



# Mechanisms of Progranulin Action and Regulation in Genitourinary Cancers

Ryuta Tanimoto<sup>1</sup>, Kuojung G. Lu<sup>1</sup>, Shi-Qiong Xu<sup>1</sup>, Simone Buraschi<sup>2</sup>, Antonino Belfiore<sup>3</sup>, Renato V. Iozzo<sup>2</sup> and Andrea Morrione<sup>1\*</sup>

<sup>1</sup> Biology of Prostate Cancer Program, Department of Urology, Kimmel Cancer Center, Thomas Jefferson University, Philadelphia, PA, USA, <sup>2</sup> Cancer Cell Biology and Signaling Program, Department of Pathology, Anatomy and Cell Biology, Kimmel Cancer Center, Thomas Jefferson University, Philadelphia, PA, USA, <sup>3</sup> Department of Health Sciences, Endocrinology, University Magna Graecia of Catanzaro, Catanzaro, Italy

The growth factor progranulin has emerged in recent years as a critical regulator of transformation in several cancer models, including breast cancer, glioblastomas, leukemias, and hepatocellular carcinomas. Several laboratories, including ours, have also demonstrated an important role of progranulin in several genitourinary cancers, including ovarian, endometrial, cervical, prostate, and bladder tumors, where progranulin acts as an autocrine growth factor thereby modulating motility and invasion of transformed cells. In this review, we will focus on the mechanisms of action and regulation of progranulin signaling in genitourinary cancers with a special emphasis on prostate and bladder tumors.

Keywords: progranulin, bladder and prostate cancer, motility, anchorage-independent growth, tumor formation in vivo

#### **OPEN ACCESS**

### Edited by:

Gabriella Castoria, Seconda Università degli Studi di Napoli, Italy

### Reviewed by:

Barbara Belletti, Centro di Riferimento Oncologico di Aviano (IRCCS), Italy Marzia Di Donato, Seconda Università degli Studi di Napoli, Italy

### \*Correspondence:

Andrea Morrione andrea.morrione@jefferson.edu

### Specialty section:

This article was submitted to Cancer Endocrinology, a section of the journal Frontiers in Endocrinology

> Received: 12 June 2016 Accepted: 08 July 2016 Published: 27 July 2016

### Citation:

Tanimoto R, Lu KG, Xu S-Q, Buraschi S, Belfiore A, lozzo RV and Morrione A (2016) Mechanisms of Progranulin Action and Regulation in Genitourinary Cancers. Front. Endocrinol. 7:100. doi: 10.3389/fendo.2016.00100

# INTRODUCTION

Progranulin, also known as proepithelin, granulin–epithelin precursor, acrogranin, and PC cell-derived growth factor, is a secreted glycoprotein with dual cellular roles, both as a growth factor regulating cell proliferation and wound repair and as a component of the transforming machinery in several cancer systems (1–3). Progranulin regulates inflammation and neurodegeneration (4) and has been linked to the development of frontotemporal dementia (FTD) (4).

Progranulin contains seven and a half granulin-like domains, which are  $\sim$ 6 kDa peptides called granulins (granulin A to G and paragranulin). These domains contain highly conserved tandem repeats of 12-cysteine residues, with the exception of granulin G, which has only 10 cysteine residues. The progranulin precursor is a 68.5-kDa protein of 593 amino acids, which migrates at  $\sim$ 88 kDa on Western immunoblot due to glycosylation (5).

Historically, granulins were first discovered as a novel class of leukocyte peptides with cytokine-like activities (6, 7). The progranulin precursor protein was later purified from PC cell-derived conditioned medium by Zhou et al. (8), who demonstrated the function of progranulin as mitogen and an autocrine growth factor for cancer cell lines. This precursor protein was identical to a growth-promoting factor later purified by Dr. Baserga's group from the conditioned media of BRL-3A rat liver cells (8), which was able to induce cell proliferation of R<sup>-</sup> cells, derived from mice with a targeted deletion of the insulin-like growth factor receptor (*IGF-IR*) gene (9). Significantly, progranulin was the only known growth factor able to bypass the requirement for the IGF-IR, thus

1

promoting growth of  $R^-$  cells (10). As demonstrated by He et al., progranulin overexpression in adrenal carcinoma and renal epithelial cells resulted in enhanced progranulin secretion, increased mitogenesis, and tumorigenesis (11). Progranulin has been shown to enhance proliferation and promote tumor growth in several cancer cell lines, such as breast, gastrointestinal, hepatic, lung, and genitourinary cancers. In this manuscript, we review progranulin's dual roles as a novel transforming growth factor and a clinical diagnostic and prognostic biomarker in genitourinary cancers.

# PROGRANULIN IN OVARIAN CANCER

Ovarian cancer (OC) is the deadliest gynecological cancer and is the fourth leading cause of cancer-related deaths in women (12). While current treatment with surgery and chemotherapy can manage early stage disease, patients with late stage disease often recur and develop distant metastases. From the initial diagnosis, the median 10-year survival is only about 40% (13). Thus, it is very important to identify novel predictors of ovarian tumor progression.

