



Mechanosensitive Ion Channel Piezo1 Regulates Diet-Induced Adipose Inflammation and Systemic Insulin Resistance

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Adipocytes function as an energy buffer and undergo significant size and volume changes in response to nutritional cues. This adipocyte plasticity is important for systemic lipid metabolism and insulin sensitivity. Accompanying the adipocyte size and volume changes, the mechanical pressure against cell membrane also changes. However, the role that mechanical pressure plays in lipid metabolism and insulin sensitivity remains to be elucidated. Here we show that Piezo1, a mechanically-activated cation channel stimulated by membrane tension and stretch, was highly expressed in adipocytes. Adipose Piezo1 expression was increased in obese mice. Adipose-specific piezo1 knockout mice (adipose-Piezo1^{-/-}) developed insulin resistance, especially when challenged with a high-fat diet (HFD). Perigonadal white adipose tissue (pgWAT) weight was reduced while pro-inflammatory and lipolysis genes were increased in the pgWAT of HFD-fed adipose-Piezo1^{-/-} mice. The adipose-Piezo1^{-/-} mice also developed hepatic steatosis with elevated expression of fatty acid synthesis genes. In cultured adipocytes, Piezo1 activation decreased, while Piezo1 inhibition elevated pro-inflammatory gene expression. TLR4 antagonist TAK-242 abolished adipocyte inflammation induced by Piezo1 inhibition. Thus, adipose Piezo1 may serve as an adaptive mechanism for adipocyte plasticity restraining pro-inflammatory response in obesity.

Keywords: adipocytes, insulin sensitivity, Piezo1, inflammation, lipolysis, TLR4

INTRODUCTION

Adipocytes are responsible for storing food energy in the form of fat. This function demands the cells to be highly plastic and dynamic in response to nutritional cues. When nutrients are sufficient, adipocytes increase their size and volume for fat storage. Under the condition of limited nutrient availability, adipocytes undergo lipolysis to release fat to provide energy for other organs (1, 2).

The balance between fat storage and lipolysis plays a major role in regulating systemic lipid metabolism and insulin sensitivity. In obesity, adipose plasticity is impaired and the balance of fat storage and lipolysis is disrupted, leading to adipose inflammation and enhanced lipolysis, which in turn cause systemic insulin resistance due to ectopic lipid deposition in other metabolic organs such as liver (3, 4). Insulin resistance is an independent risk factor for type 2 diabetes, cardiovascular disease and multiple cancers (5–8).

The molecular mechanisms for the impaired adipocyte plasticity in obesity are not well understood. The increased triglycerides in adipocytes in obesity apply significant pressure to the plasma membrane from inside out. At the meantime, adipose fibrosis developed in obesity also exerts pressure from outside in. Therefore, the plasma membrane of adipocytes is under constant mechanical pressures, which make adipocytes “stiffer” and less plastic. Adipocyte stiffness is one major mechanism for adipose inflammation and systemic insulin resistance in obesity (9–11). However, the mechanistic link between adipocyte mechanical pressure and plasticity-mediated inflammation and insulin resistance remains to be elucidated.

Piezo1 is recently identified as a novel mechanically-gated ion channel that transduces mechanical stimuli into electrical and chemical signals in mammalian cells (12, 13). Piezo1 is highly expressed in tissues with high mechanical pressures and plays an essential role in vascular development (14, 15), blood pressure regulation (16), and red blood cell volume control (17, 18). Piezo1 expression is also increased in bladder carcinoma (19), glioma (20), and breast cancer (21). However, the roles of Piezo1 in regulating adipocyte function have not been reported. In the current study, we found that Piezo1 is highly expressed in adipose tissue. Deletion of Piezo1 specifically in adipocytes reduces perigonadal fat mass and causes adipose inflammation, insulin resistance, and hepatic steatosis. Therefore, Piezo1 plays a key role in regulating adipose plasticity and insulin resistance in obesity.

MATERIALS AND METHODS

Mice

Adiponectin-Cre mice were purchased from the Jackson Laboratory (Stock No: 010803). The Piezo1-flox and Piezo1-tdTomato were generously provided by Dr. Ardem Patapoutian at The Scripps Research Institute. To generate adipose-specific Piezo1 knockout mice, Piezo1-flox/flox mice were crossed to adiponectin-Cre mice. Mice were housed four or five per cage and maintained under a 12 h light/12 h dark cycle at constant temperature (23°C) with *ad libitum* access to normal laboratory chow (#2920X; Harlan Teklad) and water. For diet-induced obesity, mice were fed a high-fat diet containing 54.8% fat calories, 24.0% carbohydrate calories, and 21.2% protein calories (4.8 kcal/g) (TD.93075; Envigo Inc.) from 4 weeks of age for 30–35 weeks. Body weight was measured biweekly. All mouse studies were conducted in accordance with federal guidelines and were approved by the Institutional Animal Care and Use Committee of the University of California, Irvine.

GTT and ITT

For the glucose tolerance test (GTT) and insulin tolerance test (ITT), food was removed at 10:00 a.m. GTTs were performed 5 h after food removal by the intraperitoneally (i.p.) injecting glucose at 1 g/kg of body weight. ITTs were performed with i.p. injections of recombinant regular human insulin (Novolin R, ReliOn) at a dose of 0.75 to 1.2 U/kg of body weight. Blood glucose was measured at the indicated time points (One Touch Ultra glucometer, Johnson and Johnson).

