

# Progress of Adipokines in the Female Reproductive System: A Focus on Polycystic Ovary Syndrome

Peipei Chen<sup>†</sup>, Rui Jia<sup>†</sup>, Yuanyuan Liu<sup>†</sup>, Mingya Cao, Liang Zhou and Zhiming Zhao<sup>\*</sup>

Department of Reproductive Medicine, The Second Hospital of Hebei Medical University, Shijiazhuang, China

Adipose tissue, one type of loose connective tissue in the human body, maintains the primary task of energy storage. Adipose tissue is not only an energy reservoir but also plays a vital role as the largest endocrine organ of the whole body *via* releasing a variety of adipokines, which participate in many pathophysiological processes, such as energy metabolism regulation, glucose and lipid metabolism, and inflammation. Polycystic ovary syndrome (PCOS) is a disorder that mainly involves the female reproductive system, affecting women of childbearing age particularly. Insulin resistance (IR) and hyperandrogenemia (HA) have been implicated as a critical link involving the etiology and outcome of PCOS. A great deal of studies has bridged the gap between adipokines (such as Adiponectin, Chemerin, Metrnl, Apelin, Resistin, Visfatin, Leptin, Vaspin, Lipocalin 2, and Omentin) and reproductive fitness. In this review, we will focus on the adipokines' functions on PCOS and come up with some points of view on the basis of current research.

## **OPEN ACCESS**

#### Edited by:

Vikas Kumar Roy, Mizoram University, India

### Reviewed by:

Ioannis Kyrou, Coventry University, United Kingdom Malini Laloraya, Rajiv Gandhi Centre for Biotechnology, India

#### \*Correspondence:

Zhiming Zhao doctor\_zhaozhao@sina.com orcid.org/0000000228443503

<sup>†</sup>These authors have contributed equally to this work

#### Specialty section:

This article was submitted to Reproduction, a section of the journal Frontiers in Endocrinology

Received: 22 February 2022 Accepted: 19 April 2022 Published: 26 May 2022

#### Citation:

Chen P, Jia R, Liu Y, Cao M, Zhou L and Zhao Z (2022) Progress of Adipokines in the Female Reproductive System: A Focus on Polycystic Ovary Syndrome. Front. Endocrinol. 13:881684. doi: 10.3389/fendo.2022.881684 Keywords: PCOS, adipokines, female reproductive system, reproductive endocrinology, insulin resistance

# INTRODUCTION

Polycystic ovary syndrome (PCOS) is characterized by heterogeneity, estimated to jeopardize 5.6% (1) nationwide and 6% - 20% (2) worldwide child-bearing women with the tendency to become an epidemic. The most commonly used criteria for PCOS come from the Rotterdam ESHRE/ASRM-Sponsored PCOS Consensus Workshop Group (3): oligo- or anovulation, clinical and/or biochemical signs of hyperandrogenism, and polycystic ovaries after exclusion of other etiologies (congenital adrenal hyperplasia, androgen-secreting tumors, or Cushing's syndrome). There has been a gradually increasing awareness that PCOS is likely to be a complicated polygenic disease, affected by external factors like lifestyle and dietary habit. Clinical manifestations of PCOS center on endocrine metabolism and involve reproduction, endocrine, substance metabolism, and psychological health although with an unclear etiology. Endocrine changes in PCOS inflict on other female systems rather than reproduction alone. With further study on adipokines, a variety of bioactive compounds released by adipose tissue have been revealed, and the signal crosstalk between adipokines and PCOS has been gradually understood. Through the paracrine and endocrine pathways, adipokines act widely on different parts of the human body, regulating and controlling glucose and fatty acid metabolism, energy expenditure, inflammatory response, cardiovascular function, reproduction, and other biological processes over the entire body or locally (4, 5). A meta-

Adipokines Associated With PCOS

analysis published in *Lancet* in 2021 consisting of 71 studies (including 2495 patients with PCOS and 2520 controls) revealed that non-obese PCOS patients have significantly higher circulating levels of Chemerin, Leptin, Resistin, and Visfatin but a significantly lower circulating level of Adiponectin when compared with non-obese healthy controls (6). A vast and expanding universe of research into the crosstalk of adipokines and PCOS has been issued, emphasizing the role of adipokines in the development of PCOS: metabolic disturbance like hyperinsulinemia, IR, HA, ovary dysfunction like granulosa cell apoptosis, and abnormal endometrial receptivity accompanied with adverse pregnancy outcomes during assisted reproductive technology (ART).

## METHODS

All documents were obtained from a systematic search of articles reported in the PubMed. The key words used were: "PCOS", "adipokines", "Adiponectin", "Chemerin", "Metrnl", "Apelin", "Resistin", "Visfatin", "Leptin", "Vaspin", "Lipocalin 2", "Omentin", "genetic variants", and "polymorphisms". Original articles, meta-analyses, and review articles in English were included in this review. Topics about adipokines, their genetic polymorphisms, biological function, and role in the physiopathology of PCOS and their relationship with the outcomes of ART of PCOS were addressed in this review.

## Adiponectin

As a collagen-like 30-kDa protein, Adiponectin (also known as acrp30, apM1, adipo-Q, or GBP28) is the most abundantly secreted adipokine by the adipose tissue, mainly the white adipose tissue, to travel through the blood (7). It is clearly a distinct member of a protein family characterized by a collagenous helical structure at the NH2 terminus and a globular domain at the COOH terminus (8). The Adiponectin gene in humans is located in the long arm of chromosome 3 (3q27), containing three exons and two introns (9). The study by Yamauchi and colleagues (10), which has discovered Adiponectin receptors 1 and 2 (AdipoR1 and AdipoR2), was a Big Bang. From then on, a vast and expanding universe of research on Adiponectin has been issued. Secreted by adipocyte as a metabolic messenger, Adiponectin targets human organs and tissues including the reproductive system by binding to its receptors (11) and participating in various physiological and pathological processes as an initiating factor or intermediary. Adiponectin is involved in the pathophysiological processes of many diseases, inhibiting tumor cell growth and metastasis through anti-proliferation and induction of apoptosis (12, 13). It has been used in the prevention and treatment of type 2 diabetes mellitus (T2DM) and atherosclerosis via anti-inflammation and improvement of insulin sensitivity (14, 15) as well as arthritis (16).

Adiponectin and its receptors (AdipoR1 and AdipoR2) have been found in human ovarian cells (oocytes, granulosa cells, follicular membrane cells, and cumulus cells) and human follicular fluid at various stages of follicular development (17), combining with each other to regulate follicular development and ovulation through increasing the gonadotropin (Gn) or insulin sensitivity directly or indirectly (18). For one thing, Adiponectin can relive the ovulatory event of porcine in vitro by directly acting on the porcine granulosa cells, accompanied by continuous and rapid induction of cyclooxygenase-2 (COX-2) and its downstream substances along with the activation of synthase (PGE synthase) in cytoplasm (19). For another, Adiponectin decreased the production of progesterone and androstenedione in insulin-induced bovine follicular theca cells (TCs) through inhibition of cytochrome P450 17A1 (CYP17A1) and cytochrome P45011A1 (CYP11A1) (20), leading to reduced steroidogenesis. As is known to all, the mid-menstrual (LH) peak induces the activation of a series of signaling pathways associated with ovulation, leading to rupture of the follicle and release of the oocyte. Given all this, Adiponectin plays a significant role in ovulation by regulating the steroid production of granulosa cells (GCs) and follicular TCs, expansion of cumulus cells, and an interaction with luteinizing hormone (LH) even though the specific mechanisms need further investigation.

In addition to being expressed in the ovaries, Adiponectin has also been found to be expressed in the hypothalamus and pituitary gland. Short-term treatment with recombinant Adiponectin can inhibit gonadotropin-releasing hormone (GnRH) secretion by activating AMP-activated protein kinase (AMPK), thereby reducing LH secretion (21). Correspondingly, Adiponectin production can be induced in human ovaries after treatment with recombinant LH (22). A hypothesis is thus suggested that there may be a regulatory mechanism of Adiponectin, similar to that of the sex hormone axis *in vivo*: GnRH and Gn induce local Adiponectin production in ovaries while circulating high levels of Adiponectin in turn inhibit GnRH and LH production in the hypothalamus and pituitary gland through negative feedback.

The latest meta-analysis shows that there is a significantly lower circulating Adiponectin level in non-obese PCOS patients (6), and the serum Adiponectin level is negatively correlated with the insulin and IR index after adjusting for body mass index (BMI) confounders (23). In addition, Adiponectin levels were lower in the non-ovulating PCOS patients than in the regular ovulating PCOS patients (24), indicating its important role in ovulation disorders of PCOS. Combined with previous animal studies which revealed Adiponectin inhibited LH and androgen secretion but promoted ovulation, it is speculated that the low Adiponectin level in PCOS may negatively increase LH secretion in the pituitary gland, resulting in a high LH and androgen level and ovulation dysfunction in PCOS patients. Abnormal hormone exposure in early pregnancy can affect metabolic status in adulthood. Most women with PCOS have unexplained high androgen levels during pregnancy (25, 26), and their offspring will exhibit HA and metabolic disorders which are centered on IR in adulthood. Animal studies have shown that embryonic Adiponectin supplementation improves the metabolic syndrome in adult PCOS offspring by activating the PI3K-Akt pathway (27). In PCOS patients treated with metformin, rosiglitazone, and pioglitazone, the serum Adiponectin levels increased, accompanied by decreased

insulin secretion, enhanced insulin action, and increased lipid oxidation (28), suggesting that Adiponectin is a beneficial factor for human body and is involved in regulating the function of the female reproductive system.

Although obese PCOS patients have a certain natural pregnancy rate after losing weight, most PCOS patients with infertility need assisted reproduction. Unfortunately, there is relatively little research in this area. Inal et al. (29) found that although PCOS patients had a low serum Adiponectin level, there was no significant difference in the embryo development quality or clinical pregnancy rate between the PCOS and non-PCOS patients during ART. This may be related to the improvement of HA and IR that benefits from physical exercise and improved diet during pregnancy medication. Although a certain relationship between the serum Adiponectin level and pregnancy outcome in PCOS patients has not been revealed, the close relationship between the Adiponectin level and the characteristic pathological changes of PCOS has been confirmed. Therefore, the specific mechanism of Adiponectin in the development of PCOS is worth studying because it has a broad prospect.

Regarding the effect of Adiponectin gene polymorphisms on PCOS, current studies showed a close correlation with race. The rs1501299 single nucleotide polymorphism (SNP) of Adiponectin was significantly correlated with PCOS risk in East Asians (30, 31), the rs17300539 showed a correlation with PCOS risk in a Chinese population (32), and rs1501299 and rs2241766 polymorphisms both had a close correlation with PCOS risk in Caucasians (33). In addition, several studies have shown the association of genetic polymorphisms of Adiponectin with endocrine metabolism and steroid hormone production in PCOS patients (34, 35). All these studies demonstrated a relationship between PCOS and Adiponectin at the genetic level.

## Chemerin

Chemerin was first discovered in 1997 while studying psoriasis and was named tazarotin-induced gene 2 (TIG2) because it could be induced by TIG2 under certain conditions (36). In 2003, the ligand was identified as the orphan receptor ChemR23 and formally named Chemerin, which turned out to be an adipokine (37-39). Located on human chromosome 7, the Chemerin gene consists of five coding exons. It is transcribed primarily by the white adipose tissue and binds to three receptors: chemokine-like receptor 1 (CMKLR1), G protein-coupled receptor 1(GPR1), and CCRL2 (e.g., 2 CC motif chemokine receptor), though binding G proteins to perform their physiological functions (40-44). Chemerin has been found to play a key role in metabolic diseases, such as obesity and T2DM. It also participates in diseases of multiple systems, such as autoimmune diseases (45), urinary diseases (46), digestive diseases (47), and psychiatric diseases (48), by regulating blood pressure, inflammation, immune response, fat cell differentiation, and carbohydrate metabolism.