Progranulin action in OC has been well characterized. Jones et al. (14) compared progranulin expression between low malignant potential and invasive ovarian tumors and demonstrated that progranulin expression was upregulated in invasive OC compared to low-grade tumors (15), suggesting that progranulin expression may be predictive of tumor progression. Progranulin mRNA and protein expression levels were significantly higher in tissue samples obtained from OC when compared to benign and normal ovarian tissues (15–17). Furthermore, progranulin expression level was correlated with higher stage and worse prognosis in OC patients (16–20).

Furthermore, numerous *in vitro* and *in vivo* studies have demonstrated the role of progranulin in promoting proliferation, resistance to apoptosis, and motility of OC cells, where progranulin activates mitogen-activated protein kinase (MAPK) and MMP-2 signaling (14, 17, 20–22).

Another potential serum marker for OC is serum leukocyte protease inhibitor (SLPI), a serine protease inhibitor protein, whose levels are elevated in serum derived from patients with OC (23). Significantly, SLPI protects progranulin from serine proteases and cleavage into mature granulins (24). Devoogdt et al. demonstrated that SLPI overexpression protected progranulin from elastase-mediated degradation and increased OC growth and survival, both *in vitro* and *in vivo* (25, 26). Han et al. reported that among progranulin, SLPI, and the FDA approved serum biomarkers HE4 and CA125, progranulin was the only biomarker independently associated with poor prognosis in OC patients in remission after initial surgical cytoreduction and chemotherapy (18).

# PROGRANULIN IN CERVICAL AND ENDOMETRIAL CANCER

Cervical cancer is the third most common cancer among women worldwide and the third most common genitourinary cancer in

United States (12). Human papillomavirus (HPV) infection is the prevalent cause of cervical cancer as in fact over 99% of cervical cancers are positive for HPV (27). However, only a small fraction of women infected by HPV develops cervical cancer suggesting that other factors contribute to its progression.

Progranulin overexpression or recombinant progranulin protein stimulated transformation of HPV16-immortalized cervical mucosa epithelial H8 cells *in vitro* and *in vivo* (28, 29). On the contrary, inhibition of progranulin expression by siRNA approaches inhibited growth (28, 30).

Endometrial cancer is the most common genital cancer among women in United States, with an estimated 60,050 new cases in 2016 (12). The risk factors are related to an increase in circulating estrogens secondary to obesity, chronic anovulation and nulliparity estrogen replacement therapy, and tamoxifen use. Endometrial cancer tissues showed upregulated progranulin expression in association with estrogen receptor (31), and progranulin expression level was increased after estradiol and/or tamoxifen treatment *in vitro* (31). Similar to cervical cancer, inhibition of progranulin with shRNAs resulted in decreased endometrial cancer cell proliferation (31).

These results suggest that in endometrial cancer progranulin may work in estradiol-dependent fashion and mediate, at least in part, estrogen receptor-dependent biological responses. This is reminiscent of progranulin action in breast cancer, where progranulin (PCDGF/granulin precursor) expression is stimulated by estradiol (32) and progranulin regulates estrogen-dependent mitogenesis (33).

# PROGRANULIN ACTION IN BLADDER CANCER

Bladder cancer is the fourth most common cancer in men in developed countries, with high recurrence rates of around 60% after treatment (12, 34, 35). The vast majority of early stages bladder tumors present as low-grade non-invasive papillary tumors (Ta stage). The remaining comprises tumors that have penetrated the basement membrane but not invaded the muscle layer of the bladder wall (T1 stage) and muscle-invasive tumors (T2–T4 stages) (36). The prognosis for low-grade tumors is generally good but about 15% of low-grade tumors progress to invasive disease. For invasive tumors, patient survival is ~50% at 5 years (37). Invasive tumors are frequently associated with metastases, which have a 5-year survival rate of 6% (37). However, despite the significant clinical impact of bladder cancer, there is still an urgent need to discover drivers of disease progression and novel therapeutic targets.

In recent years, our laboratories have established that progranulin plays a critical role in bladder cancer by regulating bladder cancer cell motility, invasion, anchorage-independent growth, and tumor formation *in vivo* (38–41).

We initially demonstrated that recombinant human progranulin did promote migration, wound closure, and invasion of urothelial cancer cells (42). Progranulin-mediated biological responses required the activation of the MAPK pathway and paxillin, which upon progranulin stimulation formed a complex

with focal adhesion kinase (FAK) and active extracellular signal-regulated kinase (ERK) at the leading edge of migrating urothelial cancer cells (42). These results suggested that progranulin-induced cell motility might be regulated by the ability of progranulin to modulate focal adhesion dynamics of motile cancer cells.

