



Maternal Diabetes and Fetal Programming Toward Neurological Diseases: Beyond Neural Tube Defects

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The purpose of this review was to search for experimental or clinical evidence on the effect of hyperglycemia in fetal programming to neurological diseases, excluding evident neural tube defects. The lack of timely diagnosis and the inadequate control of diabetes during pregnancy have been related with postnatal obesity, low intellectual and verbal coefficients, language and motor deficits, attention deficit with hyperactivity, problems in psychosocial development, and an increased predisposition to autism and schizophrenia. It has been proposed that several childhood or adulthood diseases have their origin during fetal development through a phenomenon called fetal programming. However, not all the relationships between the outcomes mentioned above and diabetes during gestation are clear, well-studied, or have been related to fetal programming. To understand this relationship, it is imperative to understand how developmental processes take place in health, in order to understand how the functional cytoarchitecture of the central nervous system takes place; to identify changes prompted by hyperglycemia, and to correlate them with the above postnatal impaired functions. Although changes in the establishment of patterns during central nervous system fetal development are related to a wide variety of neurological pathologies, the mechanism by which several maternal conditions promote fetal alterations that contribute to impaired neural development with postnatal consequences are not clear. Animal models have been extremely useful in studying the effect of maternal pathologies on embryo and fetal development, since obtaining central nervous system tissue in humans with normal appearance during fetal development is an important limitation. This review explores the state of the art on this topic, to help establish the way forward in the study of fetal programming under hyperglycemia and its impact on neurological and psychiatric disorders.

Keywords: diabetes, pregnancy, fetal programming, neurological disorders, psychiatric disease

INTRODUCTION

Diabetes is a group of metabolic diseases characterized by deficient insulin secretion and/or action which leads to hyperglycemia, and, in turn, to abnormal metabolism of carbohydrates, fats, and proteins in insulin target tissues (1). Depending on pathogenesis, diabetes can be classified as type 1, type 2, gestational diabetes mellitus (GDM), or other types of diabetes. Worldwide, \sim 5–10%

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Márquez-Valadez B, Valle-Bautista R, García-López G, Díaz NF and Molina-Hernández A (2018) Maternal Diabetes and Fetal Programming Toward Neurological Diseases: Beyond Neural Tube Defects. Front. Endocrinol. 9:664. doi: 10.3389/fendo.2018.00664 of diabetic patients have type 1 diabetes, in which complete insulin deficiency stems from β -cell autoimmune destruction as a result of genetic susceptibility or viral antigens. Type 2 affects 90–95% of diabetic patients and is characterized by either insulin resistance or defective secretion. Other, less common, types of diabetes are associated with monogenetic defects in β -cell function, pancreatic injuries, and drug abuse, among other causes (1).

GDM is commonly diagnosed in the second or third trimester of pregnancy, and its prevalence varies between 1 and 17% depending on the studied population and the diagnostic test used (1, 2). Due to the ongoing epidemic of obesity and diabetes in women of childbearing age, the American Diabetes Association (ADA) has established that women with GDM risk factors must be tested for diabetes during the second trimester of pregnancy, using standard diagnostic criteria. Furthermore, that diabetes in women in the first trimester should be classified as type 2 diabetes (1). However, pregnant women with no risk factors can develop diabetes as early as the first trimester (3–5), making it evident that earlier testing is required in some cases.

As a result of GDM, women and their offspring confront a series of problems including fetal death, spontaneous abortion, congenital malformations, fetal-placental abnormalities, and altered fetal programming (6–8). Fetal programming is defined as the development of pathologies during childhood and adulthood that originate during fetal development (9). In this sense, maternal diabetes has been linked to offspring that develop obesity, diabetes, neurodegenerative and psychiatric diseases, as well as low intellectual and verbal coefficients, language and motor impairments, attention deficit with hyperactivity disorder, and problems in psychosocial development (10–15). However, many of these relationships remain unclear, have not been well-studied and have not been related to fetal programming (**Figure 1**).

During this review, we realized that existing information regarding the mechanisms of action between hyperglycemia during fetal development and the outcomes is limited. Most reports are focused on *in utero* hyperglycemia and neural tube defects (NTD). Also, studies on viable "normal" offspring and strategies to prevent the effects of maternal diabetes are scarce; an understandable problem given that several extrinsic and intrinsic factors (including embryo susceptibility, among others), may contribute to CNS fetal programming in specific cell types, locations, and times.

Bibliographic Search

We searched for experimental and clinical evidence regarding the effect of hyperglycemia on the development of the CNS and fetal programming related to neurological diseases, but excluding evident NTD. We searched PubMed (https:// www.ncbi.nlm.nih.gov/pubmed) for studies on humans or other animals, published in English in a variety of article



types (classical article, clinical study, comparative study, evaluation studies, journal article, meta-analysis, and technical report) published between 1990/01/01 and 2018/01/31. We used a combination of the following keywords: maternal diabetes or pregnancy+hyperglycemia, in combination with neurological fetal programming, neurological outcomes, fetal neural development, or neural tube.

From a total of 525 articles, 76 remained after we eliminated those that contained the phrase "neural tube defect," those that were not related to CNS development and/or function, duplicated articles, and those published in a language other than English.

OBESITY AND DIABETES

Childhood obesity and type 2 diabetes are closely related to GDM. A systematic review that included 20 observational studies involving a total of 26,509 children showed that maternal hyperglycemia (GDM and type 1 diabetes) was associated with obesity and abnormal glucose tolerance in offspring. Interestingly, while higher body mass index was reported for the children of GDM mothers during childhood, the same was reported from prepuberty to adolescence in children from mothers with type 1 diabetes. Furthermore, offspring from GDM mothers had high 2-h plasma glucose from prepuberty to early adulthood, and those from mothers with type 1 diabetes had a high rate of type 2 diabetes from years 2 to 5 and early adulthood (16). On the other hand, the effect of diabetes during gestation in offspring can be generationally transmissive through the maternal line. Hanafi et al. (17) showed that rats with grand-maternal diabetes showed impaired glucose sensing, increased oxidative stress, insulin resistance, and impaired

Abbreviations: GDM, gestational diabetes mellitus; NTD, neural tube defects; NPY, neuropeptide Y; POMC, pro-opiomelanocortin; AgRP, agouti-related peptide; ARC, arcuate nuclei; PVN, paraventricular nuclei; CART, cocaine and amphetamine-related transcripts; Shh, Sonic hedgehog; Nkx2.1, thyroid transcription factor 1; NSCs, neural stem cells; ASD, Autism spectrum disorder.

glucose tolerance in F1 and F2, with more prominent effects in F2.

In an effort to study the mechanisms involved in the development of obesity in offspring from diabetic mothers, the plasma content of hormones involved in food intake and energy expenditure were measured in an Austrian cohort of children with a mean age of 6 years (male:female = 36:40) born from mothers with GDM, pre-gestational diabetes, and nondiabetic women. No differences were found in the plasma content of hormones involved in food intake and energy expenditure such as ghrelin, leptin, adiponectin, neuropeptide Y (NPY), peptide YY, and growth differentiation factor 15. However, using multiple regression analysis, the authors found that body mass index, leptin, and GDF-15 had independent effects on insulin resistance (18). It is worth mentioning that both GDF-15 and leptin are synthesized by white adipose tissue; the first decreases food intake, while the latter suppresses appetite and increases energy expenditure (19, 20). Furthermore, leptin is a hormone that acts via receptors in the hypothalamus, and the activation of leptin receptors in pro-opiomelanocortin (POMC) and agouti-related peptide (AgRP)/NPY neurons within the arcuate nuclei (ARC) lead to increased AgRP and POMC expression reducing food intake (19, 21). The absence of changes in plasma leptin in obese children from GDM, but its relation with body mass index may be explained through an impaired function in the leptin receptor expressed in the CNS, a phenomenon called, leptin resistance.

