



Lactate: The Mediator of Metabolism and Immunosuppression

Yuanyuan Zhang¹, Zhao Zhai¹, Jiali Duan¹, Xiangcai Wang¹, Jinghua Zhong¹, Longqiu Wu¹, An Li¹, Miao Cao¹, Yanyang Wu¹, Huaqiu Shi^{1*}, Jianing Zhong^{2*} and Zhenli Guo^{1*}

¹ The First Affiliated Hospital of Gannan Medical University, Ganzhou, China, ² Key Laboratory of Prevention and Treatment of Cardiovascular and Cerebrovascular Diseases of Ministry of Education, Gannan Medical University, Ganzhou, China

OPEN ACCESS

Edited by:

Dragana Nikitovic, University of Crete, Greece

Reviewed by:

Monica Neagu, Victor Babes National Institute of Pathology (INCDBV), Romania

*Correspondence:

Huaqiu Shi
shq3677274@163.com
Jianing Zhong
zhongning_003@163.com
Zhenli Guo
wycg1984@163.com

Specialty section:

This article was submitted to Cancer Endocrinology, a section of the journal Frontiers in Endocrinology

Received: 22 March 2022

Accepted: 27 April 2022

Published: 09 June 2022

Citation:

Zhang Y, Zhai Z, Duan J, Wang X, Zhong J, Wu L, Li A, Cao M, Wu Y, Shi H, Zhong J and Guo Z (2022) Lactate: The Mediator of Metabolism and Immunosuppression. *Front. Endocrinol.* 13:901495. doi: 10.3389/fendo.2022.901495

The Warburg effect, one of the hallmarks of tumors, produces large amounts of lactate and generates an acidic tumor microenvironment *via* using glucose for glycolysis. As a metabolite, lactate not only serves as a substrate to provide energy for supporting cell growth and development but also acts as an important signal molecule to affect the biochemical functions of intracellular proteins and regulate the biological functions of different kinds of cells. Notably, histone lysine lactylation (Kla) is identified as a novel post-modification and carcinogenic signal, which provides the promising and potential therapeutic targets for tumors. Therefore, the metabolism and functional mechanism of lactate are becoming one of the hot fields in tumor research. Here, we review the production of lactate and its regulation on immunosuppressive cells, as well as the important role of Kla in hepatocellular carcinoma. Lactate and Kla supplement the knowledge gap in oncology and pave the way for exploring the mechanism of oncogenesis and therapeutic targets. Research is still needed in this field.

Keywords: the Warburg effect, lactate, histone lysine lactylation, metabolic reprogramming, immunosuppression

INTRODUCTION

The Warburg effect, proposed by Otto Warburg in the 1920s, suggests that cancer cells predominantly relied on increased glycolysis in the presence of oxygen, accompanied by the generation of lactate and the acidic microenvironment (1). This process is also known as aerobic glycolysis. It has been thought that it is only the cancer cells that consume lots of glucose, but accumulating evidence has found that immune cells, predominantly myeloid cells, also do (2, 3). What is more interesting is that these cells continue to proliferate and establish immunosuppressive networks in the presence of lactate to gain unlimited immune escape potential (4). Thus, lactate acts as a mediator between metabolic reprogramming and immunosuppression (5). It is worth noting that in 2019, Zhang et al. proposed histone lysine lactylation (Kla), a mechanism with global influence, which provides a new research direction for exploring carcinogenesis and effective therapeutic targets (6). The liver is an organ that constantly metabolizes glucose, fat, and amino acids according to the needs of the body. The changes of related enzymes and metabolites are valuable to be used to evaluate liver function and predict the development of cancer in clinic (7–9).

METABOLIC REPROGRAMMING TO PRODUCE LACTATE

Glucose and glutamine are important sources of energy for tumor cells and immune cells, with glucose predominating. Nutritional competition occurs between two types of cells. Tumor cells consume large amounts of both substances, resulting in immune cell starvation and decreased antitumor immunity (10).

The Warburg Effect

The end point of the Warburg effect is lactate regulated by many key enzymes. First, the glucose transporter (GLUT), which is expressed in most cells, is involved in transporting glucose into cells in metabolically active tissue. A variety of tumors highly express GLUT1 and have many regulatory factors (11). Hypoxia-inducible factor-1 α (HIF-1 α) and CD147 have positive relations with GLUT1, respectively (12, 13). MiR-455-5p also promotes the upregulation of GLUT1 by the IGF-1R/Akt/GLUT1 pathway (14). Long non-coding RNAs (lncRNAs), which were previously considered as “transcriptional noise” because of the non-protein-coding function, play a significant role in the development of tumors (15). For example, HOXA transcriptional antisense RNA (HOTAIR) increases the expression of GLUT1 by activating the mTOR signaling pathway (16). Upregulating the GLUT1 and expediting glucose absorption are only the first step in lactate production. Second, hexokinase (HK) handles the conversion of glucose to glucose-6-phosphate. There are four types of HK, HK1–4, of which HK2 trumps others in hepatocellular carcinoma. Caveolin1 (CAV-1), miR-199a-5p, and miR-125a take effect (17–20). HOTAIR is also one factor by adsorbing miR-130a-3p without oxygen (21). Glucokinase, an isoenzyme of HK, is required for migration of regulatory T cells (Treg) (22). There are few studies on HK2, and focusing on it may lead to special findings. Moreover, pyruvate kinase 2 (PKM2) is the final rate-limiting enzyme regulating pyruvate accumulation. The Lamc1/PTEN/AKT pathway causes an increase in PKM2 expression, as does the downregulation of miR-122 (23, 24). The interaction between PKM2, heat shock protein 90 (HSP90), and HIF-1 α is to stabilize PKM2 and induce aerobic glycolysis to inhibit cell apoptosis (25). In addition, lactate dehydrogenase (LDH), pyruvate dehydrogenase kinase (PDK), and pyruvate dehydrogenase (PDH) are the key enzymes that determine the destination of pyruvate. LDHA and LDHB encode LDH and form five types in different proportions: LDH1 (B4), LDH2 (AB3), LDH3 (A2B2), LDH4 (A3B), and LDH5 (A4) (26). Interestingly, LDH5 is more significantly expressed than other types in hepatocellular carcinoma. Thus, inhibition of LDHB transcription or induction of LDHA transcription is an essential mechanism that promotes the generation of lactate (27). Decreased miR-142-3p and N-myc downstream-regulated gene 2 (NDRG2) and elevated miR-34c-3p stimulate the upregulation of LDHA (28–31). The generated lactate continues to mediate the activity of PDK to phosphorylate PDH, causing a blocked entry of pyruvate into the tricarboxylic acid cycle (TAC) (27). On the one hand, it reduces the consumption of glucose by oxidative phosphorylation (OXPHOS), and on the other hand, it can cause

