Check for updates

OPEN ACCESS

EDITED BY Dimitra Kiritsi, University of Freiburg Medical Center, Germany

REVIEWED BY Hongbin Lv, Southwest Medical University, China Kitiwan Rojnueangnit, Thammasat University Hospital, Thailand

*CORRESPONDENCE Ke Wu, 754299058@qq.com

[†]These authors have contributed equally to this work

SPECIALTY SECTION

This article was submitted to Genetics of Common and Rare Diseases, a section of the journal Frontiers in Genetics

RECEIVED 20 March 2022 ACCEPTED 01 July 2022 PUBLISHED 15 August 2022

CITATION

Cong Y, Jin H, Wu K, Wang H and Wang D (2022), Case Report: Chinese female patients with a heterozygous pathogenic RPS6KA3 gene variant c.898C>T and distal 22q11.2 microdeletion. *Front. Genet.* 13:900226. doi: 10.3389/fgene.2022.900226

COPYRIGHT

© 2022 Cong, Jin, Wu, Wang and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License

(CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Case Report: Chinese female patients with a heterozygous pathogenic *RPS6KA3* gene variant c.898C>T and distal 22q11.2 microdeletion

Yan Cong^{1†}, Hongxing Jin^{2†}, Ke Wu³*[†], Hao Wang¹ and Dong Wang¹

¹Rehabilitation Department, Yiwu Maternity and Child Health Care Hospital, Yiwu, China, ²Pediatric Department, Yiwu Maternity and Child Health Care Hospital, Yiwu, China, ³Prenatal Diganosis Center, Yiwu Maternity and Child Health Care Hospital, Yiwu, China

Background: Coffin–Lowry syndrome (CLS) [OMIM#303600] is a rare X-linked dominant syndrome. CLS is caused by highly heterogeneous loss-of-function mutations in the RPS6KA3 gene (OMIM*300,075). CLS is characterized by intellectual disability (ID), short stature, tapered fingers, characteristic facial features, and progressive skeletal changes. Distal 22g11.2 microdeletion syndrome (OMIM#611867) is an autosomal dominant and recurrent genomic disorder. It mainly includes three types [distal type I (D-E/F), type II (E-F), and type III (F-G)] and exhibits variable clinical phenotypes (mild, moderate, or even normal): preterm birth, pre- and/or postnatal growth restriction, development delay, ID, behavioral problems, cardiovascular defects, skeletal anomalies, and dysmorphic facial features. We investigated the genetic etiology of a Chinese pedigree with ID, short stature, digit abnormalities, facial dysmorphism, and menstrual disorder. A heterozygous RPS6KA3 gene variant c.898C>T (p.R300X) was identified in this familial case. Two female CLS patients with distal 22g11.2 microdeletion presented with more severe clinical phenotypes. We provided clinical characteristics of these Chinese female CLS patients.

Case presentation: We described a Chinese family with three affected females (the mother, the elder sister, and the proband). The mother and the elder sister had more severe clinical phenotypes (moderate facial dysmorphism, more severe cognitive impairment, and shorter stature). The common characteristic phenotypes are ID, short stature, facial dysmorphism, irregular menstruation, and cardiovascular disorders. Peripheral blood samples were collected from the pedigree. Whole-exome sequencing (WES) identified a heterozygous nonsense *RPS6KA3* gene variant c.898C>T (p.R300X). It was verified by Sanger sequencing. Copy number variation sequencing

Abbreviations: CLS, Coffin–Lowry syndrome; RPS6KA3, ribosomal protein S6 kinase A3; gDNA, genomic DNA; Trio-WES, trio-based whole-exome sequencing; NV, copy number variation; MMSE, the mini-mental state examination; MRI, magnetic resonance imaging; SIDAs, stimulus-induced drop attacks; XCI, X-chromosome inactivation; COS, childhood-onset schizophrenia (COS).