POMC is the precursor of α -melanocortin stimulating hormone, which in turn (through the activation of type 3 and 4 melanocortin receptors) is the most important component of the network responsible for controlling appetite, energy expenditure, glucose homeostasis and lipid metabolism in the hypothalamus (19, 22, 23). This peptide functions as an anorexigenic factor in ARC and paraventricular nuclei (PVN), predominantly reducing appetite (24). AgRP and NPY are orexigenic factors inducing hyperphagia and obesity (25–27).

The interaction between peripheric leptin and the melanocortin systems in the ARC and PVN is essential for the circuits that regulate food intake, energy expenditure, glucose, and lipid metabolism. Furthermore, it can be suggested that changes in hypothalamic fetal ontogenesis could be taking place in the diencephalon exposed to hyperglycemia, affecting the postnatal function of hypothalamic circuits because of the effects reported at early and late development in offspring from GDM and Type 2 diabetes mothers on leptin level, glucose homeostasis and BMI, mentioned above (16, 18). Indeed, several studies have shown that high glucose levels promote an inadequate organization of hypothalamic ARC and PVN, as well as malformation of the ventromedial hypothalamic nucleus in rats (28-31). Moreover, chicken embryos exposed to high glucose concentrations (30 mM) have lower glucose tolerance in neurons located in the hypothalamic infundibulum the equivalent anatomic area of the ARC (32).

On the other hand, the expression of hypothalamic neuropeptides in sheep ARC exposed to intrauterine hyperglycemia increases the expression of POMC and the cocaine and amphetamine-related transcripts (CART) at 81 and 140 days of gestation (33, 34). Such changes may affect



In normal conditions (gray lines and arrows), leptin activates POMC/CART neurons (pro-opiomelanocortin/cocaine- and amphetamine-regulated transcript) and inhibits NPY/AgRP neurons (neuropeptide Y/agouti-related protein) promoting satiety and a balance between food intake and energy expenditure. In diabetic offspring (black lines and arrows), a decrease in the activation of POMC/CART neurons takes place due to leptin resistance, which reduces melanocortin release at the PVN (paraventricular nuclei), promoting an increase in food intake leading to obesity and diabetes. 3V, third ventricle.

energy balance regulation in later life, affecting food intake and energy balance. To our knowledge, there is no postnatal evidence of an altered pathway for central energy balance in sheep, but there are some clues provided in the murine model. Offspring from diabetic rats and mice have increased susceptibility to body weight dysregulation and obesity due to: increased expression of the orexigenic (NPY and AgRP) and decreased expression of the anorexigenic (α -MSH) peptides in the ARC (30, 35), leptin resistance in 10-day old pups, decreased fiber density of AgRP and α -MSH peptides, as well as in the PVN, and increased food intake and body weight (31). These findings suggest that offspring born from diabetic dams showed leptin resistance in first-order neurons within the ARC, less synaptic transmission into the PVN and, consequently, obesity (**Figure 2**).

Altered levels of morphogens and transcription factors important in hypothalamic organization may affect hypothalamic fetal development under hyperglycemic conditions. During early fetal development, the hypothalamus emerges from the diencephalon, and its adequate formation depends on the precise regulation of molecular and cellular mechanisms orchestrated by regional morphogenetic organizers in the neural tube. The diencephalon is separated from surrounding regions by the influence of organizing signals (morphogens) such as Wingless/integrins, Sonic hedgehog (Shh), bone morphogenetic proteins and fibroblast growth factors, and later by the expression of the thyroid transcription factor 1 (Nk \times 2.1, a downstream gene of Shh) and a key factor for hypothalamic neuron development (36, 37).

Interestingly, Shh has been evaluated by *in situ* hybridization and qRT-PCR during brain development in embryos from streptozotocin-induced diabetic mice, at embryo day (E) 8.5 and E11.5. Normally, at E8.5 Shh expression is restricted to the ventral medial plate in the forebrain, but its domain is expanded and its expression increases in embryos from diabetic mice. At E11.5 in control animals, Shh expression is localized in the ventral diencephalon and telencephalon, but in the diencephalon from diabetic animals its expression is stronger and expanded toward the dorsal telencephalon. Thus, there is an increased and expanded expression of Nk×2.1 to the telencephalon (38). In addition, an increase in cell proliferation and neurogenesis has been reported in ventral telencephalon/diencephalon in E11.5 to diabetic mice (39).

These data suggest that early changes in the ventral-dorsal patterning and increased neurogenesis are contributing to defects in the fetal and postnatal hypothalamus in murine, chicken and sheep models; an aspect worthy of further study.

COGNITION

Cognition is involved in the regulation of emotional and social cues, including the formal measures of intelligence such as memory and attention. Cognitive functions studied in infants from diabetic mothers and offspring from different animal models include language (animal communication), learning, memory, motor coordination, perception, and problem-solving. All of these are functions that are coordinated in a complex manner by different anatomical structures such as the cerebral cortex, amygdala, hippocampus, and basal ganglia. However, the relationship between diabetes during pregnancy and impaired cognitive function after birth remains controversial. Some studies in humans have shown that maternal diabetes contributed to cognitive dysfunction in school-age children, which has been associated with changes in cell migration and differentiation during brain development (12, 40, 41). For example, Ornoy et al. (12) reported that children under 9 years old born to GDM women had lower scores in verbal tasks and fine and gross motor skills. Bolaños et al. (42) found that in utero hyperglycemia was associated with a lower average IQ and poor performance in working memory skills, such as graphic and visuospatial tasks, in children from 7 to 9years-old born to control and GDM women: They concluded that GDM leads to minor neurological deficits in children (42).

Working memory is the result of proper coordination between the prefrontal cortex and hippocampus; structures that arise from the dorsal telencephalon during embryo development.

Hyperglycemia during hippocampal development decreases synaptic plasticity and reduces memory durability in male rats

(43, 44), presumably due to a delay in normal hippocampal development regulated by insulin and insulin growth factor-1 receptors that lead to structural, behavioral, and cognitive abnormalities (45).

It is worth mentioning that the hippocampus is very vulnerable to several neurotoxic insults, including fetal hypoxia and iron deficiency, both of which are phenomena reported in fetuses from diabetic mothers (46–49). One way to evaluate early postnatal hippocampal function is through the measurement of event-related potentials (ERPs) generated by visual or auditory stimuli. ERPs are divided into two categories: sensory, which are early waves, peaking within the first 100 ms after the stimulus, and cognitive, which are waves generated later due to information processing (50). Using this tool, recognition memory (visual and auditory stimuli) was evaluated in 6-month-old infants from diabetic mothers, finding robust evidence of a memory deficit (51, 52).