Chemerin is expressed in basal cells of the human uterus, stromal cells, and extravillus trophoblast cells of pregnant women and the human placenta (49–51). Furthermore, Chemerin and CMKLR1exist and play an active role in human GCs (42). It is an indisputable fact that local IR exists in GCs of human ovary from

PCOS patients. Excessive insulin in serum can promote Chemerin production (52), and PCOS patients have elevated levels of Chemerin which are associated with IR, as compared with non-PCOS patients. Follicular fluid and luteinized granulosa cells of PCOS patients with IR have higher Chemerin levels, which can be induced by over-expressed insulin and in turn damage the glucose uptake capacity through affecting Insulin Receptor Substrate 1/2 Irs1/2, Akt phosphorylation, and GLUT4 translocation, finally aggravating IR (53). This is a vicious cycle between Chemerin and IR. A low progesterone level observed in PCOS serum indicates unexpected ART outcomes. Estienne et al. demonstrated that the over-expression of Chemerin system in human luteinized granulosa cells (hIGCs) in the case of PCOS could be a reason for progesterone secretion disturbances through CMKLR1, STAR, MAPK3/1, and PRKAA (54). They observed a significantly higher concentration of Chemerin and CMKLR1 within the follicular fluid and hIGCs at both the gene and protein levels from PCOS patients. Furthermore, incubation with the CA4910 nanobody, a biological fragment raised against CMKLR1, abolished the inhibition of Chemerin-induced STAR mRNA expression as well as progesterone secretion in human granulosa cell line KGN and primary hIGCs from PCOS patients. However, no changes were found about the CYP17A1, a key enzyme that converts cholesterol into progesterone, suggesting that Chemerin may lead to a low progesterone level in PCOS patients by inhibiting cholesterol production. Chemerin has been shown in vitro to reduce IGF-1-induced estrogen and progesterone production and cell proliferation by binding with CMKLR1, thus reducing the activation of the IGF-1R signaling pathway in primary cultured human granulosa cells (42).

As the most abundant immune cells in the ovary, macrophages play a crucial role in the inflammatory state and clearance of senescence and apoptotic cells. HA in PCOS patients leads to increased Chemerin levels in the ovaries. In rats treated with dihydrotestosterone (DHT) for 15 days, researchers demonstrated that high levels of Chemerin as a ligand for CMKLR1-expressing monocytes in the blood resulted in local ovarian inflammation, leading to granulosa cell apoptosis, follicular growth arrest, and anovulatory infertility (55). In addition, a retrospective study confirmed the association between Chemerin and ovarian polycystic changes in PCOS patients, and the results showed that serum Chemerin concentration could reflect the severity of ovarian polycystic changes (56). All these studies taken together suggest that Chemerin plays a role in follicular dysplasia in PCOS patients, but the specific mechanism behind this phenomenon needs to be further studied. The serum Chemerin level of PCOS women with HA was higher than that of women with increased androgen level alone, consistent with the change in level of free androgen index and total testosterone level. Prenatal androgen overload can cause early life changes that affect the reproductive axis and metabolic state, contributing to the development of PCOS. The female offspring of prenatal androgen hyperactive (PH) rat models exhibit two phenotypes: irregular ovulation phenotype (PHiov) and anovulation phenotype (PHanov). Both phenotypes have lower serum Chemerin protein levels (57), which suggests

that excessive androgen exposure during pregnancy may be involved in the development of PCOS by affecting the Chemerin level of female offspring.

By studying the effect of Chemerin levels on intracytoplasmic sperm injection (ICSI) results of lean patients with PCOS, Kabil Kucur et al. found that compared with the control group, the serum and follicular fluid Chemerin levels of PCOS patients were significantly higher. Among the PCOS patients under ART, patients with the successful ART had lower Chemerin levels in the serum and follicular fluid than those who failed the ART (58), indicating that Chemerin is not only involved in the pathogenesis of PCOS, but also a risk factor for the failure of the ART cycle.

Although affected by obesity status, Chemerin rs17173608 polymorphism had been shown to have a certain relationship with PCOS among Iranian women (59).

#### Metrnl

Originally reported as a homologous transcription of Meteorin with a role in the central nervous system as a neurotrophic factor (60, 61), Metrnl was identified in 2014 as a new adipocytokine in a caloric restriction model. Metrnl gene was located on human chromosome 17q25.3, encoding a secreted protein containing 266 amino acids (62). Different from the high expression of Meteorin in the nervous system (63), Metrnl is more likely to be expressed in the white adipose tissue and other tissues like heart and skeletal muscle (62, 64). Current studies have shown that Metrnl can affect the nervous system, digestive system, and cardiovascular system by inducing the browning of white fat and muscle regeneration as well as antagonizing IR and antiinflammatory as a neurotrophic factor, muscle factor, and adipokine (61, 65, 66).

Two case-control studies revealed lower serum Metrnl levels in PCOS patients than in normal controls, contrary to the level change of IR markers, FSH, and an independent correlation between Metrnl and PCOS (67, 68). A study from Iran found that serum levels of Metrnl in the infertile PCOS (PCOS-inf) and PCOS with recurrent pregnancy loss (PCOS-RPL) subgroups were significantly lower than in the control group, which remained significant after adjustment for confounding factors but disappeared when compared between the PCOS subgroups. Interestingly, after stratification by BMI, the significant difference in the serum Metrnl levels between overweight and normal-weight individuals in the PCOS-inf and PCOS-RPL groups disappeared. There was a significant rising of Metrnl levels in serum at 24 and 28 weeks of gestation in pregnant women with gestational diabetes compared with normal pregnancies, and similar results were observed in maternal peripheral blood and umbilical cord blood collected at late pregnancy. After delivery, the significant increase observed at 24-28 weeks of gestation did not exist (69), this covarying performance suggesting that Metrnl may be involved in the occurrence of gestational diabetes.

#### Leptin

Leptin, encoded by a gene on the human chromosome 7 mainly coming from the white adipose tissue, was discovered as an adipokine in 1994 (70, 71). Leptin receptor, a member of the class I cytokine receptor family, is encoded by diabetes (db) gene (72). Leptin is secreted mostly by the adipose tissue into the blood and binds mainly the Leptin receptor to activate Janus kinase signal transduction transcriptional activator signal (JAK-STAT) pathway to regulate food intake and energy homeostasis (73), participating in physiological activities such as immune response, neuroendocrine response, systemic inflammatory response, and reproductive function.

Leptin can regulate reproductive function at the central level. Studies have shown that Leptin regulates GnRH secretion through neurons expressing Kisspeptin and NO, thus playing a role in the initiation of puberty and periodic secretion of Gn even though there is no Leptin receptor in GnRH neurons (74). Kucera and colleagues (75) found that increased Leptin in follicular fluid is a sensitive marker of anovulatory fertility disorders by comparing the levels of Leptin in follicular fluid in fertility disorders with different etiologies. One of the major problems confusing PCOS patients of reproductive age is anovulatory infertility. A meta-analysis showed that the circulating Leptin level in non-obese PCOS patients was significantly higher than that in obese PCOS patients (6). Besides, the relevance between homeostatic model assessmentinsulin resistance (HOMA-IR) and serum Leptin level has also been elucidated (76). Insulin has been shown to enhance Leptin gene expression and elevate circulating Leptin levels (77). IR in PCOS patients leads to increased insulin content, which may induce the white adipose tissue to secrete more Leptin to participate in the development of PCOS. Studies have shown that Leptin plays a role in the occurrence and development of PCOS by regulating the reproductive endocrine axis and local steroid production of ovary as well as participating in IR. Circulating Leptin level is positively correlated with increased IFN- $\alpha$ , followed by apoptosis of human granulosa cells (KGN), which has been demonstrated in cell experiments (78). Obesity is another problem faced by PCOS patients, Liu et al. (79) have found that hyperandrogenism may increase feeding and lead to obesity by suppressing Leptin levels in the cerebrospinal fluid thus inhibiting Leptin signaling in the hypothalamus in a rat model.

Studies have shown that Leptin enhanced the expression of aromatase genes by activating classic signal transduction pathways MAPK and PI3K, whereas this phenomenon was significantly attenuated in granulosa cells of PCOS patients, together with significantly increased plasma Leptin and decreased soluble Leptin receptor (s OB-R) in PCOS patients (80). This may suggest that Leptin resistance in PCOS patients may be possibly mediated by Sam68, an RNA-binding protein that is widely expressed in the body and has signaling functions (81). A study aiming to illustrate the effects of aerobic exercise on PCOS rats with HA showed the effect of Leptin on inflammation in PCOS (82). In this study, the researchers divided the study subjects into normal control group with or without aerobic exercise and PCOS group with and without aerobic exercise (called NC, EC, PC, and PE group, respectively), and they were surprised to find that levels of TNF- $\alpha$ , IL-6, and Leptin were

reduced in the PE group compared to the PC group. Thus, they speculated that aerobic exercise improved the inflammatory status of PCOS by reducing Leptin resistance. Aerobic exercise reduces internal inflammation by reducing Leptin resistance (82), and high-intensity interval training (HIT) reduces circulating Leptin levels in obese women and adolescents (both men and women) with risk factors for type 2 diabetes (83, 84). But things change when it comes to the obese PCOS (85). This may be due to the interaction between Leptin and PCOS, and the formation of PCOS will regulate the expression of Leptin, making it inconsistent with physiological conditions.

Similar to Adiponectin, contribution of ethnicity to the association of gene variants of Leptin receptor with PCOS had been clarified in a case control study (86), which showed that the rs1137100 was negatively associated with PCOS in Tunisians and Bahraini women, whereas rs2025804 was associated with PCOS only in Tunisians.

# Apelin

Boucher et al. demonstrated that Apelin secreted from human adipocytes, first extracted from bovine stomachs and so named in 1998 (87), is essentially an adipokine in 2005 (88). Located on the long arm of the human X chromosome, Apelin gene expresses a precursor peptide consisting of 77 amino acid residues, which is formed by proteolytic enzymes into different protein subtypes. The main biologically active forms are Apelin-36 and Apelin13 (89), with the Apelin-13 as the most common subtype in human plasma (90). Apelin is widely distributed in central and peripheral tissues, such as pituitary, heart, lung, kidney, breast, and adipose tissues, and can be transmitted by putative receptor protein related to AT1 in these tissues, regulating a wide range of physiological activities such as energy metabolism, body fluid balance, and regulation of food intake and angiogenesis (91).

Apelin regulates human gonads by initiating signaling pathways, such as mitogen-activated protein kinase3/1 (MAPK3/1), protein kinase B(AKT), and AMPK *via* binding the Apelin receptors. Animal and cell line studies to date have revealed the role of Apelin in regulating the development of gonadal axis, ovarian angiogenesis, and follicles development (92–96). Apelin and its receptor are mainly expressed in granulosa cells, cumulus cells, and follicular membrane cells of the human ovary, but less in oocytes. In addition, the concentration of Apelin in plasma is lower than that in follicular fluid, and there is a reason to speculate that follicular Apelin is partly derived from granulosa cells to regulate granulosa cell function by paracrine and/or autocrine (97).

Bongrani et al. (98) showed that the protein and mRNA level of Apelin and its receptors in ovarian follicular fluid and granulosa cells of PCOS patients increased compared with the normal control group, with the elevated Apelin and its receptor being positively correlated with the number of preantral follicles. Insulin *in vivo* inhibits the production of IGF-1 binding protein and thus increases the biological activity of IGF-1. In PCOS patients, hyperinsulinemia caused by IR increases the activity of IGF-1 in the ovary, and IGF-1 promotes production of estrogen by stimulating Apelin. It was also reported that Apelin had a high expression in granulosa cells and follicular fluid of women with PCOS (99). Recombinant human Apelin-13 and Apelin-17 promote ovarian basal estrogen, progesterone, and insulin-like growth factor 1(IGF-1) secretion. The same study also showed that APLN increases IGF1-induced steroid production in human primary luteinized granulosa cells by increasing the hydroxysteroid dehydrogenase (HSD-3B) protein expression and activating the mapK3/1, and Akt pathways (99), implicating the effect of Apelin in the development of follicles. Vascular endothelial growth factor (VEGF) is critical for ovarian follicular angiogenesis and normal reproductive function. IGF-1 and VEGF directly or indirectly affect the ovary function of patients with PCOS in terms of cyst formation and angiogenesis by the Apelin system (100-102). Progesterone stimulated the expression of Apelin receptors in granulosa cells and hyperinsulinemia could stimulate Apelin and VEGF expression in PCOS. The expressions of insulin, Apelin, and VEGF (93, 103) were increased in this case (104), whereas Apelin receptors were significantly decreased because of the luteal phase defect (LPD) due to IR. Therefore, inefficient combination of APLN to the receptors is related to the pathogenesis of LPD in patients with PCOS. Two recent studies have shown that exercise affects Apelin levels in vivo (105, 106). So, we do not deny that some studies have reached different conclusions, possibly due to the age assessment methods for genetic characteristics of exercise populations and the heterogeneity of PCOS itself.

