



# Functional characterization of hypertrehalosemic hormone receptor in relation to hemolymph trehalose and to oxidative stress in the cockroach *Blattella germanica*

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Hypertrehalosemic hormone (HTH) is a peptide hormone that belongs to the adipokinetic hormone/red pigment concentrating hormone (AKH/RPCH) family, which exerts pleiotropic actions related to catabolic reaction and stress response. AKH peptides have been demonstrated to participate in stress response including oxidative stress in several insects. In order to study the signaling pathway of HTH involved in anti-oxidative stress, we have characterized a HIH receptor cDNA in *Blattella germanica* (*Blage-HTHR*) in structural and in functional terms using RNA interference (RNAi). *Blage-HTHR* is expressed in various female adult tissues (brain–CC–CA, ventral nerve cord, midgut, fat body, oviduct), but maximal expression is observed in the fat body. RNAi-mediated knockdown of *Blage-HTHR* expression results in a significantly lower level of hemolymph trehalose, even though HTH is exogenously administered. Paraquat elicits lethal oxidative stress in *B. germanica*, and co-injection of paraquat and HTH reduces this detrimental effect and extends the median survival time. Interestingly, the “rescue” effect of HTH on mortality caused by paraquat is diminished in specimens with depleted expression of *Blage-HTH* and *Blage-HTHR*. Finally, lipid peroxidation in the hemolymph increases 4 h after paraquat treatment, in comparison with control specimens or with HTH-treated specimens. However, lipid peroxidation induced by paraquat was not “rescued” by HTH in *Blage-HTH* and *Blage-HTHR* knockdown specimens. Our results demonstrate that HTH acts as a stress hormone mediating anti-oxidative protection in *B. germanica*, and that its receptor, *Blage-HTHR*, is essential for this action.

**Keywords:** adipokinetic hormone, paraquat, RNA interference, hypertrehalosemic hormone receptor, German cockroach

## INTRODUCTION

The adipokinetic hormone/red pigment concentrating hormone (AKH/RPCH) family is one of the extensively characterized group of neuropeptides whose members play important roles on energy homeostasis, as well as on other physiological and behavioral processes in insects (Schooley et al., 2005). AKH peptides are 8- to 10-amino acid long, with a pyroglutamate blocked N-terminus and an amidated C-terminus, aromatic residues at positions 4 and 8, and a glycine residue at position 9, when present (Gäde, 2009). Among them, the peptides that primarily induce trehalose efflux into hemolymph are referred to as a hypertrehalosemic hormone (HTH).

Recently, AKHs have been related to stress response, through mechanisms involving the stimulation of catabolic reactions while inhibiting synthetic reactions by increasing available energy (Kodrik, 2008). Earlier studies have related the release of AKHs as a result of insecticide poisoning through recording the increase of metabolites such as trehalose and lipids in the hemolymph of the cockroach *Periplaneta americana* (Granett and Leeling, 1972) and the locust *Schistocerca gregaria* (Samaranayaka, 1974). Direct measurement of AKH titer increase after insecticide exposure has

been further reported in a number of insects, including the locust *S. gregaria* and the bug *Pyrrhocoris apterus* (Candy, 2002; Kodrik and Socha, 2005; Velki et al., 2011). In addition, AKHs participate in anti-oxidative mechanisms by increasing reduced glutathione efflux and decreasing protein carbonyl levels in response to treatment with paraquat (PQ), a bipyridylum herbicide that is currently used to elicit oxidative stress in animals through redox-cycling reactions (Hassan, 1984); it has been used in this way, for example, in the potato beetle *Leptinotarsa decemlineata* (Kodrik et al., 2007) and in *P. apterus* (Vecera et al., 2007).

Oxidative stress is a type of stressor that causes neurodegeneration of brain cells under severe stress condition (Chaudhuri et al., 2007); it also affects a number of physiological processes, including an adaptive compensatory specific response of the organism that is activated to sustain homeostasis (Valko et al., 2007). Reactive oxygen species, such as superoxide anion radicals, hydroxyl radical, and hydrogen peroxide, represent a class of molecules that are derived from the normal metabolism of oxygen in the majority of aerobic organisms. Reactive oxygen species arise as endogenous by-products of metabolic reactions in the cells, and from exogenous exposure to environmental pollutants, ionizing radiation,

and others. Excess of reactive oxygen species and/or deficit of scavenging antioxidants results in oxidative stress, which may impair correct regulation of lipids, proteins, and DNA (Halliwell and Gutteridge, 2007).

Most of the previous studies show that AKHs are involved in antioxidant protection mechanisms used biochemical experimental approaches such as monitoring AKH titer changes or exogenously applying AKH under oxidative challenges (Kodrik et al., 2007; Vecera et al., 2007). Thus, little information is available at the molecular level on the relation of AKH with antioxidant protection mechanisms. In this context, we aim to provide molecular evidence that the signaling pathway of HTH is involved in the antioxidant protection mechanisms, using the cockroach *Blattella germanica* as model. Previously, we had cloned the hypertrehalosemic hormone (*Blage-HTH*) cDNA in *B. germanica* (Huang and Lee, 2011). In the present study we report the cloning of the cDNA of the HTH receptor (*Blage-HTHR*) and the effect of PQ when both *Blage-HTH* and *Blage-HTHR* expression are silenced by RNA interference (RNAi) in *B. germanica*, a species that is particularly sensitive to this knockdown technique (Belles, 2010).

## MATERIALS AND METHODS

### INSECTS

A colony of the German cockroach, *B. germanica* (L.), was maintained in environmental chambers at 28°C under L:D = 16:8 h conditions. Dog chow and water were provided *ad libitum*. Nymphs and adults were reared in groups, but adults were separated by sex to keep them in virgin state. Detailed information about rearing has been described previously (Lee and Wu, 1994).

### RNA EXTRACTION AND REVERSE TRANSCRIPTION TO cDNA

All RNA extractions were performed using TRIzol Reagent (Invitrogen) following the supplier's protocol. Possible genomic DNA contamination was removed by DNase I (Promega) treatment. One microgram of total RNA was reverse transcribed using High Capacity cDNA Reverse Transcription Kits (ABI).

### CLONING OF THE HTH RECEPTOR

The first primer set was designed based on the known AKH receptor sequences of other insects, including *P. americana* (DQ217786), *Apis mellifera* (AY898652), *Tribolium castaneum* (DQ422965), and *Bombyx mori* (AF403542). The primer sequences were as follows: 5'-TCC ACG TGG AGG AGC ACC C-3' forward; and, 5'-TTC TTC CAS STG GAG GRR CAC CCC-3' reverse. The amplified fragment (420 bp) was subcloned into the pGEMT-easy vector (Promega) and sequenced. To complete the sequence, the GeneRacer Kit (Invitrogen) for rapid amplification of cDNA ends (RACE) was used following the manufacturer's protocol. For 5'-RACE, the reverse primer was 5'-GAC TCA GCT GTA CCA GTT CTG GTA CAG GA-3', and for 3'-RACE, the forward primer was 5'-GTC TGA ACT GAG CTG ATA TCT GCG A-3'. All PCR products were subcloned into pGEMT-easy vector (Promega) and sequenced.

### PHYLOGENETIC ANALYSIS

The amino acid sequences of all known insect AKH receptors were obtained from GenBank and aligned using the Clustal X

1.18 program (Thompson et al., 1997). The resulting alignment was manually edited using the GeneDoc program (Nicholas et al., 1997) and analyzed by the Neighbor-joining method for construction of a phylogenetic tree. The AKH receptor of the insect species with the GenBank accession numbers are *Drosophila melanogaster* (AAF52426), *Anopheles gambiae* (ABD60146), *Aedes aegypti* (CAY77164 and CAY77166), *Glossina morsitans* (AEH25943), *B. mori* (AAL95712), *Manduca sexta* (ACE00761), *A. mellifera* (AAX83121), *Nasonia vitripennis* (NP\_001161243), *T. castaneum* (ABE02225), *Acyrtosiphon pisum* (XP\_001945436), *Pediculus humanus* (EEB15485), *P. americana* (ABB20590), and *B. germanica* (ADL60118). Five-letter abbreviations used for these AKH receptors and species are indicated in **Figure 1B**. Phylogenetic trees based on the resulting alignment were constructed using MEGA version 5 (Tamura et al., 2011). The gonadotropin-releasing hormone receptor of *Caenorhabditis elegans* (Cael-GnRHR, GenBank accession number NP\_506566) was used as outgroup. The robustness of phylogenetic analysis was evaluated by bootstrapping using 1000 replicates.

