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Dissolved nitrous oxide emissions associated with agricultural drainage water as influenced by manure application

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Nitrous oxide (N₂O) is highly water soluble and can be readily transported in waters draining from agricultural fields. Relatively few studies have quantified N₂O losses through agricultural tile drainage systems and none have compared the effect of different sources of applied nitrogen or their timing of application. While IPCC guidelines provide estimates of emissions from agricultural drainage water, the uncertainty in these estimates is relatively high. This research quantifies N₂O loss in tile drainage water, as influenced by nitrogen source and timing. The study site was located at Agriculture Canada research station, Harrington, PE, Canada and consisted of 12 plots with subsurface drainage systems installed at approximately 80 cm, separated by buffer drains. Three swine manure treatments were considered with inorganic fertilizer (ammonium nitrate) as a control, each replicated three times. Manure treatments included fall and spring application of solid swine manure and spring application of liquid swine manure, all applied to supply 120 kg N ha⁻¹. The magnitude of N₂O loss, as measured from samples collected at the tile outlets, demonstrated significant episodic emissions. Annual cumulative dissolved N₂O emissions ranged from 0.1 to 5.69 kg N ha⁻¹ (mean 0.83 kg N ha⁻¹), while emissions from the soil surface were 0.09–1.16 kg N ha⁻¹ (mean 0.33 kg N ha⁻¹). N₂O emissions in tile water were not significantly affected by the form of N applied, however tile drain length significantly impacted tile water N₂O concentration. IPCC coefficients for N₂O emissions from agricultural drainage water would underestimate actual N₂O emissions at this site.

KEYWORDS

nitrous oxide-N₂O, drainage water, nitrogen loss by drainage, nitrate, greenhouse gas emissions, nitrogen source, nitrogen timing

Introduction

Nitrous oxide (N₂O) is a long-lived and potent GHG with a global warming potential 298 times that of carbon dioxide and is also the leading anthropogenic cause of stratospheric ozone depletion (IPCC, 2021; WMO, 2014). Agriculture is the largest anthropogenic source of N₂O (Janzen et al., 1999; Flessa et al., 2002; IPCC, 2023; Syakila and Kroeze, 2011) producing over 60% of N₂O emissions globally from inorganic (fertilizer) and organic (manure) sources (IPCC, 2022), with annual contributions of 2.8 Gt CO₂-eq yr⁻¹ (IPCC, 2023). In Prince Edward Island (PEI), Canada, agricultural activities are responsible for

approximately 90% of human induced N_2O emissions, with direct emissions from denitrification on agricultural soils, and indirect emissions that occur after nitrogen has been transported off agricultural fields, accounting for 80% of N_2O released (Environment and Climate Change Canada, 2022). In humid and sub-humid climates such as PEI microbial denitrification is the dominant process responsible for N_2O production in agricultural soils due to anoxic conditions that frequently develop as a result of high precipitation rates (Aulakh et al., 1984; Gauder et al., 2012; Granli and Bockman, 1994; Kavdir et al., 2008).

Our understanding of global N_2O dynamics remains incomplete due to the existence of unknown or poorly quantified sources and sinks of N_2O and the complexity and interaction of mechanisms controlling the known N_2O sources (Baggs, 2011; Chapuis-Lardy et al., 2007). N_2O is produced in soils as a result of ammonia oxidation, nitrification and denitrification, and its production is affected by climatic factors such as soil temperature, moisture, and freeze-thaw cycles (Shakoor et al., 2021; Choudhary et al., 2002; Maag and Vinther, 1996; Granli and Bockman, 1994; Gillam et al., 2007), as well as management practices such as nitrogen addition, tillage, and cropping system (Venterea et al., 2011). There is a need to measure N_2O emissions from a greater range of agricultural sources and from a greater diversity of areas, both regionally and within the landscape, to allow improved quantification of national N_2O emissions inventories (Rochette and McGinn, 2008). Understanding the spatial and temporal nature of emission processes is also important in refining and accurately estimating emissions. The IPCC recognizes indirect N_2O emissions from agriculture, including those resulting from nitrogen leaching and runoff (N_2O_L). There is a greater degree of uncertainty in indirect N_2O emissions, as fewer studies have quantified these sources (Nevison, 2000; Quesnel et al., 2019; ECCC, 2022). Furthermore, indirect sources can be difficult to measure, since emissions are removed from their point of origin (Reay et al., 2003; Roper et al., 2013) and, in the case of dissolved N_2O emissions, may be confounded by subsurface soil interactions (Quesnel et al., 2019). The steps outlined by the IPCC in calculating indirect N_2O emissions involve several assumptions, relying on general nitrogen cycling principles and a limited amount of field data (ECCC, 2022). There is a need for further quantification and study of the factors influencing indirect N_2O emissions.

Leaching and run off are estimated to account for more than 75% of indirect N_2O emissions from agriculture, with leaching being the largest contributor, and represents one of largest sources of uncertainty in estimates of agricultural N_2O emissions (Nevison, 2000; Tian et al., 2019). In Canada's National Inventory Report to the IPCC the fraction of nitrogen that is lost through leaching and runoff ($\text{FRAC}_{\text{LEACH}}$) is estimated as a function of the ratio of precipitation to evapotranspiration and ranges from 5% in the more arid regions of the country to 30% in the more humid regions of the country (ECCC, 2022). The percentage of this nitrogen lost as N_2O (N_2O_L) from N leached from agricultural soils is estimated using leaching/runoff emission factor (EF_5) of $0.0075 \text{ kg N}_2\text{O-N kg}^{-1} \text{ N leached}$ (IPCC, 2019). Nevison (2000) indicated several uncertainties in the proposed IPCC methodology for calculating the N_2O emissions through leaching and run off. Since emissions from this source are likely to be significantly influenced by climate there is a need for data from a broader

range of agroecosystems. The factor further assumes a relationship between N_2O and NO_3^- concentration in agricultural drainage water. This relationship has not been well documented with some studies (Sawamoto et al., 2003) showing little or no relationship between N_2O and NO_3^- in drainage waters, with high dissolved N_2O concentrations in the fall corresponding with low NO_3^- levels. This disconnect was explained by spatial differences in where these processes were occurring; increased dissolved N_2O concentrations were attributed to subsoil denitrification which occurred during or after NO_3^- leaching had occurred (Sawamoto et al., 2003). As a result, there is a need to develop appropriately documented regional emission coefficients.

Subsurface tile drainage is a management practice used in imperfectly and poorly drained agricultural fields to prevent water logging, lower a perched water table, and drain excess water from fields (Valayamkunnath et al., 2022). Subsurface tile drainage also decreases soil structural damage, erosion, and increases the length of the effective growing season by allowing earlier access to fields (Valayamkunnath et al., 2022). In Atlantic Canada, tile drainage water flows intermittently during year and drainage events can occur during the winter period, when temperatures in the soil profile rise above freezing. Peak drainage events typically occur during spring thaw, which may coincide with peak surface N_2O emissions. In northern climates, up to 70% of N_2O emissions from agricultural soils are thought to occur during spring thaw, as a result of enhanced microbial activity and the release of N_2O trapped under a frozen ice layer (Risk et al., 2013; 2014; Wagner-Riddle et al., 2017). Tile drains intercept the flow of the water draining from the root zone and its composition is often considered to be representative of the nutrients draining from an agricultural field (Valayamkunnath et al., 2022). Several studies have documented that loss of nitrogen through subsurface tile drainage is substantial (Nangia et al., 2010; Quesnel et al., 2019; Milburn et al., 1997; Drury et al., 1993; Randall et al., 1997). In more humid agroecosystems such as PEI, it has been concluded that 15%–50% of the applied nitrogen reaches surface water via subsurface drainage (Jiang et al., 2012). Most studies of nitrogen loss in tile drainage have focused on the losses of NO_3^- and NH_4^+ (Drury et al., 1993; Klavicko et al., 1991; Randall et al., 1997), however fewer studies have examined dissolved N_2O (Bruun et al., 2017; Hack and Kaupenjohann, 2002; Hama-Aziz et al., 2017; Reay et al., 2004; Reay et al., 2009; Quesnel et al., 2019).

Nitrous oxide produced below the soil surface results in the accumulation of N_2O in the soil profile due to impeded gas transport by water-filled pores and/or formation of frozen layer during winter and spring thaw (Burton and Beauchamp, 1994; Heincke and Kaupenjohann, 1999). Nitrous oxide is highly soluble in water and its solubility increases as the water temperature decreases (DelVecchia et al., 2023). During winter and spring, cold water moves through the soil profile when N_2O concentrations are elevated (Burton and Beauchamp, 1985). These conditions result in a high potential for N_2O transport and loss in tile drainage water (Valayamkunnath et al., 2022). Moreover, agricultural fields with tile drainage expedite the movement of water from the field and therefore decrease the likelihood of further N_2O reduction in the subsurface (Hack and Kaupenjohann, 2002; Valayamkunnath et al., 2022). Dowdell et al. (1979) noted significant amounts of dissolved N_2O ($8.1\text{--}277.3 \mu\text{g N}_2\text{O-N L}^{-1}$) in water draining from agricultural

fields. Similarly, other studies have variously reported dissolved N_2O concentrations in tile drainage water from agricultural systems ranging from 0.3–1,108 $\mu\text{g N L}^{-1}$ (Bruun et al., 2017; Hama-Aziz et al., 2017; Quesnel et al., 2019; Reay et al., 2003; 2004; 2009; Roper, 2008).