the accumulation of pyruvate and indirectly facilitate the production of lactate. Differently, it is FoxP3 in Treg cells that impairs glycolysis and promotes OXPHOS that confer migratory and inhibitory functions on Treg cells (32). Finally, the acidic environment also depends on the pH-dependent monocarboxylate transport system (MCT), a tool for lactate to shuttle between cells and the microenvironment. The flow of lactate into and out of cells relies on MCT1 and MCT4, respectively (33). The transfer of lactate disrupts extracellular pH homeostasis, which not only affects the activity of enzymes but also participates in regulating immune cells. Lactate can also act as a substrate to energize cells, which is referred to as the “reverse Warburg effect” (34). In this way, the role of lactate is too diverse and complex to be ignored. **Figure 1** shows the production and regulation mechanism of lactate in hepatocellular carcinoma cells.

Glutamine Metabolism

Glutamine metabolism has two main ways *in vivo*. On the one hand, it provides C–N (amide group) for the synthesis of protein, lipid, and nucleotide (35); on the other hand, metabolites such as aspartate and oxaloacetate can be converted to substances for glycolysis (36). Cancer cells contain high levels of glutaminase 1 (GLS1), which decomposes glutamine into glutamate. Targeting GLS1 can inhibit not only cell growth but also stemness characteristics (37–39). In addition, there was a possible inhibitory relationship between glutamine metabolism and glycolysis. However, glucose is not severely limited because of the presence of resident cells (such as immune cells and endothelial cells), which can enhance glucose intake (2). More unexpectedly, glutamine metabolism also promotes cell growth

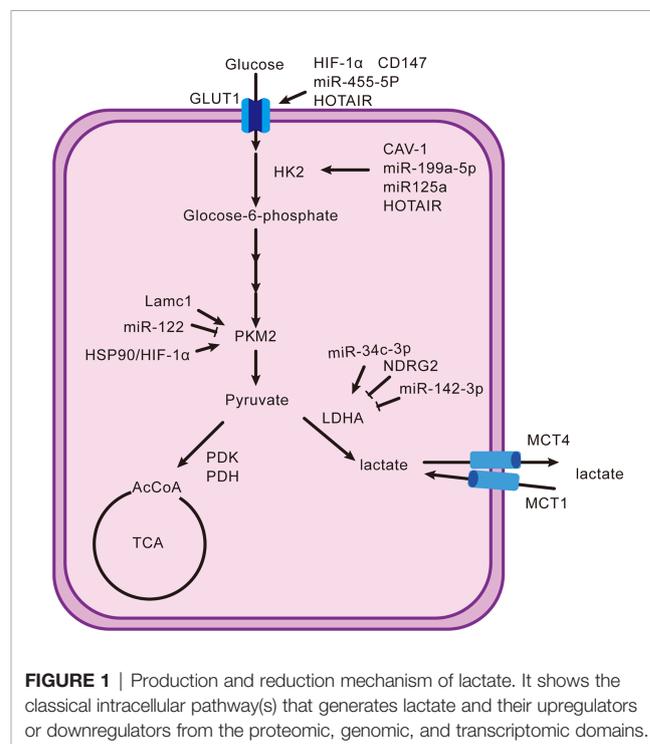


FIGURE 1 | Production and reduction mechanism of lactate. It shows the classical intracellular pathway(s) that generates lactate and their upregulators or downregulators from the proteomic, genomic, and transcriptomic domains.

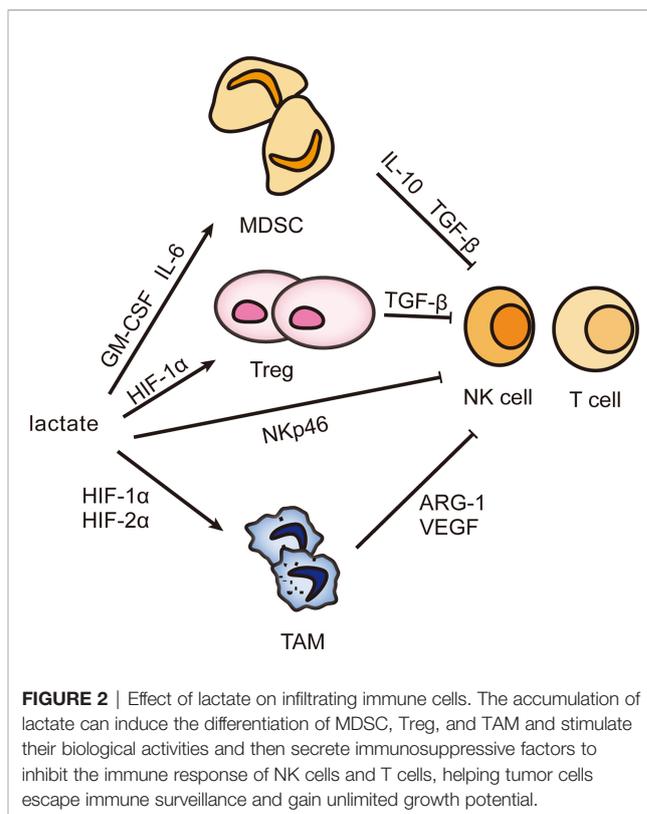
and suppresses immunity because of lactate (40). Therefore, targeting glutamine is a meaningful therapeutic strategy that deserves to explore more mechanisms.

LACTATE FACILITATES THE ASSEMBLY OF IMMUNE ESCAPE NETWORK

The liver possesses unique immune properties containing large populations of NK cells, cytotoxic CD8⁺ T cells, and others, but the incidence of hepatocellular carcinoma remains high, most possibly because an external environment suppresses immunity (41). Lactate can facilitate the assembly of an immune escape network to make the proliferation of tumor cells uncontrolled (42, 43). Lactate-mediated immune escape networks are shown in **Figure 2**.

