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RECEIVED 08 September 2024 ACCEPTED 12 December 2024 PUBLISHED 07 January 2025

CITATION

Yao G, Dou X, Chen X, Qi H, Chen J, Wu P, Li J, Liang S, Han Z, Bai S and Hu X (2025) Association between sperm DNA fragmentation index and recurrent pregnancy loss: results from 1485 participants undergoing fertility evaluation. *Front. Endocrinol.* 15:1493186. doi: 10.3389/fendo.2024.1493186

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Association between sperm DNA fragmentation index and recurrent pregnancy loss: results from 1485 participants undergoing fertility evaluation

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Objective: Several male factors have been reported to play a role in recurrent pregnancy loss (RPL). The aim of this study is to explore the relationship between semen parameters, sperm DNA fragmentation index (DFI) and RPL.

Method: A total of 1485 participants were recruited from a university hospital between April 2020 and August 2022. Six hundred and thirtyfour men from couples with RPL were assigned to the case group, while 851 men from couple without RPL who underwent fertile evaluation were assigned to the control group. Semen parameters including sperm DNA fragmentation, were assessed.

Results: No statistically significant differences in semen parameters, sperm kinematics and DFI were observed between the case group and the control group. A higher proportion of men in the case group had a DFI > 30% compared to those in the control group; however, this difference was not statistically significant. Restricted cubic spline analysis revealed no significant non-linear relationships between continuous DFI and risk of RPL.

Conclusion: Our study indicates that there is no significant relationship between DFI and RPL risk. Further prospective studies are needed to explore the impact of DFI on fertility outcomes in couples experiencing RPL.

KEYWORDS

DNA fragmentation index, recurrent pregnancy loss, semen quality, DFI, RPL

Introduction

Approximately 15% of reproductive-age couples suffer from infertility globally, with male factors accounting for half of these cases (1, 2). Conventional semen analysis is a standard method for evaluating semen quality, including parameters such as volume, sperm concentration, motility and morphology. However, this method is limited to evaluating basic parameters and additional sperm functions, such as DNA integrity, acrosome reaction and capacitation, require further assessment. Sperm DNA integrity is a critical parameter for male reproduction. Due to the complex organization of haploid genome, the DNA within the sperm nucleus is often altered by protamine-mediated compaction (3). Previous studies have demonstrated that sperm DNA fragmentation can adversely affect fertilization and embryo development (4-6). DNA Fragmentation Index (DFI) is a parameter used to assess the integrity of sperm DNA. Specifically, DFI measures the percentage of spermatozoa exhibiting DNA fragmentation, which can be an indicator of sperm quality and fertility potential. High DFI values are associated with increased sperm DNA damage, which may result from factors like oxidative stress, apoptosis, or autophagy (7).

Recurrent pregnancy loss (RPL) is defined as two or more clinically recognized pregnancy losses before 20-24 weeks of gestation, including both embryo and fetal loss (8). According to previous epidemiological studies, the prevalence of RPL is 1-4% among all patients who achieve pregnancy (9). Several factors have been suggested to contribute to pathogenesis of RPL, including maternal age, genetic defects, uterine pathology, endocrine disorders, and infectious agents. However, the exact etiology of RPL in approximately 50-75% of patients remains unclear (10). Although most known causes of RPL are related to female factors, recent studies have also indicated that semen quality may influence the risk of PRL. A systematic review and meta-analysis including 16 studies and 2969 couples suggested that increased DFI was associated with an increased risk of RPL (11). Additionally, several studies reported male partners of couples with RPL were more likely to have high DFI compared with those with low DFI (12, 13). Importantly, new guidelines from the European Society of Human Reproduction and Embryology (ESHRE), the American Urological Association (AUA), and the American Society for Reproductive Medicine (ASRM) recommend sperm DFI as a parameter for evaluating RPL (14-16).