# Resistin

Originally found in the white adipose tissue of mice, Resistin was so named because of its insulin-resistant effect, and was induced to produce and inhibit adipocyte differentiation during adipocyte differentiation (107). Located on the chromosome 19p13.3 with a span of 1369 bp, the Resistin gene consists of three introns and four exons (108), encoding a 12.5 kDa polypeptide with 108 aa mature fragments (109). Subsequent studies have shown that human macrophages, adipose tissue, oocytes, and granulosa cells at all developmental stages and follicular membrane cells of large follicles express Resistin at the mRNA and protein levels (110, 111). Thus far, four kinds of Resistin receptors have been identified: a subtype of Adenylate cyclase-associated protein 1 (CAP1), transmembrane Toll-like receptor 4 (TLR4), subtype of decorin ( $\Delta$ DCN), and receptor tyrosine kinase-like orphan receptor 1 (ROR1), involved in the pathophysiological processes such as IR, inflammation, and apoptosis (112) through CAP1-cAMP-PKA/NF-kβ and TLR4-TIRAP/MyD88-JNK/p38 (113).

Although results of published studies are still inconclusive, the analysis by Raeisi et al. (114) revealed that, overall, the Resistin level was significantly higher in PCOS women compared with healthy controls, independent of obesity status. As a nonnegligible clinical manifestation of PCOS patients, HA may be related to the increased level of Resistin in serum (115). Further *in vitro* experiments showed that serum Resistin could directly promote formation of testosterone in follicular membrane cells of PCOS patients by enhancing the activity of 17 $\alpha$  hydroxylase (116). Moreover, the follicular membrane cells of normal developing follicles also respond to Resistin

stimulation. Therefore, the lack of correlation between Resistin and serum total testosterone in normal controls may be due to the existence of a substance inhibiting Resistin in normal ovaries.

Messini et al. (117) found that Resistin inhibited the estradiol and progesterone secretion of human corpus luteum granulosa cells stimulated by follicle stimulating hormone under normal physiological conditions. We hypothesized that a high level of Resistin in PCOS patients may enhance inhibition of estrogen progesterone secretion and thus cause the clinically observed low progesterone state in PCOS patients based on the results above. Nevertheless, the mechanism via which Resistin can affect the production of estradiol and/or progesterone in the granulosa cells is not clear. Combination of Resistin and decorin induced cell proliferation and migration, resulting in the expansion of white adipose tissue (118). Increased white adipose tissue increases the secretion of various adipokines, such as Chemerin, which may influence the occurrence and development of PCOS from another direction. It has become appreciated that PCOS encompasses chronic inflammatory state all over the body, including the ovaries, and Toll-like receptor-4 has also been shown to be expressed in bovine ovarian granulosa cells (119). Moreover, it was found that human macrophages generated Resistin and directly participated in the regulation of inflammation. When the level of serum Resistin increased, macrophages were activated to produce TNF- $\alpha$ , IL-6, IL-12, and other pro-inflammatory factors (120, 121). Regrettably, there have been very few studies on the expression of TLR4 in ovarian granulosa cells as well as the level of Resistin and local inflammation of ovary in PCOS patients. Therefore, further welldesigned studies with large sample sizes should be performed to examine the circulating levels of Resistin and its role in PCOS.

### Visfatin

First cloned from human peripheral blood lymphocyte cDNA library in 1994 as a cytokine, Visfatin was also named pre-B-cell colony enhancer factor (PBEF) (122). As a 52-kDa secreted protein, Visfatin is expressed mainly in human visceral fat, as well as bone marrow, liver, and muscle (123). In mammals, Visfatin exists in two forms, intracellular and extracellular, namely iNAMPT and eNAMPT (124), identified as an important enzyme (nicotinamide phosphoribosyl transferase) involved in metabolism (125, 126).

Studies showed that granulosa cells, cumulus cells, oocytes, and follicular membrane cells of human ovary can secret Visfatin (127). It has also been proved that the number of oocytes retrieved was increased with increasing levels of Visfatin in follicular fluid (128), further substantiating a positive impact of Visfatin on female reproduction. There was no association in Visfatin levels between plasma and follicular fluid, and circulating Visfatin concentration seems to have little effect on Visfatin concentration in follicular fluid (128), suggesting little communication between blood and follicular fluid Visfatin and an important role of the local Visfatin in ovary. Visfatin is involved in local ovarian energy metabolism as a key enzyme in nicotinamide adenine dinucleotide (NAD) biosynthesis and may thus affect follicular development. In addition, recombinant human Visfatin promotes human granulosa cell proliferation and IGF-1-dependent steroid hormone production (127). In bovines, Visfatin is involved in GC steroidogenesis, proliferation, and oocyte maturation, as well as increasing the secretion of E2 secretion accompanied by an increase in StAR and HSD-3 $\beta$  expression (129).

There are contradictory reports on the levels and expression of Visfatin in PCOS subjects. More studies tend to favor a higher level of ovarian Visfatin in PCOS (130–132). In addition to adipocytes, inflammatory cells, such as macrophages, have also been shown to secrete Visfatin, which may suggest PCOS as a chronic low-grade inflammatory condition (133). Its elucidated insulin-like effects were consistent with the high levels observed in PCOS (134). A recent study found that FK866, an inhibitor of Visfatin, improved letrozole-induced pathological state of hyperandrogen PCOS mice, inhibiting androstendione and testosterone secretion, inhibiting ovarian cyst formation and apoptosis, but promoting luteal formation, and increasing local ovarian glucose content (131). In summary, whether Visfatin plays a role in the pathogenesis of PCOS is still under discussion and needs more reliable studies.

## Vaspin

Visceral adipose tissue-derived serpin (Vaspin) is a member of the serine protease inhibitor family, encoded by the SERPINA12 gene on the long arm of human chromosome 14 (14q32.1), and it is constituted by 395 amino acids as a 47-kDa protein secreted mainly by visceral and subcutaneous adipose tissue (135). Vaspin is considered to be closely related to lipid metabolism and IR (136) through binding to a 78-kDa glucose-regulated protein (GRP78) on the cell surface (137).

Several studies have shown that the circular Vaspin level is elevated in PCOS patients and plays a role in the process of IR among PCOS patients (138-140). In 2008, the first study about Vaspin in PCOS women was published (141), showing that circulating Vaspin and the transcript as well as translation levels of Vaspin in omental adipose tissue are significantly elevated in women with PCOS. In the same study, the in vitro experiments demonstrated that glucose stimulates the secretion of Vaspin in omental adipose tissue. Also, the serum Vaspin levels in PCOS women decreased significantly after 6 months of metformin treatment. In a study of serum Vaspin levels in different types of PCOS (142), researchers found that serum Vaspin level has an independent correlation with BMI and HOMA-IR. Moreover, they observed the phenomenon that PCOS patients diagnosed with all three criteria (also named the "classical" phenotype) have the highest serum Vaspin levels, indicting the correlation between PCOS severity and Vaspin. It was shown that the Vaspin rs2236242 variant in the Iranian population was associated with metabolism in obese PCOS patients, and the A allele reduced the risk of PCOS compared to the T allele, even though the association disappeared after adjusting for BMI (143). Bongrani et al. (144) detected Vaspin for the first time in the human ovary to improve the granulosa cells function concentration-dependently, which gives us evidence that Vaspin may have a regulatory role on granulosa cells in PCOS. In addition, a prospective case control study showed that those infertile PCOS patients who had successfully induced

ovulation after clomiphene treatment had significantly lower serum Vaspin levels, suggesting that serum Vaspin level may be a useful marker for prediction of ovulation induction (140). Taken together, we can conclude that elevated serum Vaspin levels in patients with PCOS are associated with their metabolism, mainly IR and BMI, and may act as a positive factor in regulating ovarian granulosa cell function, whereas the effect of Vaspin on the outcome of assisted conception in infertile patients with PCOS remains to be further investigated.

# Lipocalin 2

Lipocalin 2 is a novel adipokine (145), a member of the Lipocalin superfamily, whose coding gene is located on human chromosome 9 with the functional protein consisting of 198 amino acids. The major source of Lipocalin 2 expression is white adipose tissue (WAT) (146). Megalin, a glycoprotein located on the surface of the cell membrane, has been proposed for human Lipocalin 2, and they bind to each other to exert biological effects (147). Lipocalin 2 has been illustrated to play a role in the pathophysiological processes such as cell differentiation, apoptosis, organogenesis, and inflammation (148). In addition, it has been shown to be significantly upregulated in obesity and T2DM (149).

No meta-analysis of Lipocalin 2 levels in serum of PCOS patients is available thus far, and there are inconsistent results in the published studies. Two of them showed an elevated serum Lipocalin 2 level in PCOS patients (150, 151), correlated with serum insulin levels, HOMA-IR, and free testosterone. Gencer et al. found that serum Lipocalin 2 levels were lower in PCOS compared to those with normal ovulation (152). Patients with PCOS are known to be at an increased cardiac risk, but no correlation has been shown between Lipocalin 2 levels and early detection of atherosclerosis in PCOS patients. Three other studies showed no significant difference between patients with PCOS and the control group (153-155), which illustrated that Lipocalin 2 levels were higher in overweight/obese PCOS patients than in normal weight PCOS and control patients. Lipocalin 2 was independently associated with BMI, and weight loss led to a decrease in Lipocalin 2 levels. In addition, the study by Martínez-García et al. added conviction to the masculinization of adipokine secretion in PCOS patients (156) when comparing the expression of Lipocalin 2 of subcutaneous (SAT) and visceral adipose tissue (VAT) between severely obese men and hyperandrogenic women presenting with PCOS submitting to bariatric surgery. They found that in men and women with PCOS, Lipocalin 2 expression was higher in VAT than in SAT; in the control group, the contrary is also the case. Two other studies yielded lower Lipocalin 2 levels in PCOS patients compared to controls (157, 158). Because Lipocalin 2 activates atherosclerotic plaques, low Lipocalin 2 levels are thought to be protective in patients with PCOS (158). Although attenuated after adjustment for waist and fasting glucose, Lipocalin 2 remained positively associated with diabetes progression in the PCOS group (157). Thus far, there are no studies on the association of Lipocalin 2 gene polymorphisms with PCOS. Despite different results, we can see that Lipocalin 2 may be present as a protective factor in PCOS. However, works on expounding the mechanisms between Lipocalin 2 and PCOS remain insufficient until now.

## Omentin

Omentin, a novel adipokine, was originally identified from the human omental adipose tissue cDNA library and thus named (159). The human Omentin gene is located on chromosome 1q21.3 and encodes a 313 amino acid protein. Of note, visceral adipose tissue rather than subcutaneous adipose tissue often expresses Omentin-1 preferentially and abundantly (160). Omentin is a secretory adipokine with pleiotropic effects, and Omentin plays a positive regulatory role in IR, inflammation, and regulation of endothelial function and so on (161, 162).

