

Investigating *CENPW* as a Novel Biomarker Correlated With the Development and Poor Prognosis of Breast Carcinoma

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Wang L, Wang H, Yang C, Wu Y, Lei G, Yu Y, Gao Y, Du J, Tong X, Zhou F, Li Y and Wang Y (2022) Investigating CENPW as a Novel Biomarker Correlated With the Development and Poor Prognosis of Breast Carcinoma. Front. Genet. 13:900111. doi: 10.3389/fgene.2022.900111 Breast invasive carcinoma (BRCA) is a carcinoma with a fairly high incidence, and the therapeutic schedules are generally surgery and chemotherapy. However, chemotherapeutic drugs tend to produce serious toxic side effects, which lead to the cessation of treatment. Therefore, it is imperative to develop treatment strategies that are more effective and have fewer side effects at the genetic level. Centromeric protein W (CENPW) is an oncogene that plays an important part in nucleosome assembly. To date, no studies have reported the prognostic significance of CENPW in breast carcinoma. In this study, we verified that CENPW expression is up-regulated in breast carcinoma and positively associated with the level of immune cell infiltration. The clinicopathological characteristics further suggest that CENPW expression is correlated with a worse prognosis of breast carcinoma. Interestingly, the CENPW mutation contributes to the poor prognosis. Next, we discovered that the genes interacting with CENPW are mainly concentrated in the cell cycle pathway, and CENPW is co-expressed with CDCA7, which is also highly expressed in breast carcinoma and leads to a worse prognosis. Our subsequent studies verified that knockdown of CENPW significantly inhibits the proliferation and migration of breast carcinoma cells and promotes their apoptosis rate. Notably, inhibition of CEMPW sensitizes breast cancer cells to chemotherapeutic drugs that have been found to induce cell cycle arrest. In summary, these results provide extensive data and experimental evidence that CENPW can serve as a novel predictor of breast cancer and may act as a prospective therapeutic target.

Keywords: CENPW, poor prognosis, biomarker, breast cancer, therapeutic strategy

INTRODUCTION

The International Agency for Research on Cancer (IARC) announced the latest global carcinoma burden data in 2020, showing that the new incidence of breast carcinoma in the world is up to 11.7%, which has overtaken lung carcinoma as the world's largest most common cancer. Breast invasive carcinoma (BRCA) is the widest tumor type in women, with a poor prognosis and high recurrence rates (Lang et al., 2017). At

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present, paclitaxel combined with carboplatin has made great progress in the therapy of breast carcinoma, the long-term use of chemotherapeutic drugs produces obvious resistance and the therapeutic efficacy is still not ideal. (Fasching et al., 2021). In the general population, the pathogenesis of breast cancer is often directly related to alterations in gene expression (van den Broek et al., 2021). Therefore, finding the driver genes in tumorigenesis may open up new strategies for early breast cancer screening and the discovery of potential therapeutic targets, which will facilitate early detection and thereby reduce morbidity and mortality (Shen et al., 2021).

Centromeric protein W (CENPW), also known as cancer upregulated gene 2 (CUG2) protein, is overexpressed in multiple tumor tissues, including cervical, colon, liver, and lung cancers (Malilas et al., 2014). The CENPW is located at human chromosome 6q22.32 and is regulated by the upstream TSS region, which can transcribe a full-length mRNA of 600 bp (Lee et al., 2007). CENPW is a representative member of the Constitutive Centromere Associated Network (CCAN) family and is of crucial importance in the formation of centromeric nucleosomes (Klare et al., 2015). CENPW is encoded by CENPW and forms a DNAbinding heterodimer with CENPT (Prendergast et al., 2016), which is able to switch centromeric chromatin to a mitotic state (Prendergast et al., 2011). Previous studies have found that the regulation of CENPW can affect the migration of colon carcinoma cells and induce the epithelial-mesenchymal transition of lung carcinoma cells via $TGF-\beta$ signaling (Malilas et al., 2013; Kaowinn et al., 2018). However, the role and value of CENPW in breast carcinoma remain unclear and require further study.