Assays for analyzing sperm DNA integrity have been developed in recent years, including the terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) assay, the comet assay, the sperm chromatin dispersion (SCD) assay, and the sperm chromatin structure assay (SCSA). In a recent review, the authors provided a comprehensive summary of the techniques, advantages, and disadvantages of the TUNEL, SCSA, SCD, and Comet assays (17). The terminal deoxynucleotidyl transferase-mediated deoxyuridine triphosphate nick end labeling (TUNEL) assay is noted for its sensitivity, reliability, and minimal interrater variability. However, its disadvantages include high cost, the need for specialized equipment, and extensive training requirements. The sperm chromatin dispersion (SCD) assay is advantageous due to its simplicity and minimal equipment requirements, though it is hindered by a high degree of interobserver variability. The comet assay offers sensitivity and reproducibility, but suffers from interobserver variability, the necessity for an experienced operator, and the use of variable protocols. Finally, the sperm chromatin structure assay (SCSA) is characterized by a standardized protocol, reproducibility, and the ability to examine a large number of cells, though it also faces limitations in terms of high cost, the need for specialized equipment, and specific training requirements. Among these tests, SCSA is utilized for assessing the DFI in several large cohorts, although there is an ongoing effort to establish a consensus approach that is both highly reliable and costeffective. Date from SCSA indicate that DFI has a strong positive correlation with pregnancy outcomes and miscarriage (18). In this study, which includes a relatively large sample size, we measured sperm DFI using SCSA to investigate the association between DFI and RPL risk.

Methods

Participants

Between April 2020 and August 2022, a total of 1485 male partners of infertile couples at the Reproductive Center of The First Affiliated Hospital of University of Science and Technology of China (USTC) and the female partners aged between 22 and 40 years, were included in this retrospective study. Couples who conceived their previous natural pregnancy within 1 year and experienced RPL were assigned to the case group, while couples without RPL and at least one previous pregnancy or live birth were assigned to the control group. All control group underwent fertility treatments for female factor infertility such as tubal factors. Semen parameters, including sperm DNA fragmentation were assessed, and clinical characteristics, such as demographic factors, lifestyle factors, and medical history, were collected for all participants. Couples diagnosed with causes of RPL, such as abnormal karyotypes and mullerian ducts, diabetes, hyperprolactinemia, thyroid disorders and positive for the antiphospholipid antibodies, antinuclear antibodies, lupus antibodies and anti ß2 glycoprotein antibodies, were excluded from the study. The exclusion criteria in male partners of infertile couples included azoospermia, testicular cancer, cryptorchidism and genetic defects related to the male reproductive tract. This study was approved by the Ethical Committee of The First Affiliated Hospital of USTC (No. 2023-RE-196).

Routing semen analysis

All semen sample collected at couples with previous pregnancy or live birth less than one year. Semen parameters (e.g., semen volume, sperm concentration, motility, and morphology) were assessed according to the WHO criteria (37). Ejaculates were collected by masturbation, followed by liquefaction at 37°C for at least 30 minutes. Semen parameters, including sperm concentration, progressive motility, and total motility, were analyzed using a computer-assisted sperm analysis (CASA) system (SAS-II, SAS Medical, Beijing, China). Sperm morphology was assessed following Diff-Quick staining (Anke Biotechnology, Hefei, China) using a light microscope (CX33, Olympus Corporation, Tokyo, Japan). Leukocytes were evaluated following peroxidase staining (Anke Biotechnology, Hefei, China). Antisperm antibodies (AsAs) were assessed using the mixed antiglobulin reaction (MAR) test (Anke Biotechnology, Hefei, China).

Detection of sperm DFI

The sperm DFI was assessed using the sperm chromatin structure assay (SCSA, Celula, Chengdu, China). Semen samples were diluted with TNE buffer (Tris-HCl, NaCl, and ethylenediaminetetraacetic acid (EDTA)), followed by treatment with an acidic detergent solution (TritonX-100, NaCl, and HCl, pH 1.2). After 30 seconds, staining buffer (acridine orange, citric acid, Na2HPO4, EDTA, NaCl, pH 6.0) was added. The samples were analyzed using a flow cytometer (Celula, Chengdu, China) and a minimum of 5000 cells were examined per sample. Chromatin damage was assessed using acridine orange staining, where green fluorescence indicated double-stranded DNA and red fluorescence indicated single-stranded and denatured DNA (19).

Statistical analysis

A systematic review and meta-analysis reported that there is a mean difference of DFI of 11% between RPL and fertile control group (20). Additionally, a recent study from China showed DFI higher than 30% in unexplained RPL group was 42% (13). By setting power at 90%, alpha error at 5%, the estimated sample size was 796 with 398 in each group. Qualitative variables were described as frequencies (percentages), and quantitative variables were described as the mean ± standard deviation (SD) if normally distributed and medians (interquartile range, IQR) if not. Pearson's chi-square test and Student's t test were used for parametric comparisons, and the Mann-Whitney U test was utilized for nonparametric comparisons. Logistic regression and multivariate regression model were used to examine the association between DFI and RPL. Covariates initially included age, BMI, education, smoking, alcohol consumption, chronic diseases, urogenital infections and varicocele. Restricted cubic spline (RCS) was performed to assess for the dose-response relationships between RPL and DFI. A P value < 0.05 was considered to indicate statistical significance. All statistical analyses were performed using Prism 9.0.

