



Genetic Differentiation of Reintroduced Père David's Deer (*Elaphurus davidianus*) Based on Population Genomics Analysis

Shumiao Zhang¹, Chao Li², Yiping Li¹, Qi Chen¹, Defu Hu³, Zhibin Cheng¹, Xiao Wang¹, Yunfang Shan¹, Jiade Bai^{1*} and Gang Liu^{2*}

OPEN ACCESS

Edited by:

Thales Renato Ochotorena De Freitas, Federal University of Rio Grande do Sul, Brazil

Reviewed by:

Fernanda Simões De Almeida, State University of Londrina, Brazil Fernando Alda, University of Tennessee at Chattanooga, United States Zhongqiu Li, Nanjing University, China

*Correspondence:

Gang Liu gangl@caf.ac.cn Jiade Bai baijiade234@aliyun.com

Specialty section:

This article was submitted to Evolutionary and Population Genetics, a section of the journal Frontiers in Genetics

Received: 05 May 2021 Accepted: 10 August 2021 Published: 07 September 2021

Citation:

Zhang S, Li C, Li Y, Chen Q, Hu D, Cheng Z, Wang X, Shan Y, Bai J and Liu G (2021) Genetic Differentiation of Reintroduced Père David's Deer (Elaphurus davidianus) Based on Population Genomics Analysis. Front. Genet. 12:705337. doi: 10.3389/fgene.2021.705337 ¹Beijing Milu Ecological Research Center, Beijing, China, ²Beijing Key Laboratory of Wetland Services and Restoration, Institute of Wetland Research, Chinese Academy of Forestry, Beijing, China, ³College of Ecology and Nature Conservation, Beijing Forestry University, Beijing, China

The reintroduction is an important conservation tool to restore a species in its historically distribution area, but the rate of reintroduction success varies across species or regions due to different reasons. Genetic evaluation is important to the conservation management of reintroduced species. Conservation concerns relate to genetic threats for species with a small population size or severely historically bottle-necked species, such as negative consequences associated with loss of genetic diversity and inbreeding. The last 40 years have seen a rapid increasing of population size for Père David's deer (Elaphurus davidianus), which originated from a limited founder population. However, the genetic structure of reintroduced Père David's deer has not been investigated in terms of population genomics, and it is still not clear about the evolutionary history of Père David's deer and to what extent the inbreeding level is. Conservation genomics methods were used to reconstruct the demographic history of Père David's deer, evaluate genetic diversity, and characterize genetic structure among 18 individuals from the captive, free-ranging and wild populations. The results showed that 1,456,457 single nucleotide polymorphisms (SNPs) were obtained for Père David's deer, and low levels of genome-wide genetic diversity were observed in Père David's deer compared with Red deer (Cervus elaphus) and Sika deer (Cervus nippon). A moderate population genetic differentiation was detected among three populations of Père David's deer, especially between the captive population in Beijing Père David's deer park and the free-ranging population in Jiangsu Dafeng National Nature Reserve. The effective population size of Père David's deer started to decline ~25.8 ka, and the similar levels of three populations' LD reflected the genetic impacts of long-term population bottlenecks in the Père David's deer. The findings of this study could highlight the necessity of individual exchange between different facilities, and genetic management should generally be integrated into conservation planning with other management considerations.

Keywords: Père David's deer, reintroduction, genetic differentiation, demographic history, population genomics

INTRODUCTION

The reintroduction has been increasingly used as an important and effective conservation tool to recover locally extirpated species, and a common tool for restoring lost biodiversity (Earnhardt, 1999; Armstrong and Seddon, 2008; Stewart et al., 2017). The rate of reintroduction success varies across species and differs from regions, and generally poor performance of reintroduction is reported due to different factors (Robert, 2009; Mowry et al., 2015; Robert et al., 2015; Thévenin, 2019). Genetic diversity is important to the viability and long term persistence of reintroduced species (Frankham, 2005; Steiner et al., 2012; Wirtz et al., 2018). In theory, the source of reintroduced populations is limited because of a small population size of founder individuals, which would result in a low level of genetic diversity and limited gene flow (Stewart et al., 2017; Ovenden et al., 2019). This means it is necessary to conduct population genetics after reintroduction, however, such post-reintroduction evaluation is unfortunately inadequate (Moseby et al., 2020), and genetic consequences following wild release still remain unknown for many cases (La Haye et al., 2017).

A key challenge of reintroduction efforts is to translocate individuals in a way that prevents loss of genetic variation, and avoids genetic differentiation relative to source populations (Barbanti et al., 2019). Such a challenge would have intensified, especially for a species that has recovered from a remnant population with historically low levels of genetic variation (Malone et al., 2018). Genetic diversity available for reintroduced populations is determined by the genetic background of the source population, which in turn is heavily impacted by the demographic history (Aspi et al., 2006). Conservation concerns relate to genetic threats for species with a small population size or severely historically bottle-necked species, such as negative consequences associated with loss of genetic diversity and inbreeding (Haig et al., 1990; Ewen et al., 2012). Genetic management plays an important role in improving the performance of reintroduction actions (Theodorou and Couvet, 2010). Several studies have evaluated genetic consequences induced by reintroduction events for some ungulates using traditional molecular markers, for example, Przewalskii's wild horse (Equus ferus; Liu et al., 2014), European bison (Bison bonasus; Olech and Perzanowski, 2002), and Arabian oryx (Oryx leucoryx; El Alqamy et al., 2012), but only very few studies use genomics data to evaluate post-reintroduction genetic consequences (Flesch et al., 2020).