We later discovered that urothelial cancer cells abundantly secrete progranulin (39). Significantly, stable depletion by shRNA approaches of endogenous progranulin expressed in T24 urothelial carcinoma-derived cell inhibited both the Akt and MAPK pathways, reduced the ability of T24 cells to proliferate in the absence of serum and inhibited motility, wound healing and invasion *in vitro* (39). Collectively, these results demonstrated that progranulin acts as a tumorigenic autocrine growth factor for bladder cancer cells and may work as a biomarker for bladder neoplasms.

In order to gain further insights into the molecular mechanisms of progranulin action in bladder cancer, we searched for novel progranulin-interacting proteins using pull-down assays with recombinant progranulin and proteomic analysis. We demonstrated that drebrin (developmentally regulated brain protein), an F-actin binding protein (43, 44), interacted with progranulin in urothelial cancer cells (40), modulated progranulin-induced MAPK and Akt signaling, and promoted progranulin-dependent motility and invasion by regulating F-actin remodeling (40). In addition, drebrin was essential for anchorage-independent growth in vitro and tumor formation in vivo, and its expression was upregulated in bladder cancer tissues compared to normal tissue controls (40). Our data demonstrated an essential functional

role for drebrin in the regulation of progranulin actions suggesting that drebrin may constitute a novel target for therapeutic intervention in bladder tumors. In addition, drebrin may work as novel biomarker for bladder cancer progression (**Figure 1**).

Given the critical role of progranulin in regulating motility and invasion of urothelial cancer cells (39, 40, 42), we then determined whether targeting progranulin could have therapeutic efficacy in bladder cancer. Thus, we very recently generated progranulin-depleted tumorigenic urothelial cancer cells and demonstrated that progranulin targeting markedly reduced in vivo tumor growth of UMUC-3 cells in both orthotopic and subcutaneous xenograft tumor models (41). The immunostaining analysis of orthotopic tumors derived from progranulin-depleted UMUC-3 cells tumors had significantly lower expression level of Ki67, a cellular marker of proliferation, compared to tumor tissues derived from control-transfected cells (41). Moreover, the F-actin network of orthotopic tumors derived from progranulindepleted cells was intact and well assembled compared to controls (41), supporting our previous in vitro results (40) and the hypothesis that progranulin might regulate invasion and motility of urothelial cancer cells by modulating F-actin remodeling.

Significantly, tumor tissues derived from UMUC-3/Control orthotopic xenografts showed reduced expression of the epithelial marker E-cadherin and increased levels of the mesenchymal protein vimentin compared to progranulin-depleted orthotopic xenografts. These results would suggest that the role of progranulin in bladder cancer tumor formation might be associated with an epithelial-to-mesenchymal transition (EMT) (45), which is

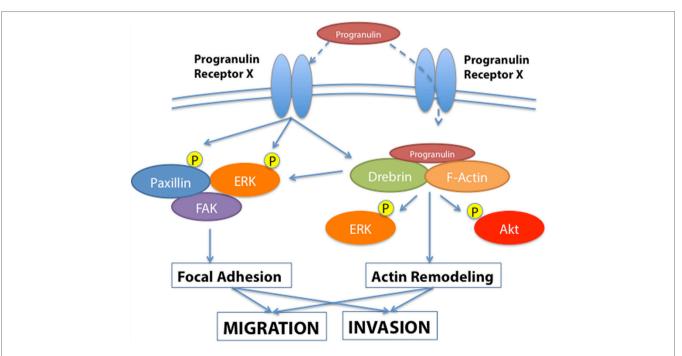


FIGURE 1 | Progranulin signaling in bladder cancer. The schematic draw summarizes the current knowledge of progranulin signaling pathways leading to motility and invasion of bladder cancer cells. Progranulin interaction with the unidentified progranulin membrane receptor mediates progranulin internalization and interaction with the F-actin protein drebrin, thus regulating MAPK and Akt pathways and cytoskeletal remodeling (40). Additionally, progranulin signaling activates paxillin, which may contribute with MAPK in regulating the dynamic of focal adhesions.

severely affected by progranulin ablation in urothelial cancer cells. These results are consistent with data showing that in OC progranulin induces motility and invasion through EMT and the activation of cancer-associated fibroblasts (20).

The standard of care for invasive and metastatic bladder cancer is based on platinum-containing anti-cancer drugs, such as cisplatin (46). Progranulin depletion significantly enhanced cisplatin-dependent cell death as compared to control bladder cancer cells (41), suggesting that progranulin targeting in combination with cisplatin could enhance the therapeutic efficacy of cisplatin-based therapy in invasive bladder tumors.

Because progranulin might be secreted in the urine either through active secretion or after cell death, initially we assessed progranulin levels in concentrated urines of healthy subjects and demonstrated that progranulin can be detected in urine using immunoblots (39). Furthermore, progranulin levels in urine were quantified by enzyme-linked immunosorbent assay (ELISA) (39). Significantly, Soukup et al. (47) tested by ELISA assay the urinary concentrations of 27 biomarkers, including progranulin, for the detection of primary and recurrent urothelial bladder cancers. Importantly, the urinary concentration of progranulin was significantly higher in patients with bladder cancer compared to healthy subjects (47). However, progranulin levels did not show significant difference between patients with cancer recurrence and patient without recurrence (47). Taking into account the sample size limitation of this study, additional analyses on larger cohorts are necessary to establish whether progranulin may work as a urine biomarker for bladder cancer.