Data is lacking on the impact of N input form (synthetic fertilizers vs. manure) and timing on dissolved N_2O losses from agricultural fields. While several studies have examined dissolved N_2O losses in agricultural fields under manure inputs (Reay et al., 2009; Quesnel et al., 2019), none have compared different manure/N application treatments. Furthermore, the timing of manure application to agricultural fields is known to impact NO_3 leaching, with fall manure applications elevating leaching risk relative to spring (van Es et al., 2006). It is unknown whether this also applies to dissolved N_2O losses. Currently, there is limited measured data documenting N_2O concentrations and loss from tile drainage systems, particularly during the winter period. The effect of soil management, such as various cropping systems and fertilizer applications, on N_2O losses via tile drainage systems also needs more study. The quantification of N_2O production during the non-growing period and its transport from the soil profile into the tile drainage water are needed to provide a more complete picture of the impacts of agricultural management on indirect N_2O emissions and identify opportunities for mitigation. Furthermore, to date only one study has considered the relative magnitude of dissolved N_2O loss compared to surface N_2O emissions and NO_3 leaching (Quesnel et al., 2019).

This paper investigates N_2O losses in agricultural tile drainage water from a 3-year potato cropping rotation in PEI, Canada. Objectives of the study were to 1) quantify and compare the annual temporal variation in N_2O and NO_3^- concentrations in tile drainage systems and surface N_2O emissions in PEI 2) determine the impact of swine manure treatment form (solid and liquid) and timing (fall and spring application) on dissolved N_2O losses in tile drainage water and 3) evaluate the EF₅ for predicting N_2O emitted from nitrogen lost through leaching in PEI.

Materials and methods

Study site

The field experiment was initiated in 2000 at Agriculture and Agri-Food Canada's Harrington Research Farm, situated 14 km north of Charlottetown, PE (46° N lat, 63° W long). The research site covers an area of 2.9-ha with a gentle slope of 1%–3%. Between 1986 and 1989 the site was divided into 13 subsurface drainage plots, each with dimensions ranging from 0.21 to 0.28 ha (Milburn et al., 1997). Tile drainage systems were installed to account for individual plot size and drainage capacity. Each plot contained two tile drains within the plot, and individual plots were separated by buffer drains to limit cross-contamination from adjacent plots due to subsurface or surface flow crossing plot boundaries. Three shorter tile drains were installed in plots 8, 9 and 10 to accommodate the irregular shape of the field. The tile drains were spaced at an interval of 12 m in plots 8, 9 and 10, and 15 m in the remaining plots. The tiles were placed at an average depth of 0.80 m and the drains discharged into tipping buckets housed in a heated discharge building.

The soils at the experimental site are Charlottetown sandy loam (Orthic Humo-Ferric Podzol), with a Malpeque sandy loam (Gleyed Eluviated Drystic Brunisol) occurring in plots 7, 8, 10, and 11 in the lower portions of the field, and a soil pH of 6.6 (Milburn et al., 1997). Charlottetown soils tend to be well drained, while Malpeque soils are imperfectly drained (MacDougall et al., 1988). Daily precipitation and soil temperature was recorded at the Harrington research farm meteorological station, located less than 1 km from the experimental site.

Nitrogen fertility treatments and cropping practices

The agronomic treatments consisted of 4 N fertility treatments applied prior to potato phase of the potato-barley-red clover rotation. The N treatments were: solid and liquid swine manure applied in the spring prior to planting of potatoes; solid manure applied in the fall prior to the potato year and NH_4NO_3 applied in the spring. Each treatment was replicated three times and the rate of application adjusted to supply 120 kg N ha⁻¹. Swine manure was generated in an adjacent facility that produced both solid and liquid manure (Campbell et al., 2003). During fall 2000 and 2003, solid swine manure was applied, while in spring 2001 and 2004 solid swine manure, liquid swine manure and ammonium nitrate were applied (Table 1). The experimental site was cropped to potato (*Solanum tuberosum* (L.) var. Shepody) in 2001 and 2004, to barley (*Hareem vulgare* (L.) var. Iona) underseeded to red clover (*Trifolium pratense* (L.) var. Marino) in 2002, and to red clover in 2003. In the spring of the barley year all plots received 60 kg N ha⁻¹ as NH_4NO_3 .

Water sampling and analysis

Water sampling for dissolved N_2O

The concentration of N_2O dissolved in tile drainage water was measured by collecting grab samples at the outlet of the drainage tiles during flow events in 2002, 2003 and 2004 (see supplementary for details on sample collection process). Samples were collected on a weekly basis or when drainage events were occurring. The diurnal pattern of N_2O emissions was examined by collecting samples every 4 hours over a 3-day flow event on April 27–29, 2004.

The NO_3^- and NH_4^+ content and temperature of tile water discharge was determined from water samples collected as part of a parallel study using ISCO 6712 automatic samplers (Teledyne ISCO Inc., Lincoln, NB) (Jiang et al., 2011) (see supplementary for details on sample collection process).

During the 2002 to 2004 observation period the frequency of sampling depended upon tile flow. During flow events samples were collected between 0800 and 0900 h. The flow rate for each tile drain was calculated by multiplying the total number of tips of the tipping bucket by the amount of water each bucket holds. The tipping buckets were wired to a data logger (Campbell Scientific, Logan, UT). Tile drainage water N_2O emissions were calculated by accounting for individual plot flow rate on each sampling date. Cumulative N_2O emissions were calculated by linear interpolation of N_2O concentrations over the sampling dates for each flow event

TABLE 1 Management practices for the experimental site from 2000 to 2004.

Year	Management practices
2000	Red clover
	Oct. 2 - Fall application of solid manure treatment at a rate to supply 120 kg N ha ⁻¹ Oct. 16 - Incorporation of manure and plow down of red clover crop
2001	Potato
	May 16 - Spring application and incorporation of ammonium nitrate, liquid manure, and solid manure treatments at a rate to supply 120 kg N ha ⁻¹ May 21 - Planting of potato crop Oct 10 - Harvesting of potato crop
2002	Barley
	May 28 - Planting of barley under-seeded with red clover May 28 - Application and incorporation of ammonium nitrate at 40 kg N ha ⁻¹ Sept. 4 - Harvest of barley crop
2003	Red clover
	July 9 - Red clover 1st cut Oct. 2 - Fall application of solid manure treatment at a rate to supply 120 kg N ha ⁻¹ Oct. 16 - Incorporation of manure and plow down of red clover crop Oct. 31 - Herbicide (Transorb @ 3 L ha ⁻¹) application
2004	Potato
	May 17 - Spring application of ammonium nitrate, liquid manure, and solid manure treatments at a rate to supply 120 kg N ha ⁻¹

and these concentrations applied to daily water flow as measured by the tipping bucket flow meters.

There is uncertainty as to the degree of degassing of N₂O that occurs as water runs along the tile drainage pipes. The influence of tile length on N₂O concentration was evaluated by sampling the three in-field tile drainage systems at plot #8, which was in addition to the 12 experimental plots where the agronomic treatments were replicated. Plot eight received the treatment of 120 kg N ha⁻¹ NH₄NO₃. The total length of each tile drain in this plot was 81.5 m (see supplementary for details on sample collection process).

Stream and spring water sampling

On 28 May 2004, water samples were collected from streams and springs to compare their dissolved N₂O concentration with that of subsurface drainage water. The stream was situated adjacent to the research site, with subsurface drainage water flowing into it. Spring samples were also collected at a nearby location. Stream and spring water samples were collected by 5 mL disposable syringe (Luer-lok tip, latex free, Becton-Dickinson, Franklin Lakes, NJ), injected immediately in an exetainer (12 mL) with sodium azide and sealed with silica sealant.

Water sample analysis

Vials containing water samples for N₂O analysis were equilibrated at room temperature (25°C) before being analyzed for headspace N₂O concentration. N₂O analysis was performed

using a Varian Star 3,800 Ga Chromatograph (Varian, Mississauga, ON) fitted with an electron capture detector with a Combi-PAL autosampler (CTC Analytics, Zwingen, Switzerland). The autosampler removes a 2.5 mL volume from the sample tube and injects this into a sample valve that delivers 0.1 mL to the ECD. The ECD was operated at 300°C, 90% Ar, 10% CH₄ carrier gas at 20 mL min⁻¹, Haysep N 80/100 pre-column (0.32 cm diameter x 50 cm length) and Haysep D 80/100 mesh analytical columns (0.32 cm diameter x 200 cm length) in a column oven operated at 70°C. Pre-column was used in combination with a four-port valve to remove water from the sample. Operational conditions and data handling was performed with Varian Star software. In each analytical run of 147 samples a single replicate of three concentrations of standard gas mixtures were included for standardization of the instrument. The lab temperature was recorded during the analysis of dissolved N₂O concentration. Total N₂O dissolved in the water sample is the sum of N₂O in the headspace and N₂O dissolved in water. The molar mass of N₂O in the headspace was calculated from the Ideal gas law (Equation 1).

$$M_{N_2O_g} = \frac{(P_{N_2O} \times 10^{-6}) \times V_{N_2O}}{R \times T} \quad (1)$$

Where: M_{N₂O_g} = moles of N₂O in vial headspace (mol); P_{N₂O} = partial pressure of gas in the headspace (atm); V_{HS} = Volume of headspace (L); R = Gas constant (0.08214 atm/mol k); and T = lab temperature (°K).