Serum Insulin, NEFA and Triglyceride Measurements

Serum insulin was measured with Ultrasensitive Insulin ELISA kit (90080, Crystal Chem). Serum non-esterified fatty acids (NEFA) were measured using NEFA Assay kit (276-76491, Wako). Total lipid from liver was isolated using Folch method (22). Serum and tissue triglyceride were measured using L-Type triglyceride Assay kit (461-09092, Wako). For the stimulated lipolysis, NEFA was measured in serum taken from mice 20 min after intraperitoneal injections of β 3-adrenergic agonist CL 316,243 (C5976, Sigma, 1 mg/kg).

Mouse Primary SVF and Adipocytes Isolation and Cell Culture

Mouse subcutaneous inguinal fat pads were removed and washed with PBS pH7.4 (Gibco) and then minced. The minced tissue was digested for 15–20 min at 37°C in the digestion buffer (10 mM CaCl₂, 2.5 unit/ml collagenase D, 2.4 units/ml Dispase II in PBS). The digested tissue was filtered through a 100 μ m mesh and centrifuged at 600 \times G for 5 min. Floating adipocyte fraction was removed. The resulting pellets were resuspended and further filtered through the 40 μ m nylon cell strainers (BD Biosciences). Stromal vascular fraction (SVF) cells were maintained in DMEM/F12 GlutaMAX (Invitrogen) containing 15%FBS, 100 U/mL penicillin, and 100 μ g/mL streptomycin. For differentiation, confluent preadipocytes were treated with medium containing 15% FBS, 0.5 mM isobutylmethylxanthine (I7018; Sigma), 1 μ M dexamethasone (D4902; Sigma), 2 μ g/mL insulin (I0546; Sigma), and 1 μ M rosiglitazone (R2408; Sigma) for 48 h. Adipocytes were then maintained in medium containing 15% FBS and 2 μ g/mL insulin. After 6–7 days of induction, differentiated cells were challenged with 5 or 20 μ M Yoda1 (SML1558, Sigma), 1 μ M TAK-242 (5.08336, Millipore) and 5 μ M GsMTx-4 (STG-100, Alomone) or vehicle for 18–20 h. Cells were collected for further analysis.

Tissue Harvest and Histology

Mice were fasted for 5 h before sacrifice. Tissues were carefully dissected to avoid contamination from surrounding tissue. Samples for RNA and protein analysis were frozen immediately in liquid nitrogen and stored at –80°C for further studies. For histology, fat and liver samples were fixed in 10% buffered formalin and embedded in the paraffin wax. A Leica AutoStainer XL (Leica Biosystems Inc., Buffalo Grove, IL) was used for automated hematoxylin and eosin staining (H&E). The histology of adipose and liver tissue ($n = 3–5$ per group) was examined in 4–6 μ m thick H&E stained sections. Adipocyte size was

measured using NIH ImageJ software (23). A total of 243–478 adipocytes per mouse were measured. Briefly, the adipose tissue imaging was calibrated for background subtraction. The excess noise was removed and the threshold was set for defining areas consisting of membrane material and empty space identified by black and white, respectively. To help define the membranes of the individual adipocytes, each membrane is uniformly enhanced. The adipocyte area was then measured using the “Measure and Label Macro” for imageJ.

Western Blot

Tissues or cells were lysed in radioimmunoprecipitation assay (RIPA) buffer containing 1mM NaF, 1mM sodium orthovanadate, and 1mM phenylmethylsulfonyl fluoride. An equal amount of total protein (25 μg/lane for cells, 50 μg/lane for tissues) was loaded into 3–8% NuPAGE Tris-acetate gels (Invitrogen) and transferred to poly-vinylidene fluoride membranes for detection with indicated antibodies. The following antibodies were used: rabbit anti-RFP (600-401-379,

Rockland) at 1:1,000 dilution; mouse anti-GADPH (60004-1-Ig, Proteintech) at 1:5,000 dilution.

RNA Extraction and Quantitative PCR

Total RNA was extracted using the NucleoSpin® RNA kit (740955, Macherey-Nagel). cDNA was synthesized using the SuperScript III first-strand synthesis supermix (11752, Invitrogen) for quantitative RT-PCR and used in real-time PCRs with Power SYBR Green PCR master mix (Applied Biosystems) on a 7900HT real-time PCR system (Applied Biosystems). The relative gene expression levels were calculated by the 2Ct method using Tata-binding protein (Tbp) or 36b4 as an internal control. Primer sequences are shown in Supplemental Table 1.

Statistical Analysis

Data are presented as mean ± SEM and were analyzed by unpaired two-tailed Student *t*-test, repeated-measures ANOVA or one-way ANOVA, as appropriate. Statistical significance was assumed at *p* < 0.05.

