



New Analysis Framework Incorporating Mixed Mutual Information and Scalable Bayesian Networks for Multimodal High Dimensional Genomic and Epigenomic Cancer Data

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Wang X, Branciamore S, Gogoshin G, Ding S and Rodin AS (2020) New Analysis Framework Incorporating Mixed Mutual Information and Scalable Bayesian Networks for Multimodal High Dimensional Genomic and Epigenomic Cancer Data. Front. Genet. 11:648. doi: 10.3389/fgene.2020.00648 We propose a novel two-stage analysis strategy to discover candidate genes associated with the particular cancer outcomes in large multimodal genomic cancers databases, such as The Cancer Genome Atlas (TCGA). During the first stage, we use mixed mutual information to perform variable selection; during the second stage, we use scalable Bayesian network (BN) modeling to identify candidate genes and their interactions. Two crucial features of the proposed approach are (i) the ability to handle mixed data types (continuous and discrete, genomic, epigenomic, etc.) and (ii) a flexible boundary between the variable selection and network modeling stages — the boundary that can be adjusted in accordance with the investigators' BN software scalability and hardware implementation. These two aspects result in high generalizability of the proposed analytical framework. We apply the above strategy to three different TCGA datasets (LGG, Brain Lower Grade Glioma; HNSC, Head and Neck Squamous Cell Carcinoma; STES, Stomach and Esophageal Carcinoma), linking multimodal molecular information (SNPs, mRNA expression, DNA methylation) to two clinical outcome variables (tumor status and patient survival). We identify 11 candidate genes, of which 6 have already been directly implicated in the cancer literature. One novel LGG prognostic factor suggested by our analysis, methylation of TMPRSS11F type II transmembrane serine protease, presents intriguing direction for the follow-up studies.

Keywords: The Cancer Genome Atlas, Bayesian networks, multimodal big data, variable selection, mixed mutual information, methylation, genomic and epigenomic molecular data

INTRODUCTION

The Cancer Genome Atlas (TCGA) resource contains genomic data compiled for more than 30 different types/subtypes of cancer (Tomczak et al., 2015). For each type, clinical outcome/progression data (e.g., tumor status and patient survival) for a considerable number of patients is matched to the large-scale molecular data. The latter is multimodal, ranging from genetic (e.g., somatic mutations) to expression (e.g., RNA-seq gene expression) to epigenetic

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(e.g., promoter methylation) data. Not surprisingly, there is substantial enthusiasm for causally linking the latter to the former using various modeling and secondary data analysis techniques (Jeong et al., 2015; Phan et al., 2016; Hou et al., 2018; Tian et al., 2018; Xu et al., 2018). The ultimate goals of these analyses are (i) to gain better mechanistic understanding of the underlying molecular biology of cancer, primarily by identifying important genes and their interactions; (ii) to construct compact and efficient clinical predictors (e.g., prognostic scores, indices and signatures); (iii) to associate the latter with the particular patient groups and subgroups, in the context of personalized/precision medicine. One of the more attractive and popular methods for such multivariate analysis is Bayesian networks (BNs) (Heckerman, 1995), a wellestablished fixture in computational systems biology (Friedman et al., 2000). Among the BN advantages are their probabilistic nature, model flexibility, ability to handle non-additive, higherorder, interactions, and ease of the result interpretation. However, applications of BNs to the TCGA (and TCGA-like) data (Gevaert et al., 2006; Xu et al., 2012, 2014; Wang et al., 2013; Huang et al., 2015; Zhu et al., 2015; Kaiser et al., 2016; Wu et al., 2017) face two principal difficulties: combining mixed data types in a single analysis framework, and achieving sufficient (for genomic data) scalability, simultaneously. (These, of course, are the two fundamental, and interconnected, BN modeling challenges in general, not just in the TCGA application). The latest developments in addressing these two challenges encompass more efficient computational approaches (Gogoshin et al., 2017; Ramsey et al., 2017), and mathematically rigorous and robust methods for handling mixed data, such as mixed local probability models and/or adaptive discretization (Gogoshin et al., 2017; Andrews et al., 2018; Sedgewick et al., 2018). Nevertheless, resolving both difficulties simultaneously in a generalizable toolkit (seamlessly applicable, for example, across the individual TCGA datasets) remains elusive. A promising approach to devising such a toolkit would be to precede the comparatively exhaustive NP-hard BN modeling with a variable selection procedure [for example (Zhang et al., 2014)], where the full dataset is pared down to a subset of variables most relevant to a particular clinical outcome or phenotype. While alleviating the scalability issue, this, however, could potentially "throw away the wheat with the chaff," especially if the variable selection process (Blum and Langley, 1997; Guyon and Elisseeff, 2003) is of a simplistic and overly too restrictive kind (e.g., a statistically conservative univariate filter). There are three possible ways to address this, namely: (i) increase the scalability of the BN modeling to genomic data levels (possible, but impractical for frequent/serial analyses), (ii) incorporate higher-order interactions into the variable selection step (thus "upgrading" it from the simple filter to the wrapper [Kohavi and John, 1997; Guyon and Elisseeff, 2003; Leng et al., 2010) — this is the solution implemented in Zhang et al. (2014)], or (iii) adjust the transition boundary between the variable selection step and the BN modeling step, depending on the investigators' computational resources and the nature (dimensionality, sparseness, heterogeneity) of the actual data. It is the third analytical strategy that we propose in this study,

with the goal to achieve the optimal compromise between the computational practicality and modeling exhaustiveness.

In our analysis pipeline, we start with the variable selection procedure based on the mixed-type Mixed Mutual Information (MMI) forward selection filter. We compute the MMI values for all available gene-outcome (specifically, tumor status and patient survival) pairs, and use the MMI frequency distribution to select top variables/genes (or, alternatively, to remove bottom variables/genes) before moving on to the BN modeling. This mixed-type measure-based approach to gene selection is the principal innovation of this paper. We then use the maximum entropy (ME) - based discretization to construct the mixed-type BNs using our previously reported scalable BN modeling algorithm and software (Gogoshin et al., 2017). Subsequently, we concentrate on the sub-networks centered around the clinical outcome variables of interest, and identify the molecular gene components belonging to these subnetworks.