Results

Clinical characteristics of 634 men from infertile couples affected by RPL and 851 men from couples who underwent fertile evaluation are reported in Table 1. Age and BMI did not show any statistically significant differences between the case and control groups. Lifestyle factors, including smoking and alcohol consumption, were similar between the two groups. No significant differences were observed in educational level, sex frequency, history of chronic diseases and urogenital infections. The duration of infertility (P < 0.001) and the prevalence of varicocele (P = 0.004) were significantly higher in the case group compared to the control group.

Semen parameters were further compared between two groups. No statistically significant differences in semen volume, sperm concentration, progressive motility, total motility, TMSC, morphology, AsAs, DFI, HDS and leukocytes were found between men in the case group and those in the control group (Table 2). In addition, the proportion of men with abnormal semen parameters did not differ significantly between the two groups. A higher proportion of men with DFI > 30% was observed in the case group compared to the control group (6.3% vs. 5.2%), but this difference did not reach statistical significance. Similarly, the study groups were comparable in terms of sperm kinematics, including VCL, VSL, VAP, BCF, ALH, MAD, LIN, WOB, and STR (Table 3).

The association between abnormal semen parameters and the risk of RPL was further investigated using logistic regression analysis (Table 4). No significant associations were observed in either the crude or adjusted models. We further performed multivariate regression models for evaluating the association between sperm DFI and RPL. However, no correlation between DFI and RPL were found (adjusted OR 1.01, 95% CI (1.00-1.03), P = 0.09, Table 5). RCS was performed to assess the dose-response relationships between RPL and continues DFI (Figure 1). The DFI was included as a natural cubic spline function, adjusting for age, BMI, education, smoking, drinking, sex frequency, chronic disease, duration of infertility, urogenital infections, varicocele, semen volume, sperm concentration, total motility, progressive motility, TMSC and normal morphology. No significant non-linear relationships were observed between continuous DFI and the risk of RPL.

Discussion

Recent studies have suggested a positive correlation between sperm DFI and abnormal pregnancy history. In this study, a total of 1485 men from couples with RPL and those without RPL was recruited, and semen quality, including sperm DFI was evaluated. However, our results indicated that no significant relationship was observed between sperm DFI and the risk of RPL.

Several clinical characteristics, including lifestyle and medical history, have been associated with RPL. A recent study by Esquerre-Lamare et al. recruited 60 participants (33 men from couples with RPL and 27 men from couples with a history of live birth less than the one year) and assessed environmental and family factors as well as semen parameters (21). The results showed that the RPL group were more likely to have a higher male BMI and a family history of infertility. They also found no significant differences in andrological histories and clinical examinations (e.g., cryptorchidism, varicoceles, vas deferens, epididymal position) between cases and controls. However, our study showed that the mean BMI of men did not differ between the study groups, which is consistent with several TABLE 1 Clinical characteristics in men from the case group and the control group.

Characteristics	Total (n=1485)	Control (n=851)	Case (n=634)	P value
Age, mean ± s.d.	32.2 ± 5.6	32.4 ± 5.8	32.0 ± 5.4	0.29
BMI ^a , mean ± s.d.	24.7 ± 3.4	24.7 ± 3.3	24.7 ± 3.5	0.78
Smoking status, n (%)				0.13
Nonsmoker ^b	865 (58.2)	510 (59.9)	355 (56.0)	
Smoker	620 (41.8)	341 (40.1)	279 (44.0)	
Drinking status, n (%)				0.60
Nondrinker ^c	677 (45.6)	393 (46.2)	284 (44.8)	
Drinker	808 (54.4)	458 (53.8)	350 (55.2)	
Education, n (%)				0.25
Middle school or lower ^d	299 (20.1)	162 (19.1)	137 (21.6)	
High school	253 (17.1)	139 (16.3)	114 (18.0)	
College/University	933 (62.8)	550 (64.6)	383 (60.4)	
Sex frequency (weekly), n (%)				0.55
<1	174 (11.7)	93 (10.9)	81 (12.8)	
1-2	973 (65.5)	563 (66.2)	410 (64.7)	
>2	338 (22.8)	195 (22.9)	143 (22.5)	
Chronic diseases, n (%) ^e				0.27
No	1309 (88.1)	757 (89.0)	552 (87.1)	
Yes	176 (11.9)	94 (11.0)	82 (12.9)	
Duration of infertility (year), n (%)				<0.001
<1	742 (50.0)	471 (55.4)	271 (42.7)	
1-2	375 (25.2)	207 (24.3)	168 (26.5)	
>2	368 (24.8)	173 (20.3)	195 (30.8)	
Urogenital infections, n (%) ^f				0.74
No	1277 (86.0)	734 (86.3)	543 (85.7)	
Yes	208 (14.0)	117 (13.7)	91 (14.3)	
Varicocele, n (%)				0.004
No	1372 (92.4)	801 (94.1)	571 (90.1)	
Yes	113 (7.6)	50 (5.9)	63 (9.9)	