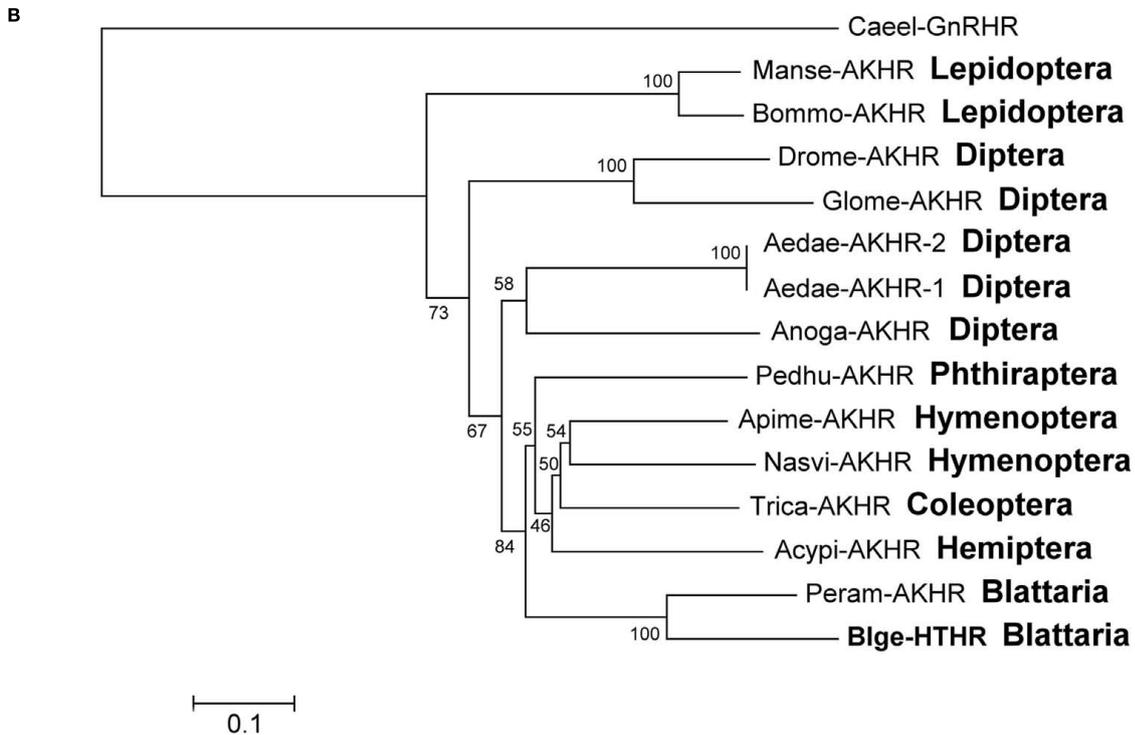
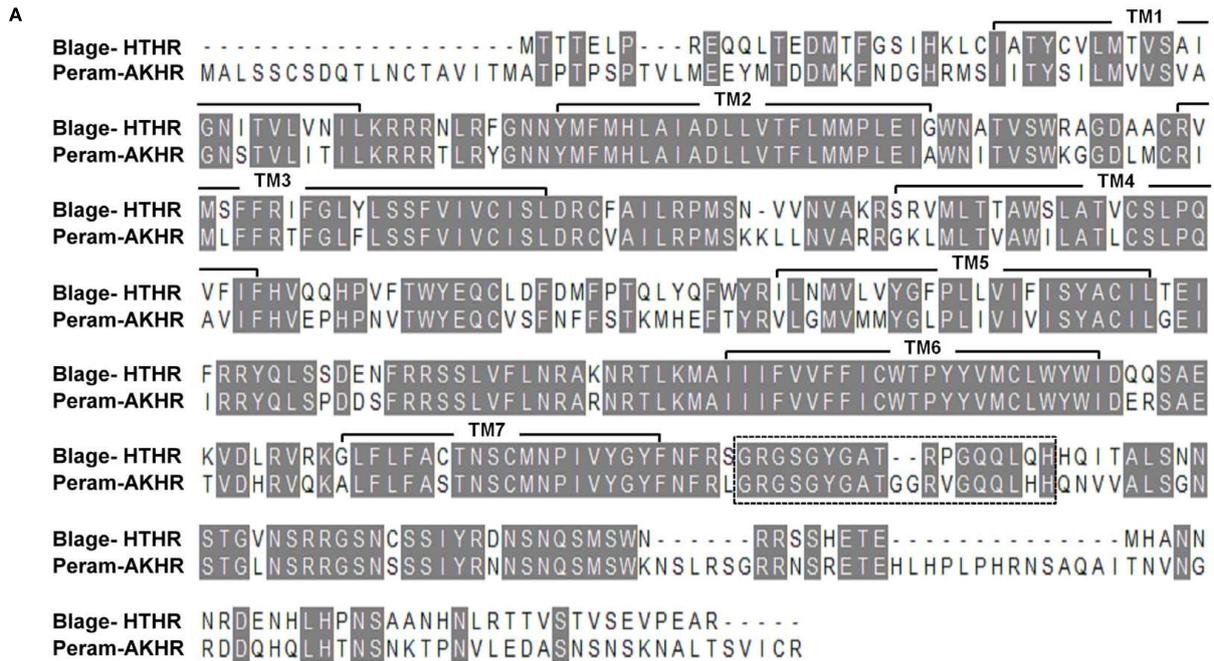
### BLAGE-HTHR EXPRESSION ANALYSIS

We performed semiquantitative RT-PCR to investigate the distribution of *Blage-HTHR* in different tissues of 7-day-old adult female *B. germanica* and to assess the effects of RNAi. Thus, the 25  $\mu$ l PCR mixture for amplifying the *Blage-HTHR* fragment included 1  $\mu$ l of cDNA, 10 pmol of forward primer (5'-GAC TCA GCT GTA CCA GTT CTG GTA-3'), 10 pmol of reverse primer (5'-GGG AAA TGT CTT GTG AAC CAG GTC-3'), and 5  $\mu$ l of Taq polymerase mixture (Protech). The PCR (30 cycles) was performed as follows: denaturation at 94°C for 30 s, annealing at 58°C for 30 s, extension at 72°C for 1 min. Expression of *BgActin* (GenBank EU514491) was used as reference. PCR products were separated and visualized on a 1.5% agarose gel.

Quantitative real-time PCR measurements were carried out in triplicate in Applied Biosystems StepOnePlus real-time PCR systems with Fast SYBR Green Master Mix (Applied Biosystems). The primers used to measure *Blage-HTHR* mRNA were as follows: 5'-TCT ATC GGG ACA ACA GCA ACC AGA-3' forward, and 5'-TGC TTC AGG AAC TTC ACT CAC AGT-3' reverse. The efficiency of this primer set was first validated by constructing a standard curve through five serial dilutions. The expression levels were normalized to *BgActin* expression using 2<sup>(-Delta Delta C(T))</sup> method (Pfaffl, 2001). The PCR program began with a single cycle at 95°C for 3 min, 40 cycles at 95°C for 15 s, and 60°C for 30 s. Afterward, PCR products were heated to 95°C for 15 s, cooled to 60°C for 30 s and increasing the temperature 0.5°C/min in order to measure the dissociation curves and to determine a unique PCR product for each gene. A template-free control was performed in each batch.