The proposed analysis strategy has been applied by us to 12 different TCGA cancer datasets. This allowed us to check for robustness, scalability and generalizability. Here, we present the results for the Brain Lower Grade Glioma (LGG), Head and Neck Squamous Cell Carcinoma (HNSC) and Stomach and Esophageal Carcinoma (STES) datasets (all three datasets being reasonably well-populated and proportionally balanced across the different outcomes and molecular data types). For the purposes of this particular analysis, we decided to concentrate on three types of molecular data, one discrete (somatic mutations) and two - continuous (RNA-seq gene expression, and promoter methylation). This selection is reflective of the recent trends in multimodal cancer data analyses (Zhang et al., 2014; Yoo et al., 2017), makes sense in the broad cancer genetics context (Phipps et al., 2016; Fang et al., 2017; Liang et al., 2017; Rajesh et al., 2017; Zhang C. et al., 2017; Koch et al., 2018), and underscores the comparative importance of the methylation molecular data (Koch et al., 2018). While focusing solely on the gene-centric modalities is inherently limiting (many disease-linked SNPs are localized in the non-coding regions), one of the primary purposes of this study was to showcase the MMI approach (enjoining three different modalities in a single measure/score), which necessitated the gene-centric analysis. In future, we plan to generalize our analytical framework to other, non-genecentric, data.

We conclude by identifying a compact list of genes potentially associated with cancer-related clinical phenotypes (tumor status and patient survival), scrutinizing these genes in light of the current literature, and discussing the generalizability of our approach to the different datasets, diseases and molecular data types.

MATERIALS AND METHODS

Data Preprocessing

The Cancer Genome Atlas, LGG, HNSC, and STES datasets were downloaded for the clinical data ["Clinical_Pick_Tier1

(MD5)"], SNP data ["Mutation_Packager_Calls (MD5)"], expression data ["mRNAseq_Preprocess (MD5)"] and promotercentric methylation data ["Methylation_Preprocess (MD5)"]. Patients were further subdivided into (i) two disease progression categories (according to the "tumor status" variable), and (ii) two patient survival categories (high death risk, with survival less than 2 years, and low death risk, with survival more than 2 years, which is a common cutoff point in recent cancer literature). We further excluded patients with ambiguous or missing outcome variable values (e.g., no survival status, survival status as "living" with survival time less than 2 years, tumor status neither "tumor-free" nor "with tumor," etc.). These clinical variables ("tumor status" and "2-year survival") were subsequently used for the variable selection purposes, and, eventually, to extract "tumor status" and "survival" centered sub-networks from the full BNs. Expression data and methylation data (designated by "E" and "M" below, for brevity) were not discretized at this stage, as both variable selection and BN construction tools in our computational pipeline can, by design, accept mixed (continuous and discreet) variable types. SNP (somatic mutation) data (designated by "S" below) were compressed into a binary variable (presence or absence of at least one non-synonymous mutation in at least one sample of the particular gene).

After filtering out patient records with incomplete, partially missing, or ambiguously labeled data, the final datasets consisted of 4782 genes (LGG), 12516 genes (HNSC) and 16164 genes (STES). 273 patient records were available for LGG/tumor status analysis (140 patients with tumor, 133 without); 213 patients – for LGG/survival (120 patients with survival less than 2 years, 93 with long-term survival). Similarly, 260 patient records were available for HNSC/tumor status analysis (94 patients with tumor, 166 without); 139 patients – for HNSC/survival (40 patients with survival less than 2 years, 99 with long-term survival). Finally, 403 patient records were available for STES/tumor status analysis (147 patients with tumor, 256 without); 258 patients – for STES/survival (191 patients with survival less than 2 years, 67 with long-term survival).

Here we would like to re-emphasize that it is possible to include other different molecular data types and outcome variables, both continuous and discrete, into the proposed framework without substantial alterations to the analysis pipeline, except for some rudimentary data preprocessing.

Variable Selection

There are very few BN algorithms/software solutions that scale up to (epi)genomic levels (tens to hundreds of thousands of variables) (Gogoshin et al., 2017; Ramsey et al., 2017). Even with these, exhaustive analyses require dedicated hardware and weeks of processing time. This might be acceptable for a one-off, "final" analysis, but is clearly impractical for the exploratory research. This is why it is a common practice to carry out variable selection (or feature selection, or feature set reduction) in order to generate a comparatively compact subset of variables to be subsequently fed into the network modeling algorithm/software (Guyon and Elisseeff, 2003). Variable selection approaches range from the very simple (univariate filters) to increasingly more sophisticated; at some point, the latter become essentially indistinguishable from the multivariate modeling methods per se. Depending on the dataset to be analyzed, different "couplings" of variable selection and multivariate modeling methods might prove to be more or less effective, and it is difficult to devise a priori the objectively optimal combination for each new dataset. For a principally network-centric data analysis approach (innate to the systems biology), it would make sense to feed as many variables into the network-building module as possible, thus "delegating" the resolution of the higher-order / non-additive interactions and conditional independence relationships to the BN algorithm itself. Therefore, for the exploratory research, we suggest that the investigators first define the upper BN scalability limit that they are comfortable with (given the available software/hardware), and then adjust the variable selection cutoff point accordingly. For more "finalized" analysis, that limit should be raised higher (and the variable selection process, consequently, be made less restrictive).

In TCGA dataset (and other similar (epi)genomic resources), there are tens of thousands of potentially predictive/relevant variables (roughly proportional to the number of genes in the human genome). The "hand off" point between the variable selection and BN analysis steps should therefore vary between 100s of variables (for the exploratory and preliminary analyses) and 1,000s of variables (for the final analyses). The actual number might also depend on the shape of the variable selection curve, or on the statistical significance criteria–we stop adding increasingly less significant variables during the forward variable selection process (or stop removing increasingly more significant variables during the backward variable elimination process) when a certain statistical significance cutoff point is reached (Rodin et al., 2009). The above considerations were taken into account in the course of this study, as detailed in the section "Results" below.