During embryo development, the cerebral cortex and hippocampus arise from the dorsal telencephalic neuroepithelium. As in other areas of the CNS, intrinsic and extrinsic factors coordinate correct patterning during development by promoting the self-renewal of neural stem cells (NSC). These then will specify into neurons, astrocytes, and oligodendrocytes, to develop the functional areas of the cerebral cortex and the characteristic hippocampus anatomy. Diabetes during pregnancy in mice promotes changes in the expression of proliferative and differentiative related genes during brain development. However, NTD embryos or complete litters have been used, which makes it difficult to relate changes in gene expression during fetal development and postnatal impaired cognitive functions in offspring without NTD. Moreover, contrasting data have been reported regarding the effect of hyperglycemia on embryo cell proliferation, differentiation, and survival. Thus, an increase in cell proliferation without affection on cell death, which promotes thickening and deformation of the dorsal telencephalon in embryos to diabetic mice, was reported at E11.5. However, increased BrdU incorporation was not observed (38, 39). On the other hand, there are several reports which suggest increased cell death using the same model and embryo data (53-57). Despite some discrepancies, changes in cell proliferation, migration, and differentiation may be taking place in a different and complex manner, and even depending on the embryos susceptibility. Thus, researchers need to separate non-NTD from NTD embryos in order to define both shared and different mechanisms, and, in turn, propose and establish specific prevention, treatment, and diagnosis strategies beyond NTD

Even though the telencephalon is the most studied structure during development in health and disease, few studies have analyzed the effect of hyperglycemia in embryos with "normal" (non-NTD) development. However, the use of cortical NSC obtained from the dorsal telencephalon can be a useful tool to study the effect of hyperglycemia. Fu et al. (39), studied the effect of high glucose (30 mM) on the proliferation and differentiation of E13 cortical NSC from normal pregnant mice, and showed that high glucose in proliferative NSC promoted increased cell death and reduced cell proliferation, together with increased neuron, astrocyte, and oligodendrocyte differentiation in differentiated cells. They concluded that NSC cultured in high glucose led to cell-cycle arrest and apoptosis and influenced lineage specification, through a mechanism in which Shh, Bmps and the Notch/Delta pathway are involved (39).

The effect of high glucose on cell phenotype must be more complex, and other factors may be involved, such as micro RNAs (miRNAs, molecules with a potential use in therapeutic as well as noninvasive biomarkers), neurotransmitters and epigenetic mechanisms (chromatin modification, histone methylation, or acetylation and DNA methylation). The level of telencephalic development-related miRNAs was evaluated in serum from control and GDM women whose fetuses were not diagnosed with NTD. Interestingly, GDM led to higher levels of proliferative and neurogenic miRNAs (miR-183-5p, miR-200b-3p, miR-125-5p, and miR-1290). Moreover, gene ontology in-silico analysis revealed alterations in cell proliferation and neuron differentiation (5). Furthermore, an array study containing \sim 2,383 probes was used to analyze the *in vitro* and *in vivo* effects of high glucose in mice forebrain NSC. Results showed that high glucose deregulated 104 and 25 miRNAs in vivo and in vitro, respectively. Neurogenic miRNAs: miR-30 family, miR-125-5p, miR-124, and miR-128 were upregulated under both conditions (58). Furthermore, miR-183, considered a proliferative miRNA, was downregulated in NSC obtained in embryos from diabetic mice (59, 60). Although the authors focused on the miRNA-30 family, because it is known to be involved in schizophrenia, ASD, axon extension and guidance, and other neurodevelopmental disorders (61-63), it is clear that changes in miRNAs expression may have an extensive regulatory effect. One miRNA may regulate hundreds or thousands of RNAm, which may, in turn, affect cell proliferation, differentiation, migration, and death, through a complex network of epitrascriptomic regulation occurring in parallel.

The expression of miRNAs under hyperglycemic conditions may be regulated by epigenetic factors. To explore this circumstance under high glucose in utero and in vitro, Shyamasundar et al. (64) isolated E13.5 dorsal telencephalic tissue from diabetic and control pregnant mice litters or NSC from control mice that were maintained under normal or highglucose conditions during 48 h (40 mM). They reported that high glucose in vivo and in vitro alters chromatin reorganization due to an increase in histone H3K9 trimethylation and global DNA methylation, and provokes a decrease in histone H3K9 acetylation. Decreased gene expression due to high glucose was expected because H3K9 can turn genes on by becoming acetylated, and can silence when methylated. However, the authors reported that high glucose in NSCs in embryos from diabetic mice promoted a significant increase in the expression of doublecortin and Pafah1b1 (Platelet-activating factor acetylhydrolase isoform 1b, subunit 1), molecules essential for neuron migration and differentiation; this effect could be explained because no change in the CpG methylation status of the gene promoter was observed (55, 64, 65). They further determined that the decreased miR-200a, miR-200b, miR-466a-3p, and miR-466d-3p miRNAs were responsible for the changes observed in Dcx and Pafah1b1 (64), suggesting the epigenetic regulation of these miRNAs. Interestingly, a lower level of mir-200b was reported in serum samples from diabetic pregnant women at the third trimester, with a negative diagnosis for NTD (5).

Neurotransmitters may also contribute to increased neurogenesis observed in the cortical neuroepithelium of embryos under hyperglycemia with non-NTD. Our group reported that histamine increases cortical and mesencephalic NSC neurogenesis through H₁-receptor (H₁R) activation. Both this neurotransmitter and H₁R showed a significant increase in the cortical neuroepithelial of embryos from diabetic pregnant rats (66–68). In mammals, histamine is a neurotransmitter/neuromodulator in the adult brain, acting through G-protein coupled receptors [H₁R, H₂RH₃R, and H₄R] (69, 70). During cortical development, histamine displays high concentration as well as a high expression of H₁R and H₂R (71–73). On the other hand, in cortical NSC, HA promotes cell proliferation and neuron differentiation through H₁R and H₂R activation, respectively (66, 74–76).

In the diabetic model, the cortical neuroepithelium of embryos without NTD showed increased neurogenesis (E14) as well as histamine concentration (E14) and H_1R expression (E12). Interestingly, the systemic administration at E12 of chlorpheniramine (H_1R antagonist/inverse agonist) partially prevented increased dorsal telencephalic neurogenesis in embryos from diabetic rats, suggesting the participation of this receptor in the impaired neurogenesis observed in embryos from the diabetic model (68). The relevance of the above findings is also supported by evidence on the effect of antihistamine drugs on controlling glycemia under diabetic conditions (77).

MOTOR BEHAVIOR

Early school-age children born from mothers with GDM present motor impairment (12). As with cognition, several regions of the brain control motor activity, including the cerebral cortex and the cerebellum. As mentioned above, the dorsal telencephalon showed increased neurogenesis during fetal development. However, to our knowledge, postnatal studies on the anatomy and functional neurochemistry of the motor cortex have not been reported. In contrast, we found a few studies regarding the effect of hyperglycemia during embryo development and its effect on the cerebellum. A reduction in cerebellum size has been observed, which correlates with a decreased number of Purkinje and granular cells, and the reduced size of the molecular and granule cell layers in offspring of the streptozotocin-induced diabetic model in rats (Figure 3) (78, 79). Under these conditions, an increase in the synaptic length and dendritic spine was detected at postnatal days 30 and 70 (78), which suggests that the reduction in cerebellum size is compensated by an increase in the size of the dendritic spine. Nevertheless, it may also indicate inadequate cerebellar synaptic maturation (80).

Although postnatal cytoarchitectonic changes are reported in diabetic offspring, to our knowledge there is no evidence that



could explain the postnatal changes in the cerebellum during fetal development.

PSYCHIATRIC DISORDERS

Autism and schizophrenia have been related to maternal diabetes. Autism spectrum disorder (ASD) is a neurodevelopmental disorder with an estimated prevalence of 1 in 68 children in 2016 (81). ASD is characterized by stereotyped behavior, severe social dysfunction, restricted attention and language impairments (82-86). Although ASD has a highly heritable factor (86), stressful environment in utero has also been implicated (83, 84, 86-91). In 2009, a two-fold increase in the risk of ASD in infants from GDM mothers was reported. However, authors have concluded that infections during pregnancy, maternal age, hypertension, and preeclampsia contribute to the incidence of the disease (92). This is supported by other studies that reported no relationship between GDM and risk of ASD (93, 94). However, a cohort study of 66,445 pregnancies examined the relationship of obstetric complications with ASD, finding that GDM presented a significant risk increment in the pathology of 1.2% of the children (89). A case-control study also reported that children from GDM mothers had a high incidence of ASD with expressive language deficits (84). Discrepancies among studies may be due to differences in the study design and ethnic populations.

