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# Methodological quality assessment of genetic studies on brain arteriovenous malformation related hemorrhage: A cross-sectional study

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**Objectives:** Rupture of a brain arteriovenous malformation (bAVM) can cause intracranial hemorrhage and severe clinical outcomes. At present, the mechanisms of bAVM-related hemorrhage are poorly understood. This study aimed to summarize the potential genetic risk factors for bAVM-related hemorrhage and appraise the methodological quality of existing genetic studies on bAVM-related hemorrhage using a cross-sectional design.

**Methods:** A systematic literature search was conducted on genetic studies associated with bAVM-related hemorrhage published in PubMed, Embase, Web of Science, China National Knowledge Internet, and Wangfang databases, up to November 2022. Subsequently, a cross-sectional study was performed to describe the potential candidate genetic variants of bAVM associated with risk of hemorrhage and to evaluate the methodological quality of the identified studies using the Newcastle–Ottawa quality assessment scale and Q-genie tool.

**Results:** Of the 1811 records identified in the initial search, nine studies met the filtering criteria and were included. Twelve single nucleotide polymorphisms (SNPs), including *IL6* rs1800795, *IL17A* rs2275913, *MMP9* rs9509, *VEGFA* rs1547651, and *EPHB4* rs314353, rs314308, and rs314313, were associated with bAVM-related hemorrhage. However, only 12.5% of the evaluated SNPs showed statistical power> 0.80 ( $\alpha$  = 0.05). Methodological quality assessment revealed significant flaws in the designs of the included studies, such as less reliable representativeness of recruited individuals, short follow-up periods in cohort studies, and less comparability between groups of hemorrhagic and non-hemorrhagic patients.

**Conclusion:** *IL1B, IL6, IL17A, APOE, MMP9, VEGFA* and *EPHB4* were potentially associated with bAVM-related hemorrhage. The methodological designs of the analyzed studies required improvement in order to obtain more reliable results.

Abbreviations: bAVM, brain arteriovenous malformation; CGAS, candidate gene association studies; CNKI, China National Knowledge Infrastructure; GWAS, genome-wide association studies; NOS, Newcastle–Ottawa scale; OR, odds ratio; SNPs, single nucleotide polymorphisms; VEGF, vascular endothelial growth factor; WES, whole exome sequencing; 95% CI, 95% confidence interval.

Regional alliances and rare disease banks need to be established to recruit large numbers of bAVM patients (especially familial and extreme-trait cases) in a multicenter, prospective cohort study with an adequate follow-up period. Furthermore, it is important to use advanced sequencing techniques and efficient measures to filter candidate genetic variants.

### KEYWORDS

brain arteriovenous malformation, intracranial hemorrhage, genetics, methodological quality, rupture

## **1** Introduction

The most common and severe manifestation of a brain arteriovenous malformation (bAVM) is its rupture, which is also the leading cause of intracranial hemorrhage in children and young adults. High pressure blood flow from the feeding arteries of the bAVM floods directly to the draining veins through the malformed nidus, causing the development of abnormal shear stress due to lack of capillary structure within the anomalous nidus, ultimately resulting in its rupture (Rutledge et al., 2014). Past observational studies have reported a 1%-3% annual incidence of bAVM-related hemorrhage in unruptured and untreated patients, whereas the reported risk was much higher in individuals with ruptured bAVM(2). Current treatments, including microsurgery, endovascular embolization, and stereotactic radiosurgery aim to reduce the risk of hemorrhage and eradicate existing lesions. Although microsurgery offers the advantage of a higher rate of complete obliteration and elimination of bAVM-related hemorrhage compared to the other treatments, craniotomy is a highly traumatic procedure resulting in a longer hospitalization as well as substantial morbidity and mortality during the perioperative period (van Beijnum et al., 2011; Derdeyn et al., 2017). Thus, it is imperative to identify risk factors for bAVM rupture as early as possible.

Prior hemorrhage has been reported to be associated with a higher rate of subsequent hemorrhage as a strongly predictive factor (Chen et al., 2020). Existing evidence suggests that angioanatomic features of bAVM, including large size, deep venous drainage, few draining veins, and coexisting arterial aneurysm, contribute to its rupture (Krithika and Sumi, 2021). Several studies have investigated and discovered genetic variants of inflammation- or angiogenesisrelated genes that could potentially influence bAVM rupture by accelerating growth and modifying lesion behavior to promote disease pathogenesis (Pawlikowska et al., 2004; Achrol et al., 2006; Pawlikowska et al., 2006; Kim et al., 2009; Weinsheimer et al., 2009; Gong et al., 2011; Li et al., 2012; Sun et al., 2012; Delev et al., 2017). However, due to the low prevalence and incidence of bAVM, most genetic studies on bAVM recruited small samples of patients and were prone to selection bias, resulting in inconsistent results. In addition, different research designs may yield conflicting results and may have varying methodological quality. Therefore, existing genetic studies on bAVM cannot always be considered as a reliable source of evidence. A well-performed research that provides reliable and high-quality information can help medical practitioners in improving their understanding of the nature of a particular disease as well as assist them in making appropriate treatment decisions. Before accepting and using any scientific evidence, healthcare professionals, medical managers, health policymakers, and even patients should evaluate the methodological quality of the referred studies.

The present study performed a cross-sectional survey to summarize the genetic factors influencing the risk of hemorrhage associated with bAVM and evaluate the methodological rigor of the included studies using the Newcastle–Ottawa quality assessment scale (NOS) and Q-genie tool. Our aim was to summarize the current information regarding genetic risk of bAVM-related hemorrhage, discuss potential research directions, and provide insights into how to improve the methodological quality in future studies investigating the risk of bAVM-related hemorrhage or hemorrhagic stroke caused by other diseases.

## 2 Materials and methods

### 2.1 Eligibility criteria

This study was registered with PROSPERO (CRD42021258353). All included articles were case-control or cohort studies recruiting individuals of any ethnic group and focused on the genetic risk factors associated with bAVM-related hemorrhage, using the methods of candidate gene association studies (CGAS), genomewide association studies (GWAS) or whole exome sequencing (WES). Individuals diagnosed with bAVM based on a recognized criteria were recruited and divided into ruptured and unruptured groups (Atkinson et al., 2001). Only original papers with accurate and sufficient genotyping data that could allow the calculation of odds ratios (ORs) and 95% confidence intervals (95% CIs) were included in the analysis. Conference abstracts, reviews, metaanalyses, protocols, case reports, and animal studies were excluded. Whenever there were duplicate or overlapping papers published by the same researcher(s), the most up-to-date version was included for evaluation.

### 2.2 Literature search strategy

We identified potential eligible studies using the search terms ("Cerebral AVM" or "cerebral arteriovenous malformation" or "brain AVM" OR "brain arteriovenous malformation") and ("gene" or "Variation" or "polymorphisms" or "SNPs") across five electronic databases: PubMed, Embase, Web of Science, China National Knowledge Internet (CNKI) and Wanfang Data. The search was limited to published citations in English or Chinese.



Thereafter, potentially relevant studies that were not obtained in the initial searches were manually retrieved from the references of the candidate papers.

## 2.3 Data extraction

The required information (e.g., author, publication year, study design, studied genes, and SNPs) was extracted from the selected studies. When genotype frequencies of variants could not be obtained from the published papers, the risk allele frequencies of SNPs were utilized to estimate the number of cases per genotype category and calculate the OR (95% CI) using STATA 14.0 (Stata Corporation, College Station, TX, USA).

### 2.4 Methodological quality assessment

Statistical power  $(1-\beta)$  was calculated according to the genetic models used in the original studies *via* a two-sided Z test, using an online software (http://powerandsamplesize.com/) to evaluate the quality of studies, with the type I error rate ( $\alpha$ ) set at 5%. The methodological quality assessment of all included studies was mainly based on the NOS (Stang, 2010) and Q-genie tool (version 1.1) (Sohani et al., 2015). NOS is a validated appraisal tool for non-randomized studies, with eight items categorized into three dimensions: selection, comparability, and outcome (in cohort studies) or exposure (in case-control studies). A score  $\geq 6$  is regarded as high quality. The Q-genie tool (version 1.1) was used to assess the quality of the genetic association studies. It is comprised of 11 items scored from 0–7, and a total score  $\leq 35$  indicates poor quality.

Study screening, data extraction, and methodological quality assessment were independently completed by two authors, and if

there was a disagreement during the process, the third senior investigator resolved the issue through re-evaluation and discussion.

## **3** Results

# 3.1 Literature selection and characteristics of included studies

The initial search indicated 1881 records through a single database check until November 2022. Among these, nine studies met the eligibility criteria and were included in the final analysis. This was followed by a manual reference-list screening; however, no additional studies were found to satisfy the filtering criteria. The detailed procedure of literature selection followed in this study is displayed in Figure 1.

