Check for updates

OPEN ACCESS

EDITED BY Yanting Wu, Fudan University, China

REVIEWED BY Ricardo Azziz, University of Alabama at Birmingham, United States Donna F. Vine, University of Alberta, Canada

*CORRESPONDENCE Licy L. Yanes Cardozo Vanes@umc.edu

SPECIALTY SECTION

This article was submitted to Reproduction, a section of the journal Frontiers in Endocrinology

RECEIVED 13 June 2022 ACCEPTED 26 January 2023 PUBLISHED 15 February 2023

CITATION

Pruett JE, Romero DG and Yanes Cardozo LL (2023) Obesity-associated cardiometabolic complications in polycystic ovary syndrome: The potential role of sodiumglucose cotransporter-2 inhibitors. *Front.* Endocrinol. 14:951099. doi: 10.3389/fendo.2023.951099

COPYRIGHT

© 2023 Pruett, Romero and Yanes Cardozo. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Obesity-associated cardiometabolic complications in polycystic ovary syndrome: The potential role of sodium-glucose cotransporter-2 inhibitors

Jacob E. Pruett¹, Damian G. Romero^{1,2,3,4} and Licy L. Yanes Cardozo^{1,2,3,4,5*}

¹Department of Cell and Molecular Biology, University of Mississippi Medical Center, Jackson, MS, United States, ²Mississippi Center of Excellence in Perinatal Research, University of Mississippi Medical Center, Jackson, MS, United States, ³Women's Health Research Center, University of Mississippi Medical Center, Jackson, MS, United States, ⁴Cardiovascular-Renal Research Center, University of Mississippi Medical Center, Jackson, MS, United States, ⁵Department of Medicine, University of Mississippi Medical Center, Jackson, MS, United States

Polycystic Ovary Syndrome (PCOS) is the most common endocrine disorder in reproductive-age women. PCOS is characterized by androgen excess, oligo/ anovulation, and polycystic appearance of the ovaries. Women with PCOS have an increased prevalence of multiple cardiovascular risk factors such as insulin resistance, hypertension, renal injury, and obesity. Unfortunately, there is a lack of effective, evidence-based pharmacotherapeutics to target these cardiometabolic complications. Sodium-glucose cotransporter-2 (SGLT2) inhibitors provide cardiovascular protection in patients with and without type 2 diabetes mellitus. Although the exact mechanisms of how SGLT2 inhibitors confer cardiovascular protection remains unclear, numerous mechanistic hypotheses for this protection include modulation of the renin-angiotensin system and/or the sympathetic nervous system and improvement in mitochondrial function. Data from recent clinical trials and basic research show a potential role for SGLT2 inhibitors in treating obesity-associated cardiometabolic complications in PCOS. This narrative review discusses the mechanisms of the beneficial effect of SGLT2 inhibitors in cardiometabolic diseases in PCOS.

KEYWORDS

polycystic ovary syndrome (PCOS), obesity, SGLT 2 inhibitors, cardiometabolic complications, hypertension, insulin resistance, mitochondrial dysfunction, women's health

Introduction

Polycystic Ovary Syndrome (PCOS) is the most common endocrine disorder in reproductive-age women, affecting 5-20% of this population (1–3). While there has been evidence of women with PCOS since the time of Hippocrates (4), the syndrome was first described by Stein and Leventhal in 1935 in their article "Amenorrhea associated with bilateral polycystic ovaries" (5). Stein and Leventhal described a series of cases focusing on amenorrhea in women with polycystic ovaries as determined by bimanual pelvic exam and/or pneumoroentgenography (5). However, they also noted physical signs of androgen excess in most cases (5). Since the 1930s, our understanding of the disease has evolved. However, the etiology of PCOS remains unknown, and there are still many gaps in our knowledge and disagreements in the field, in part due to three coexisting sets of criteria for PCOS diagnosis (6–8).

PCOS, a diagnosis of exclusion, can be characterized by using three different sets of diagnostic criteria: the National Institutes of Health (NIH) 1990 criteria, the Rotterdam 2003 criteria, and the Androgen Excess-PCOS Society (AE-PCOS) 2006 criteria (6-8) (Figure 1). The NIH and AE-PCOS criteria consider androgen excess a requirement for diagnosis. Androgen excess can be defined as either biochemical or clinical evidence of high levels of androgens (6-8). Therefore, if a woman presents to the clinic with hirsutism (as defined by the Ferriman-Gallwey Scale) (9), excessive acne after puberty, or male-pattern balding, then measurement of serum androgens is not necessary for the PCOS diagnosis per se. However, biochemical testing, including testosterone levels, is required to eliminate other possible causes of androgen excess, as PCOS is a diagnosis of exclusion. Workup to exclude elevated prolactin, Cushing syndrome, non-classic congenital adrenal hyperplasia, thyroid dysfunction, and rogen-producing tumors, and exogenous administration of androgens should be done before conferring the PCOS diagnosis (6-8). In particular, if total serum testosterone is above 200 ng/dL, if dehydroepiandrosterone sulfate is above 800 µg/dL, if signs of excess androgen progress rapidly, or if the voice changes, then the possibility of an androgen-producing tumor in the adrenal gland or ovary should be thoroughly investigated before diagnosing the patient with PCOS (10). Women with PCOS will have elevated levels of androgens, but they should not approach male levels (10, 11). The Rotterdam criteria, the most common criteria used in the clinic, requires the presence of two of the three PCOS' characteristics (Figure 1) generating 4 different phenotypes. As a result, there are women diagnosed with PCOS with and without hyperandrogenism. In recent years, several lines of evidence suggest that women with hyperandrogenic PCOS have worsened cardiovascular profiles (12). Unfortunately, there are no safe and effective therapeutic agents to decrease the levels or block the action of androgens in women. Furthermore, the heterogeneity of the clinical manifestation of PCOS suggests that the involvement of multiple pathophysiological pathways as suggested by GWAS studies (13, 14). Thereby, novel and effective therapeutic agents are needed for safe and effective PCOS clinical management.

Sodium-glucose cotransporter-2 (SGLT2) inhibitors provide cardiovascular protection in patients with and without type 2 diabetes mellitus (T2DM). Although the exact mechanisms of how SGLT2 inhibitors confer cardiovascular protection remains unclear, numerous mechanistic hypotheses have been postulated, including modulation of the renin-angiotensin system and/or the sympathetic nervous system and improvement in mitochondrial function. Data from recent clinical trials and basic research show a potential role for SGLT2 inhibitors in treating obesity-associated cardiometabolic complications targeting those abnormal pathophysiological systems in PCOS. This narrative review discusses the mechanisms of the beneficial effect of SGLT2 inhibitors in cardiometabolic diseases in PCOS.

Cardiovascular disease in PCOS

Cardiovascular disease is the number one cause of death among females in the United States (15). Unfortunately, PCOS is associated with multiple cardiovascular risk factors, such as obesity, hypertension, insulin resistance, dyslipidemia, and renal injury (16–19). While it is clear that women with PCOS have increased prevalence of cardiovascular risk factors, there is debate about whether or not these cardiovascular risk factors translate into increased cardiovascular events (20). In 2006, a study including \sim 11,000 women with PCOS from California (USA) showed no



increased risk of coronary artery disease, cerebrovascular disease, or peripheral arterial disease in PCOS (17). Later, in 2015, a study including ~20,000 Danish women with PCOS also showed no increased risk of cardiovascular disease (21). However, when the same Danish population was reanalyzed broadening the definition of CVD to include hypertension and dyslipidemia, women with PCOS showed a ~2-fold increase in CVD events (22). The negative findings of the American and some of the Danish studies were in spite of both studies showing increased prevalence of cardiovascular risk factors, such as hypertension, dyslipidemia, and T2DM, in women with PCOS (17, 21). Conversely, in 2020, a meta-analysis showed that women with PCOS have an increased risk of cardiovascular disease and stroke (23). Furthermore, in 2021, a study including ~175,000 British women with PCOS demonstrated an increased risk for cardiovascular events, including myocardial infarction, angina, and revascularization in young women with PCOS (24). Participants were matched with controls for body mass index (BMI) on a 1:1 ratio in a total of 350,000 women, giving unprecedented strength to the work of Berni et al. (20, 24). Although there are multiple pharmacological agents used to manage the cardiometabolic complications in PCOS, their safety and effectiveness to prevent or ameliorate cardiovascular disease and mortality in PCOS are limited (25).

Although the etiology of the syndrome remains unknown, hyperandrogenemia may constitute a key mechanism underlying the cardiovascular risk factors in PCOS. We have demonstrated that hyperandrogenemia in female rats elicits several cardiovascular risk factors also present in women with PCOS (Figure 2). More recently, we demonstrated the potential benefit of SGLT2 inhibitors in body composition and blood pressure in such PCOS experimental model. Therefore, novel pharmacotherapies, such as sodium-glucose cotransporter-2 inhibitors, could simultaneously target multiple mechanisms of the pathophysiology of the cardiometabolic complications associated with PCOS.