### RNA INTERFERENCE

The *Blage-HTHR* cDNA of the German cockroach was first cloned into the pGEMT-easy vector (Promega, Madison, WI, USA). Then, a 528 bp fragment targeting the 3'-UTR of *Blage-HTHR* mRNA was amplified by PCR. The primers (which contained the T7 promoter sequence: TAATACGACTCACTATAG) used in this PCR reaction were as follows: 5'-TGG ATA GAC CAG CAG TCT GCT



**FIGURE 1 | (A)** Alignment of the deduced amino acid sequences of the *Blattella germanica* HTH receptor (Blage-HTHR, GenBank GU591493) with the *Periplaneta americana* AKH receptor (Peram-AKHR, GenBank ABB20590). Identical amino acid residues are highlighted in gray. The seven transmembrane  $\alpha$ -helices are indicated by TM1-7. The dotted-line rectangle indicates an insertion present in both cockroach species. **(B)** Neighbor joining tree of AKH receptor insect sequences. The tree was generated using MEGA 5 with 1000 bootstrap replicates. The evolutionary distance is given in units of the number of amino acid substitutions per site. The

abbreviated names of insect AKHR are: Acypi-AKHR (*Acyrtosiphon pisum*), Aedae-AKHR-1 and Aedae-AKHR2 (*Aedes aegypti*), Anoga-AKHR (*Anopheles gambiae*), Apime-AKHR (*Apis mellifera*), Blage-HTHR (*Blattella germanica*), Bommo-AKHR (*Bombyx mori*), Drome-AKHR (*Drosophila melanogaster*), Glomo-AKHR (*Glossina morsitans*), Manse-AKHR (*Manduca sexta*), Nasvi-AKHR (*Nasonia vitripennis*), Pedhu-AKHR (*Pediculus humanus*), Peram-AKHR (*Periplaneta americana*), and Trica-AKHR (*Tribolium castaneum*). Gonadotropin-releasing hormone receptor of *Caenorhabditis elegans* (Caeel-GnRHR) was used as outgroup.

GAA-3' forward primer, and 5'-GGG AAA TGT CTT GTG AAC CAG GTC-3' reverse. The MEGAscript RNAi Kit (Ambion) was used to generate the dsRNA following the manufacturer's protocol. The dsRNA solution was stored at  $-20^{\circ}\text{C}$  until use. A dose of 1.5  $\mu\text{g}$  of the 528 bp dsRNA targeting the *Blage-HTHR* mRNA was injected into the abdomen of newly emerged adults. As dsRNA control, the enhanced green fluorescence protein (EGFP) sequence was used at the same dose. *Blage-HTHR* expression in the fat body was determined by semiquantitative RT-PCR within the following days after dsRNA treatment.

To investigate the role of HTH against oxidative stress, we injected 1.5  $\mu\text{g}$  of both, dsHTHR and dsHTH, the latter being prepared according to the protocol previously described (Huang and Lee, 2011), into newly emerged male *B. germanica*. *Blage-HTHR* expression in the fat body and *Blage-HTH* expression in the whole head were determined by semiquantitative RT-PCR at the 10th day after dsRNA treatment.

### TREHALOSE DETERMINATION

Trehalose in the hemolymph was determined following the protocol previously described (Huang and Lee, 2011). Briefly, 5  $\mu\text{l}$  hemolymph was obtained from the hind leg coxae. Then, the hemolymph was incubated overnight at  $37^{\circ}\text{C}$  with 2  $\mu\text{l}$  porcine kidney trehalase (Sigma) to convert trehalose into glucose. The amount of glucose in 50  $\mu\text{l}$  of the above solution was measured using the Glucose Assay Kit (Sigma). Glucose concentration was corrected by deducting the amount of glucose from the same supernatant prepared under identical conditions but without trehalase.

### OXIDATIVE STRESS TREATMENTS

The insects were individually injected with 1  $\mu\text{l}$  solution of 20 mM PQ (paraquat: 1,1'-dimethyl-4,4'-bipyridylum dichloride hydrate) to cause oxidative stress, or with 1  $\mu\text{l}$  Ringer saline as control. To study the effect of HTH against oxidative stress, the insects were injected with 1  $\mu\text{l}$  solution containing synthetic HTH peptide (pGlu-Val-Asn-Phe-Ser-Pro-Gly-Trp-Gly-Thr-NH<sub>2</sub>) of *B. germanica* at various concentrations (10, 40, and 80 pmol) in combination with 20 nmol PQ (1:1 in volume). The number of dead cockroaches were recorded every day. All chemicals used in the experiments and assays were obtained from Sigma-Aldrich Chemical Company (St. Louis, MO, USA) unless otherwise stated.

### LIPID PEROXIDATION MEASUREMENT

We used lipid peroxidation as specific test to measure the level of oxidative stress. A total of 15  $\mu\text{l}$  hemolymph from a pool of eight specimens was collected at 4 h post inoculation of oxidative stress treatment and diluted with 30  $\mu\text{l}$  PBS containing 1% butylated hydroxytoluene to prevent further oxidation of lipid during further processing. The diluted hemolymph was shaken vigorously and centrifuged ( $10,000 \times g$  for 10 min at  $4^{\circ}\text{C}$ ). To determine the levels of malondialdehyde as indicator of lipid peroxidation, the supernatants were processed using the OxiSelect TBARS (thiobarbituric acid reactive substances) assay kit (Cell Biolabs) following the manufacturer's instructions.

### STATISTICAL ANALYSIS

The results in the figures represent the mean  $\pm$  SEM. Statistical analyses were performed using computing software R. Statistical significance of results were evaluated using unpaired *t*-test between two groups and one-way ANOVA followed by Tukey's HSD *post hoc* test for multiple comparisons. The analyses of survival curves were carried out using Kaplan–Meier test for median survival time, the Log-rank test for testing the differences among treatment groups, and the Kruskal–Wallis rank sum test for post-test calculations.

## RESULTS

### CLONING AND EXPRESSION PATTERN OF *BLAGE-HTH* RECEPTOR

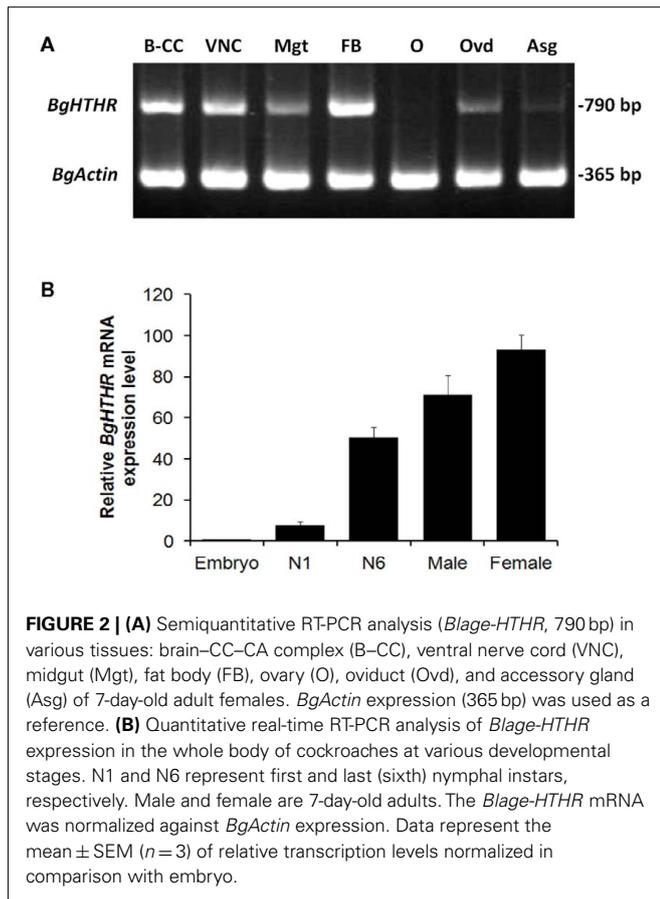
Cloning of the HTH receptor (*Blage-HTHR*) cDNA of *B. germanica* was accomplished by a RT-PCR approach, combining the use of degenerate primers based on AKHR conserved motifs to obtain a partial sequence, and 5'-RACE and 3'-RACE experiments to complete it. These amplifications rendered a full-length cDNA of 1,769 bp encoding a predicted 406 amino acid protein (*Blage-HTHR*; GenBank accession number GU591493; **Figure A1** in Appendix). According to BLAST searches, cDNA sequence of *Blage-HTHR* was very close (71.0% identity, 81.5% similarity) to *P. americana* AKH receptor (*Peram-AKHR*; GenBank accession number ABB20590). In addition, the alignment of receptor proteins of the two cockroach species showed values of 62.4% identity and 72% similarity (**Figure 1A**). The deduced amino acid sequence of *Blage-HTHR* exhibits the typical structural features of G protein-coupled receptors including an extracellular N-terminus, seven transmembrane regions, and an intracellular C-terminus.