Myeloid-Derived Suppressor Cells

Myeloid-derived suppressor cells (MDSCs) comprise a group of heterogeneous cells derived from immature bone marrow. MDSCs are the precursor of mature monocytes, dendritic cells, and granulocytes. However, they are an important assistant to favor tumor proliferation and inhibit immune response (44, 45). Multiple cytokines prevent MDSCs from differentiating and maturing, then MDSCs expand and migrate. Lactate is one (Table 1 shows the mechanism of other factors) (55–57). M6A demethylase ALKBH5 ensures the m6A methylation level and RNA stability of MCT4, followed by lactate transporting out of cells with the help of MCT4 and stimulating MDSC increase (58). The knockdown of LDHA reconfirms the role of lactate with reduced MDSCs and weakened cytolytic function of NK cells (59). How does lactate assume this responsibility? Lactate induces the accumulation of MDSCs through granulocyte-macrophage colony-stimulating factor (GM-CSF) and IL-6. Besides, lactate inhibits the expression of activated receptor NKp46 in NK cells (59). This shows that lactate can directly and indirectly inhibit the antitumor activity, and thus the more vigorous the tumor metabolism, the poorer the immune function. More interestingly, MDSCs are necessary for tumor-associated macrophage (TAM) accumulation. When MDSCs migrate to the tumor site, its CD45 phosphatase activity increases because of hypoxia and local acidification, which inhibits STAT3 and thus promotes the conversion of MDSCs to TAMs (60). In addition, lactate-induced HIF1 α can also promote the differentiation of MDSCs into TAMs by regulating the expression of inducible nitric oxide synthase (iNOS) and arginase-1 (ARG1), which is helpful to inhibit adaptive immunity (61–63). In summary, lactate



is involved in multiple pathways to suppress the antitumor immunity, so could lactate be a target for tumor treatment? During radiotherapy for pancreatic cancer, lactate plays an important role in enhancing the immunosuppressive phenotype of MDSC through the G-protein-coupled receptor 81 (GPR81)/mTOR/HIF-1 α /STAT3 pathway. Therefore, targeting lactate may provide a unique prospect for improving the radiosensitivity of pancreatic cancer (64).

Tumor-Associated Macrophage

In the 1980s, the fact that monocytes differentiated into macrophages under the recruitment of chemokines was confirmed (65). Solid tumors exhibit hypoxic areas within the tumor mass. Macrophages are attracted to these hypoxic tumor sites by various chemodynamic stimuli secreted by tumor cells under hypoxic pressure, and their movement is directly impaired, causing tumor-associated macrophage (TAM) to become trapped at the site of the ischemic tumor (66, 67). “Activated macrophages”

TABLE 1 | Other factors stimulate MDSC accumulation in hepatocellular carcinoma.

Molecule	Primary mechanism of action	Reference
HSC	Activation of the COX2/PGE2/EP4 and SDF-1/CXCR4 signaling pathways	(46, 47)
TAF	Activation of the SDF-1 α /CXCR4 and IL-6/STAT3 pathways	(48)
AMPK	Suppression of the JAK/STAT, NF- κ B, C-EBP β , and CHOP signaling pathways	(49)
HIF-1 α	Conversion of extracellular ATP to 5'-AMP through its direct transcriptional target, ENTPD2, and promotion of the interaction between CCL26 and CX3CR1	(50, 51)
PIWIL1	Induction of P38MAPK signaling	(52)
PKM2	Activation of PI3K-AKT and JNK signaling pathways and upregulation of HIF-1 α expression	(53)
RIP3	Promotion of the interaction between CXCL1 and the cognate receptor CXCR2	(54)

induce vascular proliferation, and “non-activated macrophages” do not (68). Now these two states have been named selectively activated macrophages (M2) and classically activated macrophages (M1), respectively. TAMs are broadly heterogeneous, and their functional status depends on the conditions of exposure (69). For example, Toll-like receptor agonists, interferon- γ (IFN- γ), and lipopolysaccharide (LPS) promote the phenotype of M1, while IL-4, IL-13, IL-10, transforming growth factor- β (TGF- β), and lactate are inducers of M2 (70–72).

Lactate promotes M2 by stabilizing the expression of HIF-1 α , and in this process ARG1 and vascular endothelial growth factor (VEGF) are increased, which are closely related to wound healing, angiogenesis, and tissue remodeling (73). In addition, HIF-2 α is also implicated. Lactate negatively regulates transcription factor EB (TFEB) by activating mTORC1 to reduce the expression of ATP6V0d2, which mediates the degradation of HIF-2 α by lysosomes, ensuring the activated state of M2 (74). Epigenetics also contributes to an important mechanism. Histone lysine lactylation (Kla) has been shown to promote M2 activation, but the exact mechanism is not well understood (6). M2 secretes IL-10 and TGF- β in response to lactate stimulation to weaken the anticancer ability of NK cells and lymphocytes (73). Thus, it is established that lactate affects immune function through M2 and lactate could be a therapeutic target. Machilin A inhibits cell growth and M2 polarization by decreasing lactate production (75). When macrophages were cultivated in acidic pH conditions, HIF-1 α , ARG1, and VEGF were found increased. So is it lactate or acidic conditions that are required to promote tumor development? This deserves further study and exploration.

Regulatory T Cells

Lactate also induces aggregation of regulatory T cell (Treg), the phenotype of CD4+CD25+FoxP3+ T cell. Its metabolic pathway varies with activity and function (13, 76). First, the high-speed glycolysis of Treg driven by the mTOR signal will not only metabolically restrict antitumor cells such as T lymphocytes, which is called T-cell starvation, but provide energy for Treg proliferation and migration from the thymus to the tumor periphery (10, 22). In addition, lactate-induced HIF-1 α activates the CCL20/CCR6 axis by inducing myeloid trigger receptor-1 (TREM-1) expression in TAM, attracting aggregation and initiating immunosuppressive effects of Treg (77). Most unexpectedly, Treg's activity requires a switch from glycolysis to OXPHOS, when forehead box P3 (FoxP3) comes into play. It not only inhibits glycolysis by suppressing the oncogene Myc but also regulates LDH to promote the conversion of lactate to pyruvate and even drives lipid oxidation metabolism to promote OXPHOS, which endows Treg with inhibitory activity (32, 78, 79). A stable expression of FoxP3 is associated with epigenetic modifications, particularly methylation. Promoters and CpG islands are the main methylation sites of FoxP3. The methylation level is negatively correlated with the proportion of Treg, which means that the lower the methylation level of FoxP3, the higher the proportion of Treg cells and the higher the tumor malignancy (80). TGF- β can induce FoxP3 demethylation (81). Treg performs glycolysis, OXPHOS, and fat metabolism. Different activities and functions utilize different metabolisms, which adds some complexity to Treg.