The unbiased genome-wide estimates generally reflect more accurately levels of genetic diversity and inbreeding (Kardos et al., 2016). The recent advances of next-generation sequencing (NGS) makes it feasible to detect genomic variations in many nonmodel species, and a new discipline, conservation genomics, is spawned in the new era, and is now expanding among conservation biologists (Angeloni et al., 2012; Xue et al., 2015; Wright et al., 2020; Hohenlohe et al., 2021). By taking a conservation genomics approach, genome-wide genetic diversity can be estimated in terms of neutral and non-neutral variation (Angeloni et al., 2012; Liu et al., 2020). In addition, genetic structure can be distinguished at a fine scale, even though the population differentiation is subtle (Lucek et al., 2019). Genomics data are useful for reintroduction projects by offering action guidance, including selecting source populations to maximize overall genomic variation, and valuable information for effective post-reintroduction monitoring (Latch, 2020). Meanwhile, the genomics data obtained also are important resources with the power to generate novel insights into the evolution of the focused species (Hohenlohe et al., 2021).

Père David's deer (Elaphurus davidianus), called "Milu" in China, went extinct in the wild as a result of over-hunting and habitat degradation in the late 19th century, but a few remnants were introduced into Europe and preserved safely in captivity in several European zoos. The remaining 18 individuals distributed in Berlin, Paris, and Antwerp were collected by the 11th Duke of Bedford, forming a breeding herd at the Woburn Abbey, England, where only 11 individuals were reported to have the ability to reproduce. By 1945, the global population size of Père David's deer reached 250. Three reintroduction attempts were made in China. The first 20 deer were reintroduced to Beijing, China from Woburn Abbey in 1985, and then 18 in 1987. The second reintroduction population was established in Dafeng, Jiangsu Province, through trans-locating 39 deer from seven British zoos in August 1986. The third trans-located program included 94 deer from Beijing to Shishou, Hubei Province by three batches in 1993, 1994, and 2002. As of 2020, the population size of worldwide Père David's deer is around 10,000, and exceeds 8,000 in China, suggesting a dramatic recovery over the last 40 years although after a severe bottleneck in the history (Figure 1). Père David's deer in the reintroduction sites serves as a multi-species conservation umbrella, and plays a key functional role in wetland ecosystems.

In recent years, wildlife protection laws and regulations have been gradually improved in China, and more suitable habitats are significantly provided. Efforts to rescue and protect endangered species have achieved remarkable results, with Père David's deer as a successful icon in wildlife reintroduction. Père David's deer provides an ideal system to evaluate the genetic consequences of reintroduction. Previous studies using mitochondrial DNA and microsatellites indicated that Père David's deer had extremely low genetic diversity (Zeng et al., 2007, 2013). A comparative genomics study revealed that Père David's deer was characterized with extensive genetic diversity (Zhu et al., 2018). However, genetic diversity and genetic structure have so far not been detected for the reintroduced populations of Père David's deer at the genomewide scale, and it is still not clear to what extent the inbreeding level is.

We used the genomics approach to reconstruct the demographic history of Père David's deer, and to evaluate genetic diversity, and to characterize genetic structure among captive, free-ranging, and wild populations. We also tested the hypothesis that the captive population in Beijing Père David's deer park is differentiated from the populations in Jiangsu Dafeng National Nature Reserve.



MATERIALS AND METHODS

Sample Collection

Blood samples from 10 Père David's deer individuals (four males and six females), classified as the captive population (MC), were collected from Beijing Père David's deer Park, and four samples (two males and two females) as the free-ranging population (MF) in Dafeng National Nature Reserve, Jiangsu Province, and another four samples (two males and two females) as the wild population (MW) in Dafeng National Nature Reserve, Jiangsu Province. Here, the free-ranging population indicates the animals live in a quite large fence, but can feed and breed naturally there. In contrast, the wild population survives and reproduces in the wild absolutely. The free-ranging and wild population was separated physically despite being in Dafeng National Nature Reserve. Blood samples were collected from two individuals of Red deer (Cervus elaphus) and two individuals of Sika deer (Cervus nippon) in Beijing Père David's deer Park. Blood samples were stored in vacuum blood collection tube (Heparin anticoagulation). All samples were collected in 2018 and 2019 (Supplementary Table 1).

Sequencing and Single Nucleotide Polymorphism Calling

Genomic DNA was extracted using QIAamp DNA Blood Mini Kit (Qiagen, Germany). The libraries were prepared as described in stLRF kit, and whole-genome resequencing was performed based on the BGISEQ-500 platform (BGI). The adapter and low quality reads were removed by SOAPnuke (Chen et al., 2017). Clean reads were aligned against the reference genome of Père David's deer (Project ID: PRJNA391565, downloaded from NCBI; Zhang et al., 2017) using Burrows-Wheeler Aligner (BWA; Li and Durbin, 2010), and then read alignments were sorted using SAMTools with the following parameters: "-q 1 -C 50 -g -t DP, SP, DP4 -I -d 250 -L 250 -m 2 -p," and duplicate reads were marked by Picard.

Multisample single nucleotide polymorphism (SNP) genotyping were carried out as follows. Variants calling was conducted using HaplotypeCaller and GenotypeGVCFs in GATK4 (Van der Auwera et al., 2013), and in order to ensure the identification accuracy, SNPs were removed if any of the following parameter was met: QD<2, MQ<30, MQRankSum<-12.5, FS>200, ReadPosRankSum<20, QUAL<30.0, and AN<40. The GVCF file from each sample was generated using HaplotypeCaller, and then GenotypeGVCFs were used to merge separate GVCF files with the aim to improve the sensitivity of mutation detection.