Neighbor-joining analysis of insect AKH receptor sequences gave a tree (**Figure 1B**) showing that *Blage-HTHR* clusters with *Peram-AKHR* from *P. americana*, as expected. Indeed, these two cockroach receptors have an additional consensus sequence in the intracellular C-terminus that is characteristic of them, and which is not present in other insect AKH receptor sequences (**Figure A2** in Appendix). The sister group of the cockroach node is composed by phylogenetically distal hemimetabolous species (Phthiraptera, Hemiptera) and basal holometabolous species (Coleoptera, Hymenoptera), and these two nodes have the more modified holometabolous species (Lepidoptera and Diptera) as sister group. In general, the topology of the tree approximately follows the phylogenetic relationships of the insect orders currently established.

The expression of *Blage-HTHR* was detected and quantified in various tissues and stages (**Figure 2**). Concerning the adult, the expression was observed in practically all studied tissues, but the strongest expression was found in the fat body (**Figure 2A**). Conversely, expression in the ovary was practically absent, and that in the accessory glands was very faint. Comparing different life cycle stages (**Figure 2B**), expression was very low in embryos and relatively low in the first instar nymph. Values were much higher in the last instar (N6) nymph, and in the adult stage of both sexes.

### RNAI-MEDIATED KNOCKDOWN OF *BLAGE-HTHR* REDUCES THE TREHALOSEMIC EFFECT OF HTH

We first studied the function of *Blage-HTHR* in *B. germanica* in relation to trehalose regulation by depleting its expression by

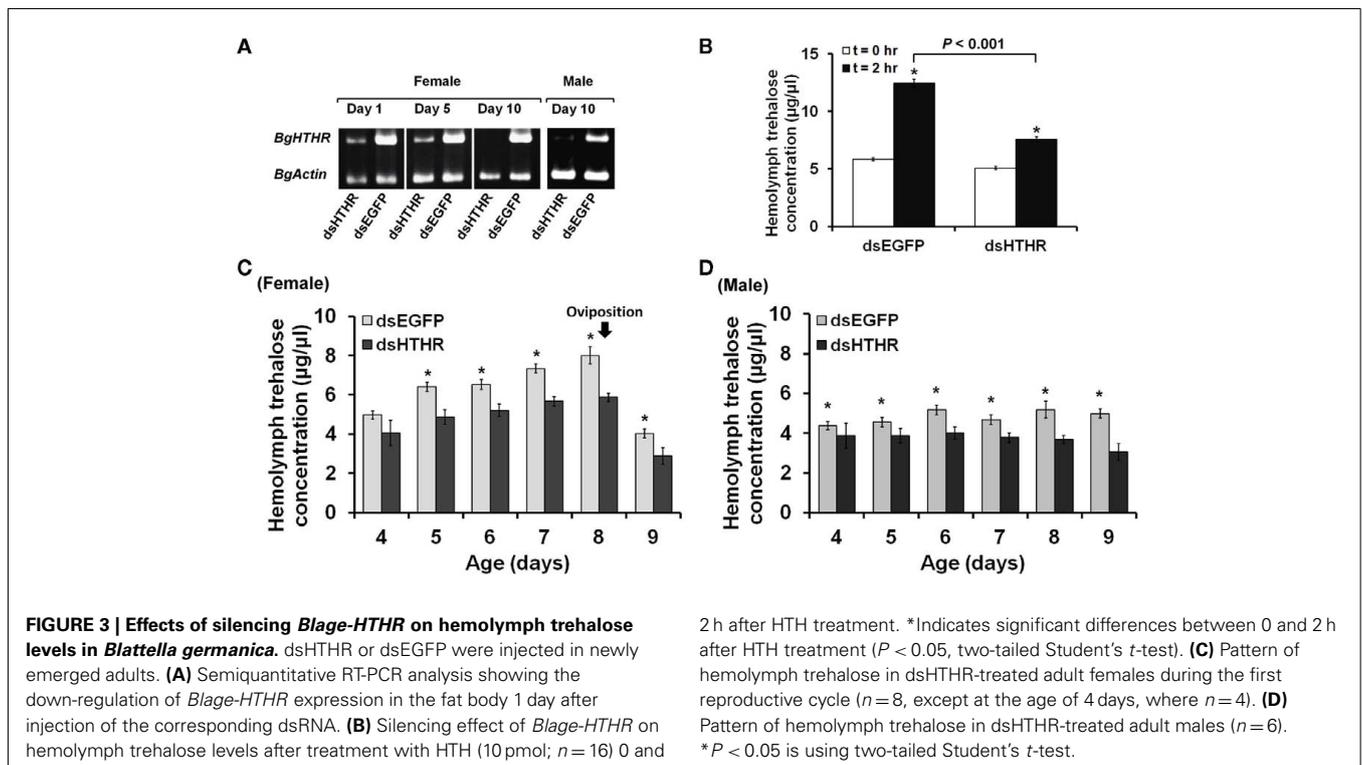


RNAi. *Blage-HTHR* mRNA was measured in the fat body 1, 5, and 10 days after the treatment with dsHTHR, and results (**Figure 3A**) showed that expression was clearly reduced in comparison with controls (dsEGFP-treated) on days 1 and 5, and it was practically undetectable on day 10, either in females as well as in males. The function of *Blage-HTHR* as HTH receptor was illustrated by the significant reduction of trehalose titers in the hemolymph after injection of HTH in dsHTHR-treated females (**Figure 3B**). The injection of 10 pmol of HTH resulted in the increase of hemolymph trehalose up to 113% 2 h later in dsEGFP-treated females. In contrast, the hemolymph trehalose increased significantly less (49%) in dsHTHR-treated females 2 h after the HTH injection. These results suggest that the remarkable increase of hemolymph trehalose elicited by HTH requires a high expression of its receptor, *Blage-HTHR*.

The titer of hemolymph trehalose displayed a steady increase and peaked at day 8 in the first reproductive cycle of female adults, either in controls (dsEGFP-treated) as in specimens treated with dsHTHR (**Figure 3C**), although the levels are lower in the latter. The hemolymph trehalose pattern is coincident with that of ovarian development. In adult males, the hemolymph trehalose levels practically do not fluctuate, either in controls (dsEGFP-treated) as in specimens treated with dsHTHR (**Figure 3D**); again, the levels were significantly lower in dsHTHR-treated specimens (**Figure 3D**).