(CNV-seq) showed that both the mother and the elder sister carried a CNVseq [hg19] del (22) (q11.22-q11.23) (22997582–23637176)×0.5. RNA from peripheral blood samples was used for measuring the relative quantification of mRNA (expressed by exon 14 of *RPS6KA3*). The levels of mRNA relative expressions were significantly lower in the mother's and the elder sister's blood samples. The levels of mRNA relative expressions were significantly higher in the proband's blood sample. X-chromosome inactivation (XCI) studies demonstrated that the proband showed extremely skewed XCI, and the XCI pattern of the elder sister was random.

Conclusion: Herein, we reported three Chinese female patients with a heterozygous nonsense *RPS6KA3* gene variant c.898C>T. Further genetic studies were performed. To our knowledge, Chinese patients with this variant have not been previously reported in the literature. The three female patients presented with variable degrees of severity. The clinical characteristics of these Chinese female CLS patients could expand the phenotypic spectrum of CLS. We helped physicians to understand the genotype–phenotype correlation further.

KEYWORDS

Coffin–Lowry syndrome, loss-of-function, intellectual disability, menstrual disorder, whole-exome sequencing, RPS6KA3 c.898C>T mutation

Background

CLS is an X-linked dominant disorder characterized by ID, craniofacial features, and skeletal abnormalities. CLS is caused by heterozygous loss-of-function mutations in the *RPS6KA3* gene. A total of 70–80% of probands of CLS had no family history, and roughly 2/3 occurred *de novo*. A total of 20–30% of CLS patients had more than one affected family member (Pereira et al., 2010). With the use of WES, we reported a Chinese pedigree with a heterozygous pathogenic variant c.898C>T (p.R300X) in *RPS6KA3* gene. Further genetic studies were performed. A CNV was found in two female CLS patients with more severe clinical phenotypes, they may be associated with distal 22q11.2 microdeletion syndrome. We described clinical characteristics of these Chinese female CLS patients. Also, we hoped to improve the comprehensive understanding of CLS for pediatricians.

Case presentation

The proband (a 17 year-old female patient) was found by us in the countryside. The proband had additional two affected family members (Figure 1). They had similar symptoms. The parents were residents of adjacent villages, and they did not have a consanguineous relation. The proband's father was phenotypically normal. The proband and her elder sister (21 years old) were born at full-term by spontaneous vaginal delivery without complication in the perinatal and neonatal period. The weight and height of the proband were 56 kg and 161 cm, respectively. The weight and height of her elder sister were 72 kg and 142 cm, respectively. The weight and height of her mother (47 years old) were 65 kg and 140 cm, respectively. The cognitive impairment tests showed that MMSE scores of the



proband, her elder sister, and mother were 13, 8, and 5, respectively. The proband had received education at a special school for 2 years. Now she can recognize some Chinese characters, and she has simple communication with people. She had a poor memory like always forgetting her own birthday. Her elder sister and mother could not recognize any Chinese characters. They had little communication with people. The three patients all had coarse craniofacial features including widely spaced downward-slanting palpebral fissures, low nasal bridge, blunt tip, broad nose, anteverted nares, and thick lips (Figure 2C). In addition, her elder sister had acne all over her face (Figure 2B), and her mother had a wide and open mouth, and thick lips with everted lower lip (Figure 2A). The elder sister's and mother's fingers tapered markedly from relatively wide proximally to narrow distally with small terminal phalanges and nails. The proband's fingers did not show anything abnormal. During their adolescent period, they all had irregular menstruation. Echocardiography showed that they all had mild mitral and tricuspid valve regurgitation. X-rays of spines showed that only the mother had scoliosis (Figure 3A). X-rays of hands did not show any signs of abnormality.

Neuropsychological tests

The mini-mental state examination (MMSE) (Pangman et al., 2000) is a 30-point questionnaire. MMSE is one of the most widely used brief screening instruments for measuring

cognitive impairment. It has been modified and translated into a Chinese version. The optimal cut-off points were determined according to the education level. The optimal cutoff points were 16/17 for illiterate individuals, 19/20 for individuals with 1–6 years of education, and 23/24 for individuals with seven or more years of education (Li et al., 2016).