HISTONE LYSINE LACTYLATION

In 2019, a new epigenetic modification of gene transcription, histone lysine lactylation (Kla), was proposed, which is based on lactate and lysine (6). This finding highlights how lactate has an advanced effect on tumor progression, and provides a mechanism by which some pathways of angiogenesis, migration, and metastasis may be activated. Kla fills an important gap in our understanding of various physiopathologies (such as cancer) that are closely related to lactate. Using M1 macrophage polarization as a model, it was confirmed that Kla regulated gene expression. Because of a low level of ARG1, M1 macrophages metabolize arginine through nitric oxide synthase (NOS) to produce nitric oxide (NO) to kill pathogens, while M2 macrophages have a higher level of ARG1, which produces ornithine to promote wound healing (82). After inhibiting LDH, the yield of lactate and the degree of Kla decreased, including the Kla markers on ARG1 promoters. Then M1 macrophages were treated with exogenous lactate to find that the global Kla and ARG1 expression increased (6). These conclusions have also been confirmed in melanoma and lung cancer (6). It scientifically and rigorously verified the positive role of lactate and Kla in inducing the polarization of M1 macrophage to M2 macrophage.

Kla can cause carcinogenic signals and represent a clear target for tumor therapy. Convincing evidence suggests that Kla promotes the transcription of YTHN6-methyladenosine RNA-binding protein 2 (YTHDF2). YTHDF2 recognizes the M⁶A modification sites on PER1 and TP53 and promotes their degradation, thus promoting the occurrence of melanoma (83). This study revealed the cancer-promoting mechanism of Kla, confirmed the interaction between Kla and M⁶A methylation for the first time, and provided a new therapeutic target to treat melanoma. In addition, Kla has also been found to regulate gene expression in non-small cell lung cancer (NSCLC). After treatment of NSCLC cells with lactate, the degree of Kla and HIF-1 α increases and the transcription of HK-1, glucose 6-phosphate dehydrogenase (G6PD), and PKM decreases, which jointly regulates the metabolism of cancer cells (84). This further confirms the important role of Kla in cancer.

CONCLUSION

To sum up, lactate is the product of aerobic glycolysis. It is used as an energy source for many kinds of cells, including tumor cells, and as a signal molecule to damage the immune response directly or indirectly. Kla, a kind of posttranslational modification, can regulate gene expression. Reducing the concentration of lactate and Kla modification contributes to restoring the physiological function of the body, so what promising strategies can achieve this effect? One is to block the production pathway of lactate by inhibiting the key enzymes of glycolysis and glutaminolysis, and the other is to block the transport of lactate (85–87). Ideally, lactate concentration and anticancer immunity return to the physiological state (88). However, in view of the diversity of enzymes, it is critical to find an enzyme that can regulate lactate to the maximum extent. In addition, how to solve the damage to normal cells is also a big problem. Therefore, targeting lactate and Kla is a frontier treatment, but there is a long way to go.

AUTHOR CONTRIBUTIONS

YZ: original draft preparation. ZZ, JD, MC, and YW: software, figure, and table. XW, JZ, LW, and AL: revising the manuscript critically. HS, JZ, and ZG: reviewing and editing. All authors contributed to the article and approved the submitted version.