SGLT2 inhibitors: Discovery and use beyond type 2 diabetes mellitus

Discovery of SGLT2 and its function

As the kidney filters blood, it must exercise precise, selective control over the reabsorption of electrolytes, carbohydrates, and amino acids while excreting waste products like urea. In the 1930s, with experiments from Shannon and Fisher, glucose in the mammalian nephron was shown to go "into reversible combination with some element in the tubule cells, present in constant but limited amount," and this mystery element was proposed to be the ratelimiting step of glucose reabsorption from the nephron (26). In the 1970s, Scriver et al. hypothesized that there might be a G1/G2 system to reabsorb hexoses in the kidney, with G1 having a low-capacity, and low affinity for glucose. In contrast, G2 would have a high-capacity, high affinity. Inspired by familial renal glucosuria, Scriver et al. also hypothesized that only the G1 system was present in the intestines while G2 was not (27). Experiments from Turner and Moran in the 1980s gave further evidence for two different sodium-dependent glucose transporters in the proximal tubule of the nephron (28). The G1 and G2 systems were later characterized as low-capacity, high-affinity, or high-capacity, low-affinity and were recognized as SGLT1 and SGLT2, respectively (29). SGLT2 expression was localized to the renal cortex, while SGLT1 was localized to the renal medulla and intestine (29, 30). As SGLT2 has a high capacity for glucose, it was later confirmed that SGLT2 is explicitly responsible for the vast majority of renal glucose reabsorption (31).

Discovery of SGLT2 inhibitors

By inhibiting SGLT2, blood glucose could be lowered independently of insulin via its glucosuric effect, thus minimizing the risk of hypoglycemia observed with other antidiabetic agents (32). However, there were multiple drawbacks to using phlorizin, an early SGLT2 inhibitor naturally found in apple trees (33, 34), such as poor absorption in the gut and its concurrent inhibition of SGLT1 (35). Lack of functioning SGLT1 can lead to diarrhea, as seen with hereditary glucose/galactose malabsorption, so the search began for selective and orally available drugs to inhibit SGLT2 (36). Out of this search came the burgeoning drug class of gliflozins, which were based on the structure of the o-glucoside phlorizin (37, 38). While some gliflozins, such as the o-glucoside sergliflozin, were not successful candidates for managing glucose homeostasis (37), many c-glucosides made it through phase III clinical trials, such as empagliflozin, dapagliflozin, and canagliflozin, which are the SGLT2 inhibitors that are widely used in clinical practice nowadays (38, 39).



SGLT2 inhibitors confer cardiovascular protection in patients with and without diabetes

Around the time that currently available SGLT2 inhibitors were progressing through clinical trials, the United States Food and Drug Administration (FDA) issued an additional requirement for phase III clinical trials of antidiabetic medications for T2DM (40). Because of evidence that thiazolidinediones, PPAR γ agonists that act as insulin sensitizers (41), may increase cardiovascular risk in patients with T2DM even while improving glycemic control, the FDA began recommending that all new antidiabetic medications also needed to demonstrate cardiovascular safety (40, 42).

Intriguingly, with the landmark EMPA-REG study (43), empagliflozin was shown to reduce the risk of cardiovascular death in T2DM, becoming the first antidiabetic medicine proven to do so (40, 43). Additionally, EMPA-REG showed preservation of renal function with empagliflozin (44). Later with the CANVAS (45), DECLARE-TIMI 58 (46), and CREDENCE (47) trials, the SGLT2 inhibitors canagliflozin and dapagliflozin were also shown to decrease the risk of cardiovascular death and renal failure. These benefits have been proposed to be independent of changes in glycemic status (47) as the reduction in blood glucose with SGLT2 inhibition is modest (43, 45, 47), suggesting that the positive benefit of SGLT2 inhibitors could be possible in patients without overt T2DM. Additionally, in terms of side effects, there was no increased risk of hypoglycemia or acute kidney injury with SGLT2 inhibition (43, 45-47). However, there are some notable side effects of this drug class. SGLT2 inhibition does increase the risk of mycotic genital infection (43, 45-47). More rarely, SGLT2 inhibition also increases the risk of diabetic ketoacidosis (46, 47), in particular euglycemic diabetic ketoacidosis (48), often occurring with surgery or illness. Euglycemic diabetic ketoacidosis is a severe and life-threatening complication that can be overlooked by providers because of the normal range blood glucose in this condition, so this is an important consideration for anyone taking or prescribing gliflozins.

More recently, with multiple trials such as EMPEROR-Reduced (49), EMPEROR-Preserved (50), DAPA-HF (51), and DAPA-CKD (52), even in the absence of T2DM, SGLT2 inhibition benefited patients with chronic kidney disease and patients with heart failure (with either reduced or preserved ejection fraction). These findings

implicated a potential role of SGLT2 inhibition in cardiovascular or renal disease for various conditions. However, exactly how SGLT2 inhibition produces this cardiovascular/renal protection independently of its effect on glycemia is uncertain and is an open question. There are multiple hypotheses concerning these mechanisms (53, 54).

SGLT2 inhibition in women with PCOS

PCOS is associated with insulin resistance, obesity, renal injury, mitochondrial dysfunction, and activation of both the Sympathetic Nervous System (SNS) and Renin-Angiotensin System (RAS). SGLT2 inhibitors have demonstrated improvements in all these disease states, suggesting they may be a promising novel therapy to improve women's healthcare in PCOS (Figure 3). Recently three small clinical trials used SGLT2 inhibitors to improve the cardiometabolic complications of patients with PCOS with exciting results (see summary in Table 1). In the trial by Javed et al., empagliflozin decreased body weight, body mass index, and fat mass in overweight women with PCOS compared to metformin (55). However, there was no decrease in insulin resistance or blood pressure, though it should be noted that the patients in this study had normal blood pressure at baseline (55). In the trial by Elkind-Hirsch et al., they explored if there were a synergistic effect between SGLT2 inhibition with dapagliflozin and glucagon-like peptide-1 receptor agonism (GLP-1RA) with exenatide in obese women with PCOS (56). Their data show that dapagliflozin and exenatide have an additive effect to further reduce body weight and fat mass than either drug individually can, which is likely due to their differing mechanisms of action. Combination therapy of SGLT2 and GLP-1RA could constitute a promising therapeutic tool to ameliorate cardiometabolic complications in PCOS women.

More recently, Tan et al. reported that licogliflozin decreased insulin resistance and circulating dehydroepiandrosterone sulfate (DHEAS) in women with PCOS, with a similar tendency in other circulating androgens (57). However, licogliflozin is not yet approved by the FDA and is more promiscuous than other SGLT2 inhibitors, having only a 30-fold selectivity for SGLT2 over SGLT1 (58), instead of the over 2,500-fold selectivity that empagliflozin has for SGLT2 (38). Higher-quality clinical trials are needed to better define the



Trial	SGLT2 Inhibitor	Number of Participants	Duration of Trial	Findings
Javed et al. 2019 (55)	Empagliflozin	39	12 weeks	\downarrow Body weight, BMI, and fat mass
Elkind-Hirsch et al. 2021 (<mark>56</mark>)	Dapagliflozin	92	24 weeks	Additive effect in \downarrow body weight and fat mass with GLP-1RA
Tan et al. 2021 (57)	Licogliflozin	20	2 weeks	↓ Insulin Resistance and DHEAS

TABLE 1 Clinical trials of SGLT2 inhibitors in PCOS.

↓, decrease.

potential role of selective SGLT2 inhibitors in treating the cardiometabolic complications in women with PCOS.

Insulin resistance and SGLT2 inhibition in PCOS

While not part of the diagnostic criteria for PCOS, insulin resistance is frequently present in lean and obese women with PCOS (16, 59). In PCOS patients using the sequential euglycemic insulin clamp technique, insulin infusion leads to elevations in testosterone (60). Mechanistically, insulin and insulin-like growth factors synergize with luteinizing hormone to increase androgen synthesis in ovarian theca cells (61). Furthermore, insulin decreases circulating sex hormone-binding globulin (SHBG) by decreasing its hepatic synthesis, leading to increased circulating free testosterone (61), which can bind and activate the androgen receptor. Moreover, testosterone can be reduced to 5α -dihydrotestosterone (DHT) in peripheral tissues to become the most potent endogenous agonist of the androgen receptor (62). However, short-term androgen administration in women also decreases insulin sensitivity, as demonstrated by both hyperglycemic and euglycemic hyperinsulinemic clamps (63). Furthermore, androgen administration in female rats leads to insulin resistance (64, 65). Thus, it is still unclear if insulin resistance causes androgen excess in women with PCOS or whether androgen excess leads to the constellation of cardiometabolic dysfunctions, including insulin resistance, in women with PCOS.

To ameliorate insulin resistance in PCOS, the first-line therapy for T2DM, metformin, is often used off-label (66, 67). However, there is an ongoing debate whether or not metformin is beneficial for insulin resistance in PCOS patients, as a recent meta-analysis showed that metformin had no impact on fasting blood glucose or insulin in overweight women with PCOS (68, 69). Small short-term clinical trials have shown that the FDA-approved anti-obesity glucagon-like peptide-1 receptor agonists (GLP-1 RAs) improve insulin resistance in PCOS patients (70) and may be superior to metformin (71). Therefore, there is a great need for new therapeutics targeting insulin resistance and T2DM in PCOS.

We previously demonstrated that hyperandrogenic female (HAF) experimental PCOS model generated with female Sprague Dawley rats chronically implanted with subcutaneous DHT pellets (7.5 mg/90 days) exhibit similar cardiometabolic complications to those in women with PCOS. The HAF rats exhibit increased food intake, obesity with an expansion of both subcutaneous and visceral fat

depots, insulin resistance, and elevated blood pressure, that closely mimic the cardiometabolic complications in women with PCOS (65, 72). This review will focus on the HAF cardiometabolic complications; however, there are multiple animal experimental models of PCOS, each one with pros and cons to model the human disease as summarized in recent reviews (73, 74).