It is difficult to integrate the multimodal, mixed-type, data into the variable selection process (filter or wrapper) as, until recently, there has been a paucity of the usable mixed-type metrics. In this study, a recently developed measure, Mixed Mutual Information (MMI) (Gao et al., 2018), was used to link the gene information (a mixed-type vector consisting of the S, E, and M molecular data components for each gene) to the clinical variable (tumor status or 2-year survival) in a "forward-selection-filter" variable selection procedure. MMI is a non-parametric and distribution-free measure [which makes it more attractive than the alternatives, such as linear correlation especially in the biological networks context (Margolin et al., 2006; Asur et al., 2007)] that is based on the entropy estimates from k-nearest neighbor (k-NN) distances (Kraskov et al., 2004). It is, therefore, sensitive to the choice of the k parameter. Lower values of k (1–4) tend to lead to higher dispersion, while much higher values (>20) are associated with unnecessarily increased computational complexity and possible overfitting [, personal communication from Gao et al. (2018)]. We have evaluated different values of k on the actual TCGA datasets by measuring the Jaccard index for the pairs of consecutive (in k) postselection variable sets as a function of k. The index appeared to stabilize in the 8-20 range in 12 different TCGA datasets

analyzed (see section "Results" below); therefore, k was set at 15 throughout this study.

Bayesian Networks Modeling

Bayesian networks modeling, in its basic form, reconstructs a sparse graphical representation of a joint multivariate probability distribution of random variables from a "flat" dataset. Nodes in the network represent random variables, edges - dependencies. Absence of an edge between the two nodes indicates conditional independence between them. Recent work in BN methodology refinement led to significant progress in scalability - our latest BN modeling software implementation (Gogoshin et al., 2017) easily processes datasets up to \sim 1 mln variables \times 1 mln datapoints. Handling mixed variable types (both continuous and discrete, in a typical application) is still not entirely seamless; it was recently suggested (Gogoshin et al., 2017; Andrews et al., 2018) that adaptive discretization (of continuous variables) might be preferable to forcing mixed local probability models. Consequently, we were using maximum entropy - based threebin discretization throughout this study - expression data ("E" molecular data component) and methylation data ("M" molecular data component) were discretized into three bins which has attractive mathematical properties, and has been shown by us earlier to maintain near-optimal over/under-fitting balance (Gogoshin et al., 2017).

Detailed description of the BN methodology in general and of our implementation (including applications to other types of high-dimensional biological data) in particular can be found in Gogoshin et al. (2017), Zhang X. et al. (2017); here we will only note that (1) our BN implementation uses a hybrid "sparse candidates" + "search-and-score" graduate descent algorithm coupled with various model scoring metrics and maximum entropy-based adaptive discretization; (2) in the resulting BN visualizations, numbers next to the edges and edge "thickness" indicate relative edge strengths (the numbers are the model scores' ratios for the models with/without corresponding edges, which are proportional to the marginal likelihood ratios); (3) directionality in the network (arrow points attached to the edges, when present) does not necessarily imply the causality flow, and is used predominantly for the mathematical convenience (to avoid cyclic dependencies); (4) when deciphering conditional dependence and independence patterns, it is useful to concentrate on the immediate Markov neighborhood (MN) of a particular variable of interest (such as a clinical outcome). This neighborhood can be roughly defined as all the nodes that are in immediate contact with ("one degree of separation" from) the node representing the aforementioned variable of interest. Under certain conditions, given its MN, the variable of interest is conditionally independent of the remaining variables (rest of the network). Therefore, deriving a MN for a variable of interest is analogous to the variable selection activity, specifically of the embedded variety (Guyon and Elisseeff, 2003). The central step in our computational analysis pipeline is using full BN reconstruction to generate the MN for the clinical outcome variable, and then ascertaining the interplay of the (small number of) gene-related variables (S, E and M

molecular data components) within that MN. (It should be noted that MN is a simplification of the more rigorous concept of Markov Blanket – meaning, for our purposes, that sometimes "two degrees of separation" are needed for encapsulating a variable/node of interest).

RESULTS

Figure 1 depicts the variable selection process for six possible combinations of two clinical variables ("tumor status" and "survival") and three TCGA cancer datasets (LGG, HNSC, and STES). MMI (mixed mutual information) between (S, E, M) and tumor status/survival was computed for 4782 genes (LGG), 12516 genes (HNSC), and 16164 genes (STES). (All six gene lists, with corresponding MMI values, are available in Supplementary Tables S1-S6). The histogram representation of the MMI distribution, as shown in Figure 1, is convenient, as it allows to evaluate (both visually and quantitatively) the relative predictive values of the top-ranking genes with respect to the outcome variable classification. For the purposes of this study, and to make the resulting full BNs "observable," we have chosen the "top genes" cutoff value of 99.5% MMI CDF (cumulative distribution function), which leads to the selection of 24 genes (72 future BN nodes/variables in total, comprising 24 S, 24 E, and 24 M components) out of 4728 for two LGG networks, 63 genes (189 nodes/variables) out of 12516 for two HNSC networks, and 81 genes (243 nodes/variables) out of 16164 for two STES networks. Note that the S, E, and M components of each gene vector were considered as the separate nodes/variables in the subsequent BN construction, as at this time we do not have a BN scoring function that can incorporate mixed multivariate distance measures. It should also be noted that although MMI, intuitively, should not be negative, due to the way it is computed it can get into the negative range when (i) continuous variables are involved, and (ii) the number of dimensions is more than two (four, in our case). This said, all the negative MMI values in Figure 1 reside well within the allowed algorithmic negative deviation range, and should not influence the variable rankings [personal communication from Gao et al. (2018)].

Interestingly, every histogram in **Figure 1** has a heavy right tail, which sometimes appears to follow a clear "knee point" – for example, at MMI $\sim = 0.08$ in **Figures 1A–C**. This suggests that MMI > 0.08 could also be used as a "natural" cutoff value, at least in these three datasets.