A meta-analysis in 2017 focusing on circulating Omentin levels in women with PCOS showed significant low levels of Omentin in patients with PCOS (163), and numerous studies have demonstrated the role of Omentin in the pathological processes of inflammatory state, IR, and steroid hormone production in PCOS (162, 164, 165). Cloix et al. identified the expression of Omentin in human ovarian granulosa cells for the first time in 2014 (165). They found similar levels of Omentin in plasma and follicular fluid in control patients but significantly higher levels of Omentin in follicular fluid than in plasma in patients with PCOS. They also demonstrated the function of human granulosa-lutein cells to secrete Omentin under insulin stimulation. In addition, the treatment of human recombinant Omentin increased nicotinamide phosphoribosyl transferase (NAMPT), IGF-1induced progesterone, and estradiol secretion, and this effect disappeared when NAMPT was knocked down. Another article aimed at studying the regulation of Omentin secretion in vivo and in vitro revealed low expression of Omentin in omental adipose tissue of women with PCOS and demonstrated that insulin can reduce Omentin expression in a dose-dependent manner (166). Another case-control study showed that after adjusting for BMI, Omentin levels in PCOS individuals with IR were lower than in those without IR. Furthermore, Omentin was negatively correlated with BMI, HOMA-IR, and fasting insulin (164). Taken together, we can speculate that insulin regulates Omentin in different ways at different sites which explains, to some extent, the opposite direction of changes in Omentin in serum and follicular fluid of PCOS patients. A cross-sectional study (162) conducted by Franik et al. revealed that plasma Omentin levels were reduced and associated with the high circulating TNF- $\alpha$  and low IL-6 levels in patients with PCOS. We know that IL6 has both pro-inflammatory and anti-inflammatory properties. While it is not clear whether Omentin has a driving or inhibitory effect in the inflammatory state of PCOS, it does affect its inflammatory state based on the data provided by Franik et al. It was shown that serum Omentin levels were negatively correlated with free testosterone levels and tended to decrease as free testosterone levels increased (167), suggesting that the increase in free testosterone in PCOS patients led to a decrease in the level of the protective factor-Omentin. PCOS patients with HA may be accompanied by hirsutism. In a study on the relationship between female hirsutism and serum Omentin (168) measuring the serum Omentin and free testosterone in 30 patients with hirsute PCOS, 30 patients with idiopathic hirsutism and 25 healthy control women, a significant inverse correlation was



FIGURE 1 | Schematic representation of adipokine expression and its role in PCOS. The solid dots of different colors represents the distribution of adipokines, with the corresponding Y as their receptors . — — represents for promotion and inhibition respectively. As shown in the figure, leptin and adiponectin regulates hypothalamic secretion of GnRH. Leptin, together with Adiponectin and Chemerin regulates progesterone production. Resistin is related to testosterone production without clear mechanism. Chemerin and Omentin are associated with inflammation. Chemerin, Apelin, Leptin, Vaspin, Omentin play a role in regulating IR. In addition, Adiponectin is closely related to ovulation and Chemerin is involved in the formation of PCOM.GnRH, gonadotropin-releasing hormone; Gn, gonadotropin; LH, luteinizing hormone; P, proges-terone; T, testosterone; E2, Estrodiol; PCOM, polycystic ovary morphology; PGE, prostaglandin E; MAPK, mitogen-activated protein kinase; COX2, cyclooxygenase-2; CYP17A1, cytochrome P45017A1; CYP11A1, cytochrome P45011A1; IGF-1, insulin-like growth factor 1; IR, insulin resistance; IRS, Insulin receptor substrate; AKT, protein kinase B; INF- y, Interferon- y; 3 8 -HSD, 3 6 -hydroxysteroid dehydrogenase; GLUT4, glucose transporter4.

revealed in the serum testosterone level with Omentin, and the idiopathic hirsutism group had the highest significant level of serum Omentin level. This may indicate that Omentin may not play a role in the development of hirsutism in PCOS patients, but is directly

related to hyperandrogenism. Additionally, treatment with metformin and oral contraceptives significantly increased serum Omentin levels in PCOS patients (169, 170). Tan et al. and colleagues (170) also found that serum VEGF levels were elevated

Adipokines	Site of production	Circulating levels	Effects in PCOS
Adiponectin	White adipose tissue Pituitary gland Ovary	ţ	Regulates gonadal axis hormone secretion, promotes ovulation, and improves insulin resistance
Chemerin	White adipose tissue Ovary	1	Aggravates insulin resistance and reduces estrogen and progesterone expression
Metrnl	White adipose tissue	$\downarrow$	Negatively correlated with fasting glucose, fasting insulin, and HOMA-IR
Leptin	White adipose tissue	Ť	Aggravates IR, promotes inflammation, induces granulocyte apoptosis, and regulates steroid hormone secretion
Apelin	White adipose tissue Hypothalamus Pituitary gland Ovary	$\rightarrow$	Promotion of IFG-1-induced estrogen and progesterone secretion
Resistin	White adipose tissue Ovary	1	Inhibits estrogen and progesterone secretion, promotes testosterone production; activates macrophages to produce pro-inflammatory factors such as TNF- $\alpha$
Visfatin	Visceral adipose tissue Hypothalamus Pituitary gland Ovary	ſ	Participates in local energy metabolism in the ovary, regulates steroid hormone production such as suppressing androgen production and promoting IFG-1-induced estrogen and progesterone secretion
Vaspin	Visceral and subcutaneous adipose tissue, ovary	Î	Improves insulin sensitivity and regulates granulocyte function
Lipocalin 2	Visceral and subcutaneous adipose tissue	conflict	Associated with diabetes progression
Omentin	Visceral adipose tissue	Ţ	Regulates inflammatory status and reduces insulin resistance

↑, Increased circulating levels, ↓, Decreased circulating levels, →, equally circulating levels. are explained in the text. In the table, we only list the main sites of production and effects in PCOS.

in PCOS patients and decreased with the reduction of Omentin levels after metformin treatment. VEGF, an essential factor for angiogenesis, plays an important role in ovarian polycystic-like changes, and this study verified that Omentin may play a role in inhibiting polycystic ovary morphology (PCOM) changes. This may give us enlightenment on improving assisted conception outcomes in patients with PCOS.

## SUMMARY AND PROSPECTS

The occurrence of PCOS involves multiple pathological processes and is influenced by more than one factor just like its various clinical manifestations. Just as there are no isolated individuals in the world, all the adipokines mentioned above may interweave and influence the development of PCOS (Figure 1). It is noteworthy that Adiponectin, Leptin, and Apelin are all beneficial under normal physiological conditions, possibly with different trends in the body fluids of PCOS patients. Adipokines exhibit compensatory overexpression due to the decline in their receptors, such as Leptin and Apelin. Although each adipokine affects more or less all aspects of PCOS, the focus is different. Chemerin worsens the IR of PCOS by affecting glucose utilization of granulosa cells. Adiponectin plays an important role in promoting ovulation and may be involved in maintaining negative feedback in the reproductive axis, but unfortunately, it may be declined in PCOS with unknown reason. Apelin maintains progesterone secretion in the luteal phase, but is limited by the reduction of its receptor caused by IR, the most common manifestation of PCOS. As for Resistin, Visfatin, and Metrnl, current studies are limited only to animals or cells, of which results are confined to the correlation of clinical or laboratory data. When it comes to Vaspin, Lipocalin 2, and Omentin, most of their current studies in PCOS patients are related to IR, even with other different priorities. Omentin plays a role in the regulation of inflammatory status and steroid hormone production in patients with PCOS, whereas Vaspin

## REFERENCES

- Li R, Zhang Q, Yang D, Li S, Lu S, Wu X, et al. Prevalence of Polycystic Ovary Syndrome in Women in China: A Large Community-Based Study. *Hum Reprod* (2013) 28:2562–9. doi: 10.1093/humrep/det262
- Jayasena CN, Franks S. The Management of Patients With Polycystic Ovary Syndrome. Nat Rev Endocrinol (2014) 10:624–36. doi: 10.1038/ nrendo.2014.102
- Rotterdam E. A.-S. P. C. W. G. Revised 2003 Consensus on Diagnostic Criteria and Long-Term Health Risks Related to Polycystic Ovary Syndrome. *Fertil Steril* (2004) 81:19–25. doi: 10.1016/j.fertnstert.2003.10.004
- Meier U, Gressner AM. Endocrine Regulation of Energy Metabolism: Review of Pathobiochemical and Clinical Chemical Aspects of Leptin, Ghrelin, Adiponectin, and Resistin. *Clin Chem* (2004) 50:1511–25. doi: 10.1373/clinchem.2004.032482
- Campos DB, Palin MF, Bordignon V, Murphy BD. The 'Beneficial' Adipokines in Reproduction and Fertility. Int J Obes (Lond) (2008) 32:223–31. doi: 10.1038/sj.ijo.0803719
- 6. Lin K, Sun X, Wang X, Wang H, Chen X. Circulating Adipokine Levels in Nonobese Women With Polycystic Ovary Syndrome and in Nonobese

has been shown to occupy an important position in the proliferation, viability, and function of ovary granulosa cells. Due to the dynamic changes in the body and the complex interplay between hormones, it is not easy to understand all the specific mechanisms involved, and hence, the underlying mechanism needs to be confirmed further.

PCOS, as a highly heterogeneous syndrome, is still controversial in its diagnosis, and its pathogenesis is not yet clear, making it a hot topic of continuous research for so many years. Adipokines, on account of their significant roles in energy metabolism, inflammation, insulin-resistance, cell senescence, and apoptosis, together or independently affect the pathological process of PCOS (**Table 1**), and have become one of the most fascinating areas of PCOS exploration. Current studies have confirmed that a variety of adipokines are differentially expressed in PCOS and non-PCOS populations, and are related to ovarian angiogenesis, steroid hormonegeneration, follicular development, and granulosa cell apoptosis. Notwithstanding, most studies are limited to defining the superficial correlation rather than clarifying the underlying mechanism, for which we still have a long way to go.

# **AUTHOR CONTRIBUTIONS**

PC and ZZ contributed to conceived and designed the review. PC wrote the paper. RJ and YL did the document retrieval. PC and MC draw the picture. LZ polished the paper. ZZ cheched the paper. All authors listed have made a substantial and intellectual contribution to the review and approved it for publication.

## FUNDING

This study was supported by Natural Science Foundation of Hebei Province (Beijing-Tianjin-Hebei Cooperation Special Project)(H2019206712).

Control Women: A Systematic Review and Meta-Analysis. Front Endocrinol (2020) 11:537809. doi: 10.3389/fendo.2020.537809

- Arita Y, Kihara S, Ouchi N, Takahashi M, Maeda K, Miyagawa J-i, et al. Paradoxical Decrease of an Adipose-Specific Protein, Adiponectin, in Obesity. *Biochem Biophys Res Commun* (1999) 425:560–4. doi: 10.1016/ j.bbrc.2012.08.024
- Hu E, Liang P, Spiegelman BM. Adipoq is a Novel Adipose-Specific Gene Dysregulated in Obesity. J Biol Chem (1996) 271:10697–703. doi: 10.1074/ jbc.271.18.10697
- Ambroziak M, Kolanowska M, Bartoszewicz Z, Budaj A. Adiponectin Gene Variants and Decreased Adiponectin Plasma Levels are Associated With the Risk of Myocardial Infarction in Young Age. *Gene* (2018) 642:498–504. doi: 10.1016/j.gene.2017.11.064
- Yamauchi T, Kamon J, Ito Y, Tsuchida A, Yokomizo T, Kita S, et al. Cloning of Adiponectin Receptors That Mediate Antidiabetic Metabolic Effects. *Nature* (2003) 423:762–9. doi: 10.1038/nature01705
- Khoramipour K, Chamari K, Hekmatikar AA, Ziyaiyan A, Taherkhani S, Elguindy NM, et al. Adiponectin: Structure, Physiological Functions, Role in Diseases, and Effects of Nutrition. *Nutrients* (2021) 13 (4):1180. doi: 10.3390/ nu13041180

