Check for updates

OPEN ACCESS

EDITED BY Ronan Padraic Murphy, Dublin City University, Ireland

REVIEWED BY Sulev Kõks, Murdoch University, Australia Maria-Christina Ungureanu, Grigore T. Popa University of Medicine and Pharmacy, Romania

*CORRESPONDENCE

Zhongze Fang fangzhongze@tmu.edu.cn Rongxiu Zheng rzheng@tmu.edu.cn Jing Li lijing_ph@tmu.edu.cn

[†]These authors share first authorship

RECEIVED 18 February 2025 ACCEPTED 21 May 2025 PUBLISHED 05 June 2025

CITATION

Zhao Z, Ma Y, Zhang X, Liu X, Li Y, Fang Z, Zheng R and Li J (2025) Association of IGF-1 and IGFBP-3 with metabolic abnormalities among children and adolescents. *Front. Endocrinol.* 16:1579107. doi: 10.3389/fendo.2025.1579107

COPYRIGHT

© 2025 Zhao, Ma, Zhang, Liu, Li, Fang, Zheng and Li. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Association of IGF-1 and IGFBP-3 with metabolic abnormalities among children and adolescents

Zhenghao Zhao^{1†}, Yuanyuan Ma^{1†}, Xinyi Zhang¹, Xiaoxiao Liu², Yang Li¹, Zhongze Fang^{1,3,4*}, Rongxiu Zheng^{2*} and Jing Li^{3,4,5*}

¹Department of Toxicology and Health Inspection and Quarantine, School of Public Health, Tianjin Medical University, Tianjin, China, ²Department of Pediatrics, Tianjin Medical University, General Hospital, Tianjin, China, ³Tianjin Key Laboratory of Environment, Nutrition and Public Health, Tianjin, China, ⁴Tianjin Center for International Collaborative Research on Environment, Nutrition and Public Health, Tianjin, China, ⁵Department of Epidemiology and Biostatistics. School of Public Health, Tianjin Medical University, Tianjin, China

Background and Objective: Insulin-like growth factor-1 (IGF-1) and insulin-like growth factor binding protein-3 (IGFBP-3) play roles in growth and development, but their association with metabolic abnormalities in children and adolescents remains unclear. This study aimed to investigate the relationship between IGF-1, IGFBP-3, and metabolic abnormalities in Chinese children and adolescents, while assessing the role of age in these associations.

Methods: Participants were categorized into low-risk and high-risk groups based on metabolic abnormality criteria. Demographic, anthropometric, and laboratory data were collected via medical records. Logistic regression was used to calculate odds ratios (ORs) and 95% confidence intervals (CIs).

Results: Data from 588 participants were analyzed. Higher IGF-1 (Q4: OR 0.24, 95% CI: 0.11–0.51) and IGFBP-3 levels (Q4: OR 0.38, 95% CI: 0.18–0.76) were associated with lower odds of metabolic abnormalities. Higher IGF-1/IGFBP-3 ratios also reduced metabolic abnormality risk. Age-related trends showed IGF-1 levels plateaued with age, while IGFBP-3 progressively increased, with the low-risk group consistently maintaining higher levels.

Conclusions: Higher IGF-1 and IGFBP-3 levels are negatively associated with metabolic abnormalities in children and adolescents. Maintaining the balance of these factors is critical for metabolic health, especially during adolescence.

KEYWORDS

insulin-like growth factor-1, insulin-like growth factor binding protein-3, adolescent, children, metabolic abnormalities

Introduction

The increasing prevalence of obesity is leading to more cases of metabolic abnormalities in youth, posing a major public health concern (1–3). In 2022, among children and adolescents aged 5-19, more than 390 million were overweight, including 160 million with obesity (4). While obesity was once considered a concern primarily for high-income countries, its prevalence is now rising rapidly in low- and middle-income nations (5). Moreover, common metabolic risk indicators such as blood pressure (6), hypertriglyceridemia (7), and low-density lipoprotein cholesterol disorders (8) are increasingly showing abnormal trends among children and adolescents. Therefore, identifying modifiable risk factors in children and adolescents is crucial for improving their metabolic health.

Insulin-like Growth Factor-1(IGF-1) is primarily secreted by the liver under the stimulation of growth hormone (GH) (9, 10) while Insulin-like Growth Factor Binding Protein-3(IGFBP-3) serves as the main binding protein for IGF-1, regulating its biological activity (11). The development of a stable complex between IGFBP-3 and IGF-1 regulates IGF-1 availability and halflife (12, 13) *in vivo*, regulating receptor activation and subsequent signaling pathways (14).

