



Regulatory Network and Prognostic Effect Investigation of PIP4K2A in Leukemia and Solid Cancers

Shouyue Zhang^{1,2†}, Zhaozhi Li^{3†}, Xinyu Yan^{4†}, Li Bao³, Yun Deng², Feier Zeng², Peiqi Wang⁴, Jianhui Zhu⁴, Dandan Yin², Fei Liao², Xueyan Zhou², Duyu Zhang², Xuyang Xia², Hong Wang⁵, Xue Yang⁵, Wanhua Zhang⁶, Hu Gao⁷, Wei Zhang⁸, Li Yang², Qianqian Hou², Heng Xu², Yan Zhang^{1*}, Yang Shu^{2*} and Yuelan Wang^{1,2*}

OPEN ACCESS

Edited by:

Luis Abel Quiñones, Universidad de Chile, Chile

Reviewed by:

Fausto Leonardo Zaruma-Torres, University of Cuenca, Ecuador Enrique Teran, Universidad San Francisco de Quito, Ecuador

*Correspondence:

Yan Zhang zhangyan915@126.com Yang Shu shuyang1986@gmail.com Yuelan Wang wangyuelan1982@sina.com †These authors have contributed equally to this work

Specialty section:

This article was submitted to Pharmacogenetics and Pharmacogenomics, a section of the journal Frontiers in Genetics

Received: 31 August 2018 Accepted: 21 December 2018 Published: 15 January 2019

Citation:

Zhang S, Li Z, Yan X, Bao L, Deng Y, Zeng F, Wang P, Zhu J, Yin D, Liao F, Zhou X, Zhang D, Xia X, Wang H, Yang X, Zhang W, Gao H, Zhang W, Yang L, Hou Q, Xu H, Zhang Y, Shu Y and Wang Y (2019) Regulatory Network and Prognostic Effect Investigation of PIP4K2A in Leukemia and Solid Cancers. Front. Genet. 9:721. doi: 10.3389/fgene.2018.00721 ¹ Department of Thoracic Oncology, Cancer Center, State Key Laboratory of Biotherapy, West China Hospital, Sichuan University, Chengdu, China, ² Department of Laboratory Medicine, Precision Medicine Center, State Key Laboratory of Biotherapy and Precision Medicine Key Laboratory of Sichuan Province, West China Hospital, Sichuan University and Collaborative Innovation Center, Chengdu, China, ³ Key Laboratory of Bio-Resources and Eco-Environment, College of Life Sciences, Sichuan University, Chengdu, China, ⁴ State Key Laboratory of Oral Diseases, West China Hospital of Stomatology, Sichuan University, Chengdu, China, ⁵ Integrated Biomedical Sciences Program, University of Tennessee Health Science Center, Memphis, TN, United States, ⁶ Department of Hematology and Hematology Research Laboratory, Sichuan University, Chengdu, China, ⁷ Department of Emergency, West China Second University Hospital, Sichuan University, Chengdu, China, China, China, China, South University, Chengdu, China, China, South University, Chengdu, China, Suthara College of Emergency, Hunan Key Laboratory of Pharmacogenetics, Xiangya Hospital, Central South University, Changsha, China

Germline variants of PIP4K2A impact susceptibility of acute lymphoblastic leukemia (ALL) through inducing its overexpression. Although limited reports suggested the oncogenic role of PIP4K2A in cancers, regulatory network and prognostic effect of this gene remains poorly understood in tumorigenesis and leukemogenesis. In this study, we conducted genome-wide gene expression association analyses in pediatric B-ALL cohorts to discover expression associated genes and pathways, which is followed by the bioinformatics analyses to investigate the prognostic role of PIP4K2A and its related genes in multiple cancer types. 214 candidates were identified to be significantly associated with PIP4K2A expression in ALL patients, with known cancer-related genes rankings the top (e.g., RAC2, RBL2, and TFDP1). These candidates do not only tend to be clustered in the same types of leukemia, but can also separate the patients into novel molecular subtypes. PIP4K2A is noticed to be frequently overexpressed in multiple other types of leukemia and solid cancers from cancer cohorts including TCGA, and associated with its candidates in subtype-specific and cancer-specific manners. Interestingly, the association status varied in tumors compared to their matched normal tissues. Moreover, PIP4K2A and its related candidates exhibit stageindependent prognostic effects in multiple cancers, mostly with its lower expression significantly associated with longer overall survival (p < 0.05). Our findings reveal the transcriptional regulatory network of PIP4K2A in leukemia, and suggest its potentially important role on molecular subtypes of multiple cancers and subsequent treatment outcomes.

Keywords: acute lymphoblastic leukemia, cancer, TCGA, TFDP1, PIP4K2A

INTRODUCTION

Acute lymphoblastic leukemia (ALL) is the most common pediatric malignancy (Greaves, 2006; Pui and Evans, 2006), induced through multifactorial mechanisms involving both environmental and genetic factors (Buffler et al., 2005). A series of genome-wide association studies (GWASs) have identified several common risk loci for genetic predisposition basis of ALL in several genes (e.g., ARID5B, IKZF1, CEBPE, CDKN2A, PIP4K2A-BMI1, and GATA3) (Papaemmanuil et al., 2009; Trevino et al., 2009; Sherborne et al., 2010; Migliorini et al., 2013; Perez-Andreu et al., 2013; Xu et al., 2013; Perez-Andreu et al., 2015; Xu et al., 2015). As more and more causal variants are considered to impact on gene expression rather than inducing amino acid changes, analyses for expression quantitative trait loci (eQTL) in ALL have also been conducted. For instance, PIP4K2A expression is strongly impacted by the genotypes of its adjacent single nucleotide polymorphisms (SNPs), which have been validated in several independent patient cohorts for ALL susceptibility (Walsh et al., 2013; Liao et al., 2016), suggesting PIP4K2A SNPs may play role on leukemogenesis through inducing its overexpression. Meanwhile, current evidence has also shown a ethnicity-specific and subtype-specific correlation between PIP4K2A SNPs and ALL susceptibility (Migliorini et al., 2013; Walsh et al., 2013; Xu et al., 2013), which is similar as ARID5B SNPs (Xu et al., 2012, 2013). However, how PIP4K2A contributes to leukemogenesis of ALL and its subtypes remains to be poorly understood.