BMI, body mass index; s.d, standard deviation.

^aCalculated as weight in kilograms divided by height in meters squared.

^bincluded never smoking and no smoking during the past 3 months.

^cincluded never drinking and no drinking during the past 3 months.

^dincluded primary school and junior high school.

eincluded diabetes, hypertension, and hyperlipidemia.

fincluded epididymitis, prostatitis, balanoposthitis, and seminal vesiculitis.

previously published results (13, 22). In addition, our study found that men from couples with RPL were more likely to have varicocele compared to men from couples without RPL. This finding is consistent with Busnelli et al., who reported a higher prevalence of varicocele in men with RPL compared to the men without RPL

(14.9% vs. 8.0%), although this difference did not reach statistical significance (23).

Traditional semen analysis provides a quantitative assessment of sperm concentration, motility and morphology (24). Several studies have shown a trend of low sperm motility and high

Semen parameters	Total (n=1485)	Control (n=851)	Case (n=634)	<i>P</i> value
Volume (ml), median (IQR)	3.2 (2.3-4.2)	3.2 (2.2-4.3)	3.1 (2.3-4.1)	0.87
< 1.5 ml, n (%)	107 (7.2)	69 (8.1)	38 (6.0)	0.12
Concentration (million/ml), median (IQR)	68.2 (38.0-120.0)	67.7 (37.3-123.2)	69.5 (39.6-118.3)	0.73
< 15 million/ml, n (%)	96 (6.5)	53 (6.2)	43 (6.8)	0.67
Progressive motility (%), median (IQR)	40.4 (28.6-52.3)	39.4 (27.3-52.0)	42.1 (29.8-52.7)	0.09
< 32%, n (%)	471 (31.7)	285 (33.4)	186 (29.3)	0.09
Total motility (%), median (IQR)	45.6 (32.6-58.6)	44.5 (31.5-58.5)	47.7 (33.6-58.8)	0.12
< 40%, n (%)	577 (38.9)	346 (40.7)	231 (36.4)	0.10
TMSC (million), median (IQR)	94.9 (39.1-204.0)	92.0 (36.1-202.4)	97.0 (44.5-205.2)	0.33
< 9 million, n (%)	91 (6.1)	53 (6.2)	38 (6.0)	0.85
Normal morphology (%), median (IQR; n)	7.0 (5.0-8.0; 1437)	7.0 (5.0-8.0; 826)	7.0 (5.0-8.0; 611)	0.38
< 4%, n (%)	106 (7.4)	61 (7.4)	45 (7.4)	0.99
Leukocytes(×10 ⁶ /ml), median (IQR; n)	0.08 (0.03-0.29; 873)	0.04 (0.01-0.08; 511)	0.06 (0.03-0.28; 362)	0.17
AsAs (%), median (IQR; n)	1.0 (0.0-3.0; 575)	1.0 (0.0-3.0; 334)	2.0 (0.0-4.0; 241)	0.20
DFI (%), median (IQR)	11.9 (7.6-17.9)	11.9 (7.6-17.8)	11.9 (7.7-18.3)	0.41
>30%, n (%)	84 (5.7)	44 (5.2)	40 (6.3)	0.35
HDS (%), median (IQR)	6.6 (4.9-8.8)	6.6 (4.8-8.8)	6.6 (5.0-8.9)	0.34

TABLE 2 Semen parameters in men from the case group and the control group.

IQR, interquartile range; TMSC, total motile sperm count; AsAs, antisperm antibodies; DFI, DNA fragmentation index; HDS, high DNA stainability.

abnormal sperm morphology in men from couples with RPL (12, 25, 26). However, similar to our findings, many studies that examined the correlation between routine semen parameters and RPL found no significant correlation (27, 28). Since normal sperm genetic material is essential for successful fertilization, as well as embryo and fetal development, sperm DNA integrity is considered a key parameter for evaluating clinical pregnancy outcomes. Several

meta-analyses have reported that sperm DFI is associated with RPL (29, 30). However, these meta-analyses may be underpowered due to the limited number of studies and sample sizes included (20). Results from our study are consistent with Zhang et al. that reported no significant difference in DFI between RPL group and control group (27). Additionally, the researchers followed up with the participants for one year and observed a trend toward higher DFI

TABLE 3 Sperm kinematics in men from the case group and the control group.