- 12. Dumesic DA, Lobo RA. Cancer Risk and PCOS. *Steroids* (2013) 78:782–5. doi: 10.1016/j.steroids.2013.04.004
- Parida S, Siddharth S, Sharma D. Adiponectin, Obesity, and Cancer: Clash of the Bigwigs in Health and Disease. *Int J Mol Sci* (2019) 20:2519. doi: 10.3390/ ijms20102519
- Spranger J, Kroke A, Möhlig M, Bergmann MM, Ristow M, Boeing H, et al. Adiponectin and Protection Against Type 2 Diabetes Mellitus. *Lancet* (2003) 361:226–8. doi: 10.1016/S0140-6736(03)12255-6
- Wild RA, Carmina E, Diamanti-Kandarakis E, Dokras A, Escobar-Morreale HF, Futterweit W, et al. Assessment of Cardiovascular Risk and Prevention of Cardiovascular Disease in Women With the Polycystic Ovary Syndrome: A Consensus Statement by the Androgen Excess and Polycystic Ovary Syndrome (AE-PCOS) Society. J Clin Endocrinol Metab (2010) 95:2038–49. doi: 10.1210/jc.2009-2724
- Chou IP, Lin YY, Ding ST, Chen CY. Adiponectin Receptor 1 Enhances Fatty Acid Metabolism and Cell Survival in Palmitate-Treated Hepg2 Cells Through the PI3 K/AKT Pathway. *Eur J Nutr* (2014) 53:907–17. doi: 10.1007/s00394-013-0594-7
- Comim FV, Hardy K, Franks S. Adiponectin and its Receptors in the Ovary: Further Evidence for a Link Between Obesity and Hyperandrogenism in Polycystic Ovary Syndrome. *PLoS One* (2013) 8:e80416. doi: 10.1371/ journal.pone.0080416
- Almabouada F, Diaz-Ruiz A, Rabanal-Ruiz Y, Peinado JR, Vazquez-Martinez R, Malagon MM. Adiponectin Receptors Form Homomers and Heteromers Exhibiting Distinct Ligand Binding and Intracellular Signaling Properties. J Biol Chem (2013) 288:3112–25. doi: 10.1074/jbc.M112.404624
- Ledoux S, Campos DB, Lopes FL, Dobias-Goff M, Palin MF, Murphy BD. Adiponectin Induces Periovulatory Changes in Ovarian Follicular Cells. *Endocrinology* (2006) 147:5178–86. doi: 10.1210/en.2006-0679
- Lagaly DV, Aad PY, Grado-Ahuir JA, Hulsey LB, Spicer LJ. Role of Adiponectin in Regulating Ovarian Theca and Granulosa Cell Function. *Mol Cell Endocrinol* (2008) 284:38–45. doi: 10.1016/j.mce.2008.01.007
- Rodriguez-Pacheco F, Martinez-Fuentes AJ, Tovar S, Pinilla L, Tena-Sempere M, Dieguez C, et al. Regulation of Pituitary Cell Function by Adiponectin. *Endocrinology* (2007) 148:401–10. doi: 10.1210/en.2006-1019
- Gutman G, Barak V, Maslovitz S, Amit A, Lessing JB, Geva E. Recombinant Luteinizing Hormone Induces Increased Production of Ovarian Follicular Adiponectin *In Vivo*: Implications for Enhanced Insulin Sensitivity. *Fertil Steril* (2009) 91:1837–41. doi: 10.1016/j.fertnstert.2008.02.006
- Mirza SS, Shafique K, Shaikh AR, Khan NA, Anwar Qureshi M. Association Between Circulating Adiponectin Levels and Polycystic Ovarian Syndrome. *J Ovarian Res* (2014) 7:18. doi: 10.1186/1757-2215-7-18
- 24. Carmina E, Bucchieri S, Mansueto P, Rini G, Ferin M, Lobo RA. Circulating Levels of Adipose Products and Differences in Fat Distribution in the Ovulatory and Anovulatory Phenotypes of Polycystic Ovary Syndrome. *Fertil Steril* (2009) 91:1332–5. doi: 10.1016/j.fertnstert.2008.03.007
- Palomba S, Marotta R, Di Cello A, Russo T, Falbo A, Orio F, et al. Pervasive Developmental Disorders in Children of Hyperandrogenic Women With Polycystic Ovary Syndrome: A Longitudinal Case-Control Study. *Clin Endocrinol (Oxf)* (2012) 77:898–904. doi: 10.1111/j.1365-2265.2012.04443.x
- Franik G, Maksym M, Owczarek AJ, Chudek J, Madej P, Olszanecka-Glinianowicz M. Estradiol/Testosterone and Estradiol/Androstenedione Indexes and Nutritional Status in PCOS Women - a Pilot Study. *Eur J Obstet Gynecol Reprod Biol* (2019) 242:166–9. doi: 10.1016/j.ejogrb.2019.05.045
- Zuo M, Liao G, Zhang W, Xu D, Lu J, Tang M, et al. Effects of Exogenous Adiponectin Supplementation in Early Pregnant PCOS Mice on the Metabolic Syndrome of Adult Female Offspring. J Ovarian Res (2021) 14:15. doi: 10.1186/s13048-020-00755-z
- Jorquera G, Echiburú B, Crisosto N, Sotomayor-Zárate R, Maliqueo M, Cruz G. Metformin During Pregnancy: Effects on Offspring Development and Metabolic Function. *Front Pharmacol* (2020) 11:653. doi: 10.3389/ fphar.2020.00653
- Inal HA, Yilmaz N, Gorkem U, Oruc AS, Timur H. The Impact of Follicular Fluid Adiponectin and Ghrelin Levels Based on BMI on IVF Outcomes in PCOS. J Endocrinol Invest (2016) 39:431–7. doi: 10.1007/s40618-015-0392-6
- 30. Zhang W, Wei D, Sun X, Li J, Yu X, Shi Y, et al. Family-Based Analysis of Adiponectin Gene Polymorphisms in Chinese Han Polycystic Ovary

Syndrome. Fertil Steril (2014) 101:1419-23. doi: 10.1016/ j.fertnstert.2014.01.035

- 31. Tiongco RE, Cabrera FJ, Clemente B, Flake CC, Salunga MA, Pineda-Cortel MR. G276T Polymorphism in the ADIPOQ Gene is Associated With a Reduced Risk of Polycystic Ovarian Syndrome: A Meta-Analysis of Asian Population. *Taiwanese J Obstet Gynecol* (2019) 58:409–16. doi: 10.1016/ j.tjog.2018.12.002
- 32. Sun X, Wu X, Duan Y, Liu G, Yu X, Zhang W. Family-Based Association Study of Rs17300539 and Rs12495941 Polymorphism in Adiponectin Gene and Polycystic Ovary Syndrome in a Chinese Population. *Med Sci Monit* (2017) 23:78–84. doi: 10.12659/MSM.901944
- Liu Z, Wang Z, Hao C, Tian Y, Fu J. Effects of ADIPOQ Polymorphisms on PCOS Risk: A Meta-Analysis. *Reprod Biol Endocrinol: RB&E* (2018) 16:120. doi: 10.1186/s12958-018-0439-6
- 34. Panidis D, Kourtis A, Kukuvitis A, Farmakiotis D, Xita N, Georgiou I, et al. Association of the T45G Polymorphism in Exon 2 of the Adiponectin Gene With Polycystic Ovary Syndrome: Role of Delta4-Androstenedione. *Hum Reprod* (2004) 19:1728–33. doi: 10.1093/humrep/deh336
- 35. Xita N, Georgiou I, Chatzikyriakidou A, Vounatsou M, Papassotiriou G-P, Papassotiriou I, et al. Effect of Adiponectin Gene Polymorphisms on Circulating Adiponectin and Insulin Resistance Indexes in Women With Polycystic Ovary Syndrome. *Clin Chem* (2005) 51:416–23. doi: 10.1373/ clinchem.2004.043109
- 36. Nagpal S, Patel S, Jacobe H, DiSepio D, Ghosn C, Malhotra M, et al. Tazarotene-Induced Gene 2 (TIG2), a Novel Retinoid-Responsive Gene in Skin. J Invest Dermatol (1997) 109:91–5. doi: 10.1111/1523-1747.ep12276660
- Bozaoglu K, Bolton K, McMillan J, Zimmet P, Jowett J, Collier G, et al. Chemerin is a Novel Adipokine Associated With Obesity and Metabolic Syndrome. *Endocrinology* (2007) 148:4687–94. doi: 10.1210/en.2007-0175
- Goralski KB, McCarthy TC, Hanniman EA, Zabel BA, Butcher EC, Parlee SD, et al. Chemerin, a Novel Adipokine That Regulates Adipogenesis and Adipocyte Metabolism. J Biol Chem (2007) 282:28175–88. doi: 10.1074/ jbc.M700793200
- Roh SG, Song SH, Choi KC, Katoh K, Wittamer V, Parmentier M, et al. Chemerin-a New Adipokine That Modulates Adipogenesis via its Own Receptor. Biochem Biophys Res Commun (2007) 362:1013–8. doi: 10.1016/ j.bbrc.2007.08.104
- Wittamer V, Franssen JD, Vulcano M, Mirjolet JF, Le Poul E, Migeotte I, et al. Specific Recruitment of Antigen-Presenting Cells by Chemerin, a Novel Processed Ligand From Human Inflammatory Fluids. *J Exp Med* (2003) 198:977–85. doi: 10.1084/jem.20030382
- Meder W, Wendland M, Busmann A, Kutzleb C, Spodsberg N, John H, et al. Characterization of Human Circulating TIG2 as a Ligand for the Orphan Receptor Chemr23. *FEBS Lett* (2003) 555:495–9. doi: 10.1016/S0014-5793 (03)01312-7
- Reverchon M, Cornuau M, Rame C, Guerif F, Royere D, Dupont J. Chemerin Inhibits IGF-1-Induced Progesterone and Estradiol Secretion in Human Granulosa Cells. *Hum Reprod* (2012) 27:1790–800. doi: 10.1093/ humrep/des089
- Li L, et al. Chemerin-Derived Peptide C-20 Suppressed Gonadal Steroidogenesis. Am J Reprod Immunol (New York NY: 1989) (2014) 71:265-77. doi: 10.1111/aji.12164
- 44. Li L, Huang C, Zhang X, Wang J, Ma P, Liu Y, et al. Expression of Chemerin and its Receptors in Rat Testes and its Action on Testosterone Secretion. *J Endocrinol* (2014) 220:155–63. doi: 10.1530/JOE-13-0275
- 45. Ahn SS, Yoon T, Song JJ, Park YB, Lee SW. Serum Adipokine Profiles in Patients With Microscopic Polyangiitis and Granulomatosis With Polyangiitis: An Exploratory Analysis. *PLoS One* (2021) 16:e0254226. doi: 10.1371/journal.pone.0254226
- Zhang RX, Zhang X. [Research Progress of Adipokine and Kidney Disease]. Zhongguo Yi Xue Ke Xue Yuan Xue Bao Acta Academiae Med Sin (2021) 43:130-5. doi: 10.3881/j.issn.1000-503X.11530
- 47. An X, Liu J, Li Y, Dou Z, Li N, Suo Y, et al. Chemerin/CMKLR1 Ameliorates Nonalcoholic Steatohepatitis by Promoting Autophagy and Alleviating Oxidative Stress Through the JAK2-STAT3 Pathway. *Peptides* (2021) 135:170422. doi: 10.1016/j.peptides.2020.170422

- Veru-Lesmes F, Guay S, Shah JL, Schmitz N, Giguère C, Joober R, et al. Adipose Tissue Dysregulation at the Onset of Psychosis: Adipokines and Social Determinants of Health. *Psychoneuroendocrinology* (2021) 123:104915. doi: 10.1016/j.psyneuen.2020.104915
- Zaitseva M, Vollenhoven BJ, Rogers PAW. Retinoids Regulate Genes Involved in Retinoic Acid Synthesis and Transport in Human Myometrial and Fibroid Smooth Muscle Cells. *Hum Reprod* (2008) 23:1076–86. doi: 10.1093/humrep/den083
- Carlino C, Trotta E, Stabile H, Morrone S, Bulla R, Soriani A, et al. Chemerin Regulates NK Cell Accumulation and Endothelial Cell Morphogenesis in the Decidua During Early Pregnancy. J Clin Endocrinol Metab (2012) 97:3603– 12. doi: 10.1210/jc.2012-1102
- 51. Kasher-Meron M, Mazaki-Tovi S, Barhod E, Hemi R, Haas J, Gat I, et al. Chemerin Concentrations in Maternal and Fetal Compartments: Implications for Metabolic Adaptations to Normal Human Pregnancy. *J Perinat Med* (2014) 42:371–8. doi: 10.1515/jpm-2013-0166
- Tan BK, Chen J, Farhatullah S, Adya R, Kaur J, Heutling D, et al. Insulin and Metformin Regulate Circulating and Adipose Tissue Chemerin. *Diabetes* (2009) 58:1971–7. doi: 10.2337/db08-1528
- 53. Li X, Zhu Q, Wang W, Qi J, He Y, Wang Y, et al. Elevated Chemerin Induces Insulin Resistance in Human Granulosa-Lutein Cells From Polycystic Ovary Syndrome Patients. *FASEB J* (2019) 33:11303–13. doi: 10.1096/ fj.201802829R
- Estienne A, Mellouk N, Bongrani A, Plotton I, Langer I, Ramé C, et al. Involvement of Chemerin and CMKLR1 in the Progesterone Decrease by PCOS Granulosa Cells. *Reproduction* (2021) 162(6):427–36. doi: 10.1530/ REP-21-0265
- 55. Lima PDA, Nivet AL, Wang Q, Chen YA, Leader A, Cheung A, et al. Polycystic Ovary Syndrome: Possible Involvement of Androgen-Induced, Chemerin-Mediated Ovarian Recruitment of Monocytes/Macrophages. *Biol Reprod* (2018) 99:838–52. doi: 10.1093/biolre/ioy096
- Huang R, Yue J, Sun Y, Zheng J, Tao T, Li S, et al. Increased Serum Chemerin Concentrations in Patients With Polycystic Ovary Syndrome: Relationship Between Insulin Resistance and Ovarian Volume. *Clin Chim Acta* (2015) 450:366–9. doi: 10.1016/j.cca.2015.09.015
- 57. Silva AF, Abruzzese GA, Ferrer MJ, Heber MF, Ferreira SR, Cerrone GE, et al. Fetal Programming by Androgen Excess Impairs Liver Lipid Content and Pparg Expression in Adult Rats. J Dev Origins Health Dis (2021) 19:1–10. doi: 10.1017/S2040174421000416
- Kabil Kucur S, Kurek Eken M, Sanli I, Kutlu T, Bilgic BE, Altuntas SL, et al. Predictive Value of Serum and Follicular Fluid Chemerin Concentrations During Assisted Reproductive Cycles in Women With Polycystic Ovary Syndrome. *Gynecol Endocrinol* (2021) 37(9):8147–8. doi: 10.1080/ 09513590.2021.1937100
- Movahed Z, Kohan L, Fallahi S, Tabiee O. Influence of Chemerin Rs17173608 Polymorphism on Polycystic Ovary Syndrome Susceptibility. *Taiwanese J Obstet Gynecol* (2015) 54:280–3. doi: 10.1016 /j.tjog.2014.05.007
- 60. Ramialison M, Bajoghli B, Aghaallaei N, Ettwiller L, Gaudan S, Wittbrodt B, et al. Rapid Identification of PAX2/5/8 Direct Downstream Targets in the Otic Vesicle by Combinatorial Use of Bioinformatics Tools. *Genome Biol* (2008) 9:R145. doi: 10.1186/gb-2008-9-10-r145
- 61. Jørgensen JR, Fransson A, Fjord-Larsen L, Thompson LH, Houchins JP, Andrade N, et al. Cometin is a Novel Neurotrophic Factor That Promotes Neurite Outgrowth and Neuroblast Migration *In Vitro* and Supports Survival of Spiral Ganglion Neurons *In Vivo. Exp Neurol* (2012) 233:172– 81. doi: 10.1016/j.expneurol.2011.09.027
- 62. Li ZY, Zheng S-L, Wang P, Xu T-Y, Guan Y-F, Zhang Y-J, et al. Subfatin is a Novel Adipokine and Unlike Meteorin in Adipose and Brain Expression. *CNS Neurosci Ther* (2014) 20:344–54. doi: 10.1111/cns.12219
- Jørgensen JR, Thompson L, Fjord-Larsen L, Krabbe C, Torp M, Kalkkinen N, et al. Characterization of Meteorin-an Evolutionary Conserved Neurotrophic Factor. J Mol Neurosci (2009) 39:104–16. doi: 10.1007/ s12031-009-9189-4
- 64. Ushach I, Burkhardt AM, Martinez C, Hevezi PA, Gerber PA, Buhren BA, et al. METEORIN-LIKE is a Cytokine Associated With Barrier Tissues and Alternatively Activated Macrophages. *Clin Immunol (Orlando Fla.)* (2015) 156:119–27. doi: 10.1016/j.clim.2014.11.006