Henry's law (Equation 2) describes the partitioning of a compound between a dilute aqueous and the gas phase:

$$C_{N_2O} = H_{N_2O} \times P_{N_2O} \quad (2)$$

Where: C_{N₂O} = Nitrous oxide concentration in the aqueous solution (moles/L); H_{N₂O} = Henry's law constant for nitrous oxide (2.5 × 10⁻² M/atm), and P_{N₂O} = partial pressure of gas in the headspace (atm).

M_{N₂O_l} (Equation 3) is calculated from C_{N₂O} present in V (5 mL) of the aqueous solution (1 mL of sodium azide and 4 mL of water sample).

$$M_{N_2O_l} = C_{N_2O} \times V \quad (3)$$

Where: M_{N₂O_l} = Moles of the nitrous oxide in dissolved in water (moles); V = Volume of the aqueous solution in the vial (L).

Therefore, the total number of moles of nitrous oxide in the vial (Equation 4), less the estimated contribution of nitrogen by sodium azide (5.3 × 10⁻⁸ mol) is,

$$T_{N_2O} = (M_{N_2O_g} + M_{N_2O_l}) - (5.3 \times 10^{-8}) \quad (4)$$

Where: T_{N₂O} = Total moles of nitrous oxide in head space and aqueous solution (moles).

The flux of dissolved N₂O (g N ha⁻¹ d⁻¹) was calculated by multiplying the dissolved N₂O concentration by the volume of tile drainage discharge per unit area (L ha⁻¹) as measured by the tipping bucket gauges and the area of the plot drained by the tile system.

Nitrate and ammonium concentration of the water samples were determined colorimetrically using a TRAACS 800 analyzer (Milburn et al., 1997). Dissolved organic carbon was analyzed by Formacs TOC analyzer (Hasegawa et al., 2000).

Nitrous oxide production from sodium azide

An experiment was carried out to determine if the sodium azide produced any N₂O during storage. Sodium azide solution was prepared at the same concentration as used in the exetainers for dissolved N₂O sample collection (0.1 M). One mL of NaN₃ was injected into a 12 mL exetainers using a 1 mL syringe (Luer-lok tip, latex free, Becton-Dickinson, Franklin Lakes, NJ), followed by flushing with helium and evacuation. After evacuation, 4 mL of nano pure water was transferred to the exetainer by means of a syringe (5 mL, Luer-lok tip, latex free, Becton-Dickinson, Franklin Lakes, NJ), ensuring a positive pressure inside the exetainer. Some vials were analyzed immediately for head space N₂O concentration, while others were kept at lab temperature and analyzed after 1 month to determine the extent of N₂O production from the sodium azide. Sodium azide was found to contribute background levels of N₂O of approximately 5.3×10^{-8} mol of N₂O.

Nitrous oxide flux at the soil surface

N₂O flux measurements were made using a non-flow-through, non-steady-state (NFT-NSS) chamber with a total volume of 1.6 L covering a soil area of 315 cm². Deployment times were kept to 30 min with samples being collected at 0, 10, 20 and 30 min. Replicate (5) samples of the atmosphere at chamber height at the time of deployment were used to estimate the ambient (time zero) concentrations. Samples were collected by removing 20 mL of gas from the headspace of the chamber and injecting into previously evacuated (to 500 millitorr) 12 mL Exetainers (Labco, UK), each containing 4 mg of Magnesium Perchlorate-a desiccant to remove water from the gas samples. Five replicates of two standard gases were collected on each sampling occasion and were used to confirm the integrity of sample storage and handling.

N₂O flux measurements were made from 9 May to 26 September 2001, 14 May to 28 August 2002 and 16 May to 27 November 2003, and 10 March to 27 May 2004. Flux sampling conducted during the snow cover period (March 10 and 25, 2004), was carried out by inserting chambers into the snow (2–3 cm depth), and with samples collected in a similar manner as in the snow free period described. Two chambers were inserted in each plot. In 2001, when cropped to potato, these two chambers were separated by location: One chamber was installed in the hill and one in the furrow row locations within each plot. The hill and furrow locations were measured separately due to the distinctly different soil, nutrient and crop growth environments within these two locations. The chambers were removed and replaced to accommodate any management practices in the experimental site.

N₂O flux (F_{N_2O}) was calculated by the following Equation 5 (Hutchinson and Livingston, 1993):

$$F_c = \left(\frac{dC}{dt} \right) \left(\frac{V_c * M_{mol}}{A * V_{mol}} \right) \quad (5)$$

Where, dC/dt is the rate of change of N₂O concentration inside the chamber (g N h⁻¹), A is the surface area of the chamber (0.0315 m²), V_c is the total volume of the chamber (1.62 L), M_{mol} is molar mass of N₂O (44 g/mol) and V_{mol} is the volume

of a mole of N₂O inside the chamber calculated from the ideal gas law based on temperature and pressure. The relative change in N₂O concentration with time (dC/dt), inside the chamber, was calculated by simple linear regression. Cumulative N₂O-N losses were calculated by linear interpolation between sampling dates.

N₂O analysis was performed using a Varian Star 3,800 Ga Chromatograph using the same method as for headspace N₂O concentration of the water samples.

Statistical analysis

An ANOVA on dissolved N₂O concentration was performed as a randomized complete block (RCB) design, according to treatment for each sampling date. Correlation analysis was done between dissolved N₂O concentration and NO₃⁻, surface N₂O flux, dissolved organic carbon, tile flow, water temperature, and plot area. Correlation analysis was also done between cumulative N₂O loss and tile length of the experimental plots. Regression analysis was performed between cumulative N₂O and NO₃⁻ loss in subsurface tile drainage water. All statistical analysis was conducted using Statistical Analysis System (SAS institute, 1990).

Results and discussion

Climatic data

Two of the three non-growing periods examined had lower than normal precipitation (Table 2). Cumulative non-growing season precipitation (November to May) for 2001–2002 and 2003–2004 was 531 and 380 mm respectively significantly lower than the long-term (1971–2000) average of 701 mm (Table 2) (Government of Canada, 2019). In 2002–2003 higher than average rainfall resulted from four high rainfall events including a 90 mm precipitation event (Table 2; Figure 1B). The duration and extent of soil freezing varied considerably between years (Figure 1) despite the average non-growing season air temperature being similar over the three winter periods. In 2001–2002 the lowest soil temperature at 5 cm was -0.2°C and the soil was only frozen for 24 days (March 6 to March 30; Figure 1A). In contrast, in 2002–2003, the soil was frozen to a depth of 5 cm for 119 days (November 27 to April 29) and was frozen to 10 cm for 93 days (January 16 to April 19; Figure 1B). In 2003–2004, the soil at 5-cm depth was frozen for 41 days (January 5 to February 15) and at 10-cm depth for 22 days (January 13 to February 4; Figure 1C). Differences in soil freezing despite small differences in average air temperatures reflect differences in snow cover with the 2002–2003 period having relatively little precipitation in the early part of the winter and therefore having little or no snow accumulation. The tile flow for the observation period ranged between <1 and 54.5 mm per day (data not shown). The cumulative tile flow for the 2002, 2003 and 2004 calendar years was 156, 99, and 68 mm respectively. The greater cumulative tile flow in 2002 corresponded to the milder temperatures during the 2001–2002 overwinter period. Peak tile flow was observed during spring thaw and following precipitation events.

TABLE 2 Mean temperature and total precipitation at the experimental site during the growing (May–October) and non-growing (November–April) periods.