Genetic tests

The guardian (the proband's father) signed an informed consent for genetic analysis. Our legal ethics committee approved this genetic study. gDNAs were extracted from peripheral blood of the patients and phenotypically normal father for WES and CNVseq. Sanger sequencing was used for further verification. CNV-seq showed that both the mother and the elder sister carried a CNVseq [hg19] del (22) (q11.22-q11.23) (22997582–23637176) \times 0.5 (Figure 4). WES identified a heterozygous nonsense *RPS6KA3* gene variant c.898C>T (p.R300X), which was verified by Sanger sequencing (Figure 5).

Levels of urine organic acids, plasma amino acids, lactate, and pyruvic acid were all normal in patients. Conventional G-banded chromosome analysis all showed a 46, XX karyotype. The results of genetic metabolic disease screening were all negative in patients.

As per the guidelines of the American College of Medical Genetics and Genomics (ACMG) for interpreting sequence variants (Richards et al., 2015), this variant was pathogenic





(PVS1+PM2+PP1+PP3+PP4+PP5). Total cellular RNA was isolated from patients' peripheral blood for measuring the relative quantification of mRNA (expressed by exon 14 of the *RPS6KA3* gene). The quantitative PCR (qPCR) data demonstrated that the levels of mRNA relative expression were significantly lower in the mother's and the elder sister's blood samples, and the levels of the mRNA relative expression were significantly higher in the proband's blood sample (Figure 6).

gDNAs from peripheral blood samples of the two sisters and their mother were used for XCI analysis. The XCI studies demonstrated that the proband showed extremely skewed XCI and the XCI pattern of the elder sister was random.

Discussion

CLS is a well-described and rare X-linked dominant syndrome. CLS is caused by highly heterogeneous loss-offunction mutations in the *RPS6KA3* gene. The ribosomal protein *S6 kinase A3* (*RPS6KA3*) gene is located in Xp22.12, and it encodes ribosomal protein S6 kinase alpha-3 (UniProtKB-P51812). This transcript of the RPS6KA3 gene has 22 coding exons, a transcript length of 7987 base pairs, and a translation length of 740 amino acids (NM_004586.3). The RPS6KA3 is one of the serine/threonine-protein kinase. RPS6KA3 acts as one of many downstream effectors of the MAP-kinase ERK in the RAS-RAF-MEK-ERK signaling pathway. RPS6KA3 plays an important role in cell-cycle progression, differentiation, and cell survival (Fischer and Raabe, 2018). RPS6KA3-deficient mice displayed spatial learning impairment (Poirier et al., 2007). In RPS6KA3 knock-out mice, proliferation of adultgenerated neurons was decreased and no pro-survival effect of learning was observed. Castillon et al. (2018) suggested that decreased young newborn neurons were associated with deficient long-term memory recall. Likewise, poor memory and learning impairment have been observed in our CLS patients.

Based on the authors' experience, the prevalence of CLS was 1:40,000–1:100,000 (Pereira et al., 2010). CLS is usually characterized by severe-to-profound ID, characteristic craniofacial features, tapered fingers, and musculoskeletal



features. Carrier females are less severely impaired. Approximately 14% of affected males and 5% of affected females had cardiovascular diseases (Hanauer and Young, 2002). A total of 13-20% of male CLS and 3-7% of female CLS presented with stimulus-induced drop episodes (SIDEs) (Rojnueangnit et al., 2014). In addition, a few CLS patients presented with diabetes type 2 (Boulos et al., 2021), epileptic seizures (Gschwind et al., 2015), growth retardation (Lv et al., 2019), and hearing or vision problems (Hunter, 2002). The risk of childhood-onset schizophrenia (COS) might be increased in CLS patients (Ambalavanan et al., 2019). Neuroimaging studies demonstrated some brain abnormalities, such as thinning and agenesis of the corpus callosum, mild dilatation of the ventricles, reduced gray- and white-matter volumes, and impacted cerebellum and hippocampus volumes (Wang et al., 2006; Kesler et al., 2007).

In this familial case, these three patients all had the ability to walk, no loss of hearing or vision. For now, there are no signs of SIDEs and epileptic seizures in them. They all had irregular menstruation at puberty. The length of their menstrual cycles kept changing, such as 45 days, 60 days, and 180 days. This symptom has not been previously reported in the literature. Ultrasonography of ovaries and uteruses all showed normal results. They refused to take brain magnetic resonance imaging (MRI) scans.