The variable selection distributions shown in **Figure 1** were derived with the MMI parameter k set at 15. **Figure 2** illustrates the motivation behind that choice, using the LGG/survival dataset example. Shown is the plot of the Jaccard index (JI, a.k.a. set "Intersection over Union," which is a common measure of sample set similarity) comparing the gene/variable sets resulting from the above variable selection procedure, with cutoff set at 99.5% MMI CDF, where JI(k) compares the sets obtained with k and k+1. It is clear that as k reaches ~15, the set composition somewhat stabilizes; further increase in k does not seem to offer any advantages. (JI plots for the other datasets exhibit a similar pattern).



FIGURE 1 | Variable selection process for six combinations of two clinical outcome variables ("tumor status" and "survival") and three TCGA cancer datasets (LGG, HNSC, STES). MMI between the (S, E, M) molecular data vector and tumor status/survival was computed for 4782 genes (LGG), 12516 genes (HNSC) and 16164 genes (STES). The histogram representation of the MMI distribution is shown with the selection of "top" (i.e., with the MMI CDF > 99.5%) genes superimposed on the right tail of the MMI frequency distribution. (A) LGG/tumor status; (B) LGG/survival; (C) HNSC/tumor status; (D) HNSC/survival; (E) STES/tumor status; (F) STES/survival.

Figures 3, 4 depict the full BNs obtained from the LGG/tumor status and LGG/survival datasets. **Supplementary Data Sheets S1–S6** depict, in PDF format, the full BNs obtained from the LGG/tumor status, LGG/survival, HNSC/tumor status, HNSC/survival, STES/tumor status, and STES/survival datasets, respectively. Six corresponding DOT (standard network / causal graphical models format) files can be found in the Supplementary Tables S7–S12.

While the resulting full BNs, in PDF format, are zoomable and searchable, and the DOT files can be exported into the specialized network-oriented software, the full BNs tend to be visually overwhelming for the number of variables/nodes >100. Consequently, **Figures 5–10** depict the immediate MNs of the clinical variables/nodes in the corresponding six BNs: LGG/tumor status (**Figure 5**), LGG/survival (**Figure 6**), HNSC/tumor status (**Figure 7**), HNSC/survival (**Figure 8**), STES/tumor status (**Figure 9**) and STES/survival (**Figure 10**).

It is noticeable in **Figures 5–10** that all three molecular data components (S, E, and M) are represented in the MNs. This testifies to the efficacy and proportionality of both the MMI measure (during the variable selection stage) and the maximum entropy - based discretization (during the BN construction stage). Also of note, for some genes, more than one component is present (HTR4 E and S for STES/tumor status, CHIA E and S for LGG/tumor status, AFP E and S for LGG/tumor status). Conversely, some genes are associated with both tumor status and survival (MUC4 for HNSC, TMPRSS11F, SLC6A18, and DEFB119 for LGG).

The performance of our BN reconstruction algorithm or software is discussed in general terms in Gogoshin et al. (2017); here, we will evaluate the statistical significance of the resulting MNs. While the edge strength estimates in Figures 5-10 are useful in the relative sense, they do not immediately translate into the statistical significance measurements (such as *p*-values). Therefore, we have augmented the edge strengths with the *p*-values obtained via two-sample Kolmogorov-Smirnov (KS) probability distribution equality test (for continuous E and M molecular component variables) and two-sided Fisher's exact test (for discrete S molecular component variable). To illustrate the KS test application, Figure 11 shows CDFs, separately for two "tumor status" groups, for seven continuous variables present in the MN depicted in Figure 5 (LGG/tumor status), in order of decreasing edge strength (Figure 11A, MMP1_M; Figure 11B, DDX4_E; Figure 11C, AFP_E; Figure 11D, CHIA_E; Figure 11E, TMPRSS11F_M; Figure 11F, KERA_E; Figure 11G, MUC16_E). Only MMP1_M and DDX4_E appear to be statistically highly significant, with TMPRSS11F_M being arguably a borderline case.

Table 1 lists the *p*-values for all 55 potentially predictive molecular gene components present in six MNs depicted in **Figures 5–10**, in order of decreasing edge strength for each network / MN. 12 gene components were found to be statistically significant (marked with an asterisk in **Table 1**), however, we decide to exclude LCT_S (marked with ** in **Table 1**) from further scrutiny because of the very low mutation counts in both survival groups.



Subsequently, we performed manual literature / database search to ascertain if any of the remaining 11 genes were previously reported in the cancer context. The following resources were used: GeneCards (Stelzer et al., 2016) and DisGeNET (Pinero et al., 2017) databases, PubMed, and Google Scholar. Six genes were found to be implicated in cancer etiology / progression / clinical outcomes with high degree of certainty: MMP1, DDX4, TRPM3, DPP6, KCNA1, and MUC17 (Senapati et al., 2010; Saied et al., 2012; Lallet-Daher et al., 2013; Kawal et al., 2016; Park et al., 2016; Schudrowitz et al., 2017). Four genes (SLC7A14, LRRIQ, SLCO1B3, and SLC9A4) were supported by weaker, circumstantial evidence (Chan-On et al., 2013; Matullo et al., 2013; Fridley et al., 2016; Tanaka et al., 2017). One gene, TMPRSS11F, has not been discussed in the cancer context before, to the best of our knowledge [see also (Kataoka et al., 2018)]. However, increased expression levels of a similar type II transmembrane serine protease, TMPRSS11D, were found to be a significant non-small cell lung cancer survival predictor (Cao et al., 2017). Therefore, we suggest that TMPRSS11F should be further investigated as a strong predictive factor playing a role in LGG patients' clinical characteristics - survival, especially. Lower TMPRSS11F methylation values correspond to a poorer long-term (2-year) survival. One possible mechanism is via the proteolysis of extracellular matrix which, in turn, is linked to the metastatic processes (Cao et al., 2017).

In summary, our analysis framework confirmed six wellknown cancer-related genes, supplied additional evidence to support four other suspected cancer-related genes, and identified one novel potentially strongly predictive factor, methylation of TMPRSS11F.

