Check for updates

OPEN ACCESS

EDITED BY Yizhe Tang, Shenzhen University Health Science Centre, China

REVIEWED BY Junfei Zhao, Guangdong Academy of Medical Sciences, China Bo Pan, Children's Hospital of Chongqing Medical University, China

*CORRESPONDENCE Yifei Li 🔀 liyfwcsh@scu.edu.cn Binlu Zhu 🔀 249308925@qq.com

 $^{\mathrm{t}}\mathrm{These}$ authors have contributed equally to this work

RECEIVED 03 November 2023 ACCEPTED 18 December 2023 PUBLISHED 08 January 2024

CITATION

Tang J, Zhao C, Lin S, Li X, Zhu B and Li Y (2024) Controversial causal association between IGF family members and osteoporosis: a Mendelian randomization study between UK and FinnGen biobanks. *Front. Endocrinol.* 14:1332803. doi: 10.3389/fendo.2023.1332803

COPYRIGHT

© 2024 Tang, Zhao, Lin, Li, Zhu and Li. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Controversial causal association between IGF family members and osteoporosis: a Mendelian randomization study between UK and FinnGen biobanks

Jie Tang[†], Chanjuan Zhao[†], Sha Lin, Xing Li, Binlu Zhu^{*} and Yifei Li^{*}

Key Laboratory of Birth Defects and Related Diseases of Women and Children of Ministry of Education (MOE), Department of Pediatrics, West China Second University Hospital, Sichuan University, Chengdu, Sichuan, China

Objectives: Osteoporosis, a prevalent skeletal disorder characterized by reduced bone strength, is closely linked to the IGF system, crucial for skeletal metabolism. However, the precise nature of this relationship remains elusive. In this study, we employed Mendelian randomization (MR) to unravel the associations between genetically predicted serum IGF system member levels and osteoporosis.

Methods: A two-sample MR approach was employed to investigate these causal associations based on two individual datasets. Predictions of 14 serum levels of IGF system members were made using 11,036,163 relevant Single Nucleotide Polymorphisms (SNPs) within a cohort of 4,301 individuals of European descent. Genetic association estimates for osteoporosis were derived from two publicly available GWAS consortia: the Finnish consortium from the FinnGen biobank, comprising 212,778 individuals of Finnish descent (3,203 cases and 209,575 controls), and the UK consortium from the UK Biobank, including 337,159 individuals of European descent (5,266 cases and 331,893 controls).

Results: According to the UK dataset, IGF-1 levels were associated with a reduced risk of osteoporosis, as indicated by the weighted median method (Odds Ratio [OR] = 0.998, 95% CI = 0.997–1.000, P = 0.032). Additionally, higher levels of IGFBP-3 were linked to a decreased risk of osteoporosis using the Inverse-Variance Weighted (IVW) method (OR = 0.999, 95% CI = 0.998–1.000, P = 0.019), and CTGF levels exhibited a negative association with osteoporosis, as determined by the weighted median method (OR = 0.998, 95% CI = 0.996–0.999, P = 0.004). In the FinnGen dataset, IGF-1 and IGFBP-3 were not identified to be associated with osteoporosis. While, IGF-LR1 levels displayed a negative association with osteoporosis, according to the MR-Egger method (OR = 0.886, 95% CI = 0.795–0.987, P = 0.036), while CYR61 was linked to an increased risk of osteoporosis based on both the weighted median and IVW methods (OR = 1.154, 95% CI = 1.009–1.319, P = 0.037, and OR = 1.115, 95% CI = 1.022–1.215, P = 0.014, respectively).

Conclusion: This study provides compelling evidence that certain IGF family members play a role in the pathogenesis of osteoporosis between different

datasets, indicating population specific causal effects between IGF family and osteoporosis. Although the results from both datasets demonstrated that IGF family involved in the pathogenesis of osteoporosis, but the responding key molecules might be various among different population. Subsequent research is warranted to evaluate the potential of these biomarkers as targets for osteoporosis prevention and treatment in specific population.

KEYWORDS

osteoporosis, IGF-1, IGF family, developmental disorders, Mendelian randomization

Introduction

Osteoporosis poses a significant public health challenge, impacting nearly 200 million individuals and resulting in 8.9 million fractures worldwide annually (1). The clinical definition of osteoporosis is primarily based on bone mineral density (BMD), a surrogate marker of bone strength that is substantially influenced by the peak bone mass attained during childhood and adolescence (2, 3). Osteoporosis results from an imbalance between bone formation and bone resorption. This imbalance is regulated by osteoblasts and osteoclasts. Disruptions in the molecular signals controlling the activity of these cells can lead to excessive bone loss. Although genetic factors influence an individual's susceptibility to osteoporosis, but the adverse environmental exposure contributes as a dominant role in the pathogenesis of osteoporosis. As a result, early life is a critical phase for establishing lifelong skeletal health, influenced by a multitude of factors (3-5). It is confirmed that maternal factors during pregnancy, such as maternal nutrition, smoking, and alcohol consumption, have been linked to bone health in offspring (6). Thus, the programmed metabolic disorders would be existed throughout lifelong. In the past two decades, the conception of fetal or early life originated diseases had been accepted by investigators. Mechanically, the environmental exposure might result in metabolic and genetic modification dysfunction, leading to long-term impacts, especially in bone and cardiovascular diseases. Importantly, the early life hormonal imbalances can impact bone health (7). Studies have shown that poor nutrition and inadequate growth during childhood can lead to lower peak bone mass, which is a major determinant of bone health later in life (8). Inadequate intake of calcium, vitamin D, and protein during childhood and adolescence can impair bone development. Conditions such as early puberty or delayed puberty may affect peak bone mass. Additionally, endocrine disorders in childhood, such as diabetes or thyroid disorders, can influence bone development (9, 10). Moreover, longitudinal epidemiological studies have provided insights into the long-term effects of early life exposures on bone health. These studies track individuals over time and assess how early life factors influence the risk of osteoporosis in adulthood (11).