Six case-control studies and three cohort studies were identified, incorporating 1214 bAVM individuals from North America, Europe, and China (Table 1). The case-control studies divided patients into two groups based on the presence or absence of hemorrhage, and genotyping was performed to test the association between SNPs and bAVM rupture. In these studies (Pawlikowska et al., 2004; Weinsheimer et al., 2009; Gong et al., 2011; Li et al., 2012; Sun et al., 2012; Delev et al., 2017), a total of 984 cases had been recruited, including 529 with hemorrhage and 455 without hemorrhage. In contrast, the cohort studies relied on a prospective follow-up of the included bAVM patients until a new intracranial hemorrhage event occurred. These studies attempted to identify the association between genetic variants and the risk of new rupture during the natural process of bAVM. Furthermore, all three cohort studies (Achrol et al., 2006; Pawlikowska et al., 2006; Kim et al., 2009) belonged to the same research team, which initially recruited 237 non-hemorrhagic and 173 hemorrhagic cases. Less than

TABLE 1 Summary of the included genetic studies on brain arteriovenous malformation related hemorrhage.

Gene	Study (Year)	Country	Language	Involved medical center	Design	Follow- up (year)	SNP Selection		nple ize	SNPs	Chromosome position, alleles, amino acid change	Gene function
				Center		(year)		ICH	non- ICH			
Genes in	volving in infl	ammatory p	athway									
IL6	Pawlikowska et al. (2004)	USA	English	Single	Case- control	-	Literature review, Unigene and dbSNP database searches	73	107	rs1800795	chr7:22727026, C>G, -	Coding a cytokine functioning in inflammation and maturation of B cells.
	Achrol et al. (2006)	USA	English	Two	Cohort	0.3	NA	18	262			
	Pawlikowska et al. (2004)	USA	English	Single	Case- control	-	Literature review, Unigene and dbSNP database searches	73	107	rs1800796	chr7:22726627, G>C, -	_
	Achrol et al. (2006)	USA	English	Two	Cohort	0.3	NA	18	262			
IL10	Pawlikowska et al. (2004)	USA	English	Single	Case- control	-	Literature review, Unigene and dbSNP database searches	73	107	rs1800896	chr1:206773552, T>C, -	Coding a cytokine mainly produced by monocytes, taking effects in immunoregulation and inflammation
TNF	Pawlikowska et al. (2004)	USA	English	Single	Case- control	-	Literature review, Unigene and dbSNP database searches	73	107	rs361525	chr6:31575324, G>A, -	Coding a multifunctional proinflammatory cytokine, involved in cell proliferation, differentiation,
	Achrol et al. (2006)	USA	English	Two	Cohort	0.3	NA	18	262			apoptosis, lipid metabolism, and coagulation.
	Pawlikowska et al. (2004)	USA	English	Single	Case- control	-	Literature review, Unigene and dbSNP database searches	73	107	rs1800629	chr6:31575254, G>A, -	_
	Achrol et al. (2006)	USA	English	Two	Cohort	0.3	NA	18	262			
APOE	Pawlikowska et al. (2006)	USA	English	Two	Cohort	0.3	NA	18	266	rs429358	chr19:45411941, T>C, p.C130R	Coding a major apoprotein of the chylomicron essential for the
										rs11542041	chr19:45411947, C>T, p.R158C	catabolism of triglyceride-rich lipoprotein.
IL1B	Kim et al.	USA	English	Two	Cohort	3.1	Putative functional	27	383	rs1143627	chr2:112836810, C>T, -	Coding a mediated cytokine produced by macrophages, involved in cell
	(2009)						effect and previous associated phenotypes			rs16944	chr2:112837290, C>T, -	proliferation, differentiation, and
										rs1143634	chr2:112832813, T>C, -	apoptosis.
IL17A	Li et al. (2012)	China	Chinese	NA	Case- control	-	NA	30	23	rs2275913	chr6:52186235, G>A, -	Coding a cytokine produced by activated T cells, eliciting crucial impacts on innated immune defenses

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Gene	Study (Year)	Country	Language	Involved medical center	Design	Follow- up (year)	SNP Selection		nple ize	SNPs	Chromosome position, alleles, amino acid change	Gene function	
				center		(year)		ICH	non- ICH				
MMP9	Sun et al. (2012)	China	English	Two	Case-	-	HapMap	181	130	rs3918241	chr20:46007096, T>A, -	Coding a protein in matrix	
					control					rs1805088	chr20:46008985, C>T, p.A20V	metalloproteinase family, playing role in the breakdown of extracellular matrix in physilogical and also diseas	
										rs17576	chr20:46011586, A>G, p.Q279R	processes.	
										rs3918254	chr20:46011752, C>T, -		
										rs3787268	chr20:46013092, G>A, -		
										rs17577	chr20:46014472, G>A, p.R668Q		
										rs13925	chr20:46016326, G>A, -	-	
										rs9509	chr20:46016514, T>C, -	-	
										rs17035945	chr3:12153128, C>T, -	-	
										rs3755724	chr3:12159406, C>T, -		
	Sun et al. (2012)	China	English	Single	Case- control	-	НарМар	181	130	rs11923383	chr3:12162637, A>G, -		
Genes in	volving in ang	iogenesis pa	athway										
ANGPT2	Pawlikowska et al. (2004)	USA	English	Single	Case- control	-	Literature review, Unigene and dbSNP database searches	73	107	rs3020221	chr8:6521242, C>T, p.245=	Coding an antagonist of angiopoietin 1 implicated in the inflammation- and angiogenesis- related signaling pathways.	
FLT4	Pawlikowska et al. (2004)	USA	English	Single	Case- control	-	Literature review, Unigene and dbSNP database searches	73	107	rs448012	chr5:180619344, G>C, p.H890Q	Coding a tyrosine kinase receptor for vascular endothelial growth factors,	
							database searches			rs1130379	chr5:180612606, C>T, p.R1146H	involved in lymphangiogenesis.	
KDR	Pawlikowska et al. (2004)	USA	English	Single	Case- control	-	Literature review, Unigene and dbSNP database searches	73	107	rs1870377	chr4:55106807, T>A, p.Q472H	Also called VEGFR, coding a tyrosin kinase receptor and the main mediato	
							uatabase searches			rs2034964	chr4:55110484, C>T, p.D392N	of endothelial proliferation, migration and other biological function induced by VEGF.	
TIE2	Pawlikowska et al. (2004)	USA	English	Single	Case- control	-	Literature review, Unigene and dbSNP database searches	73	107	rs682632	chr9:27183465, A>C, p.Q346P	Coding a tyrosine kinase receptor, binding its ligand angiopoietin-1 to	

(Continued on following page)

Gene	Study (Year)	Country	Language	Involved medical center	Design	Follow- up (year)	SNP Selection		mple ize	SNPs	Chromosome position, alleles, amino acid change	Gene function	
				center		(year)		ICH	non- ICH				
										rs3837240	chr9:27109316, dupG, -	mediate the signaling in embryonic	
										rs10967719	chr9:27108812, G>T, -	vascular development.	
EPHB4	Weinsheimer	USA	English	Two	Case-	-	НарМар	56	90	rs314346	chr7:100800895, C>T, -	Coding a Ephrin receptor, binding its	
	et al. (2009)				control					rs314353	chr7:100808522, A>G, -	ligand Ephrins to mediate numerous developmental process including	
										rs2230585	chr7:100812975, G>A, -	nervous system and angiogenesis.	
										rs144173	chr7:100818628, A>G, -	-	
										rs314308	chr7:100823256, C>T, -	-	
										rs2250818	chr7:100824534, G>A, -	-	
										rs314313	chr7:100825743, T>C, -	-	
										rs2247445	chr7:100829641, G>A, -	-	
VEGFA	Pawlikowska et al. (2004)	USA	English	Single	Case- control	-	Literature review, Unigene and dbSNP database searches	73	107	rs699947	chr6:43768652, A>C, -	Coding the vascular endothelial growth factor A, a heparin-binding protein, inducing endothelial proliferation and	
	Gong et al.	China	English	Single	Case-	-	НарМар	181	130	rs1547651	chr6:43762907, A>T, -	migration in vessels in both both physiological and pathological	
	(2011)				control					rs2010963	chr6:43770613, C>G, -	angiogenesis.	
										rs1413711	chr6:43772941, T>C, -	-	
										rs833069	chr6:43774842, T>C, -	_	
										rs3024994	chr6:43775770, C>T, -		
										rs3025010	chr6:43779840, T>C, -		
										rs3025030	chr6:43782850, G>C, -		
										rs3025035	chr6:43783622, C>T, -		
										rs3025039	chr6:43784799, C>T, -		
TGFβ1	Li et al. (2012)	China	Chinese	NA	Case- control	-	NA	30	23	rs1800469	chr19:41354391, A>G, -	Coding a ligand transforming growth factor- $\beta$ , regulating cell proliferation, differentiation and growth, including vascular endothelial cells.	

