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SPECIALTY SECTION

This article was submitted to Gut Endocrinology, a section of the journal Frontiers in Endocrinology

RECEIVED 23 January 2023 ACCEPTED 14 February 2023 PUBLISHED 01 March 2023

CITATION

Li D, Tang W, Wang Y, Gao Q, Zhang H, Zhang Y, Wang Y, Yang Y, Zhou Y, Zhang Y, Li H, Li S and Zhao H (2023) An overview of traditional Chinese medicine affecting gut microbiota in obesity. *Front. Endocrinol.* 14:1149751. doi: 10.3389/fendo.2023.1149751

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An overview of traditional Chinese medicine affecting gut microbiota in obesity

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Obesity, a chronic metabolic disease with a complex pathophysiology, is caused by several variables. High-fat diets lead to the disruption of the gut microbiota and impaired gut barrier function in obese people. The dysbiosis and its metabolites through the intestinal barrier lead to an imbalance in energy metabolism and inflammatory response, which eventually contributes to the development of chronic diseases such as diabetes, hypertension, and cardiovascular disease. Current medicines are therapeutic to obesity in the short term; however, they may bring significant physical and emotional problems to patients as major side effects. Therefore, it is urgent to explore new therapeutic methods that have definite efficacy, can be taken for a long time, and have mild adverse effects. Numerous studies have demonstrated that traditional Chinese medicine (TCM) can control the gut microbiota in a multitargeted and comprehensive manner, thereby restoring flora homeostasis, repairing damaged intestinal mucosal barriers, and eventually curbing the development of obesity. The active ingredients and compounds of TCM can restore the normal physiological function of the intestinal mucosal barrier by regulating gut microbiota to regulate energy metabolism, inhibit fat accumulation, affect food appetite, and reduce intestinal mucosal inflammatory response, thereby effectively promoting weight loss and providing new strategies for obesity prevention and treatment. Although there are some studies on the regulation of gut microbiota by TCM to prevent and treat obesity, all of them have the disadvantage of being systematic and comprehensive. Therefore, this work comprehensively describes the molecular mechanism of obesity mediated by gut microbiota based on the research state of obesity, gut microbiota, and TCM. A comprehensive and systematic summary of TCM targeting the regulation of gut microbiota for the treatment of obesity should be conducted in order to provide new strategies and ideas for the treatment of obesity.

KEYWORDS

traditional Chinese medicine, gut microbiota, obesity, short-chain fatty acids (SCFAs), active ingredients

1 Introduction

Obesity is a chronic metabolic disease that is caused by excessive accumulation and abnormal distribution of body fat due to the imbalance of energy intake and consumption (1). Research shows that obesity has become a worldwide public health problem. More than 2 billion adults are overweight or obese worldwide, and the prevalence of obesity continues to rise globally (2, 3). Obesity will increase the risk of heart disease, diabetes, digestive disorders, and several types of cancer (4). At present, there are two main types of drugs that have been clinically proven to help with weight loss. One type is the central nervous system drugs, such as cloxacillin, but these drugs have side effects including insomnia, constipation, and anxiety (5). Another type of drug works on the non-central nervous system, such as orlistat, which has the same side effects (insomnia, constipation, and anxiety) (6). According to modern medicine, the pathogenesis of obesity is complex and involves dietary habits, genetic factors, and environmental factors (7). An increasing number of studies have shown that gut microbiota, as a key environmental factor, could contribute to the occurrence and development of obesity (8).

The gut microbiota refers to the microorganisms inhabiting the human gastrointestinal tract. The gut microbiota consists of many interacting symbiotic bacteria, which is considered to be an endocrine organ involved in maintaining energy homeostasis and host immunity. The bioactive metabolites of the gut microbiota can affect the physiological effects of the host (9–12). In humans, the gut microbiota is a complex and dynamic ecosystem that represents approximately 1 kg of our body weight (13). Under normal conditions, the gut microbiota works in harmony with the host and participates in the regulation of many physiological functions of the host, such as nutrient and substance metabolism, food digestion and absorption, the formation of a biological barrier in the intestinal mucosal epithelium, and boosting host immune function (14, 15). Furthermore, healthy gut microbiota produces short-chain fatty acids (SCFAs) to repair and promote intestinal function, and to effectively inhibit the growth of spoiled bacteria in the gut and improve the intestinal environment (16). However, dietary habits or environmental factors affect the composition of gut microbiota, and long-term consumption of improper diet may affect metabolism (17). Several studies have found that antibiotics can change the intestinal mucosa bacterial composition, reduce colonization resistance, and damage the intestinal mucosal barrier by reducing the abundance of gut microbiota (18–20). The high-fat and high-protein diet altered the community structure of lactase bacteria in the intestinal mucosa and decreased the abundance of the critical lactase bacteria (21). The dysbiosis of the gut microbiota may be involved in the pathogenesis of obesity through multiple mechanisms, including disruption of energy homeostasis, lipid synthesis, and storage, central regulation of appetite and feeding behavior, and chronic low-grade inflammation (13, 22, 23). Therefore, regulation of gut microbiota and improvement of intestinal dysbiosis are regarded as key directions for the treatment of obesity, as shown in Figure 1.

Traditional Chinese medicine (TCM) has thousands of years of clinical experience in the treatment of obesity, and its benefits are safe, mild, and long-lasting. The majority of TCM is taken orally, and the active ingredients in the TCM interact with the gut microbiota as they enter the gastrointestinal tract. The growth and proliferation of certain microbiota can be stimulated or inhibited by TCM. For example, the Wu et al. study found that Lilium lancifolium can promote the growth of Lactobacillus spp. and Bifidobacteria spp., and inhibit the growth of total bacteria in the intestines of normal mice (24). Qiweibaizhu powder could overcome the influence of dysbacteriosis and could lead to the recovery of intestinal mucosal microbiota homeostasis (25). Folium senna decoction gavage increased intestinal microbiota diversity; Bacteroides vulgatus, Helicobacter ganmani, Lactobacillus murinus, Microbacterium dextranolyticum, and Klebsiella pneumoniae were significantly enriched, while Candidatus arthromitus sp. and Lactobacillus johnsonii were significantly depleted (26, 27). Qiweibaizhu powder has a positive effect on the recovery of



bacterial lactase gene diversity to normal levels and increases the abundance of *Lysobacter* and *Eukaryota* (28). The Qiwei Baizhu Powder decoction and QWBZP-TG promoted the proliferation of *Lactobacillus* and inhibited the growth of *Proteus*, *Clostridium*, *Eubacterium*, *Facklamia*, and *Escherichia* (29). The active ingredients in the Gegenqinlian decoction may increase the microbial activity of the intestinal mucosa of mice and reduce the microbial activity of the intestinal contents through multiple targets (30). This review will focus on recent advances in the connection between gut microbiota and obesity, and the role of gut microbiota in the treatment of obesity is clarified, in order to find new ideas and research directions for the treatment of obesity and related diseases.

2 Mechanisms of obesity mediation by gut microbiota

The gut microbiota colonizes the intestines and has important physiological functions. Studies have shown that obesity is associated with a decrease in gut microbiota diversity, gene expression of gut microbiota, and alteration of metabolic pathways (31, 32). Compared with healthy people, the gut microbiota of obese people usually shows changes at the phylum, family, and genus levels (33, 34). The imbalance of gut microbiota may lead to obesity, which may be related to energy balance, regulation of feeding behavior, fat storage, and inflammation. The mechanisms of gut microbiota-mediated obesity are discussed in this review.

2.1 Characteristics of obese gut microbiota

The first evidence for a link between gut microbiota and obesity was from studies of germ-free mice. In 1983, Wostmann et al. first discovered that germ-free rats needed more energy to maintain their body weight compared to wild rats, but the precise mechanism was unknown at the time (35). In 2004, Gordon et al. observed that gut microbiota can affect energy absorption from the diet and energy storage in mice, and found that the proportion of *Firmicutes* was higher and the proportion of *Bacteroidetes* was lower in the guts of obese people (36). Numerous studies have demonstrated that the structure and abundance of gut microbiota are altered in obese people compared to healthy people. Obese people show lower gut microbiota diversity than healthy people and show significant overall obesity and dyslipidemia (37–39).