Genetic Diversity Estimation

In order to estimate the genetic diversity of reintroduced populations of Père David's deer, VCFtools (Danecek et al., 2011) was used to calculate the nucleotide diversity π at the whole-genome scale for each population. The Watterson's estimator (w) of each population was calculated using VariScan v2.0.3 (Hutter et al., 2006). Genome-wide Tajima's *D* values for each population were calculated using VCFtools. A widely used approach for detecting selection is to apply a neutrality test statistic based on allele frequencies, with Tajima's D being the most frequently one (Korneliussen et al., 2013).

Population Structure Analysis

The population structure was mainly revealed through three different methods, including principal component analysis (PCA), phylogenetic tree (NJ tree), and ancestral component analysis (fastSTRUCTURE). Based on the filtered set of 1,456,457 SNPs, we used Genome-wide Complex Trait Analysis (GCTA; Yang et al., 2011) to perform principal component analysis (PCA), filter a few principal components through a dimensionality reduction algorithm, and use these principal components. Components can be used to explain the differences between individuals to the greatest extent, and at the same time, the clustering relationship of each sample based on the distribution of PCs can be estimated. A phylogenetic tree was constructed using Neighbor-joining (NJ) in Tree Building guided by Species Tree (TreeBeST; Price et al., 2010) in order to study the genetic distance relationship between individuals, with a bootstrap value of 1,000. The population genetic structure was inferred from genomics data using STRUCTURE with default settings (Raj et al., 2014). The most likely value of K was identified from 20 independent runs for each value of K ranging from 2 to 8, with 100 bootstraps. The posterior probability K was estimated, and the log likelihood was used to choose the optimal K (Evanno et al., 2005). The web-based Structure Harvester (Earl and vonHoldt, 2012) was used to assess and visualize likelihood values across multiple values of K. The genetic differentiation index, F_{ST} was usually used to detect the genetic composition variance between all populations, and we used VCFtools to calculate these values. The level of genetic differentiation can be defined as low ($F_{ST} < 0.05$), moderate $(0.05 < F_{ST} < 0.15)$, high $(0.15 < F_{ST} < 0.25)$, and extremely high (F_{ST}>0.25; Wright, 1978).

Demographic History and Inbreeding Pattern

Multiple sequential Markovian coalescence (MSMC) model (Schiffels and Durbin, 2014) can be used to reconstruct each population's demographic history based on genomic information. MSMC breaks through the limitation that PSMC can only analyze one sample at a time. In addition, MSMC can integrate and analyze the nearest common ancestor time between multiple allele sequences at the same time, thereby improving the accuracy and efficiency of effective population size (Ne) prediction (Schiffels and Durbin, 2014). The parameters used were as follows: "-t 6 -p 1*2+15*1+1*2." A mutation rate (μ) of 1.5×10^{-8} mutations per site per generation for the three Cervidae species in this study was according to the reference (Chen et al., 2019). The generation time was estimated to be 6 years based on the report of mammal generation length (Pacifici et al., 2013). Detection of linkage disequilibrium (LD) patterns is necessary to infer population historical changes as well as inbreeding events in the species evolutionary history (Karimi et al., 2020). To assess the genomic extent of inbreeding inPère David's deer, genome-wide LD was estimated for the three populations using PopLDdecay (Zhang et al., 2019). To assess the LD of Père David's deer, the correlation coefficient (R^2) between any two loci from each population was calculated using vcftools v0.1.14, with following parameters: "--ld –window -bp 500,000 –geno -r2," and average R^2 values were calculated for pairwise SNPs by keeping the same distance.

RESULTS

Genetic Diversity and Genetic Drift

A total of 12,744 G clean reads were obtained among 22 samples from Père David's deer, Red deer, and Sika deer (Supplementary Table 1). The results showed that the average genome coverage rate was 98.77%, and the average sequencing depth was 19.93X, with an average mismatch rate of 0.90% (Supplementary Table 2). After filtering, an average of 485.76 million clean reads remained for each Père David's deer sample, with a clean data ratio of 99.31%, of which 99.12% reads (98.18-99.67%) were mapped to the Père David's deer reference genome, and the mean GC content was 43.33%. Totally, 1,456,457 SNPs were obtained for all the datasets for further analysis, which had 33.92% homozygotes and 66.08% heterozygotes, meaning a heterozygosity rate of 0.38 kilobase in the Père per pair David's deer (Supplementary Figure 1). The heterozygote percentage of the free-ranging and wild populations of Père David's deer $(70.47 \pm 0.81\%)$ was significantly higher than the captive population (64.21 \pm 0.76%; p < 0.01). Across all 18 samples from Père David's deer, 367.266 indels were identified, with 54.61% insertions and 45.39% deletions, but neither the percentage of insertion nor deletion showed significant difference across Père David's deer populations (p > 0.05).

Whole genome variation based on re-sequencing revealed that free-ranging Père David's deer (MF) had the highest genomic diversity (π =4.03×10⁻⁵, w=3.58×10⁻⁵), whereas captive individuals (MC) had the lowest genetic diversity (π =4.03×10⁻⁵, w=2.99×10⁻⁵). Père David's deer had lower genetic diversity than Red deer (π =1.04×10⁻⁴, w=1.04×10⁻⁴) and Sika deer (π =1.47×10⁻⁴, w=1.45×10⁻⁴; **Table 1**). The genome-wide Tajima's *D* values were positive in three populations of Père David's deerr (**Table 1**), indicating that Père David's deer experienced balancing selection and population contraction in the history.

Population Genetic Structure

The captive population of Père David's deer from Beijing Milu Park showed moderate population genomic differentiation

TABLE 1 The genetic diversity index for three populations of Père David's deer
and its related species.