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Coding a transmembrane protein and a menber of NOTCH family, playing a forming a complex with TGF- $\beta$ R1 and regulate gene transcription related to cell proliferation, differentiation and then binding its ligand TGF- $\beta$  to role in vascular, renal and hepatic Coding a transmembrane protein other biological function. Gene function development. mino acid chang chr6:32222629, A>G, chr3:30605668, A>G, chr6:32222440, T>C, Chromosome osition, allel rs3087465 rs443198 rs915895 SNPs -uou E 23 65 Sample £ 64 30 **SNP Selection** HapMap NΑ Followyear dn ratio; 95% CI,95%confidence interval; NA, not available Design Case-control Case-control Involved cal center Single NA Language Chinese English single nucleotide polymorphisms; OR, odd Country German China Li et al. (2012) et al. (2017) Study (Year) Delev NOTCH4 TGFBR2 Gene SNP,

25% of the patients were followed-up for over 2 years, at the end of which twenty-seven patients had experienced bAVM rupture and new intracranial hemorrhage events. All included studies were CGAS, including four single-center studies and four multi-center studies. The last study did not mention the source of patient recruitment.

## 3.2 Genetic characteristics

The nine CGASs evaluated SNPs in seven genes (IL6, IL10, TNF, APOE, IL1B, IL17A, MMP9) of the inflammatory pathway and nine genes (ANGPT2, FLT4, KDR, TIE2, EPHB4, VEGFA, *TGF* $\beta$ *1*, *TGF* $\beta$ *R2*, *NOTCH4*) of the angiogenic pathway (Table 1; Figure 2). Seven SNPs in five inflammatory genes were reported to be significantly associated with bAVM rupture, including APOE rs429358 (OR, 5.09; 95% CI, 1.46-17.70), IL1B rs1143627 (OR, 4.01; 95% CI, 1.31-12.29), and IL17A rs2275913 (OR, 0.20; 95% CI, 0.05-0.66) in dominant models, and IL6 rs1800795 (OR, 2.43; 95% CI, 1.04-5.68), IL1B rs16944 (OR, 3.23; 95% CI, 1.70-6.14), IL1B rs1143634 (OR, 1.79; 95% CI, 1.21-2.66), and MMP9 rs9509 (OR, 0.19; 95% CI, 0.05-0.66) in recessive models. Five SNPs in two angiogenic genes were discovered to contribute to bAVM-related hemorrhage: VEGFA rs1547651 (OR, 2.11; 95% CI, 1.01-4.42) in dominant models, and rs314346 (OR, 1.67; 95% CI, 1.04-2.68), rs314353 (OR, 1.79; 95% CI, 1.11-2.89), rs314308 (OR, 0.36; 95% CI, 0.20-0.65), and rs314313 (OR, 0.45; 95% CI 0.25-0.79) in allelic models of EPHB4.

### 3.3 Methodological quality assessment

The statistical power of each polymorphism was calculated using the respective genotype model. The powers of the three SNPs in *VGEFA* could not be calculated as their ORs and 95% CIs were derived using additive models, with  $\alpha$  set at 0.05. The 44 SNPs that were not associated with bAVM-related hemorrhage failed to reach sufficient statistical power (range, 0.050–0.783). Among the remaining 12 candidate variants, *APOE* rs429358 (power, 0.248), *IL1B* rs1143627 (power, 0.532), *IL1B* rs16944 (power, 0.451), *IL1B* rs1143634 (power, 0.400) and *EPHB4* rs314346 (power, 0.765), there were risks of false negatives. Only seven SNPs (12.5%) demonstrated powers greater than 0.80, with *IL6* rs1800795, *IL17A* rs2275913, *EPHB4* rs314308, and rs314313 reaching powers >0.90.

Although two-thirds of the included studies were judged to be of high quality (average 6.1 stars) after assessment by NOS (Table 2), all of them were classified as high-bias studies using the Q-Genie tool (Figure 3). Regarding methodology, most studies performed well in identification of patients with bAVM, as well as the methods for identification of genetic variants. However, there was still much room for improvement in the following areas: representativeness for the gene and SNP selection procedure as well as patients with bAVM; comparability between hemorrhagic and non-hemorrhagic individuals as confounding factors, as these were hardly taken into sufficient consideration; and a longer follow-up period in cohort studies.