Numerous studies have demonstrated that the meat-based Western diet has a negative impact on the richness and function of gut microbiota. Being high in animal-derived protein and low in vegetables and fruits, the Western-style diet leads to a significant decrease in the numbers of total bacteria and commensal *Bifidobacterium* and *Eubacterium* species (40). A diet rich in fat, with high consumption of red meat and refined carbohydrates, may have a direct effect on the immune system causing a structural and functional modification of the gut microbiota (41). Lard and vegetable blend oil diet affected the composition of the intestinal

microorganisms and the functions of digestive enzymes (42). It was found that more Firmicutes than Bacteroidetes caused the body to absorb more calories from food and more easily convert calories into fat to accumulate under the skin (43, 44). The gut of obese people is rich in archaea, which can oxidize hydrogen produced by Prevotella, thus accelerating the fermentation of polysaccharides and causing the body to absorb more energy (45). According to Petersen et al. (46), Clostridium was reduced in obese people and some of the Clostridium were able to absorb fat and reduce obesity in mice. The levels of Ackermanella are significantly decreased in obese people, which reduces metabolic disturbances caused by highfat diets, improves fat accumulation, and reduces inflammatory responses (47). In summary, the gut microbiota changes in obese people are mainly shown as the increase in bacteria that cause inflammation and fat production and the decrease in bacteria that inhibit obesity and inflammation. Therefore, it can play a role in the treatment of obesity by regulating the diversity and abundance of the dominant gut microbiota, inhibiting adipogenesis, and the inflammatory response.

2.2 Regulation of energy absorption

Gut microbiota participates in energy metabolism and absorption under normal and pathological conditions. The obesity-associated gut microbiome can increase the capacity for energy harvest from the diet, thus contributing to increased adipose tissue storage of the host, especially the white adipose tissue (WAT) (48). SCFAs, mainly acetate, propionate, and butyrate, are produced by the gut microbiota through the metabolism of complex dietary plant polysaccharides (49, 50). As one of the primary metabolites of the intestinal microbiota, SCFAs can lower intestinal pH to a certain extent, thereby inhibiting the viability of pathogenic microbiota from promoting intestinal microecological balance and maintaining the integrity of the intestinal mucosal barrier (51). SCFAs are an important source of energy for the host, contributing up to 70% of the daily energy supply for herbivores and 10% of the total daily energy needed for omnivores (13). Among them, butyric acid is a significant source of energy for the epithelial cells of the human colon and cecum (52). As depicted in Figure 2, SCFAs not only provide direct energy but also raise energy expenditure and encourage lipolysis. Acetate can have a beneficial effect on host energy metabolism by reducing levels of pro-inflammatory cytokines, increasing energy expenditure and lipid oxidation, and promoting lipolysis (53). Butyrate promotes fatty acid oxidation and increases energy consumption by acting on G protein-coupled receptor 43 (GPR43) and upregulating the expression of peroxisome proliferator-activated receptor coactivator-1 alpha (PGC-1 α) and uncoupling protein 1 (UCP-1) expression in brown adipose tissue (BAT) (54). SCFAs are typically considered to have beneficial effects on the body. However, some research suggests that excessive SCFAs may have a negative impact on the body. Clinical studies by Tirosh et al. (55) showed that consuming a propionate-containing diet causes the levels of glucagon in the blood, which raises the risk of obesity. Therefore, gut microbiota metabolites can have both positive and negative effects on the host.



If we want to regulate the energy absorption of the body by regulating the metabolites of the intestinal flora, the concentration should be taken into consideration.

2.3 Regulation of feeding behavior

The brain is the main part of the appetite regulation neural pathway, which directly regulates feeding behavior. Gut microbiota produces specific metabolic compounds, which interact with the central nervous system, such as glutamic acid, bile acids, 5-hydroxytryptamine (5-HT), SCFAs, and GABA (Glu) (Figure 3) (56, 57). These neuroendocrine components and their receptors have a role in controlling central appetite and feeding behavior by mediating the bidirectional communication between the brain and

the digestive system (58). Through the "brain-gut-bacteria" axis, SCFAs can control the body's energy metabolism to control the GPRs pathway, activate the MAPK signal pathway in intestinal epithelial cells, promote the secretion of 5-HT and gastrointestinal peptides (GIPs) in the intestine, increase satiety, decrease gastric emptying and intestinal movement, affect energy intake, and decrease body weight (53). Propionate in SCFAs activates GPR41, increases leptin (LP) levels, reduces the content of neuropeptide Y (NPY), which has the effect of enhancing appetite, and inhibits the secretion of peptide YY (PYY), which has the effect of appetite (59, 60). Butyrate reduces obesity by acting on GPR43 to increase plasma GLP-1 levels, controlling appetite through the central nervous system, and reducing food intake (60, 61). In addition, oral butyric acid also inhibits the hypothalamic expression of NPY and pro-appetitive neural activity, and decreases food intake (62).



According to Frost et al. (63), acetate can reduce hunger by increasing the amount of GABA in mice's brains. Therefore, the production of SCFAs can be controlled by regulating the structure and activity of the intestinal flora to achieve obesity treatment.

2.4 Regulation of fat accumulation

The regulation of genes related to fat accumulation is greatly influenced by intestinal flora and its metabolites (Figure 4) (64). Fasting-induced adipokines (FIAFs) inhibit adipose tissue storage by inhibiting lipoprotein lipase (LPL) activity, reducing extracellular triglyceride hydrolysis and fatty acid uptake (65, 66). Studies have shown that SCFAs can inhibit fat accumulation in the body, which may be related to their ability to activate the expression of FIAF and inhibit the expression of LPL, thus preventing the triglyceride cycle (67). In addition, acetate also prevents fat accumulation by stimulating GLP-1 production and activating GPR43 in the gut (68). Propionate regulates glucose and lipid metabolism by binding to and activating neuronal GPRs, causing the release of vasoactive intestinal peptide (VIP) from the submucosal plexus, activating adenylate cyclase (AC), and reducing cAMP levels (69).

2.5 The chronic mild inflammatory response

At present, obesity and obesity-related diseases are recognized as systemic chronic low inflammation by the medical community. Recent research has demonstrated that Gram-negative bacteria in the gut microbiota of obese people can create lipopolysaccharides (LPS) and bind to toll-like receptor 4 (TLR-4), activating nearby and distant pro-inflammatory cascades and releasing inflammatory factors (70, 71). TLR4 is an important binding regulator of LPS, which is overexpressed in intestinal mucosal epithelial cells after stimulation of activation, inducing the production of various proinflammatory factors and causing and maintaining obesity-type low-level inflammation in the body (72). In addition, damage to the intestinal mucosal barrier can also lead to inflammation. The changes in intestinal flora increase intestinal permeability and promote LPS to enter the blood to activate the NF- κ B signal pathway, which leads to chronic inflammation, endotoxemia, and increased fat accumulation, and ultimately leads to metabolic syndrome and obesity (73). The high-fat diet could induce the interactions between *Thermoactinomyces*, *Staphylococcus*, and intestinal inflammation (74).