Abnormalities in the IGF-1 axis may be associated with the development of obesity, which in turn may affect metabolic health (15). Research has demonstrated a strong association of IGF-1 and its binding proteins (including GHR and IGFBP-3) in adipose tissue cells with metabolic dysfunction among obese children (16). Furthermore, there are substantial association of IGF-1 and its binding proteins (IGFBP-1, IGFBP-2, and IGFBP-3) with cardiovascular disease risk in children aged 7 to 9 (17). Low IGF-1 is linked to insulin resistance and related metabolic disorders (18). Moreover, lower IGF-1 is associated with the risk of cardiovascular disease, such as hypertension and vascular dysfunction (19). Evidence indicates that the WFS1 gene plays a pivotal role in modulating the insulin-like growth factor-1 (IGF-1) and growth hormone (GH) signaling pathway, with genetic perturbations predisposing to impaired somatic growth and metabolic disturbances, including diabetes mellitus (20, 21). Intriguingly, WFS1 was first characterized in conditioned fear paradigms, linking it to emotional and behavioral regulation (22) and suggesting a mechanistic bridge between metabolic control and affective processes via WFS1mediated regulation of IGF-1/GH activity. IGFBP-3 is associated with all five elements of the metabolic syndrome among the older population (23). Other studies have shown that children who are overweight or obese have higher serum IGFBP-3 levels and that IGFBP-3 concentrations are linked to several cardiovascular risk factors, including obesity and insulin levels (16) (24),. Most of these studies have focused on older populations or metabolic diseases, and no studies have explored the association of IGF-1 and IGBP-3 with metabolic abnormalities assessed by clinical parameters among children and adolescents.

In this cross-sectional study, we aimed to 1) assess the association of IGF-1 and IGFBP-3 and their ratios with metabolic

abnormality in adolescents; and 2) explore the trends of IGF-1 and IGFBP-3 with age.

Methods

Exclusion criteria

The exclusion criteria for our study were: 1) children who had previously suffered from kidney disease, liver failure, cancer, or other severe systemic diseases; 2) children with hypothalamic diseases, pituitary disorders, thyroid dysfunctions, diabetes, chromosomal abnormalities, or various syndromes; 3) children who had a history of smoking, long-term alcohol consumption, or use of medications that could affect lipid metabolism, blood pressure, liver function, insulin action, blood glucose levels, or body weight.

Study populations

This cross-sectional study was conducted between January 2022 and March 2024 at the Department of Pediatrics, Tianjin Medical University General Hospital, Tianjin, China. From January 2022 to March 2023, a total of 607 children were consecutively admitted to the Department of Pediatrics at Tianjin Medical University General Hospital in Tianjin, China, and agreed to participate in this study. Among them, 19 participants were excluded due to missing IGF-1 and IGFBP-3 data. Finally, A total of 588 participants aged 6–17 years were ultimately included in the study (Figure 1).

Measurement of IGF-1 and IGFBP-3

Serum levels of Insulin-like Growth Factor-1 (IGF-1) and Insulin-like Growth Factor Binding Protein-3 (IGFBP-3) were measured using standardized enzyme-linked immunosorbent assay (ELISA) kits. Blood samples were collected from participants after an overnight fast and immediately processed. The serum was separated by centrifugation and stored at -80°C until analysis. The assays were performed in accordance with the manufacturer's instructions, and each sample was measured in duplicate to ensure accuracy. The concentrations of IGF-1 and IGFBP-3 were reported in nanograms per milliliter (ng/mL). Quality control procedures were followed to minimize variability, and intra- and inter-assay coefficients of variation were maintained within acceptable ranges.

Assessment of metabolic risk

The diagnosis of metabolic abnormalities is based on the criteria for metabolic syndrome in Chinese children and adolescents proposed by the Pediatric Branch of the Chinese Medical Association in 2012 (25), which includes: 1. Central obesity. 2.



Low high-density lipoprotein cholesterol (HDL-C) or high nonhigh-density lipoprotein cholesterol (non-HDL-C).3. Hypertriglyceridemia. 4. Hypertension. 5. Impaired fasting glucose. Central obesity is diagnosed using waist circumference percentile cutoffs for Chinese children and adolescents aged 7 to 18 years. A low HDL-C is defined as HDL-C < 1.03 mmol/L, or high non-HDL-C as non-HDL-C \geq 3.76 mmol/L. Hypertriglyceridemia is defined as triglycerides (TG) \geq 1.47 mmol/L. Hypertension is defined as having a systolic blood pressure (SBP) and/or diastolic blood pressure (DBP) \geq the 95th percentile for age and sex. Impaired fasting glucose is defined as fasting blood glucose \geq 5.6 mmol/L. Low-risk (\leq 1) and high-risk (\geq 2) groups were categorized based on the number of metabolic risks.

Data collection

This study's covariates included anthropometric measurements and data collected through questionnaires. Anthropometric

measurements for each participant included weight, height, systolic blood pressure (SBP), diastolic blood pressure (DBP), waist circumference, and hip circumference. All measurements were conducted with participants wearing light clothing and without shoes. Weight was recorded to the nearest 0.1 kilogram, and height was measured to the nearest 0.1 centimeter in the morning. Obesity was assessed using the Body Mass Index (BMI), which was calculated as weight in kilograms divided by height in meters squared (kg/m²). Blood pressure was measured three times after participants had rested in a seated position for 5 minutes, and the average value was used. The definitions of overweight and obesity are based on the "Classification Criteria for Overweight and Obesity Screening in School-age Children and Adolescents" (WS/T 586-2018), which applies to children and adolescents aged 6 to 18 years. Overweight is defined as a BMI between the 85th and 95th percentiles for the same age and gender, while obesity is defined as a BMI at or above the 95th percentile (26).