DISCUSSION

Systems biology approach to the complex genetic and epigenetic cancer data analysis is arguably superior to the simpler singlegene (or even single-data type) alternatives. However, it is



part of the Supplementary Material.

intrinsically linked to the fundamental, interrelated, challenges – scalability, "curse of dimensionality," accounting for nonadditive, higher-order interactions, and visualization of the results (i.e., translation of the massive network graphs into concrete biomedical insights). In this study we propose a flexible and generalizable approach to the BN-based systems biology analysis of the multi-modal cancer data, using the TCGA database as an example. It consists of the variable selection step (which is not computationally demanding) and the BN reconstruction step (which is substantially computationally demanding). Ideally, the investigators would simply feed the complete dataset (all variables) into the BN software, obtain the full graphical model (no matter how large and complex), and then "zoom in" on the MN of the variable(s) of interest, such as a clinical outcome or a cancer phenotype. However, this is impractical for most real datasets and available hardware configurations.

Consequently, we propose starting with the variable selection step to select a (relatively) small subset of genes that are associated with the variable(s) of interest (tumor status and 2year survival in the present study). The principal novelty of our approach lies in using the MMI measure for the variable/gene selection, in which all possible types of molecular information (discrete and continuous, genetic and epigenetic) are considered



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simultaneously. The other innovative aspect of our approach lies in the adjustability of the "hand-off" point between the variable selection and BN modeling steps. This hand-off point can depend on the investigators' computational resources, the shape of the variable selection curves, or the predefined statistical cutoff points. For example, ~20 K genes can be reduced to 100-200 genes for the subsequent BNs construction, in which case the complete analysis takes less than an hour on a midlevel PC. When feeding the complete datasets (10,000-15,000 genes, in case of TCGA and similar genomic resources) into our BN software (Gogoshin et al., 2017), without the preliminary variable selection step, it takes about 3 days to build a full BN on a dedicated multi-core workstation. Therefore, the investigators can choose the appropriate balance depending on whether they are interested in a quick, exploratory analysis or a finalized, exhaustive one.

In our analyses, the final predictive gene sets (such as shown in **Table 1**) were different from the sets (of comparable sizes) of "top" genes obtained in the variable selection step alone (otherwise there would be no need to invoke the computationally expensive BN modeling step). This was to be expected, because BN modeling is a multivariate modeling tool (which aims to reconstruct the most fitting pattern of conditional independencies in the MN of a clinical variable), while MMI ranking is a univariate variable selection "filter" that does not account for the dependencies between the (top) genes. Another reason that the two corresponding gene sets tend to be different has to do with the fact that the first analysis stage is genecentric, whereas the second analysis stage separates the three molecular modalities. Limiting our analysis pipeline to just the first stage (MMI filter/ranking) would therefore miss the strong one-modality (but week multiple-modalities) predictors. In future, we plan to study the extent of intersection of such two sets as a function of the "hand-off" point (between MMI pre-ranking and full BN analysis) parameter.

Our computational pipeline is inherently generalizable, as it can be directly applied to any large multimodal genetic/epigenetic dataset with minimal preprocessing. The only two changeable parameters are the aforementioned variable selection / BN modeling hand-off point, and the BN discretization mechanism. The latter is currently set as the 3-bin maximum entropy-based discretization coupled with the multinomial local probability



model (Gogoshin et al., 2017). This is not the most elegant, or universally applicable, solution. In future, we plan to develop a novel BN model scoring function derived from a mixed distance measure (such as the MMI), or a similar metric that expresses divergence between the current network model and the data *via* mixed-type distances. The resulting two-stage analytical strategy will thus fully automatically deal with the mixed variables, in both of its stages. This has not been done before, so we plan to implement and test the MMI-based BN algorithm alongside the more established mixed-type BN solutions (hybrid local (A)

TABLE 1 | *P*-values for 55 potentially predictive molecular gene components present in six MNs depicted in **Figures 5–10**, subdivided by six datasets, in order of decreasing edge strength for each dataset/MN.

Gene_component	BN edge strength	p-value
LGG/tumor status		
MMP1_M	14.13	0.0001*
DDX4_E	12.19	0.0003*
AFP_E	4.41	0.8208
CHIA_E	4.09	0.9813
TMPRSS11F_M	2.76	0.1330
KERA_E	2.23	0.8928
MUC16 E	0.66	0.7521
AFP_S	0.61	0.4985
SLC6A18_S	0.51	~1.0
VIL1_S	0.45	0.2364
DEFB119_S	0.44	0.4872
C16orf11_S	0.41	~1.0
CHIA_S	0.41	~1.0
	0.41	
(B)		
LGG/survival		
TMPRSS11F_M	6.92	0.0003*
PKHD1L1_S	1.83	0.5818
FCRL5_S	1.55	0.1895
DEFB119_S	0.28	~1.0
SLCO6A1_S	0.28	~1.0
ABCG8_S	0.24	0.4366
TRPM1_S	0.02	0.1895
(C)		
HNSC/tumor status		
SLC7A14_E	14.48	2.1230e-05
MUC7_M	13.98	0.3432
ASB4_M	10.05	0.2832
CSMD1_S	5.07	~1.0
CNTNAP5_M	2.73	0.5261
MUC4_S	1.85	0.1496
CDH10_S	0.52	~1.0
WDR49_S	0.44	~1.0
(D)		
HNSC/survival		
FLG_M	6.07	0.2427
MUC4_S	5.42	0.4103
LCT_S	2.84	0.0226**
PCDH10_S	1.82	0.3544
KRT24_S	1.3	0.4942
ERN2_M	0.99	0.4832
CDH19_S	0.53	0.5787
(E)		
STES/tumor status		
LRRIQ_E	20.15	0.0004*
HTR4_E	7.18	0.2960