Gut microbiota and its metabolites mediate the chronic inflammatory response and inhibit obesity (Figure 5). The main mechanisms are as follows: (1) Promote the proliferation of intestinal epithelial cells, reduce the apoptosis of intestinal epithelial cells, and maintain the intestinal mucosal barrier. (2) Regulation of inflammation-related gene expression and reduction of inflammatory response. SCFAs are important signaling molecules to regulate intestinal mucosal immunity, inhibiting LPS or TNF-α-induced inflammatory responses, in which the mechanism is related to the regulation of NF-KB and MAPK signaling pathways (75). The study also discovered that additional SCFA supplementation could enhance mitochondrial activity in brown fat cells, inhibit chronic inflammation, increase the expression of G protein-coupled receptors (GPR43 and GPR41), promote the oxidation of free fatty acids and the hydrolysis of triglycerides, and decrease the body weight of obese mice (76). SCFAs inhibit the transport of toxic substances by maintaining the stability of the intestinal barrier, reducing the concentration of LPS in the blood, and reducing the inflammatory response (77). Acetate improves epithelial cell-mediated intestinal defense and enhances intestinal integrity, thereby protecting the host from lethal infections (78). Butyrate inhibits the activity of macrophages, dendritic cells, neutrophils, and T cells, which lessens inflammation of the intestinal tissue (79, 80). In addition, butyric acid can also block the activity of the transcription factor NF-B and decrease the expression of inflammatory factors like IL-6 and IL-12, which inhibits the inflammatory response in the intestine (81).

In conclusion, the development of obesity is closely correlated with gut microbiota. Human obesity and several numbers of other disorders are brought about by gut microbial dysbiosis. Healthy





intestinal flora and its metabolites can play a potential role in preventing or treating obesity by regulating energy absorption, feeding behavior, fat accumulation, and inflammatory reaction.

3 The mechanism of TCM affecting gut microbiota to inhibit obesity

The role of regulation of gut microbiota by TCM in the treatment of obesity and related diseases has been confirmed. Several studies have found that the TCM compound, single Chinese medicine, and its active ingredients act on gut microbiota and its metabolites, and play a role in controlling energy metabolism absorption, feeding behavior, and fat storage, and in assuaging chronic delicate inflammation; therefore, it can be used in the treatment of obesity and other related diseases, as shown in Table 1.

3.1 Study on the mechanism of active ingredients of TCM affecting gut microbiota to inhibit obesity

3.1.1 Regulation of energy metabolism and absorption

The anti-obesity effect of TCM and its active ingredients is closely related to the regulation of gut microbiota. The active ingredients of TCM can affect the absorption of energy metabolism in the body by improving the abundance of gut microbiota and the expression of related signal pathway proteins. Zhang et al. (82) found that ginsenosides, the active ingredient of ginseng, improved the structural composition of the gut microbiota, increased the concentration of SCFAs and receptor proteins (GPR41, GPR43, and GPR109A), and enhanced the gut barrier completeness. Song et al. (83) found that the root of *Atractylodes macrocephala* Koidzumi extract can prevent diet-induced obesity and glucose intolerance in mice. Its mechanisms are related to increasing the number of brown fat cells, increasing PGC-1 α and

UCP1 expression, and promoting the energy metabolism of brown fat tissue. Propolis ethanol extract reduced high-fat diet-induced obesity by regulating the abundance of gut microbiota and improving glucose tolerance and lipid distribution in high-fat diet rats (84). According to Wang et al. (85), Polygala tenuifolia extract inhibits high-fat diet-induced lipid accumulation in obese mice by modulating the composition of gut microbiota and regulating the levels of transcription factors of lipid oxidation. Li et al. (106) found that Dendrobium officinale can improve the diversity of intestinal mucosal flora and influence the gut microbiota to positively affect high-fat diet-induced negative effects in mice. Asparagus can affect the diversity of bacteria in the intestinal mucosa of mice fed a highfat diet, and achieve a lipid-lowering effect by regulating the intestinal microecology of mice fed a high-fat diet (107). Baohe pills can restore the number of intestinal flora to a certain extent and improve the activities of various digestive enzymes including protease and amylase in mice fed a high-fat and high-protein diet (108).

3.1.2 Regulation of feeding behavior

The active components of TCM can improve the abundance of gut microbiota and the expression of associated signaling pathway protein in obese mice to regulate feeding behavior and thus play a role in treating obesity. The active components of TCM can alter feeding behavior, increase microbiota abundance, and consequently increase the production of proteins involved in linked signaling pathways in obese mice. According to Neyrinck et al. (93), berberine and curcumin can improve intestinal barrier function, increase the number of intestinal probiotics, upregulate the expression of the innate immunity genes Pla2g2a and PYY, and decrease food intake in obese mice. Casanova-Martí et al. (94) discovered that grape seed proanthocyanidins can suppress appetite and promote weight reduction by controlling the quantity of gut microbiota in rats, raising serum GLP-1 levels, and upregulating GLP-1 mRNA and protein expression in ileum and colonic tissues. According to Lin et al. (95), diet-induced obese mice's appetites were suppressed and their weight was decreased because Rb1, the active component of

TABLE 1 Study on the intervention mechanism of TCM affecting gut microbiota in obesity.

Mechanism	Chinese medicine and formula	Target/ medium	Reference
The expression of genes that control energy absorption such as GPR43, TC, and TG	Ginsenoside	SCFAs, GPR41, GPR43, GPR109A	(82)
	Root of Atractylodes macrocephala Koidzumi	PGC-1α, UCP1	(83)
	Extract of propolis	IL-1β, IL-6, IL-10	(84)
	Polygala tenuifolia extract	SREBP1C, TG, SAA1	(85)
	Cordyceps guangdongensis	SCFAs, TC, TG, ALT	(86)
	Lingguizhugan formula	TC, TG, FFA	(87–89)
	Erchen decoction	IRS1, AKT, PKA, HSL	(9 0)
	Xiexin tang	SCFAs, ACK, BUT, PGC-1α, AMPK	(91)
	Shenlian decoction	/	(92)
The levels of PYY and GLP-1 increase, improve diversity of gut microbiota and the intestinal barrier function, and decrease food intake	Berberine Curcumin	Pla2g2a, PYY, Muc2	(93)
	Grape seed proanthocyanidins	GLP-1	(94)
	Ginsenoside	PYY, NPY, Y2 receptor	(95)
Increases the level of SCFA increase, regulates the abundance of gut microbiota, improves disorders of lipid metabolism, and inhibits fat accumulation	Ganoderma amboinense polysaccharide	SCFAs, TC, TG	(96)
	Rutin	IWAT, UCP1, SCFAs, BUT	(97)
	Heat-treated adzuki	SCFAs	(98)
Inhibits pathways leading to the production of pro-inflammatory cytokines; reduces LPS, thereby reducing chronic low-grade inflammation	Rhizoma Atractylodis Macrocephalae	HDL, IL-6, TNF-α	(99)
	Berberine	CD14, IL-1, IL-6, TNF-α	(100)
	Polysaccharide from the sclerotium of Poria cocos	PPAR-γ	(101)
	Polysaccharide from sporoderm-broken spore of <i>Ganoderma lucidum via</i> gut microbiota regulation	SCFAs, GPR43, GPR41, IL-1β	(102)
	Polysaccharide from Sijunzi decoction	SCFAs	(103)
	Effects of shenling baizhu	SCFAs, TLR4, IL- 1, TNF-α	(104)
	JinQi Jiangtang	SCFAs, MCP-1, IL-6, TNF-α	(105)

ginseng, was able to upregulate the expression of PYY mRNA in the intestine and downregulate the expression of NPY in the gut.

3.1.3 Regulation of fat accumulation

The active components of TCM effectively reduce fat formation by high-fat diets by controlling the homeostasis of gut microbiota and microbial metabolites. In mice given a high-fat diet, Ren et al. (96) discovered that *Ganoderma lucidum* polysaccharides efficiently lower blood lipid concentrations, regulate the abundance of gut bacteria and various critical pathways, and successfully relieve fat formation. According to Cheng et al. (97), Rubin may greatly raise the amounts of SCFAs and SCFA-producing enzymes in the feces of obese mice, prevent fat accumulation, treat glucolipid metabolic problems, and cause obese animals to lose weight. Heat-treated adzuki bean protein hydrolysates can assist to prevent and treat obesity and its associated consequences, as well as increase the diversity of gut bacteria in mice fed a high-fat diet (98).