PIP4K2A encodes a major component of phosphatidylinositol 5-phosphate 4-kinase type-II (Rameh et al., 1997). As actively expressed in peripheral blood cells (Divecha et al., 1995), PIP4K2A may facilitate the conversion of phosphorylation of phosphatidylinositol 5-phosphate (PtdIns5P) to generate various phosphoinositide species, which regulate crucial cellular functions including proliferation, survival, glucose uptake and migration (Fiume et al., 2015). A few reports have demonstrated that PIP4K2A may play a critical role on development and cellular activities maintaining. For instance, PIP4K2A expression can influence cell response to oxidative stress by regulating the level of PtdIns5P (Wilcox and Hinchliffe, 2008; Jones et al., 2013). Depletion of PIP4K2A is associated with decreased mTOR signaling, and induces developmental defects in Drosophila and zebrafish (Elouarrat et al., 2013; Gupta et al., 2013). Besides, PIP4K2A is considered to control neuronal KCNQ channels, disorders of which are implicated in schizophrenia (Fedorenko et al., 2008, 2009), and may also involve in insulin signaling by regulating PI3/AKT pathway. Knocking out of PIP4K2B, another component of phosphatidylinositol 5-phosphate 4-kinase type-II, can increase insulin sensitivity and reduce adiposity (Carricaburu et al., 2003; Lamia et al., 2004). However, the role of PIP4K2A on tumorigenesis (including leukemogenesis) has been mentioned in limited reports. Recently, a cell-line based experiment indicates that knockdown of PIP4K2A in acute myeloid leukemia (AML) results in accumulation of the cyclin-dependent kinase inhibitors (i.e., CDKN1A and CDKN1B), G1 cell cycle arrest and apoptosis (Jude et al., 2015). Nevertheless, PIP4K2A knockdown in normal hematopoietic stem cells and progenitor cells doesn't impair cell growth. Since knockdown of *PIP4K2A* only exerts inhibitory effects on tumor cells, this selective effect implies that *PIP4K2A* might be a candidate target for cancer treatment. In fact, *PIP4K2A* is noted to be up-regulated in breast tumor tissue and high expression of *PIP4K2A* is correlated with loss of p53. Knockdown of both *PIP4K2A* and *PIP4K2B* in a p53-deficient breast cell line dramatically abrogates cell proliferation (Emerling et al., 2013). This could be explained by impaired glucose metabolism and consequently increased reactive oxygen species (ROS), which leads to senescence. Also, *PIP4K2A* is considered to impact on sensitivity of some cancer drugs (e.g., paclitaxel), thus may be associated with overall survival of the treated cancer patients (Niu et al., 2012).

Single nucleotide polymorphisms around *PIP4K2A* are associated with ALL susceptibility through impacting expression level of *PIP4K2A*, and accumulating evidence indicates the potential role of *PIP4K2A* on cancer cells, however, it is poorly understood that how *PIP4K2A* impact on tumorigenesis (including leukemogenesis). In this study, we conducted array based genome-wide gene expression association analyses to investigate the potential regulatory network of *PIP4K2A* in leukemia, which is proved to be efficient in our previous work (Hou et al., 2017). Besides, we will also systematically examine the *PIP4K2A* status in multiple cancers and evaluate its prognostic effect.

RESULTS

Screening of PIP4K2A-Related Genes in B-ALL and Their Association Status in B-ALL Subtypes

As the expression of PIP4K2A was considered to impact on leukemogenesis, we thus downloaded array-based expression data of eight independent pediatric B-ALL patient cohorts from public resource (Supplementary Table S1), and evaluated the expression correlation of PIP4K2A with the rest of other genes using a linear regression model. A series of filter criteria (e.g., *p*-value cutoff, r^2 , and correlation direction) were applied to determine more reliable PIP4K2A-related candidates for regulation network construction (Supplementary Figure S1). Altogether, 234 probes in 214 genes were significantly correlated to *PIP4K2A* expression with $p \le 0.05$ in all patient cohorts in the same direction (Supplementary Figure S1 and Supplementary **Table S2**). Through a stricter filter (i.e., $p \le 2 \times 10^{-6}$ and association $r^2 \ge 0.1$ in at least five cohorts), 23 genes were eventually identified as the strongest candidates (Supplementary Table S3).

Since *PIP4K2A* is a major component phosphate kinase, we sought to reveal its regulatory network by focusing on 10 transcription factors we screened out (**Table 1**). For instance, *RBL2* is a member of retinoblastoma-susceptibility (RB) anti-oncogene family, which regulates cell cycle through inhibiting E2F-mediated transcription. Dysfunction of this gene is commonly involved in evolution of ALL (Ahuja et al., 1991; Stock et al., 2000; Schmitz et al., 2005), thus indicating a