We recently analyze by immunohistochemistry progranulin expression levels in a human bladder tumor microarray containing 69 validated cases, including various types and stages of bladder cancers. Progranulin was barely detectable in normal bladder. In contrast, all malignant bladder tumors exhibited a significantly increased progranulin expression with the only exception of mucinous carcinoma where progranulin levels were comparable to normal tissues (41). Progranulin upregulation was not statistically different between low-grade and high-grade invasive urothelial carcinoma tissues (41). Progranulin was also upregulated in metastatic bladder tissues suggesting that progranulin expression levels might be associated with bladder cancer metastases. We additionally tested progranulin expression levels in archived paraffin-embedded tissues derived from noninvasive carcinoma in situ (Tis), non-invasive low-grade papillary carcinomas (Ta), non-invasive high-grade papillary carcinomas (Ta) using as control normal, and invasive high-grade carcinoma. Progranulin expression levels were significantly upregulated in non-invasive low-grade papillary carcinomas (Ta) compared to benign tissues and enhanced in both non-invasive high-grade papillary carcinomas (Ta) and high-grade carcinoma tissues compared to benign tissues, while in non-invasive carcinoma in situ (Tis) progranulin expression was at levels similar to benign tissues (41). These results were somewhat surprising considering the importance of progranulin in regulating motility and invasion and suggest that progranulin expression is predictive of bladder tumor formation but do not discriminate between superficial and invasive urothelial carcinomas. On the contrary, drebrin expression correlated with bladder tumor progression (40) indicating

that expression levels of downstream progranulin effectors may be more predictive of bladder tumor progression than progranulin itself.

# PROGRANULIN ACTION IN PROSTATE CANCER

Prostate cancer is the most common cancer in men and is also the second leading cause of cancer-related deaths in men (12). In the developed world, most prostate cancers are diagnosed at an early stage where the cancer is confined to the prostate or local organs, with minimal risk of disease progression after treatment. However, a significant proportion of men with clinically localized prostate cancer treated with radical prostatectomy will develop biochemical recurrence, subsequent local recurrence, and distant metastases (48–50). Furthermore, the mechanisms promoting progression to the castration-resistant stage of prostate cancer (CRPC) are still very poorly characterized.

We previously established that progranulin is not expressed in androgen-responsive LNCaP prostate cancer cells but instead acts as an autocrine growth factor for DU145 cells and promotes castration-resistant prostate cancer cell motility, invasion, and anchorage-independent growth (38). This study supported the hypothesis that progranulin was not important for prostate cancer initiation but might play an important role in prostate cancer progression and CRPC.

Sortilin, a single-pass type I transmembrane protein of the Vps10 family, binds progranulin in neurons and mediates progranulin targeting for lysosomal degradation (51). However, the role of sortilin in cancer is still very poorly characterized. In addition, whether sortilin is expressed in prostate cancer cells and plays any role in regulating progranulin action in prostate cancer has not previously established. Androgen-responsive LNCaP cells express very low levels of endogenous progranulin, do not respond to exogenous recombinant progranulin and do not express sortilin (52), suggesting that the progranulin/sortilin functional interaction may have a prevalent role in castration-resistant prostate cancer. Accordingly, in castration-resistant DU145 and PC3 cells, we detected very low levels of sortilin, which was associated with high levels of progranulin production and enhanced motility and invasion (52). Restoring sortilin expression in PC3 and DU145 cells decreased progranulin levels and inhibited motility, invasion, and anchorage-independent growth (52). These results were recapitulated by depleting endogenous progranulin in PC3 and DU145 cells. On the contrary, targeting endogenous sortilin by shRNA approaches enhanced progranulin levels and sustained motility, invasion, and anchorage-independent growth (52).

Similar to OC, Zheng et al. (53) reported that SLPI is overexpressed in CRPC patients and supports CRPC cell growth and invasion by functionally interacting with progranulin (53). SLPI overexpression in prostate cancer cells provides a proliferative advantage after castration by its anti-protease activity toward elastase and protecting progranulin from elastase-dependent degradation (53).

These results suggest that the functional interaction between sortilin, SLPI, and progranulin levels may be critical for prostate

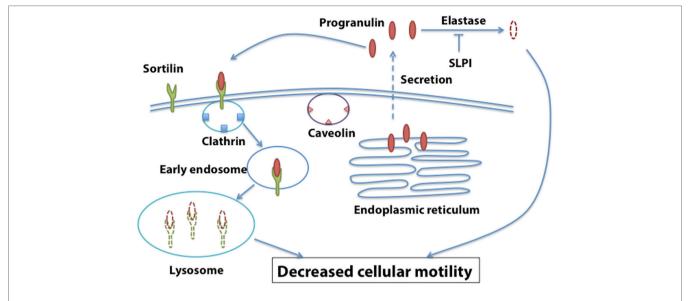


FIGURE 2 | Sortilin-dependent regulation of progranulin action in prostate cancer. Sortilin promotes progranulin uptake through clathrin-dependent endocytosis and targets progranulin for degradation in the lysosomes. Sortilin loss is associated with enhanced progranulin levels and action in castration-resistant prostate cancer cells.

cancer, and sortilin/SLPI loss may contribute to progranulindependent action in prostate cancer progression (Figure 2).