- 65. Zhang SL, Li ZY, Wang DS, Xu TY, Fan MB, Cheng MH, et al. Aggravated Ulcerative Colitis Caused by Intestinal Metrnl Deficiency is Associated With Reduced Autophagy in Epithelial Cells. *Acta Pharmacol Sin* (2020) 41:763– 70. doi: 10.1038/s41401-019-0343-4
- Ruperez C, Ferrer-Curriu G, Cervera-Barea A, Florit L, Guitart-Mampel M, Garrabou G, et al. Meteorin-Like/Meteorin-Beta Protects Heart Against Cardiac Dysfunction. J Exp Med (2021) 218:e20201206. doi: 10.1084/ jem.20201206
- Deniz R, Yavuzkir S, Ugur K, Ustebay DU, Baykus Y, Ustebay S, et al. Subfatin and Asprosin, Two New Metabolic Players of Polycystic Ovary Syndrome. J Obstet Gynaecol (2021) 41:279-84. doi: 10.1080/ 01443615.2020.1758926
- Fouani FZ, Fadaei R, Moradi N, Zandieh Z, Ansaripour S, Yekaninejad MS, et al. Circulating Levels of Meteorin-Like Protein in Polycystic Ovary Syndrome: A Case-Control Study. *PLoS One* (2020) 15:e0231943. doi: 10.1371/journal.pone.0231943
- 69. Yavuzkir S, Ugur K, Deniz R, Ustebay DU, Mirzaoglu M, Yardim M, et al. Maternal and Umbilical Cord Blood Subfatin and Spexin Levels in Patients With Gestational Diabetes Mellitus. *Peptides* (2020) 126:170277. doi: 10.1016/j.peptides.2020.170277
- Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional Cloning of the Mouse Obese Gene and its Human Homologue. *Nature* (1994) 372:425–32. doi: 10.1038/372425a0
- Halaas JL, Gajiwala KS, Maffei M, Cohen SL, Chait BT, Rabinowitz D, et al. Weight-Reducing Effects of the Plasma Protein Encoded by the Obese Gene. *Sci (New York N.Y.)* (1995) 269:543–6. doi: 10.1126/science.7624777
- Tartaglia LA, Dembski M, Weng X, Deng N, Culpepper J, Devos R, et al. Identification and Expression Cloning of a Leptin Receptor, OB-R. *Cell* (1995) 83:1263–71. doi: 10.1016/0092-8674(95)90151-5
- Sahu A. Leptin Signaling in the Hypothalamus: Emphasis on Energy Homeostasis and Leptin Resistance. Front Neuroendocrinol (2003) 24:225– 53. doi: 10.1016/j.yfrne.2003.10.001
- Zhang Y, Chua S. Leptin Function and Regulation. Compr Physiol (2017) 8:351–69. doi: 10.1002/cphy.c160041
- Kucera R, Babuska V, Ulcova-Gallova Z, Kulda V, Topolcan O. Follicular Fluid Levels of Anti-Müllerian Hormone, Insulin-Like Growth Factor 1 and Leptin in Women With Fertility Disorders. *Syst Biol Reprod Med* (2018) 64:220–3. doi: 10.1080/19396368.2018.1450906
- 76. Yildizhan R, Ilhan GA, Yildizhan B, Kolusari A, Adali E, Bugdayci G. Serum Retinol-Binding Protein 4, Leptin, and Plasma Asymmetric Dimethylarginine Levels in Obese and Nonobese Young Women With Polycystic Ovary Syndrome. *Fertil Steril* (2011) 96:246–50. doi: 10.1016/ j.fertnstert.2011.04.073
- Leroy P, Dessolin S, Villageois P, Moon BC, Friedman JM, Ailhaud G, et al. Expression of Ob Gene in Adipose Cells. Regulation by Insulin. *J Biol Chem* (1996) 271:2365–8. doi: 10.1074/jbc.271.5.2365
- Wang J, Gong P, Li C, Pan M, Ding Z, Ge X, et al. Correlation Between Leptin and IFN-Gamma Involved in Granulosa Cell Apoptosis in PCOS. *Gynecol Endocrinol* (2020) 36:1051–6. doi: 10.1080/09513590. 2020.1760817
- 79. Liu Y, Xu Y-C, Cui Y-G, Jiang S-W, Diao F-Y, Liu J-Y, et al. Androgen Excess Increases Food Intake in a Rat Polycystic Ovary Syndrome Model by Down-Regulating Hypothalamus Insulin and Leptin Signaling Pathways Preceding Weight Gain. *Neuroendocrinology* (2021). doi: 10.1159/ 000521236
- Kargasheh FB, Ansaripour S, Borumandnia N, Moradi N, Zandieh Z, Maleki M, et al. Association of Leptin G2548A and Leptin Receptor Q223R Polymorphisms and Their Serum Levels With Infertility and Recurrent Pregnancy Loss in Iranian Women With Polycystic Ovary Syndrome. *PLoS One* (2021) 16:e0255920. doi: 10.1371/journal.pone.0255920
- Vilariño-García T, Pérez-Pérez A, Santamaría-López E, Prados N, Fernández-Sánchez M, Sánchez-Margalet V. Sam68 Mediates Leptin Signaling and Action in Human Granulosa Cells: Possible Role in Leptin Resistance in PCOS. *Endocr Connect* (2020) 9:479–88. doi: 10.1530/EC-20-0062
- Li N, Yang C, Xie H, Liu Y, Liao Y. Effects of Aerobic Exercise on Rats With Hyperandrogenic Polycystic Ovarian Syndrome. *Int J Endocrinol* (2021) 2021:5561980. doi: 10.1155/2021/5561980

- Vardar SA, Karaca A, Güldiken S, Palabıyık O, Süt N, Demir AM. High-Intensity Interval Training Acutely Alters Plasma Adipokine Levels in Young Overweight/Obese Women. Arch Physiol Biochem (2018) 124:149– 55. doi: 10.1080/13813455.2017.1369998
- 84. Torres-García R, Camarillo-Romero EdS, Majluf-Cruz A, Vázquez deAnda GF, Loe-Ochoa AM, Montenegro-Morales LP, et al. [Effect of Exercise on the Serum Concentrations of Leptin and Adiponectin in Adolescents With Risk Factors of Developing Diabetes]. *Rev Med Inst Mex Seguro Soc* (2017) 55:708–14.
- Aktaş HŞ, Uzun YE, Kutlu O, Pençe HH, Özçelik F, Çil EÖ, et al. The Effects of High Intensity-Interval Training on Vaspin, Adiponectin and Leptin Levels in Women With Polycystic Ovary Syndrome. *Arch Physiol Biochem* (2019) 128: 37–42. doi: 10.1080/13813455.2019.1662450
- Dallel M, Douma Z, Finan RR, Hachani F, Letaifa DB, Mahjoub T, et al. Contrasting Association of Leptin Receptor Polymorphisms and Haplotypes With Polycystic Ovary Syndrome in Bahraini and Tunisian Women: A Case-Control Study. *Biosci Rep* (2021) 41(1):BSR20202726. doi: 10.1042/ BSR20202726
- Tatemoto K, Hosoya M, Habata Y, Fujii R, Kakegawa T, Zou MX, et al. Isolation and Characterization of a Novel Endogenous Peptide Ligand for the Human APJ Receptor. *Biochem Biophys Res Commun* (1998) 251:471–6. doi: 10.1006/bbrc.1998.9489
- Boucher J, Masri B, Daviaud D, Gesta S, Guigné C, Mazzucotelli A, et al. Apelin, a Newly Identified Adipokine Up-Regulated by Insulin and Obesity. *Endocrinology* (2005) 146:1764–71. doi: 10.1210/en.2004-1427
- Kawamata Y, Habata Y, Fukusumi S, Hosoya M, Fujii R, Hinuma S, et al. Molecular Properties of Apelin: Tissue Distribution and Receptor Binding. *Biochim Biophys Acta* (2001) 1538:162–71. doi: 10.1016/S0167-4889(00) 00143-9
- Cheng J, Luo X, Huang Z, Chen L. Apelin/APJ System: A Potential Therapeutic Target for Endothelial Dysfunction-Related Diseases. J Cell Physiol (2019) 234:12149–60. doi: 10.1002/jcp.27942
- Antushevich H, Wójcik M. Review: Apelin in Disease. Clin Chim Acta (2018) 483:241–8. doi: 10.1016/j.cca.2018.05.012
- 92. Shimizu T, Kosaka N, Murayama C, Tetsuka M, Miyamoto A. Apelin and APJ Receptor Expression in Granulosa and Theca Cells During Different Stages of Follicular Development in the Bovine Ovary: Involvement of Apoptosis and Hormonal Regulation. *Anim Reprod Sci* (2009) 116:28–37. doi: 10.1016/j.anireprosci.2009.01.009
- Schilfarth S, Antoni B, Schams D, Meyer HH, Berisha B. The Expression of Apelin and its Receptor APJ During Different Physiological Stages in the Bovine Ovary. Int J Biol Sci (2009) 5:344–50. doi: 10.7150/ijbs.5.344
- Roche J, Ramé C, Reverchon M, Mellouk N, Rak A, Froment P, et al. Apelin (APLN) Regulates Progesterone Secretion and Oocyte Maturation in Bovine Ovarian Cells. *Reproduction* (2017) 153:589–603. doi: 10.1530/REP-16-0677
- 95. Rak A, Drwal E, Rame C, Knapczyk-Stwora K, Słomczyńska M, Dupont J, et al. Expression of Apelin and Apelin Receptor (APJ) in Porcine Ovarian Follicles and In Vitro Effect of Apelin on Steroidogenesis and Proliferation Through APJ Activation and Different Signaling Pathways. *Theriogenology* (2017) 96:126–35. doi: 10.1016/j.theriogenology.2017.04.014
- 96. Taheri S, Murphy K, Cohen M, Sujkovic E, Kennedy A, Dhillo W, et al. The Effects of Centrally Administered Apelin-13 on Food Intake, Water Intake and Pituitary Hormone Release in Rats. *Biochem Biophys Res Commun* (2002) 291:1208–12. doi: 10.1006/bbrc.2002.6575
- 97. Estienne A, Bongrani A, Reverchon M, Ramé C, Ducluzeau PH, Froment P, et al. Involvement of Novel Adipokines, Chemerin, Visfatin, Resistin and Apelin in Reproductive Functions in Normal and Pathological Conditions in Humans and Animal Models. *Int J Mol Sci* (2019) 20(18):4431. doi: 10.3390/ ijms20184431
- Bongrani A, Mellouk N, Rame C, Cornuau M, Guérif F, Froment P, et al. Ovarian Expression of Adipokines in Polycystic Ovary Syndrome: A Role for Chemerin, Omentin, and Apelin in Follicular Growth Arrest and Ovulatory Dysfunction? *Int J Mol Sci* (2019) 20(15):3778. doi: 10.3390/ijms20153778
- 99. Roche J, Rame C, Reverchon M, Mellouk N, Cornuau M, Guerif F, et al. Apelin (APLN) and Apelin Receptor (APLNR) in Human Ovary: Expression, Signaling, and Regulation of Steroidogenesis in Primary Human Luteinized Granulosa Cells. *Biol Reprod* (2016) 95:104. doi: 10.1095/biolreprod.116.141754