Gene GSE7440 (<i>n</i> = 99)		202098_s_at	202449_s_at	204529_s_at	205038_at	209107_x_at	209930_s_at	210249_s_at	212330_at	212331_at	216241_s_at	43544_at
GSE7440 (n = 99)		PRMT2	RXRA	тох	IKZF1	NCOA1	NFE2	NCOA1	TFDP1	RBL2	TCEA1	MED16
	p-Value	7.49×10^{-10}	7.10×10^{-13}	8.58×10^{-7}	4.89×10^{-5}	1.69×10^{-10}	2.64×10^{-9}	2.93×10^{-13}	2.94×10^{-12}	8.99×10^{-10}	8.56×10^{-8}	3.64×10^{-11}
	Coefficient	0.64	0.61	0.33	0.38	0.84	0.38	0.92	0.51	0.58	0.65	0.74
	Se	0.09	0.07	0.06	0.09	0.12	0.06	0.11	0.06	0.09	0.11	0.10
	Adjusted r ²	0.32	0.41	0.21	0.15	0.34	0.30	0.42	0.39	0.32	0.25	0.36
GSE10255 (<i>n</i> = 161)	p-Value	0.01	0.01	7.56×10^{-4}	0.03	4.10×10^{-3}	4.80×10^{-15}	1.84×10^{-3}	1.35×10^{-3}	4.50×10^{-3}	3.65×10^{-3}	9.86×10^{-9}
	Coefficient	0.40	0.20	0.16	0.26	0.57	0.26	0.51	0.15	0.37	0.49	0.72
	Se	0.15	0.08	0.05	0.12	0.20	0.03	0.16	0.05	0.13	0.17	0.12
	Adjusted r ²	0.04	0.03	0.06	0.02	0.04	0.32	0.05	0.06	0.04	0.05	0.18
GSE10792 (n = 160)	p-Value	5.74×10^{-7}	0.01	0.02	0.03	0.02	1.67×10^{-3}	0.04	8.94×10^{-7}	6.12×10^{-4}	2.96×10^{-3}	5.79×10^{-4}
	Coefficient	0.53	0.22	0.16	0.33	0.48	0.24	0.44	0.29	0.64	0.84	0.51
	Se	0.10	0.08	0.07	0.14	0.21	0.07	0.21	0.06	0.18	0.28	0.14
	Adjusted r ²	0.26	0.07	0.05	0.05	0.05	0.11	0.04	0.26	0.13	0.10	0.13
GSE11877 (n = 207)	p-Value	4.93×10^{-9}	1.65×10^{-3}	2.19×10^{-3}	1.03×10^{-6}	1.02×10^{-4}	1.60×10^{-5}	1.21×10^{-4}	2.24×10^{-3}	8.65×10^{-13}	1.96×10^{-4}	2.71×10^{-7}
	Coefficient	0.39	0.17	0.17	0.33	0.49	0.15	0.46	0.20	0.59	0.43	0.34
	Se	0.06	0.05	0.05	0.07	0.12	0.03	0.12	0.06	0.08	0.11	0.06
	Adjusted r ²	0.15	0.04	0.04	0.11	0.07	0.08	0.07	0.04	0.22	0.06	0.12
GSE13351 (n = 107)	p-Value	4.53×10^{-9}	3.81×10^{-4}	3.94×10^{-3}	1.02×10^{-3}	1.80×10^{-5}	1.89×10^{-6}	3.54×10^{-5}	1.72×10^{-3}	3.46×10^{-12}	4.43×10^{-5}	9.53×10^{-6}
	Coefficient	0.58	0.24	0.19	0.31	0.91	0.28	0.82	0.20	0.67	0.58	0.45
	Se	0.09	0.07	0.07	0.09	0.20	0.06	0.19	0.06	0.08	0.14	0.10
	Adjusted r ²	0.31	0.12	0.08	0.10	0.18	0.22	0.16	0.09	0.41	0.16	0.19
GSE13425 (<i>n</i> = 190)	p-Value	9.14×10^{-5}	1.01×10^{-6}	5.30×10^{-3}	1.18×10^{-3}	2.07×10^{-4}	2.99×10^{-3}	3.79×10^{-3}	5.78×10^{-3}	1.32×10^{-6}	3.31×10^{-4}	1.47×10^{-3}
	Coefficient	0.36	0.39	0.25	0.31	0.66	0.18	0.52	0.20	0.54	0.54	0.33
	Se	0.09	0.08	0.09	0.09	0.17	0.06	0.18	0.07	0.11	0.15	0.10
	Adjusted r ²	0.09	0.14	0.04	0.06	0.08	0.05	0.05	0.04	0.14	0.08	0.06
GSE33315 (<i>n</i> = 575)	p-Value	4.32×10^{-17}	9.08×10^{-5}	3.21×10^{-3}	1.88×10^{-9}	1.38×10^{-12}	1.32×10^{-26}	8.79×10^{-14}	3.64×10^{-16}	1.81×10^{-23}	8.81×10^{-11}	5.83×10^{-21}
	Coefficient	0.56	0.16	0.11	0.34	0.75	0.22	0.67	0.24	0.68	0.57	0.60
	Se	0.06	0.04	0.04	0.05	0.10	0.02	0.09	0.03	0.06	0.09	0.06
	Adjusted r ²	0.13	0.03	0.02	0.07	0.10	0.21	0.11	0.13	0.19	0.08	0.17
GSE635 (n = 173)	p-Value	2.01×10^{-6}	1.38×10^{-4}	5.41×10^{-7}	0.02	3.65×10^{-3}	1.23×10^{-3}	2.97×10^{-3}	2.86×10^{-3}	3.44×10^{-5}	6.07×10^{-5}	1.01×10^{-3}
	Coefficient	0.41	0.28	0.29	0.20	0.51	0.14	0.49	0.18	0.44	0.64	0.30
	Se	0.08	0.07	0.05	0.09	0.17	0.04	0.16	0.06	0.10	0.16	0.09
	Adjusted r ²	0.12	0.08	0.13	0.02	0.04	0.05	0.04	0.05	0.09	0.08	0.06

January 2019 | Volume 9 | Article 721

potentially significant role of *RBL2* on B-ALL development. Pathway analysis was further conducted using DAVID Functional Annotation Tools, and three annotation clusters of gene sets exhibited significance with at least one term having Bonferroni *p*-value < 0.05 (**Supplementary Table S4**), including leukocyte migration pathway. Additionally, we illustrated the interactions between candidates by constructing a protein–protein interaction network based on STRING (**Figure 1B**), and identified multiple *PIP4K2A* related proteins, including the top gene of *RAC2*, which is highly expressed in immune organs. Since *RAC2* belongs to the Rho small GTPase subfamily, which can regulate cell movement, proliferation, and survival, involvement of *PIP4K2A* in leukemogenesis may be partially explained by their interactions. In this part, we firstly built the regulatory network of *PIP4K2A* in B-ALL.

Next, the association between *PIP4K2A* and its candidates were investigated in different molecular subtypes of the largest pediatric B-ALL cohort with available subtype information (GSE33315) (**Supplementary Table S5**). To exclude the impact of small sample size of some subtypes, such as BCR-ABL and MLL rearrangement, we mainly focused on four large patient groups, including hyperdiploid, ETV6-RUNX1, TCF3-PBX1 molecular subtypes as well as B-others. Totally 85 out of 214 genes are significantly associated with *PIP4K2A* expression in all four patient groups with the same direction, including seven transcription factors (**Table 1** and **Figure 2**). Although





similar association status was observed in different subtypes in terms of r^2 and coefficients of these transcription factors (e.g., *TFDP1*, *RBL2*, and *NFE2*), subtype-specificity was also noticed. For instance, *NCOA1* is significantly associated with *PIP4K2A* in all four patient groups with varied coefficients in different subtypes, while *TCEA1* is even unrelated to *PIP4K2A* in TCF3-RUNX1 subtypes, indicating regulation network of *PIP4K2A* may vary in subtype-specific manner.