We recently demonstrated that SGLT2 inhibition did not improve insulin resistance in the HAF rats, but it did improve adiposity and elevated blood pressure, suggesting that this agent could exhibit cardioprotective effects in women with PCOS. Whether SGLT2 inhibitors could be used to prevent the progression of insulin resistance to diabetes in women with PCOS remains unknown.

Hypertension and SGLT2 inhibition in PCOS

The systemic renin-angiotensin system

Multiple studies have demonstrated an increased prevalence of hypertension in women with PCOS (17, 18). Moreover, blood pressure could be only mildly elevated as demonstrated in 24-hour ambulatory blood pressure measurements in women with PCOS (75). Obesity is associated with hypertension, but women with PCOS, even with normal weight, can have elevated blood pressure (76). The specific mechanism(s) that lead to elevated blood pressure or hypertension in PCOS remain a matter of debate.

One promising mechanism is dysregulation of the reninangiotensin system (77, 78). The RAS plays a significant role in long-term blood pressure control, and it may lead to hypertension when inappropriately activated (79). The systemic RAS begins with the macula densa in the distal portion of the nephron (80). When it senses low sodium, the macula densa releases the enzyme renin from juxtaglomerular cells. Renin will then cleave angiotensinogen, released mainly from the liver, into angiotensin I (Ang I). Angiotensin-converting enzyme (ACE), which is expressed primarily in the lung, will then convert Ang I into angiotensin II (Ang II). Then, Ang II can bind to the Ang II Type 1 Receptor (AT1R) or the Ang II Type 2 Receptor (AT2R) (80, 81). Through AT1R, Ang II constricts the efferent arteriole, causing an increase in glomerular filtration rate to increase sodium delivery to the macula densa. Furthermore, Ang II stimulates aldosterone release by the adrenal gland via AT1R, which will increase sodium reabsorption by principal cells of the collecting duct of the nephron. Alternatively, via AT2R, Ang II causes vasodilation to decrease blood pressure. Furthermore,

Ang II can be converted by angiotensin-converting enzyme 2 (ACE2) into the heptapeptide angiotensin (1–7) that reduces blood pressure and insulin resistance *via* the Mas receptor (81, 82).

The intrarenal renin-angiotensin system

In addition to the systemic RAS, several tissue-specific reninangiotensin systems exist, such as in the kidneys, heart, and adipose tissue (83). We will focus on the intrarenal RAS for this review as SGLT2, an essential player in this work, is predominantly expressed in the kidney but not in the heart or the adipose tissue (84). In the nephron, AT1R lines the lumen of the proximal tubule, distal tubule, and collecting duct (85) and can stimulate angiotensinogen synthesis in the proximal tubule with Ang II-activation (86). Renin is produced in the proximal tubule (87, 88), distal tubule (89), and principal cells of the collecting duct (89), allowing for angiotensin I to be produced within the nephron. Meanwhile, ACE is expressed along the brush border of the proximal tubule (90) and within cells of the collecting duct (91), allowing for Ang I to be produced within the nephron. When Ang II binds to the AT1R of the principal cells of the collecting duct, it increases the activity of epithelial sodium channels, which can further increase blood pressure (86, 91, 92). The detailed mechanism of the regulation of blood pressure by epithelial sodium channels can be found elsewhere (93). Thus, the intrarenal RAS appears to have a positive feedback loop where intratubular Ang II can lead to the formation of more intratubular Ang II, which can lead to increases in blood pressure (86).

Hypertension in PCOS: Targeting the reninangiotensin system

The RAS, a central regulator of blood pressure, is modulated by androgens. Women with PCOS have dysregulation of the RAS with high circulating levels of renin (77, 94), the rate-limiting enzyme of the RAS (95). Plasma ACE2, which converts Ang II into the vasodilator angiotensin (1-7), is also decreased in women with PCOS (94). In a case series of four women with PCOS, treatment with telmisartan, an AT1R blocker, normalized blood pressure, reduced androgen levels, and improved the menstrual cycle (78). In a preclinical hyperandrogenemic female (HAF) rat model of PCOS, mRNA expression of renal angiotensinogen and ACE is increased (65). Furthermore, the ACE inhibitor enalapril reduces blood pressure in aged HAF rats more than in controls (96). However, compensatory alterations in the RAS have been shown in PCOS, as circulating angiotensinogen is decreased in women with PCOS (94). Moreover, intrarenal ACE2 is upregulated while intrarenal renin is downregulated in HAF rat model of PCOS (97). The upregulation of the ACE2 could be a protective mechanism to counteract the activation of the classical arm of the RAS; however, this hypothesis needs to be tested. Altogether, these data implicate the RAS is at least partially responsible for the increased blood pressure observed in PCOS. ACE inhibitors or AT1R blockers are widely used as antihypertensive drugs in the general population. However, due to their potential teratogenic, fetotoxic, and miscarriage-associated risks during pregnancy, ACE inhibitors and AT1R blockers are rarely used in the clinic in PCOS women of reproductive age (98–100). Thereby, agents that impact the RAS safely and effectively are needed to treat hypertension in women with PCOS.

SGLT2 and the renin-angiotensin system

RAS blockers are part of the standard of care for chronic kidney disease (both with and without diabetes mellitus) and heart failure. In landmark clinical trials showing cardiovascular and renal protection by SGLT2 inhibition, most patients were on some form of RAS blocker at baseline (43, 45–47, 49–52). In other words, the benefit of SGLT2 inhibition in clinical trials typically occurred on a background of RAS blockade. How might RAS blockade be working with SGLT2 inhibition, though?

As SGLT2 inhibition decreases sodium reabsorption in the proximal tubule, one would expect that there would be increased sodium delivery to the macula densa, thus reducing renin release and RAS activation. However, what has been found experimentally is more complex. Concerning the first part of the systemic RAS cascade, in a retrospective analysis of patients with hypertension and T2DM, no significant change was observed in plasma renin activity (PRA) with SGLT2 inhibition (101). Meanwhile, diabetic male mice treated with empagliflozin had decreased PRA; however, empagliflozin caused no change in PRA in control mice (102). Furthermore, a small observational study showed in diabetic patients that PRA was initially increased after one month of SGLT2 inhibition but returned to normal after three months of treatment (103). The variability observed in PRA with SGLT2 inhibition could be partly due to volume contraction from SGLT2 inhibition. The glucosuria from SGLT2 inhibition causes osmotic diuresis, which may decrease extracellular volume to trigger the release of renin (53). However, with time, the elevated antidiuretic hormone can compensate for the decrease in volume from SGLT2 inhibition (104), which may restore renin levels to normal.

Renin is far from the only component of the RAS reported to respond to SGLT2 inhibition, and the results are equally as mixed as those of renin. In a mouse model of T2DM, Woods et al. found that renal cortex angiotensinogen mRNA and protein expression was decreased by SGLT2 inhibition; however, renal ACE and AT1R mRNA expression was unchanged (105). Meanwhile, in a rat model of T2DM, Shin et al. found that AT1R protein expression was decreased in the renal cortex with SGLT2 inhibition (106). Furthermore, in diabetic Dahl salt-sensitive rats, SGLT2 inhibition was shown to work synergistically, instead of additively, with ACE inhibition to reduce blood pressure (107). Meanwhile, Bautista et al. found in male rats that Ang II increases renal SGLT2 independent of blood pressure changes and that inhibiting Ang II formation or AT1R decreases renal SGLT2 (108). Therefore, at least in male rodents, SGLT2 inhibitors and RAS blockade may work together synergistically, which may translate to the importance of patients having both types of pharmacotherapies to treat their cardiometabolic disease.

Recently, we reported that SGLT2 inhibition in HAF rats downregulates intrarenal ACE and AT1R mRNA, which was accompanied by a slight decrease in mean arterial pressure (97). However, intrarenal ACE2 mRNA, which is part of the vasodilatory arm of the RAS, was also downregulated by SGLT2 inhibition in HAF rats (97). If the upregulation of the ACE2 in the kidney is a compensatory mechanism to combat the androgen deleterious effect, one can speculate that due to the beneficial effect of SGLT2 inhibitors, this is not further needed. These data suggest that SGLT2 inhibition could work synergistically with RAS blockers to reduce blood pressure in HAF rats, similar as in male rodents (107).

Sympathetic nervous system and SGLT2 in PCOS

Another possible mechanism for hypertension in women with PCOS is an activation of the SNS. Using heart rate variability to measure autonomic dysfunction, women with PCOS matched with controls for body mass index and blood pressure have increased SNS activity and decreased parasympathetic activity (109). Furthermore, adrenergic blockade with terazocin and propranolol in young HAF rats effectively decreases blood pressure (110). Renal denervation, which reduces sympathetic activity in the kidney, also reduces blood pressure in HAF rats and in women with PCOS (110, 111). The SNS may be activated in PCOS women and HAF rats because of elevations in the adipokine leptin in the circulation (65, 112). Leptin secretion by the adipose tissue is upregulated in obesity. When leptin is chronically elevated, it increases blood pressure by stimulating the sympathetic nervous system via melanocortin 4 receptor (MC4R) in proopiomelanocortin (POMC) neurons (113). Activation of the renal SNS can also activate the RAS (113) and promote sodium retention, actions that, if sustained over time, can increase blood pressure. Consequently, it is possible to speculate that adrenergic blockade could reduce RAS activation in PCOS. The detailed mechanism by which MC4R regulates blood pressure has been recently reviewed (114).