According to the ACMG standards and guidelines for the interpretation of sequence variants, we interpreted c.898C>T (p.R300X) as a pathogenic variant. This variant caused a premature termination of codon, resulting in strong protein truncation (more than 50% of protein length was missing) (PVS1). It was absent from normal population databases (Exome Sequencing Project, 1000 Genomes Project, the Genome Aggregation Database, and Exome Aggregation Consortium) (PM2). Silico predictive algorithms (SIFT, PolyPhen-2, and Mutation Taster) of pathogenicity all showed that this variant was damaging. This variant was segregated from the disease in multiple family members (PP1). The analysis of conserved sequences suggested that this variant was located in highly conserved sequences across several species (Figure 7) (PP3). Three female patients' phenotypes and family histories were highly specific for CLS (PP4). Tos et al. (2015) reported that a familial CLS case was caused by the same variant (PP5).

As shown in Table 1, the mother and the elder sister had the similar symptoms. They were more severely impaired than the proband. They presented with shorter stature, coarser facial features, tapered fingers, and poorer cognition. We speculated that more severe phenotypes may be connected with the CNV [del (22) (q11.22-q11.23) (22997582–23637176)]. According to a proposed categorization system (Mikhail et al., 2014), they might be involved with distal 22q11.2 microdeletion syndrome. A cluster of low-copy



FIGURE 5

Results of Sanger sequencing. (A) Father (B-D). Heterozygous *RPS6KA3* gene variant (C) 898C>T (NM_004,586.3) in the mother, the elder sister, and the proband (the variant was marked with a red arrow).



FIGURE 6

Fold change was used for calculating mRNA relative expression. The levels of mRNA relative expression were significantly lower in the mother's and the elder sister's blood samples, and the levels of mRNA relative expression were significantly higher in the proband's blood sample. repeats (LCRs) from A to H in chromosome 22q11.2 (chr22: 17,900,001-25,900,000) mediated nonallelic homologous recombination and 22q11.2 chromosomal rearrangements. In the light of a systematic clinical overview of ClinGen curation (Xue et al., 2021), this CNV interval region contained the distal type II (E-F) region (chr22:23,119,414-23,649,111), and overlapped with the distal type I (D-E/F) region (chr22:21,917,117-23,649,111). Deletion of distal type I (D-E/F) or type II (E-F) regions may exhibit variable clinical phenotypes (mild, moderate, or even normal): prenatal growth restriction, short stature, ID, language delay, dysmorphic facial features, skeletal anomalies, cardiovascular defects, behavior problems, genitourinary anomalies, feeding problems, hypotonia, immune deficiency, and hypocalcemia (Burnside, 2015). Taken together, it was tempting to speculate that the CNV in the mother and elder sister made their phenotypes worse than the proband.

The two sisters showed different XCI patterns. We did not find a correlation between XCI patterns and phenotypic severity. It should