Varied Association Status of PIP4K2A With Candidate Genes in Different Leukemia Types

Next, dataset GSE13204 (3248), which contains a large number of patients with B-ALL, T-ALL, AML, chronic lymphoblastic leukemia (CLL), chronic myeloid leukemia (CML) as well as healthy bone marrow (HBM), was selected for validation and further analyses. Expression of all PIP4K2A-related genes in B-ALL tends to be clustered in terms of leukemia types, illustrating distinct expression patterns of these candidates in different leukemia types and their different roles on leukemogenesis (Figure 1A). Not surprisingly, 99.07% (212/214) of the candidates were significantly associated with PIP4K2A expression in B-ALL of this cohort with the same direction, proving the reliability of their correlations. Importantly, the consistent rate for significant association between PIP4K2A and its candidates declined to 96.26% (206/214), 91.1% (195/214), 87.85% (188/214), 66.36% (142/214) and 49.53% (106/214) in CLL, AML, T-ALL, CML and HBM, respectively, even with

opposite association directions to that in B-ALL for some genes, which account for 0.97% (2/206), 3.08% (6/195), 0% (0/188), 2.11% (3/142) and 4.72% (5/106) of in CLL, AML, T-ALL, CML and HBM, respectively (**Supplementary Table S6**). While the transcription factors are more stable, with higher consistent rates of significant association and no opposite direction (**Figure 3**). Above results implies that the association of PIP4K2A with its candidates discovered in B-ALL may also exists in other leukemia types at different levels. Notably, association status of *PIP4K2A* and its candidates exhibit the lowest consistent rate in health bone marrow in terms of *p*-value as well as r^2 (**Supplementary Table S6**), indicating the dysfunction of PIP4K2A regulation network in leukemia compared to normal tissues.

PIP4K2A Expression Profile and Potential Genes Associated With PIP4K2A Expression in Multiple Cancer Types

Although higher *PIP4K2A* expression may contribute to higher risk of leukemogenesis, normal B cells are unavailable as matched control for further analyses. Therefore, we further investigated the expression profile of *PIP4K2A* and its associated genes in other cancer types with paired tumor/normal tissues from The Cancer Genome Atlas (TCGA) (**Supplementary Figure S6**). With implicated expression information in multiple types of cancers, up to 8% of patients exhibited up-regulated *PIP4K2A* in tumors compared with their matched normal tissues in all available cancer types except a few down-regulations (e.g., KIRC) (**Supplementary Figure S2**), suggesting the potential widespread oncogenic role of *PIP4K2A* in tumorigenesis apart from leukemia.

Next, we downloaded expression data from TCGA to investigate the association of PIP4K2A with its associated transcription factors in tumors and normal separately in each cancer type (Supplementary Tables S7, S8), Among the 23 types of cancers each with at least 100 patients having available followup information, nearly 70% of these transcription factors exhibit consistent association in both tumor and normal tissues, vet with varied coefficients and r^2 , even opposite directions (Figure 4). Interestingly, tumor-specific or control-specific associations were noticed. The candidates exhibited stronger association in normal tissues (e.g., normal tissues of KIRP, KICH, KIRC, ACC, and COAD) than tumors, whereas at least half of genes lost their correlation, or even turn to be negatively associated with PIP4K2A preferentially in normal tissues of OV, TGCT, UCEC, BLCA, and CHOL. For instance, NCOA1 is more correlated to PIP4K2A in normal tissues than that in tumors in terms of r^2 . The correlations of *PIP4K2A* with *TFDP1* tend to exhibit tumor-specific (i.e., significant in 21/23 tumors but 9/20 normal tissues) (Figure 4 and Supplementary Figure S3), while that with

MED16 is tend to be in control-specific manner (i.e., significant in 14/23 tumors but 15/20 normal tissues) (**Figure 4**). Additionally, opposite association directions of tumor vs. normal were also observed, such as *MED16* in THCA (**Figure 4**), indicating the varied regulatory status and role of *PIP4K2A* on tumor vs. normal tissues in different types of cancers.

Impact of PIP4K2A and Its Correlated Gene on Overall Survival Rate

Due to the potentially important role of *PIP4K2A* in tumors, we further explored the possible role of *PIP4K2A* expression on survival rate in different cancer types. We focused on the transcription factors because of their important roles on *PIP4K2A* network described above. Totally, significant correlations were observed in 6 out of 23 cancer types (i.e., BLCA, KIRC, LAML, LIHC, SARC, and STAD) (**Supplementary Figure S4**). In particular, lower expression of *PIP4K2A* in tumors is associated with longer overall survival in all these cancer types except KIRC (**Figure 5**), suggesting the potential prognostic role of *PIP4K2A* overexpression in multiple cancer types, which is





frequently observed as described above. Moreover, we also evaluated the correlation of survival status in these six cancer types with the expression levels of *PIP4K2A* related transcription factors (**Supplementary Table S9**). Strikingly, *TFDP1* exhibits the strongest consistency with *PIP4K2A* in the same direction in five out of six cancer types (i.e., KIRC, LAML, SARC, BLCA, and LIHC) but not STAD (**Supplementary Figure S4** and **Supplementary Table S9**). On the contrary, such consistency is lower for other transcription factors. For non-transcription factors, an opposite direction was even identified, such as *RAC2* (**Supplementary Figure S4** and **Supplementary Table S9**). Hence, *TFDP1* may play an important role on the transcriptional regulatory network of *PIP4K2A* to impact on the survival rate in multiple cancer types.

DISCUSSION

PIP4K2A is largely implicated in basal cellular activities such as proliferation, survival, glucose uptake and migration, and plays an essential role for tumor growth. Due to extensive expression and pleiotropic effect of PIP4K2A in multiple tissues, it is important to unravel the transcriptional regulatory network of PIP4K2A in different cancer types, which is yet time/effectconsuming. Based on the assumption that genes participating in the same regulatory network are associated in terms of expression level, and closer transcriptional factors exhibit more significant correlation, it will be much easier to screen potential PIP4K2Arelated genes by genome-wide gene expression association analyses by using available public microarray datasets. A series of GWASs and subsequent validations have revealed that SNPs in PIP4K2A locus are significantly associated with ALL susceptibility, possibly through impact on PIP4K2A expression, indicating the potential role of PIP4K2A over-expression in leukemogenesis. Therefore, 214 genes were identified to be associated with PIP4K2A expression with the similar pipeline we used previously (Hou et al., 2017). Although association strength varied in different types of leukemia, or even subtypes of B-ALLs, almost all of these candidates can be validated in B-ALL of another big patient cohort, while a large proportion of them can also be validated in other types of leukemia in terms of expression association. These findings strongly indicate the consistency and stability of PIP4K2A network. Since PIP4K2A encodes a kinase, it is desirable to focus on characteristic regulatory network of PIP4K2A in different B-ALL subtypes, respectively, by analyzing larger samples. Additionally, the over-expression of PIP4K2A could be influenced by the chemotherapeutic treatment applied, so we have searched all the datasets to see if the influence of chemotherapeutic treatment existed in these cohorts. Among eight patient cohorts analyzed in Table 1, the patients from GSE7440 and GSE11877 received chemotherapy, while the patients from GSE10255 and GSE13425 were newly diagnosed with ALL without any initial therapy. Other cohorts didn't tell what treatment the patients received. Therefore, we compared the association status of transcription factors with PIP4K2A in abovementioned four cohorts. It turns out that no significant difference in correlation exists between the cohorts who received

chemotherapy and those who did not, indicating that the chemotherapeutic treatment does not significantly influence the association between *PIP4K2A* and candidates, at least in the analyzed cohorts.