Gene	Gene Study (Year) Si		Genetype (Wildtype / Heterozygote / S Homozygote)		Model			Odds ratio (95% Confidence	Statistical
			Hemorrage	10/001 00				Interval)	power
Genes involv	ring in inflammtory pathway		-						
IL6	Pawlikowskaet al. (2004)	rs1800795	2/19/48	10/45/48	Recessive	•	1	2.43(1.04-5.68)*	0.967
	Achrolet al.(2006)		2001/6/10	14/89/148	Recessive	<b></b>		0.99(0.37-2.70)	0.050
	Pawlikowskaet al. (2004)	rs1800796	46/22/3	74/27/3	Dominant			1.34(0.70-2.56)	0.212
	Achrolet al.(2006)	131000130	2011/5/1	187/66/6	Dominant			1.42(0.51-3.97)	0.106
IL10	Pawlikowskaet al. (2004)	rs1800896	28/32/9	43/47/13	Dominant			1.05(0.56-1.95)	0.055
TNF	Pawlikowskaet al. (2004)	rs361525	60/10/0	92/12/0	Dominant			1.28(0.52-3.14)	0.107
	Achrolet al.(2006)		14/4/0	199/55/0	Dominant			1.03(0.33-3.27)	0.051
	Pawlikowskaet al. (2004)	rs1800629	60/9/0	78/24/2	Dominant	H.		0.45(0.20-1.31)	0.683
	Achrolet al.(2006)		14/4/0	199/55/0	Dominant			1.03(0.33-3.27)	0.051
APOE	Pawlikowskaet al.(2006)	rs429358	14/4/0	234/31/1	Dominant	•	-	5.09(1.46-17.70)*	0.248
		rs11542041	13/5/0	197/64/5	Dominant			1.10(0.67-1.80)	0.054
IL1B	Kimet al.(2009)	rs1143627	2010/6/10	80/178/116	Dominant	•	-	4.01(1.31-12.29)*	0.532
		rs16944	2011/5/9	116/177/76	Recessive	+ •	-	3.23(1.70-6.14)*	0.451
		rs1143634	2000/3/21	8/100/268	Recessive	<b></b>		1.79(1.21-2.66)*	0.400
IL17A	Liet al. (2012)	rs2275913	13/14/3	3/15/5	Dominant	H <b>e</b>		0.20(0.05-0.81)	0.930
MMP9	Sun et al. (2012)	rs3918241	125/39/1	87/31/1	Dominant	<b></b>		0.87(0.51-1.49)	0.123
		rs1805088	169/10/0	122/7/0	Dominant	· · · · · · · · · · · · · · · · · · ·		1.03(0.38-2.79)	0.051
		rs17576	10/57/65	10/29/56	Recessive	<b>⊢</b> ∎−−1		0.68(0.40-1.15)	0.608
		rs3918254	118/46/4	78/35/6	Dominant			0.81(0.49-1.33)	0.258
		rs3787268	66/89/25	53/54/20	Dominant			1.24(0.78-1.97)	0.287
		rs17577	135/40/2	96/31/2	Dominant			0.91(0.54-1.53)	0.089
		rs13925	133/43/2	93/29/3	Dominant			0.98(0.58-1.66)	0.051
		rs9509	113/58/5	79/36/10	Recessive	H <b>e</b>		0.19 (0.05-0.66)*	0.855
		rs17035945	138/36/2	99/31/0	Dominant			0.88(0.51-1.51)	0.111
		rs3755724	29/60/82	16/61/49	Dominant	▶ <b></b>		1.37(0.61-3.08)	0.342
	Sun et al. (2012)	rs11923383	114/54/8	73/42/10	Dominant	H.		0.76(0.48-1.22)	0.415
Genes involv	ving in angiogenesis pathway								
ANGPT2	Pawlikowskaet al. (2004)	rs3020221	24/30/9	36/44/14	Dominant			1.01(0.52-1.95)	0.051
FLT4	Pawlikowskaet al. (2004)	rs448012	71/1/0	102/1/0	Dominant			1.44(0.09-23.35)	0.063
AL-2017 11-2017		rs1130379	55/14/1	83/14/1	Dominant			1.51(0.68-3.33)	0.267
KDR	Pawlikowskaet al. (2004)	rs1870377	4/25/41	7/39/59	Recessive		_	1.10(0.60-2.03)	0.069
		rs2034964	72/0/0	104/2/0	Dominant			0.29(0.01-6.09)	0.050
TIE2	Pawlikowskaet al. (2004)	rs682632	0/2/69	0/1/104	Recessive			0.33(0.03-3.73)	0.213
		rs3837240	52/17/1	77/24/2	Dominant			1.03(0.51-2.06)	0.051
		rs10967719	49/13/1	74/18/1	Dominant			1.11(0.51-2.06)	0.064
			15/34/7	16/48/26	Allelic				0.765
EPHB4	Weinsheimeret al. (2009)	rs314346			Allelic			1.67(1.04-2.68)*	011.00
EPHB4	Weinsheimeret al. (2009)	rs314346 rs314353	10/37/9	12/42/36	Allelic			1.67(1.04-2.68)* 1.79(1.11-2.89)*	0.860
EPHB4	Weinsheimeret al. (2009)		10/37/9 10/28/18	12/42/36 9/43/38					
EPHB4	Weinsheimeret al. (2009)	rs314353			Allelic			1.79(1.11–2.89)*	0.860
EPHB4	Weinsheimeret al. (2009)	rs314353 rs2230585	10/28/18	9/43/38	Allelic Allelic			1.79(1.11–2.89)* 1.46(0.90–2.38)	0.860 0.501
EPHB4	Weinsheimeret al. (2009)	rs314353 rs2230585 rs144173 rs314308 rs2250818	10/28/18 10/32/14 39/15/2 25/27/4	9/43/38 10/43/37 41/33/16 51/33/6	Allelic Allelic Allelic Allelic Allelic	· · · · · · · · · · · · · · · · · · ·		1.79(1.11–2.89)* 1.46(0.90–2.38) 1.61(0.99–2.61) 0.36(0.20–0.65) 1.36(0.81–2.30)	0.860 0.501 0.695 0.992 0.317
EPHB4	Weinsheimeret al. (2009)	rs314353 rs2230585 rs144173 rs314308 rs2250818 rs314313	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3	9/43/38 10/43/37 41/33/16 51/33/6 40/41/9	Allelic Allelic Allelic Allelic Allelic Allelic			1.79(1.11–2.89)* 1.46(0.90–2.38) 1.61(0.99–2.61) 0.36(0.20–0.65) 1.36(0.81–2.30) 0.45(0.25–0.79)	0.860 0.501 0.695 0.992 0.317 0.939
ЕРНВ4	Weinsheimeret al. (2009)	rs314353 rs2230585 rs144173 rs314308 rs2250818	10/28/18 10/32/14 39/15/2 25/27/4	9/43/38 10/43/37 41/33/16 51/33/6	Allelic Allelic Allelic Allelic Allelic	· · · · · · · · · · · · · · · · · · ·		1.79(1.11–2.89)* 1.46(0.90–2.38) 1.61(0.99–2.61) 0.36(0.20–0.65) 1.36(0.81–2.30)	0.860 0.501 0.695 0.992 0.317
EPHB4 VEGFA	Weinsheimeret al. (2009) Pawlikowskaet al. (2004)	rs314353 rs2230585 rs144173 rs314308 rs2250818 rs314313	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3	9/43/38 10/43/37 41/33/16 51/33/6 40/41/9	Allelic Allelic Allelic Allelic Allelic Allelic	· · · · · · · · · · · · · · · · · · ·		1.79(1.11–2.89)* 1.46(0.90–2.38) 1.61(0.99–2.61) 0.36(0.20–0.65) 1.36(0.81–2.30) 0.45(0.25–0.79)	0.860 0.501 0.695 0.992 0.317 0.939
		rs314353 rs2230585 rs144173 rs314308 rs2250818 rs314313 rs2247445	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3 25/27/4	9/43/38 10/43/37 41/33/16 51/33/6 40/41/9 52/33/5	Allelic Allelic Allelic Allelic Allelic Allelic			1.79(1.11-2.89)* 1.46(0.90-2.38) 1.61(0.99-2.61) 0.36(0.20-0.65) 1.36(0.81-2.30) 0.45(0.25-0.79) 1.45(0.86-2.45)	0.860 0.501 0.695 0.992 0.317 0.939 0.421
	Pawlikowskaet al. (2004)	rs314353 rs2230585 rs144173 rs314308 rs2250818 rs314313 rs2247445 rs699947	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3 25/27/4 16/36/20 149/30/0 52/82/39	9/43/38 10/43/37 41/33/16 51/33/6 40/41/9 52/33/5 21/51/33 115/11/0 40/66/19	Allelic Allelic Allelic Allelic Allelic Allelic Allelic Recessive			1.79(1.11-2.89)* 1.46(0.90-2.38) 1.61(0.99-2.61) 0.36(0.20-0.65) 1.36(0.81-2.30) 0.45(0.25-0.79) 1.45(0.86-2.45) 0.84(0.43-1.62)	0.860 0.501 0.695 0.992 0.317 0.939 0.421 0.104
	Pawlikowskaet al. (2004)	rs314353 rs2230585 rs144173 rs314308 rs2250818 rs314313 rs2247445 rs699947 rs1547651	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3 25/27/4 16/36/20 149/30/0	9/43/38 10/43/37 41/33/16 51/33/6 40/41/9 52/33/5 21/51/33 115/11/0	Allelic Allelic Allelic Allelic Allelic Allelic Recessive Dominant			1.79(1.11-2.89)* 1.46(0.90-2.38) 1.61(0.99-2.61) 0.36(0.20-0.65) 1.36(0.81-2.30) 0.45(0.25-0.79) 1.45(0.86-2.45) 0.84(0.43-1.62) 2.11(1.01-4.42)*	0.860 0.501 0.695 0.992 0.317 0.939 0.421 0.104 0.873
	Pawlikowskaet al. (2004)	rs314353 rs2230585 rs144173 rs314308 rs2250818 rs314313 rs2247445 rs699947 rs1547651 rs2010963	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3 25/27/4 16/36/20 149/30/0 52/82/39 9/73/98 44/88/40	9/43/38 10/43/37 41/33/16 51/33/6 40/41/9 52/33/5 21/51/33 115/11/0 40/66/19 6/43/78 36/67/18	Allelic Allelic Allelic Allelic Allelic Allelic Recessive Dominant Dominant			$\begin{array}{l} 1.79(1.11-2.89)^{*}\\ 1.46(0.90-2.38)\\ 1.61(0.99-2.61)\\ 0.36(0.20-0.65)\\ 1.36(0.81-2.30)\\ 0.45(0.25-0.79)\\ 1.45(0.86-2.45)\\ 0.84(0.43-1.62)\\ 2.11(1.01-4.42)^{*}\\ 0.63(0.34-1.16)\\ 0.75(0.47-1.19)\\ 1.73(0.94-3.20)\\ \end{array}$	0.860 0.501 0.695 0.992 0.317 0.939 0.421 0.104 0.873 0.086
	Pawlikowskaet al. (2004)	rs314353 rs2230855 rs144173 rs314308 rs2250818 rs314313 rs2247445 rs699947 rs1547651 rs2010963 rs1413711 rs833069 rs3024994	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3 25/27/4 16/36/20 149/30/0 52/82/39 9/73/98 44/88/40 153/22/1	9/43/38 10/43/37 41/33/16 51/33/6 40/41/9 52/33/5 21/51/33 115/11/0 40/66/19 6/43/78 36/67/18 116/11/0	Allelic Allelic Allelic Allelic Allelic Allelic Recessive Dominant Dominant Recessive Addictive			$\begin{array}{l} 1.79(1.11-2.89)^*\\ 1.46(0.90-2.38)\\ 1.61(0.99-2.61)\\ 0.36(0.20-0.65)\\ 1.38(0.81-2.30)\\ 0.45(0.25-0.79)\\ 1.45(0.86-2.45)\\ 0.84(0.43-1.62)\\ 2.11(1.01-4.42)^*\\ 0.63(0.34-1.16)\\ 0.75(0.47-1.19)\\ 1.73(0.94-3.20)\\ 0.62(0.30-1.30)\\ \end{array}$	0.860 0.501 0.695 0.992 0.317 0.939 0.421 0.104 0.873 0.086 0.054 0.783 -