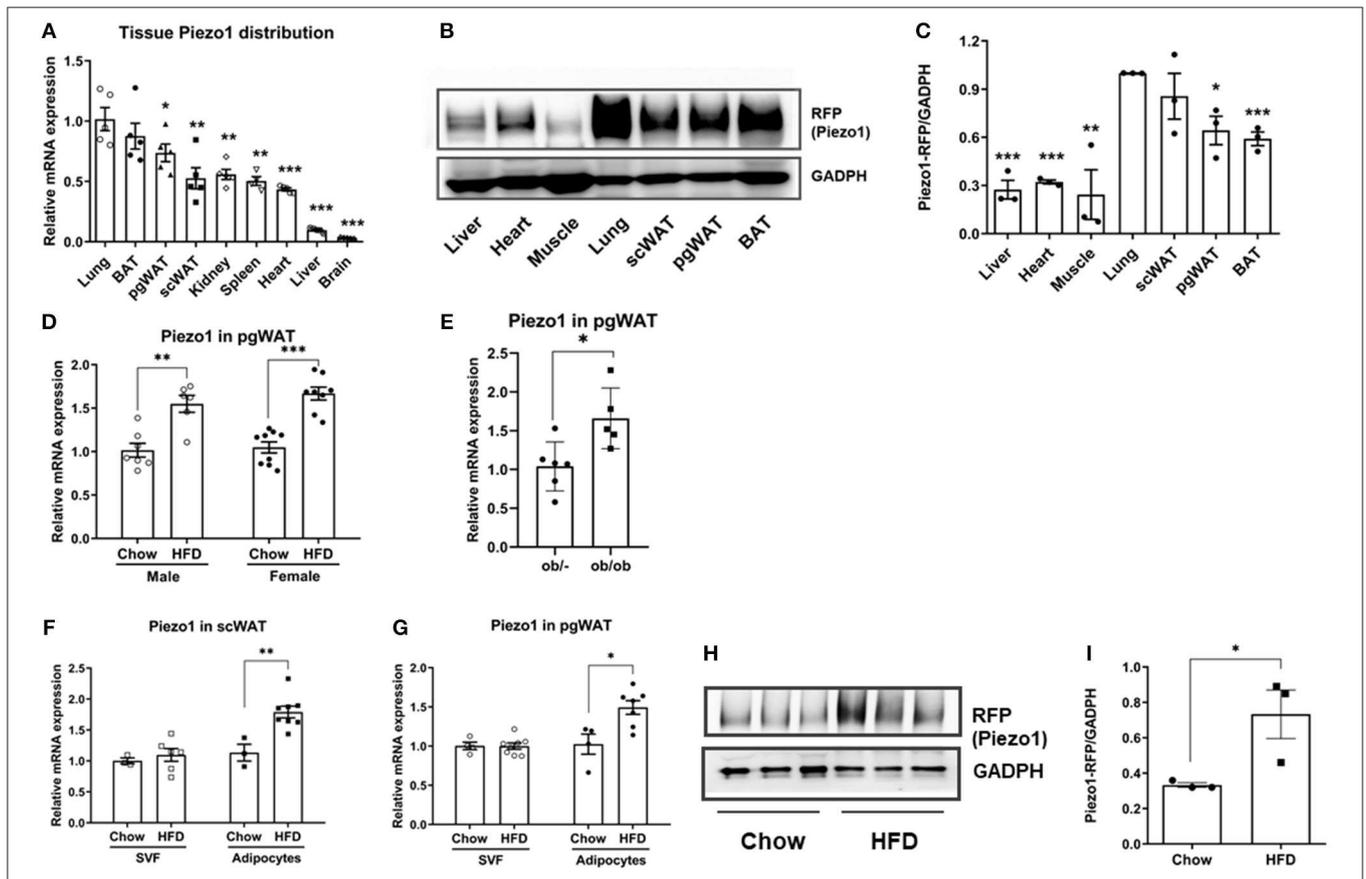


FIGURE 1 | Piezo1 expression in tissues, isolated adipocytes and SVFs. **(A)** Tissue distribution of Piezo1 mRNA levels normalized to lung in chow-fed male mice (*n* = 5). **(B,C)** Piezo1-RFP protein expression in male mice fed a chow diet. Relative protein levels were normalized to GADPH. (*n* = 3, lung was set as 1). **(D)** Piezo1 mRNA expression in pgWAT of chow and HFD-fed mice (*n* = 6–9), all values were normalized to chow-diet fed mice. **(E)** Piezo1 mRNA in pgWAT of *ob/ob* and control mice (*n* = 5–6). **(F,G)** Piezo1 mRNA in adipocytes and SVF isolated from pgWAT and scWAT of chow or HFD-fed male mice (*n* = 3–8). All values were normalized to Chow-SVF. **(H,I)** Piezo1 protein expression in adipocytes isolated from pgWAT of chow and HFD-fed male mice. Relative protein levels were normalized to GADPH (*n* = 3). **P* < 0.05, ***P* < 0.01, ****P* < 0.001. mRNA expression was corrected to Tbp. Data are expressed as mean ± SEM. SVF, stromal vascular fraction; pgWAT, perigonadal white adipose tissue; scWAT, subcutaneous white adipose tissue; BAT, brown adipose tissue; HFD, high fat diet; Tbp, TATA-Box Binding Protein.