Noteworthily, our findings identified some genes that have been reported to be related to leukemogenesis or ALL development. As one of the strongest PIP4K2Arelated candidates, the transcription factor RBL2 belongs to retinoblastoma-susceptibility (RB) anti-oncogene family. The RB gene is an anti-oncogene regulating cell cycle through inhibiting E2F-mediated transcription, which is frequently altered and thus quite commonly involved in evolution of ALL (Ahuja et al., 1991; Stock et al., 2000; Schmitz et al., 2005). RBL2 demonstrates statistically significant and positive correlation with PIP4K2A expression in four relatively larger samples of B-ALL subtypes (i.e., hyperdiploid, ETV6-RUNX1, B-other and TCF3-PBX1 subtype). RBL2 also exhibits significant correlation exclusively in tumor tissues of OV, BLCA, and CHOL, indicating a cancer type-specific correlation between RBL2 and PIP4K2A. Another example is TFDP1, this gene exhibits extraordinarily strong association with PIP4K2A. Same as RBL2, TFDP1 also shows evident association with PIP4K2A in four large samples of B-ALL subtypes as well as all types of leukemia. And in 23 other cancer types, TFDP1 remains the significance in 21 types of cancer. Specifically, in KIRC and PAAD, PIP4K2A expression is prominently affected by TFDP1. Besides, the impact of PIP4K2A and TFDP1 expression on survival status in various cancers demonstrates great consistency, and these two genes are also significantly correlated in these cancers. Above findings reveal quite close interactions between PIP4K2A and TFDP1 as well as the important role of TFDP1 in regulatory network of PIP4K2A. However, no clear binding motif information is available due to the limited studies on TFDP1. Some other candidates are linked to PIP4K2A according to PPI prediction, and we considered it is appropriate to start with these leukemia-related or cancer-related genes in order to reveal the transcriptional regulatory network of PIP4K2A in B-ALL. With the limited database, some of the candidate transcription factors have the predicted binding sites in PIP4K2A promoter or enhancer region, including RXRA (Supplementary Figure S5 and Supplementary Table S10). However, further experimental analyses are largely needed to test whether PIP4K2A is regulated by these transcription factors through direct binding. Unfortunately, we didn't screen out the well-known genes in mTOR pathway or KCNQ channel, probably because PIP4K2A may be involved in these pathways through post-transcriptional regulation or protein interactions, which can't be detected through expression association approaches.

Importantly, the candidate transcription factors are also significant associated with *PIP4K2A* expression in multiple cancer types apart from leukemia. Generally, candidate genes demonstrate much stronger association in normal tissues than tumors. Among 10 transcription factors, some genes exhibit significant associations in all 23 cancer types such as *RBL2* and *IKZF1*, indicating their important role on *PIP4K2A* and ubiquitous interactions with *PIP4K2A* in the regulatory network among different cancers, as well as the reliability of our screening process. Interestingly, *IKZF1* is one of most important transcription factors involved in hematopoietic development, mutations and deletions of this gene are frequently observed in leukemia cells of B-ALL patients. Meanwhile, SNPs in IKZF1 are also significantly associated with ALL susceptibility across multiple ethnicities, probably through impacting on the expression level of IKZF1. Therefore, the expression correlation of IKZF1 with PIP4K2A indicates that these two genes may interact with each other to facilitate leukemogenesis as well as tumorigenesis. Although insignificant in some cancer types, more than 70% of associations between PIP4K2A and the rest of the transcription factors exhibit statically significant, including cancer-specific or normal-specific manner. Moreover, loss or even opposite direction of correlations were observed in quite a few genes between tumor and normal tissues, suggesting tumor-specific dysfunction of PIP4K2A regulatory network may contribute to tumorigenesis.

Due to the lack of survival information of the ALL patients, we can't estimate the impact of SNP genotypes and subsequent SNP-induced expression changes on survival rate. However, we noticed that PIP4K2A overexpression is related to worse treatment outcomes of multiple cancer types of TCGA, except higher PIP4K2A expression related to better survival status in KIRC. Consistently, down-regulation of PIP4K2A in tumors is much more frequently observed in KIRC than any other types of cancer in TCGA (Supplementary Figure S2). Not surprisingly, some of the PIP4K2A-related genes are also related to survival rate in the same direction, among which TFDP1 exhibits the highest consistency, suggesting that TFDP1 may interact with PIP4K2A to impact on both tumorigenesis as well as treatment outcomes. However, the prognostic value of PIP4K2A and its regulatory partners should be evaluated in independent cancer cohorts.

To sum up, we have utilized available public microarray datasets and developed a systematic screening to find expressionbased regulatory network of *PIP4K2A*. Because very limited studies have reported to explore the functional role of *PIP4K2A* on leukemogenesis despite of its significant association with ALL susceptibility as well as overexpression in multiple types of cancers, our findings may shed a new light on epigenetic regulation of *PIP4K2A*, and provide a clue for the further functional studies on this gene in cancer.

MATERIALS AND METHODS

Construction of Regulatory Network of PIP4K2A

Expression level of all genes in B-ALL patients were downloaded from Gene Expression Omnibus [GSE7440 (Bhojwani et al., 2008), GSE11877 (Kang et al., 2010), GSE10792 (Bungaro et al., 2009), GSE13351 (Den Boer et al., 2009), GSE13425 (Holleman et al., 2004), GSE635 (Sorich et al., 2008), GSE10255 (Kirschner-Schwabe et al., 2006), GSE4698 (Zhang et al., 2012), GSE33315 (Haferlach et al., 2010), and GSE13204 (Curtis et al., 2012)]. Associations of PIP4K2A expression with all the rest genes were calculated using linear regression model. Afterward, we used multiple steps to conduct candidate screening and determine the strong candidates (**Supplementary Figure S1**).

All the PIP4K2A-related candidates were imported into the STRING¹ to show the known protein–protein interaction network (Szklarczyk et al., 2015). We used line thickness to indicate the strength of data support, and the nodes disconnected with the main network were hidden.