	Pawlikowskaet al. (2004)	rs314353 rs2230855 rs144173 rs314308 rs2250818 rs314313 rs2247445 rs699947 rs1547651 rs2010963 rs1413711 rs833069 rs3024994 rs3025010	10/28/18 10/32/14 39/15/2 25/27/4 16/36/20 149/30/0 52/82/39 9/73/98 44/88/40 153/22/1 97/72/11	9(43/38 10/43/37 41/33/16 51/33/6 40/41/9 52/33/5 21/51/33 115/11/0 40/86/19 6/43/78 36/67/18 116/11/0 67/54/6	Allelic Allelic Allelic Allelic Allelic Allelic Allelic Recessive Dominant Dominant Recessive Addictive Recessive			$\begin{array}{c} 1.79(1.11-2.89)^{*}\\ 1.46(0.90-2.38)\\ 1.61(0.99-2.61)\\ 0.36(0.20-0.65)\\ 1.36(0.81-2.30)\\ 0.45(0.28-0.79)\\ 1.45(0.86-2.45)\\ 0.84(0.43-1.62)\\ 2.11(1.01-4.42)^{*}\\ 0.63(0.34-1.16)\\ 0.75(0.47-1.19)\\ 0.75(0.47-1.19)\\ 1.73(0.94-2.09)\\ 0.62(0.30-1.30)\\ 1.31(0.47-3.65)\\ \end{array}$	0.860 0.501 0.695 0.992 0.317 0.939 0.421 0.104 0.873 0.086 0.054 0.783 - 0.783 -
	Pawlikowskaet al. (2004)	rs314353 rs2230585 rs144173 rs314308 rs2250818 rs2450818 rs247445 rs699947 rs1547651 rs2010963 rs1413711 rs833069 rs3024994 rs3025010 rs3025030	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3 25/27/4 16/36/20 149/30/0 52/82/39 9/73/98 44/88/40 153/22/1 97/72/11 13/1/42/4	9(43/38 10/43/37 41/33/16 51/33/6 40/41/9 52/33/5 21/51/33 115/11/0 40/66/7/8 6/43/78 36/67/18 116/11/0 67/54/6 81/39/6	Allelic Allelic Allelic Allelic Allelic Allelic Allelic Recessive Dominant Dominant Recessive Recessive Addictive			$\begin{array}{c} 1.79(1.11-2.89)^*\\ 1.46(0.90-2.38)\\ 1.61(0.99-2.61)\\ 0.36(0.20-0.65)\\ 1.36(0.81-2.30)\\ 0.45(0.28-0.79)\\ 1.45(0.88-2.45)\\ 0.84(0.43-1.62)\\ 2.11(1.01-4.42)^*\\ 0.63(0.34-1.16)\\ 0.75(0.47-1.19)\\ 1.73(0.94-3.20)\\ 0.62(0.30-1.30)\\ 1.31(0.47-3.65)\\ 1.47(0.96-2.26)\\ \end{array}$	0.860 0.501 0.695 0.992 0.317 0.939 0.421 0.104 0.873 0.086 0.054 0.783 - 0.129
	Pawlikowskaet al. (2004)	rs314353 rs2230585 rs144173 rs214308 rs214308 rs2143133 rs2247445 rs2010963 rs1547651 rs2010963 rs1547651 rs2010963 rs30254094 rs3025403 rs3025030	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3 25/27/4 16/36/20 149/30/0 52/82/39 9/73/98 44/88/40 153/22/1 97/72/11 131/42/4 122/47/5	9(43/38 10/43/37 41/33/16 51/33/6 40/41/9 52/33/5 21/51/33 115/11/0 40/66/19 6/43/78 116/11/0 67/54/6 81/39/6 95/30/1	Allelic Allelic Allelic Allelic Allelic Allelic Allelic Cominant Dominant Dominant Recessive Addictive Recessive Addictive		<b>→</b>	$\begin{array}{l} 1.79(1.11-2.89)^*\\ 1.46(0.90-2.38)\\ 1.61(0.99-2.61)\\ 0.36(0.20-0.65)\\ 1.36(0.81-2.30)\\ 0.45(0.25-0.79)\\ 0.45(0.25-0.79)\\ 1.45(0.85-2.45)\\ 0.84(0.43-1.62)\\ 2.11(1.01-4.42)^*\\ 0.63(0.34-1.16)\\ 0.75(0.47-1.19)\\ 1.73(0.47-1.19)\\ 1.73(0.47-3.65)\\ 1.47(0.96-2.26)\\ 3.70(0.43-32.05)\\ \end{array}$	0.860 0.501 0.695 0.992 0.317 0.939 0.421 0.104 0.873 0.086 0.054 0.783 - 0.129 - 0.450
VEGFA	Pawlikowskaet al. (2004) Gonget al. (2011)	rs314353 rs2230585 rs144173 rs314308 rs2450818 rs314303 rs2450814 rs2447445 rs699947 rs1547651 rs2010963 rs1413711 rs833069 rs3025030 rs3025030 rs3025035 rs3025039	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3 25/27/4 16/36/20 149/30/0 52/82/39 9/73/98 44/88/40 153/22/1 97/72/11 131/42/4 122/47/5 129/42/4	9(43/38 10)43/37 41/33/16 51/33/6 40)41/9 52/33/6 21/51/33 115/11/0 40)66/19 6/43/78 36)67/18 116/11/0 67/54/6 95/30/1 83/35/5	Allelic Allelic Allelic Allelic Allelic Allelic Allelic Recessive Dominant Dominant Recessive Recessive Recessive Addictive Recessive		→	$\begin{array}{l} 1.79(1.11-2.89)^*\\ 1.46(0.90-2.38)\\ 1.61(0.99-2.61)\\ 0.38(0.20-0.65)\\ 1.36(0.81-2.30)\\ 0.45(0.25-0.79)\\ 0.45(0.25-0.79)\\ 0.45(0.25-0.79)\\ 0.84(0.43-1.62)\\ 2.11(1.01-4.42)^*\\ 0.63(0.34-1.16)\\ 0.75(0.47-1.19)\\ 1.73(0.94-3.20)\\ 0.62(0.30-1.30)\\ 1.31(0.47-3.65)\\ 1.47(0.96-2.66)\\ 3.70(0.43-32.05)\\ 1.28(0.82-1.99)\\ \end{array}$	0.860 0.501 0.695 0.992 0.317 0.939 0.421 0.104 0.873 0.086 0.054 0.763 - 0.129 - 0.450 -
VEGFA TGFβ1	Pawlikowskaet al. (2004) Gonget al. (2011) Liet al. (2012)	rs314353 rs2230585 rs144173 rs314308 rs2250818 rs2250818 rs2247445 rs659947 rs1547651 rs2010963 rs1413711 rs832069 rs30225030 rs30225030 rs3025030 rs3025039 rs3025039 rs3025039	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3 25/27/4 16/36/20 149/30/0 52/82/39 9/73/98 44/88/20/1 153/22/1 131/42/4 122/47/5 129/42/4 13/13/4	9(43/38 10)(43)(37) 41)(33)(16 51)(33)(6 51)(33)(6 21)(51)(33) 21)(51)(33) 21)(51)(33) 21)(51)(33) 6)(6)(19) 6)(43)(78) 30)(6)(19) 6)(10)(10)(10)(10)(10)(10)(10)(10)(10)(10	Allelic Allelic Allelic Allelic Allelic Allelic Allelic Recessive Dominant Dominant Recessive Addictive Recessive Addictive		<b>→</b>	$\begin{array}{l} 1.79(1.11-2.89)^*\\ 1.46(0.90-2.38)\\ 1.61(0.99-2.61)\\ 0.36(0.20-0.65)\\ 1.36(0.81-2.30)\\ 0.45(0.25-0.79)\\ 1.45(0.86-2.45)\\ 0.84(0.43-1.62)\\ 2.11(1.01-4.42)^*\\ 0.63(0.34-1.16)\\ 0.75(0.47-1.19)\\ 1.73(0.94-3.20)\\ 1.31(0.47-3.65)\\ 1.47(0.96-2.26)\\ 3.70(0.43-3.05)\\ 1.28(0.82-1.99)\\ 1.01(0.34-3.01)\\ \end{array}$	0.860 0.501 0.695 0.992 0.317 0.939 0.421 0.104 0.873 0.086 0.054 0.783 - 0.783 - 0.783 - 0.450 - 0.050
VEGFA ΤGFβ1 ΤGFβR2	Pawlikowskaet al. (2004) Gonget al. (2011) Liet al. (2012) Liet al. (2012)	rs314353 rs2230585 rs144173 rs21314308 rs2450818 rs2450818 rs247445 rs599947 rs147371 rs147751 rs832069 rs3025030 rs3025030 rs3025030 rs3025039 rs3026039 rs3026489	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3 25/27/4 16/36/20 52/82/39 9/73/98 44/88/40 153/22/1 97/72/11 131/42/4 122/47/5	9(43/38 10)(43)(37 41)(33)(6 51)(33)(6 40)(41)(9 22)(33)(5 21)(51)(33) 115/(1)/0 40)(66)(19 6)(43)(78 36)(67)(18 116/(1)/0 67)(54)(6 81)(39)(6)(39)(6)(6)(6)(6)(6)(6)(6)(6)(6)(6)(6)(6)(6)	Allelic Allelic Allelic Allelic Allelic Allelic Allelic Cominant Dominant Dominant Recessive Addictive Recessive Recessive Addictive Dominant Dominant		→ →	$\begin{array}{l} 1.79(1.11-2.89)^*\\ 1.46(0.90-2.38)\\ 1.61(0.99-2.61)\\ 0.36(0.20-0.65)\\ 1.36(0.81-2.30)\\ 0.45(0.28-0.79)\\ 1.45(0.86-2.45)\\ 0.84(0.43-1.62)\\ 2.11(1.01-4.42)^*\\ 0.63(0.34-1.16)\\ 0.75(0.47-1.19)\\ 0.63(0.34-1.16)\\ 0.75(0.47-1.30)\\ 1.31(0.47-3.65)\\ 1.47(0.96-2.26)\\ 3.70(0.43-32.05)\\ 1.28(0.82-1.99)\\ 1.28(0.82-1.99)\\ 1.30(0.43-3.01)\\ 4.35(0.42-44.88)\\ \end{array}$	0.860 0.501 0.695 0.992 0.317 0.939 0.421 0.104 0.873 0.066 0.066 0.783 - 0.129 - 0.129 - 0.129 - 0.129 - 0.501
VEGFA TGFβ1	Pawlikowskaet al. (2004) Gonget al. (2011) Liet al. (2012)	rs314353 rs2230585 rs144173 rs314308 rs2250818 rs2250818 rs2247445 rs659947 rs1547651 rs2010963 rs1413711 rs832069 rs30225030 rs30225030 rs3025030 rs3025039 rs3025039 rs3025039	10/28/18 10/32/14 39/15/2 25/27/4 39/14/3 25/27/4 16/36/20 149/30/0 52/82/39 9/73/98 44/88/20/1 153/22/1 131/42/4 122/47/5 129/42/4 13/13/4	9(43/38 10)(43)(37) 41)(33)(16 51)(33)(6 51)(33)(6 21)(51)(33) 21)(51)(33) 21)(51)(33) 21)(51)(33) 6)(6)(19) 6)(43)(78) 30)(6)(19) 6)(10)(10)(10)(10)(10)(10)(10)(10)(10)(10	Allelic Allelic Allelic Allelic Allelic Allelic Allelic Recessive Dominant Dominant Recessive Addictive Recessive Addictive		→ →	$\begin{array}{l} 1.79(1.11-2.89)^*\\ 1.46(0.90-2.38)\\ 1.61(0.99-2.61)\\ 0.36(0.20-0.65)\\ 1.36(0.81-2.30)\\ 0.45(0.25-0.79)\\ 1.45(0.86-2.45)\\ 0.84(0.43-1.62)\\ 2.11(1.01-4.42)^*\\ 0.63(0.34-1.16)\\ 0.75(0.47-1.19)\\ 1.73(0.94-3.20)\\ 1.31(0.47-3.65)\\ 1.47(0.96-2.26)\\ 3.70(0.43-3.05)\\ 1.28(0.82-1.99)\\ 1.01(0.34-3.01)\\ \end{array}$	0.860 0.501 0.695 0.992 0.317 0.939 0.421 0.104 0.873 0.086 0.054 0.783 - 0.783 - 0.783 - 0.450 - 0.050