		Mean temperature (°C)	Total precipitation (mm)
2001	Growing	15.4	394
	Non-Growing	-2.4	531
2002	Growing	13.5	643
	Non-Growing	-2.2	746
2003	Growing	14.7	503
	Non-Growing	-3.3	380
2004	Growing	13.9	363
	Non-Growing	-3.3	470

Dissolved N₂O emissions

Effect N treatments on dissolved N₂O

Dissolved N₂O concentrations in tile water discharge exhibited significant differences between sampling dates ($p = 0.001$) but were not significantly affected by nitrogen source (NH₄NO₃, solid manure and liquid manure) or timing (spring and fall application) of N application to the potato rotation (Table 3A; Table 3B; Figure 2). There are several possible explanations for this. In their meta-analysis examining the impact of manure applications on surface N₂O emissions in agricultural soils, Xia et al. (2020) found that the application of manure and synthetic fertilizers induced similar magnitude N₂O emissions when applied at the same N rate. While the overall emission factor following manure application (1.11%) is slightly lower than that of synthetic N fertilizer (1.25%), manure N₂O emissions were also impacted by factors such as C:N ratio and C content (Xia et al., 2020), which resulted in similar levels of emissions produced. It is reasonable to assume that this would also apply to dissolved N₂O below the soil surface. Additionally, while N fertilizer application has been found to be associated with elevated concentrations of dissolved N₂O and NO₃⁻ in water draining from agricultural fields (Hasegawa et al., 2000), since the processes responsible for producing dissolved N₂O in drainage water are spatially and temporally complex, the relationship between dissolved N₂O and N application rate may not be straightforward (Reay et al., 2004; Quesnel et al., 2019). For example, fertilizers and manures are applied and measured at or near the soil surface and can subsequently be transported by leaching and surface runoff. In contrast, N₂O can be produced throughout the soil profile; therefore, even after NO₃⁻ has been depleted from the soil surface, N₂O can still be being produced in the soil, resulting in a temporal disconnect in the presence of these substances due to spatial differences in where they are measured and/or produced. It is possible that these same processes could make it difficult to detect differences in N source treatments. Failure to detect differences in treatments may have also resulted from differences in the length of drainage pipe from the plot to the collection station. We noted an inverse relationship between the lengths of the tile lines and averaged observed dissolved N₂O concentrations (Figure 3). To confirm this observation air and water sampling access ports were inserted at three locations along the perforated portion of the tile line and at one location just outside the plot on an adjacent site. Average dissolved

N₂O concentrations over ten sampling events were observed to decrease from 1.1 mg N₂O-N L⁻¹ to 0.3 mg N₂O-N L⁻¹ (Figure 4). These observations suggest that degassing of N₂O was occurring in the tile line and that concentrations measured at the sampling hut may underestimate total N₂O leaving the field. Normalizing all results for length of tile line did not, however, reveal significant N treatment effects. For the remainder of the discussion, we will consider trends observed across all 12 experimental units independent of N treatment.

Cumulative dissolved and surface N₂O losses

Annual cumulative dissolved N₂O emissions ranged from 0.01 to 5.69 kg N ha⁻¹ (mean 0.83 kg N ha⁻¹) which were slightly higher than annual cumulative N₂O emissions occurring from the soil surface of 0.09–1.16 kg N ha⁻¹ (mean 0.33 kg N ha⁻¹) (Table 3A). The dissolved N₂O-N loss through subsurface drainage water was an order of magnitude less than NO₃⁻-N, which ranged from 0.26 to 29.3 kg N ha⁻¹ (mean 8.7 kg N ha⁻¹; Table 3A). The dissolved N₂O results in our study are higher than other values of dissolved N₂O in tile drains reported in the literature, which ranged from 0.03–0.38 kg N₂O-N ha⁻¹yr⁻¹ (Dowdell et al., 1979; Hack and Kaupenjohann, 2002; Reay et al., 2004; 2009; Quesnel et al., 2019). Similarly, Reay et al. (2009), observed annual dissolved losses of N₂O in agricultural drainage that were higher than the amount being emitted at the soil surface. In contrast, Bruun et al. (2017) found that the main export pathway was dissolved N₂O, with negligible N₂O produced at the soil surface, and both Dowdell et al. (1979) and Hack & Kaupenjohann, (2002) concluded that N₂O loss in agricultural drainage water was a minor component of the soil N balance. However, Quesnel et al. (2019) found that cumulative annual soil surface N₂O emissions (13.8 kg N₂O-N ha⁻¹) were much higher than dissolved N₂O losses (0.03 kg N₂O-N ha⁻¹) in tile drainage systems. Hama-Aziz et al. (2016) observed concentrations of dissolved N₂O in agricultural tile drainage systems were comparable to NO₃⁻ concentrations. Our results indicate that dissolved N₂O emissions from potato cropping systems can be a major source of agricultural N₂O emissions in PEI—comparable or higher than surface N₂O emissions. This is likely because the majority of precipitation occurs in the non-cropping season in PEI, when conditions at the soil surface may be less favorable for denitrification and may prevent the release of N₂O produced in deeper soil layers to the atmosphere. However, large amounts of NO₃⁻ are still lost from these tile drainage systems.

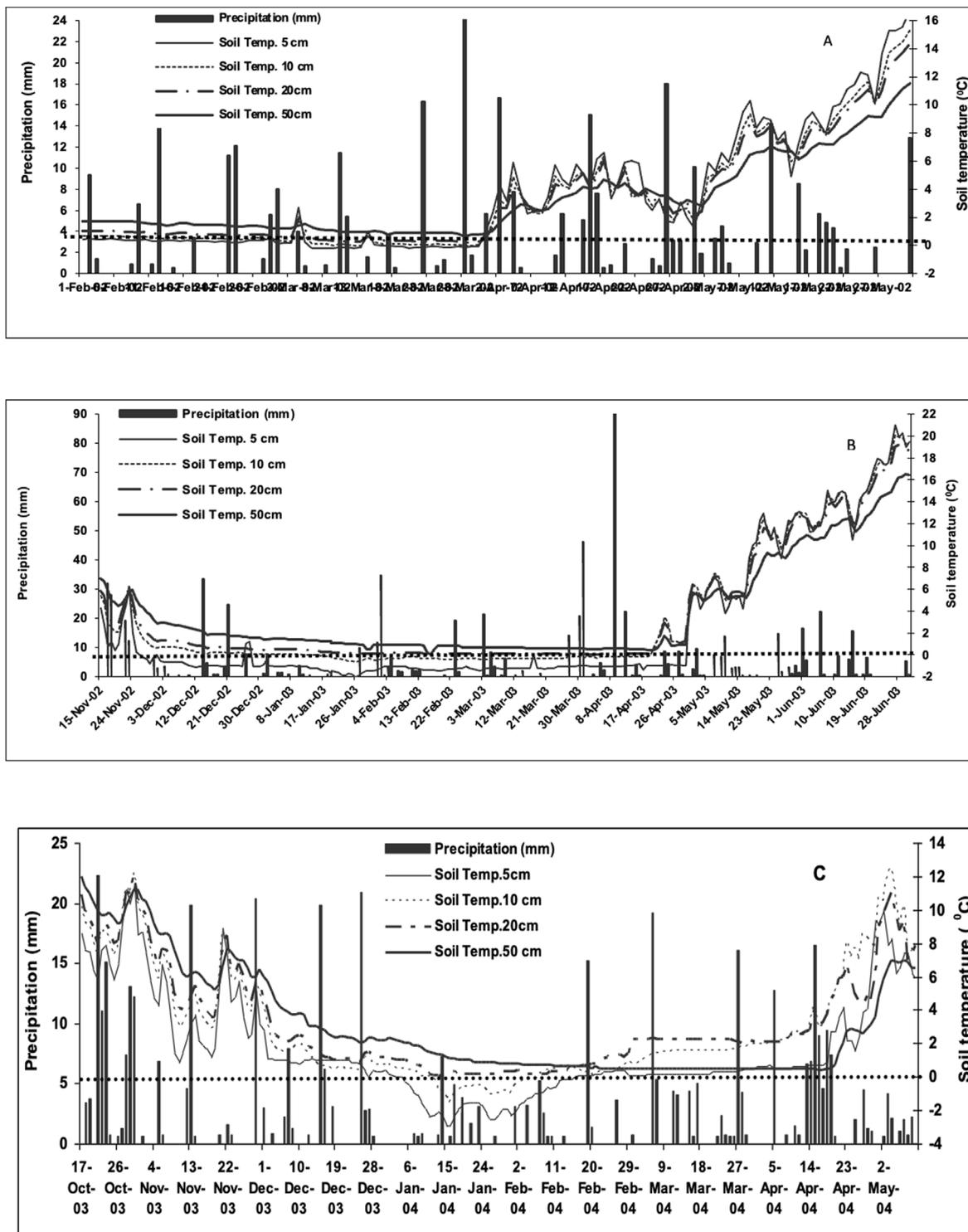


FIGURE 1 Precipitation and soil temperature during fall to spring periods at the Harrington Research station from (A) February - May 2002, (B) November 2002 to June 2003 and (C) October 2003 to May 2004.

Temporal variation in dissolved N₂O

N₂O loss in tile drainage water was strongly influenced by season with up to 62% of N₂O emissions occurred during the non-growing season (Figure 2A). One or two high N₂O emission events can contribute a significant amount for that particular year (Figure 2B).

Dissolved N₂O concentrations exhibited high temporal variability and were independent of the volume of tile flow (data not shown). For instance, in 2003, the highest and lowest dissolved N₂O concentrations (3.8 mg N L⁻¹, 0.01 mg N L⁻¹) were observed within a timespan of several days during the same flow event. A study of dissolved N₂O

TABLE 3 (Panel A) Mean and range of values (12 plots) for cumulative annual N₂O and NO₃ dissolved in drainage water and surface N₂O flux from research plots as measured during the 2002, 2003 and 2004 monitoring periods and (Panel B) mean and range of values (3 replicates) for cumulative annual N₂O dissolved in drainage water from each N treatment as measured during the 2002, 2003 and 2004 monitoring periods.