Functional enrichment was determined using the DAVID functional annotation tool² (Huang da et al., 2009a,b). The functional categories used were GO term related to biological process (BP), cellular component (CC), and molecular function (MF)³. Multiple criteria were applied for pathway analyses of candidates, including *P*-value, FDR, etc. (Huang da et al., 2009b). Various top-related pathways were selected (i.e., lysosome, porphyrin biosynthesis and adhesion, and diapedesis of granulocytes).

With limited information of transcription factors binding motif, we predicted some candidate transcription factors by importing the specific sequence of PIP4K2A in to JASPAR⁴. Some of these transcription factors got high relative scores and had predicted binding sites in PIP4K2A promoter or enhancer regions.

Exploration of Differences Between PIP4K2A Regulatory Networks in Different Subtypes

We divided the patients into multiple groups according to the subtypes of B-ALL and the subtypes of leukemia from the large two cohorts, GSE33315 and GSE13204, respectively. To study the difference of *PIP4K2A* regulatory networks in various subtypes, we calculated the association of *PIP4K2A* with all the rest genes by linear regression model in diverse subtypes. The proportions of *PIP4K2A*-related candidates in all significant related genes were computed.

We selected 133 *PIP4K2A*-related probes by various criteria (i.e., $p \le 2 \times 10^{-6}$ and association $r^2 \ge 0.1$ in at least five cohorts, and $r^2 \ge 0.3$ in at least one cohort). Then, the expression levels of these candidates in GSE13204 were used to generate the heatmap, clustering by the subtypes of leukemia and the subtypes of B-ALL, respectively.

Discovery of Association of PIP4K2A With Its Related Genes in Multiple Cancers and Corresponding Normal Tissues

We compared the expression level of PIP4K2A in various cancer types in COSMIC dataset⁵, and computed the frequency of high-regulated and low-regulated PIP4K2A samples in different status.

⁵http://cancer.sanger.ac.uk/cosmic

¹https://string-db.org/

²http://david.abcc.ncifcrf.gov/

³http://geneontology.org/

⁴http://jaspar.genereg.net/

Some functional *PIP4K2A*-related transcription factors were picked out using the Animal TFDB database $(42)^6$. Gene expression RNAseq [log₂(tpm+0.001)] of the candidates in 23 TCGA cancers (THCA, LIHC, LUAD, STAD, LUSC, OV, PAAD, PRAD, TGCT, UCEC, KICH, KIRC, ACC, BLCA, COAD, DLBC, CHOL, BRCA, THYM, LAML, CESC, PCPG, SARC) (**Supplementary Table S7**) and corresponding normal tissues in both TCGA and GTEX were acquired from UCSC Xena (Vivian et al., 2017)⁷. The association of the expression level of the selected transcription factors with *PIP4K2A* was subsequently calculated using linear regression models as well.

To further detect the association of *PIP4K2A* with these genes, the r^2 of *PIP4K2A* and its related transcription factors as well as *RAC2*, that we calculated by linear model as we described above, were imported to generate the heatmap, in which white indicates non-significance.

Comparison of PIP4K2A and Its Correlated Gene on Overall Survival Rate

The clinical information for survival analyses was also acquired from UCSC Xena, including patient ID, overall survival information, etc. We divided the expression levels of *PIP4K2A*-related transcription factors into two groups, high-expression level and low-expression level, by the significant low point of their histogram, and used this standard to conduct survival analysis in different cancer types.

⁷ https://xenabrowser.net

REFERENCES

- Ahuja, H. G., Jat, P. S., Foti, A., Bar-Eli, M., and Cline, M. J. (1991). Abnormalities of the retinoblastoma gene in the pathogenesis of acute leukemia. *Blood* 78, 3259–3268.
- Bhojwani, D., Kang, H., Menezes, R. X., Yang, W., Sather, H., Moskowitz, N. P., et al. (2008). Gene expression signatures predictive of early response and outcome in high-risk childhood acute lymphoblastic leukemia: a Children's Oncology Group Study [corrected]. J. Clin. Oncol. 26, 4376–4384. doi: 10.1200/ JCO.2007.14.4519
- Buffler, P. A., Kwan, M. L., Reynolds, P., and Urayama, K. Y. (2005). Environmental and genetic risk factors for childhood leukemia: appraising the evidence. *Cancer Invest* 23, 60–75. doi: 10.1081/CNV-46402
- Bungaro, S., Dell'Orto, M. C., Zangrando, A., Basso, D., Gorletta, T., Lo Nigro, L., et al. (2009). Integration of genomic and gene expression data of childhood ALL without known aberrations identifies subgroups with specific genetic hallmarks. *Genes Chromos. Cancer* 48, 22–38. doi: 10.1002/gcc.20616
- Carricaburu, V., Lamia, K. A., Lo, E., Favereaux, L., Payrastre, B., Cantley, L. C., et al. (2003). The phosphatidylinositol (PI)-5-phosphate 4-kinase type II enzyme controls insulin signaling by regulating PI-3,4,5-trisphosphate degradation. *Proc. Natl. Acad. Sci. U.S.A.* 100, 9867–9872. doi: 10.1073/pnas. 1734038100
- Curtis, C., Shah, S. P., Chin, S. F., Turashvili, G., Rueda, O. M., Dunning, M. J., et al. (2012). The genomic and transcriptomic architecture of 2,000 breast tumours reveals novel subgroups. *Nature* 486, 346–352. doi: 10.1038/nature10983
- Den Boer, M. L., van Slegtenhorst, M., De Menezes, R. X., Cheok, M. H., Buijs-Gladdines, J. G., Peters, S. T., et al. (2009). A subtype of childhood acute lymphoblastic leukaemia with poor treatment outcome: a genome-wide

DATA AVAILABILITY STATEMENT

Datasets analyzed for this study are available in GEO repository (http://www.ncbi.nlm.nih.gov/gds): GSE635, GSE10792, GSE13351, GSE13425, GSE10255, GSE4698, GSE33315, GSE7440, GSE11877, and GSE13204, and TCGA database (https://cancergenome.nih.gov/).

AUTHOR CONTRIBUTIONS

SZ and ZL designed the study. XiY, YD, FZ, PW, JZ, DY, FL, XZ, and DZ wrote the manuscript. LB, XX, HW, XuY, WaZ, HG, and WeZ performed the data analyses. LY, QH, HX, YS, YZ, and YW contributed to the conception of the study and drafted the manuscript. All authors contributed significantly in writing the manuscript and read and approved the final manuscript.