FIGURE 2

Summary and forest plots for the reported variants; \*, calculated by multivariant analysis.

## 4 Discussion

This cross-sectional study systematically reviewed the published studies on genetic factors associated with bAVM-related hemorrhage

and identified nine correlative research works, which were examined for statistical power as well as methodological quality using the NOS and Q-Genie tool. This led to the identification of statistically significant association between bAVM-related hemorrhage and twelve heritable



### FIGURE 3

Methodological quality assessment using Q-genie tool (version 1.1), including 11 items: Item 1, rationale for study; Item 2, selection and definition of outcome of interest; Item 3, selection and comparability of comparison groups; Item 4, technical classification of the exposure; Item 5, non-technical classification of the exposure; Item 6, other sources of bias; Item 7, sample size and power; Item 8, a priori planning of analyses; Item 9, statistical methods and control for confounding; Item 10, testing of assumptions and inferences for genetic analyses; and Item 11, appropriateness of inferences drawn from results.

variants of seven genes (*IL6*, *APOE*, *IL1B*, *IL17A*, *MMP9*, *EPHB4*, and *VEGFA*) involved in the inflammatory and angiogenic signaling pathways. After methodological assessment, limitations were noted in the study designs of the included research works, indicating that the quality and reliability of these studies needed to be improved.

The biological functions of the identified candidate genes are known to be involved in inflammatory and angiogenic signaling pathways. We summarized that SNPs of five inflammatory genes (IL6, APOE, IL1B, IL17A, MMP9) were reported to be associated with bAVM-related hemorrhage. Additionally, it has been reported that these genes can increase the expression of inflammatory cytokines in bAVM tissues, leading to endothelial dysfunction and malformation of vasculature (Krithika and Sumi, 2021). It has been shown that inflammatory infiltration can be observed even in unruptured bAVM lesions, proving the role of inflammation in the development and rupture of the disease (Liu et al., 2022). As the disease progresses, endothelial lesions would weaken the vasculature, and once patients are exposed to a trigger, an acute hemorrhage event occurs. Many studies have investigated genes involved in angiogenesis signaling and their associations with bAVM(18). The reported genes (EPHB4, VEGFA, and also MMP9) are involved in the signaling of vascular endothelial growth factor (VEGF), a representative signaling molecule of angiogenesis. VEGF is highly expressed in endothelial cells of bAVM, especially in ruptured bAVM lesions (McDonald et al., 2015). Activation of this pathway could promote endothelial cell migration and recruitment of smooth muscle cells, resulting in pathological angiogenesis (Liu et al., 2022). A recent study using mouse bAVM models demonstrated that an elevated VEGF level could contribute to bAVM hemorrhage by exposure to variable degrees of higher intraluminal flow and hypertension in the venous system (Cheng et al., 2019).

Almost all of the included studies discussed the limitation of their small sample size (mean, 223.78; standard deviation, 106.40). Only seven (12.50%) of the 52 calculated statistical powers for each SNP were more than 0.80, indicating a lower risk of Type II error. Therefore, a large study cohort would be preferable in order to achieve a statistical power 0.80 or more and to detect relatively reliable association of bAVM-related hemorrhage with the genetic variants of bAVM. All nine included

studies used a CGAS design for the genotyping of individuals. The selection of genes and SNPs was based on their known biological functions. Four studies selected tagging SNPs from the HapMap project data (http://www.hapmap.org), which takes the initiative of genotyping human populations around the world and narrows down the significant loci associated with reviewed diseases (Patnala et al., 2013). The other five studies chose SNPs located in the promoters or exomes of inflammatory/angiogenesis genes to explore their associations with bAVM-related hemorrhage. However, the researchers ignored the possibility that several genes with unknown functions may also be involved in the pathogenic process of bAVM. Thus, with the development of sequencing platforms and techniques, advanced methods should be used to detect the increasing number of genetic associations of bAVMs involving GWAS and WES (McCarthy et al., 2008; Wijmenga and Zhernakova, 2018; Tam et al., 2019). After obtaining sufficiently large whole-genome sequencing datasets, machine learning can be a practical tool to extract key information efficiently (Powell et al., 2021). Based on a variatiy of statistical approaches and biological processes of genes involved in development, signaling, disease, and homeostasis, unsupervised machine learning approaches are not biased by allele frequencies, even without reliance on prior knowledge, to identify heretofore unrecognized genetic risk factors (Powell et al., 2021; Mizikovsky et al., 2022). In addition, the representativeness of the included patients with bAVM should be mentioned. Although most of the studies recruited their cohorts consecutively, the limited number of patients who only came from one or two medical centers failed to represent the populations of their regions or countries, contributing to selection bias. Therefore, we advocate for multi-center cohorts of bAVM patients with large sample sizes to improve the representativeness of the studies.

Another issue that demands greater attention is how to avoid confounding risk factors to ensure comparability. During the study design process, four studies used the strategy of matching and taking baseline characteristics (sex, age, and race) into consideration to reduce confounding bias and improve reliability (Zeldow and Hatfield, 2021). Seven studies were able to achieve statistically significant results by performing multivariate analysis to adjust for not only for the above