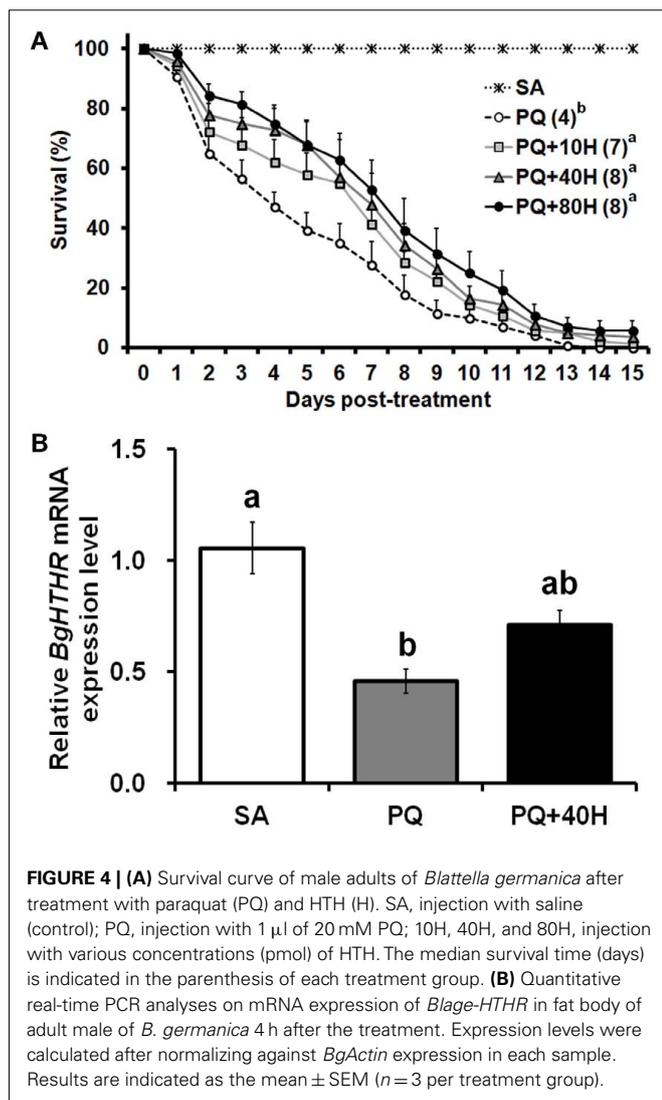
**ROLE OF HTH AS RELIEVER OF OXIDATIVE STRESS**

Subsequently, we studied the potential roles of HTH as reliever of oxidative stress. Given that female *B. germanica* exhibits a varied ability against oxidative stress during the reproductive cycle



(unpublished data), we chose only males to conduct the following experiments. The injection of 20 nmol of PQ on 10-day-old male adults caused a mortality that rapidly increased with time, thus median survival time was attained on day 4 (**Figure 4A**). However, exogenous HTH at 10, 40, and 80 pmol concentrations significantly reduced the speed of mortality increase, with median survival time attained around day 7 (**Figure 4A**), thus, the protective effect of HTH did not show significant differences among the three concentrations used.

To further assess the possible protective role of HTH against oxidative stress, the expression of *Blage-HTHR* after PQ treatment was studied (**Figure 4B**). Injection of PQ (20 nmol) caused a significant reduction of the *Blage-HTHR* expression in the fat body 4 h later. Mean value of *Blage-HTHR* mRNA was also decreased in the specimens injected with PQ and 40 pmol HTH, which extended the life span under the provoked oxidative stress. But the level of *Blage-HTHR* in PQ co-injected with HTH was not significantly different from that in either control or PQ treatment.



## RNAi-SILENCED HTH SIGNALING REDUCES ANTI-OXIDATIVE PROTECTION

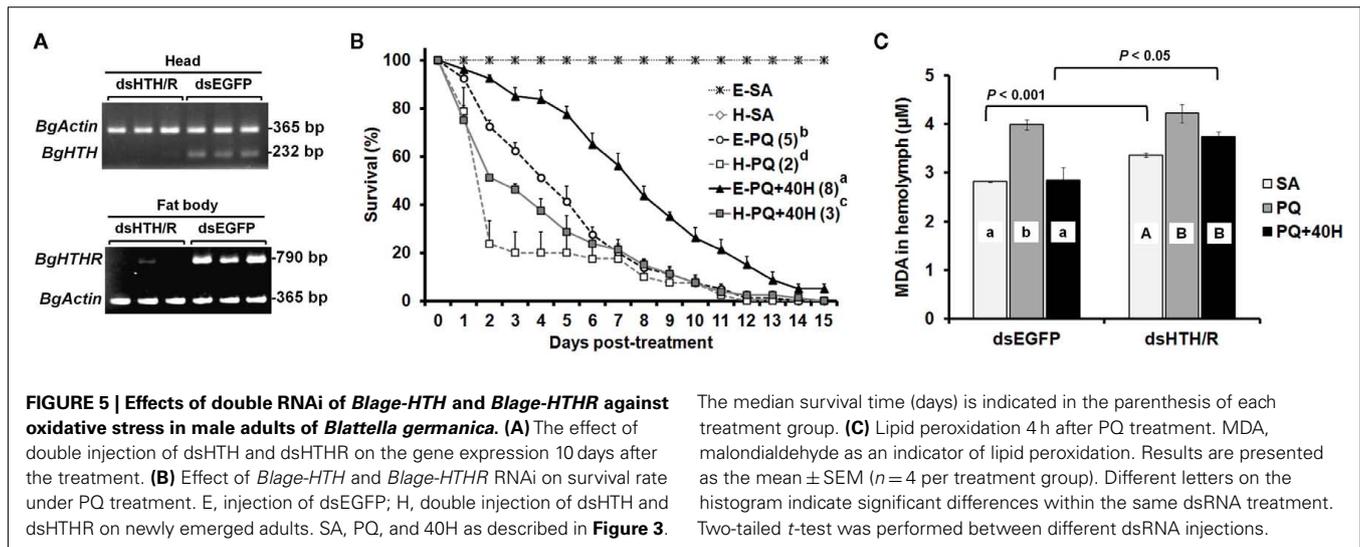
We further assessed the protective role of HTH on oxidative stress in specimens where *Blage-HTH* and *Blage-HTHR* mRNA levels had been simultaneously depleted by RNAi. A treatment of 1.5  $\mu$ g of dsHTH and 1.5  $\mu$ g of dsHTHR (dsHTH/R) was administered to newly emerged males of *B. germanica*, and the corresponding expression of *Blage-HTH* and *Blage-HTHR* was undetectable 10 days later in the tissues where these transcripts are most expressed: *Blage-HTH* in the head and *Blage-HTHR* in the fat body (**Figure 5A**). There were no lethal effects when the expression of both genes was knocked down, as all specimens survived in apparently healthy conditions at least during 25 days after the dsRNA injections. However, PQ treatment elicited a faster mortality in the specimens knockdown for *Blage-HTH* and *Blage-HTHR* (median survival time was attained on day 2 after PQ treatment), than in controls (dsEGFP-treated; median survival time attained on day 5; **Figure 5B**). Moreover, a dose of 40 pmol of HTH significantly relieved the detrimental effect elicited by PQ in control specimens (median survival time attained on day 8), but this “rescue” effect of HTH was milder in the specimens knockdown for *Blage-HTH* and *Blage-HTHR* (median survival time attained on day 3; **Figure 5B**).

Finally, the anti-oxidative stress function of *Blage-HTH* and *Blage-HTHR* was further demonstrated by measuring malondialdehyde as indicator of lipid peroxidation under oxidative stress caused by PQ, using specimens where *Blage-HTH* and *Blage-HTHR* expression had been depleted by RNAi, and applying exogenous HTH. Results (**Figure 5C**) show that without PQ, the malondialdehyde titer in the group of specimens knockdown for *Blage-HTH* and *Blage-HTHR* was higher than that of the dsEGFP-treated group. PQ elicited and increased the titer of malondialdehyde in both groups, but differences between them were not statistically significant. Finally, exogenous HTH significantly counteracted the effects of PQ in the dsEGFP-treated group, but not in the group of *Blage-HTH* and *Blage-HTHR* knockdowns.