The role of progranulin as a prostate cancer biomarker has not been clearly defined, and the data are also partially discordant. We investigated progranulin expression levels in available prostate cancer microarray studies using the ONCOMINE database (42). We found that progranulin mRNA expression was significantly increased in six primary prostate cancer data sets compared to controls (54–59). Additionally, primary tumors that have been treated with hormone therapy showed decreased progranulin mRNA levels (57). However, three studies showed contradictory results demonstrating that metastatic cancers had decreased progranulin expression levels compared to non-metastatic prostate cancers (54, 55, 60).

We evaluated by immunohistochemical analysis progranulin expression levels in normal versus prostate cancer sections and demonstrated that progranulin was expressed at low levels, in normal prostate but was instead upregulated in tumor glands (42), confirming the data by Pan et al. (61), who assessed progranulin (GP88) in normal prostate, prostatic intraepithelial neoplasia, and prostatic adenocarcinoma tissues. However, due to the limited sample size of both studies, additional analyses are indeed required to clearly define progranulin as a biomarker for prostate cancer progression.

# CONCLUSION

A vast body of evidence has emerged in support of the critical role of progranulin in promoting transformation of several cancer models. However, the mechanisms of progranulin action are still poorly characterized. Experiments are currently under way to identify novel regulators of progranulin signaling, which may constitute novel targets for therapy in tumors where progranulin plays a role.

### **AUTHOR CONTRIBUTIONS**

RT, KL, AB, RVI, and AM wrote the paper and designed experiments. RT and KL performed experiments and discussed data. SB and S-QX performed experiments and discussed data.

### **ACKNOWLEDGMENTS**

We are sorry with all authors and work we may have missed in this manuscript. We are deeply grateful to all formers AM and RVI's laboratory members who contributed through the years to the progranulin projects. A sincere thank to Dr. Baserga who initially involved me (AM) in the progranulin project.

### **FUNDING**

This work was supported in part by the Benjamin Perkins Bladder Cancer Fund, The Schorsch Family Fund for Innovative Medical Research and Care (AM), and National Institutes of Health Grants RO1 CA164462 (AM and RVI), and grants from the Associazione Italiana per la Ricerca sul Cancro (AIRC) (IG 14066/2013 to AB).