Alterations in the cerebellum have also been related to autism (95–97). One of the most consistently abnormal findings in the postmortem brains of autistic individuals, regardless of age, sex, and cognitive ability is the significant decrease in the number of Purkinje cells in the posterolateral-neocerebellar and the adjacent archicerebellar cortices (98, 99). Both Yamano et al. and Razi et al. have shown that *in utero* hyperglycemia reduces the size of the cerebellum due to a reduction in the number of Purkinje and granular cells (78, 79). Although changes in cerebellum architecture in autism have been observed, the physiological relevance of these relationships remains unsolved, probably due to the clinical heterogeneity within the broad behavioral phenotype (100).

Genetic studies have revealed three promising ASDimplicated genes: engrailed homeobox 2 (EN2), gammaaminobutyric acid type A receptor beta3 subunit (GABRB3), and MET proto-oncogene, receptor tyrosine kinase (MET). All have specific roles in cerebellar development (101, 102). The expression of En2 is restricted to midbrain and cerebellum. The loss of its function causes abnormal cerebellar foliation, with deficits in motor and social behavior (102). Interestingly, in embryos from diabetic mice a decrease in the expression of En2 has been reported (56, 103).

Several studies have shown a positive association between oxidative stress during fetal development and ASD, suggesting that oxidative damage is an important factor in the etiology of ASD (104, 105) and schizophrenia (106, 107). The release of reactive oxygen species (ROS) and the generation of oxidative stress under GDM is a critical aspect that may contribute.

On the other hand, schizophrenia is a chronic and severe psychiatric incapacitating disorder that affects a wide range of cognitive, emotional, and motor functions that have been associated with GDM. The symptoms include hallucinations or paranoid delusions, disorganized speech, and socialization, cognition, and memory impairment (108). The etiology of this disorder is unknown. However, epidemiological evidence suggests prenatal and perinatal complications as antecedents (109–112).

In a Swedish case-control cohort study, an increased susceptibility of developing schizophrenia in females from GDM mothers was reported, suggesting a gender susceptibility (113). A meta-analysis study reported that among the obstetric complications for schizophrenia, GDM was found to have a significant participation (114). Other studies described that there is no evidence of the association between GDM and the disease (109, 115). This inconsistency could be explained by the fact that most studies used questionnaires or scales where GDM or a history of maternal diabetes were not included.

On the other hand, it has been suggested that an increased level in plasma of pro-inflammatory cytokines such as IL-8 (interleukin-8) and tumor necrosis factor- α (TNF- α) during pregnancy could be related to offspring schizophrenia in adulthood (116). Thus, a hyperglycemic condition increases the expression of inflammatory cytokines in macrophages

by decreasing H3K9me3 levels in inflammatory cytokines promoters, such as TNF- α , and chemoattractant protein-1 and IL-6 (117).

Changes in the DNA methylation pattern have been reported in term placenta and cord-blood samples from GDM. Interestingly, the analysis showed that 57 genes associated with schizophrenia or schizoaffective disorders were affected (118). Moreover, several lines of evidence have shown that alterations in the degree of DNA and histone methylation are related to ASD and schizophrenia (119-122). All these data suggest that epigenetic changes during GDM may be related to offspring schizophrenia. A "two-hit" hypothesis has been proposed for this and other diseases, where the onset of disease cannot clearly be linked to a specific genetic or environmental insult. This is also applicable to ASD. For schizophrenia, it has been hypothesized that the first hit may affect neurogenic and cell specification pathways, while the second hit may have a greater effect on functional integration (123), two processes that have been suggested to be altered in embryos and early born diabetic animals (31, 38, 39, 56).

CONCLUSION

Morphological and functional alteration in the CNS and its relationship with changes in gene expression and maternal diabetes is complex. Particularly because other phenomena may be participating in the final function of a gene or group of genes, such as post-transcription, translation, or post-translational regulatory processes that define cell commitment, differentiation, migration, death, integration, and function during specific moments in development.

REFERENCES

- American Diabetes A. 2. Classification and diagnosis of diabetes: standards of medical care in diabetes-2018. *Diabetes Care* (2018) 41(Suppl 1):S13–27. doi: 10.2337/dc18-S002
- Linnenkamp U, Guariguata L, Beagley J, Whiting DR, Cho NH. The IDF Diabetes Atlas methodology for estimating global prevalence of hyperglycaemia in pregnancy. *Diabetes Res Clin Pract.* (2014) 103:186–96. doi: 10.1016/j.diabres.2013.11.004
- Reyes-Munoz E, Castellanos-Barroso G, Ramirez-Eugenio BY, Ortega-Gonzalez C, Parra A, Castillo-Mora A, et al. The risk of gestational diabetes mellitus among Mexican women with a history of infertility and polycystic ovary syndrome. *Fertil Steril.* (2012) 97:1467–71. doi: 10.1016/j.fertnstert.2012.03.023
- Reyes-Munoz E, Parra A, Castillo-Mora A, Ortega-Gonzalez C. Effect of the diagnostic criteria of the International Association of Diabetes and Pregnancy Study Groups on the prevalence of gestational diabetes mellitus in urban Mexican women: a cross-sectional study. *Endocr Pract.* (2012) 18:146–51. doi: 10.4158/EP11167.OR
- Lamadrid-Romero M, Solis KH, Cruz-Resendiz MS, Perez JE, Diaz NF, Flores-Herrera H, et al. Central nervous system development-related microRNAs levels increase in the serum of gestational diabetic women during the first trimester of pregnancy. *Neurosci Res.* (2017) 130:8–22. doi: 10.1016/j.neures.2017.08.003
- Dunne F, Brydon P, Smith K, Gee H. Pregnancy in women with Type 2 diabetes: 12 years outcome data 1990-2002. *Diabet Med.* (2003) 20:734–8. doi: 10.1046/j.1464-5491.2003.01017.x

Although fetal programming due to maternal diabetes may be evident, and a lot of information regarding the intrinsic and extrinsic factors that participate in the development of several areas of the CNS has been generated and related to NTD, we have not developed proper experimental protocols to generate information related to how hyperglycemia affects the development of specific areas of the CNS during critical time windows. It is also clear that, depending on the timing of insult in relation to the stages of brain development, different cell populations may be selectively affected. Furthermore, it is likely that early defects may be exclusively due to hyperglycemia, while defects occurring in late development may be due to high glucose and/or hyperinsulinemia. Finally, it is important to bear in mind that the problem is complex, and that no single molecular mechanism can fully explain the effects of maternal diabetes on fetal programming during neurodevelopment. Global approaches are needed.

AUTHOR CONTRIBUTIONS

All the authors contributed in the same way for the realization of this review. Additionally, AM-H and ND reviewed and edited the final version.