## DISCUSSION

About 50 peptides belonging to the AKH family have been identified in insects (Gäde, 2009) since the first hyperglycemic factor was discovered in the cockroach *P. americana* (Steele, 1961). The physiological function of these peptides on energy homeostasis as well as on signal transduction at the cellular level have been thoroughly reviewed by a number of authors (Van der Horst et al., 2001; Gäde and Auerswald, 2003; Schooley et al., 2005). However, studies at molecular level of AKH receptors have been conducted more recently in the fruit fly *D. melanogaster* and in the silkworm *B. mori* (Staubli et al., 2002). In cockroaches, only the AKH receptor cDNA of *P. americana* has been characterized (Hansen et al., 2006; Wicher et al., 2006). The present contribution reports the characterization of the HTH receptor in another cockroach, *B. germanica*, which is structurally close to that of *P. americana* AKH receptor (**Figure 1**).

Alignment of all known AKH/HTH receptor protein sequences shows a large region of evolutionary conserved residues in the transmembrane domain, but a highly diversified region in the C-terminus, which is associated to intracellular signaling (**Figure A2**



in Appendix). The phylogenetic analysis indicates that the intracellular C-terminus has considerably diverged in different insect orders, and this might have derived in different signal transduction mechanisms in different orders. Interestingly, an additional sequence at the beginning of the intracellular C-terminus in cockroaches might be related to the export of the receptor to the cell membrane, as an equivalent sequence in the C-terminal end of the AKH receptor of *B. mori* serves for this function (Huang et al., 2011).

The most abundant expression of *Blage-HTHR* in the fat body among the tissues tested (**Figure 2A**) is in agreement with the fact that HTH is the major regulator of energy metabolism in insects. This is also supported by the increasing expression level of *Blage-HTHR* from nymphal to adult stage which coincides with increasing locomotion which occurs in parallel (Yang et al., 2009), and which requires increasing energy expenditure. Our previous results showed that depletion of *Blage-HTH* expression results in a delay of oviposition in virgin females of *B. germanica* (Huang and Lee, 2011). In the present study, we observed *Blage-HTHR* expression in the oviduct, but not in ovary (**Figure 2A**), which suggests that *Blage-HTH* release could stimulate the contraction of oviduct muscles during basal oocyte extrusion. Indeed, a myoactive effect of AKHs on the dorsal vessel has been demonstrated in *P. americana* (Scarborough et al., 1984).

The canonical function of *Blage-HTHR* as HTH receptor in *B. germanica* has been demonstrated by injecting HTH in specimens where *Blage-HTHR* had been depleted by RNAi, and observing that mobilization of trehalose in the hemolymph was impaired (**Figure 3B**). Equivalently, the malaria mosquito *A. gambiae* with lowered expression of AKH receptor (Anoga-AKHR) also fails to mobilize the glycogen reserves in the fat body after injection of Anoga-AKH-I (Kaufmann and Brown, 2008). However, the occurrence of a significant elevation of hemolymph trehalose in dsHTHR-treated group was likely considered as incomplete silencing of *Blage-HTHR* expression because RNAi does not completely knockout gene expression. In addition, studies *in vitro* demonstrate that reduced

expression of the *Bombyx* AKHR on the cell membrane results in lower levels of intracellular cAMP (cyclic adenosine monophosphate) formation after AKH stimulation (Huang et al., 2011). Therefore, a maximal action of HTH would require an adequate expression of its receptor to trigger signal transduction.

RNA interference-mediated depletion of *Blage-HTHR* mRNA was found to decrease hemolymph trehalose levels in the adult of both sexes (**Figures 3C,D**), although the female displays fluctuations of trehalose related to the reproductive cycle. Our present results of *Blage-HTHR* RNAi (**Figure 3C**) supports our previous hypothesis postulating that HTH induces hemolymph trehalose mobilization during the reproductive cycle in virgin females (Huang and Lee, 2011). The results suggest that HTH plays a critical role in carbohydrate homeostasis, as occurs in AKH-cell-deficient *D. melanogaster* that has relatively low hemolymph trehalose levels (Lee and Park, 2004).

The involvement of AKH peptides in antioxidant protection in insects was inferred from the increase in AKH titer after exposure to oxidative stress (Kodrik et al., 2007; Vecera et al., 2007). The same authors showed that exogenous AKH application enhanced the efflux of reduced glutathione to the hemolymph and maintained the normal levels of protein carbonyl, a marker of oxidative stress. In the present study, we have fully demonstrated the protective role of HTH upon oxidative stress under different experimental approaches, first, through experiments of co-injection of PQ and HTH on control specimens (**Figure 4A**), and secondly, in specimens with the expression levels of *Blage-HTH* and *Blage-HTHR* depleted by RNAi (**Figure 5B**). These results provide a new and relevant information supporting that *Blage-HTH* potentiates a stress hormone on anti-oxidative stress response in *B. germanica*. The promoter of oxidative stress used in our experiments has been PQ, which acts on the redox-cycling reactions, thus eliciting free radicals as well as causes oxidative modification of macromolecules, such as lipid peroxidation (Hassan, 1984). Accordingly, we examined lipid peroxidation in the hemolymph 4 h after PQ treatment (**Figure 5C**). Levels of lipid

peroxidation in the hemolymph increased significantly after PQ injection in dsEGFP and dsHTH/R groups. However, the co-injection of HTH maintained the lipid peroxidation at a normal level in controls but not in the *Blage-HTH* and *Blage-HTHR* knockdowns, which suggests that HTH lowers the oxidative damage in *B. germanica*. A likely mechanism might be that HTH induces the efflux of glutathione into the hemolymph, as demonstrated in other insect AKHs (Kodrik et al., 2007; Vecera et al., 2007). However, the significant reduction of *Blage-HTHR* expression after PQ treatment (Figure 4B), suggests that the oxidative stress directly impairs its expression. The less severe reduction of *Blage-HTHR* expression observed after co-injecting PQ and HTH (Figure 4B) might be the outcome of downstream response induced by HTH to relieve the oxidative damage. Moreover, genome-wide analysis in *D. melanogaster* after PQ treatment have shown that cytochrome P450 genes become up- and

down-regulated (Girardot et al., 2004), and it has been additionally reported that HTH stimulates cytochrome P450 expression in the cockroach *Blaberus discoidalis* (Lu et al., 1995). Therefore, we further speculate that HTH might induce the expression of P450 genes, thus contributing to decrease the oxidative stress in *B. germanica*.