3.1.4 Alleviation of chronic mild inflammatory response

TCM and its active components can significantly improve the intestinal environment, enhance intestinal barrier function, and

10.3389/fendo.2023.1149751

control the production of inflammatory markers associated with obesity during intestinal metabolism. Wang et al. (99) showed that fermented Rhizoma Atractylodis Macrocephalae can regulate the gut microbiota, improve the growth of probiotics, alleviate the inflammatory response, and restore the intestinal epithelial barrier function and exert weight loss effects. According to Cao et al. (100), berberine may help people lose weight by controlling the abundance of their gut bacteria and lowering levels of pro-inflammatory cytokines like IL-1, IL-6, and TNF-a. Insoluble polysaccharides from Poria cocos sclerotium can significantly reduce the weight of obese mice by increasing the abundance of intestinal flora, improving the integrity of intestinal mucosa, and activating the intestinal PPAR-y pathway to reduce inflammatory response (101). G. lucidum polysaccharides improve dysbiosis of the gut microbiota and maintain intestinal barrier function to inhibit TLR4/Myd88/ NF-kB signaling pathway expression in adipose tissue, thereby exerting treatment of obesity (102).

3.2 Mechanism of TCM compounds affecting gut microbiota to control obesity

3.2.1 Regulation of gut microbiota abundance and lipid metabolism

The TCM compounds can regulate the abundance of gut microbiota and lipid metabolism to treat obesity. In the mechanism involving the regulation of gut microbiota abundance, SCFAs, and genes involved in lipid metabolism, the Cordyceps guangdongensis lipid-lowering formula dramatically reduced weight and fat accumulation in high-fat diet mice (86). By lowering total cholesterol (TC) and triglyceride (TG) levels and increasing the quantity and variety of gut microbiota, the Jian Pi Tiao Gan Yin medicine helps mice with obesity (109). By controlling the distribution and relative abundance of gut microbiota, enhancing the small intestinal villi, and altering the metabolic pathways linked to obesity, the Lingguizhugan decoction (consisting of Poria, Ramulus Cinnamomi, Rhizoma Atractylodis Macrocephalae, and Radix Glycyrrhizae) aids in weight loss (87-89). By changing the composition of the gut microbiota and the relative abundance of bacteria in obese rats, the Erchen decoction (consisting of Radix et Rhizoma Panax, Radix tangerine, Poria, and Radix Glycyrrhizae) can improve lipid metabolism disorders and change the function of the gut microbiota and cause weight loss (90). In addition to improving obesity, Xiexin Tang (a combination of rhubarb, Huang Lian, and Scutellaria) increases the activity of crucial enzymes involved in the synthesis of SCFAs, enhances the gut microbiota's capacity to metabolize SCFAs, and decreases energy intake while increasing energy expenditure (91). The Shenlian (SL) decoction (composed of Coptis and ginseng) is considered a potential drug for weight loss, which can decrease the expression of flora associated with LPS biosynthesis, increase the diversity and abundance of gut microbiota, and regulate metabolic disorders (92). Daesiho-Tang is an effective herbal formulation in attenuation of obesity in HFD-fed mice through the alteration of gene expressions and modulation of intestinal microbiota (110). The TCM formula (CoTOL) (composed of Glabrous Greenbrier Rhizome, Dioscorea septemloba Thunb, Curcuma Longa, and so on) has beneficial effects on hyperuricemia and overweight, which may be attributed to regulating material metabolism and improving the structure or function of gut microbiota (111). The TCM Formula Kang Shuai Lao Pian (composed of glutinosa, ginseng, japonicas, and so on) could improve HFD-induced obesity, glucose tolerance disorder, and gut dysbiosis (112).

3.2.2 Regulation of gut microbiota abundance and reduction of chronic inflammation

The Sijunzi decoction, which contains ginseng, Atractylodes macrocephala, Poria, and Glycyrrhiza glabra, can have immunomodulatory effects by regulating the number of gut microbiota and SCFA levels (103). The shenling baizhu powder herbal formula (composed of lotus seed meat, coix kernel, sand kernel, bellflower, poria, ginseng, and licorice) reduced weight and serum total cholesterol levels and repaired intestinal mucosa in high-fat diet rats, and the mechanism may be related to reducing TLR4-related protein expression, decreasing the levels of inflammatory factors such as TNF- α and IL-1, and increasing the relative abundance of gut microbiota (104). Jinqi Hypoglycemic Tablets (composed of Huang Lian, Huang Qi, and Jin Yin Hua) regulate the gut microbiota and promote the production of SCFAs in HFD mice by a mechanism related to enhancing intestinal barrier function and reducing host inflammatory response (105). The Shen-Yan-Fang-Shuai formula (composed of Astragali Radix, Radix Angelicae Sinensis, Rheum Officinale Baill, and four other herbs) treatment prevented weight gain, low-grade inflammation, and insulin resistance in HFD mice (113). The Gegenqinlian decoction could intervene from inflammatory response through multiple targets and multiple channels to adjust the balance of intestinal mucosa flora (114).

4 Conclusion

Gut microbiota is closely related to the occurrence and development of obesity and related metabolic diseases. TCM can improve the metabolic disorder of obesity by regulating the imbalance of gut microbiota. Gut microbiota can be used as a new target for the prevention and treatment of obesity and to improve obesity and its metabolic diseases by correcting gut microbiota disorders. This study primarily summarizes how TCM treats obesity by regulating the flora in the intestines. The active ingredients of TCM and its compound can lower blood lipid levels and fat accumulation in patients by controlling the distribution of gut microbiota, enhancing the capacity of gut microbiota metabolism to generate SCFAs, reducing inflammatory damage, and improving the energy metabolism of the body, thereby improving obesity. Further study of the relationship between TCM and gut microbiota will help to enrich the theory of TCM and develop TCM preparations that target the gut microbiota. However, the research on the interaction between TCM and gut microbiota is complex and still in the initial stage. Many key scientific issues need to be further explored. The internal

environment of gut microbiota is complex, and different types of bacteria produce different metabolic enzymes, which makes it difficult to explain the biotransformation of active ingredients of TCM and gut microbiota. In the future, the molecular mechanism of gut microbiota regulation should be further explored. It is helpful for us to understand that TCM directly acts on gut microbiota or indirectly regulates the structure of gut microbiota and maintains the balance of intestinal micro-ecology, in order to promote the use of herbal medicine in the prevention and treatment of difficult-totreat diseases such as obesity.

Author contributions

DL and WT reviewed a large amount of literature, conducted relevant data analysis, and drafted the manuscript. YYW and QG completed the mechanism diagram. YY, HWZ, YMZ and YW analyzed the relevance of the literature and provided the analysis report. YZ supervised the project and revised the manuscript. YKZ, HL and SL analyzed the data and revised the manuscript. HZ reviewed the article and provided revisions, formal analysis, and manuscript drafting. All authors contributed to the article and approved the submitted version.