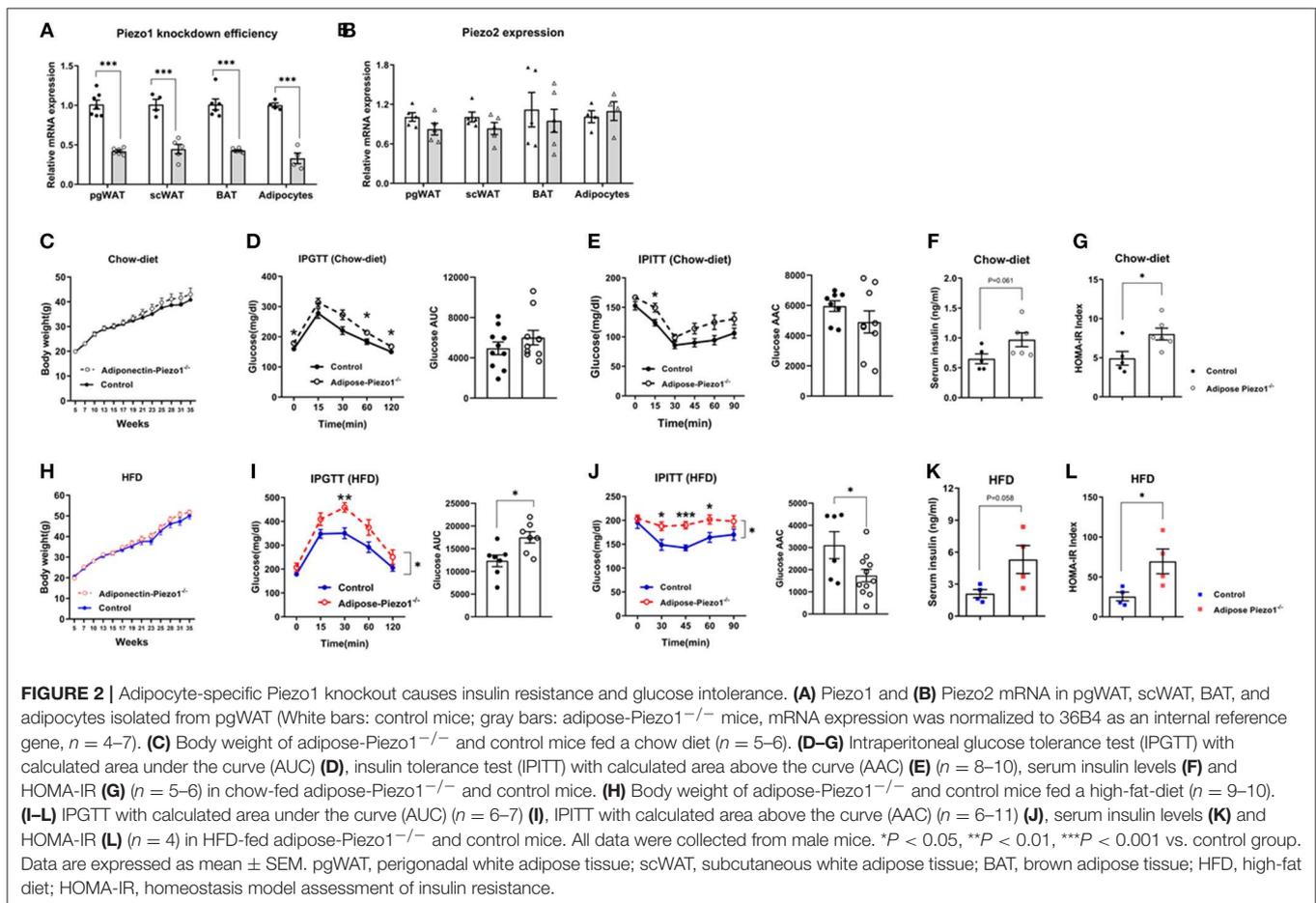
RESULTS

Piezo1 Is Highly Expressed in Adipocytes

To investigate the roles of Piezo1 in regulating adipose function, we first measured Piezo1 expression. Real-time qPCR showed that mouse Piezo1 mRNA levels were higher in brown adipose tissue (BAT), perigonadal white adipose tissue (pgWAT) and subcutaneous white adipose tissue (scWAT) than that in other organs except for lung (Figure 1A). Consistently, database searches revealed that Piezo1 mRNA in adipose tissue appeared to be the highest among 37 human tissues in the Human Protein Atlas RNA-seq data analysis (Supplementary Figure 1). Similarly, large-scale analyses of the mouse transcriptome (GSE9954) also showed that Piezo1 was highly expressed in adipose tissue (Supplementary Figure 2). Since there was no reliable Piezo1 antibody to detect Piezo1 protein, we measured the Piezo1-tdTomato fusion protein controlled by the endogenous Piezo1 promoter in the Piezo1-tdTomato transgenic mice using an RFP antibody (24). Consistent with the mRNA expression, Piezo1 protein was most abundant in lung, followed by scWAT, pgWAT, and BAT when normalized to GAPDH (Figures 1B,C). Western blot showed relatively

diffuse bands which may be caused by Piezo1 posttranslational modifications (25–27).

We then investigated whether adipose Piezo1 would be regulated in obesity. Piezo1 mRNA levels in pgWAT were elevated in both male and female HFD-fed mice and in the morbidly obese ob/ob mice compared with controls (Figures 1D,E). We further separated adipocytes from SVF in adipose tissue and measured Piezo1 mRNA levels. Piezo1 expression was increased in adipocytes but not in SVFs isolated from pgWAT and scWAT of HFD-fed obese mice compared with that from chow-fed mice (Figures 1F,G). Consistently, when the Piezo1-tdTomato reporter mice were placed on a HFD, Piezo1-tdTomato protein in adipocytes was also elevated (Figures 1H,I). We also measured the expression of Piezo2, a homolog of Piezo1. Piezo2 mRNA was very abundant in lung, but the expression was much lower in other organs including adipose tissue (Supplementary Figure 3A). Although direct comparison was not possible, Piezo2 expression was likely much lower than Piezo1 in adipose tissue because the cycle threshold (Ct) values of Piezo2 were about 5 cycles higher than that of Piezo1 using three different sets of primers in the qPCR assays (not shown). Interestingly, adipose Piezo2 was also upregulated in obesity



(Supplementary Figures 3B–D). These results show that Piezo1 is highly expressed in adipocytes and the expression is elevated in obesity.