TABLE 1	Continued
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Gene_component	BN edge strength	<i>p</i> -value
DSG1_M	6.73	0.1204
TRPM3_S	3.80	0.0245*
APOB_S	3.50	0.2723
CNTNAP4_S	2.76	0.7784
KPRP_S	2.69	~1.0
DUSP27_S	1.72	0.5480
PCDHA1_S	1.72	0.8074
SBSN_S	1.54	0.6248
PNLDC1_S	1.43	~1.0
LOC100190940_S	0.83	~1.0
KCNV1_S	0.45	~1.0
HTR4_S	0.19	0.3594
CHRNA4_S	0.16	0.3968
ZNF716_S	0.10	~1.0
(F)		
STES/survival		
SLITRK1_M	7.12	0.1082
DPP6_S	4.37	0.0280
SLCO1B3_S	2.95	0.0059
PCLO_S	2.28	0.7238
LPA_S	2.25	0.0549
SLC9A4_S	1.96	0.0059
KCNA1_S	1.93	0.0108
COL11A1_E	1.83	0.2016
PGLYRP3_S	1.83	0.1119
PGC_S	1.43	0.4527
C20orf114_S	1.32	~1.0
MUC17_S	1.13	0.0008
FAM83C_S	1.10	0.6515
DCDC1_S	0.87	0.2051
KRT6B_S	0.64	~1.0

Twelve gene components were found to be statistically significant (marked with *); LCT_S (marked with **) was excluded from further analysis because of the very low mutation counts (zero mutations in >2-year survival group, three mutations in <2-year survival group).

probability models, adaptive discretization), and use both real and simulated data to investigate which method is preferable.

Another limitation of the present study has to do with its primary focus on the clinical outcomes / phenotypes; at this time, we decided to largely concentrate on the MNs of the clinical variables/nodes. In future, we intend to analyze the resulting full BNs more "holistically," paying attention to the general network topological properties, gene clusters, hub and bottleneck genes, etc. Consequently, one useful extension of our analytical framework would be to incorporate multiple clinical outcomes / phenotypes into the network analyses, to see if the inter-outcome dependencies are reflected in the resulting networks, and if they are mediated by other nodes/variables.

Application of our pipeline to TCGA data resulted in the identification of a number of candidate genes for the different clinical cancer characteristics, *via* varied molecular components. It is well known that epigenetic processes / DNA methylation play

an important role in many cancers' diagnosis, progression, and outcome; our results support that notion, as many of the most statistically significant predictors generated in the present study were in fact the methylation molecular components (**Table 1**). Notably, the one novel candidate gene pinpointed in this study, TMPRSS11F, likely would not have been identified *via* any other (non-epigenetic) modality. Our results, therefore, underscore the essentiality of the simultaneous analysis of different molecular modalities, including the epigenetic ones, for the precision or personalized medicine to be effective in cancer treatment.

DATA AVAILABILITY STATEMENT

All the intermediate datasets generated for this study are available on request to the corresponding author.

AUTHOR CONTRIBUTIONS

XW, SB, and AR conceptualized the study, carried out the analyses, and wrote the manuscript. GG and SD contributed to carrying out the analyses. All authors contributed to the article and approved the submitted version.

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REFERENCES

- Andrews, B., Ramsey, J., and Cooper, G. F. (2018). Scoring Bayesian Networks of Mixed Variables. Int. J. Data Sci. Anal. 6, 3–18. doi: 10.1007/s41060-017-0085-7
- Asur, S., Ucar, D., and Parthasarathy, S. (2007). An ensemble framework for clustering protein-protein interaction networks. *Bioinformatics* 23, i29–i40. doi: 10.1093/bioinformatics/btm212
- Blum, A. L., and Langley, P. (1997). Selection of relevant features and examples in machine learning. Artif. Intell. 97, 245–271. doi: 10.1016/s0004-3702(97) 00063-5
- Cao, X., Tang, Z., Huang, F., Jin, Q., Zhou, X., Shi, J., et al. (2017). High TMPRSS11D protein expression predicts poor overall survival in non-small cell lung cancer. *Oncotarget* 8, 12812–12819. doi: 10.18632/oncotarget.14559
- Chan-On, W., Nairismagi, M. L., Ong, C. K., Lim, W. K., Dima, S., Pairojkul, C., et al. (2013). Exome sequencing identifies distinct mutational patterns in liver fluke-related and non-infection-related bile duct cancers. *Nat. Genet.* 45, 1474–1478. doi: 10.1038/ng.2806
- Fang, X., Yin, Z., Li, X., Xia, L., Quan, X., Zhao, Y., et al. (2017). Multiple functional SNPs in differentially expressed genes modify risk and survival of non-small cell lung cancer in chinese female non-smokers. *Oncotarget* 8, 18924–18934. doi: 10.18632/oncotarget.14836
- Fridley, B. L., Ghosh, T. M., Wang, A., Raghavan, R., Dai, J., Goode, E. L., et al. (2016). Genome-wide study of response to platinum. taxane, and combination therapy in ovarian cancer: in vitro phenotypes, inherited variation, and disease recurrence. *Front Genet* 7:37. doi: 10.3389/fgene.2016.00037
- Friedman, N., Linial, M., Nachman, I., and Pe'er, D. (2000). Using Bayesian networks to analyze expression data. J. Comput. Biol. 7, 601–620. doi: 10.1089/ 106652700750050961

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fgene. 2020.00648/full#supplementary-material

TABLES S1–S6 | Data files, in Excel format, listing the genes analyzed in this study, together with the corresponding MMI values, in order of decreasing MMI values. (1) LGG/tumor status, (2) LGG/survival, (3) HNSC/tumor status, (4) HNSC/survival, (5) STES/tumor status, (6) STES/survival. Genes with the MMI values >99.5% MMI CDF (i.e., genes selected for further BN analyses) are shown in bold.

TABLES S7–S12 | Full Bayesian networks, in Word / DOT format, for the six datasets in this study. (1) LGG/tumor status, (2) LGG/survival, (3) HNSC/tumor status, (4) HNSC/survival, (5) STES/tumor status, (6) STES/survival.

DATA SHEETS S1–S6 | Full Bayesian networks, in PDF format, for the six datasets in this study. (1) LGG/tumor status, (2) LGG/survival, (3) HNSC/tumor status, (4) HNSC/survival, (5) STES/tumor status, (6) STES/survival. Designations are as in main text Figure 3.