- 100. Abramovich D, Irusta G, Bas D, Cataldi NI, Parborell F, Tesone M. Angiopoietins/TIE2 System and VEGF are Involved in Ovarian Function in a DHEA Rat Model of Polycystic Ovary Syndrome. *Endocrinology* (2012) 153:3446–56. doi: 10.1210/en.2012-1105
- 101. Basini G, Bussolati S, Santini SE, Bianchi F, Careri M, Mangia A, et al. Antiangiogenesis in Swine Ovarian Follicle: A Potential Role for 2-Methoxyestradiol. Steroids (2007) 72:660-5. doi: 10.1016/ j.steroids.2007.05.002
- 102. Wang W, McKinnie SMK, Patel VB, Haddad G, Wang Z, Zhabyeyev P, et al. Loss of Apelin Exacerbates Myocardial Infarction Adverse Remodeling and Ischemia-Reperfusion Injury: Therapeutic Potential of Synthetic Apelin Analogues. J Am Heart Assoc (2013) 2:e000249. doi: 10.1161/ JAHA.113.000249
- 103. Shirasuna K, Shimizu T, Sayama K, Asahi T, Sasaki M, Berisha B, et al. Expression and Localization of Apelin and its Receptor APJ in the Bovine Corpus Luteum During the Estrous Cycle and Prostaglandin F2alpha-Induced Luteolysis. *Reproduction* (2008) 135:519–25. doi: 10.1530/REP-07-0409
- 104. Stanek MB, Borman SM, Molskness TA, Larson JM, Stouffer RL, Patton PE. Insulin and Insulin-Like Growth Factor Stimulation of Vascular Endothelial Growth Factor Production by Luteinized Granulosa Cells: Comparison Between Polycystic Ovarian Syndrome (PCOS) and non-PCOS Women. *J Clin Endocrinol Metab* (2007) 92:2726–33. doi: 10.1210/jc.2006-2846
- 105. Dundar A, Kocahan S, Sahin L. Associations of Apelin, Leptin, Irisin, Ghrelin, Insulin, Glucose Levels, and Lipid Parameters With Physical Activity During Eight Weeks of Regular Exercise Training. Arch Physiol Biochem (2021) 127:291–5. doi: 10.1080/13813455.2019.1635622
- 106. Kon M, Ebi Y, Nakagaki K. Effects of Acute Sprint Interval Exercise on Follistatin-Like 1 and Apelin Secretions. Arch Physiol Biochem (2021) 127:223–7. doi: 10.1080/13813455.2019.1628067
- 107. Steppan CM, Bailey ST, Bhat S, Brown EJ, Banerjee RR, Wright CM, et al. The Hormone Resistin Links Obesity to Diabetes. *Nature* (2001) 409:307–12. doi: 10.1038/35053000
- 108. Wang H, Chu WS, Hemphill C, Elbein SC. Human Resistin Gene: Molecular Scanning and Evaluation of Association With Insulin Sensitivity and Type 2 Diabetes in Caucasians. J Clin Endocrinol Metab (2002) 87:2520–4. doi: 10.1210/jcem.87.6.8528
- 109. Yang R-Z, Huang Q, Xu A, McLenithan JC, Eisen JA, Shuldiner AR, et al. Comparative Studies of Resistin Expression and Phylogenomics in Human and Mouse. *Biochem Biophys Res Commun* (2003) 310:927–35. doi: 10.1016/ j.bbrc.2003.09.093
- 110. Patel L, Buckels AC, Kinghorn IJ, Murdock PR, Holbrook JD, Plumpton C, et al. Resistin is Expressed in Human Macrophages and Directly Regulated by PPAR Gamma Activators. *Biochem Biophys Res Commun* (2003) 300:472–6. doi: 10.1016/S0006-291X(02)02841-3
- 111. Reverchon M, Cornuau M, Ramé C, Guerif F, Royère D, Dupont J. Resistin Decreases Insulin-Like Growth Factor I-Induced Steroid Production and Insulin-Like Growth Factor I Receptor Signaling in Human Granulosa Cells. *Fertil Steril* (2013) 100(1):247–55.e1–3. doi: 10.1016/j.fertnstert.2013.03.008
- 112. Lee S, Lee HC, Kwon YW, Lee SE, Cho Y, Kim J, et al. Adenylyl Cyclase-Associated Protein 1 is a Receptor for Human Resistin and Mediates Inflammatory Actions of Human Monocytes. *Cell Metab* (2014) 19:484– 97. doi: 10.1016/j.cmet.2014.01.013
- 113. Benomar Y, Gertler A, De Lacy P, Crépin D, Ould Hamouda H, Riffault L, et al. Central Resistin Overexposure Induces Insulin Resistance Through Toll-Like Receptor 4. *Diabetes* (2013) 62:102–14. doi: 10.2337/db12-0237
- 114. Raeisi T, Rezaie H, Darand M, Taheri A, Garousi N, Razi B, et al. Circulating Resistin and Follistatin Levels in Obese and non-Obese Women With Polycystic Ovary Syndrome: A Systematic Review and Meta-Analysis. *PLoS One* (2021) 16: e0246200. doi: 10.1371/journal.pone.0246200
- 115. Yilmaz M, Bukan N, Demirci H, Oztürk C, Kan E, Ayvaz G, et al. Serum Resistin and Adiponectin Levels in Women With Polycystic Ovary Syndrome. *Gynecol Endocrinol* (2009) 25:246–52. doi: 10.1080/ 09513590802653833
- 116. Munir I, Yen H-W, Baruth T, Tarkowski R, Azziz R, Magoffin DA, et al. Resistin Stimulation of 17alpha-Hydroxylase Activity in Ovarian Theca Cells *In Vitro*: Relevance to Polycystic Ovary Syndrome. *J Clin Endocrinol Metab* (2005) 90:4852–7. doi: 10.1210/jc.2004-2152

- 117. Messini CI, Vasilaki A, Korona E, Anifandis G, Georgoulias P, Dafopoulos K, et al. Effect of Resistin on Estradiol and Progesterone Secretion From Human Luteinized Granulosa Cells in Culture. Syst Biol Reprod Med (2019) 65:350– 6. doi: 10.1080/19396368.2019.1615151
- Daquinag AC, Zhang Y, Amaya-Manzanares F, Simmons PJ, Kolonin MG. An Isoform of Decorin is a Resistin Receptor on the Surface of Adipose Progenitor Cells. Cell Stem Cell (2011) 9:74–86. doi: 10.1016/j.stem.2011.05.017
- 119. Herath S, Williams EJ, Lilly ST, Gilbert RO, Dobson H, Bryant CE, et al. Ovarian Follicular Cells Have Innate Immune Capabilities That Modulate Their Endocrine Function. *Reproduction* (2007) 134:683–93. doi: 10.1530/ REP-07-0229
- 120. Filková M, Haluzík M, Gay S, Senolt L. The Role of Resistin as a Regulator of Inflammation: Implications for Various Human Pathologies. *Clin Immunol* (*Orlando Fla.*) (2009) 133:157–70. doi: 10.1016/j.clim.2009.07.013
- 121. Silswal N, Singh AK, Aruna B, Mukhopadhyay S, Ghosh S, Ehtesham NZ. Human Resistin Stimulates the Pro-Inflammatory Cytokines TNF-Alpha and IL-12 in Macrophages by NF-Kappab-Dependent Pathway. *Biochem Biophys Res Commun* (2005) 334:1092–101. doi: 10.1016/j.bbrc.2005 .06.202
- 122. Samal B, Sun Y, Stearns G, Xie C, Suggs S, McNiece I. Cloning and Characterization of the Cdna Encoding a Novel Human Pre-B-Cell Colony-Enhancing Factor. *Mol Cell Biol* (1994) 14:1431–7. doi: 10.1128/ MCB.14.2.1431
- 123. Fukuhara A, Matsuda M, Nishizawa M, Segawa K, Tanaka M, Kishimoto K, et al. Visfatin: A Protein Secreted by Visceral Fat That Mimics the Effects of Insulin. Sci (New York N.Y.) (2005) 307:426–30. doi: 10.1126/ science.1097243
- 124. Revollo JR, Grimm AA, Imai S. the Regulation of Nicotinamide Adenine Dinucleotide Biosynthesis by Nampt/Pbef/Visfatin in Mammals. *Curr Opin Gastroenterol* (2007) 23:164–70. doi: 10.1097/MOG.0b013e32801b3c8f
- 125. Revollo JR, Grimm AA, Imai S. the Nad Biosynthesis Pathway Mediated by Nicotinamide Phosphoribosyltransferase Regulates Sir2 Activity in Mammalian Cells. J Biol Chem (2004) 279:50754–63. doi: 10.1074/ jbc.M408388200
- 126. Wang T, Zhang X, Bheda P, Revollo JR, Imai S, Wolberger C. Structure of Nampt/PBEF/Visfatin, a Mammalian NAD+ Biosynthetic Enzyme. Nat Struct Mol Biol (2006) 13:661–2. doi: 10.1038/nsmb1114
- 127. Reverchon M, Cornuau M, Cloix L, Ramé C, Guerif F, Royère D, et al. Visfatin is Expressed in Human Granulosa Cells: Regulation by Metformin Through AMPK/SIRT1 Pathways and its Role in Steroidogenesis. *Mol Hum Reprod* (2013) 19:313–26. doi: 10.1093/molehr/gat002
- 128. Shen CJ, Tsai EM, Lee J-N, Chen Y-L, Lee C-H, Chan T-F. The Concentrations of Visfatin in the Follicular Fluids of Women Undergoing Controlled Ovarian Stimulation are Correlated to the Number of Oocytes Retrieved. Fertil Steril (2010) 93:1844-50. doi: 10.1016/ j.fertnstert.2008.12.090
- 129. Reverchon M, Rame C, Bunel A, Chen W, Froment P, Dupont J. Visfatin (Nampt) Improves In Vitro IGF1-Induced Steroidogenesis and IGF1 Receptor Signaling Through SIRT1 in Bovine Granulosa Cells. Biol Reprod (2016) 94:54. doi: 10.1095/biolreprod.115.134650
- 130. Nejabati HR, Samadi N, Shahnazi V, Mihanfar A, Fattahi A, Latifi Z, et al. Nicotinamide and its Metabolite N1-Methylnicotinamide Alleviate Endocrine and Metabolic Abnormalities in Adipose and Ovarian Tissues in Rat Model of Polycystic Ovary Syndrome. *Chemico-biol Interact* (2020) 324:109093. doi: 10.1016/j.cbi.2020.109093
- 131. Annie L, Gurusubramanian G, Roy VK. Inhibition of Visfatin by FK866 Mitigates Pathogenesis of Cystic Ovary in Letrozole-Induced Hyperandrogenised Mice. *Life Sci* (2021) 276:119409. doi: 10.1016/ j.lfs.2021.119409
- 132. Tsouma I, Kouskouni E, Demeridou S, Boutsikou M, Hassiakos D, Chasiakou A, et al. Correlation of Visfatin Levels and Lipoprotein Lipid Profiles in Women With Polycystic Ovary Syndrome Undergoing Ovarian Stimulation. *Gynecol Endocrinol* (2014) 30:516–9. doi: 10.3109/ 09513590.2014.896896
- 133. Kowalska I, Straczkowski M, Nikolajuk A, Adamska A, Karczewska-Kupczewska M, Otziomek E, et al. Serum Visfatin in Relation to Insulin Resistance and Markers of Hyperandrogenism in Lean and Obese Women

With Polycystic Ovary Syndrome. Hum Reprod (2007) 22:1824–9. doi: 10.1093/ humrep/dem118