### TABLE 2 Summary of the included genetic studies on brain arteriovenous malformation related hemorrhage.

Author (Year)	Study design							Compa	arability					Outcome#		Tota
		Case definition*	Case or exposed cohort representativeness**	Selection of controls or unexposed cases*#	Definition of controls*	Ascertainment of exposure <sup>#</sup>	Demonstration of unhappened outcome of interest <sup>#</sup>	Important factors* <sup>#</sup>	Additional factors* <sup>#</sup>		Same method of ascertainment for cases and controls*	Non- response rate*	Assessment of outcome"	Follow- up period#	Adequacy of follow up of cohorts"	
Pawlikowska et al. (2004)	Case- control	According to cerebral angiography by an attending interventional neuroradiologist. (*)	Consecutively recrruited in single medical center. (*)	Unruptured bAVM patients recruited in the same medical center. (*)	Patients had clinical presentation with ICH and signs of new intracranial hemorrhage on CT or MRI. (*)	-	-	Including race and gender, but not diagnosed age. (-)	Including bAVM size and venous drainage. (*)	Template-directed dye- terminator incorporation assay with fluorescence polarization detection. (*)	Yes(*)	NA (-)	-	_	_	7
Achrol et al. (2006)	Cohort	_	Consecutively recruited in two medical center. (*)	Unruptured bAVM patients recruited in the same medical centers. (*)	_	Template-directed dye- terminator incorporation assay with fluorescence polarization detection. (*)	New ICH (*)	Including diagnosed age, race and gender. (*)	Including bAVM size, venous drainage and initial ICH presentation. (*)	_	_	_	Patients had clinical presentation with ICH and signs of new intracranial hemorrhage on CT or MRI. (*)	Median follow-up time was 0.31 years and interquartile range was 1.40 years (-)	NA (-)	7
Pawlikowska et al. (2006)	Cohort	_	Consecutively recruited in two medical center. (*)	Unruptured bAVM patients recruited in the same medical centers. (*)	_	Template-directed dye- terminator incorporation assay. (*)	New ICH (*)	Including diagnosed age, race and gender. (*)	Including bAVM size, venous drainage and initial ICH presentation. (*)	_	_	_	Patients had clinical presentation with ICH and signs of new intracranial hemorrhage on CT or MRI. (*)	Median follow-up time was 0.30 years and interquartile range was 1.36 years (-)	NA (-)	7
Kim et al. (2009)	Cohort	_	Consecutively recrruited in two medical center. (*)	Unruptured bAVM patients recruited in the same medical centers. (*)	_	Template-directed dye- terminator incorporation assay with fluorescence polarization detection. (*)	New ICH (*)	NA (-)	NA (-)	_	_	_	Patients had clinical presentation with ICH and signs of new intracranial hemorrhage on CT or MRI. (*)	Mean follow-up time was 3.1 years and standard deviation was 7.5 years (-)	NA (-)	5
Weinsheimer et al. (2009)	Case- control	According to cerebral angiography by an attending interventional neuroradiologist, or checked in records by a trained medical records analyst. (*)	Consecutively recruited in two medical center. (*)	Unruptured bAVM patients recruited in the same medical centers. (*)	Patients had clinical presentation with ICH and signs of new intracranial hemorrhage on CT or MRL (*)	_	_	Including race and gender, but not diagnosed age. (-)	Including venous drainage, but not bAVM size. (-)	Beckman Coulter SNPstream 48plex technology or by template-directed primer extension with fluorescence polarization detection. (*)	Yes (*)	NA (-)	_	_	_	6
Gong et al. (2011)	Case- control	According to pathology or angiography. (*)	Consecutively recrruited in single medical center. (*)	Unruptured bAVM patients recruited in the same medical center. (*)	Patients had clinical presentation with ICH and signs of new intracranial hemorrhage on CT or MRI. (*)	_	_	Including diagnosed age, race, and gender. (*)	Neither bAVM size and venous drainage were not matched. (-)	Allele-specific MALDI- TOF mass spectrometry assay with MassARRAY iPLEX platform. (*)	Yes (*)	NA (-)	-	_	_	7
.i et al. (2012)	Case control	According to angiography. (*)	NA (-)	Unruptured bAVM patients recruited in the same medical center. (*)	Patients had clinical presentation with ICH and signs of new intracranial hemorrhage on CT or MRI. (*)	_	_	Including race, but not diagnosed age and gender. (-)	Neither bAVM size and venous drainage were not matched. (-)	Sanger sequencing. (*)	Yes (*)	NA (-)	_	_	_	5

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Outcome# Total score	Follow- Adequacy up of follow period" up of cohorts"	•   	un   
	Non- Assessment response of outcome <sup>t</sup> rate*	лч (-)	NA (.)
Exposure*	int* Same method of ascertainment for cases and controls*	RRAV Yes (*) B	ator Yes (*) V1.1 1 t and 3100 - (*)
	onal Ascertainment* rs**	bAVM Sequenom MasARRAV venous SNP genatyping were not platform. (*) et. (.)	<ul> <li>venous Big Dye Terninator but not Cycle Sequencing VI.11 lize. (-) Ready Reaction kit and automated ABI-3100 DNA sequencer. (*)</li> </ul>
Comparability	Important Additional factors** factors**	Including race Neither bAVM and gender, but size and venous not diagnosed drainage were not age. (.) matched. (.)	Including Including verous diagnosed age, drainage, but not race and bAVM size. (-) gender. (*)
	Demonstration of unhappened outcome of interest <sup>®</sup>	1	I
	Ascertainment of exposure <sup>®</sup>	1	1
Selection	f Definition of controls*	Patients had clinical presentation with ICH and signs of new intracranial hemorrhage on CT or MRL (*)	NA (-)
Sel	Selection of controls or unexposed cases**	Unruptured bAVM patients recruited in the same medical center. (*)	Unruptured bAVM patients recruited in the same medical center. (*)
	Case or exposed cohort representativeness**	Consentively recruited in single medical center. (*)	() NA ()
	Case definition*	According to angiography. (*)	According to pathology or angiography. (*)
Study design		Case- control	Case-
Author (Year)		Sun et al. (2012)	Delev et al. (2017)

mentioned baseline characteristics, but also for the identified morphological risk factors, such as the size and draining veins of bAVM. Three included cohort studies were conducted by the same research team; two of which considered prior hemorrhage an independent risk and confounding factor to assess the associations between genetic factors and bAVM-related hemorrhage, therefore, only new hemorrhagic events were evaluated their association with genetic risk factors, but not the overall risk of hemorrhage. Hence, it would be preferable to recruit patients with unruptured bAVM in future cohort studies. Additionally, it should be noted that the follow-up period was too short (median follow-up period was 4 months in two studies, and average follow-up was 3.1 years in one) to obtain reliable results, since some patients may have experienced hemorrhagic events after the last follow-up, resulting in a misclassification. Furthermore, the researchers used the time point and clinical information of the last follow-up, instead of regarding them as lost subjects. Calculating the rate of loss to follow-up and evaluating the adequacy of the follow-up stage are challenging tasks. Thus, in order to improve the research quality and accuracy of results, it is necessary to set an appropriate follow-up duration so that outcome events can be observed as much as possible, while preventing excessive environmental confounding factors.

To promote the genetic study on bAVM-related hemorrhage, the methodological issues in other diseases are also worth referring to. Two strategies are usually performed to identify the disease-causing variants: sequencing affected individuals with a family history and extreme-trait sequencing (Cirulli and Goldstein, 2010). The first strategy is widely adopted for rare neurological and cerebrovascular diseases, such as moyamoya disease and hereditary hemorrhagic telangiectasia (Liu et al., 2011; Benzinou et al., 2012). It is easier to detect co-effected members within families to identify overlapping variants. Considering a relatively low incidence of bAVM and its related hemorrhage, as well as its rare familial cases, it is of more value for clinicians to consciously collect these individuals for genetic study in their clinical practice (Chen et al., 2020). Extreme-trait sequencing is based on a hypothesis that the enrichment of rare variants would result in an extreme phenotype (Cirulli and Goldstein, 2010). Thus, it could be efficient to filter damaging variants by recruiting individuals with extreme-traits, including: large lesions, epileptic symptoms or an early onset of hemorrhage.

This study had several limitations. First, owing to the low incidence of bAVM, only a small number of studies with small sample sizes and insufficient statistical powers were included in the assessment. Second, the included studies published only in English or Chinese, resulting in a reduction of representativeness. Third, we calculated the statistical powers assuming  $\alpha = 0.05$ , while the results would be more reliable by using the Bonferroni correction and multiple comparisons similar to those used in GWAS. However, the sample sizes of the existing studies did not reach a statistical significance. Thus, multi-center studies are warranted to increase the sample size and improve comparability.

### 5 Conclusion

Several genes were identified to be associated with bAVM-related hemorrhage, including: IL6, IL17A, MMP9, VEGFA, EPHB4. Efforts are needed to investigate the mechanism of bAVM-related hemorrhage in the future. We call for the establishment of regional alliances and rare disease banks in order to perform a

of NOS; (-), studies did not win the star in each item of NOS

multicenter prospective cohort study with a large sample size of patients with bAVM and adequate follow-up. Familial and extreme-trait cases are precious genetic resources for highthroughput sequencing. Furthermore, it may be more efficient to investigate genetic risks capitalizing on machine learning, Multi-omic systems-based approaches should be utilized to uncover the roles of candidate genes in bAVM development. Designers and researchers should strive to improve the methodological quality of their studies according to the demands of assessment tools, especially avoiding confounding risk factors, to ensure comparability.

### Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding authors.