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- Salge AK, Rocha KM, Xavier RM, Ramalho WS, Rocha EL, Guimaraes JV, et al. Macroscopic placental changes associated with fetal and maternal events in diabetes mellitus. *Clinics* (2012) 67:1203–8. doi: 10.6061/clinics/2012(10)13
- Lehnen H, Zechner U, Haaf T. Epigenetics of gestational diabetes mellitus and offspring health: the time for action is in early stages of life. *Mol Hum Reprod.* (2013) 19:415–22. doi: 10.1093/molehr/gat020
- Entringer S, Buss C, Swanson JM, Cooper DM, Wing DA, Waffarn F, et al. Fetal programming of body composition, obesity, and metabolic function: the role of intrauterine stress and stress biology. J Nutr Metab. (2012) 2012:632548. doi: 10.1155/2012/632548
- Ornoy A, Ratzon N, Greenbaum C, Peretz E, Soriano D, Dulitzky M. Neurobehaviour of school age children born to diabetic mothers. *Arch Dis Child Fetal Neonatal Ed.* (1998) 79:F94–99. doi: 10.1136/fn.79.2.F94
- Stenninger E, Flink R, Eriksson B, Sahlen C. Long-term neurological dysfunction and neonatal hypoglycaemia after diabetic pregnancy. *Arch Dis Child Fetal Neonatal Ed.* (1998) 79:F174–9. doi: 10.1136/fn.79.3.F174
- Ornoy A, Wolf A, Ratzon N, Greenbaum C, Dulitzky M. Neurodevelopmental outcome at early school age of children born to mothers with gestational diabetes. *Arch Dis Child Fetal Neonatal Ed.* (1999) 81:F10–14. doi: 10.1136/fn.81.1.F10
- Nazer Herrera J, Garcia Huidobro M, Cifuentes Ovalle L. [Congenital malformations among offspring of diabetic women]. *Rev Med Chil.* (2005) 133:547–54. doi: 10.4067/S0034-98872005000500006
- Dionne G, Boivin M, Seguin JR, Perusse D, Tremblay RE. Gestational diabetes hinders language development in offspring. *Pediatrics* (2008) 122:e1073–9. doi: 10.1542/peds.2007-3028

- Nomura Y, Marks DJ, Grossman B, Yoon M, Loudon H, Stone J, et al. Exposure to gestational diabetes mellitus and low socioeconomic status: effects on neurocognitive development and risk of attentiondeficit/hyperactivity disorder in offspring. *Arch Pediatr Adolesc Med.* (2012) 166:337–43. doi: 10.1001/archpediatrics.2011.784
- Kawasaki M, Arata N, Miyazaki C, Mori R, Kikuchi T, Ogawa Y, et al. Obesity and abnormal glucose tolerance in offspring of diabetic mothers: a systematic review and meta-analysis. *PLoS ONE* (2018) 13:e0190676. doi: 10.1371/journal.pone.0190676
- Hanafi MY, Abdelkhalek TM, Saad MI, Saleh MM, Haiba MM, Kamel MA. Diabetes-induced perturbations are subject to intergenerational transmission through maternal line. *J Physiol Biochem.* (2016) 72:315–26. doi: 10.1007/s13105-016-0483-7
- Bozkurt L, Gobl CS, Rami-Merhar B, Winhofer Y, Baumgartner-Parzer S, Schober E, et al. The cross-link between adipokines, insulin resistance and obesity in offspring of diabetic pregnancies. *Horm Res Paediatr.* (2016) 86:300–8. doi: 10.1159/000448076
- Varela L, Horvath TL. Leptin and insulin pathways in POMC and AgRP neurons that modulate energy balance and glucose homeostasis. *EMBO Rep.* (2012) 13:1079–86. doi: 10.1038/embor.2012.174
- Adela R, Banerjee SK. GDF-15 as a target and biomarker for diabetes and cardiovascular diseases: a translational prospective. J Diabetes Res. (2015) 2015:490842. doi: 10.1155/2015/490842
- Lima LB, Metzger M, Furigo IC, Donato JJr. Leptin receptor-positive and leptin receptor-negative proopiomelanocortin neurons innervate an identical set of brain structures. *Brain Res.* (2016) 1646:366–76. doi: 10.1016/j.brainres.2016.06.024
- Ghamari-Langroudi M, Srisai D, Cone RD. Multinodal regulation of the arcuate/paraventricular nucleus circuit by leptin. *Proc Natl Acad Sci USA*. (2011) 108:355–60. doi: 10.1073/pnas.1016785108
- Sohn JW. Network of hypothalamic neurons that control appetite. BMB Rep. (2015) 48:229–33. doi: 10.5483/BMBRep.2015.48.4.272
- Schwartz MW, Woods SC, Porte D Jr, Seeley RJ, Baskin DG. Central nervous system control of food intake. *Nature* (2000) 404:661–71. doi: 10.1038/35007534
- Hahn TM, Breininger JF, Baskin DG, Schwartz MW. Coexpression of Agrp and NPY in fasting-activated hypothalamic neurons. *Nat Neurosci.* (1998) 1:271–2. doi: 10.1038/1082
- Woods SC, Schwartz MW, Baskin DG, Seeley RJ. Food intake and the regulation of body weight. *Annu Rev Psychol.* (2000) 51:255–77. doi: 10.1146/annurev.psych.51.1.255
- 27. Morton GJ, Meek TH, Schwartz MW. Neurobiology of food intake in health and disease. *Nat Rev Neurosci.* (2014) 15:367–78. doi: 10.1038/nrn3745
- Plagemann A, Harder T, Rake A, Melchior K, Rittel F, Rohde W, et al. Hypothalamic insulin and neuropeptide Y in the offspring of gestational diabetic mother rats. *Neuroreport* (1998) 9:4069–73. doi: 10.1097/00001756-199812210-00012
- Plagemann A, Harder T, Janert U, Rake A, Rittel F, Rohde W, et al. Malformations of hypothalamic nuclei in hyperinsulinemic offspring of rats with gestational diabetes. *Dev Neurosci.* (1999) 21:58–67. doi: 10.1159/000017367
- Franke K, Harder T, Aerts L, Melchior K, Fahrenkrog S, Rodekamp E, et al. 'Programming' of orexigenic and anorexigenic hypothalamic neurons in offspring of treated and untreated diabetic mother rats. *Brain Res.* (2005) 1031:276–83. doi: 10.1016/j.brainres.2004.11.006
- Steculorum SM, Bouret SG. Maternal diabetes compromises the organization of hypothalamic feeding circuits and impairs leptin sensitivity in offspring. *Endocrinology* (2011) 152:4171–9. doi: 10.1210/en.2011-1279
- Tzschentke B, Bogatyrev S, Schellong K, Rancourt RC, Plagemann A. Temporary prenatal hyperglycemia leads to postnatal neuronal 'glucoseresistance' in the chicken hypothalamus. *Brain Res.* (2015) 1618:231–40. doi: 10.1016/j.brainres.2015.05.037
- 33. Muhlhausler BS, Adam CL, Marrocco EM, Findlay PA, Roberts CT, McFarlane JR, et al. Impact of glucose infusion on the structural and functional characteristics of adipose tissue and on hypothalamic gene expression for appetite regulatory neuropeptides in the sheep fetus during late gestation. J Physiol. (2005) 565(Pt 1), 185–195. doi: 10.1113/jphysiol.2004.079079