In the current study, several databases were used to evaluate the role of *CENPW* in breast carcinoma. We discovered that *CENPW*, which is highly expressed in breast carcinoma, is associated with a poor prognosis of breast carcinoma. We further found that *CENPW* mainly influences the cell cycle pathway, and the downregulation of *CENPW* expression could inhibit the migration and proliferation of breast cancer cells and promote apoptosis. Notably, we believe that *CENPW* could be a biomarker in the development process of breast carcinoma and propose that inhibition of *CENPW* may be a prospective strategy for inhibiting breast carcinoma development.

MATERIALS AND METHODS

GEPIA Analysis Dataset

GEPIA is an interactive website server based on TCGA and GTEx databases, which provides us with customizable functional modules (http://gepia.cancer-pku.cn/). In this study, the *CENPW* expression was analyzed using GEPIA in different tumors and paracancerous tissues. The association between *CENPW* and the highly coexpressed gene (*CDCA7*) was evaluated according to the function of correlation analysis. In addition, the OS of *CDCA7* gene was evaluated by the survival plots module.

Oncomine Database Analysis

The Oncomine Database is an openly available database for the analysis of *CENPW* expression levels in multiple tumor types. The difference of mRNA expression and co-expression of

CENPW gene in breast carcinoma were studied by this database. The thresholds were set as two-fold change, top 10% gene rank and p-value = 1E-4.

UALCAN Cancer Database Analysis

The UALCAN Cancer Database comprehensively contains TCGA dataset, CPTAC dataset, and CBTTC dataset. The *CENPW* expression in BRCA was verified and the relationship between clinicopathological features and *CENPW* expression was identified by the UALCAN Cancer Database.

Bc-GenEx Miner Analysis

Bc-GenEx Miner is a full-featured online access database, which is specialized for breast cancer. According to this database, the *CENPW* expression in different clinical stages was mined, and the correlation between *CENPW* and the related genes was clustered.

Kaplan–Meier Survival Analysis

To evaluate the prognostic significance of a particular gene, the patient samples were divided into two groups on the basis of the median expression. The prognostic significance of *CENPW* in breast carcinoma was evaluated according to overall survival (OS), distant metastasis-free survival (DMFS), and relapse-free survival (RFS) by Kaplan–Meier plotter. Log-rank *p*-value and HRs with 95% confidence intervals were set.

cBioPortal Website Analysis

cBioPortal is a comprehensive and open website based on the TCGA database, which provides us with visual cancer gene mutation data (http://www.cbioportal.org/). The *CENPW* gene mutations were analyzed using seven breast cancer databases available on the cBioPortal website.

UCSC Xena Website Analysis

The UCSC Xena website contains multiple genomic databases for researching correlations between genomic or phenotypic variables (https://xenabrowser.net/). The TCGA database related to breast cancer provided by UCSC Xena was used for data mining to construct a heat map of *CENPW* and *CDCA7*.

GeneMANIA Website Analysis

The GeneMANIA site comprises a wealth of gene information, and can be used for functional analysis (http://www.genemania.org). Its prediction algorithm has high accuracy (Warde-Farley et al., 2010). Therefore, the gene clusters of mutual value with *CENPW* were predicted using this website.

Metascape Online Analysis

Metascape Online (https://metascape.org/gp/index.html) can be employed for function enrichment of *CENPW*-related gene sets and PPI network analysis (Zhou et al., 2019). And we used MCODE1 to further analyze densely connected regions. *p*-value < 0.05 was set as a cutoff value (Lu et al., 2021).

TIMER Website Analysis

The new version of TIMER 2.0 has seven main analysis systems, with gene and survival functions designed to assess the infiltration of multiple immune cells (https://cistrome.shinyapps.io/timer/) (Li et al., 2017). *CENPW* was selected to generate a scatter map through a gene module to observe the relevance between the expression of *CENPW* and the immune infiltration level in breast carcinoma.