Waist circumference was measured with participants standing, feet shoulder-width apart. A non-elastic measuring tape was placed around the waist at the narrowest part of the torso, or midway between the iliac crest and the lower rib margin. The tape was kept flat against the skin (or clothing) but not so tight as to compress the skin. Measurements were taken at the end of a natural exhalation and recorded to the nearest 0.1 centimeter. Hip circumference was similarly measured while standing, with feet shoulder-width apart. The tape measure was positioned around the fullest part of the hips, generally just above the upper thighs. The tape was ensured to be level around the body, and measurements were taken at the end of a natural exhalation, and recorded to the nearest 0.1 centimeter. All measurements were performed by trained personnel under standardized conditions to ensure reliability. Multiple measurements were taken, and the average was calculated. The waist-to-hip ratio was determined by dividing waist circumference by hip circumference, expressed as Waist-Hip Ratio (WHR)= waist circumference (cm)/hip circumference (cm).

Data collection also included questionnaires completed by parents at registration. Parents provided information on the family's socioeconomic status and personal details for both parents, including lifestyle factors such as smoking and drinking. The questionnaire also included questions on the child's lifestyle habits, sleep patterns, dietary habits, and physical activity. Physical activity was defined as engaging in exercise at least three times a week for more than 30 minutes per session; participants not meeting this criterion were considered not to engage in regular physical activity. Biochemical blood tests were conducted in a specialized diagnostic laboratory using a fully automated biochemical analyzer (Hitachi 7150, Tokyo, Japan). Fasting venous blood samples were collected from each participant between 8:00 AM and 9:30 AM after an 8-hour fasting period. The biochemical examination information of the participants included alanine Aminotransferase (ALT), aspartate aminotransferase (AST), triglycerides (TG), high-density lipoprotein cholesterol (HDL-C), lowdensity lipoprotein cholesterol (LDL-C), total bilirubin (TBIL), direct bilirubin (DBIL), fasting glucose, fasting insulin, and other related parameters.

Statistical analysis

The chi-square test for categorical variables and t-test for continuous variables were used to compare the characteristics of the study participants based on metabolic risk.

The odds ratio (OR) and 95% confidence interval (CI) for the associations of IGF-1, IGFBP-3, and IGF-1/IGFBP-3 (continuous and quartile) with metabolic risk were estimated using logistic regression models. In the multivariable models, we adjusted for age, sex, BMI, WHR, ALT, AST, TBIL, DBIL, HOMA, TG, HDL-C, and LDL. The relationships of IGF-1, IGFBP-3, and their ratio with metabolic risk were visualized using the local estimation scatterplot smoothing (LOESS) technique.

In sensitivity analysis, we further evaluated the association between IGF-1, IGFBP-3, and their ratio with metabolic risk by dividing them into tertiles. Additionally, we further adjusted the household income and parental education level. Two-tailed *P*-values <0.05 were considered statistically significant. All statistical analyses were performed using R Version 4.3.2.

Results

The characteristics of the study population

Of the 588 participants included in the analysis (mean [SD]: 11.01 years; 51.19% of female). Compared to the low-risk participants, the high-risk participants had higher age (11.4 \pm 2.3 years vs. 10.8 \pm 2.2 years), height (153.7 \pm 15.9 cm vs. 145.9 \pm 15.2 cm), weight (61.1 \pm 22.8 kg vs. 42.3 \pm 16.4 kg), BMI (25.0 \pm 6.27 kg/ m² vs. 19.2 \pm 4.59 kg/m²), additionally, WHR (0.86 [0.82, 0.89] vs. 0.85 [0.81, 0.88], p = 0.002) and TG (1.51 mmol/L [1.04, 1.91] vs. 0.91 mmol/L [0.67, 1.20], p < 0.001) were also higher in the high-risk group, and HOMA-IR (3.962 [2.418, 6.359] vs. 2.129 [1.321, 3.472], p < 0.001), were also significantly elevated in the high-risk group. Furthermore, ALT (19 U/L (13, 32) vs. 15 U/L (12, 20), p < 0.001) was higher in the high-risk group, while TBIL (8.8 µmol/L [6.9, 12.2] vs. 9.9 µmol/L [7.7, 13.4], p = 0.032) and DBIL (2.7 µmol/L [2.1, 3.6] vs. 3.0 µmol/L [2.2, 4.2], p = 0.004) were lower. (Table 1).

Association of IGF and IGFBP-3 with metabolic abnormalities

In the multivariable model, compared to the lowest quartile of IGF-1 and IGFBP-3, the highest quartile of IGF-1 (OR: 0.24, 95% CI: 0.11 to 0.51) and IGFBP-3 (OR: 0.38, 95% CI: 0.18 to 0.76) were negatively associated with metabolic abnormalities (Table 2). In the multivariable model, compared with the lowest quartile, Q3 (OR: 0.43; 95% CI: 0.21 to 0.86) and Q4 (OR: 0.47, 95% CI: 0.22 to 0.96) levels of IGF-1/IGFBP-3 were negatively associated with metabolic abnormalities (Table 3).Age- and sex-stratified subgroup analyses similarly revealed the same trends(Supplementary Table S1, Supplementary Table S2).