Funding

This study was supported by the Heilongjiang Province in Special Funding for Postdoctoral (LBH-Q20185), the Heilongjiang

Provincial North Medicine and Function Food for Featured Discipline of Heilongjiang Province (2018-TSXK-02), the Technological Innovation Team Construction Project of Heilongjiang Provincial Department of Education (2021-KYYEF-0638), the Key Program of the Natural Science Foundation of Heilongjiang Province of China (ZD2022H006), the Joint Guidance Project of the Natural Science Foundation of Heilongjiang Province (LH2022H093), and the Basic Research Project of Fundamental Research Business Expenses of Education Department in Heilongjiang Province (No. 2022-KYYWF-0615).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Zhang C, Sheng J, Xie W, Luo X, Xue Y, Xu G, et al. Mechanism and basis of traditional Chinese medicine against obesity: Prevention and treatment strategies. *Front Pharmacol* (2021) 12:615895. doi: 10.3389/fphar.2021.615895

2. Aaseth J, Ellefsen S, Alehagen U, Sundfør T, Alexander J. Diets and drugs for weight loss and health in obesity - an update. *BioMed Pharmacother* (2021) 140:111789. doi: 10.1016/j.biopha.2021.111789

3. Wang L, Yu C, Li J, Tian Q, Du Y. Mechanism of action of acupuncture in obesity: A perspective from the hypothalamus. *Front Endocrinol (Lausanne)* (2021) 12:632324. doi: 10.3389/fendo.2021.632324

4. Piché M, Tchernof A, Després J. Obesity phenotypes, diabetes, and cardiovascular diseases. *Circ Res* (2020) 126(11):1477–500. doi: 10.1161/circresaha.120.316101

5. Fan Q, Xu F, Liang B, Zou X. The anti-obesity effect of traditional Chinese medicine on lipid metabolism. *Front Pharmacol* (2021) 12:696603. doi: 10.3389/fphar.2021.696603

6. Krentz A, Fujioka K, Hompesch M. Evolution of pharmacological obesity treatments: Focus on adverse side-effect profiles. *Diabetes Obes Metab* (2016) 18 (6):558-70. doi: 10.1111/dom.12657

7. Lin X, Li H. Obesity: Epidemiology, pathophysiology, and therapeutics. *Front Endocrinol (Lausanne)* (2021) 12:706978. doi: 10.3389/fendo.2021.706978

8. Cheng Z, Zhang L, Yang L, Chu H. The critical role of gut microbiota in obesity. *Front Endocrinol (Lausanne)* (2022) 13:1025706. doi: 10.3389/fendo.2022.1025706

9. Clarke G, Stilling R, Kennedy P, Stanton C, Cryan J, Dinan T. Minireview: Gut microbiota: the neglected endocrine organ. *Mol Endocrinol* (2014) 28(8):1221–38. doi: 10.1210/me.2014-1108

10. Jia Q, Xie Y, Lu C, Zhang A, Lu Y, Lv S, et al. Endocrine organs of cardiovascular diseases: Gut microbiota. J Cell Mol Med (2019) 23(4):2314–23. doi: 10.1111/jcmm.14164

11. Chen Y, Zhou J, Wang L. Role and mechanism of gut microbiota in human disease. Front Cell Infect Microbiol (2021) 11:625913. doi: 10.3389/fcimb.2021.625913

12. Wu Y, Peng X, Li X, Li D, Tan Z, Yu R. Sex hormones influence the intestinal microbiota composition in mice. *Front Microbiol* (2022) 13:964847. doi: 10.3389/fmicb.2022.964847

13. Gérard P. Gut microbiota and obesity. Cell Mol Life Sci (2016) 73(1):147–62. doi: 10.1007/s00018-015-2061-5

14. Thursby E, Juge N. Introduction to the human gut microbiota. *Biochem J* (2017) 474(11):1823–36. doi: 10.1042/bcj20160510

15. Fan Y, Pedersen O. Gut microbiota in human metabolic health and disease. *Nat Rev Microbiol* (2021) 19(1):55–71. doi: 10.1038/s41579-020-0433-9

16. Li C, Zhou K, Xiao N, Peng M, Tan Z. The effect of qiweibaizhu powder crude polysaccharide on antibiotic-associated diarrhea mice is associated with restoring intestinal mucosal bacteria. *Front Nutr* (2022) 9:952647. doi: 10.3389/fnut.2022.952647

17. Qiao B, Li X, Peng M, Hui H, Tan Z. Alteration of intestinal mucosal microbiota in mice with Chinese dampness-heat syndrom diarrhea by improper diet combined with high temperature and humidity environments. *Front Cell Infect Microbiol* (2022) 12:1096202. doi: 10.3389/fcimb.2022.1096202

18. He L, Long C, Liu Y, Guo Y, Xiao N, Tan Z. Effects of debaryomyces hansenii treatment on intestinal microorganisms in mice with antibiotics-induced diarrhea. *3 Biotech* (2017) 7(5):347. doi: 10.1007/s13205-017-0953-9

19. Long C, Liu Y, He L, Tan Q, Yu Z, Xiao N, et al. Bacterial lactase genes diversity in intestinal mucosa of mice with dysbacterial diarrhea induced by antibiotics. *3 Biotech* (2018) 8(3):176. doi: 10.1007/s13205-018-1191-5

20. Xie G, Tan K, Peng M, Long C, Li D, Tan Z. Bacterial diversity in intestinal mucosa of antibiotic-associated diarrhea mice. *3 Biotech* (2019) 9(12):444. doi: 10.1007/s13205-019-1967-2

21. Zhou K, Peng M, Deng N, Tan Z, Xiao N. Lactase bacteria in intestinal mucosa are associated with diarrhea caused by high-fat and high-protein diet. *BMC Microbiol* (2022) 22(1):226. doi: 10.1186/s12866-022-02647-2

22. Liu B, Liu X, Liang Z, Wang J. Gut microbiota in obesity. *World J Gastroenterol* (2021) 27(25):3837–50. doi: 10.3748/wjg.v27.i25.3837

23. Li X, Deng N, Zheng T, Qiao B, Peng M, Xiao N, et al. Importance of dendrobium officinale in improving the adverse effects of high-fat diet on mice associated with intestinal contents microbiota. *Front Nutr* (2022) 9:957334. doi: 10.3389/fnut.2022.957334

24. Wu Y, Zhang C, Shao H, Luo H, Tan Z. Characteristics of intestinal microbiota and enzyme activities in mice fed with lily bulb. *3 Biotech* (2021) 11(1):17. doi: 10.1007/s13205-020-02597-4

25. Shao H, Zhang C, Wang C, Tan Z. Intestinal mucosal bacterial diversity of antibiotic-associated diarrhea (AAD) mice treated with debaryomyces hansenii and qiweibaizhu powder. 3 Biotech (2020) 10(9):392. doi: 10.1007/s13205-020-02383-2

26. Yuan Z, Zhang C, Peng X, Shu L, Long C, Tan Z. Intestinal microbiota characteristics of mice treated with folium senna decoction gavage combined with restraint and tail pinch stress. *3 Biotech* (2020) 10(4):180. doi: 10.1007/s13205-020-02172-x

27. Zhang C, Shao H, Peng X, Liu T, Tan Z. Microbiotal characteristics colonized in intestinal mucosa of mice with diarrhoea and repeated stress. *3 Biotech* (2020) 10 (8):372. doi: 10.1007/s13205-020-02368-1

28. Long C, Liu Y, He L, Yu R, Li D, Tan Z, et al. Bacterial lactase genes diversity in intestinal mucosa of dysbacterial diarrhea mice treated with qiweibaizhu powder. *3 Biotech* (2018) 8(10):423. doi: 10.1007/s13205-018-1460-3

29. Xie G, Deng N, Zheng T, Peng X, Zhang S, Tan Z. Total glycosides contribute to the anti-diarrheal effects of qiwei baizhu powder *via* regulating gut microbiota and bile acids. *Front Cell Infect Microbiol* (2022) 12:945263. doi: 10.3389/fcimb.2022.945263

30. Li X, Zhang C, Tan Z, Yuan J. Network pharmacology-based analysis of gegenqinlian decoction regulating intestinal microbial activity for the treatment of diarrhea. *Evid Based Complement Alternat Med* (2021) 2021:5520015. doi: 10.1155/2021/5520015

31. Turnbaugh P, Hamady M, Yatsunenko T, Cantarel B, Duncan A, Ley R, et al. A core gut microbiome in obese and lean twins. *Nature* (2009) 457(7228):480-4. doi: 10.1038/nature07540

32. Wang P, Gao J, Ke W, Wang J, Li D, Liu R, et al. Resveratrol reduces obesity in high-fat diet-fed mice *via* modulating the composition and metabolic function of the gut microbiota. *Free Radic Biol Med* (2020) 156:83–98. doi: 10.1016/j.freeradbiomed.2020.04.013

33. Gomes A, Hoffmann C, Mota J. The human gut microbiota: Metabolism and perspective in obesity. *Gut Microbes* (2018) 9(4):308–25. doi: 10.1080/19490976.2018. 1465157

34. Muscogiuri G, Cantone E, Cassarano S, Tuccinardi D, Barrea L, Savastano S, et al. Gut microbiota: A new path to treat obesity. *Int J Obes Suppl* (2019) 9(1):10–9. doi: 10.1038/s41367-019-0011-7

35. Wostmann B, Larkin C, Moriarty A, Bruckner-Kardoss E. Dietary intake, energy metabolism, and excretory losses of adult male germfree wistar rats. *Lab Anim Sci* (1983) 33(1):46–50.