REFERENCES

- Warburg O, Wind F, Negelein E. The Metabolism of Tumors in the Body. *J Gen Physiol* (1927) 8:519–30. doi: 10.1085/jgp.8.6.519
- Reinfeld BI, Madden MZ, Wolf MM, Chytil A, Bader JE, Patterson AR, et al. Cell-Programmed Nutrient Partitioning in the Tumour Microenvironment. *Nature* (2021) 7858: 282–8. doi: 10.1038/s41586-021-03442-1
- Kelly B, O'Neill LA. Metabolic Reprogramming in Macrophages and Dendritic Cells in Innate Immunity. *Cell Res* (2015) 25:771–84. doi: 10.1038/cr.2015.68
- Ganapathy-Kanniappan S. Linking Tumor Glycolysis and Immune Evasion in Cancer: Emerging Concepts and Therapeutic Opportunities. *Biochim Biophys Acta Rev Cancer* (2017) 1868:212–20. doi: 10.1016/j.bbcan.2017.04.002
- Neagu M. Metabolic Traits in Cutaneous Melanoma. *Front Oncol* (2020) 10:851. doi: 10.3389/fonc.2020.00851
- Zhang D, Tang Z, Huang H, Zhou G, Cui C, Weng Y, et al. Metabolic Regulation of Gene Expression by Histone Lactylation. *Nature* (2019) 574:575–80. doi: 10.1038/s41586-019-1678-1
- De Matteis S, Ragusa A, Marisi G, De Domenico S, Casadei Gardini A, Bonafè M, et al. Aberrant Metabolism in Hepatocellular Carcinoma Provides Diagnostic and Therapeutic Opportunities. *Oxid Med Cell Longev* (2018) 2018:7512159. doi: 10.1155/2018/7512159
- Rui L. Energy Metabolism in the Liver. *Compr Physiol* (2014) 4:177–97. doi: 10.1002/cphy.c130024
- Feng Y, Xiong Y, Qiao T, Li X, Jia L, Han Y. Lactate Dehydrogenase A: A Key Player in Carcinogenesis and Potential Target in Cancer Therapy. *Cancer Med* (2018) 7:6124–36. doi: 10.1002/cam4.1820
- Rivadeneira DB, Delgoffe GM. Antitumor T-Cell Reconditioning: Improving Metabolic Fitness for Optimal Cancer Immunotherapy. *Clin Cancer Res* (2018) 24:2473–81. doi: 10.1158/1078-0432.Ccr-17-0894
- Mueckler M, Thorens B. The SLC2 (GLUT) Family of Membrane Transporters. *Mol Aspects Med* (2013) 34:121–38. doi: 10.1016/j.mam.2012.07.001
- Amann T, Maegdefrau U, Hartmann A, Agaimy A, Marienhagen J, Weiss TS, et al. GLUT1 Expression is Increased in Hepatocellular Carcinoma and Promotes Tumorigenesis. *Am J Pathol* (2009) 174:1544–52. doi: 10.2353/ajpath.2009.080596
- Li X, Zhang Y, Ma W, Fu Q, Liu J, Yin G, et al. Enhanced Glucose Metabolism Mediated by CD147 Contributes to Immunosuppression in Hepatocellular Carcinoma. *Cancer Immunol Immunother* (2020) 69:535–48. doi: 10.1007/s00262-019-02457-y
- Hu Y, Yang Z, Bao D, Ni JS, Lou J. miR-455-5p Suppresses Hepatocellular Carcinoma Cell Growth and Invasion via IGF-1r/AKT/GLUT1 Pathway by Targeting IGF-1r. *Pathol Res Pract* (2019) 215:152674. doi: 10.1016/j.prp.2019.152674
- Kopp F, Mendell JT. Functional Classification and Experimental Dissection of Long Noncoding RNAs. *Cell* (2018) 172:393–407. doi: 10.1016/j.cell.2018.01.011
- Wei S, Fan Q, Yang L, Zhang X, Ma Y, Zong Z, et al. Promotion of Glycolysis by HOTAIR Through GLUT1 Upregulation via mTOR Signaling. *Oncol Rep* (2017) 38:1902–8. doi: 10.3892/or.2017.5840
- Patra KC, Wang Q, Bhaskar PT, Miller L, Wang Z, Wheaton W, et al. Hexokinase 2 is Required for Tumor Initiation and Maintenance and its Systemic Deletion is Therapeutic in Mouse Models of Cancer. *Cancer Cell* (2013) 24:213–28. doi: 10.1016/j.ccr.2013.06.014
- Chai F, Li Y, Liu K, Li Q, Sun H. Caveolin Enhances Hepatocellular Carcinoma Cell Metabolism, Migration, and Invasion *In Vitro* via a Hexokinase 2-Dependent Mechanism. *J Cell Physiol* (2019) 234:1937–46. doi: 10.1002/jcp.27074
- Guo W, Qiu Z, Wang Z, Wang Q, Tan N, Chen T, et al. MiR-199a-5p is Negatively Associated With Malignancies and Regulates Glycolysis and Lactate Production by Targeting Hexokinase 2 in Liver Cancer. *Hepatology* (2015) 62:1132–44. doi: 10.1002/hep.27929
- Jin F, Wang Y, Zhu Y, Li S, Liu Y, Chen C, et al. The miR-125a/HK2 Axis Regulates Cancer Cell Energy Metabolism Reprogramming in Hepatocellular Carcinoma. *Sci Rep* (2017) 7:3089. doi: 10.1038/s41598-017-03407-3
- Hu M, Fu Q, Jing C, Zhang X, Qin T, Pan Y. LncRNA HOTAIR Knockdown Inhibits Glycolysis by Regulating miR-130a-3p/HIF1A in Hepatocellular Carcinoma Under Hypoxia. *BioMed Pharmacother* (2020) 125:109703. doi: 10.1016/j.biopha.2019.109703
- Kishore M, Cheung KCP, Fu H, Bonacina F, Wang G, Coe D, et al. Regulatory T Cell Migration Is Dependent on Glucokinase-Mediated Glycolysis. *Immunity* (2017) 47:875–889.e810. doi: 10.1016/j.immuni.2017.10.017
- Ye G, Qin Y, Wang S, Pan D, Xu S, Wu C, et al. Lamc1 Promotes the Warburg Effect in Hepatocellular Carcinoma Cells by Regulating PKM2 Expression Through AKT Pathway. *Cancer Biol Ther* (2019) 20:711–9. doi: 10.1080/15384047.2018.1564558
- Liu AM, Xu Z, Shek FH, Shek FH, Wong KF, Lee NP, et al. miR-122 Targets Pyruvate Kinase M2 and Affects Metabolism of Hepatocellular Carcinoma. *PLoS One* (2014) 9:e86872. doi: 10.1371/journal.pone.0086872
- Feng J, Wu L, Ji J, Chen K, Yu Q, Zhang J, et al. PKM2 is the Target of Proanthocyanidin B2 During the Inhibition of Hepatocellular Carcinoma. *J Exp Clin Cancer Res* (2019) 38:204. doi: 10.1186/s13046-019-1194-z
- Drent M, Cobben NA, Henderson RF, Wouters EF, van Dieijen-Visser M. Usefulness of Lactate Dehydrogenase and its Isoenzymes as Indicators of Lung Damage or Inflammation. *Eur Respir J* (1996) 9:1736–42. doi: 10.1183/09031936.96.09081736
- Hong SM, Lee YK, Park I, Kwon SM, Min S, Yoon G. Lactic Acidosis Caused by Repressed Lactate Dehydrogenase Subunit B Expression Down-Regulates Mitochondrial Oxidative Phosphorylation via the Pyruvate Dehydrogenase (PDH)-PDH Kinase Axis. *J Biol Chem* (2019) 294:7810–20. doi: 10.1074/jbc.RA118.006095
- Hua S, Liu C, Liu L, Wu D. miR-142-3p Inhibits Aerobic Glycolysis and Cell Proliferation in Hepatocellular Carcinoma via Targeting LDHA. *Biochem Biophys Res Commun* (2018) 496:947–54. doi: 10.1016/j.bbrc.2018.01.112
- Guo Y, Li X, Sun X, Wang J, Yang X, Zhou X, et al. Combined Aberrant Expression of NDRG2 and LDHA Predicts Hepatocellular Carcinoma Prognosis and Mediates the Anti-Tumor Effect of Gemcitabine. *Int J Biol Sci* (2019) 15:1771–86. doi: 10.7150/ijbs.35094
- Weng Q, Chen M, Yang W, Li J, Fan K, Xu M, et al. Integrated Analyses Identify miR-34c-3p/MAGI3 Axis for the Warburg Metabolism in Hepatocellular Carcinoma. *FASEB J* (2020) 34:5420–34. doi: 10.1096/fj.201902895R
- Sheng SL, Liu JJ, Dai YH, Sun XG, Xiong XP, Huang G. Knockdown of Lactate Dehydrogenase A Suppresses Tumor Growth and Metastasis of Human Hepatocellular Carcinoma. *FEBS J* (2012) 279:3898–910. doi: 10.1111/j.1742-4658.2012.08748.x
- Angelin A, Gil-de-Gómez L, Dahiya S, Jiao J, Guo L, Levine MH, et al. Foxp3 Reprograms T Cell Metabolism to Function in Low-Glucose, High-Lactate Environments. *Cell Metab* (2017) 25:1282–1293.e1287. doi: 10.1016/j.cmet.2016.12.018
- Zhao Y, Li W, Li M, Hu Y, Zhang H, Song G, et al. Targeted Inhibition of MCT4 Disrupts Intracellular pH Homeostasis and Confers Self-Regulated Apoptosis on Hepatocellular Carcinoma. *Exp Cell Res* (2019) 384:111591. doi: 10.1016/j.yexcr.2019.111591

FUNDING

This project was supported by the Major Program of the National Natural Science Foundation of China (No. 82060363) and Science and Technology Plan of Jiangxi Provincial Health and Health Commission (SKJP220203570).