Trends in IGF-1 and IGFBP-3 with age

In both the low-risk and high-risk groups, IGF-1 levels initially increase with age and then plateau. In the low-risk group, IGF-1 levels rise steadily until approximately 12 to 14 years of age, after which they stabilize, while IGF-1 levels in the high-risk group consistently remain lower than those in the low-risk group and also stabilize around 14 years of age. During mid-puberty (12 to 14 years), changes in IGF-1 levels in both groups are relatively stable, whereas greater variability is observed in the early and late stages of puberty (Figure 2a). IGFBP-3 levels progressively increase with age in both the low-risk and high-risk groups. Across all age ranges, IGFBP-3 levels are consistently higher in the low-risk group

Characteristics	Low metabolic risk (n = 382)	High metabolic risk (n = 206)	<i>P</i> -Value
Age, y	10.8 (2.2)	11.4 (2.3)	0.002
Female	198 (51.83)	103 (50.00)	0.736
BMI (kg/m ²)	19.2 (4.59)	25.0 (6.27)	<0.001
Normal	319 (83.51)	127 (61.65)	< 0.001
Overweight	23 (6.02)	15 (7.28)	
Obesity	40 (10.47)	64 (31.07)	
SBP (mmHg)	106 (99, 112)	120 (109, 128)	<0.001
DBP (mmHg)	67 (63,72)	75 (69, 81)	<0.001
WHR	0.85 (0.81, 0.88)	0.86 (0.82, 0.89)	0.002
TG (mmol/l)	0.91 (0.67, 1.20)	1.51 (1.04, 1.91)	<0.001
HDL-C (mmol/l)	1.30 (1.14, 1.55)	1.09 (0.96, 1.27)	<0.001
Non-HDL-C (mmol/l)	2.91 (2.48, 3.38)	3.23 (2.69, 3.82)	<0.001
LDL-C (mmol/l)	2.35 (1.97, 2.76)	2.61 (2.21, 3.15)	< 0.001
FBG (mmol/L)	4.81 (4.54, 5.05)	4.87 (4.58, 5.21)	0.031
Insulin (mU/L)	9.9 (6.3, 15.7)	18.1 (11.0, 28.8)	<0.001
HOMA-IR	2.129 (1.321, 3.472)	3.962 (2.418, 6.359)	<0.001
ALT (U/L)	15 (12, 20)	19 (13, 32)	<0.001
AST (U/L)	22 (19, 27)	22 (17, 29)	0.866
TBIL (umol/L)	9.9 (7.7, 13.4)	8.8 (6.9, 12.2)	0.032
DBIL (umol/L)	3.0 (2.2, 4.2)	2.7 (2.1, 3.6)	0.004
IGF-1 (ng/ml)	334.5 (235.30, 458.50)	276.0 (194.00, 391.80)	0.001
IGFBP-3 (ug/ml)	5.82 (4.99, 6.75)	5.45 (4.59, 6.29)	0.003

TABLE 1 The characteristics of the study population among children and adolescents (n = 588).

Data are presented as mean ± standard deviations, median (interquartile range), or n (%). BMI, Body Mass Index; SBP, Systolic Blood Pressure; DBP, Diastolic Blood Pressure; AO, Abdominal obesity; IGF-1, Insulin-like Growth Factor-1; IGFBP-3, Insulin-like Growth Factor Binding Protein-3; TC, Total Cholesterol; TG, Triglycerides; HDL-C, High-Density Lipoprotein Cholesterol; Non-HDL-C, Non-High-Density Lipoprotein Cholesterol; LDL-C, Low-Density Lipoprotein Cholesterol; FBG, Fasting Blood Glucose; Insulin, Fasting Insulin; HOMA-IR, Homeostatic Model Assessment of Insulin Resistance; ALT, Alanine Transaminase; AST, Aspartate Transaminase; TBIL, Total Bilirubin; DBIL, Direct Bilirubin.

compared to the high-risk group, with the differences becoming particularly pronounced between the ages of 12 and 15 years. After age 15, IGFBP-3 levels in the low-risk group plateau, while the highrisk group exhibits a slower rate of increase, with levels remaining noticeably lower after age 14 (Figure 2b).

Sensitivity analyses

In the sensitivity analysis, the results were not substantially altered when we repeated the analysis by 1) further adjusting for

TABLE 2 The association of IGF-1 and IGFBP-3 with metabolic abnormalities among children and adolescents.

	OR (95% CI)	P-value				
IGF-1 (ng/ml)						
Model 1						
Q1	Reference					
Q2	0.55 (0.34, 0.89)	0.014				
Q3	0.62 (0.38, 0.99)	0.045				
Q4	0.39 (0.23, 0.63)	<0.001				
Model 2						
Q1	Reference					
Q2	0.50 (0.26, 0.96)	0.038				
Q3	0.37 (0.19, 0.74)	0.005				
Q4	0.24 (0.12, 0.49)	< 0.001				
Model 3						
Q1	Reference					
Q2	0.52 (0.26, 1.06)	0.073				
Q3	0.33 (0.16, 0.68)	0.003				
Q4	0.24 (0.11, 0.51)	< 0.001				
IGFBP-3 (µg/ml)						
Model 1						
Q1	Reference					
Q2	0.65 (0.41, 1.04)	0.074				
Q3	0.59 (0.37, 0.95)	0.031				
Q4	0.53 (0.32, 0.85)	0.009				
Model 2						
Q1	Reference					
Q2	0.56 (0.29, 1.06)	0.076				
Q3	0.46 (0.24, 0.88)	0.020				
Q4	0.42 (0.21, 0.83)	0.013				
Model 3						
Q1	Reference					
Q2	0.50 (0.25, 1.00)	0.051				
Q3	0.42 (0.21, 0.82)	0.013				
Q4	0.38 (0.18, 0.76)	0.007				

$$\begin{split} & IGF-1; \ Q1:<214.8\ ng/ml; \ Q2:214.8-433.5\ ng/ml; \ Q3:433.5-983.00\ ng/ml; \ Q4: \ge 983.00\ ng/ml. \\ & IGFBP-3: \ Q1:<4.84\ \mug/ml; \ Q2:4.84-5.76\ \mug/ml; \ Q3:5.76-12.00\ \mug/ml; \ Q4: \ge 12.00\ \mug/ml. \\ & Model 1 \ is the un-variable model. \end{split}$$

Model 2 adjusted for age, sex, BMI, WHR, ALT, AST, TBIL, DBIL, TG, and HOMA. Model 3 was further adjusted for HDL-C and LDL-C based on Model 2.

variables (such as household income, parental education level, Supplementary Table S3), and Supplementary Table S2) categorizing IGF-1 and IGFBP-3 into tertiles (Supplementary Table S4).