Cell Culture

Human breast carcinoma cell lines (MDA-MB-231, BT-549) were purchased from the Cell Bank of the Chinese Academy of Science (Shanghai, China). MDA-MB-231 cell was cultured in DMEM (Hyclone, Logan, UT, United States) with 10% FBS (Gibco, Grand Island, United States) at 37°C and 5% CO₂. BT-549 cell was cultured in RPMI-1640 (Hyclone, Logan, UT, United States) with 10% FBS (Gibco, Grand Island, United States) at 37°C and 5% CO₂.

Cell Transfection

Two pairs of siRNAs targeting *CENPW*, and negative control siRNA-NC (si-NC) were synthesized (RiboBio, Guangzhou, China). MDA-MB-231 and BT-549 cells were plated with appropriate cell density, and further transfected corresponding siRNA (si-NC, si1-*CENPW* and si2-*CENPW*) by LipofectamineTM 3000 (Invitrogen, Carlsbad, CA, United States).

Western Blot Analysis

The transfected cells were lysed with RIPA buffer (Sigma-Aldrich, United States) to extract protein solution. The protein concentration was gauged with BCA kit (Boxbio Science and vernight with primary antibodies, then removed for (HRP)conjugated secondary antibody incubation. The blots were visualized by ECL-Plus kit (Thermo Scientific, United States).

Cell Proliferation Detection

Cell proliferation was detected by CCK-8 (Meilunbio, China). 5,000 transfected cells were inoculated into a 96-well plate (NEST Biotechnology). After culture for 0, 24, 48 and 72 h, 10 μ L CCK-8 reagent was added into detecting well, and the optical density at 450 nm wavelength was detected.

Clone Formation Assay

Transfected cells (500 cells/well) were seeded into 6-well plates. A second transfection was performed 7 days after the first one to improve the transfection efficiency. After 14 days, the colonies were fixed with paraformaldehyde and stained with crystal violet for 20 min. Whereafter, the field of vision was photographed by a light microscope (Olympus, Japan).

Transwell Invasion Assay

The transfected cells were seeded into the upper chamber (Corning, United States), and the lower chamber was added with 500 μ L 5% FBS complete medium. After 24 h of culture, the cells were fixed with paraformaldehyde solution and stained with crystal violet for 20 min. Finally, the invading cells were captured by a light microscope (Olympus, Japan).

Wound Healing Assay

The transfected cells were seeded into 6-well plates and grew to around 90% confluence. The monolayer cells were then scratched with the tip of a 10 μ L pipette. After 24 h of culture, the wound healing area was captured by a light microscope (Nikon, Japan).

Flow Cytometry Analysis

The cell apoptosis was detected by flow cytometry using the Annexin V-FITC/PI apoptosis kit (MultiSciences, Hangzhou, China). The transfected cells (6×10^5 cells/well) were cultured in 6-well plates for 48 h and collected in 1×Binding buffer, then Annexin V-FITC (5μ L), and PI (10μ L) were added for 10 min incubation in the dark and subjected to the flow cytometry (Beckman coulter, CA, United States).

Statistical Analysis

All statistical computing was performed by GraphPad Prism 7.0. Results are expressed as mean \pm SD. The discrepancies between two groups were accomplished by the student's *t*-test. Statistical significance was established at **p* < 0.05, ***p* < 0.01, compared to the corresponding control group.

RESULTS

The Evaluation of *CENPW* Expression in Multiple Cancer Tissues

To clarify the molecular biological characteristics of CENPW in different tumors, we first investigated CENPW expression in cancers and adjoining normal tissues from various databases. The data from the Oncomine database verified that the expression of CENPW was enhanced in numerous tumors including brain, breast, colorectal, head and neck, lung carcinomas compared with normal samples, while CENPW expression was lower in leukemia than in normal samples (Figure 1A). GEPIA database was further employed to evaluate the expression differences of CENPW in 33 kinds of tumor types (Figure 1B). We found that the CENPW was highly expressed in many tumors such as in BLCA, BRCA, COAD, HNSC, LUAD and STAD. On the contrary, the CENPW expression was lower than that of normal controls in acute myeloid leukemia. Collectively, these results from different databases together revealed that CENPW is excessively upregulated in most tumors and may act as a potential indicator of cancer progression.

