth use, dive duration, dive shapes as well as movement, side fidelity and haul-out behaviour and are most likely linked to the annual reproduction cycle (Thompson et al., 1989; Baechler et al., 2002; Dietz et al., 2012; Wilson et al., 2015). Seasonal changes in diet composition indicated by gut content, spew/scat samples and stable isotopes have been linked to periodic shifts in prey availability and in movement patterns of harbour seals (Pierce et al., 1991; Hall et al., 1998; Dietz et al., 2012; de la Vega et al., 2016).

Considering dietary analysis in predatory taxa, DMTA has been successfully applied to terrestrial carnivorans (Schubert et al., 2010; De Santis, 2016), microbats (Purnell et al., 2013), fishes (Purnell et al., 2012; Purnell and Darras, 2015), odontocete whales (Purnell et al., 2017), and reptiles (Bestwick et al., 2021, 2019; Winkler et al., 2019b), but not yet to pinnipeds. Therefore, our goal is to establish a workflow for applying DMTA in pinnipeds and determine suitable tooth positions for measuring the dental microwear texture in harbour seals. We aim to develop a standardised sampling protocol for targeting tooth sampling positions in order to reduce variation in texture roughness due to differences in feeding biomechanics, which could possibly obscure dietary signals (Gordon, 1982). For this study, archived material from museum collections was used to establish DMTA as sensible tool to assess long term dietary and ecosystem changes in vulnerable marine top predators. We tested the following hypotheses.

(a) The dental microwear texture in harbour seals should differ between frontal and distal tooth positions because of their dissimilar demands. (b) The dental microwear texture should differ between age classes, sexes, and seasons according to reported observed differences in foraging behaviour.

MATERIALS AND METHODS

Specimen Selection

This study focused on the upper dentition of skeletal material from adult harbour seal specimens (Figure 1) (41 females, 37 males), curated at the osteological collection of the Zoological Institute of Kiel University, Germany. We concentrated on the dental record of harbour seals, which were collected in the year 1988 between June and October along the German Wadden Sea coast (Figure 1). Since the North European harbour seal population of 1988/89 was highly decimated by the phocine distemper virus (PDV), we were provided with a large number of individuals from both sexes across all age groups, which had died within a short period of time (4 month). The period of time reflects two seasons, summer = June, July, August; fall = September, October. This gave us a snapshot of the population and minimised the effect of other seasonal and annual prey availability. Due to the fragile preservation of the dental enamel of harbour seals, not all age groups were available for sampling and only two adult age groups (\geq 5 years and \geq 10 years were available in sample sizes with $n \ge 10$). Further, due to damage of single teeth (Supplementary Figure 1), we selected one side of the jaw only and chose the tooth of the betterpreserved jaw side (left or right). Information on sex, age at death, date, and location of collection as well as data on body length and weight was available for each animal. Age at death was determined by using cement-layer-analysis of the upper canine teeth (Abt, 2002). The 49 examined specimens from the North Friesian Wadden Sea (Figure 1) were 5 years (n = 12)and 10 years old at death (n = 37) (Table 1). An additional 30 examined specimens stem from the East Frisian Wadden Sea, ranging from 5- to 16-year old individuals (Table 1). We focused on the upper dentition (Figure 2) since during the epizootic the skulls were more often retrieved from the carcasses and curated than mandibles. The upper permanent dentition of harbour seals consists of tooth classes according to the dental formula (I 3/2, C 1/1, P 4/4, M 1/1 = 34) (Jefferson et al., 1993; Figures 2A,B). The premolar and molar teeth in pinnipeds are not clearly distinguishable. Therefore, those teeth are often referred to as postcanine teeth (Hillson, 1986). In harbour seals, these postcanines are small, uniform, of simple shape and have a main cusp and smaller cusps located mesial and distal (Hillson, 1986; Figure 2). Along the upper dentition, the following tooth positions (Figure 2) were measured: third incisor (I3), canine tooth (C), second (PC2), third (PC3) and fourth postcanine teeth (PC4). The first and second incisor as well as the first and the fifth postcanine teeth were excluded from this study. The first two incisors (I1, I2) of the upper dentition were too small to take adequate moulds. In most cases, those teeth were glued into the maxillary bone with an incorrect orientation. The first postcanine (PC1) exhibited no wear as it possibly experienced no tooth-tooth contact and/or limited contact to ingesta due to its



small size and its situation between much larger teeth. The fifth postcanine (PC5) was often calcified and/or exhibited sections were the thin enamel was flaked off, potentially due to excessive heating and drying during skull preparation (Aalderink et al., 2015). Specimens with dental anomalies (f. e. supernumerary teeth, abnormal tooth morphologies) and lesions as well as strong pathological changes of the temporomandibular joint (Kahle et al., 2018; Ludolphy et al., 2018) were excluded from this study.

Moulding

After cleaning the dentition with acetone-drenched soft cotton, moulds of upper teeth surfaces were made, using a high resolution silicone (Provil novo Light C.D.2 regular set; Type 3; Heraeus Kulzer, Dormagen, Germany) following established procedures (Schulz et al., 2010). To ensure measurements on intact enamel regardless of the wear stage, the silicone was applied in all specimens on an enamel surface not prone to chipping. On the second to fourth postcanine teeth of older animals, the thin enamel was occasionally abraded mesially along the narrow ridge of the main cusp. It was also worn down distal-lingually on the smaller cusps exposing areas of dentine (**Supplementary Figure 1**). Therefore, the silicone was applied not on the occluding dental facets between upper and lower dentition, but instead buccally on the main cusp of the postcanines on the straight surface between the maximum convexity at the cervical third and the tip (**Figures 2C–E**). The third incisors were moulded on the labial side, level with the postcanine moulds (**Figure 2D**). The canines of older individuals showed signs of lateral wear, most likely caused by attritional contacts between upper and lower canine teeth. Therefore, all moulds of the canines were taken on the labial side (**Figure 2D**).