TABLE 3	The	associa	tion	betv	veen	IGF-1/	IGFBP-3	ratio	and	metabo	lic
abnormal	lities	among	child	dren	and	adoles	cents.				

	OR (95% CI)	<i>P</i> -value			
Model 1					
Q1	Reference				
Q2	0.62 (0.38, 0.99)	0.045			
Q3	0.53 (0.32, 0.85)	0.009			
Q4	0.56 (0.35, 0.90)	0.018			
Model 2					
Q1	Reference				
Q2	0.72 (0.37, 1.39)	0.328			
Q3	0.45 (0.24, 0.87)	0.017			
Q4	0.45 (0.22, 0.88)	0.020			
Model 3					
Q1	Reference				
Q2	0.76 (0.38, 1.54)	0.451			
Q3	0.43 (0.21, 0.86)	0.018			
Q4	0.47 (0.22, 0.96)	0.040			

IGF-1/IGFBP-3 ratio: Q1:<41.165; Q2: 41.165-55.963; Q3: 55.963-73.337; Q4: ≥73.337. Model 1 is the un-variable model.

Model 2 adjusted for age, sex, BMI, WHR, ALT, AST, TBIL, DBIL, TG, and HOMA. Model 3 was further adjusted for HDL-C and LDL-C based on Model 2.

Discussion

In this cross-sectional study, we identified that high levels of IGF-1, IGFBP-3, and their ratio were associated with metabolic abnormalities among children and adolescents. Additionally, within the observed age range, the levels of IGF-1 and IGFBP-3 were higher in the low-risk group compared to the high-risk group.

IGF-1 is an important biomarker for the assessment of growth abnormalities (27–29) such as growth retardation or dwarfism (30) in children and adolescent populations. Notably, recent population

studies have shown that IGF-1 levels are lower when individuals suffer from conditions such as obesity (31), lipid metabolism disorders (32), hypertension (33), and metabolic syndrome (34). Meanwhile, some studies have shown that low levels of IGF-1 are associated with the components of metabolic syndrome, including elevated triglycerides (35) and reduced HDL cholesterol levels (34), which is consistent with our findings. Most of these studies have focused on older people or people suffering from other diseases. To our knowledge, only a few studies have evaluated the association of IGF-1 and IGFBP3 with metabolic abnormalities in children and adolescents, finding an association of lower IGF-1 and IGFBP-3 with metabolic abnormalities (36) and that improving the status of obesity and insulin resistance in children and adolescents may further augment the functionality of the IGF-1 axis (37). Our study found that high IGF-1 and IGBP-3 were related to metabolic abnormalities in children and adolescent populations.

Several mechanisms explain the association of IGF-1 and IGFBP-3 with metabolic abnormalities. Firstly, In the body, insulin not only regulates energy metabolism by promoting peripheral glucose uptake but also acts at high concentrations in the portal vein on hepatocytes to upregulate GH receptor expression and enhance GH-driven IGF-1 synthesis (38). Most synthesized IGF-1 circulates bound in a ternary complex with IGFBP-3 and the acid-labile subunit, which prolongs its half-life and regulates its delivery to tissues (39). IGFBP-3 levels are coregulated by GH and insulin, allowing it to both stabilize IGF-1 and, via IGF-independent mechanisms-including inhibition of IRS-1/ IR-β tyrosine phosphorylation, reduction of Akt phosphorylation and GLUT-4 translocation, and downregulation of adiponectin-to antagonize insulin signaling in peripheral tissues and promote insulin resistance (40). Second, IGF-1 affects lipid metabolism and fat storage in the body by regulating the expression of key enzymes involved in lipid metabolism such as fatty acid synthase (FAS) (41) and lipoprotein lipase (LPL) (42). In in vitro studies, IGFBP-3 significantly inhibited insulin-stimulated glucose transport and the expression of adiponectin in adipocytes, showing an insulin-antagonistic effect. Adiponectin is a hormone related to insulin sensitivity, and inhibition of its expression with



Frontiers in Endocrinology

IGFBP-3 may further exacerbate insulin resistance (43). In animal study, managing IGFBP-3 expression influences the growth and differentiation of brown preadipocytes, suggesting that IGFBP-3 plays a key role in regulating brown adipocyte fate (44). Thirdly, IGF-1 stimulates the endothelial cells to produce nitric oxide (NO), which can improve vasodilation and decrease vascular stiffness (45). Finally, IGF-1 and IGFBP-3 reduce TNF- α , IL-6, and other inflammatory indicators, which are critical for decreasing chronic low-grade inflammation in the progression of metabolic syndrome (46). Although our research lacked formal assessments of mood disorders or seasonal hormone measurements, prior studies have documented pronounced seasonal fluctuations in GH secretion (47), underscoring the importance of temporal hormonal dynamics in endocrine–metabolic research.