The hormone insulin-like growth factor (IGF) family comprises two ligands (IGF-1 and IGF-2), two receptors (IGF-1R and IGF-2R), seven high-affinity binding proteins (IGFBPs 1-7), a substantial group of IGFBP proteases, and a novel category of proteins known as low-affinity IGFBP-related proteins (IGFBP-rPs) (12). This family is widely recognized for its pivotal roles in growth and development, regulating processes such as proliferation, differentiation, metabolism, and cell survival across various tissues, including bone development and hemostasis (13). Among them, CCN1 (also named IGFBP-10), CCN2 (IGFBP-8) and CCN3 (IGFBP-9) which were identified as low-affinity IGFBPs, along with those high-affinity IGFBPs 1-6, together constitute an IGFBP superfamily whose products function in IGF-dependent or IGFindependent modes to regulate skeletal metabolism (13). Fundamental research suggests that the IGF regulatory system plays crucial roles in bone acquisition and maintenance (14, 15). Nevertheless, observational studies and randomized controlled trials examining the association between the IGF regulatory system and osteoporosis (or fracture risk) have yielded conflicting results (16-24). Growth hormone and IGF family members had been identified to be served as important regulators in bone remodeling and metabolism, while the fundamental role had been well established in bone mass maintenance. Previous studies introduced that the impairment of IGF family members would increase the risk of fractures (15, 25, 26). Besides, several single nucleotide polymorphisms had been identified to be involved in osteoporosis patients (15, 25, 26). Moreover, some FDA approved medication had been issued for particular treatment, some integrative therapeutic management would be evaluated since the causal effect between IGF family and osteoporosis was addressed.

If a particular plasma biomarker is directly involved in an underlying pathological process, then inherited variation changing plasma concentrations of this biomarker should affect risk of disease in the direction and magnitude predicted by the plasma concentrations (27). Mendelian randomization (MR) is a valuable analytical approach for establishing causal links between exposures and specific outcomes, which is just such an analytical method to reach a causal inference between a genetically predicted exposure and an outcome, which uses genetic variants that are strongly and solely associated with exposure as instrumental variables (IVs) hence avoiding confounding factors and reverse causality. In this study, we employed a two-sample MR analysis to explore a potential causal relationship between genetically predicted serum IGF system components and osteoporosis.

Methods

Study design

This study was design to assess the causal effects of IGF family members in the risk of osteoporosis (OP). The related traits of IGF family members had been identified, and fourteen IGF family members traits included: IGF-1, IGF-1 sR, IGF-2R, IGFBP-1, IGFBP-2, IGFBP-3, IGFBP-4, IGFBP-5, IGFBP-6, IGFBP-7, IGF-LR1, CTGF, WISP-1 and CYR61. First, the effects of fourteen IGF family members and their serum concentration were evaluated to identify the potential Single nucleotide polymorphisms (SNPs) as one sample MR analysis. Then two-sample MR analysis had been completed among OP traits to measure the causal effects of IGF family members in OP pathogenesis in the large sample size trait (finn-b-M13_OSTEOPOROSIS and ukb-a-87). Then, further confirmation had been performed among two OP traits to validate the results.

Outcome data sources

Data for the genetic associations on osteoporosis were obtained from the publicly available GWAS summary datasets, the FinnGen biobank (Risteys FinnGen R6 - M13_OSTEOPOROSIS) and the Neale lab secondary analysis of UK Biobank phenotypes (https:// pheweb.org/UKB-Neale/pheno/20002_1309), which had no sample overlap with each other. GWAS data of osteoporosis from the FinnGen biobank consisted of 212,778 Finnish-descent individuals (3,203 cases and 209,575 controls). Osteoporosis here was determined from hospital episode statistics, including osteoporosis (further divided into postmenopausal, postoophorectomy, drug-induced, idiopathic osteoporosis and so on) with pathological fracture, osteoporosis without pathological fracture and osteoporosis in other diseases (e.g., multiple myelomatosis, endocrine disorders), all of which were classified by the International Statistical Classification of Diseases and Related Health Problems (ICD) 10 codes. Another osteoporosis GWAS obtained from the UK Biobank involved 337,159 European-descent individuals (5,266 cases and 331,893 controls) for self-reported osteoporosis without cancer illness. This study only utilized publicly available summarized results from published genome-wide association studies. No individual-level data were involved.

Genetic instrument selection

Single nucleotide polymorphisms (SNPs) associated with IGF system members were obtained and selected from the summary

statistics of the genome-wide association study (GWAS) of 14 IGF system members in 4,301 participants of European descent from two cohort studies, the KORA study (28) and the INTERVAL study (29). Genetic associations were adjusted for relevant covariates. We extracted SNPs that strongly predicted exposures at the genomewide significance threshold (P < 1.00E-5) as instrument variants, then clumped based on 1,000 Genomes Project linkage disequilibrium (LD) structure to omit the superposition effect of correlated SNPs (R2 < 0.01 with any other associated SNP within 5Mb).

Statistical analysis

We hypothesized that there exist causal association (whether inverse or positive) of genetically predicted serum IGF system members with osteoporosis. To obtain a reliable foundation for the MR analysis, the following assumptions were satisfied: the genetic variants used as instrumental variables were associated with the exposure; the genetic variants were not associated with any confounders; and the genetic variants were associated with osteoporosis through the exposure (serum IGF system members) only.

In this study, we used TwoSampleMR packages (version 0.5.6) in R (version 4.0.4) to estimate the effect of each IGF system member on osteoporosis by applying MR Egger regression (30), inverse variance weighted (IVW) (31) and weighted median methods (32). And the codes used for R were available in Supplementary file 1. Odds ratios (OR) and 95% confidence intervals (CIs) for osteoporosis were estimated, and a P < 0.05 was considered as statistically significant. To avoid weak IVs, average SNP-specific F-statistics were calculated, and IVs with Fstatistics > 10 were considered as strong IVs for MR analysis. All the results of F-statistics and P values for included SNPs had been listed in Supplementary Table 1. Regarding directional pleiotropy analyses, we conducted MR-Egger regression methods to evaluate the possible pleiotropic effect by the intercept in MR-Egger regression model (P for intercept < 0.05) (30). Heterogeneity was tested for by applying Cochran's Q test on the IVW and MR-Egger estimates. We also performed the leave-one-out analysis with the IVW method to evaluate whether the overall estimate was driven by single SNP.