Adrenergic blockade is part of the standard of care in particular clinical conditions such as heart failure with reduced ejection fraction (115). To the best of our knowledge, randomized clinical trials with adrenergic blockade have not been yet performed in women with PCOS. However, while used to treat some individuals with essential hypertension, adrenergic blockade is not the gold standard, with some studies showing a lack of cardiovascular protection with this drug class (116, 117). Therefore, while the SNS appears to be upregulated in overweight or obese women with PCOS, whether or not direct adrenergic blockade would be beneficial in attenuating their hypertension is still unclear.

There are some evidence that SGLT2 inhibitors could target the SNS to exhibit their cardioprotective effects. Activation of the SNS leads to vasoconstriction and an increase in heart rate, leading to an increase in blood pressure (80). With the potential volume contraction from the osmotic diuresis caused by SGLT2 inhibition, one would not be surprised to observe a compensatory increase in heart rate. However, data from phase II/III clinical trials in patients with T2DM show that SGLT2 inhibition is associated with a decrease in heart rate (118, 119). Why might that be? A study by Herat and colleagues recently demonstrated in Schlager mice, a model of neurogenic hypertension with sympathetic activation, that SGLT2 inhibition decreases SNS innervation of the kidney, accompanied by a

reduction of renal norepinephrine (120). However, precisely how SGLT2 regulates renal SNS is uncertain. A recent meta-analysis in patients with T2DM demonstrated that SGLT2 inhibition was associated with decreased circulating leptin (121), and we recently found that SGLT2 inhibition decreases plasma leptin in HAF rats (97). As leptin is known to stimulate the renal SNS, reduced circulating leptin may contribute to how SGLT2 regulates the renal SNS.

Mitochondrial dysfunction and SGLT2 inhibitors in PCOS

The mitochondrion is an essential organelle for eukaryotic organisms. It is known as "the powerhouse of the cell" because it conducts oxidative phosphorylation, a process necessary to generate enough energy for complex organisms to function (80). Mitochondrial dysfunction can be defined as when mitochondria cannot provide ATP for the cell while minimizing an overflow of naturally formed reactive oxygen species (ROS) from damaging the rest of the cell (122, 123). Methods to measure ROS in humans or animals include assessing total antioxidant capacity in serum or measuring markers of oxidative damage, such as lipid peroxidation, through 2-thiobarbituric acid reactive substances assay (124). Mitochondrial dysfunction has been linked to diabetes (122, 125), metabolic syndrome (125), heart failure (126), chronic kidney disease (127), and PCOS (128).

Women with PCOS have decreased mitochondrial DNA, a marker of mitochondrial content or volume, in circulating leukocytes (129, 130). Decreased inner mitochondrial membrane potential, altered mitochondrial structure, and increased ROS have also been demonstrated in the oocytes of a PCOS mouse model (131). Furthermore, lean women with PCOS have decreased circulating total antioxidant capacity and increased malondialdehyde, indicating increased oxidative stress (59). The mitochondrion is an organizing center for cellular metabolism (80), so there are significant implications for insulin resistance and obesity in PCOS. Additionally, excess oxidative stress can lead to inflammation, further worsening insulin resistance, obesity, and blood pressure in PCOS (132, 133). Therefore, targeting mitochondrial dysfunction may be a promising therapeutic avenue in patients with PCOS. Yilmaz et al. demonstrated in lean women with PCOS that rosiglitazone, a peroxisome proliferatoractivated receptor-y (PPARy) agonist that stimulates mitochondrial biogenesis (41), increases circulating total antioxidant capacity, decreases circulating malondialdehyde, and decreases insulin resistance (59). However, PPARy has multiple functions outside of stimulating mitochondrial biogenesis (134), so it is uncertain if specifically increasing mitochondrial content improves these parameters in women with PCOS. Furthermore, body mass index increases with the thiazolidinedione rosiglitazone in women with PCOS (59). However, there are concerns about thiazolidinediones and cardiovascular risk (42), limiting excitement for using PPARy agonists in women with PCOS. Exploring the potential role of other pharmacotherapeutics that improve mitochondrial function, such as SGLT2 inhibitors (105, 135-138), is a promising new direction in the field.

10.3389/fendo.2023.951099

SGLT2 inhibition has been found to improve mitochondrial function in various ways. In the heart of male rodents with T2DM, SGLT2 inhibition decreases ROS production (135) and also increases the expression of nuclear respiratory factor 1 (NRF1) and PPARy coactivator 1- α (PGC1 α) (136), which positively regulate mitochondrial biogenesis (134). SGLT2 inhibition also decreases markers of oxidative stress in the blood while increasing the activity of the antioxidant enzyme superoxide dismutase (136). Meanwhile, in the kidney of male rodents with T2DM, SGLT2 inhibition normalizes mitochondrial morphology (preventing mitochondria from becoming excessively round or fragmented) while decreasing urinary 8isoprostane and 8-hydroxydeoxyguanosine, which are markers of oxidative stress (105, 137). In white adipose tissue of male rodents with T2DM, SGLT2 inhibition similarly increased the expression of NRF1 and PGC1 α as it did in the heart, which was associated with an increase in mitochondrial DNA, a marker of mitochondrial content (136, 138). The diseases currently indicated for SGLT2 inhibitor use, such as T2DM, heart failure, and chronic kidney disease, have been linked to mitochondrial dysfunction (125-127), so improving mitochondrial function may be an essential pathway for the beneficial effects seen in patients on SGLT2 inhibitors. However, more studies are needed to explore the therapeutic potential of SGLT2 inhibition on mitochondrial dysfunction in women, especially those with PCOS. We recently demonstrated that hyperandrogenemia in the HAF rat model of PCOS causes the expansion of white adipose tissue, which is associated with decreases in mitochondrial content and function in both subcutaneous and visceral adipose tissue (139). Treatment with SGLT2 inhibitors increased the frequency of small adipocytes in visceral adipose tissue without affecting mitochondrial dysfunction in white adipose tissue, oxidative stress, or insulin resistance in the HAF rat model (139). Our study suggests that targeting mitochondrial dysfunction in PCOS may be necessary to improve insulin resistance and that hyperandrogenemia blunts the beneficial effect of SGLT2 inhibitors in the HAF rat model of PCOS.

Perspective and clinical implications

PCOS is the most common endocrine disorder in reproductiveage women (1–3). Patients with PCOS have an increased incidence of major adverse cardiovascular events (24), likely driven by the increased incidence of cardiovascular risk factors in this population, such as hypertension, insulin resistance, renal injury, and obesity (16– 19) (Figure 2). Unfortunately, there is a lack of effective, evidencebased pharmacotherapeutics targeted at cardiometabolic disease (25). Meanwhile, SGLT2 inhibitors have been rapidly expanding their clinical indications because of their cardiovascular protection in patients with and without T2DM (43, 47, 51, 52). However, whether and exactly how SGLT2 inhibitors confer cardiovascular protection in PCOS women, with and without diabetes remains to be elucidated pending high quality large clinical trials. Limited clinical data have suggested that women with PCOS have renal and cardiac target organ injury (140). Moreover, a recent study has shown that women with PCOS have a higher risk of preeclampsia/eclampsia, peripartum cardiomyopathy, and heart failure during hospitalizations for delivery (141). Women with PCOS who have increased cardiovascular risk factors as the conditions mentioned above could benefit from SGLT2 inhibitors, pending confirmation with clinical trials. The numerous mechanistic hypotheses for cardiovascular protection include deactivation of the RAS and/or the SNS as well as improvement in mitochondrial function, all of which are abnormal in women with PCOS. Data from recent small clinical trials and basic research show promise for SGLT2 inhibitors in treating some of the cardiometabolic complications in PCOS.

Author contributions

JP and LY drafted the manuscript. JP, DR, and LY reviewed and edited the manuscript. All authors contributed to the article and approved the submitted version.

Funding

This work was supported by National Institutes of Health National Institute of General Medical Sciences grant P20GM121334 (LY and DR) and National Institute of Diabetes and Digestive and Kidney Diseases grant R21DK113500 (DR). JP was supported by National Institutes of Health National Institute of Diabetes and Digestive and Kidney Diseases fellowship F30DK127527. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Acknowledgments

Figures generated with BioRender.com.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Bozdag G, Mumusoglu S, Zengin D, Karabulut E, Yildiz BO. The prevalence and phenotypic features of polycystic ovary syndrome: a systematic review and meta-analysis. *Hum Reprod* (2016) 31(12):2841–55. doi: 10.1093/humrep/dew218

2. March WA, Moore VM, Willson KJ, Phillips DI, Norman RJ, Davies MJ. The prevalence of polycystic ovary syndrome in a community sample assessed under contrasting diagnostic criteria. *Hum Reprod* (2010) 25(2):544–51. doi: 10.1093/humrep/ dep399

3. Knochenhauer ES, Key TJ, Kahsar-Miller M, Waggoner W, Boots LR, Azziz R. Prevalence of the polycystic ovary syndrome in unselected black and white women of the southeastern united states: a prospective study. *J Clin Endocrinol Metab* (1998) 83 (9):3078–82. doi: 10.1210/jc.83.9.3078

4. Hippocrates. The aphorisms of Hippocrates: With a translation into Latin and English. (1822) Coar T, editor. London: Longman p. 140.