The strength of this study included exploring the role of IGF-1 and IGFBP-3 in childhood metabolism. However, some limitations should be acknowledged. Firstly, since the crosssectional design of our study, we cannot infer causation. Secondly, given that the study population was from China, caution should be taken in extrapolating the results to other racial or ethnic groups. Also, Our cohort did not collect Tanner staging or other formal pubertal assessments; therefore, we were unable to perform analyses stratified by pubertal status. Finally, although we adjusted for several confounders, we did not incorporate other confounders such as genetic variation that potentially affects the association.

Conclusion

In conclusion, this study found that association of higher IGF-1 and IGFBP-3 with a lower risk of metabolic abnormalities in children and adolescents. Our findings highlight that maintaining IGF-1 and IGBP-3 homeostasis is negatively associated with the risk of metabolic abnormalities in children and adolescents. Future studies are needed to explore the mechanisms as well as to further verify this association through cohort studies.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

This study was approved by Tianjin Medical University, and all research activities were conducted in accordance with the principles of the Declaration of Helsinki. Prior to any research activity, we provided all participants (or their guardians) with detailed information regarding the purpose, procedures, potential risks, and benefits of the study, and obtained their written informed consent. The privacy and personal information of participants were protected, and any published research data does not contain information that could identify individual participants.

Author contributions

ZZ: Writing – original draft, Writing – review & editing. YM: Writing – original draft. XZ: Writing – original draft. XL: Writing – review & editing. YL: Writing – review & editing. ZF: Writing – review & editing. RZ: Writing – review & editing. JL: Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. The study was supported by the National Key Research and Development Program of China (2021YFA1301202) and the National Natural Science Foundation of China (82473677, 82273676, 32400963, 92357305).

Acknowledgments

We would like to express our sincere gratitude to everyone who contributed to the writing and development of this manuscript. Their valuable insights, collaborative efforts, and continuous support were instrumental in the successful completion of this work. We appreciate the dedication and hard work of all involved throughout the process.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fendo.2025. 1579107/full#supplementary-material

10.3389/fendo.2025.1579107

References

1. Fu J, Prasad HC. Changing epidemiology of metabolic syndrome and type 2 diabetes in Chinese youth. *Curr Diabetes Rep.* (2014) 14:447. doi: 10.1007/s11892-013-0447-z

2. Magge SN, Goodman E, Armstrong SC, Committee On Nutrition, Section On Endocrinology and Section On Obesity. The metabolic syndrome in children and adolescents: shifting the focus to cardiometabolic risk factor clustering. *Pediatrics.* (2017) 140:e20171603. doi: 10.1542/peds.2017-1603

3. Camhi SM, Katzmarzyk PT. Tracking of cardiometabolic risk factor clustering from childhood to adulthood. *Int J Pediatr Obes.* (2010) 5:122–9. doi: 10.3109/17477160903111763

4. World Health Organization. *Obesity and overweight*. Available online at: https:// www.who.int/news-room/fact-sheets/detail/obesity-and-overweight (Accessed November 18, 2024).

5. Ng M, Fleming T, Robinson M, Thomson B, Graetz N, Margono C, et al. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980–2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet.* (2014) 384:766–81. doi: 10.1016/S0140-6736(14)60460-8

6. Song P, Zhang Y, Yu J, Zha M, Zhu Y, Rahimi K, et al. Global prevalence of hypertension in children: A systematic review and meta-analysis. *JAMA Pediatr.* (2019) 173:1154–63. doi: 10.1001/jamapediatrics.2019.3310

7. Jung MK, Yoo EG. Hypertriglyceridemia in obese children and adolescents. J Obes Metab Syndr. (2018) 27:143–9. doi: 10.7570/jomes.2018.27.3.143

8. Kavey RE. Combined dyslipidemia in childhood. J Clin Lipidol. (2015) 9:841-56. doi: 10.1016/j.jacl.2015.06.008

9. Aguirre GA, De Ita JR, de la Garza RG, Castilla-Cortazar I. Insulin-like growth factor-1 deficiency and metabolic syndrome. *J Transl Med.* (2016) 14:3. doi: 10.1186/s12967-015-0762-z

10. Romero CJ, Ng Y, Luque RM, Kineman RD, Koch L, Bruning JC, et al. Targeted deletion of somatotroph insulin-like growth factor-I signaling in a cell-specific knockout mouse model. *Mol Endocrinol.* (2010) 24:1077–89. doi: 10.1210/me.2009-0393

11. Yamada PM, Lee KW. Perspectives in mammalian IGFBP-3 biology: local vs. systemic action. *Am J Physiol Cell Physiol.* (2009) 296:C954–76. doi: 10.1152/ajpcell.00598.2008

12. Muzumdar RH, Ma X, Fishman S, Yang X, Atzmon G, Vuguin P, et al. Central and opposing effects of IGF-I and IGF-binding protein-3 on systemic insulin action. *Diabetes.* (2006) 55:2788–96. doi: 10.2337/db06-0318

13. Mohseni-Zadeh S, Binoux M. Insulin-like growth factor (IGF) binding protein-3 interacts with the type 1 IGF receptor, reducing the affinity of the receptor for its ligand: an alternative mechanism in the regulation of IGF action. *Endocrinology.* (1997) 138:5645–8. doi: 10.1210/endo.138.12.5714