Results

In accordance with our study design strategy, we investigated the potential causal effects of serum IGF family members' concentrations on the risk of osteoporosis. We included fourteen molecules in the initial one-sample MR analysis to identify SNPs that might influence their serum concentrations. These molecules were IGF-1 (prot-c-2952_75_2), IGF-1 sR (prot-c-4232_19_2), IGF-2R (prot-c-3676_15_3), IGFBP-1 (prot-c-2771_35_2), IGFBP-2 (prot-c-2570_72_5), IGFBP-3 (prot-c-2571_12_3), IGFBP-4 (prot-c-2950_57_2), IGFBP-5 (prot-c-2685_21_2), IGFBP-6 (prot-c-2686_67_2), IGFBP-7 (prot-c-3320_49_2), IGF LR1 (prot-a-1455), CTGF (prot-c-2975_19_2), WISP-1 (prot-c-3057_55_1), and CYR61 (prot-a-758).

Out of these fourteen serum concentration traits related to IGF family members, which had previously been substantiated in published studies, thirteen of them displayed more than one genome-wide significant SNP site. Further details, including the outcomes of the clumping process for LD-independent SNPs related to the exposure, are provided in the supplementary figures. Notably, all calculated F-statistics exceeded a value of ten, indicating that the results were less susceptible to the bias associated with weak instruments. The essential information regarding the enrolled traits has been summarized in Table 1.

In the initial one-step MR analysis, we employed both the MR-Egger and IVW methods. Subsequently, we identified multiple SNPs that reached genome-wide significance (P<1×10-5) among the fourteen IGF family molecules, which were employed to assess their causal effects on osteoporosis. Upon pooling the data, three IGF family molecules were found to be associated with osteoporosis. IGF-1 level is associated with a reduced risk of osteoporosis, as indicated by the weighted median method (odds ratio [OR] = 0.998, 95% CI = 0.997-1.000, P = 0.032) (Figure 1). Furthermore, higher levels of IGFBP-3 were also associated with a decreased risk of osteoporosis using the IVW method (OR = 0.999, 95% CI = 0.998-1.000, P = 0.019) (Figure 1). Additionally, the CTGF level exhibited a negative association with osteoporosis, according to the weighted median method (OR = 0.998, 95% CI = 0.0998, P = 0.004)

TABLE 1 All GWAS datasets selected in this	article.
--	----------

Trait	GWAS id	Sample size	Number of SNPs
Non-cancer illness code self- reported: osteoporosis	ukb-a-87	337,159	10,894,596
Osteoporosis	finn- b- M13_OSTEOPOROSIS	212,778	16,380,452
IGF-1	prot-c-2952_75_2	1,000	501,428
IGF-1 sR	prot-c-4232_19_2	1,000	501,428
IGF-2R	prot-c-3676_15_3	1,000	501,428
IGF-LR1	prot-a-1455	3,301	10,534,735
IGFBP-1	prot-c-2771_35_2	1,000	501,428
IGFBP-2	prot-c-2570_72_5	1,000	501,428
IGFBP-3	prot-c-2571_12_3	1,000	501,428
IGFBP-4	prot-c-2950_57_2	1,000	501,428
IGFBP-5	prot-c-2685_21_2	1,000	501,428
IGFBP-6	prot-c-2686_67_2	1,000	501,428
IGFBP-7	prot-c-3320_49_2	1,000	501,428
CTGF	prot-c-2975_19_2	1,000	501,428
WISP-1	prot-c-3057_55_1	1,000	501,428
CYR61	prot-a-758	3,301	10,534,735

(Figure 1). We conducted a series of sensitivity analyses to validate the causal associations between each trait and osteoporosis. Heterogeneity was not detected for IGF-1 (MR Egger, Cochran P value = 0.449; IVW, Cochran P value = 0.585) and IGFBP-3 (MR Egger, Cochran P value = 0.978; IVW, Cochran P value = 0.961). However, there appeared to be moderate heterogeneity in the analysis of CTGF (MR Egger, Cochran P value = 0.027; IVW, Cochran P value = 0.049). Notably, we did not find any evidence of potential horizontal pleiotropy for IGF-1, CTGF, and IGFBP-3 (intercepts = -0.000, P = 0.814; intercept = -3.242E-5, P = 0.977; intercept = -0.000, P = 0.377, respectively) when employing the MR-Egger method. Leave-one-out analyses for these two traits suggested that the estimated causal effects were not significantly influenced by any single instrumental variable. Scatter plots depicting the MR analyses of the causal effects of IGFs on osteoporosis with statistical significance are presented in Figure 2 (A for IGF-1, B for IGFBP-3, and C for CTGF, respectively). All the involved funnel plots, scatter plots and "leave-one out analysis" plots in assessing the association between IGFs family and osteoporosis in UK trait were shown in Supplementary file 2.

Moreover, in our analysis using data from the FinnGen consortium, we also explored the associations of other IGF family members with osteoporosis. The IGF-LR1 level was negatively associated with osteoporosis according to the MR-Egger method (OR = 0.886, 95% CI = 0.795-0.987, P = 0.036) (Figure 3). On the other hand, CYR61 was linked to an increased risk of osteoporosis based on both the weighted median and IVW methods (OR = 1.154, 95% CI = 1.009-1.319, P = 0.037, and OR = 1.115, 95% CI = 1.022-1.215, P = 0.014, respectively) (Figure 3). We observed no heterogeneity of effects for IGF-LR1 (MR Egger, Cochran P value = 0.702; IVW, Cochran P value = 0.297) and CYR61 (MR Egger, Cochran P value = 0.709; IVW, Cochran P value = 0.735) when utilizing Cochran's Q test. Additionally, no clear evidence of horizontal pleiotropy was found for CYR61 (intercept = -0.012, P = 0.499). However, we did detect possible pleiotropy for IGF-LR1 (intercept = 0.044, P = 0.004). The results remained robust as indicated by the leave-one-out analysis. Scatter plots illustrating the MR analyses of the causal effects of IGFs on osteoporosis with statistical significance are presented in Figure 4 (A for IGF-LR1, and B for CYR61, respectively). MR Egger regression tests suggested no significant horizontal pleiotropy in this part. All the involved funnel plots, scatter plots and "leave-one out analysis" plots in assessing the association between IGFs family and osteoporosis in FinnGen trait were shown in Supplementary file 3.

Discussion

This MR study conducted a comprehensive assessment of the causal relationship between genetically predicted IGF system members and osteoporosis, utilizing extensive GWAS summary statistics. Our findings indicate that genetically proxied higher serum levels of IGF-1, IGFBP-3, IGF-LR1, and CTGF, as well as lower levels of CYR61, are associated with a decreased risk of osteoporosis. However, it's worth noting that these results did not consistently withstand all sensitivity analyses.