Sperm kinematics	Total (n=1485)	Control (n=851)	Case (n=634)	P value
VCL (µm/sec), median (IQR)	32.5 (23.3-43.9)	32.0 (22.7-43.5)	33.3 (24.0-44.7)	0.18
VSL (µm/sec), median (IQR)	13.7 (9.1-18.5)	13.2 (8.8-18.5)	14.2 (9.6-18.6)	0.19
VAP (µm/sec), median (IQR)	17.5 (12.0-23.5)	16.9 (11.7-23.4)	18.1 (12.6-23.7)	0.18
BCF (Hz), median (IQR)	8.3 (5.7-11.6)	8.0 (5.4-11.6)	8.7 (5.9-11.6)	0.13
ALH (µm/sec), median (IQR)	2.3 (2.0-2.4)	2.2 (2.0-2.4)	2.3 (2.0-2.5)	0.25
MAD (degrees), median (IQR)	13.9 (11.1-16.6)	13.7 (10.9-16.6)	13.9 (11.3-16.7)	0.30
LIN, median (IQR)	38.7 (32.8-43.9)	38.4 (32.5-43.8)	39.1 (33.2-44.2)	0.26
WOB, median (IQR)	50.8 (45.3-55.1)	50.6 (45.1-55.0)	51.2 (45.5-55.4)	0.25
STR, median (IQR)	74.8 (64.4-79.4)	74.4 (63.9-79.2)	75.3 (65.1-79.6)	0.14

VCL, curvilinear velocity; VSL, straight-line velocity; VAP, average path velocity; BCF, frequency of beat cross; ALH, mean amplitude of lateral head displacement; MAD, mean angular displacement; LIN, linearity coefficient; WOB, wobble coefficient; STR, straightness coefficient.

Semen parameters	Case		
	OR (95% CI)	Р	
Crude			
Volume < 1.5 ml	0.72 (0.48-1.08)	0.12	
Concentration < 15 million/ml	1.10 (0.72-1.66)	0.67	
Progressive motility <32%	0.82 (0.66-1.03)	0.09	
Total motility <40%	0.84 (0.68-1.03)	0.10	
TMSC < 9 million	0.96 (0.62-1.47)	0.85	
Normal morphology < 4%	1.00 (0.67-1.48)	0.99	
DFI > 30%	1.24 (0.79-1.92)	0.35	
Adjusted			
Volume < 1.5 ml	0.75 (0.49-1.13)	0.17	
Concentration < 15 million/ml	1.00 (0.96-1.00)	0.99	
Progressive motility <32%	0.80 (0.64-1.01)	0.06	
Total motility <40%	0.80 (0.64-1.00)	0.05	
TMSC < 9 million	0.89 (0.57-1.38)	0.60	
Normal morphology < 4%	0.91 (0.60-1.38)	0.67	
DFI > 30%	1.24 (0.78-1.95)	0.36	

TABLE 4 Odds ratios (95% confidence intervals) for abnormal semen parameters in men from couples with RPL.

RPL, recurrent pregnancy loss; TMSC, total motile sperm count; DFI, DNA fragmentation index; OR, odds ratio; CI, confidence interval.

Adjusted model: adjusted for age, BMI, education, smoking, alcohol consumption, chronic diseases, urogenital infections, and varicocele.

in men from couples with subsequent abnormal pregnancy outcomes compared to those from couples with ongoing pregnancies. This suggests that DFI may be a potential semen parameter for predicting future reproductive outcomes. TABLE 5 Multivariate regression models for the association between sperm DFI and RPL.

DFI	RPL	
	<i>OR</i> (95% CI)	Р
Crude	1.01(1.00-1.02)	0.23
Adjusted	1.01 (1.00-1.03)	0.09

DFI, DNA fragmentation index; RPL, recurrent pregnancy loss; OR, odds ratio; CI, confidence interval. Adjusted model: adjusted for age, BMI, education, smoking, alcohol consumption, chronic diseases, urogenital infections, varicocele, semen volume, sperm concentration, total motility, progressive motility, TMSC and normal morphology.