5. Stein IF. Amenorrhea associated with bilateral polycystic ovaries. Am J Obstet Gynecol (1935) 29:181–91. doi: 10.1016/S0002-9378(15)30642-6

6. Rotterdam ESHRE/ASRM-Sponsored PCOS consensus workshop group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome (PCOS). *Hum reproduction* (2004) 19(1):41–7. doi: 10.1093/humrep/ deh098

7. Azziz R, Carmina E, Dewailly D, Diamanti-Kandarakis E, Escobar-Morreale HF, Futterweit W, et al. Criteria for defining polycystic ovary syndrome as a predominantly hyperandrogenic syndrome: an androgen excess society guideline. *J Clin Endocrinol Metab* (2006) 91(11):4237–45. doi: 10.1210/jc.2006-0178

8. Zawadski JK, Dunaif A. Diagnostic criteria for polycystic ovary syndrome: towards a rational approach. In: Dunaif A, Givens JR, Haseltine FP, Merrian GR editors. *Polycystic ovary syndrome*. Boston: Blackwell Scientific Publications (1992). p. 377–84.

9. Bode DV, Seehusen DA, Baird D. Hirsutism in women. Am Fam Physician (2012) 85(4):373-80.

10. Sheehan MT. Polycystic ovarian syndrome: diagnosis and management. *Clin Med Res* (2004) 2(1):13–27. doi: 10.3121/cmr.2.1.13

11. Travison TG, Vesper HW, Orwoll E, Wu F, Kaufman JM, Wang Y, et al. Harmonized reference ranges for circulating testosterone levels in men of four cohort studies in the united states and Europe. *J Clin Endocrinol Metab* (2017) 102(4):1161–73. doi: 10.1210/jc.2016-2935

12. Yanes Cardozo LL, Romero DG, Reckelhoff JF. Cardiometabolic features of polycystic ovary syndrome: role of androgens. *Physiology* (2017) 32(5):357-66. doi: 10.1152/physiol.00030.2016

13. Dapas M, Lin FT, Nadkarni GN, Sisk R, Legro RS, Urbanek M, et al. Distinct subtypes of polycystic ovary syndrome with novel genetic associations: An unsupervised, phenotypic clustering analysis. *PLoS Med* (2020) 17(6):e1003132. doi: 10.1371/journal.pmed.1003132

14. Day F, Karaderi T, Jones MR, Meun C, He C, Drong A, et al. Large-Scale genomewide meta-analysis of polycystic ovary syndrome suggests shared genetic architecture for different diagnosis criteria. *PLoS Genet* (2018) 14(12):e1007813. doi: 10.1371/ journal.pgen.1007813

15. Heron M. Deaths: Leading causes for 2019. Natl Vital Stat Reports: Natl Center Health Stat (2021) 70(9):1–114. doi: 10.15620/cdc:104186

16. Dumesic DA, Oberfield SE, Stener-Victorin E, Marshall JC, Laven JS, Legro RS. Scientific statement on the diagnostic criteria, epidemiology, pathophysiology, and molecular genetics of polycystic ovary syndrome. *Endocr Rev* (2015) 36(5):487-525. doi: 10.1210/er.2015-1018

17. Lo JC, Feigenbaum SL, Yang J, Pressman AR, Selby JV, Go AS. Epidemiology and adverse cardiovascular risk profile of diagnosed polycystic ovary syndrome. *J Clin Endocrinol Metab* (2006) 91(4):1357–63. doi: 10.1210/jc.2005-2430

18. Elting MW, Korsen TJ, Bezemer PD, Schoemaker J. Prevalence of diabetes mellitus, hypertension and cardiac complaints in a follow-up study of a Dutch PCOS population. *Hum Reprod* (2001) 16(3):556–60. doi: 10.1093/humrep/16.3.556

19. Song Y, Ye W, Ye H, Xie T, Shen W, Zhou L. Serum testosterone acts as a prognostic indicator in polycystic ovary syndrome-associated kidney injury. *Physiol Rep* (2019) 7(16):e14219. doi: 10.14814/phy2.14219

20. Romero DG, Yanes Cardozo LL. Cardiovascular events in polycystic ovary syndrome: Is the debate settled for good? *J Clin Endocrinol Metab* (2021) 106(12): e5258–e9. doi: 10.1210/clinem/dgab509

21. Glintborg D, Hass Rubin K, Nybo M, Abrahamsen B, Andersen M. Morbidity and medicine prescriptions in a nationwide Danish population of patients diagnosed with polycystic ovary syndrome. *Eur J Endocrinol* (2015) 172(5):627–38. doi: 10.1530/EJE-14-1108

22. Glintborg D, Rubin KH, Nybo M, Abrahamsen B, Andersen M. Cardiovascular disease in a nationwide population of Danish women with polycystic ovary syndrome. *Cardiovasc diabetology* (2018) 17(1):1–12. doi: 10.1186/s12933-018-0680-5

23. Zhang J, Xu J-H, Qu Q-Q, Zhong G-Q. Risk of cardiovascular and cerebrovascular events in polycystic ovarian syndrome women: a meta-analysis of cohort studies. *Front Cardiovasc Med* (2020) 7:552421. doi: 10.3389/fcvm.2020.552421

24. Berni TR, Morgan CL, Rees DA. Women with polycystic ovary syndrome have an increased risk of major cardiovascular events: a population study. *J Clin Endocrinol Metab* (2021) 106(9):e3369–e80. doi: 10.1210/clinem/dgab392

25. Yanes Cardozo LL, Romero DG. Management of cardiometabolic complications in polycystic ovary syndrome: Unmet needs. *FASEB J* (2021) 35(11):e21945. doi: 10.1096/fj.202002526RR

26. Shannon JA, Fisher S. The renal tubular reabsorption of glucose in the normal dog. *Am J Physiology-Legacy Content* (1938) 122(3):765-74. doi: 10.1152/ajplegacy.1938.122.3.765

27. Scriver CR, Chesney RW, McInnes RR. Genetic aspects of renal tubular transport: diversity and topology of carriers. *Kidney Int* (1976) 9(2):149–71. doi: 10.1038/ki.1976.18

28. Turner RJ, Moran A. Heterogeneity of sodium-dependent d-glucose transport sites along the proximal tubule: evidence from vesicle studies. *Am J Physiology-Renal Physiol* (1982) 242(4):F406–F14. doi: 10.1152/ajprenal.1982.242.4.F406

29. You G, Lee W-S, Barros EJ, Kanai Y, Huo T-L, Khawaja S, et al. Molecular characteristics of na+-coupled glucose transporters in adult and embryonic rat kidney. J Biol Chem (1995) 270(49):29365–71. doi: 10.1074/jbc.270.49.29365

30. Hediger M, Kanai Y, You G, Nussberger S. Mammalian ion-coupled solute transporters. J Physiol (1995) 482(suppl):7–17. doi: 10.1113/jphysiol.1995.sp020559

31. Vallon V, Platt KA, Cunard R, Schroth J, Whaley J, Thomson SC, et al. SGLT2 mediates glucose reabsorption in the early proximal tubule. *J Am Soc Nephrology* (2011) 22(1):104–12. doi: 10.1681/ASN.2010030246

32. Filippas-Ntekouan S, Filippatos TD, Elisaf MS. SGLT2 inhibitors: are they safe? Postgrad Med (2018) 130(1):72-82. doi: 10.1080/00325481.2018.1394152

33. Ehrenkranz JR, Lewis NG, Ronald Kahn C, Roth J. Phlorizin: a review. *Diabetes Metab Res Rev* (2005) 21(1):31–8. doi: 10.1002/dmrr.532

34. Rieg T, Vallon V. Development of SGLT1 and SGLT2 inhibitors. *Diabetologia* (2018) 61(10):2079-86. doi: 10.1007/s00125-018-4654-7

35. Pajor AM, Randolph KM, Kerner SA, Smith CD. Inhibitor binding in the human renal low- and high-affinity na+/glucose cotransporters. *J Pharmacol Exp Ther* (2008) 324:985–91. doi: 10.1124/jpet.107.129825

36. Turk E, Martín MG, Wright EM. Structure of the human na+/glucose cotransporter gene SGLT1. J Biol Chem (1994) 269:15204–9. doi: 10.1016/S0021-9258 (17)36592-4

37. White JR. Apple trees to sodium glucose co-transporter inhibitors: a review of SGLT2 inhibition. *Clin Diabetes* (2010) 28(1):5–10. doi: 10.2337/diaclin.28.1.5

38. Grempler R, Thomas L, Eckhardt M, Himmelsbach F, Sauer A, Sharp D, et al. Empagliflozin, a novel selective sodium glucose cotransporter-2 (SGLT-2) inhibitor: characterisation and comparison with other SGLT-2 inhibitors. *Diabetes Obes Metab* (2012) 14(1):83–90. doi: 10.1111/j.1463-1326.2011.01517.x

39. Braunwald E. SGLT2 inhibitors: the statins of the 21st century. *Eur Heart J* (2022) 43(11):1029–30. doi: 10.1093/eurheartj/ehab765

40. Smith RJ, Goldfine AB, Hiatt WR. Evaluating the cardiovascular safety of new medications for type 2 diabetes: time to reassess? *Diabetes Care* (2016) 39(5):738–42. doi: 10.2337/dc15-2237

41. Bogacka I, Xie H, Bray GA, Smith SR. Pioglitazone induces mitochondrial biogenesis in human subcutaneous adipose tissue in vivo. *Diabetes* (2005) 54(5):1392–9. doi: 10.2337/diabetes.54.5.1392

42. Rosen CJ. The rosiglitazone story-lessons from an FDA advisory committee meeting. New Engl J Med (2007) 357(9):844–6. doi: 10.1056/NEJMp078167