14. Devi GR, Yang DH, Rosenfeld RG, Oh Y. Differential effects of insulin-like growth factor (IGF)-binding protein-3 and its proteolytic fragments on ligand binding, cell surface association, and IGF-I receptor signaling. *Endocrinology.* (2000) 141:4171–9. doi: 10.1210/endo.141.11.7781

15. Shamardl HAMA, Ibrahim NA, Merzeban DH, Elamir AM, Golam RM, Elsayed AM. Resveratrol and Dulaglutide ameliorate adiposity and liver dysfunction in rats with diet-induced metabolic syndrome: Role of SIRT-1/adipokines/PPARγ and IGF-1. *Daru.* (2023) 31:13–27. doi: 10.1007/s40199-023-00458-y

16. Kempf E, Landgraf K, Vogel T, Spielau U, Stein R, Raschpichler M, et al. Associations of GHR, IGF-1 and IGFBP-3 expression in adipose tissue cells with obesity-related alterations in corresponding circulating levels and adipose tissue function in children. *Adipocyte*. (2022) 11:630-42. doi: 10.1080/21623945.2022.2148886

17. Vera S, Figueroa T, Aranzalez LH, Mockus I. Cardiovascular disease risk markers in children under 10 years of age and their relationship with serum concentrations of IGF-1, IGFBP-1, IGFBP-2 and IGFBP-3. *Rev la Facultad Medicina*. (2020) 68:51–8. doi: 10.15446/revfacmed.v68n1.69979

 De Santi M, Annibalini G, Marano G, Biganzoli G, Venturelli E, Pellegrini M, et al. Association between metabolic syndrome, insulin resistance, and IGF-1 in breast cancer survivors of DIANA-5 study. J Cancer Res Clin Oncol. (2023) 149:8639–48. doi: 10.1007/s00432-023-04755-6

19. Lin J, Yang L, Huang J, Liu Y, Lei X, Chen R, et al. Insulin-like growth factor 1 and risk of cardiovascular disease: results from the UK biobank cohort study. *J Clin Endocrinol Metab.* (2023) 108:e850–60. doi: 10.1210/clinem/dga105

20. Köks S, Soomets U, Paya-Cano JL, Fernandes C, Luuk H, Plaas M, et al. Wfs1 gene deletion causes growth retardation in mice and interferes with the growth hormone pathway. *Physiol Genomics*. (2009) 37:249–59. doi: 10.1152/ physiolgenomics.90407.2008

21. Kõks S. Genomics of wolfram syndrome 1 (WFS1). Biomolecules. (2023) 13:1346. doi: 10.3390/biom13091346

22. Köks S, Luuk H, Nelovkov A, Areda T, Vasar E. A screen for genes induced in the amygdaloid area during cat odor exposure. *Genes Brain Behav.* (2004) 3:80–9. doi: 10.1046/j.1601-183x.2003.00047.x

23. Yeap BB, Chubb SA, Ho KK, Setoh JW, McCaul KA, Norman PE, et al. IGF1 and its binding proteins 3 and 1 are differentially associated with metabolic syndrome in older men. *Eur J Endocrinol.* (2010) 162:249–57. doi: 10.1530/EJE-09-0852

24. Kong AP, Choi KC, Wong GW, Ko GT, Ho CS, Chan MH, et al. Serum concentrations of insulin-like growth factor-I, insulin-like growth factor binding protein-3 and cardiovascular risk factors in adolescents. *Ann Clin Biochem.* (2011) 48:263–9. doi: 10.1258/acb.2011.010267

25. Pediatric Endocrinology and Genetic Metabolism Group of the Chinese Medical Association, Pediatric Cardiology Group of the Chinese Medical Association and Pediatric Health Care Group of the Chinese Medical Association. Definition and recommendations for prevention and treatment of metabolic syndrome in Chinese children and adolescents. *Chin J Pediatr.* (2012) 50:420–2. doi: 10.3760/cma.j.issn.0578-1310.2012.06.005

26. National Health Commission of the People's Republic of China. Screening for overweight and obesity in school-age children and adolescents (Standard No. WS/T 586-2018) [in Chinese]. Standards Press of China (2018).

27. Liu HJ, Wang LH, Chen L. Evaluation of safety and efficacy of growth hormone therapy by IGF-1 Z score in children with short stature. *Adv Ther.* (2019) 36:2374–83. doi: 10.1007/s12325-019-01021-5

28. Yoon JS, Hwang IT. Microdeletion in the IGF-1 receptor gene of a patient with short stature and obesity: a case report. *J Pediatr Endocrinol Metab.* (2021) 34:255–9. doi: 10.1515/jpem-2020-0478

29. Zhang Y, Zhang M, Chu Y, Ji B, Shao Q, Ban B. Association between growth hormone-insulin-like growth factor-1 axis gene polymorphisms and short stature in chinese children. *BioMed Res Int.* (2018) 2018;7431050. doi: 10.1155/2018/7431050

30. Baron J, Sävendahl L, De Luca F, Dauber A, Phillip M, Wit JM, et al. Short and tall stature: a new paradigm emerges. *Nat Rev Endocrinol.* (2015) 11:735–46. doi: 10.1038/nrendo.2015.165