Data Acquisition

Surface scans of enamel facets were conducted by the highresolution disk scanning confocal microscope μ surf Custom with a blue LED (470 nm) and a high-speed progressive-scan digital camera (984 × 984 pixels) (NanoFocus AG, Oberhausen, Germany) following the established protocol (Schulz et al., 2010). Pre-processing and filtering following the established protocol (Schulz et al., 2013, 2010); the template used is given as **Supplementary Material 1**. Where possible, four nonoverlapping measurement fields per facet with a square area of 160 μ m² were collected. We used a 100 × long distance objective (numerical aperture of 0.8, a resolution in *x*, *y* = 0.16 μ m and *z* = 0.06 μ m). Measurements with < 95% surface points or a vertical displacement range of $\delta z > 40 \ \mu$ m were rejected as well as surface areas with defects or adherent dirt. The dental microwear texture (DMT) was quantified by employing

TABLE 1 | Specimens included in the sample.

Specimen	Age at death*	Sex			Teeth			Collection date	Region	Total length	Girth	Weight
	[years]		13	С	PC2	PC3	PC4			[mm]	[mm]	[g]
27846	10	m	I	I	rl	r	r	25.06.1988	NF WS	1770	n.a.	n.a.
27941	10	m	1	Ι	r	r		12.08.1988	NF WS	1570	1300	61500
27952	5	f			I			23.08.1988	NF WS	1460	870	42000
28008	5	f	r	r	rl	rl	r	03.10.1988	NF WS	1400	820	37700
28052	5	m	rl	rl	rl	r	rl	01.07.1988	NF WS	1580	1000	62700
28066	5	m	r	r	rl	rl	rl	13.07.1988	NF WS	1560	n.a.	48500
28107	10	m						01.07.1988	NF WS	1630	1080	72500
28108	10	m	r	rl	I		I	01.07.1988	NF WS	1570	n.a.	64000
28184	10	m	I.	r	r	r	r	11.10.1988	NF WS	1610	n.a.	46000
28205	10	f	r	r	r	r	rl	11.10.1988	NF WS	1610	n.a.	47000
28212	5	m					I	11.10.1988	NF WS	1540	n.a.	43000
28213	5	f	I.	I		rl		11.10.1988	NF WS	1360	n.a.	34000
28297	10	m						29.09.1988	NF WS	n.a.	n.a.	n.a.
28327	10	f	r	I	r	rl		13.09.1988	NF WS	1460	n.a.	47000
28328	10	m						13.09.1988	NF WS	1500	n.a.	60000
28329	5	m	I	r	rl	rl	rl	13.09.1988	NF WS	1480	n.a.	52000
28338	5	m					I	15.09.1988	NF WS	1480	n.a.	46000
28344	10	m						15.09.1988	NF WS	1630	n.a.	52000
28348	10	m	1	I	rl	rl	rl	15.09.1988	NF WS	n.a.	n.a.	n.a.
28354	5	f	rl	Ì	1	rl	1	15.09.1988	NF WS	n.a.	n.a.	n.a.
28364	10	m	1	r	r	r	r	15 09 1988	NF WS	na	na	na
28426	5	f	rl	rl	rl		r	13 09 1988	NF WS	1420	na	41000
28464	5	f		r				30 07 1988	NF WS	1440	860	40700
28509	5	m	r	rl		r	rl	04 08 1988	NF WS	1540	940	53000
28802	10	f	1	1	I		rl	18 08 1988	NF WS	1720	na	54000
28812	10	m	i i	r				18 08 1988	NF WS	1600	na	60000
28814	10	f	r	r	r	rl		18 08 1988	NF WS	1500	na	40000
28850	10	f					r	15.07.1988	NE WS	1480	na	57000
28966	10	f			rl			09.08.1988	NE WS	1500	880	42000
28976	10	f						08.09.1988	NE WS	1410	na	42000
28981	10	f		I	r	rl	r	08 09 1988	NF WS	1360	n a	35000
28983	10	m	r	·		r	r	01 09 1988	NE WS	1610	na	47000
29043	10	f	rl					31.08.1988	NE WS	na	n a	na
29151	10	f				r		05.09.1988	NE WS	1520	n a	n a
29154	10	f					r	05.09.1988	NE WS	1460	n a	42000
29216	10	f						31.08.1988	NE WS	1560	n a	40000
29219	10	f				r	,	31.08.1988	NE WS	1560	n a	48000
20210	10	f			r			22.08.1988	NE WS	1570	n a	47000
20202	10	f						04.08.1988	NE WS	1570	n a	n a
29355	10	f				1		08.08.1988	NE WS	1550	na.	54000
29330	10	m						25.08.1988	NE WS	1570	na.	55000
29370	10	f			r			08.00.1088		1400	n.a.	00000
29403	10	n n	1		1			01.00.1088		1620	n.a.	77000
29423	10	f	1	I		1	1	01.09.1900		1660	n.a.	58000
20402	10	ı f				I		26 00 1000	NE MO	1000	n.a.	00000
23400	10	і т						20.03.1300	NE WO	11.ä.	n.a.	11.d.
23000	10	·11	ы	ч	-	1	I	31.00.1900	NE WO	1000	n.a.	77000
23000	10	111 F	11	ſI	r I	I	I	13.07.1900	NE WO	1030	n.a.	000
23100	10	۱ ۲			I			01.09.1900	NE WO	1.8.	n.a.	11.d.
29930	IU E	Ĩ	1			r		01.09.1988	INF WS	1000	n.a.	00000
30010	э 7	T	1			 		01.09.1988	EF WS	1220	n.a.	28000
30022	1	m	I		r	ri	I	06.09.1988	EF WS	1340	n.a.	40000