## REFERENCES

- He Z, Ismail A, Kriazhev L, Sadvakassova G, Bateman A. Progranulin (PC-cell-derived growth factor/acrogranin) regulates invasion and cell survival. Cancer Res (2002) 62:5590-6.
- He Z, Bateman A. Progranulin (granulin-epithelin precursor, PC-cell-derived growth factor, acrogranin) mediates tissue repair and tumorigenesis. J Mol Med (2003) 81:600–12. doi:10.1007/s00109-003-0474-3
- 3. He Z, Ong CH, Halper J, Bateman A. Progranulin is a mediator of the wound response. *Nat Med* (2003) 9:225–9. doi:10.1038/nm816
- Cenik B, Sephton CF, Kutluk Cenik B, Herz J, Yu G. Progranulin: a proteolytically processed protein at the crossroads of inflammation and neurodegeneration. J Biol Chem (2012) 287:32298–306. doi:10.1074/jbc.R112.399170
- Bhandari V, Palfree RG, Bateman A. Isolation and sequence of the granulin precursor cDNA from human bone marrow reveals tandem cysteine-rich granulin domains. *Proc Natl Acad Sci U S A* (1992) 89:1715–9. doi:10.1073/ pnas.89.5.1715
- Bateman A, Belcourt D, Bennett H, Lazure C, Solomon S. Granulins, a novel class of peptide from leukocytes. *Biochem Biophys Res Commun* (1990) 173:1161–8. doi:10.1016/S0006-291X(05)80908-8
- Shoyab M, Mcdonald VL, Byles C, Todaro GJ, Plowman GD. Epithelins 1 and 2: isolation and characterization of two cysteine-rich growth-modulating proteins. Proc Natl Acad Sci U S A (1990) 87:7912–6. doi:10.1073/pnas.87.20.7912
- Zhou J, Gao G, Crabb JW, Serrero G. Purification of an autocrine growth factor homologous with mouse epithelin precursor from a highly tumorigenic cell line. J Biol Chem (1993) 268:10863–9.
- Sell C, Dumenil G, Deveaud C, Miura M, Coppola D, Deangelis T, et al. Effect of a null mutation of the insulin-like growth factor I receptor gene on growth and transformation of mouse embryo fibroblasts. *Mol Cell Biol* (1994) 14:3604–12. doi:10.1128/MCB.14.6.3604
- Xu SQ, Tang D, Chamberlain S, Pronk G, Masiarz FR, Kaur S, et al. The granulin/epithelin precursor abrogates the requirement for the insulin-like growth factor 1 receptor for growth in vitro. *J Biol Chem* (1998) 273:20078–83. doi:10.1074/jbc.273.32.20078
- 11. He Z, Bateman A. Progranulin gene expression regulates epithelial cell growth and promotes tumor growth in vivo. *Cancer Res* (1999) 59:3222–9.
- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. CA Cancer J Clin (2016) 66:7–30. doi:10.3322/caac.21332
- Jayson GC, Kohn EC, Kitchener HC, Ledermann JA. Ovarian cancer. Lancet (2014) 384:1376–88. doi:10.1016/S0140-6736(13)62146-7
- Jones MB, Michener CM, Blanchette JO, Kuznetsov VA, Raffeld M, Serrero G, et al. The granulin-epithelin precursor/PC-cell-derived growth factor is a growth factor for epithelial ovarian cancer. Clin Cancer Res (2003) 9:44–51.
- Jones MB, Spooner M, Kohn EC. The granulin-epithelin precursor: a putative new growth factor for ovarian cancer. Gynecol Oncol (2003) 88:S136–9. doi:10.1006/gyno.2002.6704
- Davidson B, Alejandro E, Florenes VA, Goderstad JM, Risberg B, Kristensen GB, et al. Granulin-epithelin precursor is a novel prognostic marker in epithelial ovarian carcinoma. Cancer (2004) 100:2139–47. doi:10.1002/cncr.20219
- Cuevas-Antonio R, Cancino C, Arechavaleta-Velasco F, Andrade A, Barron L, Estrada I, et al. Expression of progranulin (acrogranin/PCDGF/granulinepithelin precursor) in benign and malignant ovarian tumors and activation of MAPK signaling in ovarian cancer cell line. Cancer Invest (2010) 28:452–8. doi:10.3109/07357900903346455
- Han JJ, Yu M, Houston N, Steinberg SM, Kohn EC. Progranulin is a potential prognostic biomarker in advanced epithelial ovarian cancers. *Gynecol Oncol* (2011) 120:5–10. doi:10.1016/j.ygyno.2010.09.006
- Carlson AM, Maurer MJ, Goergen KM, Kalli KR, Erskine CL, Behrens MD, et al. Utility of progranulin and serum leukocyte protease inhibitor as diagnostic and prognostic biomarkers in ovarian cancer. *Cancer Epidemiol Biomarkers Prev* (2013) 22:1730–5. doi:10.1158/1055-9965.EPI-12-1368
- Dong T, Yang D, Li R, Zhang L, Zhao H, Shen Y, et al. PGRN promotes migration and invasion of epithelial ovarian cancer cells through an epithelial mesenchymal transition program and the activation of cancer associated fibroblasts. Exp Mol Pathol (2016) 100:17–25. doi:10.1016/j.yexmp.2015.11.021
- Miyanishi M, Mandai M, Matsumura N, Yamaguchi K, Hamanishi J, Higuchi T, et al. Immortalized ovarian surface epithelial cells acquire tumorigenicity by acrogranin gene overexpression. Oncol Rep (2007) 17:329–33.