Adipose-Specific Piezo1 Knockout Mice Are Insulin Resistant

We generated adipose-specific Piezo1 knockout mice (adipose-Piezo1^{-/-}) by crossing Piezo1-flox/flox mice to adiponectin-Cre mice (28, 29). Piezo1 expression was reduced by approximately 60% in adipose tissue and by about 70% in isolated adipocytes from adipose-Piezo1^{-/-} compared with that from control mice (Figure 2A). The expression of Piezo2 was not altered by Piezo1 knockout (Figure 2B). Body weight (Figures 2C,H), body weight gain (Supplementary Figure 4A), and body composition (Supplementary Figure 4B) were not different between adipose-Piezo1^{-/-} and control mice under either chow-fed

or HFD-challenged conditions. Chow-fed adipose-Piezo1^{-/-} mice showed mildly elevated glucose levels in the intraperitoneal glucose tolerance test (GTT) and insulin tolerance test (ITT), but the areas under (GTT), or above (ITT) curves were not different (Figures 2D,E). The fold changes corrected to the basal glucose in GTT and ITT were comparable between chow-fed adipose-Piezo1^{-/-} mice and controls (Supplementary Figures 5A,B). Nevertheless, the chow-fed adipose-Piezo1^{-/-} mice may be mildly insulin resistant since insulin levels tended to be higher and HOMA-IR index was elevated (Figures 2F,G). When challenged with a HFD, adipose-Piezo1^{-/-} mice showed exacerbated insulin resistance as evidenced by significantly increased glucose levels in GTT and ITT, and elevated insulin and HOMA-IR index (Figures 2H–L). These data indicate that adipose Piezo1 is required for maintaining systemic glucose homeostasis and insulin sensitivity.

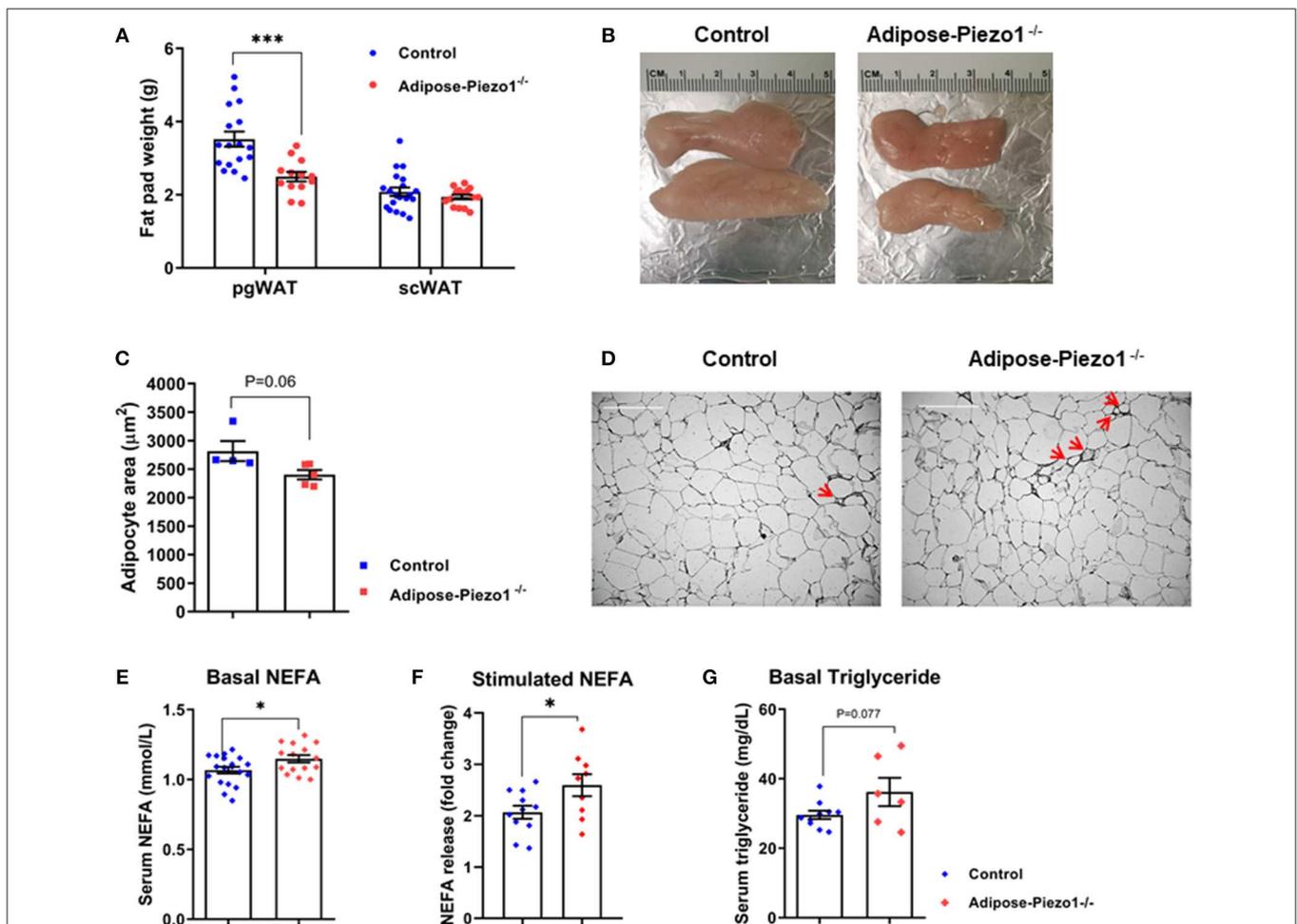


FIGURE 3 | Effects of adipose Piezo1 knockout on adipocyte size and inflammation. **(A)** Fat pad weight of pgWAT and scWAT of HFD-fed adipose-Piezo1^{-/-} and control mice ($n = 16$ – 20). **(B)** Representative pgWAT pads. **(C)** Adipocyte area of pgWAT ($n = 5$). **(D)** H&E staining of pgWAT. Arrow: crown-like structures. Scale Bar, $200\ \mu\text{m}$. **(E)** Serum NEFA after 5 h food removal ($n = 15$ – 19). **(F)** Serum NEFA elevation after treatment of beta3 adrenergic agonist CL 316,243 stimulation for 20 min ($n = 9$ – 11). **(G)** Serum triglyceride levels in HFD-fed adipose-Piezo1^{-/-} and control mice ($n = 6$ – 10). All data were collected from male mice. * $P < 0.05$, *** $P < 0.001$ vs. control mice. Data are expressed as mean \pm SEM. SVF, stromal vascular fraction; pgWAT, perigonadal white adipose tissue; scWAT, subcutaneous white adipose tissue; BAT, brown adipose tissue; HFD, high-fat diet; NEFA, non-esterified fatty acids.