(Continued)

TABLE 1 | Continued

Specimen	Age at death* [years]	Sex	Teeth					Collection date	Region	Total length	Girth	Weight
			13	С	PC2	PC3	PC4			[mm]	[mm]	[g]
30026	6	f	I		I	r		29.08.1988	EF WS	n.a.	n.a.	45000
30038	16	m	rl		r			23.09.1988	EF WS	1510	n.a.	51000
30040	5	m	r	I		r	r	29.08.1988	EF WS	1390	n.a.	65000
30041	10	f				I.		01.09.1988	EF WS	1350	n.a.	45000
30042	7	f	rl	r	r	rl	rl	28.08.1988	EF WS	1480	n.a.	55000
30046	6	f			r			07.09.1988	EF WS	1410	n.a.	30000
30047	10	m	r		I			22.08.1988	EF WS	1460	n.a.	51000
30049	11	f				I		31.08.1988	EF WS	1390	n.a.	n.a.
30050	8	f			r		r	02.10.1988	EF WS	1410	n.a.	62000
30051	5	m	r		r		r	25.08.1988	EF WS	1480	n.a.	47000
30056	12	f	rl		r		rl	20.08.1988	EF WS	1290	n.a.	34000
30058	6	m			I			18.09.1988	EF WS	1280	n.a.	41000
30061	10	m			I			27.08.1988	EF WS	1450	n.a.	45000
30063	9	f	r				1	01.09.1988	EF WS	1410	n.a.	34000
30064	5	f	r			r	r	23.08.1988	EF WS	1380	n.a.	52000
30496	13	m						18.08.1988	EF WS	1500	n.a.	52000
30642	9	m			I			09.08.1988	EF WS	1580	n.a.	80000
30644	9	f					r	15.08.1988	EF WS	1310	n.a.	34000
30646	10	m	r					19.08.1988	EF WS	1430	n.a.	60000
30651	8	m			I			12.09.1988	EF WS	1500	n.a.	50000
30657	6	m	r	r				22.09.1988	EF WS	1290	n.a.	41000
30660	13	m	I			1		22.09.1988	EF WS	1400	n.a.	64000
30662	10	f				r		05.10.1988	EF WS	1330	n.a.	50000
30666	5	m						19.08.1988	EF WS	1400	n.a.	70000
30670	11	m			I			24.08.1988	EF WS	1500	n.a.	48000
30678	10	f				r		05.09.1988	EF WS	1450	n.a.	49000
30679	5	f	rl	r				24.08.1988	EF WS	1420	n.a.	52000

ZIK, Zoological Institute of Kiel University, Germany; I, from left maxilla; r, from right maxilla; I3, third incisor teeth; C, canine teeth; PC2, second postcanine teeth; PC3, third postcanine teeth; PC4, fourth postcanine teeth; m, male; f, female; NF WS, North Frisian Wadden Sea; EF WS, East Frisian Wadden Sea; n. a., not available. *Age at death was determined by cementochronology on canine roots (Abt, 2002).

two methods: (1) scale-sensitive fractal analysis (SSFA, four parameters, **Table 2**) using length-scale and area-scale fractal analyses (Ungar et al., 2003; Scott et al., 2006) and (2) three-dimensional surface texture analysis (3DST, 46 parameters, **Table 2**) using standardised roughness (ISO 25178) and flatness (ISO 12781) parameters combined with non-standardised further motif, furrows, direction and isotropy parameters (Schulz et al., 2010, 2013; Calandra et al., 2012). A full description of the 49 microwear texture parameters used is given in **Table 2**. The analysis was conducted using the μ soft analysis premium v.7.0.6672 software (NanoFocus AG, Oberhausen, Germany; a derivative of Mountains[®] Analysis software by Digital Surf, Besançon, France).

Statistics

The statistical analyses were carried out using the open-source software R versions 2.15.1 (R Development Core Team, 2011) following an established statistical procedure (Calandra et al., 2012). The R packages xlsx version 0.3.0 (Dragulescu, 2011), rJava (Urbanek, 2016), doBy version 4.5.9 (Højsgaard and Halekoh, 2013), grDevices version 2.15.1 (R Development Core Team,

2011) and R.utils version 1.9.11 (R Development Core Team, 2011) were used. All statistical tests were carried out using functions (Wilcox et al., 2005) which are included in the package WRS (Wilcox and Schönbrodt, 2010, v. 0.12.1). The groups tested were defined in four datasets according to the hypotheses (Table 3): sex (females versus males), age (5 years old versus 10 year old), season (summer = June/July versus fall = September/October); in each group sorted according to tooth position (I3/C/PC2/PC3/PC4); and additionally a pooled dataset with all specimens sorted according to tooth position (I3/C/PC2/PC3/PC4). All data were trimmed 15% in each tail to compensate for non-normality and heteroscedastic tests were applied due to heterogeneity of variances. The robust Welch-Yuen omnibus test (Welch, 1938; Yuen, 1974) was performed to test for significant differences (Supplementary Table 3) between the tested groups (Wilcox, 2003; Wilcox et al., 2005). Subsequently, the source of significant differences within the groups (arithmetical means) was determined using a heteroscedastic pairwise comparison test analogue to Dunnett's T3 ("Dunnett test" hereafter) (Dunnett, 1980). The results were accepted if Cliff's ordinal method ("Cliff test" hereafter)



FIGURE 2 | Harbour seal (*Phoca vitulina*) dentition and moulding process. (A) Skull in ventral and (B) lateral view showing the upper dentition. Tooth positions from mesial to distal: first incisor (I1), second incisor (I2), third incisor (I3), canine (C), first postcanine (PC1), second postcanine (PC2), third postcanine (PC3), fourth postcanine (PC4), fifth postcanine (PC5). Specimen 9343, curated at Center of Natural History, Hamburg, Germany. (C) Moulding process on the buccal side of PC2, PC3 and PC4 and (D) on C and I3. The short wire is oriented towards cusp tip, the long wire towards mesial. Specimen 30670, curated at the skeletal collection of the Zoological Institute, University of Kiel, Germany. Scale = 1 cm. (E) Buccal view of measurement area (coloured) on the third upper postcanine tooth of *Phoca vitulina*.