Exposure IGF-1	e Method	nSNP	b	se		p OR (95% CI)
	MR Egger	6	-0.001	0.002		0.700 0.999 (0.996 - 1.003)
	Weighted median	6	-0.002	0.001	H B	0.032 0.998 (0.997 - 1.000)
	Inverse variance weighted	6	-0.001	0.001		0.052 0.999 (0.998 - 1.000)
IGF-1 sR						
	MR Egger	4	0.000	0.002	-	0.943 1.000 (0.997 - 1.003)
	Weighted median	4	0.001	0.001	⊢-≣ +	0.430 1.001 (0.999 - 1.002)
	Inverse variance weighted	4	0.001	0.001	⊨ +	0.438 1.001 (0.999 - 1.002)
IGF-2R						
	MR Egger	6	0.000	0.001	⊢	0.749 1.000 (0.998 - 1.001)
	Weighted median	6	0.000	0.000		0.475 1.000 (0.999 - 1.001)
	Inverse variance weighted	6	0.000	0.000	.	0.657 1.000 (0.999 - 1.001)
IGFLR1	· ·					
	MR Egger	33	-0.001	0.001		0.247 0.999 (0.998 - 1.000)
	Weighted median	33		0.001		0.367 1.000 (0.998 - 1.001)
	Inverse variance weighted			0.000		0.379 1.000 (0.999 - 1.000)
IGFBP-1					T	,
	MR Egger	4	0.002	0.003		0.560 1.002 (0.996 - 1.008)
	Weighted median	4		0.001		0.639 1.001 (0.998 - 1.003)
	Inverse variance weighted	4		0.001		0.341 1.001 (0.999 - 1.003)
IGFBP-2	myerse vanance welghteu	-	0.001	0.001		0.041 1.001 (0.000 - 1.003)
IGI DF-2	MR Egger	4	-0.005	0.004		0.340 0.995 (0.988 - 1.003)
	WR Egger Weighted median	4		0.004		0.340 0.995 (0.988 - 1.003) 0.567 1.001 (0.998 - 1.003)
	Inverse variance weighted	4		0.001		0.832 1.000 (0.998 - 1.003)
10555	inverse variance weighted	4	0.000	0.001		0.832 1.000 (0.998 - 1.002)
IGFBP-3		9	0.000	0.004		0.444 0.000 (0.005 . 4.000)
	MR Egger	-		0.001		0.144 0.998 (0.995 - 1.000)
	Weighted median	9		0.001	-	0.086 0.999 (0.998 - 1.000)
	Inverse variance weighted	9	-0.001	0.000		0.019 0.999 (0.998 - 1.000)
IGFBP-4						
	Inverse variance weighted	2	0.001	0.001		0.414 1.001 (0.999 - 1.003)
IGFBP-5						
	MR Egger	4		0.003		0.737 1.001 (0.996 - 1.006)
	Weighted median	4		0.001		0.398 1.001 (0.999 - 1.003)
	Inverse variance weighted	4	0.000	0.001	* * -*	0.598 1.000 (0.999 - 1.002)
IGFBP-6					_	
	MR Egger	3		0.002	┝╾╪╋╾┥	0.629 1.002 (0.997 - 1.006)
	Weighted median	3		0.001	► -	0.594 1.001 (0.998 - 1.003)
	Inverse variance weighted	3	0.001	0.001	+- - -	0.495 1.001 (0.999 - 1.002)
IGFBP-7						
	MR Egger	4		0.003		0.841 0.999 (0.993 - 1.006)
	Weighted median	4		0.001	⊢ ∎+	0.167 0.999 (0.997 - 1.000)
	Inverse variance weighted	4	0.000	0.001	F=-	0.978 1.000 (0.998 - 1.002)
CYR61						
	MR Egger	25		0.001		0.898 1.000 (0.997 - 1.003)
	Weighted median	25	0.000		+− 1	0.902 1.000 (0.999 - 1.002)
	Inverse variance weighted	25	0.000	0.001	+ + •	0.615 1.000 (0.999 - 1.001)
CTGF						
	MR Egger	7	-0.001	0.004		0.757 0.999 (0.990 - 1.007)
	Weighted median	7	-0.002	0.001		0.004 0.998 (0.996 - 0.999)
	Inverse variance weighted	7	-0.002	0.001	H E	0.051 0.998 (0.997 - 1.000)
WISP-1						
	MR Egger	3	0.000	0.005	·+	0.972 1.000 (0.990 - 1.010)
	Weighted median	3	0.000	0.001	·••	0.700 1.000 (0.999 - 1.002)
	Inverse variance weighted	3	0.000	0.001	→ →	0.601 1.000 (0.999 - 1.002)
				0.975	1	1.025
				0.010	The Estimates	

IGF-1, which reaches its peak during puberty, a critical period for acquiring peak bone mass (33), has been extensively studied in relation to bone strength and, consequently, osteoporosis. In our analysis using data from the UK trait, IGF-1 level is associated with a reduced risk of osteoporosis, as indicated by the weighted median method (OR=0.998, 95% CI = 0.997-1.000, P = 0.032). While analyzing data from the FinnGen consortium, the OR value of IGF-1 failed to reach a significant association between IGF-1 and osteoporosis with all p value above 0.05. Although the result from UK trait demonstrated the IGF-1 level is associated with reduced risk, but the 95%CI is near 1.00. Thus, we considered there is only some mild difference between the two traits, which indicates the causal effects of IGF-1 in reducing the prevalence of osteoporosis held population or ethic specificity. Also, we consider the differences in interpreting the results of IGF-1and osteoporosis might be related with the sample sizes and population internal differences of the two datasets, which may include some potential confounding factors, including dietary, living environment, and other genetic background. Observational studies have consistently shown that lower-to-normal circulating levels of IGF-1 correlate with an increased risk of fractures in older individuals. Notably, research involving a sizable cohort of 425 women and 257 men (aged 72–94 years) from the Framingham Heart Study provided compelling evidence that IGF-1 levels are associated with higher BMD in older women, even after adjusting for various confounding variables, including weight, height, protein intake, smoking,



mobility, weight changes, and BMI (17). Similarly, a separate cohort of 2,902 older men also identified low serum IGF-1 levels as a potential risk factor for fractures in older individuals (18). This association mirrors the situation observed in men with idiopathic osteoporosis (IOM), who develop osteoporosis at a young age, further highlighting the potential pathogenic role of IGF-1 in the development of low bone mass (34). Previous whole-genome scan studies in human populations have also suggested a link between the IGF-1 gene and BMD (35, 36). Furthermore, a recent MR study aligns with the notion that IGF-1 plays a role in fracture prevention (37). In our own findings, we observed that a higher circulating level of IGF-1 is associated with a decreased risk of osteoporosis (OR = 0.998, 95% CI = 0.997–1.000, P = 0.032), in line with previous research indicating that elevated IGF-1 levels may serve as a protective factor against osteoporosis.