- Gao, W., Kannan, S., Oh, S., and Viswanath, P. (2018). Estimating mutual information for discrete-continuous mixtures. arXiv [Preprint]. Available online at: https://arxiv.org/abs/1709.06212
- Gevaert, O., De Smet, F., Timmerman, D., Moreau, Y., and De Moor, B. (2006). Predicting the prognosis of breast cancer by integrating clinical and microarray data with Bayesian networks. *Bioinformatics* 22, e184–e190. doi: 10.1093/ bioinformatics/btl230
- Gogoshin, G., Boerwinkle, E., and Rodin, A. S. (2017). New Algorithm and Software (BNOmics) for Inferring and Visualizing Bayesian Networks from Heterogeneous Big Biological and Genetic Data. J. Comput. Biol. 24, 340–356. doi: 10.1089/cmb.2016.0100
- Guyon, I., and Elisseeff, A. (2003). An introduction to variable and feature selection. J. Mach. Learn. Res. 3, 1157–1182.
- Heckerman, D. (1995). *Tutorial on Learning with Bayesian Networks*. Technical Report MSR-TR-95–96. Redmond, WA: Microsoft Research.
- Hou, X., He, X., Wang, K., Hou, N., Fu, J., et al. (2018). Genome-wide networkbased analysis of colorectal cancer identifies novel prognostic factors and an integrative prognostic index. *Cell Physiol. Biochem.* 49, 1703–1716. doi: 10. 1159/000493614
- Huang, T., Yang, J., and Cai, Y. D. (2015). Novel candidate key drivers in the integrative network of genes, microRNAs, methylations, and copy number variations in squamous cell lung carcinoma. *Biomed. Res. Int.* 2015, 358125.
- Jeong, H., Leem, S., Wee, K., and Sohn, K. A. (2015). Integrative network analysis for survival-associated gene-gene interactions across multiple genomic profiles in ovarian cancer. J. Ovarian. Res. 8:42.
- Kaiser, J. L., Bland, C. L., and Klinke, D. J. II (2016). Identifying causal networks linking cancer processes and anti-tumor immunity using Bayesian network inference and metagene constructs. *Biotechnol. Prog.* 32, 470–479. doi: 10.1002/ btpr.2230

- Kataoka, H., Kawaguchi, M., Fukushima, T., and Shimomura, T. (2018). Hepatocyte growth factor activator inhibitors (HAI-1 and HAI-2): emerging key players in epithelial integrity and cancer. *Pathol. Int.* 68, 145–158. doi: 10.1111/pin.12647
- Kawal, P., Chandra, A., Rajkumar, Dhole, T. N., and Ojha, B. (2016). Correlations of polymorphisms in matrix metalloproteinase-1, -2, and -7 promoters to susceptibility to malignant gliomas. *Asian J. Neurosurg.* 11, 160–166.
- Koch, A., Joosten, S. C., Feng, Z., de Ruijter, T. C., Draht, M. X., Melotte, V., et al. (2018). Analysis of DNA methylation in cancer: location revisited. *Nat. Rev. Clin. Oncol.* 15, 459–466. doi: 10.1038/s41571-018-0004-4
- Kohavi, R., and John, G. H. (1997). Wrappers for feature subset selection. Artif. Intell. 97, 273–324. doi: 10.1016/s0004-3702(97)00043-x
- Kraskov, A., Stögbauer, H., and Grassberger, P. (2004). Estimating mutual information. *Phys. Rev. E* 69, 066138.
- Lallet-Daher, H., Wiel, C., Gitenay, D., Navaratnam, N., Augert, A., Calvé, B. L., et al. (2013). Potassium channel KCNA1 modulates oncogene-induced senescence and transformation. *Cancer Res.* 73, 5253–5265. doi: 10.1158/0008-5472.can-12-3690
- Leng, J., Valli, C., and Armstrong, L. (2010). "A wrapper-based feature selection for analysis of large data sets," in *Proceedings of the Conference: 3rd International Conference on Computer and Electrical Engineering (ICCEE 2010)*, Chengdu.
- Liang, A., Zhou, B., and Sun, W. (2017). Integrated genomic characterization of cancer genes in glioma. *Cancer Cell Int.* 17:90.
- Margolin, A. A., Nemenman, I., Basso, K., Wiggins, C., Stolovitzky, G., Favera, R. D., et al. (2006). ARACNE: an algorithm for the reconstruction of gene regulatory networks in a mammalian cellular context. *BMC Bioinformatics* 7(Suppl. 1):S7. doi: 10.1186/1471-2105-7-S1-S7
- Matullo, G., Guarrera, S., Betti, M., Fiorito, G., Ferrante, D., Voglino, F., et al. (2013). Genetic variants associated with increased risk of malignant pleural mesothelioma: a genome-wide association study. *PLoS One* 8:e61253. doi: 10. 1371/journal.pone.0061253
- Park, Y. R., Chun, J. N., So, I., Kim, H. J., Baek, S., Jeon, J. H, et al. (2016). Datadriven Analysis of TRP channels in cancer: linking variation in gene expression to clinical significance. *Cancer Genom. Proteom.* 13, 83–90.
- Phan, J. H., Hoffman, R., Kothari, S., Wu, P. Y., and Wang, M. D. (2016). Integration of multi-modal biomedical data to predict cancer grade and patient survival. *IEEE EMBS Int. Conf. Biomed. Health Inform.* 2016, 577–580.
- Phipps, A. I., Passarelli, M. N., Chan, A. T., Harrison, T. A., Jeon, J., Hutter, C. M., et al. (2016). Common genetic variation and survival after colorectal cancer diagnosis: a genome-wide analysis. *Carcinogenesis* 37, 87–95.
- Pinero, J., Bravo, A., Queralt-Rosinach, N., Gutierrez-Sacristan, A., Deu-Pons, J., Centeno, E., et al. (2017). DisGeNET: a comprehensive platform integrating information on human disease-associated genes and variants. *Nucleic Acids Res.* 45, D833–D839.
- Rajesh, Y., Pal, I., Banik, P., Chakraborty, S., Borkar, S. A., Dey, G., et al. (2017). Insights into molecular therapy of glioma: current challenges and next generation blueprint. *Acta Pharmacol. Sin.* 38, 591–613. doi: 10.1038/aps.2016. 167
- Ramsey, J., Glymour, M., Sanchez-Romero, R., and Glymour, C. (2017). A million variables and more: the Fast Greedy Equivalence Search algorithm for learning high-dimensional graphical causal models, with an application to functional magnetic resonance images. *Int. J. Data Sci. Anal.* 3, 121–129. doi: 10.1007/ s41060-016-0032-z
- Rodin, A. S., Litvinenko, A., Klos, K., Morrison, A. C., Woodage, T., Coresh, J., et al. (2009). Use of wrapper algorithms coupled with a random forests classifier for variable selection in large-scale genomic association studies. *J. Comput. Biol.* 16, 1705–1718. doi: 10.1089/cmb.2008.0037
- Saied, M. H., Marzec, J., Khalid, S., Smith, P., Down, T. A., Rakyan, V. K., et al. (2012). Genome wide analysis of acute myeloid leukemia reveal leukemia specific methylome and subtype specific hypomethylation of repeats. *PLoS One* 7:e33213. doi: 10.1371/journal.pone.0033213
- Schudrowitz, N., Takagi, S., Wessel, G. M., and Yajima, M. (2017). Germline factor DDX4 functions in blood-derived cancer cell phenotypes. *Cancer Sci.* 108, 1612–1619. doi: 10.1111/cas.13299