(Cliff, 1996), and the combination of the Welch-Yuen test and the Dunnett's test showed a significant output ($p \le 0.05$). We applied the combination of the three tests using robust means to control

over type I error (probability of detecting a false difference). This combination is similar to the *F*-test to test for group differences. Whenever a significant difference was found, the heteroscedastic

TABLE 2 Description of applied microwear texture parameters according to ISO 25178, ISO 12781, motif, furrow, texture direction, isotropy and SSFA.

ST	Description (condition)	Functional group	Standard	Unit
Asfc	Area-scale fractal complexity	complexity	SSFA	no unit
epLsar	Length-scale anisotropy of relief	direction	SSFA	no unit
FLTp	Peak to reference flatness deviation (Gaussian Filter,0.025 mm)	height	ISO 12781	μm
FLTq	Root mean square flatness deviation (Gaussian Filter, 0.025 mm)	height	ISO 12781	μm
FLTt	Peak to valley flatness deviation of the surface (Gaussian Filter, 0.025 mm)	height	ISO 12781	μm
FLTv	Reference to valley flatness deviation (Gaussian Filter, 0.025 mm)	height	ISO 12781	μm
Hasfc	Heterogeneity of area-scale fractal complexity	complexity	SSFA	no unit
IsT	Texture isotropy	direction	Isotropy	%
madf	Maximum depth of furrows	height	Furrow	μm
mea	Mean area	area	Motif	μm^2
medf	Mean density of furrows	density	Furrow	cm/cm ²
meh	Mean height	height	Motif	μm
metf	Mean depth of furrows	height	Furrow	μm
mev	Mean volume	volume	Motif	μm ³ /μm ²
nMotif	Number of motifs	complexity	Motif	no unit
S10z	Ten-point height	height	ISO 25178	μm
S5p	Five-point peak height	height	ISO 25178	μm
S5v	Five-point valley height	height	ISO 25178	μm
Sa	Mean surface roughness	height	ISO 25178	μm
Sal	Auto-correlation length (s = 0.2)	density	ISO 25178	μm
Sda	Closed dale area	area	ISO 25178	μm ²
Sda	Root mean square gradient	slope	ISO 25178	no unit
Sdr	Developed interfacial area ratio	complexity	ISO 25178	%
Sdv	Closed dale volume	volume	ISO 25178	μm ³
Sha	Closed hill area	area	ISO 25178	μm ²
Shv	Closed hill volume	volume	ISO 25178	µm ³
Sku	Kurtosis of the height distribution	height	ISO 25178	no unit
Smc	Inverse areal material ratio ($\rho = 10\%$)	plateau	ISO 25178	μm
Smfc	Scale of maximum complexity	complexity	SSFA	no unit
Smr	Areal material ration	plateau	ISO 25178	μm
Sp	Maximum peak height	height	ISO 25178	μm
, Spc	Arithmetic mean peak curvature	peak sharpness	ISO 25178	1/μm
Spd	Density of peaks	density	ISO 25178	1/µm ²
Sa	Standard deviation of the height distribution	height	ISO 25178	μm
, Ssk	Skewness of the height distribution	height	ISO 25178	no unit
Std	Texture direction	direction	ISO 25178	_
Str	Texture aspect ratio ($s = 0.2$)	direction	ISO 25178	no unit
Sv	Maximum pit height	height	ISO 25178	μm
Sxp	Peak extreme height	height	ISO 25178	μm
Sz	Maximum height	height	ISO 25178	μm
Tr1R	First direction	direction	Direction	-
Tr2R	Second direction	direction	Direction	no unit
Tr3R	Third direction	direction	Direction	-
Vm	Material volume at a given material ratio ($\rho = 10\%$)	volume	ISO 25178	μm ³ /μm ²
Vmc	Material volume of the core at given material ratio ($p = 10\%$, $q = 80\%$)	volume	ISO 25178	μm ³ /μm ²
Vmp	Material volume of the peaks	volume	ISO 25178	μm ³ /μm ²
Vv	Void volume at a given material ratio ($\rho = 10\%$)	volume	ISO 25178	μm ³ /μm ²
Vvc	Void volume of the core ($p = 10\%$, $q = 80\%$)	volume	ISO 25178	μm ³ /μm ²
Vvv	Void volume of the valley at a given material ratio ($p = 80\%$)	volume	ISO 25178	μm ³ /μm ²

pair-wise Dunnett's test was subsequently employed to reveal the source of difference. In order to control for type II error (probability of not detecting a genuine difference, related to the power of the test) we applied a heteroscedastic rank-based test that performs Cliff's test for all pairs, in which the family-wiseerror is controlled via Hochberg's method (Hochberg, 1988).

RESULTS

Descriptive statistical values, including the mean and standard derivation (SD) for the surface texture parameters are given in **Supplementary Tables 1–3**. The parameters are described in **Table 2**.