43. Zinman B, Wanner C, Lachin JM, Fitchett D, Bluhmki E, Hantel S, et al. Empagliflozin, cardiovascular outcomes, and mortality in type 2 diabetes. *New Engl J Med* (2015) 373(22):2117–28. doi: 10.1056/NEJMoa1504720

44. Wanner C, Inzucchi SE, Lachin JM, Fitchett D, von Eynatten M, Mattheus M, et al. Empagliflozin and progression of kidney disease in type 2 diabetes. *New Engl J Med* (2016) 375(4):323–34. doi: 10.1056/NEJMoa1515920

45. Neal B, Perkovic V, Mahaffey KW, De Zeeuw D, Fulcher G, Erondu N, et al. Canagliflozin and cardiovascular and renal events in type 2 diabetes. *New Engl J Med* (2017) 377(7):644–57. doi: 10.1056/NEJMoa1611925

46. Wiviott SD, Raz I, Bonaca MP, Mosenzon O, Kato ET, Cahn A, et al. Dapagliflozin and cardiovascular outcomes in type 2 diabetes. *New Engl J Med* (2019) 380(4):347–57. doi: 10.1056/NEJMoa1812389

47. Perkovic V, Jardine MJ, Neal B, Bompoint S, Heerspink HJ, Charytan DM, et al. Canagliflozin and renal outcomes in type 2 diabetes and nephropathy. *New Engl J Med* (2019) 380(24):2295–306. doi: 10.1056/NEJMoa1811744

48. Blau JE, Tella SH, Taylor SI, Rother KI. Ketoacidosis associated with SGLT2 inhibitor treatment: analysis of FAERS data. *Diabetes Metab Res Rev* (2017) 33(8):e2924. doi: 10.1002/dmrr.2924

49. Packer M, Anker SD, Butler J, Filippatos G, Pocock SJ, Carson P, et al. Cardiovascular and renal outcomes with empagliflozin in heart failure. *New Engl J Med* (2020) 383(15):1413–24. doi: 10.1056/NEJMoa2022190

50. Anker SD, Butler J, Filippatos G, Ferreira JP, Bocchi E, Böhm M, et al. Empagliflozin in heart failure with a preserved ejection fraction. *New Engl J Med* (2021) 385(16):1451-61. doi: 10.1056/NEJMoa2107038

51. McMurray JJ, Solomon SD, Inzucchi SE, Køber L, Kosiborod MN, Martinez FA, et al. Dapagliflozin in patients with heart failure and reduced ejection fraction. *New Engl J Med* (2019) 381(21):1995–2008. doi: 10.1056/NEJMoa1911303

52. Heerspink HJ, Stefánsson BV, Correa-Rotter R, Chertow GM, Greene T, Hou F-F, et al. Dapagliflozin in patients with chronic kidney disease. *New Engl J Med* (2020) 383 (15):1436–46. doi: 10.1056/NEJMoa2024816

53. Vallon V, Verma S. Effects of SGLT2 inhibitors on kidney and cardiovascular function. *Annu Rev Physiol* (2021) 83:503–28. doi: 10.1146/annurev-physiol-031620-095920

54. Filippatos TD, Liontos A, Papakitsou I, Elisaf MS. SGLT2 inhibitors and cardioprotection: a matter of debate and multiple hypotheses. *Postgraduate Med* (2019) 131(2):82–8. doi: 10.1080/00325481.2019.1581971

55. Javed Z, Papageorgiou M, Deshmukh H, Rigby AS, Qamar U, Abbas J, et al. Effects of empagliflozin on metabolic parameters in polycystic ovary syndrome: A randomized controlled study. *Clin Endocrinol (Oxf)* (2019) 90(6):805–13. doi: 10.1111/cen.13968

56. Elkind-Hirsch KE, Chappell N, Seidemann E, Storment J, Bellanger D. Exenatide, dapagliflozin, or Phentermine/Topiramate differentially affect metabolic profiles in polycystic ovary syndrome. *J Clin Endocrinol Metab* (2021) 106(10):3019–33. doi: 10.1210/clinem/dgab408

57. Tan S, Ignatenko S, Wagner F, Dokras A, Seufert J, Zwanziger D, et al. Licogliflozin versus placebo in women with polycystic ovary syndrome: A randomized, double-blind, phase 2 trial. *Diabetes Obes Metab* (2021) 23(11):2595–9. doi: 10.1111/dom.14495

58. Bays HE, Kozlovski P, Shao Q, Proot P, Keefe D. Licogliflozin, a novel SGLT1 and 2 inhibitor: body weight effects in a randomized trial in adults with overweight or obesity. *Obesity* (2020) 28(5):870–81. doi: 10.1002/oby.22764

59. Yilmaz M, Bukan N, Gk A, Karakoç A, Toruner F, Çakir N, et al. The effects of rosiglitazone and metformin on oxidative stress and homocysteine levels in lean patients with polycystic ovary syndrome. *Hum Reproduction* (2005) 20(12):3333–40. doi: 10.1093/ humrep/dei258

60. Micic D, Popovic V, Nesovic M, Sumarac M, Dragasevic M, Kendereski A, et al. Androgen levels during sequential insulin euglycemic clamp studies in patients with polycystic ovary disease. *J Steroid Biochem* (1988) 31(6):995–9. doi: 10.1016/0022-4731 (88)90344-5

61. Nestler JE. Insulin regulation of human ovarian androgens. *Hum Reprod* (1997) 12 (suppl_1):53-62. doi: 10.1093/humrep/12.suppl_1.53

62. Swerdloff RS, Dudley RE, Page ST, Wang C, Salameh WA. Dihydrotestosterone: biochemistry, physiology, and clinical implications of elevated blood levels. *Endocrine Rev* (2017) 38(3):220–54. doi: 10.1210/er.2016-1067

63. Diamond MP, Grainger D, Diamond MC, Sherwin RS, DeFronzo RA. Effects of methyltestosterone on insulin secretion and sensitivity in women. *J Clin Endocrinol Metab* (1998) 83(12):4420–5. doi: 10.1210/jc.83.12.4420

64. Manneras L, Cajander S, Holmaüng A, Seleskovic Z, Lystig T, Loünn M, et al. A new rat model exhibiting both ovarian and metabolic characteristics of polycystic ovary syndrome. *Endocrinology* (2007) 148(8):3781–91. doi: 10.1210/en.2007-0168

65. Yanes LL, Romero DG, Moulana M, Lima R, Davis DD, Zhang H, et al. Cardiovascular-renal and metabolic characterization of a rat model of polycystic ovary syndrome. *Gender Med* (2011) 8(2):103–15. doi: 10.1016/j.genm.2010.11.013

66. McCartney CR, Marshall JCCLINICAL PRACTICE. Polycystic ovary syndrome. N Engl J Med (2016) 375(1):54–64. doi: 10.1056/NEJMcp1514916

67. Association AD. 7. approaches to glycemic treatment. *Diabetes Care* (2015) 38 (Supplement 1):S41-S8. doi: 10.2337/dc15-S010

68. Guan Y, Wang D, Bu H, Zhao T, Wang H. The effect of metformin on polycystic ovary syndrome in overweight women: a systematic review and meta-analysis of randomized controlled trials. *Int J Endocrinol* (2020) 2020:5150684. doi: 10.1155/2020/5150684

69. Diamanti-Kandarakis E, Kouli C, Tsianateli T, Bergiele A. Therapeutic effects of metformin on insulin resistance and hyperandrogenism in polycystic ovary syndrome. *Eur J Endocrinology* (1998) 138(3):269–74. doi: 10.1530/eje.0.1380269

70. Jensterle M, Kocjan T, Kravos NA, Pfeifer M, Janez A. Short-term intervention with liraglutide improved eating behavior in obese women with polycystic ovary syndrome. *Endocrine Res* (2015) 40(3):133–8. doi: 10.3109/07435800.2014.966385

71. Han Y, Li Y, He B. GLP-1 receptor agonists versus metformin in PCOS: a systematic review and meta-analysis. *Reprod Biomed Online* (2019) 39(2):332-42. doi: 10.1016/j.rbmo.2019.04.017

72. Dalmasso C, Maranon R, Patil C, Bui E, Moulana M, Zhang H, et al. Cardiometabolic effects of chronic hyperandrogenemia in a new model of postmenopausal polycystic ovary syndrome. *Endocrinology* (2016) 157(7):2920-7. doi: 10.1210/en.2015-1617

73. Stener-Victorin E, Padmanabhan V, Walters KA, Campbell RE, Benrick A, Giacobini P, et al. Animal models to understand the etiology and pathophysiology of polycystic ovary syndrome. *Endocrine Rev* (2020) 41(4):bnaa010. doi: 10.1210/endrev/bnaa010

74. Stener-Victorin E. Update on animal models of polycystic ovary syndrome. Endocrinology (2022) 163(12):bqac164. doi: 10.1210/endocr/bqac164

75. Holte J, Gennarelli G, Berne C, Bergh T, Lithell H. Elevated ambulatory day-time blood pressure in women with polycystic ovary syndrome: A sign of a pre-hypertensive state? *Hum Reprod* (1996) 11(1):23–8. doi: 10.1093/oxfordjournals.humrep.a019028

76. Mellembakken JR, Mahmoudan A, Mørkrid L, Sundström-Poromaa I, Morin-Papunen L, Tapanainen JS, et al. Higher blood pressure in normal weight women with PCOS compared to controls. *Endoc Connect* (2021) 10(2):154–63. doi: 10.1530/EC-20-0527