A cut-off DFI > 30% was reported as a severe DNA fragmentation in several studies (13, 31, 32). Consistent with other studies, our study also showed a trend of a higher proportion of sperm $DFI \ge 30\%$ in men from the RPL group compared to those from the control group (22, 33). Interestingly, the RCS model indicated that when DFI exceeded 30%, the risk of RPL appeared to increase. Damaged DNA in the sperm may cause embryos to fail in their early stages, contributing to pregnancy loss (34). In addition, sperm DNA contributes to the development of placenta. Fragmented DNA can impair trophoblast development, leading to inadequate placental function, which is a known cause of pregnancy loss (35). Furthermore, DNA fragmentation may disrupt epigenetic marks in sperm, leading to errors in gene expression regulation during embryogenesis. This can result in developmental abnormalities and increase the risk of miscarriage (36). These findings suggest that a threshold DFI, rather than continuous DFI, may be more useful for assessing the correlation between DFI and RPL.

This study has several strengths. First, the relatively large sample size provides substantial statistical power. Additionally, our study included potential confounding factors (e.g. lifestyles and medical history) that have been previously reported to be associated with RPL. However, this study has several limitations. First, the study is limited by its



FIGURE 1

Dose-response curves between DFI and risk of RPL. The DFI was included as a natural cubic spline function, adjusting for age, BMI, education, smoking, drinking, sex frequency, chronic disease, duration of infertility, urogenital infections, varicocele, semen volume, sperm concentration, total motility, progressive motility, TMSC and normal morphology.

retrospective design and potential selection bias. Second, all participants were recruited from couples undergoing fertility evaluation at a single clinic. Third, although several potential confounders were adjusted for, residual confounding and unknown factors could not be ruled out. It is note that the relationship between sperm DNA fragmentation and recurrent pregnancy loss cannot be fully understood without considering female factors. Maternal age, uterine environment, oocyte quality, and immune response all play significant roles in moderating the impact of DFI on pregnancy outcomes. Forth, the SCSA assay cannot measure DNA strand breaks directly. Finally, different cut-off values for DFI may produce biased results.

In conclusion, our study found no significant relationship between sperm DFI and the risk of RPL. Further prospective studies are needed to investigate the impact of DFI on fertility outcomes in couples experiencing RPL.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

This study was approved by the Ethical Committee of The First Affiliated Hospital of USTC (No. 2023-RE-196). The studies were conducted in accordance with the local legislation and institutional requirements. The ethics committee/institutional review board waived the requirement of written informed consent for participation from the participants or the participants' legal guardians/next of kin because this study was a retrospective data analysis, the written informed consent was waived.

References

1. Mclachlan RI, De Kretser DM. Male infertility: the case for continued research. Med J Aust. (2001) 174:116-7. doi: 10.5694/j.1326-5377.2001.tb143180.x

2. Tremellen K. Oxidative stress and male infertility-a clinical perspective. *Hum Reprod Update*. (2008) 14:243–58. doi: 10.1093/humupd/dmn004

3. Yazdanpanah Ghadikolaei P, Ghaleno LR, Vesali S, Janzamin E, Gilani M, Sajadi H, et al. Epidemiology of sperm DNA fragmentation in a retrospective cohort of 1191 men. *Andrology*. (2023) 11:1663–72. doi: 10.1111/andr.13472

 Panner Selvam MK, Baskaran S, O'connell S, Almajed W, Hellstrom WJG, Sikka SC. Association between seminal oxidation-reduction potential and sperm DNA fragmentation-A meta-analysis. *Antioxidants (Basel)*. (2022) 11:1563. doi: 10.3390/antiox11081563

5. Hussain T, Kandeel M, Metwally E, Murtaza G, Kalhoro DH, Yin Y, et al. Unraveling the harmful effect of oxidative stress on male fertility: A mechanistic insight. *Front Endocrinol (Lausanne).* (2023) 14:1070692. doi: 10.3389/fendo.2023.1070692

6. Liu K, Mao X, Pan F, Chen Y, An R. Correlation analysis of sperm DNA fragmentation index with semen parameters and the effect of sperm DFI on outcomes of ART. *Sci Rep.* (2023) 13:2717. doi: 10.1038/s41598-023-28765-z

7. Will MA, Swain J, Fode M, Sonksen J, Christman GM, Ohl D. The great debate: varicocele treatment and impact on fertility. *Fertil Steril.* (2011) 95:841–52. doi: 10.1016/j.fertnstert.2011.01.002