Intra-Individual Variation

The microwear texture along the tooth rows of individuals differentiated into two groups: Firstly, the mesial teeth (I3, C) with a less rough surface texture (only difference in one out of 49 parameters), and, secondly, the more rough distal

teeth (PC2, PC3, PC4) (no difference in 49 parameters), while I3 and PC3/4 differ in 33 out of 49 parameters. The distal teeth displayed a complex, anisotropic texture (*Asfc, Smfc, Sdr*) with a higher surface profile (*meh, FLTp, FLTq, FLTt, FLTv, S10z, S5p, S5v, Sa, Sp, Sq, Sv, Sxp, Sz, Smc, Smr*) featuring voluminous, more steep, pointed peaks (*Sdv, Sdq, Shv, Spc*) and deeper, denser furrows (*medf, metf, madf*) of greater volume (*Vm, Vmc, Vmp, Vv, Vvc, Sha*) (**Table 3** and **Figure 3**). The frontal tooth positions I3 and C were similar in their texture roughness. The only significant difference between I3 and C was the higher anisotropy (*epLsar*) in the texture of the canine teeth, rather resembling the postcanine

TABLE 3 Overview table of the tested hypotheses and the given statistical significant microwear texture parameters. **Hypotheses** Group 1 Group 2 Significant parameters nsp sex 13 male (n = 22)female (n = 22)Asfc. metf. S5p. Sda. Sdr. Sp 6 С male (n = 16)female (n = 16)PC₂ male (n = 21)female (n = 21)PC3 male (n = 16)female (n = 25)PC4 male (n = 18)female (n = 20)13 5 years (n = 14) 10 years age (n = 21)С 10 vears 5 vears (n = 11)(n = 19)PC₂ 10 years 5 years (n = 8)(n = 24)PC3 5 years (n = 12)10 years (n = 23)PC4 10 years 5 years (n = 13) (n = 19)13 season summer (n = 20)fall (n = 15)С summer (n = 16)fall (n = 14)PC2 summer (n = 17)fall (n = 15)PC3 summer (n = 16) fall (n = 19)PC4 summer (n = 17)fall (n = 15)intraindividual all specimens 13 (n = 35)C(n = 30)epLsar 1 I3 (n = 35)PC2 (n = 32)epLsar, FLTp, FLTq, FLTt, FLTv, madf, medf, meh, metf, S10z, S5p, 32 S5v, Sa, Sda, Sdq, Sdr, Sdv, Shv, Smc, Smr, Sp, Spd, Sq, Sv, Sxp, Sz, Vm, Vmc, Vmp, Vv, Vvc, Vvv 13 (n = 35)PC3 (n = 35)Asfc, epLsar, FLTp, FLTq, FLTt, FLTv, madf, medf, meh, metf, S10z, 33 S5v, Sa, Sda, Sdq, Sdr, Sdv, Shv, Smc, Smfc, Smr, Sp, Spd, Sq, Sv, Sxp, Sz, Vm, Vmc, Vmp, Vv, Vvc, Vvv 13 (n = 35)Asfc, epLsar, FLTp, FLTq, FLTt, FLTv, HAsfc, madf, medf, meh, PC4 (n = 32)33 metf, S10z, S5p, S5v, Sa, Sda, Sdq, Sdr, Sdv, Shv, Smc, Smr, Sp, Sq. Sv. Sxp. Sz. Vm. Vmc. Vmp. Vv. Vvc. Vvv C(n = 30)PC2 (n = 32) Asfc, FLTq, FLTt, FLTv, madf, medf, metf, S10z, S5p, S5v, Sa, Sda, 27 Sdq, Sdr, Sdv, Shv, Smc, Smr, Sp, Sq, Sv, Sxp, Sz, Vmc, Vv, Vvc, Vvv PC3 (n = 35)Asfc, FLTp, FLTq, FLTt, FLTv, madf, medf, metf, S10z, S5p, S5v, 31 C(n = 30)Sa, Sdq, Sdr, Sdv, Shv, Smc, Smfc, Smr, Sp, Spd, Sq, Sv, Sxp, Sz, Vm, Vmc, Vmp, Vv, Vvc, Vvv C(n = 30)PC4 (n = 32)Asfc, FLTp, FLTq, FLTt, FLTv, madf, medf, metf, S10z, S5p, S5v, 29 Sa, Sdq, Sdr, Sdv, Shv, Smc, Smr, Sp, Sq, Sv, Sxp, Sz, Vm, Vmc, Vmp, Vv, Vvc, Vvv PC2 (n = 32) PC3 (n = 35)PC4 (n = 32)PC2 (n = 32)PC3 (n = 35)PC4 (n = 32)

I3, third incisor teeth; C, canine teeth; PC2, second postcanine teeth; PC3, third postcanine teeth; PC4, fourth postcanine teeth; n, number of specimens; n_{sp}, number of significant parameters. For descriptions of microwear texture parameter, see **Table 2**.

dentition. A trend of increasing surface roughness along the tooth row from mesial to distal was found, with most voluminous furrows and highest peaks in the postcanine teeth (**Table 3**, **Figure 2**).

Individual Age

The microwear texture of the frontal dentition in harbour seals of different age classes appeared to be similar (Table 3 and Figure 3). The variability within the age classes is larger than the difference between 5- and 10-year old specimens (Supplementary Table 3). Regarding the intra-individual variation in the 10-year old group, it is notable that microwear textures were undistinguishable between PC2, PC3 and PC4. In contrast, in 5-year old individuals the microwear texture of the second postcanine was significantly less complex (Asfc) than in the third postcanine (Supplementary Table 3). In contrast to the I3 in 10 year olds, the second postcanines exhibited a higher surface profile (FLTq, FLTv, S10z, S5v, Sa, Smc, Smr, Sq, Sv, Sxp, Sz) with pointed peaks (Sdv, Shv) and deeper, denser furrows of greater volume (madf, metf, Vmc, Vv, Vvv) (differences in 19 out of 49 parameters), whereas in 5 year olds, no differences between PC2 and I3 were detected (Supplementary Table 3).