34. Liu HT, Huang DA, Li MM, Liu HD, Guo K. HSF1: A Mediator in Metabolic Alteration of Hepatocellular Carcinoma Cells in Cross-Talking With Tumor-Associated Macrophages. *Am J Transl Res* (2019) 11:5054–64.
35. Zhang J, Pavlova NN, Thompson CB. Cancer Cell Metabolism: The Essential Role of the Nonessential Amino Acid, Glutamine. *EMBO J* (2017) 36:1302–15. doi: 10.15252/embj.201696151
36. Yang L, Venneti S, Nagrath D. Glutaminolysis: A Hallmark of Cancer Metabolism. *Annu Rev BioMed Eng* (2017) 19:163–94. doi: 10.1146/annurev-bioeng-071516-044546
37. Yang L, Moss T, Mangala LS, Marini J, Zhao H, Wahlig S, et al. Metabolic Shifts Toward Glutamine Regulate Tumor Growth, Invasion and Bioenergetics in Ovarian Cancer. *Mol Syst Biol* (2014) 10:728. doi: 10.1002/msb.20134892
38. Xiang Y, Stine ZE, Xia J, Lu Y, O'Connor RS, Altman BJ, et al. Targeted Inhibition of Tumor-Specific Glutaminase Diminishes Cell-Autonomous Tumorigenesis. *J Clin Invest* (2015) 125:2293–306. doi: 10.1172/jci75836
39. Li B, Cao Y, Meng G, Qian L, Xu T, Yan C, et al. Targeting Glutaminase 1 Attenuates Stemness Properties in Hepatocellular Carcinoma by Increasing Reactive Oxygen Species and Suppressing Wnt/beta-Catenin Pathway. *EBioMedicine* (2019) 39:239–54. doi: 10.1016/j.ebiom.2018.11.063
40. Leone RD, Zhao L, Englert JM, Sun IM, Oh MH, Sun IH, et al. Glutamine Blockade Induces Divergent Metabolic Programs to Overcome Tumor Immune Evasion. *Science* (2019) 366:1013–21. doi: 10.1126/science.aav2588
41. Piñero Fernández J, Luddy KA, Harmon C, O'Farrelly C. Hepatic Tumor Microenvironments and Effects on NK Cell Phenotype and Function. *Int J Mol Sci* (2019) 20(17):4131. doi: 10.3390/ijms20174131
42. Scott KE, Cleveland JL. Lactate Wreaks Havoc on Tumor-Infiltrating T and NK Cells. *Cell Metab* (2016) 24:649–50. doi: 10.1016/j.cmet.2016.10.015
43. Brand A, Singer K, Koehl GE, Kolitzus M, Schoenhammer G, Thiel A, et al. LDHA-Associated Lactic Acid Production Blunts Tumor Immunosurveillance by T and NK Cells. *Cell Metab* (2016) 24:657–71. doi: 10.1016/j.cmet.2016.08.011
44. Gabrilovich DI, Nagaraj S. Myeloid-Derived Suppressor Cells as Regulators of the Immune System. *Nat Rev Immunol* (2009) 9:162–74. doi: 10.1038/nri2506
45. Kohanbash G, Okada H. Myeloid-Derived Suppressor Cells (MDSCs) in Gliomas and Glioma-Development. *Immunol Invest* (2012) 41:658–79. doi: 10.3109/08820139.2012.689591
46. Xu Y, Zhao W, Xu J, Li J, Hong Z, Yin Z, et al. Activated Hepatic Stellate Cells Promote Liver Cancer by Induction of Myeloid-Derived Suppressor Cells Through Cyclooxygenase-2. *Oncotarget* (2016) 7:8866–78. doi: 10.18632/oncotarget.6839
47. Xu Y, Fang F, Jiao H, Zheng X, Huang L, Yi X, et al. Activated Hepatic Stellate Cells Regulate MDSC Migration Through the SDF-1/CXCR4 Axis in an Orthotopic Mouse Model of Hepatocellular Carcinoma. *Cancer Immunol Immunother* (2019) 68:1959–69. doi: 10.1007/s00262-019-02414-9
48. Deng Y, Cheng J, Fu B, Liu W, Chen G, Zhang Q, et al. Hepatic Carcinoma-Associated Fibroblasts Enhance Immune Suppression by Facilitating the Generation of Myeloid-Derived Suppressor Cells. *Oncogene* (2017) 36:1090–101. doi: 10.1038/ncr.2016.273
49. Salminen A, Kauppinen A, Kaarniranta K. AMPK Activation Inhibits the Functions of Myeloid-Derived Suppressor Cells (MDSC): Impact on Cancer and Aging. *J Mol Med (Berl)* (2019) 97:1049–64. doi: 10.1007/s00109-019-01795-9
50. Chiu DK, Tse AP, Xu IM, Di Cui J, Lai RK, Li LL, et al. Hypoxia Inducible Factor HIF-1 Promotes Myeloid-Derived Suppressor Cells Accumulation Through ENTPD2/CD39L1 in Hepatocellular Carcinoma. *Nat Commun* (2017) 8:517. doi: 10.1038/s41467-017-00530-7
51. Chiu DK, Xu IM, Lai RK, Tse AP, Wei LL, Koh HY, et al. Hypoxia Induces Myeloid-Derived Suppressor Cell Recruitment to Hepatocellular Carcinoma Through Chemokine (C-C Motif) Ligand 26. *Hepatology* (2016) 64:797–813. doi: 10.1002/hep.28655
52. Wang N, Tan HY, Lu Y, Chan YT, Wang D, Guo W, et al. PIWIL1 Governs the Crosstalk of Cancer Cell Metabolism and Immunosuppressive Microenvironment in Hepatocellular Carcinoma. *Signal Transduct Target Ther* (2021) 6:86. doi: 10.1038/s41392-021-00485-8
53. Liu WR, Tian MX, Yang LX, Lin YL, Jin L, Ding ZB, et al. PKM2 Promotes Metastasis by Recruiting Myeloid-Derived Suppressor Cells and Indicates Poor Prognosis for Hepatocellular Carcinoma. *Oncotarget* (2015) 6:846–61. doi: 10.18632/oncotarget.2749
54. Li YM, Liu ZY, Wang JC, Yu JM, Li ZC, Yang HJ, et al. Receptor-Interacting Protein Kinase 3 Deficiency Recruits Myeloid-Derived Suppressor Cells to Hepatocellular Carcinoma Through the Chemokine (C-X-C Motif) Ligand 1-Chemokine (C-X-C Motif) Receptor 2 Axis. *Hepatology* (2019) 70:1564–81. doi: 10.1002/hep.30676
55. Tian X, Shen H, Li Z, Wang T, Wang S. Tumor-Derived Exosomes, Myeloid-Derived Suppressor Cells, and Tumor Microenvironment. *J Hematol Oncol* (2019) 12:84. doi: 10.1186/s13045-019-0772-z
56. Veglia F, Perego M, Gabrilovich D. Myeloid-Derived Suppressor Cells Coming of Age. *Nat Immunol* (2018) 19:108–19. doi: 10.1038/s41590-017-0022-x
57. Tian J, Rui K, Tang X, Ma J, Wang Y, Tian X, et al. MicroRNA-9 Regulates the Differentiation and Function of Myeloid-Derived Suppressor Cells via Targeting Runx1. *J Immunol* (2015) 195:1301–11. doi: 10.4049/jimmunol.1500209
58. Li N, Kang Y, Wang L, Huff S, Tang R, Hui H, et al. ALKBH5 Regulates Anti-PD-1 Therapy Response by Modulating Lactate and Suppressive Immune Cell Accumulation in Tumor Microenvironment. *Proc Natl Acad Sci U S A* (2020) 117:20159–70. doi: 10.1073/pnas.1918986117
59. Husain Z, Huang Y, Seth P, Sukhatme VP. Tumor-Derived Lactate Modifies Antitumor Immune Response: Effect on Myeloid-Derived Suppressor Cells and NK Cells. *J Immunol* (2013) 191:1486–95. doi: 10.4049/jimmunol.1202702
60. Kumar V, Cheng P, Condamine T, Mony S, Languino LR, McCaffrey JC, et al. CD45 Phosphatase Inhibits STAT3 Transcription Factor Activity in Myeloid Cells and Promotes Tumor-Associated Macrophage Differentiation. *Immunity* (2016) 44:303–15. doi: 10.1016/j.immuni.2016.01.014
61. Corzo CA, Condamine T, Lu L, Cotter MJ, Youn JI, Cheng P, et al. HIF-1 α Regulates Function and Differentiation of Myeloid-Derived Suppressor Cells in the Tumor Microenvironment. *J Exp Med* (2010) 207:2439–53. doi: 10.1084/jem.20100587
62. Sica A, Bronte V. Altered Macrophage Differentiation and Immune Dysfunction in Tumor Development. *J Clin Invest* (2007) 117:1155–66. doi: 10.1172/jci31422
63. Melillo G, Musso T, Sica A, Taylor LS, Cox GW, Varesio L. A Hypoxia-Responsive Element Mediates a Novel Pathway of Activation of the Inducible Nitric Oxide Synthase Promoter. *J Exp Med* (1995) 182:1683–93. doi: 10.1084/jem.182.6.1683
64. Yang X, Lu Y, Hang J, Zhang J, Zhang T, Huo Y, et al. Lactate-Modulated Immunosuppression of Myeloid-Derived Suppressor Cells Contributes to the Radioresistance of Pancreatic Cancer. *Cancer Immunol Res* (2020) 8:1440–51. doi: 10.1158/2326-6066.Cir-20-0111
65. Bottazzi B, Polentarutti N, Balsari A, Boraschi D, Ghezzi P, Salmons M, et al. Chemotactic Activity for Mononuclear Phagocytes of Culture Supernatants From Murine and Human Tumor Cells: Evidence for a Role in the Regulation of the Macrophage Content of Neoplastic Tissues. *Int J Cancer* (1983) 31:55–63. doi: 10.1002/ijc.2910310110
66. Grimshaw MJ, Balkwill FR. Inhibition of Monocyte and Macrophage Chemotaxis by Hypoxia and Inflammation—a Potential Mechanism. *Eur J Immunol* (2001) 31:480–9. doi: 10.1002/1521-4141(200102)31:2<480::aid-immu480>3.0.co;2-I
67. Kes MMG, Van den Bossche J, Griffioen AW, Huijbers EJM. Oncometabolites Lactate and Succinate Drive Pro-Angiogenic Macrophage Response in Tumors. *Biochim Biophys Acta Rev Cancer* (2020) 1874:188427. doi: 10.1016/j.bbcan.2020.188427
68. Polverini PJ, Cotran PS, Gimbrone MA Jr., Unanue ER. Activated Macrophages Induce Vascular Proliferation. *Nature* (1977) 269:804–6. doi: 10.1038/269804a0
69. Xue J, Schmidt SV, Sander J, Draffehn A, Krebs W, Quester I, et al. Transcriptome-Based Network Analysis Reveals a Spectrum Model of Human Macrophage Activation. *Immunity* (2014) 40:274–88. doi: 10.1016/j.immuni.2014.01.006
70. Locati M, Mantovani A, Sica A. Macrophage Activation and Polarization as an Adaptive Component of Innate Immunity. *Adv Immunol* (2013) 120:163–84. doi: 10.1016/b978-0-12-417028-5.00006-5