Year	N ₂ O surface flux	N ₂ O dissolved in drainage water	Drainage as % of total N ₂ O loss	NO ₃ dissolved in drainage water
(kg N ha ⁻¹)				
Potato	0.46 (0.16–1.16)			
Winter 2002		0.54 (0.01–1.82) [†]	54%	16.9 (1.0–29.3)
Barley	0.15 (0.09–0.23)			
Winter 2003		1.73 (0.02–5.69)	82%	4.5 (0.14–12.6)
Red Clover	0.37 (0.18–0.48)			
Winter 2004		0.23 (0.02–0.42)	38%	4.8 (0.26–11.1)
Year	NH ₄ NO ₃	Spring liquid manure	Spring solid manure	Fall solid manure
(kg N ha ⁻¹)				
Potato				
Winter 2002	0.30 (0.17–0.53)	0.25 (0.01–0.67)	0.97 (0.01–1.82)	0.63 (0.06–1.3)
Barley				
Winter 2003	1.67 (1.22–2.13)	0.96 (0.02–1.47)	1.32 (0.13–2.34)	3.00 (0.54–5.69)
Red Clover				
Winter 2004	0.19 (0.11–0.24)	0.19 (0.01–0.31)	0.27 (0.03–0.42)	0.26 (0.10–0.41)

[†]Mean (range of value) for 12 plots.

[‡]Mean (range of value) for 3 replications.

emissions over a 36-h monitoring period observed a mean concentration of 0.48 mg N L⁻¹, a coefficient of variation of 46%, with values ranging from 0.13 to 0.81 mg N L⁻¹. Reay et al. (2003) have observed similar variation in dissolved N₂O in agricultural drainage waters. Peak dissolved N₂O concentrations (4.2 mg N L⁻¹) recorded in this study were higher than those reported by others (Dowdell et al., 1979; Hack and Kaupenjohann, 2002; Hasegawa et al., 2000; Minami and Fukushi, 1984; Minami and Oshawa, 1990; Reay et al., 2003; Sawamoto et al., 2002; Sawamoto et al., 2003).

Over the monitoring period dissolved N₂O concentrations of water collected from tile lines ranged from less than 0.01 mg N L⁻¹–4.2 mg N L⁻¹ (Figure 2B). The highest dissolved N₂O concentrations occurred during winter 2003 (Figure 2B). A continuous frozen layer was present from 27 November 2002 to 29 April 2003, which may have prevented gas phase transport of N₂O to the soil surface and resulted in an accumulation of N₂O in the atmosphere of subsurface layers (Burton and Beauchamp, 1994). Elevated N₂O concentrations in the soil atmosphere would result in greater N₂O dissolution in soil water and a supersaturated soil solution. As N₂O is highly soluble in cold water, there is a high probability that the water draining through the soil profile at this time would contain a high concentration of dissolved N₂O (Goodroad and Keeney, 1984). In the tile flow of 2001–2002 and 2003–2004, N₂O emissions were much lower than 2002–2003. The winter pattern is similar for both years, with the soil at 5 cm depth freezing only for a few days at a time. Furthermore, there was precipitation at regular intervals from November to March in both 2001–2002 and 2003–2004 (Figures

1A, C). The lack of a continuous frozen layer and regular precipitation would favor more rapid NO₃⁻ leaching, reducing the opportunity for N₂O production and accumulation in the subsurface. The relative contribution of surface and sub-surface pathways appears to site-specific and may vary significantly from year to year.

Relationship between dissolved NO₃ and dissolved N₂O

During the observation period, dissolved NO₃⁻ emissions from tile lines fluctuated between 0.09 and 39 mg N L⁻¹ (Figure 2B). In general, the NO₃⁻ concentrations observed in this study are comparable to previous studies conducted at this site (Milburn et al., 1997) and to values reported by others (Dowdell et al., 1979; Hack and Kaupenjohann, 2002; Hasegawa et al., 2000; Minami and Fukushi, 1984; Minami and Oshawa, 1990; Quesnel et al., 2019; Reay et al., 2003; Sawamoto et al., 2002; Sawamoto et al., 2003). The timing of manure and fertilizer applications to the potato crop (fall 2000, spring 2001, fall 2003, spring 2004) influenced the magnitude of NO₃⁻ loss in tile water. After the potato harvest in 2001, the research plots were left fallow. The NO₃⁻ not utilized by the potato crop and accumulating in the soil increased the loss of NO₃⁻ in the tile flow during 2001–2002 non-growing period (Figure 2B). Low rates of nitrogen fertilizer (40 kg N ha⁻¹) were applied to barley crop across all N treatments in 2002, reducing fall NO₃⁻ accumulation in the soil and the loss of NO₃⁻ via tile drainage flow during the 2002–2003 non-growing period (Figure 2B).

There was a significant ($p < 0.01$) negative correlation between dissolved N₂O-N and NO₃⁻-N concentration for individual

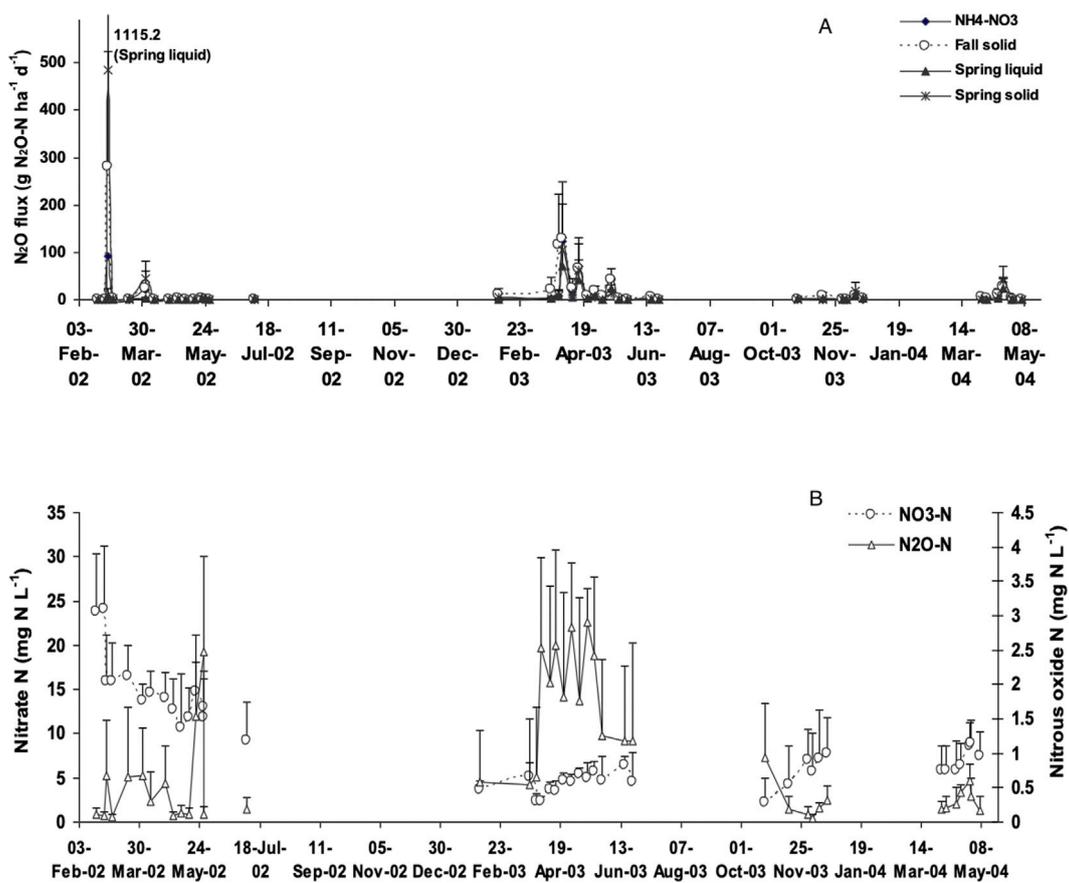


FIGURE 2 (A) Treatment effect on N₂O flux from agricultural drainage water, and (B) N₂O-N and NO₃-N concentration in weekly samples of agricultural drainage water.

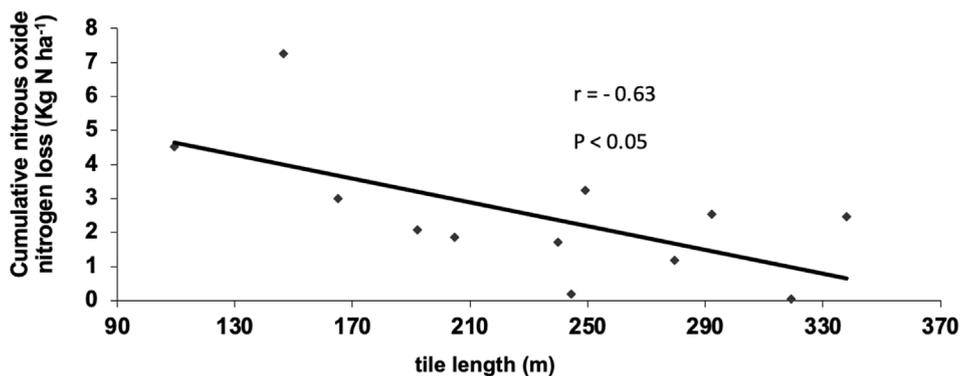


FIGURE 3 Relationship between cumulative N₂O-N loss and tile length.

observations, but this accounted for less than 10% of the total variation in the data ($R^2 = 0.05$; Figure 5A). These results are similar to Sawamoto et al. (2003) and Reay et al. (2004), who both found little or no relation between N₂O-N and NO₃⁻-N concentration in agricultural drainage water. In contrast others have observed a positive relationship between dissolved N₂O-N and NO₃⁻-N in agricultural drainage waters (Hack and

Kaupenjohann, 2002; Minami and Oshawa, 1990; Reay et al., 2003). Dowdell et al. (1979) and Hack and Kaupenjohann (2002) have noted NO₃⁻ concentrations are typically one to three orders of magnitude higher than N₂O concentrations. In our study N₂O and NO₃⁻ concentration were one to two orders of magnitude greater than dissolved N₂O concentrations, depending on season (Table 3A).