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## REFERENCES

- Belles, X. (2010). Beyond *Drosophila*: RNAi in vivo and functional genomics in insects. *Annu. Rev. Entomol.* 55, 111–128.
- Candy, D. J. (2002). Adipokinetic hormones concentrations in the haemolymph of *Schistocerca gregaria*, measured by radioimmunoassay. *Insect Biochem. Mol. Biol.* 32, 1361–1367.
- Chaudhuri, A., Bowling, K., Funderburk, C., Lawal, H., Inamdar, A., Wang, Z., and O'Donnell, J. M. (2007). Interaction of genetic and environmental factors in a *Drosophila* parkinsonism model. *J. Neurosci.* 27, 2457–2467.
- Gäde, G. (2009). Peptides of the adipokinetic hormone/red pigment-concentrating hormone family: a new take on biodiversity. *Ann. N. Y. Acad. Sci.* 1163, 125–136.
- Gäde, G., and Auerswald, L. (2003). Mode of action of neuropeptides from the adipokinetic hormone family. *Gen. Comp. Endocrinol.* 132, 10–20.
- Girardot, F., Monnier, V., and Tricoire, H. (2004). Genome wide analysis of common and specific stress responses in adult *Drosophila melanogaster*. *BMC Genomics* 5, 74. doi:10.1186/1471-2164-5-74
- Granett, J., and Leeling, N. C. (1972). A hyperglycemic agent in the serum of DDT-prostrate American cockroaches, *Periplaneta americana*. *Ann. Entomol. Soc. Am.* 65, 299–302.
- Halliwell, B., and Gutteridge, J. M. C. (2007). *Free Radicals in Biology and Medicine*, 4th Edn. Oxford: Clarendon Press.
- Hansen, K. K., Hauser, F., Cazzamali, G., Williamson, M., and Grimmelikhuijzen, C. J. P. (2006). Cloning and characterization of the adipokinetic hormone receptor from the cockroach *Periplaneta americana*. *Biochem. Biophys. Res. Commun.* 343, 638–643.
- Hassan, H. M. (1984). Exacerbation of superoxide radical formation by paraquat. *Meth. Enzymol.* 105, 523–532.
- Huang, H., Deng, X., He, X., Yang, W., Li, G., Shi, Y., Shi, L., Mei, L., Gao, J., and Zhou, N. (2011). Identification of distinct c-terminal domains of the Bombyx adipokinetic hormone receptor that are essential for receptor export, phosphorylation and internalization. *Cell. Signal.* 23, 1455–1465.
- Huang, J.-H., and Lee, H.-J. (2011). RNA interference unveils functions of the hypertrehalosemic hormone on cyclic fluctuation of hemolymph trehalose and oviposition in the virgin female *Blattella germanica*. *J. Insect Physiol.* 57, 858–864.
- Kaufmann, C., and Brown, M. R. (2008). Regulation of carbohydrate metabolism and flight performance by a hypertrehalosaemic hormone in the mosquito *Anopheles gambiae*. *J. Insect Physiol.* 54, 367–377.
- Kodrik, D. (2008). Adipokinetic hormone functions that are not associated with insect flight. *Physiol. Entomol.* 33, 171–180.
- Kodrik, D., Krishnan, N., and Habus-tova, O. (2007). Is the titer of adipokinetic peptides in *Leptinotarsa decemlineata* fed on genetically modified potatoes increased by oxidative stress? *Peptides* 28, 974–980.
- Kodrik, D., and Socha, R. (2005). The effect of insecticide on adipokinetic hormone titre in the insect body. *Pest Manag. Sci.* 61, 1077–1082.
- Lee, G., and Park, J. H. (2004). Hemolymph sugar homeostasis and starvation-induced hyperactivity affected by genetic manipulations of the adipokinetic hormone-encoding gene in *Drosophila melanogaster*. *Genetics* 167, 311–323.
- Lee, H. J., and Wu, Y. L. (1994). Mating effects on the feeding and locomotion of the German cockroach, *Blattella germanica*. *Physiol. Entomol.* 19, 39–45.
- Lu, K.-H., Bradfield, J. Y., and Keeley, L. L. (1995). Hypertrehalosemic hormone-regulated gene expression for cytochrome P4504C1 in the fat body of the cockroach *Blaberus discoidalis*. *Arch. Insect Biochem. Physiol.* 28, 79–90.
- Nicholas, K. B., Nicholas, H. B., and Deerfield, D. W. (1997). GeneDoc: analysis and visualization of genetic variation. *Embnet News* 4, 14.
- Pfaffl, M. W. (2001). A new mathematical model for relative quantification in real-time RT-PCR. *Nucleic Acids Res.* 29, e45.
- Samaranayaka, M. (1974). Insecticide-induced release of hyperglycaemic and adipokinetic hormones of *Schistocerca gregaria*. *Gen. Comp. Endocrinol.* 24, 424–436.
- Scarborough, R. M., Jamieson, G. C., Kalish, F., Kramer, S. J., McEnroe, G. A., Miller, C. A., and Schooley, D. A. (1984). Isolation and primary structure of two peptides with cardioacceleratory and hyperglycemic activity from the corpora cardiaca of *Periplaneta americana*. *Proc. Natl. Acad. Sci. U.S.A.* 81, 5575–5579.
- Schooley, D. A., Horodyski, F. M., and Coast, G. M. (2005). "Hormones controlling homeostasis in insects," in *Comprehensive Molecular Insect Science*, eds L. I. Gilbert, K. Iatrou, and S. S. Gill (Amsterdam: Elsevier Press), 493–550.
- Staubli, F., Jorgensen, T. J. D., Cazzamali, G., Williamson, M., Lenz, C., Sondergaard, L., Roepstorff, P., and Grimmelikhuijzen, C. J. P. (2002). Molecular identification of the insect adipokinetic hormone receptors. *Proc. Natl. Acad. Sci. U.S.A.* 99, 3446–3451.
- Steele, J. E. (1961). Occurrence of a hyperglycaemic factor in the corpus cardiacum of an insect. *Nature* 192, 680–681.
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., and Kumar, S. (2011). MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol. Biol. Evol.* 28, 2731–2739.
- Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F., and Higgins, D. G. (1997). The CLUSTAL\_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res.* 25, 4876–4882.
- Valko, M., Leibfritz, D., Moncol, J., Cronin, M. T., Mazur, M., and Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem. Cell Biol.* 39, 44–84.
- Van der Horst, D. J., Van Marrewijk, W. J. A., and Diederens, J. H. B. (2001). "Adipokinetic hormones of insect: release, signal transduction, and responses," in *International Review of Cytology – A Survey of Cell Biology*, ed. K. W. Jeon (San Diego, CA: Academic Press), 179–240.
- Vecera, J., Krishnan, N., Alquicer, G., Kodrik, D., and Socha, R. (2007). Adipokinetic hormone-induced enhancement of antioxidant capacity of *Pyrrhocoris apterus* hemolymph in response to oxidative stress. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* 146, 336–342.

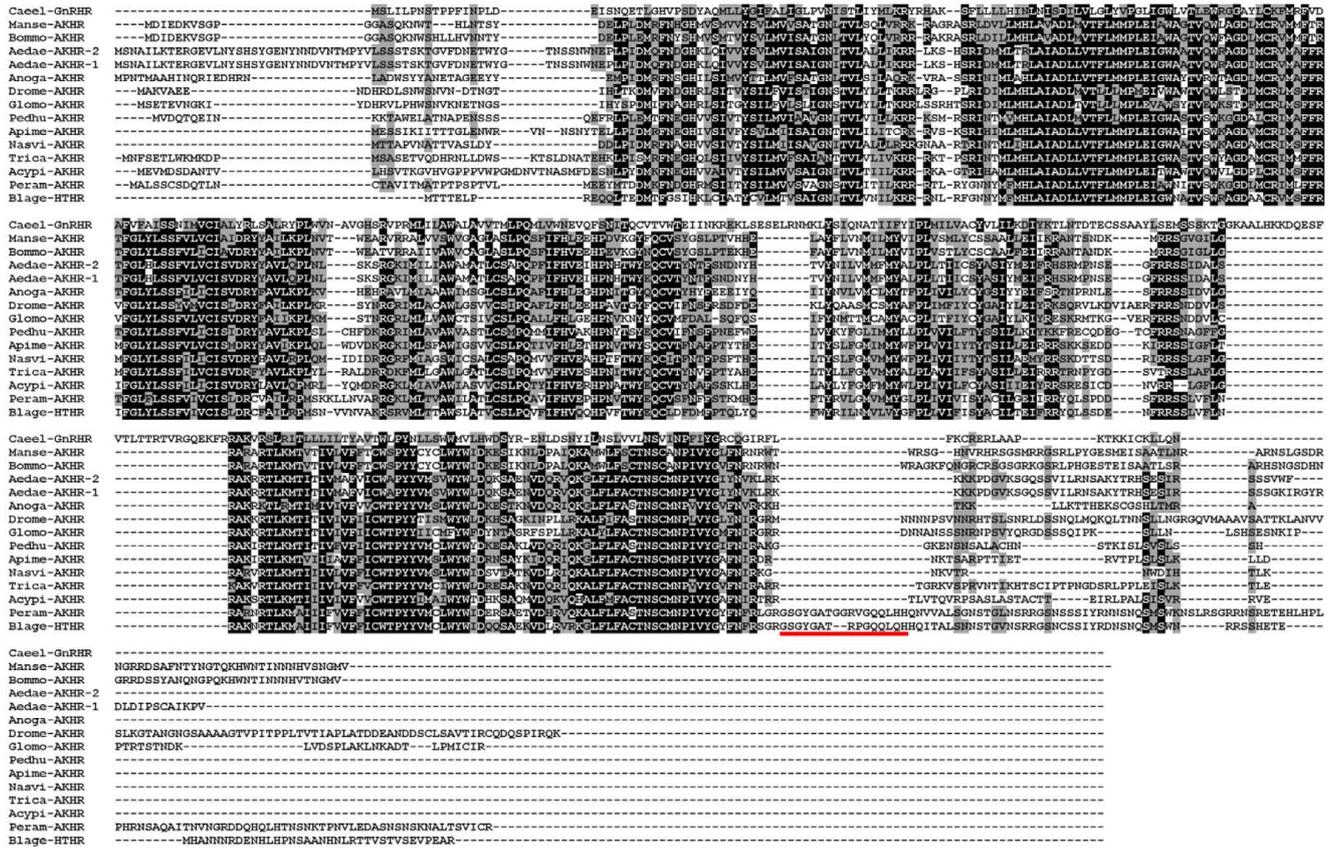
- Velki, M., Kodrik, D., Vecera, J., Hackenberger, B. K., and Socha, R. (2011). Oxidative stress elicited by insecticides: a role for the adipokinetic hormone. *Gen. Comp. Endocrinol.* 172, 77–84.
- Wicher, D., Agricola, H. J., Sohler, S., Gundel, M., Heinemann, S. H., Wollweber, L., Stengl, M., and Derst, C. (2006). Differential receptor activation by cockroach adipokinetic hormones produces differential effects on ion currents, neuronal activity, and locomotion. *J. Neurophysiol.* 95, 2314–2325.
- Yang, Y. Y., Wen, C.-J., Mishra, A., Tsai, C.-W., and Lee, H.-J. (2009). Development of the circadian clock in the German cockroach, *Blattella germanica*. *J. Insect Physiol.* 55, 469–478.
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## APPENDIX