- 34. Adam CL, Findlay PA, Chanet A, Aitken RP, Milne JS, Wallace JM. Expression of energy balance regulatory genes in the developing ovine fetal hypothalamus at midgestation and the influence of hyperglycemia. *Am J Physiol Regul Integr Comp Physiol.* (2008) 294:R1895–900. doi: 10.1152/ajpregu.00163.2008
- Grove KL, Smith MS. Ontogeny of the hypothalamic neuropeptide Y system. *Physiol Behav.* (2003) 79:47–63. doi: 10.1016/S0031-9384(03)00104-5
- Ishii Y, Bouret SG. Embryonic birthdate of hypothalamic leptinactivated neurons in mice. *Endocrinology* (2012) 153:3657–67. doi: 10.1210/en.2012-1328
- Martinez-Ferre A, Martinez S. Molecular regionalization of the diencephalon. Front Neurosci. (2012) 6:73. doi: 10.3389/fnins.2012.00073
- Liao DM, Ng YK, Tay SS, Ling EA, Dheen ST. Altered gene expression with abnormal patterning of the telencephalon in embryos of diabetic Albino Swiss mice. *Diabetologia* (2004) 47:523–31. doi: 10.1007/s00125-004-1351-5
- Fu J, Tay SS, Ling EA, Dheen ST. High glucose alters the expression of genes involved in proliferation and cell-fate specification of embryonic neural stem cells. *Diabetologia* (2006) 49:1027–38. doi: 10.1007/s00125-006-0153-3
- Clausen TD, Mortensen EL, Schmidt L, Mathiesen ER, Hansen T, Jensen DM, et al. Cognitive function in adult offspring of women with gestational diabetes-the role of glucose and other factors. *PLoS ONE* (2013) 8:e67107. doi: 10.1371/journal.pone.0067107
- Camprubi Robles M, Campoy C, Garcia Fernandez L, Lopez-Pedrosa JM, Rueda R, Martin MJ. Maternal diabetes and cognitive performance in the offspring: a systematic review and meta-analysis. *PLoS ONE* (2015) 10:e0142583. doi: 10.1371/journal.pone.0142583
- Bolanos L, Matute E, Ramirez-Duenas Mde L, Zarabozo D. Neuropsychological impairment in school-aged children born to mothers with gestational diabetes. *J Child Neurol.* (2015) 30:1616–24. doi: 10.1177/0883073815575574
- Delascio Lopes C, Sinigaglia-Coimbra R, Mazzola J, Camano L, Mattar R. Neurofunctional evaluation of young male offspring of rat dams with diabetes induced by streptozotocin. *ISRN Endocrinol.* (2011) 2011:480656. doi: 10.5402/2011/480656
- 44. Hami J, Kheradmand H, Haghir H. Gender differences and lateralization in the distribution pattern of insulin-like growth factor-1 receptor in developing rat hippocampus: an immunohistochemical study. *Cell Mol Neurobiol.* (2014) 34:215–26. doi: 10.1007/s10571-013-0005-x
- Hami J, Sadr-Nabavi A, Sankian M, Balali-Mood M, Haghir H. The effects of maternal diabetes on expression of insulin-like growth factor-1 and insulin receptors in male developing rat hippocampus. *Brain Struct Funct.* (2013) 218:73–84. doi: 10.1007/s00429-011-0377-y
- Georgieff MK, Widness JA, Mills MM, Stonestreet BS. The effect of prolonged intrauterine hyperinsulinemia on iron utilization in fetal sheep. *Pediatr Res.* (1989) 26:467–9. doi: 10.1203/00006450-198911000-00021
- Georgieff MK, Landon MB, Mills MM, Hedlund BE, Faassen AE, Schmidt RL, et al. Abnormal iron distribution in infants of diabetic mothers: spectrum and maternal antecedents. *J Pediatr.* (1990) 117:455–61. doi: 10.1016/S0022-3476(05)81097-2
- Georgieff MK, Schmidt RL, Mills MM, Radmer WJ, Widness JA. Fetal iron and cytochrome c status after intrauterine hypoxemia and erythropoietin administration. *Am J Physiol.* (1992) 262(3 Pt 2):R485–91. doi: 10.1152/ajpregu.1992.262.3.R485
- Petry CD, Eaton MA, Wobken JD, Mills MM, Johnson DE, Georgieff MK. Iron deficiency of liver, heart, and brain in newborn infants of diabetic mothers. J Pediatr. (1992) 121:109–14. doi: 10.1016/S0022-3476(05)82554-5
- Sur S, Sinha VK. Event-related potential: an overview. *Ind Psychiatry J.* (2009) 18:70–3. doi: 10.4103/0972-6748.57865
- Deregnier RA, Nelson CA, Thomas KM, Wewerka S, Georgieff MK. Neurophysiologic evaluation of auditory recognition memory in healthy newborn infants and infants of diabetic mothers. *J Pediatr.* (2000) 137:777– 84. doi: 10.1067/mpd.2000.109149
- Nelson CA, Wewerka S, Thomas KM, Tribby-Walbridge S, deRegnier R, Georgieff M. Neurocognitive sequelae of infants of diabetic mothers. *Behav Neurosci.* (2000) 114:950–6. doi: 10.1037/0735-7044.114.5.950
- Jiang Y, He SK. [The effect of oxidative stress on retinal pigment epithelial cell migration]. *Zhonghua Yan Ke Za Zhi* (2005) 41:100–5. doi: 10.3760/j:issn:0412-4081.2005.02.002

- Moores CA, Perderiset M, Kappeler C, Kain S, Drummond D, Perkins SJ, et al. Distinct roles of doublecortin modulating the microtubule cytoskeleton. *EMBO J.* (2006) 25:4448–57. doi: 10.1038/sj.emboj.7601335
- Jiang B, Kumar SD, Loh WT, Manikandan J, Ling EA, Tay SS, et al. Global gene expression analysis of cranial neural tubes in embryos of diabetic mice. *J Neurosci Res.* (2008) 86:3481–93. doi: 10.1002/jnr.21800
- Pavlinkova G, Salbaum JM, Kappen C. Maternal diabetes alters transcriptional programs in the developing embryo. *BMC Genomics* (2009) 10:274. doi: 10.1186/1471-2164-10-274
- 57. Wentzel P, Eriksson UJ. Altered gene expression in rat cranial neural crest cells exposed to a teratogenic glucose concentration *in vitro*: paradoxical downregulation of antioxidative defense genes. *Birth Defects Res B Dev Reprod Toxicol.* (2011) 92:487–97. doi: 10.1002/bdrb.20321
- Ramya S, Shyamasundar S, Bay BH, Dheen ST. Maternal diabetes alters expression of MicroRNAs that regulate genes critical for neural tube development. *Front Mol Neurosci.* (2017) 10:237. doi: 10.3389/fnmol.2017.00237
- Barca-Mayo O, De Pietri Tonelli D. Convergent microRNA actions coordinate neocortical development. *Cell Mol Life Sci.* (2014) 71:2975–95. doi: 10.1007/s00018-014-1576-5
- Diaz NF, Cruz-Resendiz MS, Flores-Herrera H, Garcia-Lopez G, Molina-Hernandez A. MicroRNAs in central nervous system development. *Rev Neurosci.* (2014) 25:675–86. doi: 10.1515/revneuro-2014-0014
- Mellios N, Sur M. The emerging role of microRNAs in schizophrenia and autism spectrum disorders. *Front Psychiatry* (2012) 3:39. doi: 10.3389/fpsyt.2012.00039
- Hancock ML, Preitner N, Quan J, Flanagan JG. MicroRNA-132 is enriched in developing axons, locally regulates Rasa1 mRNA, and promotes axon extension. *J Neurosci.* (2014) 34:66–78. doi: 10.1523/JNEUROSCI.3371-13.2014
- Sun T, Li S, Yang J, Yin Y, Ling S. Identification of a microRNA regulator for axon guidance in the olfactory bulb of adult mice. *Gene* (2014) 547:319–28. doi: 10.1016/j.gene.2014.06.063
- 64. Shyamasundar S, Jadhav SP, Bay BH, Tay SS, Kumar SD, Rangasamy D, et al. Analysis of epigenetic factors in mouse embryonic neural stem cells exposed to hyperglycemia. *PLoS ONE* (2013) 8:e65945. doi: 10.1371/journal.pone.0065945
- Hirotsune S, Fleck MW, Gambello MJ, Bix GJ, Chen A, Clark GD, et al. Graded reduction of Pafah1b1 (Lis1) activity results in neuronal migration defects and early embryonic lethality. *Nat Genet.* (1998) 19:333–9. doi: 10.1038/1221
- Molina-Hernandez A, Velasco I. Histamine induces neural stem cell proliferation and neuronal differentiation by activation of distinct histamine receptors. J Neurochem. (2008) 106:706–17. doi: 10.1111/j.1471-4159.2008.05424.x
- Escobedo-Avila I, Vargas-Romero F, Molina-Hernandez A, Lopez-Gonzalez R, Cortes D, De Carlos JA, et al. Histamine impairs midbrain dopaminergic development in vivo by activating histamine type 1 receptors. *Mol Brain* (2014) 7:58. doi: 10.1186/s13041-014-0058-x
- Solis KH, Mendez LI, Garcia-Lopez G, Diaz NF, Portillo W, De Nova-Ocampo M, et al. The histamine H1 receptor participates in the increased dorsal telencephalic neurogenesis in embryos from diabetic rats. *Front Neurosci.* (2017) 11:676. doi: 10.3389/fnins.2017.00676
- Haas HL, Sergeeva OA, Selbach O. Histamine in the nervous system. *Physiol Rev.* (2008) 88:1183–241. doi: 10.1152/physrev.00043.2007
- Connelly WM, Shenton FC, Lethbridge N, Leurs R, Waldvogel HJ, Faull RL, et al. The histamine H4 receptor is functionally expressed on neurons in the mammalian CNS. *Br J Pharmacol.* (2009) 157:55–63. doi: 10.1111/j.1476-5381.2009.00227.x
- Vanhala A, Yamatodani A, Panula P. Distribution of histamine-, 5hydroxytryptamine-, and tyrosine hydroxylase-immunoreactive neurons and nerve fibers in developing rat brain. *J Comp Neurol.* (1994) 347:101–14. doi: 10.1002/cne.903470108
- Kinnunen A, Lintunen M, Karlstedt K, Fukui H, Panula P. In situ detection of H1-receptor mRNA and absence of apoptosis in the transient histamine system of the embryonic rat brain. J Comp Neurol. (1998) 394:127–37. doi: 10.1002/(SICI)1096-9861(19980427)394:1<127::AID-CNE10>3.0.CO;2-L