Adipose Piezo1 Knockdown Reduces Perigonadal Fat Mass

Despite no difference in body weight (**Figure 2H**), pgWAT mass was reduced in HFD-fed adipose-Piezo1^{-/-} mice compared with controls (**Figures 3A,B**). scWAT mass was not altered (**Figure 3A**). The pgWAT adipocyte size tended to be decreased (**Figures 3C,D**). The basal and beta3 adrenergic agonist-stimulated lipolysis were enhanced in adipose-Piezo1^{-/-} mice (**Figures 3E,F**). Furthermore, serum triglyceride levels also tended to be higher in adipose-Piezo1^{-/-} mice (**Figure 3G**). Therefore, Piezo1 appears to be necessary for adipocyte plasticity and fat storage in pgWAT of obese mice.

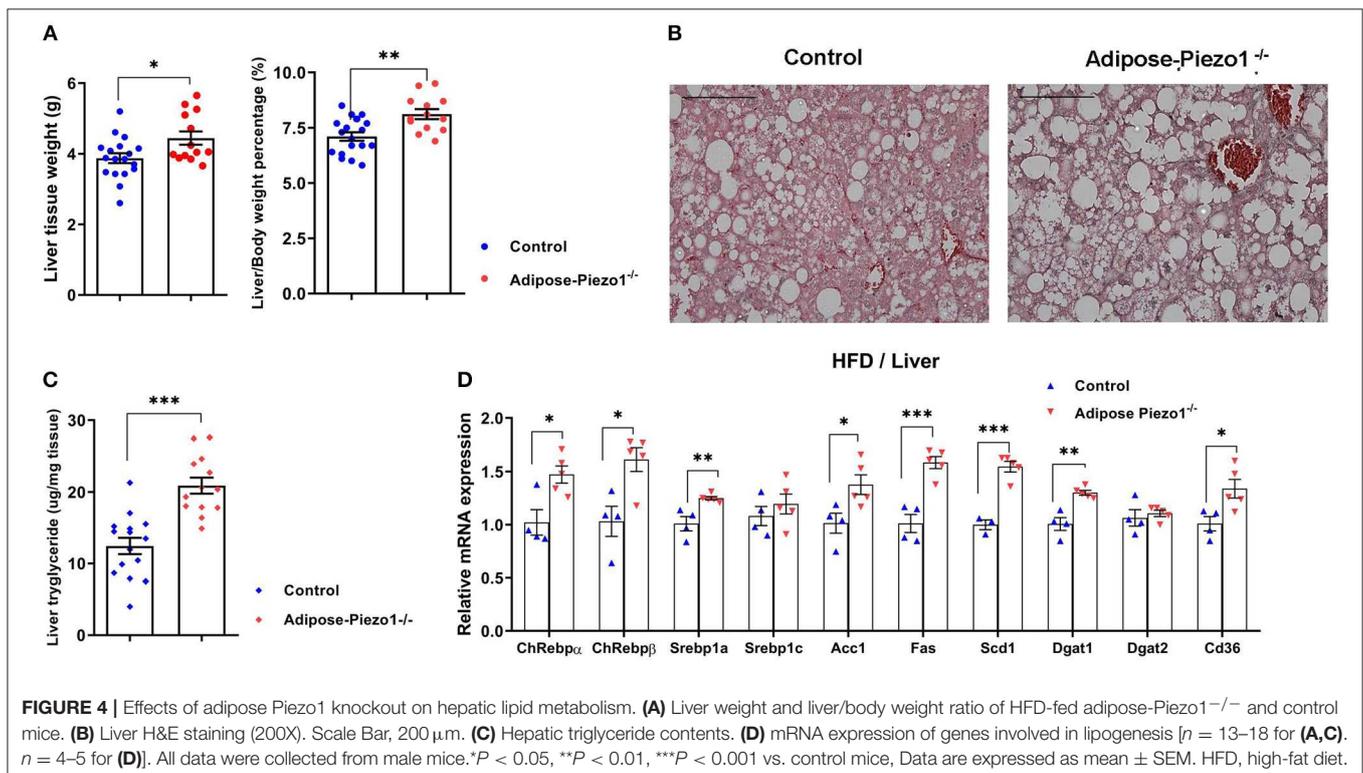
Adipose-Specific Piezo1 Knockout Mice Develop Fatty Liver