P51812 KS6A3_Human/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P P18654 KS6A3_Mouse/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P A0A212U9Z1 A0A212U9Z1_Cat/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P F6RET3 F6RET3_Horse/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P K7DQL9 K7DQL9_Chimpanzee/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P A0A1D5NWT4 A0A1D5NWT4_CHICK/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P A0A286XBX2 A0A286XBX2_Guinea pig/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P A0A452FNF2 A0A452FNF2_Goat/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P
A0A212U9ZI A0A212U9ZI _Cat/1-740 L GMPOFL SPEADSLLRMLFKRMPANRLGAGP F6RET3 F6RET3_Horse/1-740 L GMPOFL SPEADSLLRMLFKRMPANRLGAGP K7DQL9 K7DQL9_Chimpanzee/1-740 L GMPOFL SPEADSLLRMLFKRMPANRLGAGP A0A1D5NWT4 A0A1D5NWT4_CHICK/1-740 L GMPOFL SPEADSLLRMLFKRMPANRLGAGP A0A286XBX2 A0A286XBX2_Guinea pig/1-740 L GMPOFL SPEADSLLRMLFKRMPANRLGAGP A0A452FNF2 A0A452FNF2_Goat/1-740 L GMPOFL SPEADSLLRMLFKRMPANRLGAGP
F6RET3 F6RET3_Horse/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P K7DQL9 K7DQL9_Chimpanzee/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P A0A1D5NWT4 A0A1D5NWT4_CHICK/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P A0A286XBX2 A0A286XBX2_Guinea pig/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P A0A452FNF2 A0A452FNF2_Goat/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P
K7DQL9 K7DQL9_Chimpanzee/1-740 L GMPOFL SPEADSLLRMLFKRMPANRLGAGP A0A1D5NWT4 A0A1D5NWT4_CHICK/1-740 L GMPOFL SPEADSLLRMLFKRMPANRLGAGP A0A286XBX2 A0A286XBX2_Guinea pig/1-740 L GMPOFL SPEADSLLRMLFKRMPANRLGAGP A0A452FNF2 A0A452FNF2_Goat/1-740 L GMPOFL SPEADSLLRMLFKRMPANRLGAGP
A0A1D5NWT4 A0A1D5NWT4_CHICK/1-740 L GMPQFL SPEADSLLRMLFKRNPANRLGAGP A0A286XBX2 A0A286XBX2_Guinea pig/1-740 L GMPQFL SPEADSLLRMLFKRNPANRLGAGP A0A452FNF2 A0A452FNF2_Goat/1-740 L GMPQFL SPEADSLLRMLFKRNPANRLGAGP
A0A286XBX2 A0A286XBX2_Guinea pig/1-740 L GMPQFL SPEAQSLLRMLFKRNPANRLGAGP A0A452FNF2 A0A452FNF2_Goat/1-740 LGMPQFLSPEAQSLLRMLFKRNPANRLGAGP
A0A452FNF2 A0A452FNF2_Goat/1-740 L G M P Q F L S P E A Q S L L R M L F K R N P A N R L G A G P
A0A4X2L1B5 A0A4X2L1B5_Wombat)/1-740 <mark>LGMPQFLSPEAQSLI</mark> RMLFKRNPANRLGAGP
A0A3Q7STG7 A0A3Q7STG7_Red fox)/1-740 L G M P G F L S P E A G S L L R M L F K R N P A N R L G A G P
FIGURE 7
In total, 30 amino acids surrounded the variant position (marked with a black box).

TABLE 1 Summary of genotype and phenotypes of the three female patients.

Genotype	Proband	Elder sister	Mother
<i>RPS6KA3</i> variant (NM_004,586.3)	c.898C>T	c.898C>T	c.898C>T
CNV	-	del (22) (q11.22-q11.23) involved with distal 22q11.2 microdeletion syndrome	del (22) (q11.22-q11.23) involved with distal 22q11.2 microdeletion syndrome
XCI pattern	Extremely skewed	Random	ND
mRNA relative expression (<i>RPS6KA3</i> exon 14)	Significantly higher	Significantly lower	Significantly lower
Phenotypes of CLS (have not been reported)			
Irregular menstruation	+	+	+
Phenotypes of CLS (previously reported)			
Cognitive impairment	Simple communication, little literacy, poor memory, ID	No communication or literacy, poor memory, ID	No communication or literacy, poor memory, ID
Dysmorphic facial features	Mild	Moderate	Moderate
Stature	Normal (161 cm)	Short stature (142 cm)	Short stature (140 cm)
Fingers	-	Markedly tapered	Markedly tapered
Skeletal anomalies	-	-	Scoliosis
SIDEs	-	-	-
Cardiovascular diseases	Mild mitral and tricuspid valve regurgitation	Mild mitral and tricuspid valve regurgitation	Mild mitral and tricuspid valve regurgitation
Diabetes	-	-	-
Seizures	-	-	-
Hearing or vision problems	-	-	-
Growth retardation	-	+	ND

-: negative; +: positive; XCI, X-chromosome inactivation; ND; no data; SIDEs, stimulus-induced drop episodes; ID, intellectual disability.

be noted that the XCI pattern might be different in different tissues. Thus, XCI analysis in lymphocytes did not rule out this possibility. Also, the connection between the levels of mRNA (expressed by mutant *RPS6KA3* in blood samples) and phenotypic severity should be studied further. As we know, mRNA of the *RPS6KA3* gene is highly expressed in skeletal muscle. But we did not perform muscle biopsies, as we thought that it was kind of hard to get some tissues such as skeletal muscle, skin, or brain.