CENPW Acts as an Oncogene in the Carcinogenesis and Progression of Breast Carcinoma

To better explore the potential value of *CENPW* in breast carcinoma patients, we further explored the expression and function of *CENPW* in breast carcinoma through several means. According to the Richardson Breast two dataset (Richardson et al., 2006), the *CENPW* expression in ductal breast cancer was distinctly higher than that in normal tissues (**Figure 2A**). In addition, the Curtis Breast dataset (Curtis et al., 2012) showed a 2.698-fold (p = 1.70E-09) increase in



CENPW mRNA expression in medullary breast carcinoma (Figure 2B). Differences in mRNA expression of *CENPW* in breast carcinoma were also confirmed using the UALCAN database (Figure 2C). In addition, the UALCAN database was devoted to analyzing the relation between *CENPW* expression and different clinicopathological characteristics in BRCA patients. The data indicated that the *CENPW* expression was higher in female patients and was positively associated with patient age, nodal metastasis, and cancer stage (Figures 2D-G). Additionally, the relation between *CENPW* expression and triple-negative breast carcinoma classification was analyzed, and the expression level of *CENPW* was the highest in TNBC-BL2 (Figure 2H). The pairwise comparison *p*-values of the above results obtained by the UALCAN database are shown in Supplementary Table S1.

We also evaluated the expression of *CENPW* with different clinicopathological features in breast carcinoma patients by the bc-GenExMiner online tool. The *CENPW* expression was higher in the less than 51-year-old group, and higher *CENPW* expression

was correlated with a more advanced Scarff–Bloom–Richardson (SBR) grade (**Figures 3A,B**). *CENPW* levels in breast carcinoma patients with positive node status (N) were higher than those in patients with negative node status (**Figure 3C**). Progesterone receptor (PR) and estrogen receptor (ER) status were negatively correlated with *CENPW* expression (**Figures 3D–F**). In contrast, human epidermal growth factor receptor-2 (HER-2) status was positively correlated with *CENPW* expression (**Figure 3G**). In addition, *CENPW* increased in base-like subtypes compared to non-base-like subtypes (**Figure 3H**). Simultaneously, *CENPW* was strongly elevated in triple-negative breast cancer (TNBC) patients (**Figure 3I**). Detailed data on the clinicopathological features are presented in **Supplementary Table S2**.

The High Level of *CENPW* is Correlated With Poor Prognosis in Breast Carcinoma Patients

We further explored the relationship between *CENPW* expression and breast carcinoma prognosis. Results of the



Kaplan-Meier plotter showed that higher expressions of *CENPW* were linked to poorer OS (**Figure 4A**). Similarly, DMFS and RFS were worse in breast cancer patients with increased *CENPW* expression (**Figures 4B,C**). To reconfirm the function of *CENPW* in breast carcinoma prognosis, we illustrated the effect of *CENPW* on OS, DMFS, and DFS by bc-GenExMiner database. Breast cancer patients with upregulated *CENPW* presented with worse OS, DMFS, and DFS (**Figures 4D-F**), which were similar to the results of the Kaplan-Meier plotter. In summary, these data showed that

CENPW expression is associated with poor prognosis in breast cancer.

The Analysis of Gene Mutation and Interaction of *CENPW* in Breast Carcinoma Patients

To further understand the influence of *CENPW* gene mutations on breast carcinoma patients, we used the cBioPortal database and detected mutations in seven breast cancer databases. It was found that in 6,801 breast cancer patients, 2% had mutations in the



CENPW gene (**Figure 5A**), and alterations with amplification were more common (**Figure 5B**). Furthermore, we predicted the 3D structure of *CENPW* and identified the mutation site using the cBioPortal online tool (**Figure 5C**). Next, we evaluated the effect of mutation on the prognosis of breast carcinoma patients and found that the high mutation group was obviously linked to shorter DFS and PFS in breast carcinoma patients (**Figures 5D,E**). In addition, the UALCAN database was used to assess the methylation levels of the *CENPW* promoters in BRCA (**Figure 5F**). The data indicated that the methylation level of the *CENPW* promoter in patients with BRCA was dramatically reduced, which may give rise to an increase in *CENPW* expression. We also revealed the relation between *CENPW* expression and TP53 mutations and found that *CENPW* was highly expressed in TP53 mutant groups (**Figure 5G**).