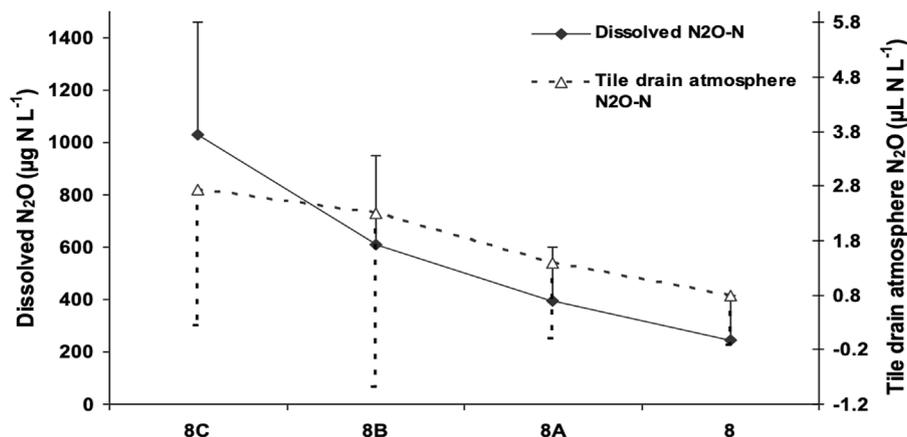


FIGURE 4
Dissolved N₂O and tile drain atmosphere N₂O concentration at three locations along a tile drain in plot# 8 (8A, 8B, 8C) and at the tile drain outlet of plot# 8 (8). Error bars represent the standard deviation of the 12 water and air samples taken during different days (between November 14 to 19 Dec 2003 and April 1 to 6 May 2004). Error bars are presented in opposite directions for the clarity of the figure.

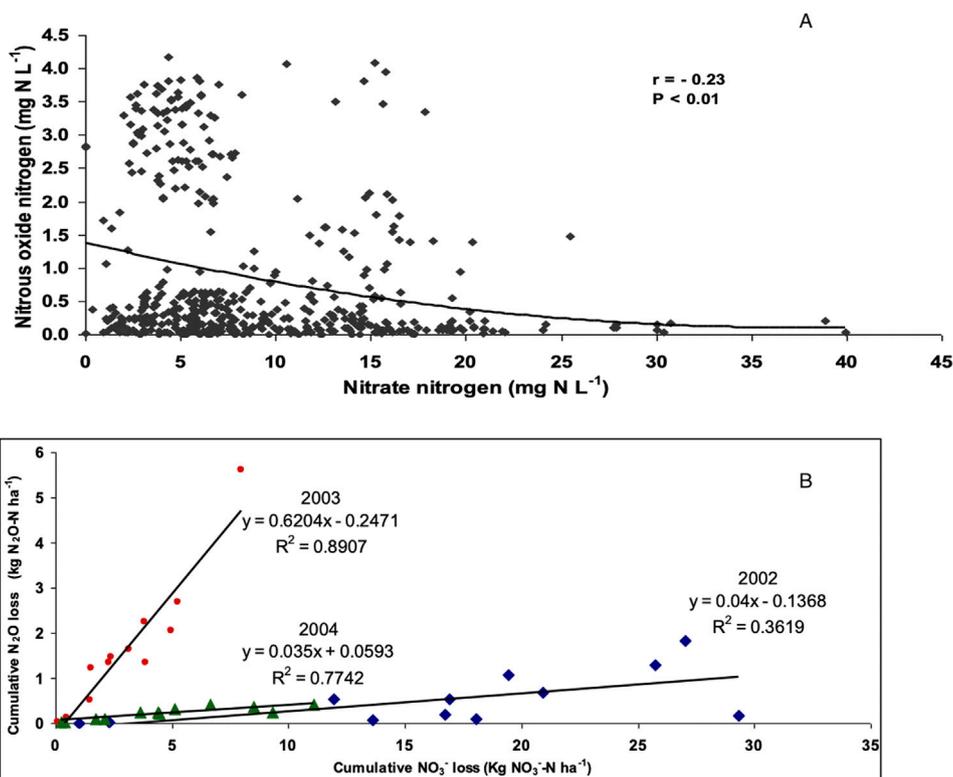


FIGURE 5
(A) Daily N₂O-N loss vs. NO₃⁻-N loss in tile drainage water from a potato - barley - red clover rotation for 2002, 2003 and 2004, and (B) Cumulative N₂O-N loss vs. cumulative NO₃⁻-N loss for 2002, 2003 and 2004.

The strength of the relationship between dissolved N₂O and NO₃⁻-N was much greater when cumulative losses were examined. A significant ($p < 0.001$) positive relationship existed between cumulative annual N₂O-N and NO₃⁻-N losses (Figure 5B). The slope of this relationship was significantly greater in 2003 than in the other 2 years of study. The relationships within individual years accounted for a much

greater percentage of the total variation ranging from 36% to 89% of the total variation (Figure 5B).

The difference in the relationship between N₂O and NO₃⁻ concentrations in drainage water for individual sampling events (Figure 5A) vs. cumulative annual N losses (Figure 5B) provides an interesting contrast. The rather weak negative relationship in the weekly samples indicates that the processes resulting in these losses are

TABLE 4 Mean and range of values of N₂O concentrations in tile drainage water, stream water and spring water.

Water system	N ₂ O concentration ($\mu\text{g N L}^{-1}$)
Tile drainage water	372 (4.0–778.2) [†]
Stream water	18.23 (8.4–33.1) [‡]
Spring water	36.08 (9.3–67.1) [‡]

[†]Mean (range of value) for 12 samples.

[‡]Mean (range of value) for 3 samples.

independent at this time scale or that these processes are occurring at different location and are thus exposed to different environmental conditions. However, the positive relationship between N₂O and NO₃⁻ for each year of observation indicates that over this longer timeframe both processes appear to be responding to similar controls. In essence the leakiness of the system with respect to nitrogen is similarly expressed in terms of both of NO₃⁻ and N₂O losses. These losses simply are not observed at the same time at the end of the tile drain. These results are similar to Quesnel et al. (2019), who observed a significant positive relationship between mean dissolved N₂O-N and NO₃⁻-N concentrations during the growing season, however they found that the relationship became weaker in the non growing season. They attributed this to the greater complexity of N₂O production and transport in the non-growing season. Overall, our results indicate that a positive relationship exists between NO₃⁻ and N₂O, but that these processes are spatially and/or temporally separate.

IPCC coefficient and dissolved N₂O emissions

Currently the IPCC coefficients for N₂O emissions from agricultural drainage is calculated as a fixed fraction of the NO₃⁻-N estimated to be lost from the soil profile ($EF_5 = 0.0075 \text{ kg N}_2\text{O-N kg}^{-1} \text{ N leached}$). Here, on an annual basis the ratio of kg N₂O-N lost per kg NO₃⁻-N leached ranged from 0.035 to 0.62 kg N₂O-N kg⁻¹ N leached. These values are significantly higher than the IPCC coefficient for this site in these years and therefore would underestimate the actual N₂O emissions at this site.

Dissolved N₂O concentration in different water systems

To determine whether the water being discharged from the agricultural fields was enriched relative to the adjacent streams, water samples were collected on 28 May 2004 from subsurface tile drainage, stream and spring water. These samples displayed significant variation in dissolved N₂O concentrations (Table 4). Dissolved N₂O concentrations in stream and spring water were 8.4–33.1 $\mu\text{g N L}^{-1}$ (mean 18.2 $\mu\text{g N L}^{-1}$) and 9.0–67.1 $\mu\text{g N L}^{-1}$ (mean 36.1 $\mu\text{g N L}^{-1}$), respectively, while N₂O concentrations of water being discharged from the tile drainage system ranged from 4.0 to 778 $\mu\text{g N L}^{-1}$ (mean 372 $\mu\text{g N L}^{-1}$). This indicates that the tile drainage water was enriched in dissolved N₂O prior to being released into the waterway. Low N₂O concentrations in stream water were likely reflect the degassing of dissolved N₂O degassing to the atmosphere following discharge, as was observed over the length of the tile line. This suggests that large amounts of indirect N₂O loss can occur after the tile drain water enters the stream.

Conclusions

Annual tile drainage losses of dissolved N₂O from this site were substantive, approaching that of surface N₂O emissions. The nature of the winter period had important implications for the magnitude and timing of dissolved N₂O emissions. The presence of a continuous period of sub-zero soil and air temperatures resulted in greater dissolved N₂O emissions than occurred under more moderate winter temperatures. There was significant spatial and temporal complexity in the nature of dissolved N₂O emissions that complicated their characterization at a field scale.