To sum up, severity of clinical features may be markedly variable. If possible, getting some tissues such as skeletal muscle or skin may be more useful for further genetic studies. These three female patients exhibited variable degrees of severity. Clinical characteristics of our CLS patients expanded the phenotypic spectrum of CLS. We hoped that our phenotypic and genetic studies could improve the comprehensive understanding of CLS for pediatricians.

Declarations

Ethics approval and consent to participate

All study procedures were approved by the ethical committee of Yiwu maternity and child health care hospital.

Consent for publication

Written informed consent of publication was obtained from the patients' guardian signed (the father). Also the guardian was informed and agreed to publish patients' photos.

Data availability statement

The datasets for this article are not publicly available due to concerns regarding participant/patient anonymity. Requests to access the datasets should be directed to the corresponding author.

Ethics statement

The studies involving human participants were reviewed and approved by the Ethical Committee of Yiwu Maternity and Child Health Care Hospital. Written informed consent to participate in this study was provided by the participants' legal guardian/next of kin. Written informed consent was obtained from the individual(s), and minor(s)' legal guardian/next of kin, for the publication of any potentially identifiable images or data included in this article.

Author contributions

YC and HJ contributed equally to the manuscript, YC and KW wrote the main manuscript text, HW carried out the molecular genetic experimental. HJ and DW prepared the

References

Ambalavanan, A., Chaumette, B., Zhou, S., Xie, P., He, Q., Spiegelman, D., et al. (2019). Exome sequencing of sporadic childhood-onset schizophrenia suggests the contribution of X-linked genes in males. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* 180 (6), 335–340. doi:10.1002/ajmgb.32683

Boulos, T., Moukarzel, Y., Yammine, T., Salem, N., Souaid, M., and Farra, C. (2021). Novel missense mutation c.1784A>G, p.Tyr595Cys in RPS6KA3 gene responsible for Coffin-Lowry syndrome in a family with variable features and diabetes 2. *Clin. Dysmorphol.* 30 (1), 32–35. doi:10.1097/MCD.00000000000343

Burnside, R. D. (2015). 22q11.21 deletion syndromes: A review of proximal, central, and distal deletions and their associated features. *Cytogenet. Genome Res.* 146 (2), 89–99. doi:10.1159/000438708

Castillon, C., Lunion, S., Desvignes, N., Hanauer, A., Laroche, S., and Poirier, R. (2018). Selective alteration of adult hippocampal neurogenesis and impaired spatial pattern separation performance in the RSK2-deficient mouse model of

clinical data and imaging data. KW contributed to the checking of revision, genetic evaluation, and genetic databases analysis. KW critically revised the manuscript. All authors reviewed the manuscript. All authors have read and approved the final manuscript.

Acknowledgments

We are grateful to the patients and their families for participation in this study, as well as the help of all the physicians in the course of the medical treatment. We wish to thank the staffs of Chigene (Beijing) Translational Medical Research Center Co. Ltd. for assisting with whole-exome sequencing and Sanger sequencing.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors, and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fgene. 2022.900226/full#supplementary-material

Gschwind, M., Foletti, G., Baumer, A., Bottani, A., and Novy, J. (2015). Recurrent nonconvulsive status epilepticus in a patient with Coffin-Lowry Syndrome. *Mol. Syndromol.* 6 (2), 91–95. doi:10.1159/000430429

Hanauer, A., and Young, I. D. (2002). Coffin-lowry syndrome: Clinical and molecular features. J. Med. Genet. 39 (10), 705–713. doi:10.1136/jmg.39. 10.705

Hunter, A. G. (2002). Coffin-lowry syndrome: A 20-year follow-up and review of long-term outcomes. Am. J. Med. Genet. 111 (4), 345-355. doi:10.1002/ajmg.10574

Coffin-Lowry syndrome. Neurobiol. Dis. 115, 69-81. doi:10.1016/j.nbd.2018. 04.007