IGFBP-3 represents another member of the IGF system that exhibited a negative association with the risk of osteoporosis (OR = 0.999, 95% CI = 0.998-1.000, P = 0.019) in UK biobank. While the results from FinnGen dataset did not find a clear association between IGFBP-3 and osteoporosis. Notably, IGFBP-3 is a prominent component of the circulating IGF complex (38). Much like other IGF binding proteins, it plays a role in modulating skeletal growth and metabolism by regulating the access of IGFs to their receptors (14). In vitro studies have shown that IGFBP-3 can either inhibit or stimulate IGF activity, the latter by enhancing the delivery of IGF-1 to cell surface receptors (38). Consistent with our findings, existing evidence suggests that IGFBP-3 may have a positive impact on bone formation. It has been demonstrated to participate in the storage of IGFs within the skeletal matrix by binding to the type I collagen molecule (39). Additionally, individuals with growth hormone deficiency who exhibit low serum levels of IGF-1 and IGFBP-3 tend to have reduced BMD and a significantly higher risk of osteoporotic fractures (40). Furthermore, concurrent administration of IGF-1 and IGFBP-3 has been shown to stimulate bone growth (41). Circulating IGF-LR1 was also identified as having a negative association with the risk of osteoporosis (OR = 0.886, 95% CI = 0.795-0.987, P = 0.036) in FinnGen consortium. IGF-LR1 is a gene that encodes a protein and is situated on chromosome 19. It exhibits widespread expression in organs such as lymph nodes, spleen, and kidney (42). While our literature review indicated limited observational studies on the relationship between IGF-LR1 and osteoporosis, it suggests the need for further investigation in this area. And the molecular function of IGF-LR1 presented population specificity as controversial results identified between UK and FinnGen biobanks.

Similarly, CTGF/CCN2 displayed an inverse association with osteoporosis (OR = 0.998, 95% CI = 0.996-0.999, P = 0.004) in UK biobank, which failed to demonstrate consistent result in FinnGen dataset. CTGF, which is expressed and secreted by osteoblasts during their proliferation, differentiation, bone formation, and fracture healing processes, has been shown to play a regulatory role in osteogenesis within osteoblasts (43, 44). For instance, Kawaki and colleagues reported that the knockout of CTGF in osteoblasts led to delayed osteoblast maturation and mineralization in cultured osteoblasts (45). These findings, in conjunction with our results, suggest that CTGF may serve as a protective factor against osteoporosis. Conversely, cysteine-rich protein 61 (CYR61/CCN1) was the sole member that we identified as being associated with an increased risk of osteoporosis (OR = 1.115, 95% CI = 1.022-1.215, P = 0.014). However, it's noteworthy that most basic research (46-48)has reported CYR61 to have a bone-stimulating effect, which appears to contrast with our findings. Therefore, future studies of higher quality investigating the relationship between CYR61 and osteoporosis may be warranted to reconcile these seemingly contradictory results.

This analysis possesses several notable strengths, primarily stemming from the MR design, which effectively mitigated bias associated with residual confounding, thereby bolstering the causal inference. Furthermore, the study benefits from the extensive availability of GWAS data on IGF system members and osteoporosis, rendering it a well-powered investigation for elucidating the observational relationship between IGF system members and osteoporosis. However, it is crucial to acknowledge certain limitations within these analyses. Firstly, a higher significance

Exposure IGF-1	Method	nSNP	b	se		p OR (95% Cl)
	MR Egger		0.407			0.143 1.503 (0.969 - 2.330)
	Weighted median	6	0.053		H - H	0.456 1.055 (0.917 - 1.213)
	Inverse variance weighted	6	0.071	0.059	•	0.231 1.073 (0.956 - 1.205)
IGF-1 sR						
	MR Egger	4	0.155		▶ 	0.744 1.168 (0.519 - 2.628)
	Weighted median	4	-0.064		H H -1	0.547 0.938 (0.761 - 1.156)
	Inverse variance weighted	4	-0.069	0.147	H R -1	0.642 0.934 (0.699 - 1.247)
IGF-2R						
	MR Egger		0.003		H # H	0.969 1.003 (0.864 - 1.165)
	Weighted median	6	-0.027		•	0.497 0.973 (0.900 - 1.053)
	Inverse variance weighted	6	-0.008	0.038	•	0.822 0.992 (0.921 - 1.068)
IGFLR1						
	MR Egger		-0.121		•	0.036 0.886 (0.795 - 0.987)
	Weighted median		-0.013		•	0.775 0.987 (0.904 - 1.078)
	Inverse variance weighted	35	0.023	0.032	•	0.474 1.023 (0.961 - 1.089)
IGFBP-1						
	MR Egger	4	0.395		·	0.325 1.484 (0.816 - 2.699)
	Weighted median	4	-0.048	0.099	H H H	0.625 0.953 (0.785 - 1.156)
	Inverse variance weighted	4	-0.021	0.084	H#H	0.804 0.979 (0.831 - 1.154)
IGFBP-2						
	MR Egger	4	0.069	0.370	▶ 	0.869 1.072 (0.519 - 2.213)
	Weighted median	4	-0.018	0.091	H H H	0.840 0.982 (0.822 - 1.173)
	Inverse variance weighted	4	0.036	0.083	⊢= →	0.664 1.036 (0.882 - 1.218)
IGFBP-3						
	MR Egger	9	0.175	0.127	 ∎1	0.212 1.191 (0.928 - 1.528)
	Weighted median	9	0.025	0.054	-	0.646 1.025 (0.922 - 1.140)
	Inverse variance weighted	9	0.022	0.042	•	0.596 1.022 (0.942 - 1.110)
IGFBP-4						
	Inverse variance weighted	2	0.203	0.119	⊷ ∎1	0.088 1.225 (0.971 - 1.547)
IGFBP-5						
	MR Egger	4	-0.272	0.213	⊷ ∎⊷•	0.330 0.762 (0.502 - 1.157)
	Weighted median	4	-0.072	0.083	Hand I have a second se	0.386 0.930 (0.790 - 1.095)
	Inverse variance weighted	4	-0.089	0.071		0.213 0.915 (0.796 - 1.052)
IGFBP-6						
	MR Egger	3	-0.083	0.351		0.851 0.920 (0.463 - 1.829)
	Weighted median	3	-0.051		Hand I have a second se	0.551 0.951 (0.805 - 1.123)
	Inverse variance weighted	3	-0.012	0.103	H H -1	0.909 0.988 (0.807 - 1.210)
IGFBP-7	0.000					· · · · ·
	MR Egger	4	-0.215	0.200	⊢∎ , −	0.395 0.807 (0.545 - 1.194)
	Weighted median		-0.115		-	0.065 0.892 (0.789 - 1.007)
	Inverse variance weighted	4	-0.073		-	0.229 0.930 (0.825 - 1.047)
CYR61	0.000					
	MR Egger	22	0.162	0.089	⊢ ∎-1	0.084 1.176 (0.987 - 1.400)
	Weighted median		0.143			0.037 1.154 (1.009 - 1.319)
	Inverse variance weighted		0.109			0.014 1.115 (1.022 - 1.215)
CTGF						
	MR Egger	7	-0.136	0.506	·	0.798 0.872 (0.323 - 2.353)
	Weighted median	7	0.068			0.410 1.070 (0.910 - 1.259)
	Inverse variance weighted	7	0.050		HHH	0.516 1.051 (0.904 - 1.221)
WISP-1			5.000		Γ	oloro ficori (oloci filmati)
	MR Egger	3	1.027	0.459		0.268 2.791 (1.135 - 6.863)
	Weighted median		-0.012			0.885 0.988 (0.843 - 1.158)
	Inverse variance weighted		-0.012			0.895 0.986 (0.798 - 1.218)
		5	0.014	Г		
				0	1 2 3 4 The Estimates	5 6 7
					s and osteoporosis from the Fin	