36. Bäckhed F, Ding H, Wang T, Hooper L, Koh G, Nagy A, et al. The gut microbiota as an environmental factor that regulates fat storage. *Proc Natl Acad Sci U.S.A.* (2004) 101(44):15718–23. doi: 10.1073/pnas.0407076101

37. Le Chatelier E, Nielsen T, Qin J, Prifti E, Hildebrand F, Falony G, et al. Richness of human gut microbiome correlates with metabolic markers. *Nature* (2013) 500 (7464):541–6. doi: 10.1038/nature12506

38. Liu R, Hong J, Xu X, Feng Q, Zhang D, Gu Y, et al. Gut microbiome and serum metabolome alterations in obesity and after weight-loss intervention. *Nat Med* (2017) 23(7):859–68. doi: 10.1038/nm.4358

39. Vallianou N, Stratigou T, Christodoulatos G, Dalamaga M. Understanding the role of the gut microbiome and microbial metabolites in obesity and obesity-associated metabolic disorders: Current evidence and perspectives. *Curr Obes Rep* (2019) 8 (3):317–32. doi: 10.1007/s13679-019-00352-2

40. Moszak M, Szulińska M, Bogdański P. You are what you eat-the relationship between diet, microbiota, and metabolic disorders-a review. *Nutrients* (2020) 12 (4):1096. doi: 10.3390/nu12041096

41. Merra G, Noce A, Marrone G, Cintoni M, Tarsitano M, Capacci A, et al. Influence of Mediterranean diet on human gut microbiota. *Nutrients* (2020) 13(1):7. doi: 10.3390/nu13010007

42. Qiao B, Li X, Zheng T, Tan Z. Different effects of lard and vegetable blend oil on intestinal microorganisms, enzyme activity and blood routine in mice. *J Oleo Sci* (2022) 71(2):301–10. doi: 10.5650/jos.ess21247

43. Turnbaugh P, Ley R, Mahowald M, Magrini V, Mardis E, Gordon J. An obesityassociated gut microbiome with increased capacity for energy harvest. *Nature* (2006) 444(7122):1027–31. doi: 10.1038/nature05414

44. Meijnikman A, Aydin O, Prodan A, Tremaroli V, Herrema H, Levin E, et al. Distinct differences in gut microbial composition and functional potential from lean to morbidly obese subjects. *J Intern Med* (2020) 288(6):699–710. doi: 10.1111/joim.13137

45. Kobyliak N, Virchenko O, Falalyeyeva T. Pathophysiological role of host microbiota in the development of obesity. *Nutr J* (2016) 15:43. doi: 10.1186/s12937-016-0166-9

46. Petersen C, Bell R, Klag K, Lee S, Soto R, Ghazaryan A, et al. T Cell-mediated regulation of the microbiota protects against obesity. *Science* (2019) 365(6451): eaat9351. doi: 10.1126/science.aat9351

47. Everard A, Belzer C, Geurts L, Ouwerkerk J, Druart C, Bindels L, et al. Cross-talk between akkermansia muciniphila and intestinal epithelium controls diet-induced obesity. *Proc Natl Acad Sci U.S.A.* (2013) 110(22):9066–71. doi: 10.1073/pnas.1219451110

48. Wu D, Wang H, Xie L, Hu F. Cross-talk between gut microbiota and adipose tissues in obesity and related metabolic diseases. *Front Endocrinol (Lausanne)* (2022) 13:908868. doi: 10.3389/fendo.2022.908868

49. Koh A, De Vadder F, Kovatcheva-Datchary P, Bäckhed F. From dietary fiber to host physiology: Short-chain fatty acids as key bacterial metabolites. *Cell* (2016) 165 (6):1332–45. doi: 10.1016/j.cell.2016.05.041

50. Canfora E, Meex R, Venema K, Blaak E. Gut microbial metabolites in obesity, NAFLD and T2DM. *Nat Rev Endocrinol* (2019) 15(5):261–73. doi: 10.1038/s41574-019-0156-z

51. Li C, Xiao N, Deng N, Li D, Tan Z, Peng M. Dose of sucrose affects the efficacy of qiweibaizhu powder on antibiotic-associated diarrhea: Association with intestinal mucosal microbiota, short-chain fatty acids, IL-17, and MUC2. *Front Microbiol* (2023) 14:1108398. doi: 10.3389/fmicb.2023.1108398

52. Zhang L, Liu C, Jiang Q, Yin Y. Butyrate in energy metabolism: There is still more to learn. *Trends Endocrinol Metab* (2021) 32(3):159-69. doi: 10.1016/j.tem.2020.12.003

53. Cani P, Van Hul M, Lefort C, Depommier C, Rastelli M, Everard A. Microbial regulation of organismal energy homeostasis. *Nat Metab* (2019) 1(1):34-46. doi: 10.1038/s42255-018-0017-4

54. Lu Y, Fan C, Li P, Lu Y, Chang X, Qi K. Short chain fatty acids prevent high-fatdiet-induced obesity in mice by regulating G protein-coupled receptors and gut microbiota. *Sci Rep* (2016) 6:37589. doi: 10.1038/srep37589

55. Tirosh A, Calay E, Tuncman G, Claiborn K, Inouye K, Eguchi K, et al. The shortchain fatty acid propionate increases glucagon and FABP4 production, impairing insulin action in mice and humans. *Sci Transl Med* (2019) 11(489):eaav0120. doi: 10.1126/scitranslmed.aav0120

56. Engevik M, Luck B, Visuthranukul C, Ihekweazu F, Engevik A, Shi Z, et al. Human-derived bifidobacterium dentium modulates the mammalian serotonergic system and gut-brain axis. *Cell Mol Gastroenterol Hepatol* (2021) 11(1):221–48. doi: 10.1016/j.jcmgh.2020.08.002

57. Generoso J, Giridharan V, Lee J, Macedo D, Barichello T. The role of the microbiota-gut-brain axis in neuropsychiatric disorders. *Braz J Psychiatry* (2021) 43 (3):293–305. doi: 10.1590/1516-4446-2020-0987

58. Torres-Fuentes C, Schellekens H, Dinan T, Cryan J. The microbiota-gut-brain axis in obesity. *Lancet Gastroenterol Hepatol* (2017) 2(10):747–56. doi: 10.1016/s2468-1253(17)30147-4

 Oduro-Donkor D, Turner M, Farnaud S, Renshaw D, Kyrou I, Hanson P, et al. Modification of fecal microbiota as a mediator of effective weight loss and metabolic benefits following bariatric surgery. *Expert Rev Endocrinol Metab* (2020) 15(5):363–73. doi: 10.1080/17446651.2020.1801412

60. Cunningham A, Stephens J, Harris D. A review on gut microbiota: a central factor in the pathophysiology of obesity. *Lipids Health Dis* (2021) 20(1):65. doi: 10.1186/s12944-021-01491-z

61. Naraoka Y, Yamaguchi T, Hu A, Akimoto K, Kobayashi H. Short chain fatty acids upregualte adipokine production in type 2 diabetes-derived human adipocytes. *Acta Endocrinol (Buchar)* (2018) 14(3):287–93. doi: 10.4183/aeb.2018.287