- Karlstedt K, Senkas A, Ahman M, Panula P. Regional expression of the histamine H(2) receptor in adult and developing rat brain. *Neuroscience* (2001) 102:201–8. doi: 10.1016/S0306-4522(00)00464-4
- Molina-Hernandez A, Diaz NF, Arias-Montano JA. Histamine in brain development. J Neurochem. (2012) 122:872–82. doi: 10.1111/j.1471-4159.2012.07863.x
- Rodriguez-Martinez G, Velasco I, Garcia-Lopez G, Solis KH, Flores-Herrera H, Diaz NF, et al. Histamine is required during neural stem cell proliferation to increase neuron differentiation. *Neuroscience* (2012) 216:10– 7. doi: 10.1016/j.neuroscience.2012.04.049
- 76. Molina-Hernandez A, Rodriguez-Martinez G, Escobedo-Avila I, Velasco I. Histamine up-regulates fibroblast growth factor receptor 1 and increases FOXP2 neurons in cultured neural precursors by histamine type 1 receptor activation: conceivable role of histamine in neurogenesis during cortical development *in vivo. Neural Dev.* (2013) 8:4. doi: 10.1186/1749-81 04-8-4
- 77. Pini A, Obara I, Battell E, Chazot PL, Rosa AC. Histamine in diabetes: is it time to reconsider? *Pharmacol Res.* (2016) 111:316–24. doi: 10.1016/j.phrs.2016.06.021
- Yamano T, Shimada M, Fujizeki Y, Kawasaki H, Onaga A. Quantitative synaptic changes on Purkinje cell dendritic spines of rats born from streptozotocin-induced diabetic mothers. *Brain Dev.* (1986) 8:269–73. doi: 10.1016/S0387-7604(86)80080-8
- Razi EM, Ghafari S, Golalipour MJ. Effect of gestational diabetes on purkinje and granule cells distribution of the rat cerebellum in 21 and 28 days of postnatal life. *Basic Clin Neurosci.* (2015) 6:6–13.
- Hashimoto K, Kano M. Functional differentiation of multiple climbing fiber inputs during synapse elimination in the developing cerebellum. *Neuron* (2003) 38:785–96. doi: 10.1016/S0896-6273(03)00298-8
- CDC (2016). Autism Spectrum Disorder (ASD) [Online]. Available online at: http://www.cdc.gov/ncbddd/autism/index.html (Accessed April 2016).
- Levy SE, Mandell DS, Schultz RT. Autism. Lancet (2009) 374:1627–38. doi: 10.1016/S0140-6736(09)61376-3
- Guinchat V, Thorsen P, Laurent C, Cans C, Bodeau N, Cohen D. Pre-, periand neonatal risk factors for autism. *Acta Obstet Gynecol Scand.* (2012) 91:287–300. doi: 10.1111/j.1600-0412.2011.01325.x
- Krakowiak P, Walker CK, Bremer AA, Baker AS, Ozonoff S, Hansen RL, et al. Maternal metabolic conditions and risk for autism and other neurodevelopmental disorders. *Pediatrics* (2012) 129:e1121–8. doi: 10.1542/peds.2011-2583
- Ousley O, Cermak T. Autism spectrum disorder: defining dimensions and subgroups. *Curr Dev Disord Rep.* (2014) 1:20–8. doi: 10.1007/s40474-013-0003-1
- Xu G, Jing J, Bowers K, Liu B, Bao W. Maternal diabetes and the risk of autism spectrum disorders in the offspring: a systematic review and meta-analysis. J Autism Dev Disord. (2014) 44:766–75. doi: 10.1007/s10803-013-1928-2
- Brimacombe M, Ming X, Lamendola M. Prenatal and birth complications in autism. *Matern Child Health J.* (2007) 11:73–9. doi: 10.1007/s10995-006-0142-7
- Zimmerman AW, Connors SL, Matteson KJ, Lee LC, Singer HS, Castaneda JA, et al. Maternal antibrain antibodies in autism. *Brain Behav Immun.* (2007) 21:351–7. doi: 10.1016/j.bbi.2006.08.005
- Lyall K, Pauls DL, Spiegelman D, Ascherio A, Santangelo SL. Pregnancy complications and obstetric suboptimality in association with autism spectrum disorders in children of the Nurses' Health Study II. *Autism Res.* (2012) 5:21–30. doi: 10.1002/aur.228
- Field SS. Interaction of genes and nutritional factors in the etiology of autism and attention deficit/hyperactivity disorders: a case control study. *Med Hypotheses* (2014) 82:654–61. doi: 10.1016/j.mehy.2014.02.021
- Maramara LA, He W, Ming X. Pre- and perinatal risk factors for autism spectrum disorder in a New Jersey cohort. *J Child Neurol.* (2014) 29:1645–51. doi: 10.1177/0883073813512899
- Gardener H, Spiegelman D, Buka SL. Prenatal risk factors for autism: comprehensive meta-analysis. Br J Psychiatry (2009) 195:7–14. doi: 10.1192/bjp.bp.108.051672
- 93. Burstyn I, Sithole F, Zwaigenbaum L. Autism spectrum disorders, maternal characteristics and obstetric complications among singletons born in Alberta, Canada. *Chronic Dis Can.* (2010) 30:125–34.