threshold of P < 1.00E-5 was employed for the selection of SNPs from the GWAS datasets on IGF family members. This decision was necessitated by the limited number of members with at least one genome-wide significant SNP when using P < 5.00E-8 as the threshold. Additionally, 13 of these members exhibited more than two genome-wide significant SNPs when P < 1.00E-5. Secondly, the precise functionality of the ultimately selected SNPs remains unclear. This lack of clarity implies that these genetic variants have not been definitively established as biologically linked to the exposure (IGF family members), introducing a potential source of bias in causal estimates. Also, the GWAS summary datasets for osteoporosis were derived from two published studies that did not account for specific factors such as age, sex, and BMI. The presence of population stratification and imbalances in age and gender between the two GWASs may introduce additional sources of bias. Moreover, it's important to note that the MR results offer insight into the direction of how concentrations of IGF system components may influence the risk of osteoporosis, but they do not provide a specific magnitude. This limitation yields a relatively broad causal inference.

Conclusion

In conclusion, our MR study provides support for causal relationships between five IGF system members and the development of osteoporosis. Specifically, our findings indicate that



lower levels of IGF-1, IGFBP-3, IGF-LR1, and CTGF, along with higher levels of CYR61, genetically contribute to an increased risk of osteoporosis. However, controversial results had been identified between different datasets indicating population specific causal effects between IGF family and osteoporosis. Although the results from both datasets demonstrated that IGF family involved in the pathogenesis of osteoporosis, but the responding key molecules might be various among different population. Further research is warranted to elucidate the underlying mechanisms governing these associations. Essentially, our results suggest that IGF system members, particularly IGF-1, may serve as potential predictive markers during early growth stages and represent promising therapeutic candidates for addressing osteoporosis in validated regional individuals.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.

Author contributions

YL: Conceptualization, Data curation, Funding acquisition, Investigation, Methodology, Project administration, Supervision, Validation, Writing – review & editing. JT: Conceptualization, Formal Analysis, Investigation, Methodology, Software, Validation, Writing – original draft. CZ: Data curation, Formal Analysis, Investigation, Methodology, Writing – original draft. SL: Investigation, Methodology, Software, Writing – original draft. XL: Formal Analysis, Investigation, Methodology, Writing – original draft. BZ: Conceptualization, Investigation, Project administration, Supervision, Validation, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by grants from Technology Project of Sichuan Province of China (2021YFQ0061) and the National Natural Science Foundation of China (82270249). The funding did not participate in the design of the study and collection, analysis, and interpretation of data and in writing the manuscript.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fendo.2023.1332803/ full#supplementary-material

SUPPLEMENTARY TABLE 1 The results of F-statistics and P values for included SNPs.

References

1. Pisani P, Renna MD, Conversano F, Casciaro E, Di Paola M, Quarta E, et al. Major osteoporotic fragility fractures: Risk factor updates and societal impact. *World J Orthop* (2016) 7(3):171–81. doi: 10.5312/wjo.v7.i3.171

2. Ensrud KE, Crandall CJ. Osteoporosis. Ann Intern Med (2017) 167(3):Itc17-itc32. doi: 10.7326/aitc201708010

3. NIH Consensus Development Panel on Osteoporosis Prevention, Diagnosis, and Therapy. Osteoporosis prevention, diagnosis, and therapy. *Jama* (2001) 285(6):785–95. doi: 10.1001/jama.285.6.785

4. Heaney RP, Abrams S, Dawson-Hughes B, Looker A, Marcus R, Matkovic V, et al. Peak bone mass. *Osteoporos Int* (2000) 11(12):985–1009. doi: 10.1007/s001980070020

5. van der Eerden BC, Karperien M, Wit JM. Systemic and local regulation of the growth plate. *Endocr Rev* (2003) 24(6):782–801. doi: 10.1210/er.2002-0033

6. Masztalerz-Kozubek D, Zielinska-Pukos MA, Hamulka J. Maternal diet, nutritional status, and birth-related factors influencing offspring's bone mineral density: A narrative review of observational, cohort, and randomized controlled trials. *Nutrients* (2021) 13(7):2302. doi: 10.3390/nu13072302