The Co-expression of *CENPW* and *CDCA7* in Breast Carcinoma

To further explore the mechanism of *CENPW* in BRCA, we employed the Oncomine database to construct the co-expression network of *CENPW* (**Figure 6A**). The co-expression profile showed that *CENPW* had the highest association coefficient



analysis of (D) OS, (E) DMFS and (F) DFS analyzed by bc-GenExMiner database in patients with breast cancer.

with CDCA7 among 129 breast carcinoma samples in the LU Breast dataset (Lu et al., 2008). We also confirmed the correlation between CENPW and CDCA7 through the GEPIA website and bc-GenExMiner database and obtained results consistent with those of Oncomine (Figures 6B,C). Furthermore, CENPW and CDCA7 expression heat maps were derived using the UCSC Xena website. The CENPW expression was positively linked to the transcription level of CDCA7 in a 50-gene qPCR assay (PAM50) for breast carcinoma subtypes (Figure 6D). These data indicated that CENPW might be associated with the CDCA7 signaling pathway in breast cancer. Then the UALCAN database was performed to verify differences in CDCA7 expression. The results revealed that CDCA7 was obviously upregulated in BRCA (Figure 6E). According to the outcome of the GEPIA database survival analysis, CDCA7 upregulation was correlated with poorer OS in breast carcinoma patients (Figure 6F), which was consistent with the effect of CENPW on breast cancer.

The Enrichment and Interaction Analysis of *CENPW*-Related Genes

Twenty genes (including CENPT, CENPH, CENPA, FOXO3, and CKS2) interacting with CENPW were excavated using the

GeneMANIA website (Figure 7A). To explore the signaling pathway associated with CENPW, we used Metascape Online for functional analysis and discovered that the genes interacting with CENPW were primarily enriched in cell cycle, cell division, and chromosome maintenance (Figure 7B). These data inspired us to explore the relationship between CENPW and the cell cycle pathway. Next, the PPI network based on Metascape Online and the MCODE1 plug-in, further helped to identify these important modules of genes interacting with CENPW (Figures 7C,D). To further verify the co-expression of CENPW and CDCA7, we also conducted the above study on CDCA7. The functional analysis of twenty genes interacting with CDCA7 showed that they were also mainly enriched in cell cycle signaling pathway (Supplementary Figure S1). This was consistent with the main biological effects of CENPW on breast cancer cells.

The Correlation Between *CENPW* Expression and Immune Cell Infiltration

The relation between *CENPW* expression and the level of immune cell infiltration in BRCA was further investigated by



TIMER2.0 database. Specifically, we investigated the association between CENPW expression and specific immune cell subsets. The data showed that CENPW expression was positively associated with most infiltrated immune cells in BRCA but negatively associated with the immune invasion levels of macrophages (Figure 8). There were noticeable discrepancies between CENPW expression and immune invasion levels of B cells, neutrophils, dendritic cells and macrophages. In addition, we analyzed the immune process of CDCA7 in the breast cancer microenvironment, and the expression level of CDCA7 was negatively correlated with macrophages, positively correlated with immune of B cells, neutrophil, dendritic invasion cell

(Supplementary Figure S2), which is consistent with the results of *CENPW*.

Knockdown of *CENPW* Inhibits the Proliferation of Breast Carcinoma Cells

The above bioinformatics data proved that under-expression of *CENPW* was correlated with better survival in breast cancer patients. To explore the role of *CENPW* in breast cancer cells, we transfected MDA-MB-231 and BT-549 cells with two specific siRNA sequences to induce knockdown of *CENPW* expression. Growth curves suggested that *CENPW* knockdown suppressed the proliferation of these two breast carcinoma cell lines (*p < 0.05, Figures 9A,B).