Sexual Segregation

Sexual differences were most prominent in the I3 with 6 out of 49 microwear texture parameters (**Table 3** and **Figure 3**). Regarding this tooth position, males (n = 22) displayed a microwear texture with higher complexity (*Asfc, Sdr*), higher pointy peaks (*S5p, Sdq, Sp*) as well as deeper furrows (*metf*) compared to females (n = 21) (**Table 3** and **Figure 3**). In both sexes, no intra-individual differences between I3 and C or between the postcanines were detected (**Supplementary Table 3**). Both within females and males, the frontal tooth positions were clearly distinguishable from the postcanines in their texture roughness. In females, the postcanines (PC3/PC4) were rougher than I3 (difference in 24/24 out of 49 parameters) (**Supplementary Table 3**). In contrast, the differences in males between I3 and PC3/PC4 were not as pronounced (difference in 9/1 out of 49 parameters).

Seasonality of Feeding Behaviour

We were unable to fully examine seasonal variations dependent on sex and age classes because of the sample size. When pooled, the results revealed no differences in texture roughness between different seasons (summer = June, July, August; fall = September, October) (Table 3). In comparison to the summer specimens (n = 28), the fall specimens (n = 24) displayed a tendency of a more exaggerated gradient from smoother surface texture in frontal teeth (I3 lower values for FLTt, FLTv, Smr, Sv) to rougher texture in distal teeth (PC2 higher values for FLTp, FLTt, Vm, *Vmp*) (Supplementary Table 3). Within the fall specimens, the most differences in microwear texture were between I3 and PC4 (23 parameters out of 49), with PC4 displaying a more complex texture (Asfc, epLsar, Sdr), a larger texture profile (FLTq, FLTt, FLTv, Smr, madf, metf, S5v, Sa, Sdq, Sq, Sv, Sxp) and more voluminous peaks and furrows (Sdv Shv, Vm, Vmc, Vmp, Vv, Vvc, Vvv) (Supplementary Table 3). Within summer specimens,

the differences between I3 and the postcanines were not as pronounced (9 parameters out of 49 between I3 and PC3).

DISCUSSION

Intra-Individual Variation

Distal teeth exhibited larger texture depths and feature volumes than frontal teeth. We interpret the gradual increase of enamel surface roughness along the dentition from mesial to distal as a biomechanical trend along the tooth row, due to the functional shifts. Our results are in line with former results in marine odontocete whales (Purnell et al., 2017), which found that dental microwear texture (especially Sa, Sg and Sku values) varies with tooth location in the jaw in marine odontocete whales. Harbour seals use their frontal teeth to capture prey, whereas the postcanines are only engaged, if prey items are too large to swallow in whole and need to be reoriented or fractured (Adam and Berta, 2002; Fahlke et al., 2013). The posterior teeth in mammals transmit higher bite forces due to their proximity to the mandibular condyle (Greaves, 2002, 2000; Santana and Dumont, 2009). This might result in the rougher microwear texture of distal teeth. In contrast, short capturing bites with the frontal dentition could have caused less roughness. Soft ingesta (with less ingesta abrasives) tend to smoothen the tooth surface of softfood feeders (Lucas et al., 2008). A similar effect for the surface topography but with a different cause was proposed (Kaiser et al., 2015) relating lower height and volume enamel surface texture parameters to a higher matrix resistance of the ingesta (independent of ingesta abrasives). The last case could be the situation we observed in harbour seals. Therefore, we consider it possible, that the piercing of soft fish might have led to more pronounced polishing in frontal teeth. We consider sediment, fish scales and exoskeletons to be the main abrasive agents in harbour seal ingesta. Previous studies on ungulates, rodents and primates attributed the main ingesta abrasiveness to airborne mineral dust and soil particles adherent to vegetation (Kaiser et al., 2018; Schulz-Kornas et al., 2019), which should play a minor role in a marine environment of the pelagic prey species. However, when hunting and capturing benthic prey species like sand eels (Ammodytidae), flatfish (Pleuronectiformes), and gobies (Gobiidae), the ingestion of additional sediment seems to be inevitable. The occasionally observed regurgitation behaviour in harbour seals has in fact been interpreted as a mechanism to expel swallowed sediment after feeding events targeting benthic prey (Bowen et al., 2002; Heithaus et al., 2009). It is known that quartz sand dominated most sediment sections of the German Wadden Sea (Volkman et al., 2000), but the indentation hardness of underwater quartz sand is unknown to us. According to nanoindentation studies, quartz dust (12.8 GPa) must be considered harder than dental enamel of several vertebrates, including humans (4.88 GPa), wild boars (Sus scrofa, 6.5 GPa) and dolphins (Steno bredanensis, 3.86 GPa, Pontoporia blainvillei, 2.36 GPa) (Loch and Simoes-Lopes, 2013; Lucas et al., 2013; Kaiser et al., 2018). However, the actual influence of sediment on the enamel microwear texture would be dependent on grain size, shape and quantity, as fine external abrasive have been reported



FIGURE 3 [Dental microwear texture in the upper dentition of *Phoca vitulina*. (**A**) Three-dimensional (3D) microwear textures in *Phoca vitulina* (ZIK-28348) for all examined tooth positions. The textures are sorted from mesial to distal along the upper dentition. Scale = 40 μ m. (**B**) Boxplots of selected surface texture parameters measured along the upper dentition of *Phoca vitulina* specimens at the age of 10 years (light green) and 5 years (dark green) as well as female (pink) and male (blue) specimens. The boxplots are sorted from mesial to distal teeth. Microwear texture parameters: *FLTt* = Peak to valley flatness deviation of the surface [μ m] (ISO 12781), *metf* = Mean depth of furrows [μ m] (Motif), *epLsar* = Length-scale anisotropy of the relief [no unit] (SSFA), *Vvv* = Void volume of the valley at a given material ratio ($\rho = 80\%$) [μ m³/ μ m²] (ISO 25178), *Smr* = Areal material ration [μ m] (ISO 25178), *Sxp* = Peak extreme height [μ m] (ISO 25178). Test statistics following the established protocol (Calandra et al., 2012) combining the Welch-Yuen-Test, the equivalent of Dunnett's T3-test, and Cliff's method; significance levels: * = $\rho \le 0.05$, ** = $\rho \le 0.05$, ** = $\rho \le 0.05$, Abbreviations for tooth positions from mesial to distal teeth: I3, third upper incisor; C, canine; PC2, second postcanine; PC3, third postcanine; PC4, fourth postcanine.

to cause a polishing effect (Ackermans et al., 2020; Winkler et al., 2020b).