Fischer, M., and Raabe, T. (2018). Animal models for coffin-lowry syndrome: RSK2 and nervous system dysfunction. *Front. Behav. Neurosci.* 12, 106. doi:10.3389/fnbeh.2018.00106

Kesler, S. R., Simensen, R. J., Voeller, K., Abidi, F., Stevenson, R. E., Schwartz, C. E., et al. (2007). Altered neurodevelopment associated with mutations of RSK2: A morphometric MRI study of coffin-lowry syndrome. *Neurogenetics* 8 (2), 143–147. doi:10.1007/s10048-007-0080-6

Li, H., Jia, J., and Yang, Z. (2016). Mini-mental state examination in elderly Chinese: A population-based normative study. *J. Alzheimers Dis.* 53 (2), 487–496. doi:10.3233/JAD-160119

Lv, Y., Zhu, L., Zheng, J., Wu, D., and Shao, J. (2019). Growth concerns in coffinlowry syndrome: A case report and literature review. *Front. Pediatr.* 6, 430. doi:10. 3389/fped.2018.00430

Mikhail, F. M., Burnside, R. D., Rush, B., Ibrahim, J., Godshalk, R., Rutledge, S. L., et al. (2014). The recurrent distal 22q11.2 microdeletions are often de novo and do not represent a single clinical entity: A proposed categorization system. *Genet. Med.* 16 (1), 92–100. doi:10.1038/gim.2013.79

Pangman, V. C., Sloan, J., and Guse, L. (2000). An examination of psychometric properties of the mini-mental state examination and the standardized mini-mental state examination: Implications for clinical practice. *Appl. Nurs. Res.* 13 (4), 209–213. doi:10.1053/apnr.2000.9231

Pereira, P. M., Schneider, A., Pannetier, S., Heron, D., and Hanauer, A. (2010). Coffin-Lowry syndrome. *Eur. J. Hum. Genet.* 18 (6), 627–633. doi:10.1038/ejhg.2009.189

Poirier, R., Jacquot, S., Vaillend, C., Soutthiphong, A. A., LibbeyM.DaviS, S., et al. (2007). Deletion of the Coffin-Lowry syndrome gene Rsk2 in mice is associated with

impaired spatial learning and reduced control of exploratory behavior. Behav. Genet. 37 (1), 31–50. doi:10.1007/s10519-006-9116-1

Richards, S., Aziz, N., Bale, S., Bick, D., Das, S., Gastier-Foster, J., et al. (2015). Standards and guidelines for the interpretation of sequence variants: A joint consensus recommendation of the American college of medical genetics and genomics and the association for molecular pathology. *Genet. Med.* 17 (5), 405–424. doi:10.1038/gim.2015.30

Rojnueangnit, K., Jones, J. R., Basehore, M. J., and Robin, N. H. (2014). Classic phenotype of Coffin-Lowry syndrome in a female with stimulus-induced drop episodes and a genotype with preserved N-terminal kinase domain. *Am. J. Med. Genet. A* 164A (2), 516–521. doi:10.1002/ajmg.a.36299

Tos, T., Alp, M. Y., Aksoy, A., and HAnAuer, A. (2015). A familial case of Coffin-Lowry syndrome caused by RPS6KA3 C.898C>T mutation associated with multiple abnormal brain imaging findings. *Genet. Couns.* 26 (1), 47–52.

Wang, Y., Martinez, J. E., Wilson, G. L., He, X. Y., Tuck-Muller, C. M., Maertens, P., et al. (2006). A novel RSK2 (RPS6KA3) gene mutation associated with abnormal brain MRI findings in a family with Coffin-Lowry syndrome. *Am. J. Med. Genet. A* 140, 1274–1279. doi:10.1002/ajmg.a.31266

Xue, J., Shen, R., Xie, M., Liu, Y., Zhang, Y., Gong, L., et al. (2021). 22q11.2 recurrent copy number variation-related syndrome: A retrospective analysis of our own microarray cohort and a systematic clinical overview of ClinGen curation. *Transl. Pediatr.* 10 (12), 3273–3281. doi:10.21037/tp-21-560