	π	w	Tajima's D	
MC	0.0000403	0.0000299	0.4908771	
MF	0.0000409	0.0000358	0.2367667	
MW	0.00004	0.0000351	0.2300057	
RD	0.0001041	0.0001037	0.0074172	
SK	0.0001466	0.0001452	0.0213304	

MC, captive Père David's deer; MF, free-ranging Père David's deer; MW, wild Père David's deer; RD, Red deer; and SK, Sika deer.

from free-ranging and wild populations from Jiangsu based on both STRUCTURE and PCA analysis. Within the populations from Jiangsu, free-ranging and wild populations were further divided into two genetic subgroups, but population admixture also existed (Figure 2). The individuals from MF population admixed with MW population, and the individuals from MW population also clustered into MF population. This genetic assignment implied relatively frequent gene flow between individuals within the reintroduced population in Jiangsu. At a cluster value of two (K=2; Supplementary Figure 2), STRUCTURE performed well to distinguish among three populations of Père David's deer, suggesting the reintroduced populations had separated into two genetic structures (Figure 2A). The NJ phylogenetic tree based on pairwise SNP differences also revealed separate genetic clusters among captive, free-ranging, and wild populations of the reintroduced populations (**Figure 2A**). The average F_{ST} value between the captive and free-ranging was 0.073906, and 0.07302 between the captive and wild populations, suggesting a moderate genetic differentiation (**Table 2**). The average F_{ST} value was negative between the free-ranging and wild population, indicating no genetic differentiation, which was consistent with the results generated by PCA (**Figure 2B**) and phylogenetic tree.

Demographic History

The results from the MSMC analysis showed that the effective population size of Père David's deer started to decline ~25.8 ka. The three populations of Père David's deer had a similar long-term demographic history. About 2.58×10^4 years ago, the effective population sizes for Père David's deer were always larger than those for Red deer and Sika deer, but over the past 1.25×10^4 years, it showed an opposite trend (**Figure 3**).



Component Analysis (PCA) plot.

Inbreeding Pattern

Genome-wide LD analysis demonstrated that the average pairwise distance for the LD to decay until $R^2=0.4$ was >900 kb for Père David's deer (**Figure 4**). The three populations of Père David's deer had similar levels of LD, to some extent reflecting the genetic impacts of long-term population bottlenecks in the Père David's deer.

DISCUSSION

°0

107

Effective population size 10⁶

105

L0⁴

RD

мw

MF

This is, to our knowledge, the first study to evaluate the genomic consequences of reintroduced Père David's deer using the approach of population genomics, covering captive, free-ranging, and wild populations. The genome-wide genetic diversity of Père David's deer was lower than that of Red deer and Sika deer. This result was consistent with other earlier studies using mitochondrial DNA and nuclear DNA (Zeng et al., 2007), and microsatellite (Wu et al., 2008; Zhang et al., 2010). In addition,

TABLE 2 The genetic differentiation index between populations base on F _s							
	SK	RD	MW	MF	МС		
МС	0.910565	0.91975	0.07302	0.073906	0		
MF	0.888112	0.904536	-0.00976	0			
MW	0.889444	0.905702	0				
RD	0.586584	0					
SK	0						

MC, the captive population; MF, the free-ranging population; MW, the wild population; RD, Red deer; and SK, Sika deer.

MC

SK

 10^{4}

the captive population had the lowest genomic diversity, while the highest was revealed in the free-ranging population. Although 18 individuals were considered as the founder source of all current populations of Père David's deer around the world, 13 of them contributed to the breeding from the start at the Woburn Abbey, England. Père David's deer underwent drastic declines in population size, during which population decline is always accompanied by loss of genomic diversity due to





 10^{6}



Time(Years ago)

10⁵

genetic drift and inbreeding. Furthermore, the 12th Duke of Bedford even suspected that all current populations of Père David's deer were descended from a single male (Sowerby, 1949). Up to now, unfortunately almost nothing is known about the composition and structure of the Père David's deer Y chromosome, and further knowledge on the Y chromosome in future studies may be crucial for adding a new dimension to our understanding of the low level of genomic diversity. Taking the Przewalski's horse as an example, only two haplotypes of Y chromosome were kept after experiencing a severe historically bottleneck (Kavar and Dovc, 2008), and the limited Y chromosome lineages partially contribute to homozygous variations of Przewalski's horse (Do et al., 2014).

To be noticed, the genomic diversity in the captive population was lower than that in the free-ranging population of Père David's deer (Table 1), which was contrary to the results based on microsatellite (Zeng et al., 2007). Such inconsistent results may be attributed to different methods applied in different studies. Compared with neutral loci (e.g., microsatellite), the genomic approach can increase the power and accuracy of estimating a set of key parameters in conservation biology (Allendorf et al., 2010; Fan et al., 2018). Conservation breeding actions play an important role to protect species by controlling threatening factors and providing reintroduction source (Ebenhard, 1995). However, a key issue associated with captive breeding is the limited gene pool, leading to the accumulation of inbreeding from the founder generation, due to small population sizes and constrained migration (Alroy, 2015; Mulvena et al., 2020). The origin of Père David's deer differs between Beijing and Jiangsu (Bai et al., 2012; Cao, 2021), which means that genetic composition varies between regions. In addition, small populations are expected to suffer from loss of genetic diversity through genetic drift and reduced fitness because of inbreeding (Kvie et al., 2019). As of 2000, the captive population size in Beijing has not exceeded 200 individuals, and in contrast, the population size in Jiangsu has nearly reached 6,000 (Figure 1).