FUNDING

This study was supported by National Key Research Development Program [Grant No. 2016YFC0905000 (2016YFC0905002)], and the National Natural Science Foundation of China (Grant Nos. 81522028 and 81673452).

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fgene. 2018.00721/full#supplementary-material

classification study. Lancet Oncol. 10, 125-134. doi: 10.1016/S1470-2045(08) 70339-5

- Divecha, N., Truong, O., Hsuan, J. J., Hinchliffe, K. A., and Irvine, R. F. (1995). The cloning and sequence of the C isoform of PtdIns4P 5-kinase. *Biochem. J.* 309(Pt 3), 715–719. doi: 10.1042/bj3090715
- Elouarrat, D., van der Velden, Y. U., Jones, D. R., Moolenaar, W. H., Divecha, N., and Haramis, A. P. (2013). Role of phosphatidylinositol 5-phosphate 4-kinase alpha in zebrafish development. *Int. J. Biochem. Cell Biol.* 45, 1293–1301. doi: 10.1016/j.biocel.2013.03.009
- Emerling, B. M., Hurov, J. B., Poulogiannis, G., Tsukazawa, K. S., Choo-Wing, R., Wulf, G. M., et al. (2013). Depletion of a putatively druggable class of phosphatidylinositol kinases inhibits growth of p53-null tumors. *Cell* 155, 844–857. doi: 10.1016/j.cell.2013.09.057
- Fedorenko, O., Strutz-Seebohm, N., Henrion, U., Ureche, O. N., Lang, F., Seebohm, G., et al. (2008). A schizophrenia-linked mutation in PIP5K2A fails to activate neuronal M channels. *Psychopharmacology (Berl)* 199, 47–54. doi: 10.1007/s00213-008-1095-x
- Fedorenko, O., Tang, C., Sopjani, M., Foller, M., Gehring, E. M., Strutz-Seebohm, N., et al. (2009). PIP5K2A-dependent regulation of excitatory amino acid transporter EAAT3. *Psychopharmacology (Berl)* 206, 429–435. doi: 10.1007/ s00213-009-1621-5
- Fiume, R., Stijf-Bultsma, Y., Shah, Z. H., Keune, W. J., Jones, D. R., Jude, J. G., et al. (2015). PIP4K and the role of nuclear phosphoinositides in tumour suppression. *Biochim. Biophys. Acta* 1851, 898–910. doi: 10.1016/j.bbalip.2015.02.014
- Greaves, M. (2006). Infection, immune responses and the aetiology of childhood leukaemia. *Nat. Rev. Cancer* 6, 193–203. doi: 10.1038/nrc1816
- Gupta, A., Toscano, S., Trivedi, D., Jones, D. R., Mathre, S., Clarke, J. H., et al. (2013). Phosphatidylinositol 5-phosphate 4-kinase (PIP4K) regulates TOR

⁶ http://bioinfo.life.hust.edu.cn/AnimalTFDB2/

signaling and cell growth during *Drosophila* development. *Proc. Natl. Acad. Sci.* U.S.A. 110, 5963–5968. doi: 10.1073/pnas.1219333110

- Haferlach, T., Kohlmann, A., Wieczorek, L., Basso, G., Kronnie, G. T., Bene, M. C., et al. (2010). Clinical utility of microarray-based gene expression profiling in the diagnosis and subclassification of leukemia: report from the international microarray innovations in leukemia study group. J. Clin. Oncol. 28, 2529–2537. doi: 10.1200/JCO.2009.23.4732
- Holleman, A., Cheok, M. H., den Boer, M. L., Yang, W., Veerman, A. J., Kazemier, K. M., et al. (2004). Gene-expression patterns in drug-resistant acute lymphoblastic leukemia cells and response to treatment. *N. Engl. J. Med.* 351, 533–542. doi: 10.1056/NEJMoa033513
- Hou, Q., Liao, F., Zhang, S., Zhang, D., Zhang, Y., Zhou, X., et al. (2017). Regulatory network of GATA3 in pediatric acute lymphoblastic leukemia. Oncotarget 8, 36040–36053. doi: 10.18632/oncotarget.16424
- Huang da, W., Sherman, B. T., and Lempicki, R. A. (2009a). Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists. *Nucleic Acids Res.* 37, 1–13. doi: 10.1093/nar/gkn923
- Huang da, W., Sherman, B. T., and Lempicki, R. A. (2009b). Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat. Protoc.* 4, 44–57. doi: 10.1038/nprot.2008.211
- Jones, D. R., Foulger, R., Keune, W. J., Bultsma, Y., and Divecha, N. (2013). PtdIns5P is an oxidative stress-induced second messenger that regulates PKB activation. FASEB J. 27, 1644–1656. doi: 10.1096/fj.12-218842
- Jude, J. G., Spencer, G. J., Huang, X., Somerville, T. D., Jones, D. R., Divecha, N., et al. (2015). A targeted knockdown screen of genes coding for phosphoinositide modulators identifies PIP4K2A as required for acute myeloid leukemia cell proliferation and survival. Oncogene 34, 1253–1262. doi: 10.1038/onc.2014.77
- Kang, H., Chen, I. M., Wilson, C. S., Bedrick, E. J., Harvey, R. C., Atlas, S. R., et al. (2010). Gene expression classifiers for relapse-free survival and minimal residual disease improve risk classification and outcome prediction in pediatric B-precursor acute lymphoblastic leukemia. *Blood* 115, 1394–1405. doi: 10.1182/ blood-2009-05-218560
- Kirschner-Schwabe, R., Lottaz, C., Todling, J., Rhein, P., Karawajew, L., Eckert, C., et al. (2006). Expression of late cell cycle genes and an increased proliferative capacity characterize very early relapse of childhood acute lymphoblastic leukemia. *Clin. Cancer Res.* 12, 4553–4561. doi: 10.1158/1078-0432.CCR-06-0235
- Lamia, K. A., Peroni, O. D., Kim, Y. B., Rameh, L. E., Kahn, B. B., and Cantley, L. C. (2004). Increased insulin sensitivity and reduced adiposity in phosphatidylinositol 5-phosphate 4-kinase beta-/- mice. *Mol. Cell. Biol.* 24, 5080–5087. doi: 10.1128/MCB.24.11.5080-5087.2004
- Liao, F., Yin, D., Zhang, Y., Hou, Q., Zheng, Z., Yang, L., et al. (2016). Association between PIP4K2A polymorphisms and acute lymphoblastic leukemia susceptibility. *Medicine (Baltimore)* 95:e3542. doi: 10.1097/MD. 000000000003542
- Migliorini, G., Fiege, B., Hosking, F. J., Ma, Y., Kumar, R., Sherborne, A. L., et al. (2013). Variation at 10p12.2 and 10p14 influences risk of childhood B-cell acute lymphoblastic leukemia and phenotype. *Blood* 122, 3298–3307. doi: 10.1182/ blood-2013-03-491316
- Niu, N., Schaid, D. J., Abo, R. P., Kalari, K., Fridley, B. L., Feng, Q., et al. (2012). Genetic association with overall survival of taxane-treated lung cancer patients – A genome-wide association study in human lymphoblastoid cell lines followed by a clinical association study. *BMC Cancer* 12:422. doi: 10.1186/1471-2407-12-422
- Papaemmanuil, E., Hosking, F. J., Vijayakrishnan, J., Price, A., Olver, B., Sheridan, E., et al. (2009). Loci on 7p12.2, 10q21.2 and 14q11.2 are associated with risk of childhood acute lymphoblastic leukemia. *Nat. Genet.* 41, 1006–1010. doi: 10.1038/ng.430
- Perez-Andreu, V., Roberts, K. G., Harvey, R. C., Yang, W., Cheng, C., Pei, D., et al. (2013). Inherited GATA3 variants are associated with Ph-like childhood acute lymphoblastic leukemia and risk of relapse. *Nat. Genet.* 45, 1494–1498. doi: 10.1038/ng.2803
- Perez-Andreu, V., Roberts, K. G., Xu, H., Smith, C., Zhang, H., Yang, W., et al. (2015). A genome-wide association study of susceptibility to acute lymphoblastic leukemia in adolescents and young adults. *Blood* 125, 680–686. doi: 10.1182/blood-2014-09-595744