77. Diamanti-Kandarakis E, Economou FN, Livadas S, Tantalaki E, Piperi C, Papavassiliou AG, et al. Hyperreninemia characterizing women with polycystic ovary

syndrome improves after metformin therapy. *Kidney Blood Press Res* (2009) 32(1):24–31. doi: 10.1159/000201791

78. Jensterle M, Janez A, Vrtovec B, Meden-Vrtovec H, Pfeifer M, Prezelj J, et al. Decreased androgen levels and improved menstrual pattern after angiotensin II receptor antagonist telmisartan treatment in four hypertensive patients with polycystic ovary syndrome: Case series. *Croat Med J* (2007) 48(6):864–70. doi: 10.3325/cmj.2007.6.864

79. Hall JE. Control of blood pressure by the renin-angiotensin-aldosterone system. *Clin Cardiol* (1991) 14(S4):6–21. doi: 10.1002/clc.4960141802

80. Hall J. Guyton and Hall textbook of medical physiology. 13th ed. Philadelphia, Pennsylvania: Elsevier (2016).

81. Chappell MC. Non-classical renin-angiotensin system and renal function. Compr Physiol (2012) 2(4):2733. doi: 10.1002/cphy.c120002

82. Muñoz MC, Giani JF, Burghi V, Mayer MA, Carranza A, Taira CA, et al. The Mas receptor mediates modulation of insulin signaling by angiotensin-(1–7). *Regul peptides* (2012) 177(1-3):1–11. doi: 10.1016/j.regpep.2012.04.001

83. Nehme A, Zouein FA, Deris Zayeri Z, Zibara K. An update on the tissue renin angiotensin system and its role in physiology and pathology. *J Cardiovasc Dev Dis* (2019) 6(2):14. doi: 10.3390/jcdd6020014

84. Chen J, Williams S, Ho S, Loraine H, Hagan D, Whaley JM, et al. Quantitative PCR tissue expression profiling of the human SGLT2 gene and related family members. *Diabetes Ther* (2010) 1(2):57–92. doi: 10.1007/s13300-010-0006-4

85. Miyata N, Park F, Li XF, Cowley AWJr. Distribution of angiotensin AT1 and AT2 receptor subtypes in the rat kidney. *Am J Physiology-Renal Physiol* (1999) 277(3):F437–46. doi: 10.1152/ajprenal.1999.277.3.F437

86. Navar LG, Kobori H, Prieto MC, Gonzalez-Villalobos RA. Intratubular reninangiotensin system in hypertension. *Hypertension* (2011) 57(3):355–62. doi: 10.1161/ HYPERTENSIONAHA.110.163519

87. Chen M, Harris M, Rose D, Smart A, He X, Kretzler M, et al. Renin and renin mRNA in proximal tubules of the rat kidney. *J Clin Invest* (1994) 94(1):237-43. doi: 10.1172/JCI117312

88. Moe OW, Ujiie K, Star RA, Miller RT, Widell J, Alpern RJ, et al. Renin expression in renal proximal tubule. J Clin Invest (1993) 91(3):774–9. doi: 10.1172/JCI116296

89. Prieto-Carrasquero MC, Harrison-Bernard LM, Kobori H, Ozawa Y, Hering-Smith KS, Hamm LL, et al. Enhancement of collecting duct renin in angiotensin IIdependent hypertensive rats. *Hypertension* (2004) 44(2):223–9. doi: 10.1161/ 01.HYP.0000135678.20725.54

90. Mezzano S, Droguett A, Burgos ME, Ardiles LG, Flores CA, Aros CA, et al. Reninangiotensin system activation and interstitial inflammation in human diabetic nephropathy. *Kidney Int* (2003) 64:S64–70. doi: 10.1046/j.1523-1755.64.s86.12.x

91. Komlosi P, Fuson AL, Fintha A, Peti-Peterdi J, Rosivall L, Warnock DG, et al. Angiotensin I conversion to angiotensin II stimulates cortical collecting duct sodium transport. *Hypertension* (2003) 42(2):195–9. doi: 10.1161/01.HYP.0000081221.36703.01

92. Sun P, Yue P, Wang W-H. Angiotensin II stimulates epithelial sodium channels in the cortical collecting duct of the rat kidney. *Am J Physiology-Renal Physiol* (2012) 302(6): F679–F87. doi: 10.1152/ajprenal.00368.2011

93. Baker EH. Ion channels and the control of blood pressure. Br J Clin Pharmacol (2000) 49(3):185–98. doi: 10.1046/j.1365-2125.2000.00159.x

94. Moin ASM, Sathyapalan T, Atkin SL, Butler AE. Renin-angiotensin system overactivation in polycystic ovary syndrome, a risk for SARS-CoV-2 infection? *Metab Open* (2020) 7:100052. doi: 10.1016/j.metop.2020.100052

95. Alreja G, Joseph J. Renin and cardiovascular disease: Worn-out path, or new direction. World J Cardiol (2011) 3(3):72–83. doi: 10.4330/wjc.v3.i3.72

96. Torres Fernandez ED, Huffman AM, Syed M, Romero DG, Yanes Cardozo LL. Effect of GLP-1 receptor agonists in the cardiometabolic complications in a rat model of postmenopausal PCOS. *Endocrinology* (2019) 160(12):2787–99. doi: 10.1210/en.2019-00450

97. Pruett JE, Torres Fernandez ED, Everman SJ, Vinson RM, Davenport K, Logan MK, et al. Impact of SGLT-2 inhibition on cardiometabolic abnormalities in a rat model of polycystic ovary syndrome. *Int J Mol Sci* (2021) 22(5):2576. doi: 10.3390/ijms22052576

98. Cooper WO, Hernandez-Diaz S, Arbogast PG, Dudley JA, Dyer S, Gideon PS, et al. Major congenital malformations after first-trimester exposure to ACE inhibitors. *New Engl J Med* (2006) 354(23):2443–51. doi: 10.1056/NEJMoa055202

99. Saji H, Yamanaka M, Hagiwara A, Ijiri R. Losartan and fetal toxic effects. *Lancet* (2001) 357(9253):363. doi: 10.1016/S0140-6736(00)03648-5

100. Martinovic J, Benachi A, Laurent N, Daïkha-Dahmane F, Gubler MC. Fetal toxic effects and angiotensin-II-receptor antagonists. *Lancet* (2001) 358(9277):241–2. doi: 10.1016/S0140-6736(01)05426-5

101. Sawamura T, Karashima S, Nagase S, Nambo H, Shimizu E, Higashitani T, et al. Effect of sodium–glucose cotransporter-2 inhibitors on aldosterone-to-renin ratio in diabetic patients with hypertension: A retrospective observational study. *BMC Endocrine Disord* (2020) 20(1):1–8. doi: 10.1186/s12902-020-00656-8

102. Kidokoro K, Cherney DZ, Bozovic A, Nagasu H, Satoh M, Kanda E, et al. Evaluation of glomerular hemodynamic function by empagliflozin in diabetic mice using *in vivo* imaging. *Circulation* (2019) 140(4):303–15. doi: 10.1161/CIRCULATIONAHA.118.037418

103. Schork A, Saynisch J, Vosseler A, Jaghutriz BA, Heyne N, Peter A, et al. Effect of SGLT2 inhibitors on body composition, fluid status and renin–angiotensin–aldosterone system in type 2 diabetes: A prospective study using bioimpedance spectroscopy. *Cardiovasc Diabetol* (2019) 18(1):1–12. doi: 10.1186/s12933-019-0852-y

104. Masuda T, Muto S, Fukuda K, Watanabe M, Ohara K, Koepsell H, et al. Osmotic diuresis by SGLT2 inhibition stimulates vasopressin-induced water reabsorption to maintain body fluid volume. *Physiol Rep* (2020) 8(2):e14360. doi: 10.14814/phy2.14360

105. Woods TC, Satou R, Miyata K, Katsurada A, Dugas CM, Klingenberg NC, et al. Canagliflozin prevents intrarenal angiotensinogen augmentation and mitigates kidney injury and hypertension in mouse model of type 2 diabetes mellitus. *Am J Nephrol* (2019) 49(4):331–42. doi: 10.1159/000499597

106. Shin SJ, Chung S, Kim SJ, Lee EM, Yoo YH, Kim JW, et al. Effect of sodiumglucose Co-transporter 2 inhibitor, dapagliflozin, on renal renin-angiotensin system in an animal model of type 2 diabetes. *PLoS One* (2016) 11(11):e0165703. doi: 10.1371/ journal.pone.0165703

107. Kojima N, Williams JM, Slaughter TN, Kato S, Takahashi T, Miyata N, et al. Renoprotective effects of combined SGLT2 and ACE inhibitor therapy in diabetic Dahl s rats. *Physiol Rep* (2015) 3(7):e12436. doi: 10.14814/phy2.12436

108. Bautista R, Manning R, Martinez F, Avila-Casado Mdel C, Soto V, Medina A, et al. Angiotensin II-dependent increased expression of na+-glucose cotransporter in hypertension. *Am J Physiol Renal Physiol* (2004) 286(1):F127-33. doi: 10.1152/ajprenal.00113.2003

109. Yildirir A, Aybar F, Kabakci G, Yarali H, Oto A. Heart rate variability in young women with polycystic ovary syndrome. *Ann Noninvasive Electrocardiology* (2006) 11 (4):306–12. doi: 10.1111/j.1542-474X.2006.00122.x