### Author contributions

JJ: Data curation, statistical analysis, resources and software, writing for original draft; ZQ: Data curation, resources and software; JY: Administration and supervision, methodology, original draft review and editing; JL: Administration and supervision,

### References

Achrol, A. S., Pawlikowska, L., McCulloch, C. E., Poon, K. Y., Ha, C., Zaroff, J. G., et al. (2006). Tumor necrosis factor-alpha-238G>A promoter polymorphism is associated with increased risk of new hemorrhage in the natural course of patients with brain arteriovenous malformations. UCSF BAVM Study Project. *Stroke* 37 (1), 231–234. doi:10.1161/01.STR.0000195133.98378.4b

Atkinson, R. P., Awad, I. A., Batjer, H. H., Dowd, C. F., Furlan, A., Giannotta, S. L., et al. (2001). Reporting terminology for brain arteriovenous malformation clinical and radiographic features for use in clinical trials. *Stroke* 32 (6), 1430–1442. doi:10.1161/01. str.32.6.1430

Benzinou, M., Clermont, F. F., Letteboer, T. G., Kim, J. H., Espejel, S., Harradine, K. A., et al. (2012). Mouse and human strategies identify PTPN14 as a modifier of angiogenesis and hereditary haemorrhagic telangiectasia. *Nat. Commun* 3, 616. doi:10.1038/ncomms1633

Chen, C. J., Ding, D., Derdeyn, C. P., Lanzino, G., Friedlander, R. M., Southerland, A. M., et al. (2020). Brain arteriovenous malformations: A review of natural history, pathobiology, and interventions. *Neurology* 95 (20), 917–927. doi:10.1212/WNL. 000000000010968

Cheng, P., Ma, L., Shaligram, S., Walker, E. J., Yang, S. T., Tang, C., et al. (2019). Effect of elevation of vascular endothelial growth factor level on exacerbation of hemorrhage in mouse brain arteriovenous malformation. *J. Neurosurg.* 132 (5), 1566–1573. doi:10. 3171/2019.1.JNS183112

Cirulli, E. T., and Goldstein, D. B. (2010). Uncovering the roles of rare variants in common disease through whole-genome sequencing. *Nat. Rev. Genet.* 11 (6), 415–425. doi:10.1038/nrg2779

Delev, D., Pavlova, A., Grote, A., Bostrom, A., Hollig, A., Schramm, J., et al. (2017). NOTCH4 gene polymorphisms as potential risk factors for brain arteriovenous malformation development and hemorrhagic presentation. *J. Neurosurg.* 126 (5), 1552–1559. doi:10.3171/2016.3.JNS151731

Derdeyn, C. P., Zipfel, G. J., Albuquerque, F. C., Cooke, D. L., Feldmann, E., Sheehan, J. P., et al. (2017). Management of brain arteriovenous malformations: A scientific statement for healthcare professionals from the American heart association/American stroke association. *Stroke* 48 (8), e200–e224. doi:10.1161/STR.000000000000134

Gong, Z. P., Qiao, N. D., Gu, Y. X., Song, J. p., Li, P. I., Qiu, H. j., et al. (2011). Polymorphisms of VEGFA gene and susceptibility to hemorrhage risk of brain arteriovenous malformations in a Chinese population. *Acta Pharmacol. Sin.* 32 (8), 1071–1077. doi:10.1038/aps.2011.76

Kim, H., Hysi, P. G., Pawlikowska, L., Poon, A., Burchard, E. G., Zaroff, J. G., et al. (2009). Common variants in interleukin-1-Beta gene are associated with intracranial hemorrhage and susceptibility to brain arteriovenous malformation. *Cerebrovasc. Dis.* 27 (2), 176–182. doi:10.1159/000185609

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### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Krithika, S., and Sumi, S. (2021). Neurovascular inflammation in the pathogenesis of brain arteriovenous malformations. J. Cell Physiol. 236 (7), 4841–4856. doi:10.1002/jcp.30226

Li, X. S., Jiang, N., Guo, S. L., Liang, F., and Qi, T. W. (2012). Correlation of single nucleotide polymorphisms of three genes and hemorrhage risk in patients with the BAVM. *J. Trop. Med.* 12 (8), 977–979.

Liu, J., Li, Y., Zhang, H., Luo, C., Yuan, D., Jiang, W., et al. (2022). Associated genetic variants and potential pathogenic mechanisms of brain arteriovenous malformation. *J. Neurointerv Surg.*, neurintsurg-2022-018776. doi:10.1136/neurintsurg-2022-018776

Liu, W., Morito, D., Takashima, S., Mineharu, Y., Kobayashi, H., Hitomi, T., et al. (2011). Identification of RNF213 as a susceptibility gene for moyamoya disease and its possible role in vascular development. *PLoS One* 6 (7), e22542. doi:10.1371/journal. pone.0022542

McCarthy, M. I., Abecasis, G. R., Cardon, L. R., Goldstein, D. B., Little, J., Ioannidis, J. P. A., et al. (2008). Genome-wide association studies for complex traits: Consensus, uncertainty and challenges. *Nat. Rev. Genet.* 9 (5), 356–369. doi:10.1038/nrg2344

McDonald, J., Wooderchak-Donahue, W., VanSant Webb, C., Whitehead, K., Stevenson, D. A., and Bayrak-Toydemir, P. (2015). Hereditary hemorrhagic telangiectasia: Genetics and molecular diagnostics in a new era. *Front. Genet.* 6, 1. doi:10.3389/fgene.2015.00001

Mizikovsky, D., Naval Sanchez, M., Nefzger, C. M., Cuellar Partida, G., and Palpant, N. J. (2022). Organization of gene programs revealed by unsupervised analysis of diverse gene-trait associations. *Nucleic Acids Res.* 50 (15), e87. doi:10. 1093/nar/gkac413

Patnala, R., Clements, J., and Batra, J. (2013). Candidate gene association studies: A comprehensive guide to useful *in silico* tools. *BMC Genet.* 14, 39. doi:10.1186/1471-2156-14-39

Pawlikowska, L., Poon, K. Y., Achrol, A. S., McCulloch, C. E., Ha, C., Lum, K., et al. (2006). Apolipoprotein E epsilon 2 is associated with new hemorrhage risk in brain arteriovenous malformations. *Neurosurgery* 58 (5), 838–843. ; discussion 838-43. doi:10. 1227/01.NEU.0000209605.18358.E5

Pawlikowska, L., Tran, M. N., Achrol, A. S., McCulloch, C. E., Ha, C., Lind, D. L., et al. (2004). Polymorphisms in genes involved in inflammatory and angiogenic pathways and the risk of hemorrhagic presentation of brain arteriovenous malformations. *Stroke* 35 (10), 2294–2300. doi:10.1161/01.STR.0000141932.44613.b1

Powell, S. K., O'Shea, C., Brennand, K. J., and Akbarian, S. (2021). Parsing the functional impact of noncoding genetic variants in the brain epigenome. *Biol. Psychiatry* 89 (1), 65–75. doi:10.1016/j.biopsych.2020.06.033

Rutledge, W. C., Ko, N. U., Lawton, M. T., and Kim, H. (2014). Hemorrhage rates and risk factors in the natural history course of brain arteriovenous malformations. *Transl. Stroke Res.* 5 (5), 538–542. doi:10.1007/s12975-014-0351-0

Sohani, Z. N., Meyre, D., de Souza, R. J., Joseph, P. G., Gandhi, M., Dennis, B. B., et al. (2015). Assessing the quality of published genetic association studies in meta-analyses: The quality of genetic studies (Q-Genie) tool. *BMC Genet.* 16, 50. doi:10.1186/s12863-015-0211-2

Stang, A. (2010). Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. *Eur. J. Epidemiol.* 25, 603–605. doi:10.1007/s10654-010-9491-z

Sun, B., Qiu, H., Zhao, F., Qiao, N., Fan, W., Lu, D., et al. (2012). The rs9509 polymorphism of MMP-9 is associated with risk of hemorrhage in brain arteriovenous malformations. *J. Clin. Neurosci.* 19 (9), 1287–1290. doi:10.1016/j. jocn.2011.09.036

Tam, V., Patel, N., Turcotte, M., Bossé, Y., Paré, G., and Meyre, D. (2019). Benefits and limitations of genome-wide association studies. *Nat. Rev. Genet.* 20 (8), 467–484. doi:10.1038/s41576-019-0127-1

van Beijnum, J., van der Worp, H. B., Buis, D. R., Al-Shahi Salman, R., Kappelle, L. J., Rinkel, G. J. E., et al. (2011). Treatment of brain arteriovenous malformations: A systematic review and meta-analysis. *JAMA* 306 (18), 2011–2019. doi:10.1001/jama.2011.1632

Weinsheimer, S., Kim, H., Pawlikowska, L., Chen, Y., Lawton, M. T., Sidney, S., et al. (2009). EPHB4 gene polymorphisms and risk of intracranial hemorrhage in patients with brain arteriovenous malformations. *Circ. Cardiovasc Genet.* 2 (5), 476–482. doi:10. 1161/CIRCGENETICS.109.883595

Wijmenga, C., and Zhernakova, A. (2018). The importance of cohort studies in the post-GWAS era. *Nat. Genet.* 50 (3), 322–328. doi:10.1038/s41588-018-0066-3

Zeldow, B., and Hatfield, L. A. (2021). Confounding and regression adjustment in differencein-differences studies. *Health Serv. Res.* 56 (5), 932–941. doi:10.1111/1475-6773.13666