FIGURE 6 Coexpression analysis of *CENPW*. (A) Coexpression profile of *CENPW* derived from the Oncomine database. (B) The relation between *CENPW* and *CDCA7* expression in breast carcinoma in GEPIA website and (C) the bc-GenExMiner database. (D) Heat map of *CENPW* and *CDCA7* expression across PAM50 subtypes by UCSC Xena website. (E) The *CDCA7* expression of breast cancer in Ualcan database. (F) The OS status according to the *CDCA7* expression in GEPIA database. *p < 0.05, **p < 0.01 between the compared groups.

In addition, we examined the effect of *CENPW* knockdown on long-term cell viability using a clonal formation survival assay in MDA-MB-231 and BT-549 cells (**Figures** **9C,D**). And then after 14 days of culture, the clonal number of the two breast carcinoma cell treated with si1-*CENPW* and si2-*CENPW* decreased compared to the control cells.



Knockdown of *CENPW* Inhibits Migration and Invasion of Breast Cancer Cells

The most concerning aspect of cancer is its ability to migrate and invade. Therefore, the invasion and migration of MDA-MB-231 and BT-549 cells were evaluated using adherent cell scratch migration and transwell invasion assays. Transwell analysis indicated that knockdown of *CENPW* reduced the number of invading cells (**p < 0.01, **Figures 10A,B**). In addition, as indicated by the scratch test in **Figures 10C,D**, the wound healing rates of MDA-MB-231 and BT-549 cells after *CENPW* silencing were significantly reduced (**p < 0.01). Moreover, these results together revealed that knockdown of *CENPW* can reduce the migration and invasion ability of breast carcinoma cells.

Knockdown of *CENPW* Blocks the Cell Cycle Process and Increases the Probability of DNA Damage and Apoptosis in Breast Carcinoma Cells

To figure out the causes of these phenomena, we discovered the protein expression levels of the related pathways. The western blot results indicated that si1-CENPW and si2-CENPW could induce the degradation of CDK6 and Cyclin-D1, suggesting that inhibition of CENPW could block the cell cycle process, which was consistent with the data of GO and KEGG analysis. Furthermore, the significantly decreased protein expression levels of Wee1 and CHK1 indicate that knockdown of CENPW may cause DNA damage, which may be closely related to the participation of CENPW in the nucleosome assembly process. Meanwhile, we discovered that the expression of the apoptosis-related gene decreased after CENPW BCL2 knockdown (Figure 11A). Further apoptosis assay detected by flow cytometry reflected that knockdown of CENPW increased the proportion of apoptotic cells (Figure 11B). Notably, we found that breast cancer cells with reduced CENPW expression were more sensitive to chemotherapeutic drugs that have been found to induce cell cycle arrest. Improved tumor cell killing effect of cisplatin and doxorubicin was found in the CENPW inhibited breast carcinoma cells (Figures 11C,D).





colony formation of breast cancer cells with or without *CENPW* knockdown. Quantitative histograms of colony formation are shown on the right. *p < 0.05, **p < 0.01 compared with si-NC group.



DISCUSSION

At present, breast cancer has become the largest number of diagnoses and the highest incidence in women, and it has serious adverse effects on health (Joko-Fru et al., 2021). Therefore, it is crucial to investigate the pathogenesis, diagnosis, treatment, and prognosis of breast carcinoma (Liang et al., 2020). In the study, we explored and revealed that abnormal CENPW expression is closely linked to breast tumors through various databases and in vitro cell experiments. Ziliang Zhou, et al. (Zhou et al., 2020) reported that the CENPW expression level in liver carcinoma was obviously higher than normal liver tissues. Similarly, we discovered that CENPW expression was up-regulated in ductal breast carcinoma and medullary breast carcinoma using two different online databases. According to the clinicopathological features of breast carcinoma, compared with normal tissue, the HER-2 status, SBR grade, basallike status, nodular status, and triple-negative status were positively associated with CENPW levels in breast carcinoma samples. In contrast, age, ER, and PR were negatively correlated with CENPW expression. These results indicated that CENPW expression was linked to poor prognosis of breast carcinoma, which was also confirmed by subsequent data from the bcGenexMiner database. Therefore, based on the above series of data analyses, we believe that *CENPW* may be a promising indicator for screening and may be used as a new prognostic factor for breast carcinoma.