- Zhang X, Lv CC, Tian J, Miao RJ, Xi W, Hertz-Picciotto I, et al. Prenatal and perinatal risk factors for autism in China. J Autism Dev Disord. (2010) 40:1311–21. doi: 10.1007/s10803-010-0992-0
- 95. Courchesne E. New evidence of cerebellar and brainstem hypoplasia in autistic infants, children and adolescents: the MR imaging study by Hashimoto and colleagues. J Autism Dev Disord. (1995) 25:19–22. doi: 10.1007/BF02178164
- 96. Schmitz C, Rezaie P. The neuropathology of autism: where do we stand? *Neuropathol Appl Neurobiol.* (2008) 34:4–11. doi: 10.1111/j.1365-2990.2007.00872.x
- van Kooten IA, Palmen SJ, von Cappeln P, Steinbusch HW, Korr H, Heinsen H, et al. Neurons in the fusiform gyrus are fewer and smaller in autism. *Brain* (2008) 131(Pt 4):987–99. doi: 10.1093/brain/ awn033
- Whitney ER, Kemper TL, Bauman ML, Rosene DL, Blatt GJ. Cerebellar Purkinje cells are reduced in a subpopulation of autistic brains: a stereological experiment using calbindin-D28k. *Cerebellum* (2008) 7:406–16. doi: 10.1007/s12311-008-0043-y
- Fatemi SH, Aldinger KA, Ashwood P, Bauman ML, Blaha CD, Blatt GJ, et al. Consensus paper: pathological role of the cerebellum in autism. *Cerebellum* (2012) 11:777–807. doi: 10.1007/s12311-012-0355-9
- 100. Miles JH. Autism spectrum disorders-a genetics review. Genet Med. (2011) 13:278-94. doi: 10.1097/GIM.0b013e3181f f67ba
- 101. Ieraci A, Forni PE, Ponzetto C. Viable hypomorphic signaling mutant of the Met receptor reveals a role for hepatocyte growth factor in postnatal cerebellar development. *Proc Natl Acad Sci USA*. (2002) 99:15200–5. doi: 10.1073/pnas.222362099
- 102. Cheh MA, Millonig JH, Roselli LM, Ming X, Jacobsen E, Kamdar S, et al. En2 knockout mice display neurobehavioral and neurochemical alterations relevant to autism spectrum disorder. *Brain Res.* (2006) 1116:166–76. doi: 10.1016/j.brainres.2006.07.086
- Eriksson UJ, Wentzel P. The status of diabetic embryopathy. Ups J Med Sci. (2016) 121:96–112. doi: 10.3109/03009734.2016.1165317
- 104. Chauhan A, Chauhan V, Brown WT, Cohen I. Oxidative stress in autism: increased lipid peroxidation and reduced serum levels of ceruloplasmin and transferrin-the antioxidant proteins. *Life Sci.* (2004) 75:2539–49. doi: 10.1016/j.lfs.2004.04.038
- 105. Ming X, Stein TP, Brimacombe M, Johnson WG, Lambert GH, Wagner GC. Increased excretion of a lipid peroxidation biomarker in autism. *Prostaglandins Leukot Essent Fatty Acids* (2005) 73:379–84. doi: 10.1016/j.plefa.2005.06.002
- 106. Meyer U. Developmental neuroinflammation and schizophrenia. Prog Neuropsychopharmacol Biol Psychiatry (2013) 42:20–34. doi: 10.1016/j.pnpbp.2011.11.003
- 107. Moller M, Swanepoel T, Harvey BH. Neurodevelopmental animal models reveal the convergent role of neurotransmitter systems, inflammation, and oxidative stress as biomarkers of schizophrenia: implications for novel drug development. ACS Chem Neurosci. (2015) 6:987–1016. doi: 10.1021/cn5003368
- 108. Insel TR. Rethinking schizophrenia. Nature (2010) 468:187–93. doi: 10.1038/nature09552
- 109. Dassa D, Sham PC, van Os J, Abel K, Jones P, Murray RM. Relationship of birth season to clinical features, family history, and obstetric complication in schizophrenia. *Psychiatry Res.* (1996) 64:11–7. doi: 10.1016/0165-1781(96)02868-5

- 110. Dalman C, Allebeck P, Cullberg J, Grunewald C, Koster M. Obstetric complications and the risk of schizophrenia: a longitudinal study of a national birth cohort. Arch Gen Psychiatry (1999) 56:234–40. doi: 10.1001/archpsyc.56.3.234
- 111. Konnecke R, Hafner H, Maurer K, Loffler W, der Heiden W. Main risk factors for schizophrenia: increased familial loading and pre- and perinatal complications antagonize the protective effect of oestrogen in women. *Schizophr Res.* (2000) 44:81–93. doi: 10.1016/S0920-9964(99)00139-5
- Preti A, Cardascia L, Zen T, Marchetti M, Favaretto G, Miotto P. Risk for obstetric complications and schizophrenia. *Psychiatry Res.* (2000) 96:127–39. doi: 10.1016/S0165-1781(00)00185-2
- 113. Hultman CM, Sparen P, Takei N, Murray RM, Cnattingius S. Prenatal and perinatal risk factors for schizophrenia, affective psychosis, and reactive psychosis of early onset: case-control study. *BMJ* (1999) 318:421–6. doi: 10.1136/bmj.318.7181.421
- 114. Cannon M, Jones PB, Murray RM. Obstetric complications and schizophrenia: historical and meta-analytic review. Am J Psychiatry (2002) 159:1080–92. doi: 10.1176/appi.ajp.159.7.1080
- 115. McNeil TF. Perinatal risk factors and schizophrenia: selective review and methodological concerns. *Epidemiol Rev.* (1995) 17:107–12. doi: 10.1093/oxfordjournals.epirev.a036165
- 116. Brown AS, Hooton J, Schaefer CA, Zhang H, Petkova E, Babulas V, et al. Elevated maternal interleukin-8 levels and risk of schizophrenia in adult offspring. Am J Psychiatry (2004) 161:889–95. doi: 10.1176/appi.ajp.161.5.889
- 117. Dinh W, Futh R, Nickl W, Krahn T, Ellinghaus P, Scheffold T, et al. Elevated plasma levels of TNF-alpha and interleukin-6 in patients with diastolic dysfunction and glucose metabolism disorders. *Cardiovasc Diabetol.* (2009) 8:58. doi: 10.1186/1475-2840-8-58
- 118. Ruchat SM, Houde AA, Voisin G, St-Pierre J, Perron P, Baillargeon JP, et al. Gestational diabetes mellitus epigenetically affects genes predominantly involved in metabolic diseases. *Epigenetics* (2013) 8:935–43. doi: 10.4161/epi.25578
- Schanen NC. Epigenetics of autism spectrum disorders. Hum Mol Genet. (2006) 15 Spec No 2:R138–50. doi: 10.1093/hmg/ddl213
- Akbarian S. Epigenetics of schizophrenia. Curr Top Behav Neurosci. (2010) 4:611–28. doi: 10.1007/7854_2010_38
- Akbarian S. The molecular pathology of schizophrenia–focus on histone and DNA modifications. *Brain Res Bull.* (2010) 83:103–7. doi: 10.1016/j.brainresbull.2009.08.018
- 122. Shorter KR, Miller BH. Epigenetic mechanisms in schizophrenia. Prog Biophys Mol Biol. (2015) 118:1–7. doi: 10.1016/j.pbiomolbio.2015.04.008
- 123. Maynard TM, Sikich L, Lieberman JA, LaMantia AS. Neural development, cell-cell signaling, and the "two-hit" hypothesis of schizophrenia. *Schizophr Bull*. (2001) 27:457–76. doi: 10.1093/oxfordjournals.schbul.a006887

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