A moderate genetic differentiation was represented among captive, free-ranging, and wild populations by both STRUCTURE and PCA analysis (Figure 2). The captive and free-ranging populations showed the highest level of genetic differentiation (Table 2), which was supported by the microsatellite based study (Zeng et al., 2007). The population in Beijing was reintroduced from the Woburn Abbey, but the population in Jiangsu was established with founders from five zoo populations (Jiang and Ding, 2011). The different lineage would be the main reason resulting in different genetic structure. In the past 40 years, 520 Père David's deer in Beijing Milu Park have been released into 40 sites through 53 translocation events covering 20 provinces around China, but unfortunately, few individuals were translocated to Dafeng, Jiangsu. The lack of translocation between Beijing and Jiangsu may hinder the gene flow, and further drive genetic differentiation.

The demographic history analysis based on MSMC2 showed that the effective population size of Père David's deer started to decline ~25.8 ka (**Figure 3**), which coincides with the inference according to PSMC (Zhu et al., 2018). During late

Pleistocene, large mammal declines became much more severe than previously expected due to the interaction effects of climate change and human activities, although the impact varies across continents or species (Barnosky et al., 2004; Chen et al., 2019). With the human population expanding dramatically (Li and Durbin, 2011), the population decline of Père David's deer might be at least partially attributable to human activities (e.g., human hunting), which was further supported by the fossil evidence (Dong et al., 2019). This is further supported by new evidence from large-scale ruminant genome sequencing, given that the demographic pattern was species-specific because of variable habitat types or feeding types (Chen et al., 2019). With the population size decreasing, inbreeding accumulated during the evolutionary history of Père David's deer (Zhu et al., 2018). The LD pattern is a useful indicator for estimating inbreeding at the individual level, and generally LD rapidly decays to very low levels in populations with low inbreeding level (Wang et al., 2021). Père David's deer exhibited extremely high LD (Figure 4), which implied that a prolonged population decline might have caused an increased overall burden of inbreeding.

CONSERVATION IMPLICATIONS

This study evaluates genomic consequences of post-reintroduction of Père David's deer using the conservation genomics approach, and the results would contribute to inform future conservation management. There are many factors influencing species reintroduction success, among which genetic is the most important (Flesch et al., 2020). The genetic diversity of an endangered species is one of the main parameters that can directly reflect the evolutionary potential. Genetic evaluation can provide decision-making reference for the relevant government departments to formulate species protection strategies and plans. Genomic evaluation of reintroduced populations can help address the uncertainties in reintroduction projects. According to IUCN guidelines, genetic management should generally be integrated into conservation planning with other management considerations (Ralls et al., 2018).

In the future, the genetic evaluation should cover more populations and sample more individuals, for example, the population size of Père David's deer has reached 1,600 in Shishou, Hubei province. The extremely low genetic diversity has always been a warning to humans that population genetic health of Père David's deer is still relatively fragile, and how to maintain the current level and prevent further loss still remains a concern in conservation management. In the specific implementation of the reintroduction, there are still some geopolitical factors that may affect the source selection and the selection of reintroduction sites. It is recommended to conduct individual exchanges among different facilities, so as to enhance gene flow and reduce inbreeding. For example, the captive individuals in Beijing should be selected to be released to the wild in Jiangsu Dafeng National Nature Reserve. Before releasing, the genetic background of the source individuals should be monitored to ensure sufficient gene flow. In addition,

the samples of Père David's deer are important resources for scientific research, especially the ones who exhibited abnormal phenotypes in the population, because they can be used to study the inbreeding depression and genetic changes over time. So, we suggest building a sample library for reintroduced species.

DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found at: https://www.ncbi.nlm. nih.gov/, Project ID: PRJNA756107.

ETHICS STATEMENT

The animal study was reviewed and approved by China Wildlife Conservation Association.

AUTHOR CONTRIBUTIONS

SZ and GL conceived the study. YL, QC, ZC, XW, and YS performed the experiments. GL and CL performed the

REFERENCES

- Allendorf, F. W., Hohenlohe, P. A., and Luikart, G. (2010). Genomics and the future of conservation genetics. *Nat. Rev. Genet.* 11, 697–709. doi: 10.1038/ nrg2844
- Alroy, J. (2015). Limits to captive breeding of mammals in zoos. Conserv. Biol. 29, 926–931. doi: 10.1111/cobi.12471
- Angeloni, F., Wagemaker, N., Vergeer, P., and Ouborg, J. (2012). Genomic toolboxes for conservation biologists. *Evol. Appl.* 5, 130–143. doi: 10.1111/j. 1752-4571.2011.00217.x
- Armstrong, D. P., and Seddon, P. J. (2008). Directions in reintroduction biology. Trends Ecol. Evol. 23, 20–25. doi: 10.1016/j.tree.2007.10.003
- Aspi, J., Roininen, E., Ruokonen, M., Kojola, I., and Vilà, C. (2006). Genetic diversity, population structure, effective population size and demographic history of the Finnish wolf population. *Mol. Ecol.* 15, 1561–1576. doi: 10.1111/j.1365-294X.2006.02877.x
- Bai, J., Zhang, L., Zhong, Z., and Dong, J. (2012). Research progress on the development status of Chinese *Elaphurus davidianus* population. *China Ani*. *Husb. Vet. Med.* 39, 225–230. doi: 10.3969/j.issn.1671-7236.2012.11.052
- Barbanti, A., Turmo, M., Blumenthal, J., Boyle, J., Broderick, A., Collyer, L., et al. (2019). Founding in action: genetic assessment of new populations from the same reintroduction program. *Front. Mar. Sci.* 6:168. doi: 10.3389/ conf.fmars.2019.08.00168
- Barnosky, A. D., Koch, P. L., Feranec, R. S., Wing, S. L., and Shabel, A. B. (2004). Assessing the causes of late Pleistocene extinctions on the continents. *Science* 306, 70–75. doi: 10.1126/science.1101476
- Cao, M. (2021). The 35th anniversary of the reintroduction of Milu deer to China: history, population status, achievements and challenges. *Biodivers. Science* 29, 160–166. doi: 10.17520/biods.2021018
- Chen, Y., Chen, Y., Shi, C., Huang, Z., Zhang, Y., Li, S., et al. (2017). SOAPnuke: a MapReduce acceleration-supported software for integrated quality control and preprocessing of high-throughput sequencing data. *GigaScience* 7, 1–6. doi: 10.1093/gigascience/gix120
- Chen, L., Qin, Q., Jiang, Y., Wang, K., Lin, Z., Li, Z., et al. (2019). Large-scale ruminant genome sequencing provides insights into their evolution and distinct traits. *Science* 364:eaav6202. doi: 10.1126/science.aav6202