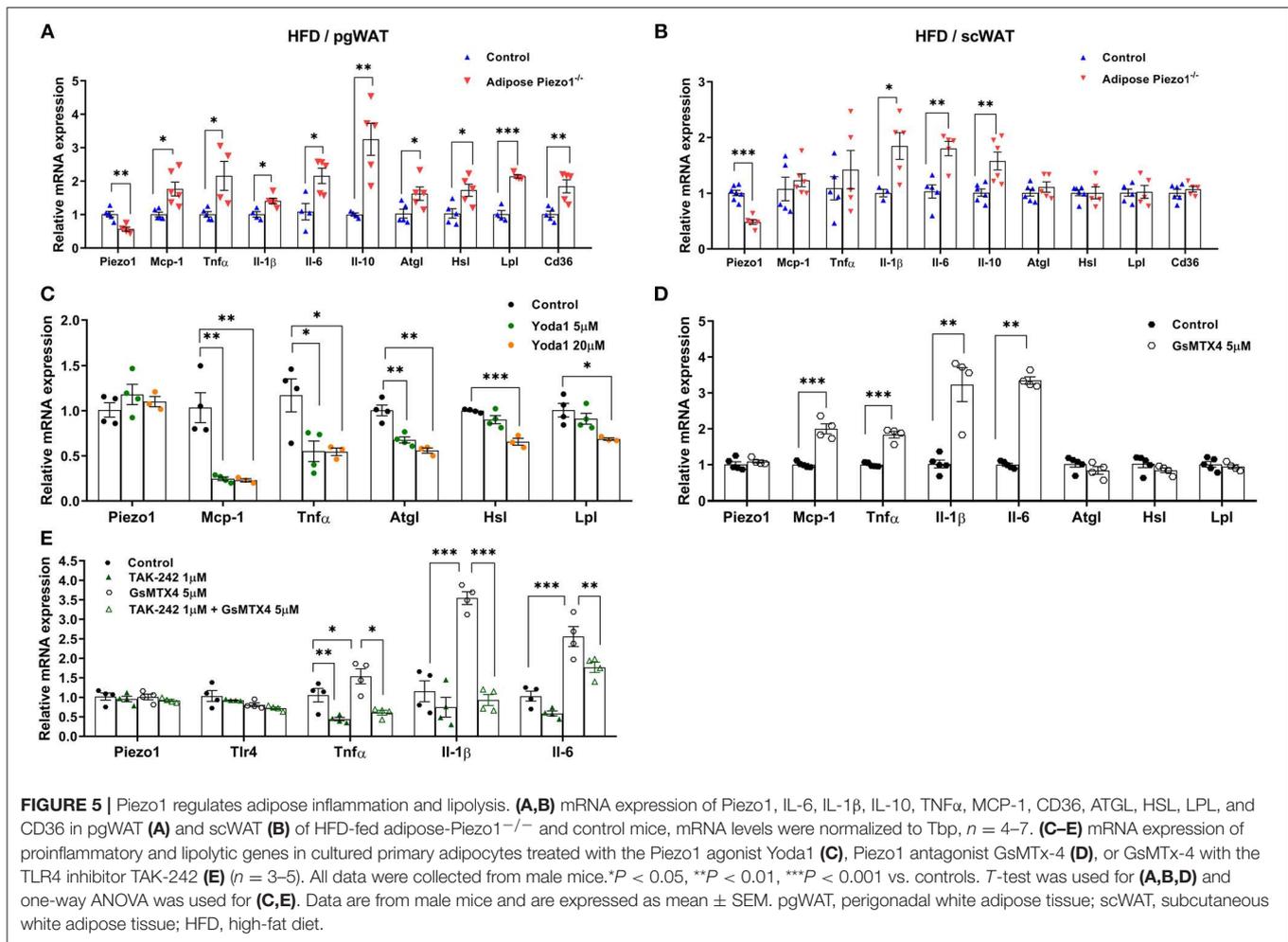
Interestingly, adipose Piezo1 knockdown in mice fed a HFD increased liver weight (**Figure 4A**), likely due to the development of fatty liver (**Figure 4B**). Liver triglyceride contents were elevated in HFD-fed adipose-Piezo1^{-/-} mice (**Figure 4C**). Consistently, hepatic lipogenic genes including carbohydrate-response element-binding protein alpha (ChREBP α) and beta (ChREBP β), sterol regulatory element binding protein-1a (SREBP-1a), acetyl-CoA carboxylase1 (Acc1), fatty acid synthase (Fas), stearoyl-CoA desaturase1 (Scd1), and diacylglycerol acyltransferase 1 (Dgat-1) were increased in adipose-Piezo1^{-/-} mice fed a HFD. In addition, the fatty acid transporter Cd36 was also elevated (**Figure 4D**). These data show that adipose Piezo1 knockdown causes lipid deposition in liver in obesity.

Adipose Piezo1 Inhibition Induces Inflammation and Lipolysis Genes

The reduced pgWAT mass and fatty liver in adipose-Piezo1^{-/-} mice may represent mild lipodystrophy (30, 31). We measured the expression of key genes involved in the lipodystrophic syndrome. Plin1 and Cidec were mildly reduced while others were not altered in adipose tissue of adipose-Piezo1^{-/-} mice (**Supplementary Figure 6A**). The second possibility is that the larger adipocytes in adipose-Piezo1^{-/-} might undergo apoptosis. However, apoptotic markers were not changed in adipose tissue with Piezo1 knockdown (**Supplementary Figure 6B**). Furthermore, fatty acid transporter Cd36 was increased rather than decreased in pgWAT of adipose-Piezo1^{-/-} mice (**Figure 5A**). These results suggest that lipodystrophy, apoptosis and reduced fatty acid import are unlikely the cause of reduced pgWAT mass in the adipose-Piezo1^{-/-} mice.

Adipose plasticity has been linked to inflammation and lipolysis. Genes that regulate lipolysis including adipose tissue triglyceride lipase (Atgl), hormone-sensitive lipase (Hsl), and lipoprotein lipase (Lpl) were increased in the highly lipolytic pgWAT, but not in the less lipolytic scWAT of HFD-fed adipose-Piezo1^{-/-} mice (**Figures 5A,B**). Furthermore, pro-inflammatory cytokines including monocyte chemoattractant protein-1 (Mcp-1), tumor necrosis factor- α (Tnf α), interleukin-1 β (Il-1 β), and Il-6 were elevated in scWAT and pgWAT of the HFD-fed adipose-Piezo1^{-/-} mice (**Figures 5A,B**). Il-10, which suppresses the inflammatory response, was also induced (**Figures 5A,B**). pgWAT of the HFD-fed adipose-Piezo1^{-/-} mice appeared to contain more crown-like structures





(Figure 3D). Although adipose inflammation is linked to fibrosis (32, 33), the expression of collagens and matrix metalloproteinases was not altered in adipose tissue with Piezo1 knockdown (Supplementary Figure 6C). The expression of the pro-inflammatory and lipolytic genes was not altered in the chow-fed adipose-Piezo1^{-/-} mice (not shown). The results suggest that Piezo1 is necessary for restraining pro-inflammatory response and lipolysis in obesity.