71. Martinez FO, Gordon S, Locati M, Mantovani A. Transcriptional Profiling of the Human Monocyte-to-Macrophage Differentiation and Polarization: New Molecules and Patterns of Gene Expression. *J Immunol* (2006) 177:7303–11. doi: 10.4049/jimmunol.177.10.7303
72. Sica A, Mantovani A. Macrophage Plasticity and Polarization: *In Vivo* Veritas. *J Clin Invest* (2012) 122:787–95. doi: 10.1172/jci59643
73. Colegio OR, Chu NQ, Szabo AL, Chu T, Rhebergen AM, Jairam V, et al. Functional Polarization of Tumour-Associated Macrophages by Tumour-Derived Lactic Acid. *Nature* (2014) 513:559–63. doi: 10.1038/nature13490
74. Liu N, Luo J, Kuang D, Xu S, Duan Y, Xia Y, et al. Lactate Inhibits ATP6V0d2 Expression in Tumor-Associated Macrophages to Promote HIF-2 α -Mediated Tumor Progression. *J Clin Invest* (2019) 129:631–46. doi: 10.1172/jci123027
75. Chung TW, Kim EY, Han CW, Park SY, Jeong MS, Yoon D, et al. Machilin A Inhibits Tumor Growth and Macrophage M2 Polarization Through the Reduction of Lactic Acid. *Cancers (Basel)* (2019) 11(7):963. doi: 10.3390/cancers11070963
76. Unitt E, Rushbrook SM, Marshall A, Davies S, Gibbs P, Morris LS, et al. Compromised Lymphocytes Infiltrate Hepatocellular Carcinoma: The Role of T-Regulatory Cells. *Hepatology* (2005) 41:722–30. doi: 10.1002/hep.20644
77. Wu Q, Zhou W, Yin S, Zhou Y, Chen T, Qian J, et al. Blocking Triggering Receptor Expressed on Myeloid Cells-1-Positive Tumor-Associated Macrophages Induced by Hypoxia Reverses Immunosuppression and Anti-Programmed Cell Death Ligand 1 Resistance in Liver Cancer. *Hepatology* (2019) 70:198–214. doi: 10.1002/hep.30593
78. Gerriets VA, Kishton RJ, Johnson MO, Cohen S, Siska PJ, Nichols AG, et al. Foxp3 and Toll-Like Receptor Signaling Balance T(reg) Cell Anabolic Metabolism for Suppression. *Nat Immunol* (2016) 17:1459–66. doi: 10.1038/ni.3577
79. Wang R, Dillon CP, Shi LZ, Milasta S, Carter R, Finkelstein D, et al. The Transcription Factor Myc Controls Metabolic Reprogramming Upon T Lymphocyte Activation. *Immunity* (2011) 35:871–82. doi: 10.1016/j.immuni.2011.09.021
80. Liu Q, Du F, Huang W, Ding X, Wang Z, Yan F, et al. Epigenetic Control of Foxp3 in Intratumoral T-Cells Regulates Growth of Hepatocellular Carcinoma. *Aging (Albany NY)* (2019) 11:2343–51. doi: 10.18632/aging.101918
81. Kim HP, Leonard WJ. CREB/ATF-Dependent T Cell Receptor-Induced FoxP3 Gene Expression: A Role for DNA Methylation. *J Exp Med* (2007) 204:1543–51. doi: 10.1084/jem.20070109
82. Rath M, Müller I, Kropf P, Closs EI, Munder M. Metabolism via Arginase or Nitric Oxide Synthase: Two Competing Arginine Pathways in Macrophages. *Front Immunol* (2014) 5:532. doi: 10.3389/fimmu.2014.00532
83. Yu J, Chai P, Xie M, Ge S, Ruan J, Fan X, et al. Histone Lactylation Drives Oncogenesis by Facilitating M(6)A Reader Protein YTHDF2 Expression in Ocular Melanoma. *Genome Biol* (2021) 22:85. doi: 10.1186/s13059-021-02308-z
84. Dichtl S, Lindenthal L, Zeitler L, Behnke K, Schlösser D, Strobl B, et al. Lactate and IL6 Define Separable Paths of Inflammatory Metabolic Adaptation. *Sci Adv* (2021) 7(26):eabg3505. doi: 10.1126/sciadv.abg3505
85. Sanità P, Capulli M, Teti A, Galatioto GP, Vicentini C, Chiarugi P, et al. Tumor-Stroma Metabolic Relationship Based on Lactate Shuttle can Sustain Prostate Cancer Progression. *BMC Cancer* (2014) 14:154. doi: 10.1186/1471-2407-14-154
86. Choi SYC, Ettinger SL, Lin D, Xue H, Ci X, Nabavi N, et al. Targeting MCT4 to Reduce Lactic Acid Secretion and Glycolysis for Treatment of Neuroendocrine Prostate Cancer. *Cancer Med* (2018) 7:3385–92. doi: 10.1002/cam4.1587
87. Choi SY, Xue H, Wu R, Fazli L, Lin D, Collins CC, et al. The MCT4 Gene: A Novel, Potential Target for Therapy of Advanced Prostate Cancer. *Clin Cancer Res* (2016) 22:2721–33. doi: 10.1158/1078-0432.Ccr-15-1624
88. Wang JX, Choi SYC, Niu X, Kang N, Xue H, Killam J, et al. Lactic Acid and an Acidic Tumor Microenvironment Suppress Anticancer Immunity. *Int J Mol Sci* (2020) 21(21):8363. doi: 10.3390/ijms21218363