bioinformatics analyses and data analysis. DH and JB revised the manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

This study was funded by Innovation Team Program of Beijing Academy of Science and Technology "Research on Genetic Resources Conservation and Fine Management of Milu" [IG201805N/C1 (newly added)].

ACKNOWLEDGMENTS

The authors would like to thank Beijing Père David's deer Park and Jiangsu Dafeng National Nature Reserve for helping with samples collection.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fgene.2021.705337/ full#supplementary-material

- Danecek, P., Auton, A., Abecasis, G., Albers, C. A., Banks, E., DePristo, M. A., et al. (2011). The variant call format and VCFtools. *Bioinformatics* 27, 2156–2158. doi: 10.1093/bioinformatics/btr330
- Do, K.-T., Kong, H.-S., Lee, J.-H., Lee, H.-K., Cho, B.-W., Kim, H.-S., et al. (2014). Genomic characterization of the Przewalski's horse inhabiting Mongolian steppe by whole genome re-sequencing. *Livest. Sci.* 167, 86–91. doi: 10.1016/j.livsci.2014.06.020
- Dong, W., Wei, Q., Bai, W., Zhang, L., Liu, W., Chen, Z., et al. (2019). New material of the Early Pleistocene Elaphurus (Artiodactyla, Mammalia) from North China and discussion on taxonomy of Elaphurus. *Quat. Int.* 519, 113–121. doi: 10.1016/j.quaint.2018.05.015
- Earl, D. A., and vonHoldt, B. M. (2012). STRUCTURE HARVESTER: a website and program for visualizing STRUCTURE output and implementing the Evanno method. *Conserv. Genet. Resour.* 4, 359–361. doi: 10.1007/ s12686-011-9548-7
- Earnhardt, J. M. (1999). Reintroduction programmes: genetic trade-offs for populations. Anim. Conserv. 2, 279–286. doi: 10.1111/j.1469-1795.1999. tb00074.x
- Ebenhard, T. (1995). Conservation breeding as a tool for saving animal species from extinction. *Trends Ecol. Evol.* 10, 438–443. doi: 10.1016/S0169-5347(00)89176-4
- El Alqamy, H., Senn, H., Roberts, M.-F., McEwing, R., and Ogden, R. (2012). Genetic assessment of the Arabian oryx founder population in the emirate of Abu Dhabi, UAE: an example of evaluating unmanaged captive stocks for reintroduction. *Conserv. Genet.* 13, 79–88. doi: 10.1007/s10592-011-0264-3
- Evanno, G., Regnaut, S., and Goudet, J. (2005). Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. *Mol. Ecol.* 14, 2611–2620. doi: 10.1111/j.1365-294X.2005.02553.x
- Ewen, J. G., Armstrong, D. P., Parker, K. A., and Seddon, P. J. (2012). Reintroduction Biology: Integrating Science and Management. New Jersey: John Wiley & Sons.
- Fan, H., Hu, Y., Wu, Q., Nie, Y., Yan, L., and Wei, F. (2018). Conservation genetics and genomics of threatened vertebrates in China. J. Genet. Genomics 45, 593–601. doi: 10.1016/j.jgg.2018.09.005
- Flesch, E. P., Graves, T. A., Thomson, J. M., Proffitt, K. M., White, P. J., Stephenson, T. R., et al. (2020). Evaluating wildlife translocations using genomics: a bighorn sheep case study. *Ecol. Evol.* 10, 13687–13704. doi: 10.1002/ece3.6942