- Sedgewick, A. J., Buschur, K., Shi, I., Ramsey, J. D., Raghu, V. K., Manatakis, D. V., et al. (2018). Mixed graphical models for integrative causal analysis with application to chronic lung disease diagnosis and prognosis. *Bioinformatics* 35, 1204–1212. doi: 10.1093/bioinformatics/bty769
- Senapati, S., Ho, S. B., Sharma, P., Das, S., Chakraborty, S., Kaur, S., et al. (2010). Expression of intestinal MUC17 membrane-bound mucin in inflammatory and neoplastic diseases of the colon. J. Clin. Pathol. 63, 702–707. doi: 10.1136/jcp. 2010.078717
- Stelzer, G., Rosen, N., Plaschkes, I., Zimmerman, S., Twik, M., Fishilevich, S., et al. (2016). The genecards suite: from gene data mining to disease genome sequence analyses. *Curr. Protoc. Bioinformatics* 54:1.30.1-1.30.33.
- Tanaka, H., Kanda, M., Shimizu, D., Tanaka, C., Kobayashi, D., Hayashi, M., et al. (2017). FAM46C serves as a predictor of hepatic recurrence in patients with resectable gastric cancer. *Ann. Surg. Oncol.* 24, 3438–3445. doi: 10.1245/ s10434-016-5636-y
- Tian, W., Li, Y., Zhang, J., Li, J., and Gao, J. (2018). Combined analysis of DNA methylation and gene expression profiles of osteosarcoma identified several prognosis signatures. *Gene* 650, 7–14. doi: 10.1016/j.gene.2018.01.093
- Tomczak, K., Czerwinska, P., and Wiznerowicz, M. (2015). The Cancer Genome Atlas (TCGA): an immeasurable source of knowledge. *Contemp. Oncol.* 19, A68–A77.
- Wang, W., Baladandayuthapani, V., Holmes, C. C., and Do, K. A. (2013). Integrative network-based Bayesian analysis of diverse genomics data. BMC Bioinformatics 14(Suppl. 13):S8. doi: 10.1186/1471-2105-14-S13-S8
- Wu, S., Law, A., and Whipple, M. E. (2017). A bayesian network model of head and neck squamous cell carcinoma incorporating gene expression profiles. *Stud. Health Technol. Inform.* 245, 634–638.
- Xu, P., Yang, J., Liu, J., Yang, X., Liao, J., Yuan, F., et al. (2018). Identification of glioblastoma gene prognosis modules based on weighted gene co-expression network analysis. BMC Med. Genomics 11:96. doi: 10.1186/s12920-018-0407-1
- Xu, Y., Zhang, J., Yuan, Y., Mitra, R., Muller, P., Ji, Y., et al. (2012). A bayesian graphical model for integrative analysis of TCGA Data. *IEEE Int. Workshop Genomic Signal Process. Stat.* 2012, 135–138.
- Xu, Y., Zhu, Y., Muller, P., Mitra, R., and Ji, Y. (2014). Characterizing Cancer-Specific Networks by Integrating TCGA Data. *Cancer Inform.* 13, 125–131.
- Yoo, B. C., Kim, K. H., Woo, S. M., and Myung, J. K. (2017). Clinical multiomics strategies for the effective cancer management. *J. Proteomics* 188, 97–106. doi: 10.1016/j.jprot.2017.08.010
- Zhang, Q., Burdette, J. E., and Wang, J. P. (2014). Integrative network analysis of TCGA data for ovarian cancer. *BMC Syst Biol* 8:1338. doi: 10.1186/s12918-014-0136-9
- Zhang, X., Branciamore, S., Gogoshin, G., Rodin, A. S., and Riggs, A. D. (2017). Analysis of high-resolution 3D intrachromosomal interactions aided by Bayesian network modeling. *Proc. Natl. Acad. Sci. U.S.A.* 114, E10359–E10368.
- Zhang, C., Cheng, W., Ren, X., Wang, Z., Liu, X., Li, G., et al. (2017). Tumor purity as an underlying key factor in Glioma. *Clin. Cancer Res.* 23, 6279–6291. doi: 10.1158/1078-0432.ccr-16-2598
- Zhu, Y., Xu, Y., Helseth, D. L. Jr., Gulukota, K., Yang, S., Pesce, L. L., et al. (2015). Zodiac: a comprehensive depiction of genetic interactions in cancer by integrating TCGA data. *J. Natl. Cancer Inst.* 107:djv129.

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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