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Zhang, Zhai, Duan, Wang, Zhong, Wu, Li, Cao, Wu, Shi, Zhong and Guo. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

GLOSSARY

Kla	histone lysine lactylation
GLUT	glucose transporter
HIF-1a	hypoxia-inducible factor-1
lncRNAs	long non-coding RNAs
HOTAIR	HOX transcriptional antisense RNA
HK	hexokinase
CAV-1	Caveolin1
Treg	regulatory T cells
PKM2	pyruvate kinase 2
HSP90	heat shock protein 90
LDH	lactate dehydrogenase
PDK	pyruvate dehydrogenase kinase
PDH	pyruvate dehydrogenase
NDRG2	N-myc downstream regulated gene 2
TAC	tricarboxylic acid cycle
OXPPOS	oxidative phosphorylation
MCT	pH-dependent monocarboxylate transport system
GLS1	glutaminase1
MDSC	myeloid-derived suppressor cell
GM-CSF	granulocyte-macrophage colony-stimulating factor
iNOS	inducible nitric oxide synthase
ARG1	arginase-1
GPR81	G-protein-coupled receptor 81
TAM	tumor-associated macrophage
M2	selectively activated macrophages
M1	classically activated macrophages
IFN-g	interferon-g
LPS	lipopolysaccharide
TGF-b	transforming growth factor-b
VEGF	vascular endothelial growth factor
TFEB	transcription factor EB
TREM-1	myeloid trigger receptor-1
FoxP3	forehead box P3
NOS	nitric oxide synthase
NO	nitric oxide
YTHDF2	YTHN6-methyladenosine RNA-binding protein 2
NSCLC	non-small cell lung cancer
G6PD	glucose 6-phosphate dehydrogenase