7. Riggs BL, Khosla S, Melton LJ. Sex steroids and the construction and conservation of the adult skeleton. *Endocrine Rev* (2002) 23(3):279–302. doi: 10.1210/edrv.23.3.0465

8. Rizzoli R, Biver E, Brennan-Speranza TC. Nutritional intake and bone health. Lancet Diabetes Endocrinol (2021) 9(9):606-21. doi: 10.1016/S2213-8587(21)00119-4

9. Williams GR, Bassett JHD. Thyroid diseases and bone health. *J Endocrinol Invest* (2018) 41(1):eng. 5.4. doi: 10.1007/s40618-017-0753-4

10. Kindler JM, Kelly A, Khoury PR, Levitt Katz LE, Urbina EM, Zemel BS. Bone mass and density in youth with type 2 diabetes, obesity, and healthy weight. *Diabetes Care* (2020) 43(10):2544–2552. eng. 16.2. doi: 10.2337/dc19-2164

11. Rubio-Gutierrez JC, Mendez-Hernández P, Guéguen Y, Galichon P, Tamayo-Ortiz M, Haupt K, et al. Overview of traditional and environmental factors related to bone health. *Environ Sci pollut Res Int* (2022) 29(21):31042–31058. eng. 5.8. doi: 10.1007/s11356-022-19024-1

12. LeRoith D, Holly JMP, Forbes BE. Insulin-like growth factors: Ligands, binding proteins, and receptors. *Mol Metab* (2021) 52:101245. eng. 8.1. doi: 10.1016/j.molmet.2021.101245

13. Kim HS, Nagalla SR, Oh Y, Wilson E, Roberts CTJr., Rosenfeld RG. Identification of a family of low-affinity insulin-like growth factor binding proteins (IGFBPs): characterization of connective tissue growth factor as a member of the IGFBP superfamily. *Proc Natl Acad Sci U S A* (1997) 94(24):12981–6. doi: 10.1073/pnas.94.24.12981

14. Clemmons DR. Role of IGF binding proteins in regulating metabolism. *Trends Endocrinol Metab* (2016) 27(6):375–91. doi: 10.1016/j.tem.2016.03.019

15. Mazziotti G, Lania AG, Canalis E. Skeletal disorders associated with the growth hormone-insulin-like growth factor 1 axis. *Nat Rev Endocrinol* (2022) 18(6):353–65. doi: 10.1038/s41574-022-00649-8

16. Yamaguchi T, Kanatani M, Yamauchi M, Kaji H, Sugishita T, Baylink DJ, et al. Serum levels of insulin-like growth factor (IGF); IGF-binding proteins-3, -4, and -5; and their relationships to bone mineral density and the risk of vertebral fractures in postmenopausal women. *Calcif Tissue Int* (2006) 78(1):18–24. doi: 10.1007/s00223-005-0163-z

17. Langlois JA, Rosen CJ, Visser M, Hannan MT, Harris T, Wilson PW, et al. Association between insulin-like growth factor I and bone mineral density in older women and men: the Framingham Heart Study. *J Clin Endocrinol Metab* (1998) 83 (12):4257–62. doi: 10.1210/jcem.83.12.5308

18. Ohlsson C, Mellström D, Carlzon D, Orwoll E, Ljunggren O, Karlsson MK, et al. Older men with low serum IGF-1 have an increased risk of incident fractures: the MrOS Sweden study. *J Bone Miner Res* (2011) 26(4):865–72. doi: 10.1002/jbmr.281

19. Garnero P, Sornay-Rendu E, Delmas PD. Low serum IGF-1 and occurrence of osteoporotic fractures in postmenopausal women. *Lancet* (2000) 355(9207):898–9. doi: 10.1016/s0140-6736(99)05463-x

20. Schürch MA, Rizzoli R, Slosman D, Vadas L, Vergnaud P, Bonjour JP. Protein supplements increase serum insulin-like growth factor-I levels and attenuate proximal

SUPPLEMENTARY FILE 1

The codes used for R software in the research.

SUPPLEMENTARY FILE 2

Funnel plots and Leave-one-out sensitive analysis of IGFs on osteoporosis based on UK trait.

SUPPLEMENTARY FILE 3

Funnel plots and Leave-one-out sensitive analysis of IGFs on osteoporosis based on FinnGen trait.

femur bone loss in patients with recent hip fracture. A randomized, double-blind, placebo-controlled trial. *Ann Intern Med* (1998) 128(10):801–9. doi: 10.7326/0003-4819-128-10-199805150-00002

21. Lloyd ME, Hart DJ, Nandra D, McAlindon TE, Wheeler M, Doyle DV, et al. Relation between insulin-like growth factor-I concentrations, osteoarthritis, bone density, and fractures in the general population: the Chingford study. *Ann Rheum Dis* (1996) 55(12):870–4. doi: 10.1136/ard.55.12.870

22. Rosen C, Donahue LR, Hunter S, Holick M, Kavookjian H, Kirschenbaum A, et al. The 24/25-kDa serum insulin-like growth factor-binding protein is increased in elderly women with hip and spine fractures. *J Clin Endocrinol Metab* (1992) 74(1):24–7. doi: 10.1210/jcem.74.1.1370164

23. Kassem M, Brixen K, Blum W, Mosekilde L, Eriksen EF. No evidence for reduced spontaneous or growth-hormone-stimulated serum levels of insulin-like growth factor (IGF)-I, IGF-II or IGF binding protein 3 in women with spinal osteoporosis. *Eur J Endocrinol* (1994) 131(2):150–5. doi: 10.1530/eje.0.1310150

24. Friedlander AL, Butterfield GE, Moynihan S, Grillo J, Pollack M, Holloway L, et al. One year of insulin-like growth factor I treatment does not affect bone density, body composition, or psychological measures in postmenopausal women. *J Clin Endocrinol Metab* (2001) 86(4):1496–503. doi: 10.1210/jcem.86.4.7377

25. Canalis E. MANAGEMENT OF ENDOCRINE DISEASE: Novel anabolic treatments for osteoporosis. Eur J Endocrinol (2018) 178(2):R33-r44. doi: 10.1530/eje-17-0920

26. Lara-Castillo N, Johnson ML. Bone-muscle mutual interactions. *Curr Osteoporos Rep* (2020) 18(4):408–21. doi: 10.1007/s11914-020-00602-6