1 5' -AATATGAATCTAACAAGCTATCATACTTCT  
 31 CTTATGGCTGGAGGAGCGGAAAAATCTGCAGTGTAGAAAAAGCGATAACATTCTAAGAACAACTGAATGATTAGACAAATCAAATGT  
 121 TCGAGAATTTATTCAACGAAGATTGAAGTGATGAACGAATTAGTTCTAACAGTGAATGGAAGAGACGAAGTGATGATTCCGCTTAGTGAA  
 211 ACAGTGGCATGGAAGAATTTGGGTGTTGGTAATCTTGATTTCATCTTTCAACTGTCAACATTTTGACATGAGCATTGACAGGAACCCAAG  
 301 **ATG**ACGACTACAGAGCTGCCCCGTGAGCAGCAGCTGACTGAGGACATGACTTTTGGCTCCATACATAAGTTATGTATCGCCACGTATTGC  
 M T T T E L P R E Q Q L T E D M T F G S I H K L C I A T Y C  
 391 GTTCTCATGACTGTATCTGCAATTGGCAACATCACGGTCTGGTCAACATACTCAAGAGACGCCGCAACCTTCGTTTTGGGAACAACTAC  
 V L M T V S A I G N I T V L V N I L K R R R N L R F G N N Y  
 481 ATGTTTCATGCATCTTGCATAGCAGACCTATTGGTGACTTTTCTCATGATGCCGCTAGAAATCGGCTGGAATGCAACAGTGTCTTGGAGA  
 M F M H L A I A D L L V T F L M M P L E I G W N A T V S W R  
 571 GCCGGTGACGCTGCTTGCAGAGTGATGCGTTCTTCAGGATATTCGGCCTCTATCTGTCTAGCTTCGTAATTGTTGTGCATCAGTCTGGAC  
 A G D A A C R V M S F F R I F G L Y L S S F V I V C I S L D  
 661 CGCTGTTTCGCCATTCTAAGGCCCATGTCGAACGTCGTCACAGTCGCCAAACGCAGCAGAGTAATGCTGACCACTGCCTGGTCATTGGCC  
 R C F A I L R P M S N V V N V A K R S R V M L T T A W S L A  
 751 ACTGTTTGTAGTTTACCTCAGGTGTTTCATCTTCCACGTACAACAGCATCCCGTTTTTCACATGGTATGAGCAGTGTGGACTTCGACATG  
 T V C S L P Q V F I F H V Q Q H P V F T W Y E Q C L D F D M  
 841 TTTCCGACTCAGCTGTACCAGTCTGGTACAGGATATTAATAATGGTCTAGTGTACGGTTTCCCACTCTCGTCATTTTCATCTCATA  
 F P T Q L Y Q F W Y R I L N M V L V Y G F P L L V I F I S Y  
 931 GCCTGTATCTCACAGAGATTTTTCGAGATATCAGCTCAGTTCAGACGAAAACCTCCGGAGATCGAGCCTTGTGTTCTGAACAGAGCC  
 A C I L T E I F R R Y Q L S S D E N F R R S S L V F L N R A  
 1021 AAAAATAGGACGCTCAAAAATGGCCATCATAATATTCGTAGTGTCTTCATCTGCTGGACTCCCTACTATGTGATGTGTCTCTGGTACTGG  
 K N R T L K M A I I I F V V F F I C W T P Y Y V M C L W Y W  
 1111 **ATAGACCAGCAGTCTGCTGAAAAGGTTGACTTGCGCGTGAGAAAAGGCTGTTCTGTTGCTTGTACCAATTCCGTGTATGAACCCGATT**  
 I D Q Q S A E K V D L R V R K G L F L F A C T N S C M N P I  
 1201 **GTGTACGGTACTTCAACTTCCGTTTCAGGACGGGGAAGTGGTTATGGTGCAACAAGACCAGGGCAGCAGTTACAGCATCATCAAATAACT**  
 V Y G Y F N F R S G R G S G Y G A T R P G Q Q L Q H H Q I T  
 1291 **GCAATTGAGCAATAACTCAACGGGAGTTAACAGCCGAAGGGGAAGCAACTGCAGCAGCATCTATCGGGACAACAGCAACCAGAGCATGTCC**  
 A L S N N S T G V N S R R G S N C S S I Y R D N S N Q S M S  
 1381 **TGGAATCGCCAAGCAGCCATGAAACAGAAAATGCACGCCAATAACAATCGAGACGAAAATCACTTACACCCCAACTCAGCTGCGAACCAC**  
 W N R R S S H E T E M H A N N N R D E N H L H P N S A A N H  
 1471 **AATTTGCGAACTACAGTATCTACTGTGAGTGAAGTTCCTGAAGCAAGATGAGGTTTACTCCTTGTGCTTCGAAATTATACTATCAGTGTAT**  
 N L R T T V S T V S E V P E A R \*  
 1561 **AAACATTGCCAATGTAATTAACATTAATGTTCTAGTCTAAGACTTGACATGACCTGGTTCACAAGACATTTCCC**AAGAGAGTGAACAAA  
 1651 ACAATTGAATTGGTATAAACATGTGTTAACTAATGTACAATACACTCACTCAACAGTTTTACACAAAAATAAATTTATTATTTTTTG  
 1741 CCACTAAAAAAAAAAAAAAAAAAAAAAAAA - 3'

**FIGURE A1 | Nucleotide sequence of cDNA encoding the open-reading frame for *B. germanica* HTHR with flanking 5'- and 3'-UTR sequences (GenBank accession number GU591493). Start and stop codon are indicated by yellow background. The primer pair for**

qRT-PCR is shown in underlined bold type. The primer pair for semiquantitative PCR is boxed. The fragment encompassed by the dsHTHR used in the RNAi experiments is shown in green background.



**FIGURE A2 |** Alignment of the AKH receptor protein sequences from insects and gonadotropin-releasing hormone receptor of *Caenorhabditis elegans* used in the phylogenetic analysis. The alignment was carried out using ClustalW program. A black background

indicates the amino acids are identical among species, whereas a gray background shows similarity among residues. The red underline indicates the insertion present in both cockroach species. The abbreviated names of sequences used are provided in **Figure 1B**.