- Pizarro GO, Zhou XC, Koch A, Gharib M, Raval S, Bible K, et al. Prosurvival function of the granulin-epithelin precursor is important in tumor progression and chemoresponse. *Int J Cancer* (2007) 120:2339–43. doi:10.1002/ijc.22559
- Thompson RC, Ohlsson K. Isolation, properties, and complete amino acid sequence of human secretory leukocyte protease inhibitor, a potent inhibitor of leukocyte elastase. *Proc Natl Acad Sci U S A* (1986) 83:6692–6. doi:10.1073/ pnas.83.18.6692
- Zhu J, Nathan C, Jin W, Sim D, Ashcroft GS, Wahl SM, et al. Conversion of proepithelin to epithelins: roles of SLPI and elastase in host defense and wound repair. Cell (2002) 111:867–78. doi:10.1016/S0092-8674(02)01141-8
- Simpkins FA, Devoogdt NM, Rasool N, Tchabo NE, Alejandro EU, Kamrava MM, et al. The alarm anti-protease, secretory leukocyte protease inhibitor, is a proliferation and survival factor for ovarian cancer cells. Carcinogenesis (2008) 29:466–72. doi:10.1093/carcin/bgm212
- Devoogdt N, Rasool N, Hoskins E, Simpkins F, Tchabo N, Kohn EC. Overexpression of protease inhibitor-dead secretory leukocyte protease inhibitor causes more aggressive ovarian cancer in vitro and in vivo. Cancer Sci (2009) 100:434–40. doi:10.1111/j.1349-7006.2009.01076.x
- 27. Wright JD, Herzog TJ. Human papillomavirus: emerging trends in detection and management. *Curr Womens Health Rep* (2002) 2:259–65.
- Lu Y, Zheng L, Zhang W, Feng T, Liu J, Wang X, et al. Growth factor progranulin contributes to cervical cancer cell proliferation and transformation in vivo and in vitro. *Gynecol Oncol* (2014) 134:364–71. doi:10.1016/j.ygyno.2014.05.025
- Wei Z, Huang Y, Xie N, Ma Q. Elevated expression of secreted autocrine growth factor progranulin increases cervical cancer growth. *Cell Biochem Biophys* (2015) 71:189–93. doi:10.1007/s12013-014-0183-2
- Zhang XY, Pan ZX, Liu H, Yu JL, Li GX, Wang HY, et al. Effect of progranulin (PGRN) on the proliferation and senescence of cervical cancer cells. *Genet Mol Res* (2015) 14:14331–8. doi:10.4238/2015.November.13.18
- Jones MB, Houwink AP, Freeman BK, Greenwood TM, Lafky JM, Lingle WL, et al. The granulin-epithelin precursor is a steroid-regulated growth factor in endometrial cancer. J Soc Gynecol Investig (2006) 13:304–11. doi:10.1016/j. jsgi.2006.03.003
- Lu R, Serrero G. Stimulation of PC cell-derived growth factor (epithelin/ granulin precursor) expression by estradiol in human breast cancer cells. Biochem Biophys Res Commun (1999) 256:204–7. doi:10.1006/bbrc.1999.0253
- Lu R, Serrero G. Mediation of estrogen mitogenic effect in human breast cancer MCF-7 cells by PC-cell-derived growth factor (PCDGF/granulin precursor). Proc Natl Acad Sci U S A (2001) 98:142-7. doi:10.1073/pnas. 98.1.142
- Burger M, Catto JW, Dalbagni G, Grossman HB, Herr H, Karakiewicz P, et al. Epidemiology and risk factors of urothelial bladder cancer. *Eur Urol* (2013) 63:234–41. doi:10.1016/j.eururo.2012.07.033
- Kluth LA, Black PC, Bochner BH, Catto J, Lerner SP, Stenzl A, et al. Prognostic and prediction tools in bladder cancer: a comprehensive review of the literature. Eur Urol (2015) 68:238–53. doi:10.1016/j.eururo.2015.01.032
- Mitra AP, Cote RJ. Molecular pathogenesis and diagnostics of bladder cancer. Annu Rev Pathol (2009) 4:251–85. doi:10.1146/annurev. pathol.4.110807.092230
- Knowles MA. Molecular pathogenesis of bladder cancer. Int J Clin Oncol (2008) 13:287–97. doi:10.1007/s10147-008-0812-0
- Monami G, Gonzalez EM, Hellman M, Gomella LG, Baffa R, Iozzo RV, et al. Proepithelin promotes migration and invasion of 5637 bladder cancer cells through the activation of ERK1/2 and the formation of a paxillin/ FAK/ERK complex. Cancer Res (2006) 66:7103–10. doi:10.1158/0008-5472. CAN-06-0633
- Lovat F, Bitto A, Xu SQ, Fassan M, Goldoni S, Metalli D, et al. Proepithelin is an autocrine growth factor for bladder cancer. *Carcinogenesis* (2009) 30:861–8. doi:10.1093/carcin/bgp050
- Xu SQ, Buraschi S, Morcavallo A, Genua M, Shirao T, Peiper SC, et al. A novel role for drebrin in regulating progranulin bioactivity in bladder cancer. Oncotarget (2015) 6:10825–39. doi:10.18632/oncotarget.3424
- Buraschi S, Xu SQ, Stefanello M, Moskalev I, Morcavallo A, Genua M, et al. Suppression of progranulin expression inhibits bladder cancer growth and sensitizes cancer cells to cisplatin. *Oncotarget* (2016). doi:10.18632/ oncotarget.9556
- 42. Monami G, Emiliozzi V, Bitto A, Lovat F, Xu SQ, Goldoni S, et al. Proepithelin regulates prostate cancer cell biology by promoting cell growth, migration,