8. Lee JY, Moon J, Hu HJ, Ryu CS, Ko EJ, Ahn EH, et al. Discovery of pathogenic variants associated with idiopathic recurrent pregnancy loss using whole-exome sequencing. *Int J Mol Sci.* (2024) 25:5447. doi: 10.3390/ijms25105447

Author contributions

GY: Writing – original draft. XD: Writing – review & editing. XC: Writing – review & editing. HQ: Writing – review & editing. JC: Writing – review & editing. PW: Writing – review & editing. JL: Writing – review & editing. SL: Writing – review & editing. ZH: Writing – review & editing. SB: Writing – original draft. XH: Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This study was supported by the National Natural Science Foundation of China (81901543).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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 Magnus MC, Wilcox AJ, Morken NH, Weinberg CR, Haberg SE. Role of maternal age and pregnancy history in risk of miscarriage: prospective register based study. *BMJ*. (2019) 364:1869. doi: 10.1136/bmj.1869

10. Turesheva A, Aimagambetova G, Ukybassova T, Marat A, Kanabekova P, Kaldygulova L, et al. Recurrent pregnancy loss etiology, risk factors, diagnosis, and management. Fresh look into a full box. J Clin Med. (2023) 12:4074. doi: 10.3390/jcm12124074

11. Robinson L, Gallos ID, Conner SJ, Rajkhowa M, Miller D, Lewis S, et al. The effect of sperm DNA fragmentation on miscarriage rates: a systematic review and metaanalysis. *Hum Reprod.* (2012) 27:2908–17. doi: 10.1093/humrep/des261

12. Bareh GM, Jacoby E, Binkley P, Chang TC, Schenken RS, Robinson RD. Sperm deoxyribonucleic acid fragmentation assessment in normozoospermic male partners of couples with unexplained recurrent pregnancy loss: a prospective study. *Fertil Steril.* (2016) 105329–336. doi: 10.1016/j.fertnstert.2015.10.033

13. Zhu XB, Chen Q, Fan WM, Niu ZH, Xu BF, Zhang AJ. Sperm DNA fragmentation in Chinese couples with unexplained recurrent pregnancy loss. *Asian J Androl.* (2020) 22:296–301. doi: 10.4103/aja.aja_60_19

14. Schlegel PN, Sigman M, Collura B, De Jonge CJ, Eisenberg ML, Lamb DJ, et al. Diagnosis and treatment of infertility in men: AUA/ASRM guideline part I. *Fertil Steril.* (2021) 115:54–61. doi: 10.1016/j.fertnstert.2020.11.015

15. Schlegel PN, Sigman M, Collura B, De Jonge CJ, Eisenberg ML, Lamb DJ, et al. Diagnosis and treatment of infertility in men: AUA/ASRM guideline part II. Fertil Steril. (2021) 115:62–9. doi: 10.1016/j.fertnstert.2020.11.016

16. Rpl EGGO, Bender Atik R, Christiansen OB, Elson J, Kolte AM, Lewis S, et al. ESHRE guideline: recurrent pregnancy loss: an update in 2022. *Hum Reprod Open.* (2023) 2023:hoad002. doi: 10.1093/hropen/hoad002

17. Farkouh A, Saleh R, Shah R, Agarwal A. Sperm DNA fragmentation in male infertility: From bench to bedside. *Arab J Urol.* (2023) 21:199–203. doi: 10.1080/20905998.2023.2278200

18. Evenson DP, Jost LK, Marshall D, Zinaman MJ, Clegg E, Purvis K, et al. Utility of the sperm chromatin structure assay as a diagnostic and prognostic tool in the human fertility clinic. *Hum Reprod.* (1999) 14:1039–49. doi: 10.1093/humrep/14.4.1039

19. Oleszczuk K, Giwercman A, Bungum M. Intra-individual variation of the sperm chromatin structure assay DNA fragmentation index in men from infertile couples. *Hum Reprod.* (2011) 26:3244–8. doi: 10.1093/humrep/der328

20. Mcqueen DB, Zhang J, Robins JC. Sperm DNA fragmentation and recurrent pregnancy loss: a systematic review and meta-analysis. *Fertil Steril.* (2019) 112:54–60e53. doi: 10.1016/j.fertnstert.2019.03.003

21. Esquerre-Lamare C, Walschaerts M, Chansel Debordeaux L, Moreau J, Bretelle F, Isus F, et al. Sperm aneuploidy and DNA fragmentation in unexplained recurrent pregnancy loss: a multicenter case-control study. *Basic Clin Androl.* (2018) 28:4. doi: 10.1186/s12610-018-0070-6