31. Haldrup D, Wei C, Holland-Fischer P, Kristensen K, Rittig S, Lange A, et al. Effects of lifestyle intervention on IGF-1, IGFBP-3, and insulin resistance in children with obesity with or without metabolic-associated fatty liver disease. *Eur J Pediatr.* (2023) 182:855–65. doi: 10.1007/s00431-022-04731-1

32. OSAIzumi S, Ribeiro-Filho FF, Carneiro G, Togeiro SM, Tufik S, Zanella MT. IGF-1 levels are inversely associated with metabolic syndrome in obstructive sleep apnea. J Clin Sleep Med. (2016) 12:487–93. doi: 10.5664/jcsm.5672

33. Ricotti R, Solito A, Mariotti Zani E, Caputo M, Genoni G, Barone-Adesi F, et al. The relationship between cortisol and IGF-I influences metabolic alteration in pediatric overweight and obesity. *Eur J Endocrinol.* (2020) 182:255–64. doi: 10.1530/EJE-19-0792

34. Liang S, Hu Y, Liu C, Qi J, Li G. Low insulin-like growth factor 1 is associated with low high-density lipoprotein cholesterol and metabolic syndrome in Chinese nondiabetic obese children and adolescents: a cross-sectional study. *Lipids Health Dis.* (2016) 15:112. doi: 10.1186/s12944-016-0275-7

35. Sesti G, Sciacqua A, Cardellini M, Marini MA, Maio R, Vatrano M, et al. Plasma concentration of IGF-I is independently associated with insulin sensitivity in subjects with different degrees of glucose tolerance. *Diabetes Care.* (2005) 28:120–5. doi: 10.2337/diacare.28.1.120

36. Ahn M-B, Lee S-H, Choi Y-J, Kim S-K, Kim S-H, Cho W-K, et al. SAT-246 insulin-like growth factor-1 and binding protein-3 in children with metabolic syndrome. *J Endocrine Soc.* (2019) 3:SAT-246. doi: 10.1210/js.2019-SAT-246

37. Succurro E, Andreozzi F, Marini MA, Lauro R, Hribal ML, Perticone F, et al. Low plasma insulin-like growth factor-1 levels are associated with reduced insulin sensitivity and increased insulin secretion in nondiabetic subjects. *Nutr Metab Cardiovasc Dis.* (2009) 19:713–9. doi: 10.1016/j.numecd.2008.12.011

38. Nijenhuis-Noort EC, Berk KA, Neggers SJCMM, Lely AJV. The fascinating interplay between growth hormone, insulin-like growth factor-1, and insulin. *Endocrinol Metab (Seoul).* (2024) 39:83–9. doi: 10.3803/EnM.2024.101

39. Dichtel LE, Cordoba-Chacon J, Kineman RD. Growth hormone and insulin-like growth factor 1 regulation of nonalcoholic fatty liver disease. *J Clin Endocrinol Metab.* (2022) 107:1812–24. doi: 10.1210/clinem/dgac088

40. Vázquez-Borrego MC, Del Río-Moreno M, Pyatkov M, Sarmento-Cabral A, Mahmood M, Pelke N, et al. Direct and systemic actions of growth hormone receptor (GHR)-signaling on hepatic glycolysis, *de novo* lipogenesis and insulin sensitivity, associated with steatosis. *Metabolism.* (2023) 144:155589. doi: 10.1016/j.metabol.2023.155589

41. Liu P, Kong F, Wang J, Lu Q, Xu H, Qi T, et al. Involvement of IGF-1 and MEOX2 in PI3K/Akt1/2 and ERK1/2 pathways mediated proliferation and differentiation of perivascular adipocytes. *Exp Cell Res.* (2015) 331:82–96. doi: 10.1016/j.yexcr.2014.09.011

42. Tsukahara H, Gordienko DV, Tonshoff B, Gelato MC, Goligorsky MS. Direct demonstration of insulin-like growth factor-I-induced nitric oxide production by endothelial cells. *Kidney Int.* (1994) 45:598–604. doi: 10.1038/ki.1994.78

43. Kim HS, Ali O, Shim M, Lee KW, Vuguin P, Muzumdar R, et al. Insulin-like growth factor binding protein-3 induces insulin resistance in adipocytes *in vitro* and in rats *in vivo*. *Pediatr Res.* (2007) 61:159-64. doi: 10.1203/pdr.0b013e31802d8a30

44. Nguyen KH, Mishra S, Nyomba BL. *In vitro* differentiation of mouse brown preadipocytes is enhanced by IGFBP-3 expression and reduced by IGFBP-3 silencing. *Obesity (Silver Spring).* (2015) 23:2083–92. doi: 10.1002/oby.21204

45. Masodsai K, Lin YY, Lin SY, Su CT, Lee SD, Yang AL. Aging additively influences insulin- and insulin-like growth factor-1-mediated endothelial dysfunction and antioxidant deficiency in spontaneously hypertensive rats. *Biomedicines*. (2021) 9:676. doi: 10.3390/biomedicines9060676

46. Martín AI, Priego T, Moreno-Ruperez Á, González-Hedström D, Granado M, López-Calderón A. IGF-1 and IGFBP-3 in inflammatory cachexia. *Int J Mol Sci.* (2021) 22:9469. doi: 10.3390/ijms22179469

47. Köks S, Männistö PT, Bourin M, Shlik J, Vasar V, Vasar E. Cholecystokinininduced anxiety in rats: relevance of pre-experimental stress and seasonal variations. *J Psychiatry Neurosci.* (2000) 25:33–42.