- 134. Kim JJ, Choi YM, Hong MA, Kim MJ, Chae SJ, Kim SM, et al. Serum Visfatin Levels in non-Obese Women With Polycystic Ovary Syndrome and Matched Controls. Obstet Gynecol Sci (2018) 61:253–60. doi: 10.5468/ ogs.2018.61.2.253
- 135. Hida K, Wada J, Eguchi J, Zhang H, Baba M, Seida A, et al. Visceral Adipose Tissue-Derived Serine Protease Inhibitor: A Unique Insulin-Sensitizing Adipocytokine in Obesity. Proc Natl Acad Sci U S A (2005) 102:10610–5. doi: 10.1073/pnas.0504703102
- 136. Zhou X, Chen Y, Tao Y, Zhang W, Xu W, Lu X. Serum Vaspin as a Predictor of Adverse Cardiac Events in Acute Myocardial Infarction. J Am Heart Assoc (2019) 8:e010934. doi: 10.1161/JAHA.118.010934
- 137. Nakatsuka A, Wada J, Iseda I, Teshigawara S, Higashio K, Murakami K, et al. Vaspin is an Adipokine Ameliorating ER Stress in Obesity as a Ligand for Cell-Surface GRP78/MTJ-1 Complex. *Diabetes* (2012) 61:2823–32. doi: 10.2337/db12-0232
- 138. Mehrabani S, Arab A, Karimi E, Nouri M, Mansourian M. Blood Circulating Levels of Adipokines in Polycystic Ovary Syndrome Patients: A Systematic Review and Meta-Analysis. *Reprod Sci (Thousand Oaks Calif)* (2021) 28 (11):3032–50. doi: 10.1007/s43032-021-00709-w
- 139. Dogan K, Helvacioglu C, Baghaki S, Ekin M. Comparison of Body Mass Index and Metabolic Parameters With Serum Vaspin Levels in Women With Polycystic Ovary Syndrome. *Diabetes Metab Syndrome* (2020) 14:137–9. doi: 10.1016/j.dsx.2020.01.008
- 140. Dogan K, Ekin M, Helvacioğlu Ç, Yaşar L. Can Serum Vaspin Levels Predict Clomiphene Resistance in Infertile Women With PCOS? Eur J Obstet Gynecol Reprod Biol (2017) 6–11 217. doi: 10.1016/j.ejogrb.2017.08.006
- 141. Tan BK, Heutling D, Chen J, Farhatullah S, Adya R, Keay SD, et al. Metformin Decreases the Adipokine Vaspin in Overweight Women With Polycystic Ovary Syndrome Concomitant With Improvement in Insulin Sensitivity and a Decrease in Insulin Resistance. *Diabetes* (2008) 57:1501–7. doi: 10.2337/db08-0127
- 142. Koiou E, Dinas K, Tziomalos K, Toulis K, Kandaraki EA, Kalaitzakis E, et al. The Phenotypes of Polycystic Ovary Syndrome Defined by the 1990 Diagnostic Criteria are Associated With Higher Serum Vaspin Levels Than the Phenotypes Introduced by the 2003 Criteria. Obes Facts (2011) 4:145–50. doi: 10.1159/000327935
- 143. Kohan L, Zarei A, Fallahi S, Tabiee O. Association Between Vaspin Rs2236242 Gene Polymorphism and Polycystic Ovary Syndrome Risk. *Gene* (2014) 539:209–12. doi: 10.1016/j.gene.2014.01.078
- 144. Bongrani A, Mellouk N, Ramé C, Cornuau M, Guerif F, Froment P, et al. Vaspin, a Novel Adipokine in Woman Granulosa Cells Physiology and PCOS Pathogenesis? J Endocrinol (2021) 249:57–70. doi: 10.1530/JOE-20-0550
- 145. Flower DR. The Lipocalin Protein Family: A Role in Cell Regulation. FEBS Lett (1994) 354(1):7–11. doi: 10.1016/0014-5793(94)01078-1
- 146. Yan QW, Yang Q, Mody N, Graham TE, Hsu CH, Xu Z, et al. The Adipokine Lipocalin 2 is Regulated by Obesity and Promotes Insulin Resistance. *Diabetes* (2007) 56:2533–40. doi: 10.2337/db07-0007
- 147. Hvidberg V, Jacobsen C, Strong R K, Cowland J B, Moestrup S K, Borregaard N. The Endocytic Receptor Megalin Binds the Iron Transporting Neutrophil-Gelatinase-Associated Lipocalin With High Affinity and Mediates its Cellular Uptake. FEBS Lett (2005) 579:773–7. doi: 10.1016/j.febslet.2004.12.031
- 148. Jaberi SA, Cohen A, D'Souza C, Abdulrazzaq YM, Ojha S, Bastaki S, et al. Lipocalin-2: Structure, Function, Distribution and Role in Metabolic Disorders. *Biomed Pharmacother = Biomed Pharmacother* (2021) 142:112002. doi: 10.1016/j.biopha.2021.112002
- 149. Wang Y, Lam KSL, Kraegen EW, Sweeney G, Zhang J, Tso AWK, et al. Lipocalin-2 is an Inflammatory Marker Closely Associated With Obesity, Insulin Resistance, and Hyperglycemia in Humans. *Clin Chem* (2007) 53:34– 41. doi: 10.1373/clinchem.2006.075614
- 150. Yilmaz Ö, Temur M, Calan M, Kume T, Özbay PÖ, Karakulak M, et al. The Relationship Between Lipocalin-2 and Free Testosterone Levels in Polycystic Ovary Syndrome. *Endokrynologia Polska* (2017) 68(1):7–12. doi: 10.5603/ EP.2017.0002

- 151. Cakal E, Ozkaya M, Engin-Ustun Y, Ustun Y. Serum Lipocalin-2 as an Insulin Resistance Marker in Patients With Polycystic Ovary Syndrome. *J Endocrinol Invest* (2011) 34(2):97–100. doi: 10.3275/7077
- 152. Gencer M, Gazi E, Hacıvelioğlu S, Binnetoğlu E, Barutçu A, Türkön H, et al. The Relationship Between Subclinical Cardiovascular Disease and Lipocalin-2 Levels in Women With PCOS. *Eur J Obstet Gynecol Reprod Biol* (2014) 181:99–103 doi: 10.1016/j.ejogrb.2014.07.032
- 153. Sahin SB, Cure MC, Ugurlu Y, Ergul E, Gur EU, Alyildiz N, et al. Epicardial Adipose Tissue Thickness and NGAL Levels in Women With Polycystic Ovary Syndrome. J Ovarian Res (2014) 7:24. doi: 10. 1186/1757-2215-7-24
- 154. Panidis D, Tziomalos K, Koiou E, Kandaraki EA, Tsourdi E, Delkos D, et al. The Effects of Obesity and Polycystic Ovary Syndrome on Serum Lipocalin-2 Levels: A Cross-Sectional Study. *Reprod Biol Endocrinol* (2010) 8:151. doi: 10.1186/1477-7827-8-151
- 155. Koiou E, Tziomalos K, Katsikis I, Kandaraki EA, Kalaitzakis E, Delkos D, et al. Weight Loss Significantly Reduces Serum Lipocalin-2 Levels in Overweight and Obese Women With Polycystic Ovary Syndrome. Gynecol Endocrinol (2012) 28:20–4. doi: 10.3109/09513590.2011. 588745
- 156. Martínez-García MÁ, Montes-Nieto R, Fernández-Durán E, Insenser M, Luque-Ramírez M, Escobar-Morreale HF. Evidence for Masculinization of Adipokine Gene Expression in Visceral and Subcutaneous Adipose Tissue of Obese Women With Polycystic Ovary Syndrome (PCOS). J Clin Endocrinol Metab (2013) 98:E388–96. doi: 10.1210/jc.2012-3414
- 157. Cheng F, Ng NYH, Tam CHT, Zhang Y, Lim CKP, Jiang G, et al. Association Between FGF19, FGF21 and Lipocalin-2, and Diabetes Progression in PCOS. *Endocr Connect* (2021) 10:1243–52. doi: 10.1530/ EC-21-0082
- 158. Diamanti-Kandarakis E, Livadas S, Kandarakis SA, Margeli A, Papassotiriou I. Serum Concentrations of Atherogenic Proteins Neutrophil Gelatinase-Associated Lipocalin and its Complex With Matrix Metalloproteinase-9 are Significantly Lower in Women With Polycystic Ovary Syndrome: Hint of a Protective Mechanism? *Eur J Endocrinol* (2008) 158:525–31. doi: 10.1530/EJE-07-0822
- 159. Schäffler A, Neumeier M, Herfarth H, Fürst A, Schölmerich J, Büchler C.. Genomic Structure of Human Omentin, a New Adipocytokine Expressed in Omental Adipose Tissue. *Biochim Biophys Acta* (2005) 1732 (1-3):96–102. doi: 10.1016/j.bbaexp.2005.11.005
- 160. Yang RZ, Lee M-J, Hu H, Pray J, Wu H-B, Hansen BC, et al. Identification of Omentin as a Novel Depot-Specific Adipokine in Human Adipose Tissue: Possible Role in Modulating Insulin Action. Am J Physiol Endocrinol Metab (2006) 290:E1253–61. doi: 10.1152/ajpendo.00572.2004
- 161. Khoshi A, Bajestani MK, Shakeri H, Goodarzi G, Azizi F. Association of Omentin Rs2274907 and FTO Rs9939609 Gene Polymorphisms With Insulin Resistance in Iranian Individuals With Newly Diagnosed Type 2 Diabetes. *Lipids Health Dis* (2019) 18:142. doi: 10.1186/s12944-019-1085-5
- 162. Franik G, Sadlocha M, Madej P, Owczarek A, Skrzypulec-Plinta V, Plinta R, et al. Circulating Omentin-1 Levels and Inflammation in Polycystic

Ovary Syndrome. Ginekologia Polska (2020) 91:308-12. doi: 10.5603/ GP.2020.0057

- 163. Tang Y-L, et al. Circulating Omentin-1 Levels in Women With Polycystic Ovary Syndrome: A Meta-Analysis. *Gynecol Endocrinol* (2017) 33:244–9. doi: 10.1080/09513590.2016.1254180
- 164. Yang HY, Ma Y, Lu X-H, Liang X-H, Suo Y-J, Huang Z-X, et al. The Correlation of Plasma Omentin-1 With Insulin Resistance in non-Obese Polycystic Ovary Syndrome. *Annales d'endocrinologie* (2015) 76:620–7. doi: 10.1016/j.ando.2015.08.002
- 165. Cloix L, Reverchon M, Cornuau M, Froment P, Ramé C, Costa C, et al. Expression and Regulation of INTELECTIN1 in Human Granulosa-Lutein Cells: Role in IGF-1-Induced Steroidogenesis Through NAMPT. *Biol Reprod* (2014) 91:50. doi: 10.1095/biolreprod.114.120410
- 166. Tan BK, Adya R, Farhatullah S, Lewandowski KC, O'Hare P, Lehnert H, et al. Omentin-1, a Novel Adipokine, is Decreased in Overweight Insulin-Resistant Women With Polycystic Ovary Syndrome: Ex Vivo and *In Vivo* Regulation of Omentin-1 by Insulin and Glucose. *Diabetes* (2008) 57:801–8. doi: 10.2337/db07-0990
- 167. Choi JH, Rhee E-J, Kim K-H, Woo H-Y, Lee W-Y, Sung K-C. Plasma Omentin-1 Levels are Reduced in non-Obese Women With Normal Glucose Tolerance and Polycystic Ovary Syndrome. *Eur J Endocrinol* (2011) 165:789– 96. doi: 10.1530/EJE-11-0375
- 168. Kenawi MZ, Akl EM, Sabry JH, Mostafa ST. Evaluation of Serum Level of Omentin-1 in Females With Hirsutism. J Cosmet Dermatol (2020) 19:535–9. doi: 10.1111/jocd.13043
- 169. Anithasri A, Ananthanarayanan PH, Veena P. A Study on Omentin-1 and Prostate Specific Antigen in Women on Treatment for Polycystic Ovary Syndrome. *Indian J Clin Biochem* (2019) 34:108–14. doi: 10.1007/s12291-017-0723-9
- 170. Tan BK, Adya R, Farhatullah S, Chen J, Lehnert H, Randeva HS. Metformin Treatment may Increase Omentin-1 Levels in Women With Polycystic Ovary Syndrome. *Diabetes* (2010) 59:3023–31. doi: 10.2337/ db10-0124

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Chen, Jia, Liu, Cao, Zhou and Zhao. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.