110. Maranon R, Lima R, Spradley FT, do Carmo JM, Zhang H, Smith AD, et al. Roles for the sympathetic nervous system, renal nerves, and CNS melanocortin-4 receptor in the elevated blood pressure in hyperandrogenemic female rats. *Am J Physiology-Regulatory Integr Comp Physiol* (2015) 308(8):R708–R13. doi: 10.1152/ajpregu.00411.2014

111. Schlaich MP, Straznicky N, Grima M, Ika-Sari C, Dawood T, Mahfoud F, et al. Renal denervation: A potential new treatment modality for polycystic ovary syndrome? *J Hypertens* (2011) 29(5):991–6. doi: 10.1097/HJH.0b013e328344db3a

112. Yildizhan R, Ilhan GA, Yildizhan B, Kolusari A, Adali E, Bugdayci G. Serum retinol-binding protein 4, leptin, and plasma asymmetric dimethylarginine levels in obese and nonobese young women with polycystic ovary syndrome. *Fertil Steril* (2011) 96 (1):246–50. doi: 10.1016/j.fertnstert.2011.04.073

113. Hall JE, do Carmo JM, da Silva AA, Wang Z, Hall ME. Obesity, kidney dysfunction and hypertension: Mechanistic links. *Nat Rev Nephrology* (2019) 15 (6):367–85. doi: 10.1038/s41581-019-0145-4

114. da Silva AA, do Carmo JM, Wang Z, Hall JE. Melanocortin-4 receptors and sympathetic nervous system activation in hypertension. *Curr Hypertension Rep* (2019) 21 (6):1–7. doi: 10.1007/s11906-019-0951-x

115. Maddox TM, Januzzi JLJr., Allen LA, Breathett K, Butler J, Davis LL, et al. 2021 Update to the 2017 ACC expert consensus decision pathway for optimization of heart failure treatment: answers to 10 pivotal issues about heart failure with reduced ejection fraction: A report of the American college of cardiology solution set oversight committee. J Am Coll Cardiol (2021) 77(6):772–810. doi: 10.1016/j.jacc.2020.11.022

116. MRC Working Party. Medical research council trial of treatment of hypertension in older adults: Principal results. *BMJ* (1992) 304(6824):405-12. doi: 10.1136/ bmj.304.6824.405

117. Messerli FH. Implications of discontinuation of doxazosin arm of ALLHAT. Lancet (2000) 355(9207):863-4. doi: 10.1016/S0140-6736(00)00092-1

118. Nojima T, Matsubayashi Y, Yoshida A, Suganami H, Abe T, Ishizawa M, et al. Influence of an SGLT2 inhibitor, tofogliflozin, on the resting heart rate in relation to adipose tissue insulin resistance. *Diabetic Med* (2020) 37(8):1316–25. doi: 10.1111/dme.14279

119. Sano M, Chen S, Imazeki H, Ochiai H, Seino Y. Changes in heart rate in patients with type 2 diabetes mellitus after treatment with luseogliflozin: Subanalysis of placebocontrolled, double-blind clinical trials. *J Diabetes Invest* (2018) 9(3):638–41. doi: 10.1111/ jdi.12726

120. Herat LY, Magno AL, Rudnicka C, Hricova J, Carnagarin R, Ward NC, et al. SGLT2 inhibitor-induced sympathoinhibition: a novel mechanism for cardiorenal protection. *JACC: Basic to Trans Science* (2020) 5(2):169–79. doi: 10.1016/j.jacbts.2019.11.007

121. Wu P, Wen W, Li J, Xu J, Zhao M, Chen H, et al. Systematic review and metaanalysis of randomized controlled trials on the effect of SGLT2 inhibitor on blood leptin and adiponectin level in patients with type 2 diabetes. *Hormone Metab Res* (2019) 51 (08):487–94. doi: 10.1055/a-0958-2441 122. Pieczenik SR, Neustadt J. Mitochondrial dysfunction and molecular pathways of disease. *Exp Mol Pathol* (2007) 83(1):84–92. doi: 10.1016/j.yexmp.2006.09.008

123. Nicolson GL. Mitochondrial dysfunction and chronic disease: Treatment with natural supplements. Integr Med (Encinitas) (2014) 13(4):35.

124. Katerji M, Filippova M, Duerksen-Hughes P. Approaches and methods to measure oxidative stress in clinical samples: Research applications in the cancer field. *Oxid Med Cell Longevity* (2019) 2019:1279250. doi: 10.1155/2019/1279250

125. Bhatti JS, Bhatti GK, Reddy PH. Mitochondrial dysfunction and oxidative stress in metabolic disorders–a step towards mitochondria based therapeutic strategies. *Biochim Biophys Acta Basis Dis* (2017) 1863(5):1066–77. doi: 10.1016/j.bbadis.2016.11.010

126. Zhou B, Tian R. Mitochondrial dysfunction in pathophysiology of heart failure. J Clin Invest (2018) 128(9):3716–26. doi: 10.1172/JCI120849

127. Zhang X, Agborbesong E, Li X. The role of mitochondria in acute kidney injury and chronic kidney disease and its therapeutic potential. *Int J Mol Sci* (2021) 22 (20):11253. doi: 10.3390/ijms222011253

128. Zhang J, Bao Y, Zhou X, Zheng L. Polycystic ovary syndrome and mitochondrial dysfunction. *Reprod Biol Endocrinol* (2019) 17(1):1–15. doi: 10.1186/s12958-019-0509-4

129. Lee S-H, Chung D-J, Lee H-S, Kim T-J, Kim M-H, Jeong HJ, et al. Mitochondrial DNA copy number in peripheral blood in polycystic ovary syndrome. *Metabolism* (2011) 60(12):1677–82. doi: 10.1016/j.metabol.2011.04.010

130. Reddy TV, Govatati S, Deenadayal M, Sisinthy S, Bhanoori M. Impact of mitochondrial DNA copy number and displacement loop alterations on polycystic ovary syndrome risk in south Indian women. *Mitochondrion* (2019) 44:35–40. doi: 10.1016/j.mito.2017.12.010

131. Chappell NR, Zhou B, Schutt AK, Gibbons WE, Blesson CS. Prenatal androgen induced lean PCOS impairs mitochondria and mRNA profiles in oocytes. *Endocrine connections* (2020) 9(3):261–70. doi: 10.1530/EC-19-0553

132. Okuno Y, Fukuhara A, Hashimoto E, Kobayashi H, Kobayashi S, Otsuki M, et al. Oxidative stress inhibits healthy adipose expansion through suppression of SREBF1mediated lipogenic pathway. *Diabetes* (2018) 67(6):1113–27. doi: 10.2337/db17-1032

133. Mouton AJ, Li X, Hall ME, Hall JE. Obesity, hypertension, and cardiac dysfunction: Novel roles of immunometabolism in macrophage activation and inflammation. *Circ Res* (2020) 126(6):789-806. doi: 10.1161/CIRCRESAHA.119.312321

134. Corona JC, Duchen MR. PPARγ as a therapeutic target to rescue mitochondrial function in neurological disease. *Free Radical Biol Med* (2016) 100:153–63. doi: 10.1016/ j.freeradbiomed.2016.06.023

135. Croteau D, Luptak I, Chambers JM, Hobai I, Panagia M, Pimentel DR, et al. Effects of sodium-glucose linked transporter 2 inhibition with ertugliflozin on mitochondrial function, energetics, and metabolic gene expression in the presence and absence of diabetes mellitus in mice. *J Am Heart Assoc* (2021) 10(13):e019995. doi: 10.1161/JAHA.120.019995

136. Shao Q, Meng L, Lee S, Tse G, Gong M, Zhang Z, et al. Empagliflozin, a sodium glucose co-transporter-2 inhibitor, alleviates atrial remodeling and improves mitochondrial function in high-fat diet/streptozotocin-induced diabetic rats. *Cardiovasc Diabetol* (2019) 18(1):1–14. doi: 10.1186/s12933-019-0964-4

137. Takagi S, Li J, Takagaki Y, Kitada M, Nitta K, Takasu T, et al. Ipragliflozin improves mitochondrial abnormalities in renal tubules induced by a high-fat diet. *J Diabetes Invest* (2018) 9(5):1025–32. doi: 10.1111/jdi.12802

138. Wei D, Liao L, Wang H, Zhang W, Wang T, Xu Z. Canagliflozin ameliorates obesity by improving mitochondrial function and fatty acid oxidation *via* PPARα *in vivo* and in vitro. *Life Sci* (2020) 247:117414. doi: 10.1016/j.lfs.2020.117414

139. Pruett JE, Everman SJ, Hoang NH, Salau F, Taylor LC, Edwards KS, et al. Mitochondrial function and oxidative stress in white adipose tissue in a rat model of PCOS: Effect of SGLT2 inhibition. *Biol Sex Differ* (2022) 13(1):45. doi: 10.1186/s13293-022-00455-x

140. Reckelhoff JF, Shawky NM, Romero DG, Yanes Cardozo LL. Polycystic ovary syndrome: Insights from preclinical research. *Kidney360* (2022) 3(8):1449–57. doi: 10.34067/KID.0002052022

141. Zahid S, Khan MZ, Gowda S, Faza NN, Honigberg MC, Vaught A, et al. Trends, predictors, and outcomes of cardiovascular complications associated with polycystic ovary syndrome during delivery hospitalizations: A national inpatient sample analysis (2002–2019). J Am Heart Assoc (2022) 11(16):e025839. doi: 10.1161/JAHA.121.025839