62. Li Z, Yi C, Katiraei S, Kooijman S, Zhou E, Chung C, et al. Butyrate reduces appetite and activates brown adipose tissue *via* the gut-brain neural circuit. *Gut* (2018) 67(7):1269–79. doi: 10.1136/gutjnl-2017-314050

63. Frost G, Sleeth M, Sahuri-Arisoylu M, Lizarbe B, Cerdan S, Brody L, et al. The short-chain fatty acid acetate reduces appetite *via* a central homeostatic mechanism. *Nat Commun* (2014) 5:3611. doi: 10.1038/ncomms4611

64. Jing N, Liu X, Jin M, Yang X, Hu X, Li C, et al. Fubrick tea attenuates high-fat diet induced fat deposition and metabolic disorder by regulating gut microbiota and caffeine metabolism. *Food Funct* (2020) 11(8):6971–86. doi: 10.1039/d0fo01282c

65. Villaret A, Galitzky J, Decaunes P, Estève D, Marques MA, Sengenès C, et al. Adipose tissue endothelial cells from obese human subjects: Differences among depots in angiogenic, metabolic, and inflammatory gene expression and cellular senescence. *Diabetes* (2010) 59(11):2755–63. doi: 10.2337/db10-0398

66. Goldberg I. 2017 George Lyman duff memorial lecture: Fat in the blood, fat in the artery, fat in the heart: Triglyceride in physiology and disease. *Arterioscler Thromb Vasc Biol* (2018) 38(4):700–6. doi: 10.1161/atvbaha.117.309666

67. Jiao A, Yu B, He J, Yu J, Zheng P, Luo Y, et al. Sodium acetate, propionate, and butyrate reduce fat accumulation in mice *via* modulating appetite and relevant genes. *Nutrition* (2021) 87-88:111198. doi: 10.1016/j.nut.2021.111198

68. Cuevas-Sierra A, Ramos-Lopez O, Riezu-Boj J, Milagro F, Martinez J. Diet, gut microbiota, and obesity: Links with host genetics and epigenetics and potential applications. *Adv Nutr* (2019) 10(suppl_1):S17–s30. doi: 10.1093/advances/nmy078

69. Steinert R, Beglinger C, Langhans W. Intestinal GLP-1 and satiation: from man to rodents and back. *Int J Obes (Lond)* (2016) 40(2):198–205. doi: 10.1038/ijo.2015.172

70. Bäckhed F, Manchester J, Semenkovich C, Gordon J. Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. *Proc Natl Acad Sci U.S.A.* (2007) 104(3):979–84. doi: 10.1073/pnas.0605374104

71. Al Bander Z, Nitert M, Mousa A, Naderpoor N. The gut microbiota and inflammation: An overview. *Int J Environ Res Public Health* (2020) 17(20):7618. doi: 10.3390/ijerph17207618

72. Soltani N, Esmaeil N, Marandi S, Hovsepian V, Momen T, Shahsanai A, et al. Assessment of the effect of short-term combined high-intensity interval training on TLR4, NF- κ B and IRF3 expression in young overweight and obese girls. *Public Health Genomics* (2020) 23(1-2):26–36. doi: 10.1159/000506057

73. Ridaura V, Faith J, Rey F, Cheng J, Duncan A, Kau A, et al. Gut microbiota from twins discordant for obesity modulate metabolism in mice. *Science* (2013) 341 (6150):1241214. doi: 10.1126/science.1241214

74. Liu J, Qiao B, Deng N, Wu Y, Li D, Tan Z. The diarrheal mechanism of mice with a high-fat diet in a fatigued state is associated with intestinal mucosa microbiota. *3 Biotech* (2023) 13(3):77. doi: 10.1007/s13205-023-03491-5

75. Li M, van Esch B, Wagenaar G, Garssen J, Folkerts G, Henricks P. Pro- and antiinflammatory effects of short chain fatty acids on immune and endothelial cells. *Eur J Pharmacol* (2018) 831:52–9. doi: 10.1016/j.ejphar.2018.05.003

76. Vatanen T, Franzosa E, Schwager R, Tripathi S, Arthur T, Vehik K, et al. The human gut microbiome in early-onset type 1 diabetes from the TEDDY study. *Nature* (2018) 562(7728):589–94. doi: 10.1038/s41586-018-0620-2

77. Kelly C, Zheng L, Campbell E, Saeedi B, Scholz C, Bayless A, et al. Crosstalk between microbiota-derived short-chain fatty acids and intestinal epithelial HIF augments tissue barrier function. *Cell Host Microbe* (2015) 17(5):662–71. doi: 10.1016/j.chom.2015.03.005

78. Fukuda S, Toh H, Hase K, Oshima K, Nakanishi Y, Yoshimura K, et al. Bifidobacteria can protect from enteropathogenic infection through production of acetate. *Nature* (2011) 469(7331):543–7. doi: 10.1038/nature09646

79. Park J, Kim M, Kang S, Jannasch A, Cooper B, Patterson J, et al. Short-chain fatty acids induce both effector and regulatory T cells by suppression of histone deacetylases and regulation of the mTOR-S6K pathway. *Mucosal Immunol* (2015) 8 (1):80–93. doi: 10.1038/mi.2014.44

80. Iyengar N, Gucalp A, Dannenberg A, Hudis C. Obesity and cancer mechanisms: Tumor microenvironment and inflammation. *J Clin Oncol* (2016) 34(35):4270–6. doi: 10.1200/jco.2016.67.4283

81. Aguilar E, Leonel A, Teixeira L, Silva A, Silva J, Pelaez J, et al. Butyrate impairs atherogenesis by reducing plaque inflammation and vulnerability and decreasing NF κ B activation. *Nutr Metab Cardiovasc Dis* (2014) 24(6):606–13. doi: 10.1016/j.numecd.2014.01.002

82. Zhang M, Zhao J, Deng J, Duan Z, Zhu C, Fan D. The protective effect of protopanaxatriol-type saponin on intestinal health in antibiotic-treated mice. *Food Funct* (2019) 10(7):4124–33. doi: 10.1039/c9fo00242a

83. Song M, Lim S, Wang J, Kim H. The root of atractylodes macrocephala koidzumi prevents obesity and glucose intolerance and increases energy metabolism in mice. *Int J Mol Sci* (2018) 19(1):278. doi: 10.3390/ijms19010278

84. Cai W, Xu J, Li G, Liu T, Guo X, Wang H, et al. Ethanol extract of propolis prevents high-fat diet-induced insulin resistance and obesity in association with modulation of gut microbiota in mice. *Food Res Int* (2020) 130:108939. doi: 10.1016/j.foodres.2019.108939

85. Wang C, Yen J, Cheng Y, Lin C, Hsieh C, Gau R, et al. Polygala tenuifolia extract inhibits lipid accumulation in 3T3-L1 adipocytes and high-fat diet-induced obese mouse model and affects hepatic transcriptome and gut microbiota profiles. *Food Nutr Res* (2017) 61(1):1379861. doi: 10.1080/16546628.2017.1379861

86. Wang G, Sun C, Xie B, Wang T, Liu H, Chen X, et al. Cordyceps guangdongensis lipid-lowering formula alleviates fat and lipid accumulation by modulating gut microbiota and short-chain fatty acids in high-fat diet mice. *Front Nutr* (2022) 9:1038740. doi: 10.3389/fnut.2022.1038740

87. Liu M, Huang Y, Zhang T, Tan L, Lu X, Qin J. Lingguizhugan decoction attenuates diet-induced obesity and hepatosteatosis *via* gut microbiota. *World J Gastroenterol* (2019) 25(27):3590–606. doi: 10.3748/wjg.v25.i27.3590

88. Wu R, Zhao D, An R, Wang Z, Li Y, Shi B, et al. Linggui zhugan formula improves glucose and lipid levels and alters gut microbiota in high-fat diet-induced diabetic mice. *Front Physiol* (2019) 10:918. doi: 10.3389/fphys.2019.00918