Nitrogen application treatments, source and timing had no effect on the magnitude of dissolved N₂O loss. On an event basis, dissolved N₂O emissions were not correlated with NO₃⁻ concentrations. Dissolved N₂O and NO₃⁻ losses appeared to be occurring at different times, however, examined on a seasonal basis, plots with high NO₃⁻ losses also exhibited high dissolved N₂O loss.

Subsurface tile length influenced N₂O discharge from subsurface tile drainage water, indicating the presence of a sink within the system. Consequently, our results likely underestimate N₂O produced at the field level. More study is needed to understand the magnitude and fate of these N₂O losses.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Author contributions

DB: Conceptualization, Formal Analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Supervision, Writing–original draft, Writing–review and editing. HW: Data curation, Formal Analysis, Visualization, Writing–original draft, Writing–review and editing. JM: Conceptualization, Investigation, Methodology, Supervision, Writing–original draft, Writing–review and editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fenvs.2024.1479754/full#supplementary-material>

References

- Aulakh, M. S., Rennie, D. A., and Paul, E. A. (1984). Gaseous nitrogen losses from soils under zero-till as compared with conventional-till management systems. *J. Environ. Qual.* 13 (1), 130–136. doi:10.2134/jeq1984.00472425001300010024x
- Baggs, E. M. (2011). Soil microbial sources of nitrous oxide: recent advances in knowledge, emerging challenges and future direction. *Curr. Opin. Environ. Sustain.* 3 (5), 321–327. doi:10.1016/j.cousust.2011.08.011
- Bruun, J., Hoffmann, C. C., and Kjaergaard, C. (2017). Convective transport of dissolved gases determines the fate of the greenhouse gases produced in reactive drainage filters. *Ecol. Eng.* 98, 1–10. doi:10.1016/j.ecoleng.2016.10.027
- Burton, D. L., and Beauchamp, E. G. (1985). Denitrification rate relationships with soil parameters in the field. *Commun. Soil Sci. Plant Anal.* 16, 539–549. doi:10.1080/00103628509367626
- Burton, D. L., and Beauchamp, E. G. (1994). Profile nitrous oxide and carbon dioxide concentrations in a soil subject to freezing. *Soil Sci. Soc. Am. J.* 58, 115–122. doi:10.2136/sssaj1994.03615995005800010016x
- Campbell, A. J., Van Lunen, T. A., MacLeod, J. A., Hurnik, D., and Linkletter, G. (2003). Design and performance of a swine finishing barn for production and manure research. *Can. Biosyst. Eng.* 45, 5.1–5.6.
- Chapuis-Lardy, L. Y. D. I. E., Wrage, N., Metay, A., Chotte, J. L., and Bernoux, M. (2007). Soils, a sink for N₂O? A review. *Glob. Change Biol.* 13 (1), 1–17. doi:10.1111/j.1365-2486.2006.01280.x
- Choudhary, M. A., Akramkhanov, A., and Saggat, S. (2002). Nitrous oxide emissions from a New Zealand cropped soil: tillage effects, spatial and seasonal variability. *Agric. Ecosyst. Environ.* 93, 33–43. doi:10.1016/s0167-8809(02)00005-1
- DelVecchia, A. G., Rhea, S., Aho, K. S., Stanley, E. H., Hotchkiss, E. R., Carter, A., et al. (2023). Variability and drivers of CO₂, CH₄, and N₂O concentrations in streams across the United States. *Limnol. & Oceanogr.* 68 (2), 394–408. doi:10.1002/lno.12281
- Dowdell, R. J., Burford, J. R., and Crees, R. (1979). Losses of nitrous oxide dissolved in drainage water from agricultural land. *Nature* 278, 342–343. doi:10.1038/278342a0
- Drury, C. F., McKenney, D. J., Findlay, W. L., and Gaynor, J. D. (1993). Influence of tillage on nitrate loss in surface runoff and tile drainage. *Soil Sci. Soc. Am. J.* 57, 797–802. doi:10.2136/sssaj1993.03615995005700030028x
- ECCC (Environment and Climate Change Canada) (2022). *National inventory Report 1990–2020: Greenhouse gas sources and sinks in Canada*. Canada: Ottawa.
- Flessa, H., Ruser, R., Schilling, R., Löffel, N., Munch, J. C., Kaiser, E. A., et al. (2002). N₂O and CH₄ fluxes in potato fields: automated measurement, management effects and temporal variation. *Geoder* 105, 307–325. doi:10.1016/s0016-7061(01)00110-0
- Gauder, M., Butterbach-Bahl, K., Graeff-Hnninger, S., Claupein, W., and Wiegel, R. (2012). *Soil-derived trace gas fluxes from different energy crops - results from a field experiment in southwest Germany*. Great Britain: Blackwell Publishing Ltd.
- Gillam, K. M., Zebbarth, B. J., and Burton, D. L. (2007). Nitrous oxide emissions from denitrification and the partitioning of gaseous losses as affected by nitrate and carbon addition and soil aeration. *Can. J. Soil Sci.* 88, 133–143. (in press). doi:10.4141/cjss06005
- Goodroad, L. L., and Keeney, D. R. (1984). Nitrous oxide emissions from soils during thawing. *Can. J. Soil Sci.* 64, 187–194. doi:10.4141/cjss84-020
- Government of Canada (2019). Canadian climate normals 1981–2010 station data. Available at: https://climate.weather.gc.ca/climate_normals/results_1981_2010_e.html?searchType=stnProv&lstProvince=PE&txtCentralLatMin=0&txtCentralLatSec=0&txtCentralLongMin=0&txtCentralLongSec=0&stnID=6526&dispBack=0 (Accessed December 1, 2023).
- Granli, T., and Bockman, O. C. (1994). Nitrous oxide from agriculture. *Norw. J. Agric. Sci. Suppl.* 12, 128.
- Hack, J., and Kaupenjohann, M. (2002). “N₂O discharge with drain water from agricultural soils of the upper Neckar region in southern Germany,” in *Proc. Third Int. Symp. Non-CO₂ Greenhouse gases: scientific understanding, control options and policy aspects, maastricht, Netherlands. 21–23 january 2002*. Editors J. Van Ham, A. P. M. Baede, R. Guicherit, and J. G. F. M. Williams-Jacobse (Millipress, Rotterdam, Netherlands), 185–190.
- Hama-Aziz, Z. Q., Hiscock, K. M., and Cooper, R. J. (2016). Indirect nitrous oxide emission factors for agricultural field drains and headwater streams. *Environ. Sci. & Technol.* 51 (1), 301–307. doi:10.1021/acs.est.6b05094
- Hama-Aziz, Z. Q., Hiscock, K. M., and Cooper, R. J. (2017). Dissolved nitrous oxide (N₂O) dynamics in agricultural field drains and headwater streams in an intensive arable catchment. *Hydrol. Process.* 31 (6), 1371–1381. doi:10.1002/hyp.11111
- Hasegawa, K., Hanaki, K., Matsuo, T., and Hidaka, S. (2000). Nitrous oxide from the agricultural water system contaminated with high nitrogen. *Chemosphere Glob. Change Sci.* 2, 335–345. doi:10.1016/s1465-9972(00)00009-x
- Heincke, M., and Kaupenjohann, M. (1999). Effects of soil solution on the dynamics of N₂O emissions: a review. *Nutr. Cycl. Agroecosys.* 55, 133–157. doi:10.1023/a:1009842011599
- Hutchinson, G., and Livingston, G. (1993). “Use of chamber systems to measure trace gas fluxes. Pages 63–78,” in *Agricultural ecosystem effects on trace gases and global climate change*. Editors L. A. Harper, A. R. Mosier, J. M. Duxbury, and D. E. Rolston (Madison, WI: ASA, CSSA, SSSA). Special Publication No 55.
- IPCC (2019). *in 2019 refinement to the 2006 IPCC guidelines for national Greenhouse gas inventories. Volume 4 agriculture, forestry and other land use*. Editors Calvo Buendia, E., Tanabe, K., Kranjc, A., Baasansuren, J., Fukuda, M., Ngarize, S., et al. Published: IPCC, Switzerland.
- IPCC (2021). *in Climate change 2021: the physical science basis. Contribution of working group I to the sixth assessment Report of the intergovernmental panel on climate change*. Editors Masson-Delmotte, V., Zhai, P., Pirani, A., Connors, S. L., Péan, C., Berger, S., et al. (Cambridge, United Kingdom and New York, NY, USA: Cambridge University Press). In press. doi:10.1017/9781009157896
- IPCC (2022). *in Climate change 2022: mitigation of climate change. Contribution of working group III to the sixth assessment Report of the intergovernmental panel on climate change*. Editors Shukla, P. R., Skea, J., Slade, R., Al Khourdajie, A., van Diemen, R., McCollum, D., et al. (Cambridge, UK and New York, NY, USA: Cambridge University Press). doi:10.1017/9781009157926
- IPCC (2023). “Climate change 2023: synthesis Report,” in *Contribution of working groups I, II and III to the sixth assessment Report of the intergovernmental panel on climate change core writing team*. Editors Lee, H., and Romero, J. (Geneva, Switzerland: IPCC), 35–115. doi:10.59327/IPCC/AR6-9789291691647
- Janzen, H. H., Desjardins, R. L., Asselin, J. M. R., and Grace, B. (1999). *The health of our air: toward a sustainable agriculture in Canada*. Ottawa, Canada: Agriculture and Agri Food Canada Publication# 1981/E. Available at: <https://publications.gc.ca/site/eng/9.686880/publication.html>
- Jiang, Y., Zebbarth, B., and Love, J. (2011). Long-term simulations of nitrate leaching from potato production systems in Prince Edward Island, Canada. *Nutrient Cycl. Agroecosyst. Former. Fertilizer Res.* 91 (3), 307–325. doi:10.1007/s10705-011-9463-z
- Jiang, Y., Zebbarth, B. J., Somers, G. H., MacLeod, J. A., and Savard, M. M. (2012). Nitrate leaching from potato production in eastern Canada. *Sustain. Potato Prod. Glob. Case Stud.*, 233–250. doi:10.1007/978-94-007-4104-1_13
- Kavdir, Y., Hellebrand, H. J., and Kern, J. (2008). Seasonal variations of nitrous oxide emission in relation to nitrogen fertilization and energy crop types in sandy soil. *Soil & Tillage Res.* 98, 175–186. doi:10.1016/j.still.2007.11.002
- Klavdiko, E. J., Van Scoyoc, G. E., Monke, E. J., Oates, K. M., and Pask, W. (1991). Pesticide and nutrient movement into subsurface tile drains on a silt loam soil in Indiana. *J. Environ. Qual.* 20, 264–270. doi:10.2134/jeq1991.00472425002000010043x
- Maag, M., and Vinther, F. (1996). Nitrous oxide emission by nitrification and denitrification in different soil types and at different soil moisture contents and temperatures. *Appl. Soil Ecol.* 4, 5–14. doi:10.1016/0929-1393(96)00106-0