To investigate whether Piezo1 regulates inflammation directly in adipocytes, we activated or inhibited Piezo1 activity in cultured adipocytes. Treating adipocytes with the Piezo1 agonist Yoda1 reduced the expression of Mcp-1 and Tnf α . Lipolysis gene Atgl, Hsl, and Lpl were also mildly inhibited (Figure 5C). Conversely, treatment of adipocytes with the Piezo1 inhibitor GsMTx4 (34) elevated the expression of Mcp-1, Tnf α , Il-1 β , and Il-6, but did not alter lipolytic genes (Figure 5D). TLR (Toll-like receptor) 4 signal pathway plays a major role in adipose pro-inflammatory cytokine expression. Treating adipocytes with TAK-242, a specific TLR4 inhibitor, abolished GsMTx4-induced Tnf α , Il-1 β , and Il-6 expression (Figure 5E). These findings suggest that Piezo1 regulates TLR4-mediated cytokine expression in adipocytes.

DISCUSSION

Piezo1 is a transmembrane mechanically gated ion channel that is highly expressed in tissues with high mechanical pressure such as lungs, adipocytes, and blood vessels (14, 35). Adipocytes are under constant mechanical pressure from stored triglycerides inside and from extracellular matrix outside (36, 37). It is therefore not surprising that Piezo1 is highly expressed in adipocytes. Importantly, Piezo1 is upregulated in isolated adipocytes but not in the SVF of adipose tissue of obese mice, providing the initial evidence that Piezo1 may regulate adipocyte biological function.

The insulin resistance in the HFD-fed adipose-Piezo1^{-/-} mice indicates that the upregulation of adipose Piezo1 in obesity is likely adaptive. Piezo1 knockdown or inhibition enhances the expression of pro-inflammatory genes in adipocytes. This appears to be mediated by TLR4 since TLR4 inhibition abolishes pro-inflammatory cytokine expression induced by Piezo1 inhibition. TLR4 activation in adipocytes causes chronic inflammation and systemic insulin resistance (3, 38). However, how Piezo1 suppresses TLR4-mediated pro-inflammatory pathway in adipocytes remains elusive. Activation of Piezo1

allows nonselective membrane permeability of cations such as monovalent Na^+ , K^+ , and divalent Ca^{2+} and Mg^{2+} (39, 40). Piezo1-mediated calcium influx has been implicated in Piezo1 biological functions including regulation of red blood cell volume, apoptosis, myotube formation, and cell lineage choice (15, 18, 41–43). However, intracellular calcium is required for LPS-induced TLR4 activation at least in macrophages (44). Further studies are needed to investigate whether other cations such as Mg^{2+} , which has been shown to inhibit TLR-stimulated TNF- α and IL6 production in monocytes (45), are involved in Piezo1-mediated inflammation suppression.

The smaller pgWAT mass and fatty liver in HFD-fed adipose-Piezo1^{-/-} mice suggest that Piezo1 plays an important role in maintaining adipose plasticity to prevent ectopic lipid deposition in liver. Our data show that the reduced adipocyte size in adipose-Piezo1^{-/-} mice is less likely caused by decreased fatty acid uptake, increased apoptosis or altered lipodystrophic genes. Rather adipose inflammation in adipose-Piezo1^{-/-} mice may enhance lipolysis and reduce adipocyte size (46, 47). This may also explain the increased hepatic lipogenesis and fatty liver in adipose-Piezo1^{-/-} mice (48, 49). Interestingly, the phenotype of smaller adipocytes and insulin resistance in adipose-Piezo1^{-/-} mice is similar to that in the volume-sensitive ion channel Swell1 knockout mice (50), although the underlying mechanisms may be different. Swell1 interacts with growth factor receptor-bound 2 (GRB2) and regulates insulin signaling. Our attempts to immunoprecipitate Piezo1-tdTomato to investigate its potential interaction with TLR4 or other proteins was unsuccessful (not shown), likely because Piezo1 is a 286 kDa membrane protein with 30–40 transmembrane domains (51, 52).

In summary, our studies add Piezo1 to a growing list of ion channels that are implicated in regulating insulin sensitivity, glucose metabolism and energy expenditure in obesity (53). We found that Piezo1 is highly expressed in adipocytes and the expression is regulated in obesity. Importantly, Piezo1 inhibition in adipocytes promotes TLR4-mediated inflammation, decreases adipocyte size, and induces insulin resistance. These results

provide the first evidence that Piezo1, a novel mechanical sensitive ion channel, may play an important role in regulating adipose plasticity and insulin resistance.

DATA AVAILABILITY

This manuscript contains previously unpublished data. The name of the repository and accession number are not available.

ETHICS STATEMENT

This study was carried out in accordance with the recommendations of the Institutional Animal Care and Use Committee of the University of California, Irvine. The protocol was approved by the Institutional Animal Care and Use Committee of the University of California, Irvine.

AUTHOR CONTRIBUTIONS

CZ, QS, LT, YC, and JN conducted experiments and analyzed results. CZ and QY wrote the manuscript. MP, XL, and QY conceived the experimental design and supervised the studies.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fendo.2019.00373/full#supplementary-material>

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