Other than incidentally ingested sediment, we propose the scales of modern teleost fish to be a potential abrasive agent in both pelagic and benthic prey species. Teleost fish scales consist of tough collagen reinforced with hydroxyapatite crystallites (Ikoma et al., 2003; Vernerey and Barthelat, 2010; Zhu et al., 2013; Naleway et al., 2016). Pleuronectiformes have ctenoid scales exhibiting complex external rugosities and surface appendices that entrap additional abrasive particles (Spinner et al., 2016; Minicozzi et al., 2019). Those surface appendices are found to be the hardest part of the ctenoid scale in the common sole (Solea solea, 0.40 GPa, Spinner et al., 2019). The contact between accumulated abrasives (ctenoid scales, overlying sediment) and harbour seal teeth would be amplified by repeated contact, which would be necessary when handling larger, unwieldy fish (Bowen et al., 2002). Therefore, we consider cryptic flatfish to be a more abrasive prey item than demersal fish with smooth cycloid scales, f. e. gadoids and gobies. Pelagic fish with cycloid scales, like smelt, we regard as less abrasive prey.

The anisotropy (*epLsar*) in the third incisor (I3) was significantly lower than all other (more distal) tooth positions. The higher anisotropy values in the canine and postcanine dentition are likely due to numerous parallel scratches (**Figure 3A**). This finding is in accordance with results from odontocete whales (Fahlke et al., 2013), who investigated 2D dental microwear of extinct whales and modern marine mammals. According to their results, the enamel surfaces on cheek teeth of harbour seals and other pinnipeds featured many parallel to subparallel scratches compared to other marine mammals, indicating a straight ortho-retractional occlusal movement.

Individual Age

No differences in dental microwear texture between 5- and 10- year old animals were detectable This could not be related to a slight shift in prey spectrum, driven by increased body mass, resulting in a preferences for larger prey (Sievers, 1989; de la Vega et al., 2016). We assume that during the (relatively short) study period both age groups used similar available dietary sources. Therefore, a pooling of adult samples in further analyses of our study was justified. It is important to note that both groups can be considered adult individuals (Sievers, 1989) and diet items are likely similar in composition and might differ only in size (larger prey size in older animals (Behrends, 1985).

Apart from the general trend toward more texture roughness in younger adults, the microwear texture between both adult age groups can be considered similar and could be explained by similar body size. For future studies we recommend to test larger age ranges that differ more widely in body size; for example juvenile animals below 2 years in comparison to 10-year old adult individuals. It would be promising to test if similar like in Californian sea lions four to seven years old individuals have highest frequency of excessive tooth wear in the species age profile, which was interpreted as an indication of deficient feeding and/or chronic malnutrition (Labrada-Martagon et al., 2007). This would allow to test if hunting success between juvenile individuals (< 2 years) and older individuals (> 5 years) that differ more clearly in body size, is more reflected in microwear texture signals.

Sexual Segregation

We found differences in microwear texture roughness between males and females in the frontal I3 position (*Asfc, metf, S5p, Sdq, Sdr, Sp*); within females, there were more pronounced differences between frontal and distal teeth (I3 – PC3: 24 differences) than within males (I3 – PC3: 9 differences).

As the texture in frontal teeth reflects the initial contact with prev and/or sediment and as males showed more roughness, this might indicate a sex-specific foraging style or aggressive behaviour during male-male competitions and mating in the breeding season. In previous studies, DMTA was successfully used to detect significant sex-related differences in the diet of terrestrial mammals (Merceron et al., 2010). For harbour seals, sexual segregation has been documented in foraging area, traveling distances and diving patterns (Baechler et al., 2002). Sexual segregation of habitat use has been reported as mechanism to minimise the effect of consumptive competition in harbour seals and other pinnipeds (Herreman et al., 2009; Wilson et al., 2015). Dietary differences between sexes in the Norwegian population of the harbour seals were found (Herreman et al., 2009), which the authors attributed to increased competition and predation-risk. This resulted in longer travel distances for males and in nearshore hunting behaviour in females during pupping and weaning. Since the Norwegian harbour seal population belongs to the same subspecies (Phoca vitulina vitulina) as our study material (Bjørge et al., 2010), we infer a similar foraging behaviour for the individuals from the German Wadden Sea. Further sex-related dietary resource partitioning was reported for two fur seal species (Arctocephalus forsteri and Arctocephalus pusillus) as a potential for reducing foraging competition in nearby waters (Page et al., 2006). We conclude that the variation in dental microwear texture between harbour seal males and females is caused by a sexual segregation to reduce forage competition.

Seasonality of Feeding Behaviour

The enamel surfaces of summer and fall specimens were indistinguishable in their microwear texture. Within fall specimens there is a microwear texture gradient along the tooth row that might reflect a seasonal shift in prey availability (prey distribution and abundance, feeding/spawning activity, prey size), which has been reported in several dietary studies (Pierce et al., 1991; Hall et al., 1998; de la Vega, 2016). During the summer, pelagic fish are the most abundant in the Wadden Sea, however it is the benthic species, which are mostly consumed by harbour seals (de la Vega et al., 2016). During fall, a higher use of North Sea dietary resources has been reported (de la Vega et al., 2016). A large moving distance and increased range size has been reported for harbour seals from the Danish Kattegat region (Dietz et al., 2012). As a result, minor seasonal variations in enamel surface roughness could reflect either no shift in prey composition or a switch in high abrasive foraging habitat from coastal to less abrasive offshore foraging grounds that are too small to be detected by microwear texture.