- and anchorage-independent growth. Am J Pathol (2009) 174:1037–47. doi:10.2353/ajpath.2009.080735
- Shirao T, Inoue HK, Kano Y, Obata K. Localization of a developmentally regulated neuron-specific protein S54 in dendrites as revealed by immunoelectron microscopy. *Brain Res* (1987) 413:374–8. doi:10.1016/0006-8993(87) 91032-8
- Shirao T, Kojima N, Kato Y, Obata K. Molecular cloning of a cDNA for the developmentally regulated brain protein, drebrin. *Brain Res* (1988) 464:71–4. doi:10.1016/0169-328X(88)90020-4
- 45. LamouilleS,XuJ,DerynckR.Molecularmechanismsofepithelial-mesenchymal transition. *Nat Rev Mol Cell Biol* (2014) 15:178–96. doi:10.1038/nrm3758
- Grossman HB, Natale RB, Tangen CM, Speights VO, Vogelzang NJ, Trump DL, et al. Neoadjuvant chemotherapy plus cystectomy compared with cystectomy alone for locally advanced bladder cancer. N Engl J Med (2003) 349:859–66. doi:10.1056/NEJMoa022148
- Soukup V, Kalousova M, Capoun O, Sobotka R, Breyl Z, Pesl M, et al. Panel of urinary diagnostic markers for non-invasive detection of primary and recurrent urothelial urinary bladder carcinoma. *Urol Int* (2015) 95:56–64. doi:10.1159/000368166
- Baillargeon J, Platz EA, Rose DP, Pollock BH, Ankerst DP, Haffner S, et al. Obesity, adipokines, and prostate cancer in a prospective population-based study. Cancer Epidemiol Biomarkers Prev (2006) 15:1331–5. doi:10.1158/1055-9965.EPI-06-0082
- Thompson IM, Tangen CM, Paradelo J, Lucia MS, Miller G, Troyer D, et al. Adjuvant radiotherapy for pathological T3N0M0 prostate cancer significantly reduces risk of metastases and improves survival: long-term followup of a randomized clinical trial. *J Urol* (2009) 181:956–62. doi:10.1016/j. juro.2008.11.032
- Bolla M, Van Poppel H, Tombal B, Vekemans K, Da Pozzo L, De Reijke TM, et al. Postoperative radiotherapy after radical prostatectomy for high-risk prostate cancer: long-term results of a randomised controlled trial (EORTC trial 22911). *Lancet* (2012) 380:2018–27. doi:10.1016/S0140-6736(12) 61253-7
- Hu F, Padukkavidana T, Vaegter CB, Brady OA, Zheng Y, Mackenzie IR, et al. Sortilin-mediated endocytosis determines levels of the frontotemporal dementia protein, progranulin. *Neuron* (2010) 68:654–67. doi:10.1016/j. neuron.2010.09.034
- Tanimoto R, Morcavallo A, Terracciano M, Xu SQ, Stefanello M, Buraschi S, et al. Sortilin regulates progranulin action in castration-resistant prostate cancer cells. *Endocrinology* (2015) 156:58–70. doi:10.1210/en.2014-1590
- Zheng D, Gui B, Gray KP, Tinay I, Rafiei S, Huang Q, et al. Secretory leukocyte protease inhibitor is a survival and proliferation factor for castration-resistant prostate cancer. *Oncogene* (2016). doi:10.1038/onc.2016.13

- Dhanasekaran SM, Barrette TR, Ghosh D, Shah R, Varambally S, Kurachi K, et al. Delineation of prognostic biomarkers in prostate cancer. *Nature* (2001) 412:822–6. doi:10.1038/35090585
- Magee JA, Araki T, Patil S, Ehrig T, True L, Humphrey PA, et al. Expression profiling reveals hepsin overexpression in prostate cancer. *Cancer Res* (2001) 61:5692–6.
- Welsh JB, Sapinoso LM, Su AI, Kern SG, Wang-Rodriguez J, Moskaluk CA, et al. Analysis of gene expression identifies candidate markers and pharmacological targets in prostate cancer. *Cancer Res* (2001) 61:5974–8.
- Holzbeierlein J, Lal P, Latulippe E, Smith A, Satagopan J, Zhang L, et al. Gene expression analysis of human prostate carcinoma during hormonal therapy identifies androgen-responsive genes and mechanisms of therapy resistance. *Am J Pathol* (2004) 164:217–27. doi:10.1016/S0002-9440(10)63112-4
- Rhodes DR, Yu J, Shanker K, Deshpande N, Varambally R, Ghosh D, et al. ONCOMINE: a cancer microarray database and integrated data-mining platform. Neoplasia (2004) 6:1–6. doi:10.1016/S1476-5586(04)80047-2
- Dhanasekaran SM, Dash A, Yu J, Maine IP, Laxman B, Tomlins SA, et al. Molecular profiling of human prostate tissues: insights into gene expression patterns of prostate development during puberty. FASEB J (2005) 19:243–5.
- Yu YP, Paranjpe S, Nelson J, Finkelstein S, Ren B, Kokkinakis D, et al. High throughput screening of methylation status of genes in prostate cancer using an oligonucleotide methylation array. *Carcinogenesis* (2005) 26:471–9. doi:10.1093/carcin/bgh310
- Pan CX, Kinch MS, Kiener PA, Langermann S, Serrero G, Sun L, et al. PC cell-derived growth factor expression in prostatic intraepithelial neoplasia and prostatic adenocarcinoma. Clin Cancer Res (2004) 10:1333–7. doi:10.1158/1078-0432.CCR-1123-03

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer MD and handling editor declared their shared affiliation, and the handling editor states that the process nevertheless met the standards of a fair and objective review.

Copyright © 2016 Tanimoto, Lu, Xu, Buraschi, Belfiore, Iozzo and Morrione. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.