89. Ning Y, Gong Y, Zheng T, Xie Y, Yuan S, Ding W. Lingguizhugan decoction targets intestinal microbiota and metabolites to reduce insulin resistance in high-fat diet rats. *Diabetes Metab Syndr Obes* (2022) 15:2427–42. doi: 10.2147/dmso.S370492

90. Zhao T, Zhan L, Zhou W, Chen W, Luo J, Zhang L, et al. The effects of erchen decoction on gut microbiota and lipid metabolism disorders in zucker diabetic fatty rats. *Front Pharmacol* (2021) 12:647529. doi: 10.3389/fphar.2021.647529

91. Xiao S, Zhang Z, Chen M, Zou J, Jiang S, Qian D, et al. Xiexin tang ameliorates dyslipidemia in high-fat diet-induced obese rats *via* elevating gut microbiota-derived short chain fatty acids production and adjusting energy metabolism. *J Ethnopharmacol* (2019) 241:112032. doi: 10.1016/j.jep.2019.112032

92. Sun R, Huang W, Xiao Y, Wang D, Mu G, Nan H, et al. Shenlian (SL) decoction, a traditional Chinese medicine compound, may ameliorate blood glucose *via* mediating the gut microbiota in db/db mice. *J Diabetes Res* (2022) 2022:7802107. doi: 10.1155/2022/7802107

93. Neyrinck A, Sánchez C, Rodriguez J, Cani P, Bindels L, Delzenne N. Prebiotic effect of berberine and curcumin is associated with the improvement of obesity in mice. *Nutrients* (2021) 13(5):1436. doi: 10.3390/nu13051436

94. Casanova-Martí À, Serrano J, Portune K, Sanz Y, Blay M, Terra X, et al. Grape seed proanthocyanidins influence gut microbiota and enteroendocrine secretions in female rats. *Food Funct* (2018) 9(3):1672–82. doi: 10.1039/c7fo02028g

95. Lin N, Cai D, Jin D, Chen Y, Shi J. Ginseng panaxoside Rb1 reduces body weight in diet-induced obese mice. *Cell Biochem Biophys* (2014) 68(1):189–94. doi: 10.1007/ s12013-013-9688-3

96. Ren F, Meng C, Chen W, Chen H, Chen W. Ganoderma amboinense polysaccharide prevents obesity by regulating gut microbiota in high-fat-diet mice. *Food Biosci* (2021) 42:101107. doi: 10.1016/j.fbio.2021.101107

97. Cheng L, Shi L, He C, Wang C, Lv Y, Li H, et al. Rutin-activated adipose tissue thermogenesis is correlated with increased intestinal short-chain fatty acid levels. *Phytother Res* (2022) 36(6):2495–510. doi: 10.1002/ptr.7462

98. Zhao Q, Fu Y, Zhang F, Wang C, Yang X, Bai S, et al. Heat-treated adzuki bean protein hydrolysates reduce obesity in mice fed a high-fat diet *via* remodeling gut microbiota and improving metabolic function. *Mol Nutr Food Res* (2022) 66(8): e2100907. doi: 10.1002/mnfr.202100907

99. Wang J, Bose S, Kim H, Han K, Kim H. Fermented rhizoma atractylodis macrocephalae alleviates high fat diet-induced obesity in association with regulation of intestinal permeability and microbiota in rats. *Sci Rep* (2015) 5:8391. doi: 10.1038/srep08391

100. Cao Y, Pan Q, Cai W, Shen F, Chen G, Xu L, et al. Modulation of gut microbiota by berberine improves steatohepatitis in high-fat diet-fed BALB/C mice. Arch Iran Med (2016) 19(3):197–203. doi: 10.1088/1475-7516/2016/03/008

101. Sun S, Wang K, Ma K, Bao L, Liu H. An insoluble polysaccharide from the sclerotium of poria cocos improves hyperglycemia, hyperlipidemia and hepatic steatosis in ob/ob mice *via* modulation of gut microbiota. *Chin J Nat Med* (2019) 17 (1):3–14. doi: 10.1016/s1875-5364(19)30003-2

102. Sang T, Guo C, Guo D, Wu J, Wang Y, Wang Y, et al. Suppression of obesity and inflammation by polysaccharide from sporoderm-broken spore of ganoderma lucidum *via* gut microbiota regulation. *Carbohydr Polymers* (2021) 256:117594. doi: 10.1016/j.carbpol.2020.117594

103. Gao B, Wang R, Peng Y, Li X. Effects of a homogeneous polysaccharide from sijunzi decoction on human intestinal microbes and short chain fatty acids *in vitro*. J Ethnopharmacol (2018) 224:465–73. doi: 10.1016/j.jep.2018.06.006

104. Zhang Y, Tang K, Deng Y, Chen R, Liang S, Xie H, et al. Effects of shenling baizhu powder herbal formula on intestinal microbiota in high-fat diet-induced NAFLD rats. *BioMed Pharmacother* (2018) 102:1025–36. doi: 10.1016/j.biopha.2018.03.158

105. Cao Y, Yao G, Sheng Y, Yang L, Wang Z, Yang Z, et al. JinQi jiangtang tablet regulates gut microbiota and improve insulin sensitivity in type 2 diabetes mice. J Diabetes Res (2019) 2019:1872134. doi: 10.1155/2019/1872134

106. Li X, Peng X, Guo K, Tan Z. Bacterial diversity in intestinal mucosa of mice fed with dendrobium officinale and high-fat diet. 3 Biotech (2021) 11(1):22. doi: 10.1007/s13205-020-02558-x

107. Guo K, Xu S, Zhang Q, Peng M, Yang Z, Li W, et al. Bacterial diversity in the intestinal mucosa of mice fed with asparagus extract under high-fat diet condition. 3 *Biotech* (2020) 10(5):228. doi: 10.1007/s13205-020-02225-1

108. Guo K, Yan Y, Zeng C, Shen L, He Y, Tan Z. Study on baohe pills regulating intestinal microecology and treating diarrhea of high-fat and high-protein diet mice. *BioMed Res Int* (2022) 2022:6891179. doi: 10.1155/2022/6891179

109. Dong W, Mao Y, Xiang Z, Zhu J, Wang H, Wang A, et al. Traditional Chinese medicine formula jian pi tiao gan yin reduces obesity in mice by modulating the gut microbiota and fecal metabolism. *Evid Based Complement Alternat Med* (2022) 2022;9727889. doi: 10.1155/2022/9727889

110. Hussain A, Yadav M, Bose S, Wang J, Lim D, Song Y, et al. Daesiho-tang is an effective herbal formulation in attenuation of obesity in mice through alteration of gene expression and modulation of intestinal microbiota. *PloS One* (2016) 11(11):e0165483. doi: 10.1371/journal.pone.0165483

111. Gao Y, Sun J, Zhang Y, Shao T, Li H, Wang M, et al. Effect of a traditional Chinese medicine formula (CoTOL) on serum uric acid and intestinal flora in obese hyperuricemic mice inoculated with intestinal bacteria. *Evid Based Complement Alternat Med* (2020) 2020:8831937. doi: 10.1155/2020/8831937

112. Gong S, Ye T, Wang M, Wang M, Li Y, Ma L, et al. Traditional Chinese medicine formula kang shuai lao pian improves obesity, gut dysbiosis, and fecal metabolic disorders in high-fat diet-fed mice. *Front Pharmacol* (2020) 11:297. doi: 10.3389/fphar.2020.00297

113. Wang Z, Lu J, Zhou J, Sun W, Qiu Y, Chen W, et al. Modulation of the gut microbiota by shen-Yan-Fang-Shuai formula improves obesity induced by high-fat diets. *Front Microbiol* (2020) 11:564376. doi: 10.3389/fmicb.2020.564376

114. Li X, Zhang C, Hui H, Tan Z. Effect of gegenqinlian decoction on intestinal mucosal flora in mice with diarrhea induced by high temperature and humidity treatment. *3 Biotech* (2021) 11(2):83. doi: 10.1007/s13205-020-02628-0