- MacDougall, J. I., Veer, C., and Wilson, F. (1988). *Soils of Prince Edward Island: Prince Edward Island soil survey. Research Branch*. Agriculture Canada.
- Milburn, P., Macleod, J. A., and Sanderson, B. (1997). Control of nitrate leaching from early harvested potatoes on Prince Edward Island. *Can. Agri. Eng.* 39, 263–271.
- Minami, K., and Fukushi, S. (1984). Methods for measurement N₂O flux from water surface and N₂O dissolved in water from agricultural land. *Soil Sci. Plant Nutr.* 30, 495–502. doi:10.1080/00380768.1984.10434716
- Minami, K., and Oshawa, A. (1990). “Emission of nitrous oxide dissolved in drainage water from agricultural land,” in *Soils and the Greenhouse effect*. Editor A. F. Bouman (Chichester, UK: John Wiley & Sons), 503–509.
- Nangia, V., Gowda, P. H., Mulla, D. J., and Sands, G. R. (2010). Modeling impacts of tile drain spacing and depth on nitrate-nitrogen losses. *Artif. Drain.* 9 (1), 61–72. doi:10.2136/vzj2008.0158
- Nevison, C. (2000). Review of the IPCC methodology for estimating nitrous oxide emissions associated with agricultural leaching and runoff. *Chemosphere Glob. Change Sci.* 2, 493–500. doi:10.1016/s1465-9972(00)00013-1
- Quesnel, J., VanderZaag, A., Crolla, A., Kinsley, C., Gregorich, E., and Wagner-Riddle, C. (2019). Surface and subsurface N₂O losses from dairy cropping systems. *Nutr. Cycl. Agroecosyst* 114, 277–293. doi:10.1007/s10705-019-10004-5
- Randall, G. W., Huggins, D. R., Russelle, M. P., Fuchs, D. J., Nelson, W. W., and Anderson, J. L. (1997). Nitrate losses through subsurface tile drainage in conservation reserve program, alfalfa and row crop systems. *J. Environ. Qual.* 26, 1240–1247. doi:10.2134/jeq1997.00472425002600050007x
- Reay, D., Edwards, A., and Smith, K. (2004). Nitrous oxide in agricultural drainage waters following field fertilisation. *Water Air Soil Pollut. Focus* 4, 437–451. doi:10.1023/b:wafo.0000028370.68472.d2
- Reay, D. S., Edwards, A. C., and Smith, K. A. (2009). Importance of indirect nitrous oxide emissions at the field, farm and catchment scale. *Agric. Ecosyst. & Environ.* 133 (3/4), 163–169. doi:10.1016/j.agee.2009.04.019
- Reay, S. D., Smith, K. A., and Edwards, C. A. (2003). Nitrous oxide emission from agricultural drainage waters. *Glob. Change Biol.* 9, 195–203. doi:10.1046/j.1365-2486.2003.00584.x
- Risk, N., Snider, D., and Wagner-Riddle, C. (2013). Mechanisms leading to enhanced soil nitrous oxide fluxes induced by freeze-thaw cycles. *Can. J. Soil Sci.* 93 (4), 401–414. doi:10.4141/cjss2012-071
- Risk, N., Wagner-Riddle, C., Furon, A., Warland, J., and Blodau, C. (2014). Comparison of simultaneous soil profile N₂O concentration and surface N₂O flux measurements overwinter and at spring thaw in an agricultural soil. *Soil Sci. Soc. Am. J.* 78 (1), 180–193. doi:10.2136/sssaj2013.06.0221
- Rochette, P., and McGinn, S. M. (2008). N₂O emissions from agricultural soils in Canada (preface) *can. J. Soil Sci.* 88 (2), 131–132. doi:10.4141/cjss07301
- Roper, J. (2008). *Influence of tillage practices on surface and subsurface nitrogen emissions from agricultural soils*. Halifax, NS: Dalhousie University. MSc thesis.
- Roper, J., Burton, D., Madani, A., and Stratton, G. (2013). A simple method for quantifying dissolved nitrous oxide in tile drainage water. *Can. J. Soil Sci.* 93 (1), 59–64. doi:10.4141/cjss2012-021
- Sawamoto, T., Kusa, K., Hu, R., and Hatano, R. (2002). Dissolved N₂O, CH₄ and CO₂ in pipe drainage, seepage and stream water in a livestock farm in Hokkaido, Japan. *Soil Sci. Plant Nutr.* 48, 433–439. doi:10.1080/00380768.2002.10409222
- Sawamoto, T., Kusa, K., Hu, R., and Hatano, R. (2003). Dissolved N₂O, CH₄ and CO₂ emissions from subsurface drainage in a structured clay soil cultivated with onion in central Hokkaido, Japan. *Soil Sci. Plant Nutr.* 49, 31–38. doi:10.1080/00380768.2003.10409976
- Shakoor, A., Shakoor, S., Rehman, A., Ashraf, F., Abdullah, M., Shahzad, S. M., et al. (2021). Effect of animal manure, crop type, climate zone, and soil attributes on greenhouse gas emissions from agricultural soils-A global meta-analysis. *J. Clean. Prod.* 278, 1–13. doi:10.1016/j.jclepro.2020.124019
- Syakila, A., and Kroeze, C. (2011). The global nitrous oxide budget revisited. *Greenh. Gas. Meas. Manage.* 1, 17–26. doi:10.3763/ghgmm.2010.0007
- Tian, L., Cai, Y., and Akiyama, H. (2019). A review of indirect N₂O emission factors from agricultural nitrogen leaching and runoff to update of the default IPCC values. *Environ. Pollut.* 245, 300–306. doi:10.1016/j.envpol.2018.11.016
- Valayamkunnath, P., Gochis, D. J., Chen, F., Barlage, M., and Franz, K. J. (2022). Modeling the hydrologic influence of subsurface tile drainage using the national water model. *Water Resour. Res.* 58 (4). doi:10.1029/2021WR031242
- van Es, H. M., Sogbedji, J. M., and Schindelbeck, R. R. (2006). Effect of manure application timing, crop, and soil type on nitrate leaching. *J. Environ. Qual.* 35 (2), 670–679. doi:10.2134/jeq2005.0143
- Venterea, R., Bijesh, M., and Dolan, M. (2011). Fertilizer source and tillage effects on yield-scaled nitrous oxide emissions in a corn cropping system. *J. Environ. Qual.* 40 (5), 1521–1531. doi:10.2134/jeq2011.0039
- Wagner-Riddle, C., Congreves, K., Abalos, D., Berg, A., Brown, S., Ambadan, J., et al. (2017). Globally important nitrous oxide emissions from croplands induced by freeze-thaw cycles. *Nat. Geosci.* 10, 279–283. doi:10.1038/ngeo2907
- World Meteorological Organization (2014). *Scientific assessment of ozone depletion: 2014 global ozone research and monitoring project-rep*, 56. Geneva, Switzerland: World Meteorological Organization.
- Xia, F., Mei, K., Zhang, C., Xu, Y., Dahlgren, R. A., and Zhang, M. (2020). Response of N₂O emission to manure application in field trials of agricultural soils across the globe. *Sci. Total Environ.* 733, 139390. doi:10.1016/j.scitotenv.2020.139390