27. Voight BF, Peloso GM, Orho-Melander M, Frikke-Schmidt R, Barbalic M, Jensen MK, et al. Plasma HDL cholesterol and risk of myocardial infarction: a mendelian randomisation study. *Lancet* (2012) 380(9841):572–80. doi: 10.1016/s0140-6736(12)60312-2

28. Suhre K, Arnold M, Bhagwat AM, Cotton RJ, Engelke R, Raffler J, et al. Connecting genetic risk to disease end points through the human blood plasma proteome. *Nat Commun* (2017) 8:14357. doi: 10.1038/ncomms14357

29. Sun BB, Maranville JC, Peters JE, Stacey D, Staley JR, Blackshaw J, et al. Genomic atlas of the human plasma proteome. *Nature* (2018) 558(7708):73–9. doi: 10.1038/s41586-018-0175-2

30. Bowden J, Davey Smith G, Burgess S. Mendelian randomization with invalid instruments: effect estimation and bias detection through Egger regression. *Int J Epidemiol* (2015) 44(2):512–25. doi: 10.1093/ije/dyv080

31. Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. *Genet Epidemiol* (2013) 37 (7):658–65. doi: 10.1002/gepi.21758

32. Hemani G, Zheng J, Elsworth B, Wade KH, Haberland V, Baird D, et al. The MR-Base platform supports systematic causal inference across the human phenome. *Elife* (2018) 7:e34408. doi: 10.7554/eLife.34408

33. Rosen CJ. Insulin-like growth factor I and bone mineral density: experience from animal models and human observational studies. *Best Pract Res Clin Endocrinol Metab* (2004) 18(3):423–35. doi: 10.1016/j.beem.2004.02.007

34. Rosen CJ, Kurland ES, Vereault D, Adler RA, Rackoff PJ, Craig WY, et al. Association between serum insulin growth factor-I (IGF-I) and a simple sequence repeat in IGF-I gene: implications for genetic studies of bone mineral density. *J Clin Endocrinol Metab* (1998) 83(7):2286–90. doi: 10.1210/jcem.83.7.4964

35. Karasik D, Myers RH, Cupples LA, Hannan MT, Gagnon DR, Herbert A, et al. Genome screen for quantitative trait loci contributing to normal variation in bone mineral density: the Framingham Study. *J Bone Miner Res* (2002) 17(9):1718–27. doi: 10.1359/jbmr.2002.17.9.1718

36. Deng HW, Xu FH, Huang QY, Shen H, Deng H, Conway T, et al. A wholegenome linkage scan suggests several genomic regions potentially containing quantitative trait Loci for osteoporosis. *J Clin Endocrinol Metab* (2002) 87(11):5151– 9. doi: 10.1210/jc.2002-020474

37. Yuan S, Wan ZH, Cheng SL, Michaëlsson K, Larsson SC. Insulin-like growth factor-1, bone mineral density, and fracture: A mendelian randomization study. *J Clin Endocrinol Metab* (2021) 106(4):e1552–8. doi: 10.1210/clinem/dgaa963

38. Giustina A, Mazziotti G, Canalis E. Growth hormone, insulin-like growth factors, and the skeleton. *Endocr Rev* (2008) 29(5):535–59. doi: 10.1210/er.2007-0036

39. Kawai M, Rosen CJ. The insulin-like growth factor system in bone: basic and clinical implications. *Endocrinol Metab Clin North Am* (2012) 41(2):323–33, vi. doi: 10.1016/j.ecl.2012.04.013

40. Wüster C, Slenczka E, Ziegler R. [Increased prevalence of osteoporosis and arteriosclerosis in conventionally substituted anterior pituitary insufficiency: need for additional growth hormone substitution]? *Klin Wochenschr* (1991) 69(16):769–73. doi: 10.1007/bf01797616

41. Williams RM, McDonald A, O'Savage M, Dunger DB. Mecasermin rinfabate: rhIGF-I/rhIGFBP-3 complex: iPLEX. *Expert Opin Drug Metab Toxicol* (2008) 4 (3):311–24. doi: 10.1517/17425255.4.3.311

42. Fagerberg L, Hallström BM, Oksvold P, Kampf C, Djureinovic D, Odeberg J, et al. Analysis of the human tissue-specific expression by genome-wide integration of transcriptomics and antibody-based proteomics. *Mol Cell Proteomics* (2014) 13(2):397–406. doi: 10.1074/mcp.M113.035600

43. Mundy C, Gannon M, Popoff SN. Connective tissue growth factor (CTGF/ CCN2) negatively regulates BMP-2 induced osteoblast differentiation and signaling. *J Cell Physiol* (2014) 229(5):672–81. doi: 10.1002/jcp.24491 44. Safadi FF, Xu J, Smock SL, Kanaan RA, Selim AH, Odgren PR, et al. Expression of connective tissue growth factor in bone: its role in osteoblast proliferation and differentiation in vitro and bone formation in vivo. J Cell Physiol (2003) 196(1):51–62. doi: 10.1002/jcp.10319

45. Kawaki H, Kubota S, Suzuki A, Yamada T, Matsumura T, Mandai T, et al. Functional requirement of CCN2 for intramembranous bone formation in embryonic mice. *Biochem Biophys Res Commun* (2008) 366(2):450–6. doi: 10.1016/ j.bbrc.2007.11.155

46. Su JL, Chiou J, Tang CH, Zhao M, Tsai CH, Chen PS, et al. CYR61 regulates BMP-2-dependent osteoblast differentiation through the {alpha}v{beta}3 integrin/ integrin-linked kinase/ERK pathway. *J Biol Chem* (2010) 285(41):31325–36. doi: 10.1074/jbc.M109.087122

47. Si W, Kang Q, Luu HH, Park JK, Luo Q, Song WX, et al. CCN1/Cyr61 is regulated by the canonical Wnt signal and plays an important role in Wnt3A-induced osteoblast differentiation of mesenchymal stem cells. *Mol Cell Biol* (2006) 26(8):2955–64. doi: 10.1128/mcb.26.8.2955-2964.2006

48. Zhao G, Huang BL, Rigueur D, Wang W, Bhoot C, Charles KR, et al. CYR61/ CCN1 regulates sclerostin levels and bone maintenance. *J Bone Miner Res* (2018) 33 (6):1076–89. doi: 10.1002/jbmr.3394