CENPW is a crucial member of the CCAN family, which plays a critical role in nucleosome assembly and is related to the deregulation of centromere proteins and carcinogenesis (Hori et al., 2008). Our findings showed that breast carcinoma patients with high CENPW expression had a worse prognosis than those with low CENPW expression. Notably, through gene mutation analysis in the cBioPortal database, we identified that mutations in CENPW were associated with poor prognosis. Interestingly, a highly correlated gene, cell division cycle-associated protein 7 (CDCA7) (Ye et al., 2018), was discovered by CENPW co-expression analysis. Cai et al. (Cai et al., 2021) reported that downregulation of CDCA7 not only inhibits cell proliferation, but also intercepts the cell cycle in ovarian carcinoma. In the current study, functional enrichment analysis indicated that genes interacting with CENPW were mainly enriched in the cell cycle. These data indicated that CENPW and CDCA7 may play a joint role in the cell cycle pathway; however, further direct evidence is required to confirm this. In addition, studies on CDCA7 have found that its high expression can help to predict



poor prognosis and tumor progression in colorectal carcinoma and kidney carcinoma (Li et al., 2020; Liu et al., 2021). Further investigation reveals that *CDCA7* expression is also up-regulated in breast carcinoma and leads to poor prognosis.

Notably, CENPW principally affects the cell cycle process of tumor cells by regulating nucleosome assembly (Sridhar et al., 2021). Therefore, we used the GeneMANIA website and Metascape online tools to analyze the gene enrichment and pathway enrichment of CENPW, and the data suggested that CENPW has a vital influence on the cell cycle pathway. In addition, Zhou et al. (Zhou et al., 2021) reported that knockdown of CENPW restrains HCC progression by inactivating E2F signaling. However, the significance of its downregulation in breast carcinoma prognosis has not yet been studied. To illustrate the specific molecular function of the CENPW gene in breast cancer, we constructed CNEPW gene silenced cells and found that CENPW suppression could inhibit the proliferation and migration of MDA-MB-231 and BT-549 cells, hindering the cell cycle signaling pathway, thereby causing apoptosis of breast tumor cells. Previous studies have confirmed that CENPW accelerates cell transformation and stemness through nuclear NPM1 protein and TGF-β signaling (Kaowinn et al., 2019a; Yawut et al., 2020). Moreover, Kaowinn et al. (Kaowinn et al., 2019b) found that CENPW elevated the expression of YAP1, resulting in the

increase of the EMT in lung cancer cells. Interestingly, we found that breast cancer cells with *CENPW* inhibited were more sensitive to chemotherapeutic drugs that have been found to induce cell cycle arrest. These effects may be resulting from the inhibition of TGF- β signaling pathway, but required further exploration.

These cellular results were consistent with the above bioinformatics analysis and fully illustrated the fact that *CNEPW* may be a crucial oncogene and accelerant for the progression of breast carcinoma, which could help in the development of new breast carcinoma treatment strategies. However, there are still some limitations in our current study. A gene regulates multiple metabolic pathways, but its specific mechanism of action cannot be clearly explained by a single signaling pathway. Therefore, further experimental studies are required to better understand its functions. Further clinical trials are indispensable for elucidating the value of *CENPW* in the treatment of breast carcinoma.

CONCLUSION

This study presents evidence that high *CENPW* expression in breast cancer generates poor prognosis and can therefore be used as a novel

biomarker for breast carcinoma. In particular, mutations in the *CENPW* gene also led to a poor prognosis. Therefore, knockdown *CENPW* expression can suppress the proliferation and migration of breast cancer cells and induce apoptosis. Our results provide bioinformatics data and experimental evidence that *CENPW* can be used as a unique biomarker for breast carcinoma, and as a hopeful therapeutic option for the development of related inhibitors to treat breast cancer at the genetic level.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding authors.

AUTHOR CONTRIBUTIONS

FZ, XT, and YW conceived and designed the experiments. LW, HW, and CY performed the experiments. YW, GL, YY, and YG

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analyzed the data. YL and LW wrote the paper. FZ, JD, and YW revised and finalized the manuscript. All authors read and approved the final manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fgene.2022.900111/full#supplementary-material

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