- Frankham, R. (2005). Genetics and extinction. *Biol. Conserv.* 126, 131–140. doi: 10.1016/j.biocon.2005.05.002
- Haig, S. M., Ballou, J. D., and Derrickson, S. R. (1990). Management options for preserving genetic diversity: reintroduction of Guam rails to the wild. *Conserv. Biol.* 4, 290–300. doi: 10.1111/j.1523-1739.1990.tb00291.x
- Hohenlohe, P. A., Funk, W. C., and Rajora, O. P. (2021). Population genomics for wildlife conservation and management. *Mol. Ecol.* 30, 62–82. doi: 10.1111/ mec.15720
- Hutter, S., Vilella, A. J., and Rozas, J. (2006). Genome-wide DNA polymorphism analyses using VariScan. BMC Bioinform. 7:409. doi: 10.1186/1471-2105-7-409
- Jiang, Z., and Ding, Y. (2011). Dafeng Père David's Deer and Biodiversity. Beijing: China Forestry Publishing House.
- Kardos, M., Taylor, H. R., Ellegren, H., Luikart, G., and Allendorf, F. W. (2016). Genomics advances the study of inbreeding depression in the wild. *Evol. Appl.* 9, 1205–1218. doi: 10.1111/eva.12414
- Karimi, K., Farid, A. H., Sargolzaei, M., Myles, S., and Miar, Y. (2020). Linkage disequilibrium, effective population size and genomic inbreeding rates in American mink using genotyping-by-sequencing data. *Front. Genet.* 11:223. doi: 10.3389/fgene.2020.00223
- Kavar, T., and Dovc, P. (2008). Domestication of the horse: genetic relationships between domestic and wild horses. *Livest. Sci.* 116, 1–14. doi: 10.1016/j. livsci.2008.03.002
- Korneliussen, T. S., Moltke, I., Albrechtsen, A., and Nielsen, R. (2013). Calculation of Tajima's D and other neutrality test statistics from low depth next-generation sequencing data. *BMC Bioinform.* 14:289. doi: 10.1186/1471-2105-14-289
- Kvie, K. S., Heggenes, J., Bardsen, B.-J., and Roed, K. H. (2019). Recent largescale landscape changes, genetic drift and reintroductions characterize the genetic structure of Norwegian wild reindeer. *Conserv. Genet.* 20, 1405–1419. doi: 10.1007/s10592-019-01225-w
- La Haye, M. J. J., Reiners, T. E., Raedts, R., Verbist, V., and Koelewijn, H. P. (2017). Genetic monitoring to evaluate reintroduction attempts of a highly endangered rodent. *Conserv. Genet.* 18, 877–892. doi: 10.1007/s10592-017-0940-z
- Latch, E. K. (2020). Integrating genomics into conservation management. *Mol. Ecol. Resour.* 20, 1455–1457. doi: 10.1111/1755-0998.13188
- Li, H., and Durbin, R. (2010). Fast and accurate long-read alignment with burrows-wheeler transform. *Bioinformatics* 26, 589–595. doi: 10.1093/ bioinformatics/btp698
- Li, H., and Durbin, R. (2011). Inference of human population history from individual whole-genome sequences. *Nature* 475, 493–U484. doi: 10.1038/ nature10231
- Liu, G., Shafer, A. B. A., Zimmermann, W., Hu, D., Wang, W., Chu, H., et al. (2014). Evaluating the reintroduction project of Przewalski's horse in China using genetic and pedigree data. *Biol. Conserv.* 171, 288–298. doi: 10.1016/j. biocon.2013.11.022
- Liu, D., Zhang, L., Wang, J., and Ma, Y. (2020). Conservation genomics of a threatened Rhododendron: contrasting patterns of population structure revealed from neutral and selected SNPs. *Front. Genet.* 11:757. doi: 10.3389/ fgene.2020.00757
- Lucek, K., Gompert, Z., and Nosil, P. (2019). The role of structural genomic variants in population differentiation and ecotype formation in Timema cristinae walking sticks. *Mol. Ecol.* 28, 1224–1237. doi: 10.1111/mec.15016
- Malone, E. W., Perkin, J. S., Leckie, B. M., Kulp, M. A., Hurt, C. R., and Walker, D. M. (2018). Which species, how many, and from where: integrating habitat suitability, population genomics, and abundance estimates into species reintroduction planning. *Glob. Chang. Biol.* 24, 3729–3748. doi: 10.1111/ gcb.14126
- Moseby, K. E., Blumstein, D. T., Letnic, M., and West, R. (2020). Choice or opportunity: are post-release social groupings influenced by familiarity or reintroduction protocols? Oryx 54, 215–221. doi: 10.1017/S0030605318000054
- Mowry, R. A., Schneider, T. M., Latch, E. K., Gompper, M. E., Beringer, J., and Eggert, L. S. (2015). Genetics and the successful reintroduction of the Missouri river otter. Anim. Conserv. 18, 196–206. doi: 10.1111/acv.12159
- Mulvena, S. R., Pierson, J. C., Farquharson, K. A., McLennan, E. A., Hogg, C. J., and Grueber, C. E. (2020). Investigating inbreeding in a free-ranging, captive population of an Australian marsupial. *Conserv. Genet.* 21, 665–675. doi: 10.1007/s10592-020-01278-2
- Olech, W., and Perzanowski, K. (2002). A genetic background for reintroduction program of the European bison (*Bison bonasus*) in the Carpathians. *Biol. Conserv.* 108, 221–228. doi: 10.1016/S0006-3207(02)00108-8