Microwear Texture as a Tool to Access Ecosystem Changes

Our data showed the importance of standard measuring protocols since the tooth position has a significant effect on variability of the dietary signal. The postcanine dentition, preferably PC4 - PC2, proved most suitable to assess dietary signals; it was the least affected by age and sex. For future studies, we recommend to test frontal C tooth position to discern a wider range of foraging behaviour. Additionally, seal populations from different geographic regions should be investigated to infer interspecific dietary preferences related to prey species abundance in different habitats. This would allow tracking the feeding ecology in various seal populations and ecosystems. Following this direction by sorting harbour seal museum material from different periods and under various anthropogenic endangerment, long term dietary monitoring of harbour seals would be possible. We propose that in harbour seals microwear texture has a very interesting potential to be an indicator of a common dietary niche or specific example of the available food in addition to gut content analyses that allow a very particular short-term snapshot of diet. This could lead to a better understanding of critical extinction events and especially shed some light on the feeding ecology's role and habitat use.

CONCLUSION

This study successfully established a combination of 49 microwear texture parameters to quantify the enamel surface texture as an indicator of dietary behaviour in harbour seals. The buccal side of the upper teeth were selected for analysis and the second to fourth postcanine tooth positions were established as a reference. Increasing enamel texture roughness from frontal to distal was found along the tooth row. Compared to the abundant intra-individual differences, only small intraspecific differences dependent on age, sex and season were detected. Variations in microwear texture between ages were limited to the posterior teeth with a tendency toward more roughness in younger adults. The variations between sexes were more pronounced, with a decreasing trend from the most frontal tooth to the middle tooth positions and with slightly rougher microwear textures in males than in females. Minor seasonal variations were reflected in a stronger texture roughness gradient along the tooth row of fall specimens. Marine mammal collections contain useful records in detecting changes in food spectrum and prey availability. A better understanding of shifts in marine environments of the past might help employ conservation measurements for marine mammals in a changing environment. Future microwear texture studies on pinnipeds should adopt defined homogeneous parameters regarding tooth position and orientation to avoid obscuring dietary patterns with high variability linked to biomechanical intra-individual variation and to ensure comparability.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article and the **Supplementary Material** (**Supplementary Tables 1–4**). Additionally, the **Supplementary Template 1** and the surface measurements are accessible through the open source data repository of the University of Hamburg (template via doi: 10.25592/uhhfdm.8981, surface measurements (*.nms files) via doi: 10.25592/uhhfdm.8979).

ETHICS STATEMENT

Ethical review and approval was not required for the animal study because the dental specimen were parts of skeletal material in a bone collection in a Natural History Museum. No animal testing of living specimens was conducted.

AUTHOR CONTRIBUTIONS

TK and ES-K designed the study. EB collected and analysed the data and wrote the manuscript with input from all co-authors (ES-K, KL, US, TK). All authors contributed to the article and approved the submitted version

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fevo.2021. 644019/full#supplementary-material

Supplementary Figure 1 | Lateral view of the upper postcanine dentition of *Phoca vitulina* displaying abraded enamel and calcification. Exposed dentine mesially along the narrow ridge of the main cusp and distal-lingually on the smaller cusps marked by white arrows on PC2, PC3 and PC4 (from left to right). Calcified area on PC5 marked by black arrow. Scale = 1 cm.

Supplementary Table 1 | Test statistics given for microwear texture data sorted. I3, third incisor teeth; C, canine teeth; PC2, second postcanine teeth; PC3, third postcanine teeth; PC4, fourth postcanine teeth; n, number of specimens; ST, surface texture parameter; Ft/ph, test statistics; p, level of significance (bold values indicate significant levels < 0.05); nu1/nu2/df, degree of freedom; pl, lower 95% confidence interval; pu, upper 95% confidence interval; pcrit, critical significance level $\alpha = 0.05$; pcomp, 1 (p < pcrit) or 0 (p > pcrit). For descriptions of microwear texture parameter, see **Table 2**.

Supplementary Table 2 | Descriptive statistics (mean \pm SD) for all DMTA parameters for the upper dentition of *Phoca vitulina*. For descriptions of microwear texture parameter, see **Table 2**.

Supplementary Table 3 Overview table of the inter-individual variation within the tested groups and the given statistical significant microwear texture parameters. I3, third incisor teeth; C, canine teeth; PC2, second postcanine teeth; PC3, third

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postcanine teeth; PC4, fourth postcanine teeth; n, number of specimens; n_{sp} , number of significant parameters. For descriptions of microwear texture parameter, see **Table 2**.

Supplementary Table 4 | Raw dental microwear data for all 242 measured teeth. phvit, *Phoca vitulina*; ZIK, Zoological Institute of Kiel University, Germany; I3, third incisor teeth; C, canine teeth; PC2, second postcanine teeth; PC3, third postcanine teeth; PC4, fourth postcanine teeth; *n* Scans, number of valid scans included in analysis; Total length [mm], Weight [g]; For descriptions of parameters, see **Table 2**. Surface texture files are stored at the database of the public natural history museum (University of Hamburg, Center of Natural History Hamburg, mammal collection) and can be accessed via the open source data repository of the University of Hamburg (doi: 10.25592/uhhfdm.8979).

Supplementary Template 1 | Template to analyse the raw files indicating a detailed batch analyses to run pre-and post-processing filtering of the surface texture files as conducted in established protocols (Schulz et al., 2013, 2010). The template is accessible through the open-source data repository Zenodo: doi: 10.25592/uhhfdm.8981.

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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