- Pui, C. H., and Evans, W. E. (2006). Treatment of acute lymphoblastic leukemia. *N. Engl. J. Med.* 354, 166–178. doi: 10.1056/NEJMra052603
- Rameh, L. E., Tolias, K. F., Duckworth, B. C., and Cantley, L. C. (1997). A new pathway for synthesis of phosphatidylinositol-4,5-bisphosphate. *Nature* 390, 192–196. doi: 10.1038/36621
- Schmitz, N. M., Leibundgut, K., and Hirt, A. (2005). CDK2 catalytic activity and loss of nuclear tethering of retinoblastoma protein in childhood acute lymphoblastic leukemia. *Leukemia* 19, 1783–1787. doi: 10.1038/sj.leu.240 3900
- Sherborne, A. L., Hosking, F. J., Prasad, R. B., Kumar, R., Koehler, R., Vijayakrishnan, J., et al. (2010). Variation in CDKN2A at 9p21.3 influences childhood acute lymphoblastic leukemia risk. *Nat. Genet.* 42, 492–494. doi: 10.1038/ng.585
- Sorich, M. J., Pottier, N., Pei, D., Yang, W., Kager, L., Stocco, G., et al. (2008). In vivo response to methotrexate forecasts outcome of acute lymphoblastic leukemia and has a distinct gene expression profile. *PLoS Med.* 5:e83. doi: 10.1371/journal.pmed.0050083
- Stock, W., Tsai, T., Golden, C., Rankin, C., Sher, D., Slovak, M. L., et al. (2000). Cell cycle regulatory gene abnormalities are important determinants of leukemogenesis and disease biology in adult acute lymphoblastic leukemia. *Blood* 95, 2364–2371.
- Szklarczyk, D., Franceschini, A., Wyder, S., Forslund, K., Heller, D., Huerta-Cepas, J., et al. (2015). STRING v10: protein-protein interaction networks, integrated over the tree of life. *Nucleic Acids Res.* 43(Database issue), D447– D452. doi: 10.1093/nar/gku1003
- Trevino, L. R., Yang, W., French, D., Hunger, S. P., Carroll, W. L., Devidas, M., et al. (2009). Germline genomic variants associated with childhood acute lymphoblastic leukemia. *Nat. Genet.* 41, 1001–1005. doi: 10.1038/ng.432
- Vivian, J., Rao, A. A., Nothaft, F. A., Ketchum, C., Armstrong, J., Novak, A., et al. (2017). Toil enables reproducible, open source, big biomedical data analyses. *Nat. Biotechnol.* 35, 314–316. doi: 10.1038/nbt.3772
- Walsh, K. M., de Smith, A. J., Chokkalingam, A. P., Metayer, C., Dahl, G. V., Hsu, L. I., et al. (2013). Novel childhood ALL susceptibility locus BMI1-PIP4K2A is specifically associated with the hyperdiploid subtype. *Blood* 121, 4808–4809. doi: 10.1182/blood-2013-04-495390
- Wilcox, A., and Hinchliffe, K. A. (2008). Regulation of extranuclear PtdIns5P production by phosphatidylinositol phosphate 4-kinase 2alpha. *FEBS Lett.* 582, 1391–1394. doi: 10.1016/j.febslet.2008.03.022
- Xu, H., Cheng, C., Devidas, M., Pei, D., Fan, Y., Yang, W., et al. (2012). ARID5B genetic polymorphisms contribute to racial disparities in the incidence and treatment outcome of childhood acute lymphoblastic leukemia. J. Clin. Oncol. 30, 751–757. doi: 10.1200/JCO.2011.38.0345
- Xu, H., Yang, W., Perez-Andreu, V., Devidas, M., Fan, Y., Cheng, C., et al. (2013). Novel susceptibility variants at 10p12.31-12.2 for childhood acute lymphoblastic leukemia in ethnically diverse populations. J. Natl. Cancer Inst. 105, 733–742. doi: 10.1093/jnci/djt042
- Xu, H., Zhang, H., Yang, W., Yadav, R., Morrison, A. C., Qian, M., et al. (2015). Inherited coding variants at the CDKN2A locus influence susceptibility to acute lymphoblastic leukaemia in children. *Nat. Commun.* 6:7553. doi: 10.1038/ ncomms8553
- Zhang, J., Ding, L., Holmfeldt, L., Wu, G., Heatley, S. L., Payne-Turner, D., et al. (2012). The genetic basis of early T-cell precursor acute lymphoblastic leukaemia. *Nature* 481, 157–163. doi: 10.1038/nature10725

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Zhang, Li, Yan, Bao, Deng, Zeng, Wang, Zhu, Yin, Liao, Zhou, Zhang, Xia, Wang, Yang, Zhang, Gao, Zhang, Yang, Hou, Xu, Zhang, Shu and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.