- Ovenden, T. S., Palmer, S. C. F., Travis, J. M. J., and Healey, J. R. (2019). Improving reintroduction success in large carnivores through individualbased modelling: how to reintroduce Eurasian lynx (Lynx lynx) to Scotland. *Biol. Conserv.* 234, 140–153. doi: 10.1016/j.biocon.2019.03.035
- Pacifici, M., Santini, L., Di Marco, M., Baisero, D., Francucci, L., Marasini, G. G., et al. (2013). Generation length for mammals. *Nat. Conserv.-Bulgaria* 5, 87–94. doi: 10.3897/natureconservation.5.5734
- Price, M. N., Dehal, P. S., and Arkin, A. P. (2010). FastTree 2 approximately maximum-likelihood trees for large alignments. *PLoS One* 5:e9490. doi: 10.1371/journal.pone.0009490
- Raj, A., Stephens, M., and Pritchard, J. K. (2014). fastSTRUCTURE: variational inference of population structure in large SNP data sets. *Genetics* 197:573. doi: 10.1534/genetics.114.164350
- Ralls, K., Ballou, J. D., Dudash, M. R., Eldridge, M. D. B., Fenster, C. B., Lacy, R. C., et al. (2018). Call for a paradigm shift in the Genetic Management of Fragmented Populations. *Conserv. Lett.* 11, 1–6. doi: 10.1111/conl.12412
- Robert, A. (2009). Captive breeding genetics and reintroduction success. Biol. Conserv. 142, 2915–2922. doi: 10.1016/j.biocon.2009.07.016
- Robert, A., Colas, B., Guigon, I., Kerbiriou, C., Mihoub, J.-B., Saint-Jalme, M., et al. (2015). Defining reintroduction success using IUCN criteria for threatened species: a demographic assessment. *Anim. Conserv.* 18, 397–406. doi: 10.1111/acv.12188
- Schiffels, S., and Durbin, R. (2014). Inferring human population size and separation history from multiple genome sequences. *Nat. Genet.* 46, 919–925. doi: 10.1038/ng.3015
- Sowerby, A. (1949). Notes on the original habitat of father David's deer.Musee Heude. Notes de Mammal. 4, 3-19.
- Steiner, C., Putnam, A., Hoeck, P., and Ryder, O. (2012). Conservation genomics of threatened animal species. Annu. Rev. Anim. Biosci. 1, 261–281. doi: 10.1146/annurev-animal-031412-103636
- Stewart, F. E. C., Volpe, J. P., Taylor, J. S., Bowman, J., Thomas, P. J., Pybus, M. J., et al. (2017). Distinguishing reintroduction from recolonization with genetic testing. *Biol. Conserv.* 214, 242–249. doi: 10.1016/j.biocon.2017.08.004
- Theodorou, K., and Couvet, D. (2010). Genetic management of captive populations: the advantages of circular mating. *Conserv. Genet.* 11, 2289–2297. doi: 10.1007/s10592-010-0114-8
- Thévenin, C. (2019). Reintroduction efficiency: a stepping stone approach to reintroduction success? Anim. Conserv. 22, 116–117. doi: 10.1111/acv.12501
- Van der Auwera, G. A., Carneiro, M. O., Hartl, C., Poplin, R., Del Angel, G., Levy-Moonshine, A., et al. (2013). From FastQ data to high confidence variant calls: the genome analysis toolkit best practices pipeline. *Curr. Protoc. Bioinformatics* 43, 11.10.11–11.10.33. doi: 10.1002/0471250953. bi1110s43
- Wang, P., Burley, J. T., Liu, Y., Chang, J., Chen, D., Lu, Q., et al. (2021). Genomic consequences of long-term population decline in brown eared pheasant. *Mol. Biol. Evol.* 38, 263–273. doi: 10.1093/molbev/msaa213
- Wirtz, S., Böhm, C., Fritz, J., Kotrschal, K., Veith, M., and Hochkirch, A. (2018). Optimizing the genetic management of reintroduction projects: genetic population structure of the captive Northern Bald Ibis population. *Conserv. Genet.* 19, 853–864. doi: 10.1007/s10592-018-1059-6
- Wright, S. (1978). Evolution and the Genetics of Populations. Chicago, University of Chicago Press.
- Wright, B. R., Farquharson, K. A., McLennan, E. A., Belov, K., Hogg, C. J., and Grueber, C. E. (2020). A demonstration of conservation genomics for threatened species management. *Mol. Ecol. Resour.* 20, 1526–1541. doi: 10.1111/1755-0998.13211
- Wu, H.-L., Ni, X.-W., Zhang, L.-Y., Xia, J.-S., Zhong, Z.-Y., Zhu, G.-P., et al. (2008). Eighteen novel polymorphic microsatellite loci developed from the Père David's deer (*Elaphurus davidianus*). Conserv. Genet. 9, 1679–1682. doi: 10.1007/s10592-008-9523-3
- Xue, Y., Prado-Martinez, J., Sudmant, P., Narasimhan, V., Ayub, Q., Szpak, M., et al. (2015). Mountain gorilla genomes reveal the impact of long-term population decline and inbreeding. *Science* 348, 242–245. doi: 10.1126/science. aaa3952
- Yang, J., Lee, S. H., Goddard, M. E., and Visscher, P. M. (2011). GCTA: A tool for genome-wide complex trait analysis. Am. J. Hum. Genet. 88, 76–82. doi: 10.1016/j.ajhg.2010.11.011
- Zeng, Y., Jiang, Z., and Li, C. (2007). Genetic variability in relocated Père David's deer (*Elaphurus davidianus*) populations—implications to

reintroduction program. Conserv. Genet. 8, 1051-1059. doi: 10.1007/s10592-006-9256-0

- Zeng, Y., Li, C., Zhang, L., Zhong, Z., and Jiang, Z. (2013). No correlation between neonatal fitness and heterozygosity in a reintroduced population of Père David's deer. *Curr. Zool.* 59, 249–256. doi: 10.1093/czoolo/59.2.249
- Zhang, C., Chen, L., Zhou, Y., Wang, K., Chemnick, L. G., Ryder, O. A., et al. (2017). Draft genome of the milu (*Elaphurus davidianus*). *GigaScience* 7:gix130. doi: 10.1093/gigascience/gix130
- Zhang, C., Dong, S.-S., Xu, J.-Y., He, W.-M., and Yang, T.-L. (2019). PopLDdecay: a fast and effective tool for linkage disequilibrium decay analysis based on variant call format files. *Bioinformatics* 35, 1786–1788. doi: 10.1093/bioinformatics/bty875
- Zhang, L., Wu, H., Zhong, Z., and Zhang, S. (2010). Microsatellite polymorphisms and population genetic structure of Elaphurus davidianus in Beijing Milu Park, Sichuan. J. Zool. 29, 505–508.
- Zhu, L., Deng, C., Zhao, X., Ding, J., Huang, H., Zhu, S., et al. (2018). Endangered Père David's deer genome provides insights into population recovering. *Evol. Appl.* 11, 2040–2053. doi: 10.1111/eva.12705

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Zhang, Li, Li, Chen, Hu, Cheng, Wang, Shan, Bai and Liu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.