22. Solanki M, Joseph T, Muthukumar K, Samuel P, Aleyamma TK, Kamath MS. Impact of sperm DNA fragmentation in couples with unexplained recurrent pregnancy loss: A cross-sectional study. *J Obstet Gynaecol Res.* (2024) 50:1687-96. doi: 10.1111/jog.16041

23. Busnelli A, Garolla A, Di Credico E, D'ippolito S, Merola AM, Milardi D, et al. Sperm DNA fragmentation and idiopathic recurrent pregnancy loss: Results from a multicenter case-control study. *Andrology*. (2023) 11:1673–81. doi: 10.1111/andr.13395

24. Bai S, Dou XC, Qi HL, Zhu YS, Zhang YT, Liu YX, et al. Association between semen collection time and semen parameters: an observational study. *Asian J Androl.* (2023) 25:339–44. doi: 10.4103/aja202268

25. Brahem S, Mehdi M, Landolsi H, Mougou S, Elghezal H, Saad A. Semen parameters and sperm DNA fragmentation as causes of recurrent pregnancy loss. *Urology*. (2011) 78:792–6. doi: 10.1016/j.urology.2011.05.049

26. Kumar K, Deka D, Singh A, Mitra DK, Vanitha BR, Dada R. Predictive value of DNA integrity analysis in idiopathic recurrent pregnancy loss following

spontaneous conception. J Assist Reprod Genet. (2012) 29:861-7. doi: 10.1007/s10815-012-9801-3

27. Zhang L, Wang L, Zhang X, Xu G, Zhang W, Wang K, et al. Sperm chromatin integrity may predict future fertility for unexplained recurrent spontaneous abortion patients. *Int J Androl.* (2012) 35:752–7. doi: 10.1111/j.1365-2605.2012.01276.x

28. Coughlan C, Clarke H, Cutting R, Saxton J, Waite S, Ledger W, et al. Sperm DNA fragmentation, recurrent implantation failure and recurrent miscarriage. *Asian J Androl.* (2015) 17:681–5. doi: 10.4103/1008-682X.144946

29. Dai Y, Liu J, Yuan E, Li Y, Shi Y, Zhang L. Relationship among traditional semen parameters, sperm DNA fragmentation, and unexplained recurrent miscarriage: A systematic review and meta-analysis. *Front Endocrinol (Lausanne)*. (2021) 12:802632. doi: 10.3389/fendo.2021.802632

30. Inversetti A, Bossi A, Cristodoro M, Larcher A, Busnelli A, Grande G, et al. Recurrent pregnancy loss: a male crucial factor-A systematic review and meta-analysis. *Andrology.* (2025) 13:130-45. doi: 10.1111/andr.13540

31. Evenson DP, Larson KL, Jost LK. Sperm chromatin structure assay: its clinical use for detecting sperm DNA fragmentation in male infertility and comparisons with other techniques. J Androl. (2002) 23:25–43. doi: 10.1002/j.1939-4640.2002.tb02599.x

32. Venkatesh S, Singh A, Shamsi MB, Thilagavathi J, Kumar R, Mitra DK, et al. Clinical significance of sperm DNA damage threshold value in the assessment of male infertility. *Reprod Sci.* (2011) 18:1005–13. doi: 10.1177/1933719111401662

33. Buck Louis GM, Sundaram R, Schisterman EF, Sweeney A, Lynch CD, Kim S, et al. Semen quality and time to pregnancy: the Longitudinal Investigation of Fertility and the Environment Study. *Fertil Steril.* (2014) 101:453–62. doi: 10.1016/j.fertnstert.2013.10.022

34. Sedova AO, Shtaut MI, Bragina EE, Sorokina TM, Shmarina GV, Andreeva MV, et al. Comprehensive semen examination in patients with pancreatic-sufficient and pancreatic-insufficient cystic fibrosis. *Asian J Androl.* (2023) 25:591–7. doi: 10.4103/aja2022115

35. Zini A, Boman JM, Belzile E, Ciampi A. Sperm DNA damage is associated with an increased risk of pregnancy loss after IVF and ICSI: systematic review and metaanalysis. *Hum Reprod.* (2008) 23:2663–8. doi: 10.1093/humrep/den321

36. Aston KI, Carrell DT. Prospects for clinically relevant epigenetic tests in the andrology laboratory. Asian J Androl. (2014) 16:782. doi: 10.4103/1008-682X.125415

37. WHO (2010). WHO Laboratory Manual for the Examination and Processing of Human Semen, 5th Edn. Geneva, Switzerland: WHO Press.