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RECEIVED 07 April 2024

ACCEPTED 18 June 2024

PUBLISHED 10 July 2024

CITATION

Spaner C, Durkee-Shock J, Weng A,
Stubbins R, Gerrie AS, Pittaluga S, Cohen JI
and Chen LYC (2024) Case report:
Aggressive natural killer cell leukemia
and refractory hemophagocytic
lymphohistiocytosis in an adolescent.
Front. Hematol. 3:1413794.
doi: 10.3389/frhem.2024.1413794

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Case report: Aggressive natural killer cell leukemia and refractory hemophagocytic lymphohistiocytosis in an adolescent

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Aggressive natural killer cell leukemia (ANKL) is a rare, aggressive hematologic malignancy which often presents as fulminant Epstein-Barr virus (EBV)- driven hemophagocytic lymphohistiocytosis (HLH). ANKL lacks a distinct immunologic and morphologic signature, making early diagnosis particularly challenging. Here we present a case of ANKL in a patient presenting with EBV-HLH. After poor treatment response to the HLH-2004 protocol (etoposide and dexamethasone), bone marrow biopsy demonstrated an atypical CD3-/CD56+ natural killer (NK) cell population with diminished CD7 expression consistent with EBV+ ANKL. Asparaginase-based chemotherapy was initiated but his disease progressed and he died from multiorgan failure. This case highlights the diagnostic challenges of ANKL given the lack of standardized diagnostic criteria, the importance of considering T/NK cell malignancies in the differential diagnosis of EBV-HLH, and adds to the literature on this rare disease.

KEYWORDS

hemophagocytic lymphohistiocytosis, aggressive natural killer cell leukemia, EBV - Epstein-Barr virus, PEG-asparaginase, T/NK cell malignant lymphoma

Introduction

Aggressive natural killer cell leukemia (ANKL) is a rare disease characterized by neoplastic proliferation of natural killer (NK) cells in the blood, bone marrow, liver, and spleen (1, 2). Approximately 90% of cases are driven by Epstein Barr Virus (EBV) (3). ANKL is classified by the World Health Organization as a “mature T/NK cell neoplasm” (4). However, it lacks a

unique immunophenotypic or molecular diagnostic signature, making early diagnosis particularly challenging. Most patients are of East Asian descent, and the prognosis is poor, with a median survival of 2 months. Moreover, ANKL commonly presents as fulminant EBV-driven hemophagocytic lymphohistiocytosis (HLH), which can be very difficult to distinguish from other discrete causes of EBV-HLH. HLH is a cytokine storm syndrome characterized by pathologic activation of T lymphocytes and macrophages, resulting in marked immune dysregulation, hyperinflammation, and multiorgan failure (5, 6). HLH can be caused by genetic mutations affecting lymphocyte cytotoxicity (primary HLH) or in the setting of underlying illness (secondary HLH), such as malignancy, infection and autoimmune disease.

The differential diagnosis for a patient presenting with EBV-HLH includes distinct malignant and non-malignant etiologies (7–10). Non-neoplastic causes include acute infection. Neoplastic causes include B cell lymphomas, due to the tropism of EBV to B cells; T- and NK-cell lymphomas, including ANKL, extranodal NK/T-cell lymphoma (ENKL), chronic active EBV disease (CAEBV), and systemic EBV positive T-cell lymphoma of childhood (11). There is significant clinicopathologic overlap between these entities. As well, the tumor burden in ANKL can be low, making it even more difficult to elucidate a precise diagnosis in an acutely ill patient. Less than 500 cases of this rare disease have been reported in the literature, and the vast majority of these are from Asia. We present the case of an adolescent who presented in British Columbia, Canada, with florid EBV-HLH and had a fatal outcome.

Case report

A 17-year-old previously healthy male of Chinese descent presented to the emergency department of a community hospital with a 5-day history of headache and fever up to 39.4°C. Physical examination on admission was significant for palpable hepatosplenomegaly without lymphadenopathy. Initial blood work (Table 1) showed pancytopenia with reactive lymphocytes and pelgeroid neutrophils seen on peripheral blood film. No blasts or large lymphoid cells were reported. Liver enzymes including ALT, AST, and bilirubin were elevated, and he had a prolonged aPTT but normal INR and fibrinogen. Ferritin was markedly elevated at 10,614 ug/L (normal range 15–300 ug/L), with elevated LDH, C-reactive peptide (CRP), and fibrin D-dimer. CXCL9 was elevated at 6,395 pg/mL (normal range ≤ 657 pg/mL), as was soluble interleukin 2 receptor (sIL2r) (16,426 U/mL, normal range 241–846 U/mL). Monospot test was positive, with Epstein-Barr virus (EBV) serology positive for reactive IgG, nonreactive IgM, and a viral load by plasma PCR of 498,000 copies/mL (Table 1). Autoimmune serology, HIV, and viral hepatitis panels were negative.

A working diagnosis of non-neoplastic EBV-HLH was made based on the HLH-2004 criteria. He was treated initially with dexamethasone 10mg/m², with the goal of avoiding cytotoxic chemotherapy. However, his fever worsened and his EBV viral load increased to 8,910,000 copies/mL by day 5 of admission so he was transferred to our tertiary care hospital. Etoposide 150 mg/m² was added, and he received intravenous immunoglobulin (IVIg)

TABLE 1 Inflammatory markers at time of diagnosis and peak values during course of disease.

Lab marker	Value at presentation	Peak value	Reference range
Ferritin (ug/L)	10,614	333,070	15–300
D-dimer (ug/L)	11,458	55,092	<500
Soluble interleukin-2 receptor (U/mL)	16,426	16,426	241–846
CXCL9 (pg/mL)	6,395	Not applicable*	≤ 657
Creatinine (umol/L)	62	94	45–115
Triglycerides (mmol/L)	3.5	6.3	<1.70
Aspartate aminotransferase (U/L)	227	332	<35
Alanine transaminase (U/L)	125	433	10–55
Lactate dehydrogenase (U/L)	1,662	15,620	90–240
C-reactive peptide (mg/L)	66.2	187.0	<3.1
Bilirubin, total (umol/L)	27	326	<20
Albumin (g/L)	29	11	32–50
International normalized ratio	1.1	2.1	0.9–1.2
Activated partial thromboplastin time (seconds)	43	63	25–38
Epstein-Barr viral load, plasma (copies/mL)	498,000	21,200,000	No reference range

*CXCL9 was measured one time on day 46 of admission.

0.5 g/kg, and rituximab 375 mg/m² targeting EBV-infected B cells (12). Cyclosporine is not typically used initially for management of HLH in our center as it is felt to have limited benefit (13). His HLH parameters remained persistently elevated, and ruxolitinib 10 mg twice daily was then initiated for refractory HLH (14, 15). His biochemical and clinical parameters subsequently improved.

Bone marrow biopsy and aspiration performed on day 2 of admission revealed increased histiocytes and occasional hemophagocytosis, and an abnormal population of large, atypical cells with round to irregular, occasionally convoluted nuclei (Figure 1). Cellularity was 80%, with trilineage hematopoiesis present. Flow cytometry demonstrated an atypical CD3-/CD56+ NK cell population with diminished CD7 expression accounting for 27% of the lymphocyte population (Figure 2) (16). The CD4/8 ratio was 1:1 and no aberrant T cells were noted. Additional immunohistochemistry showed frequent scattered large, atypical lymphoid cells positive for CD56 (Figure 3A), cytoplasmic CD3, CD2, CD56, CD7 (small subset),

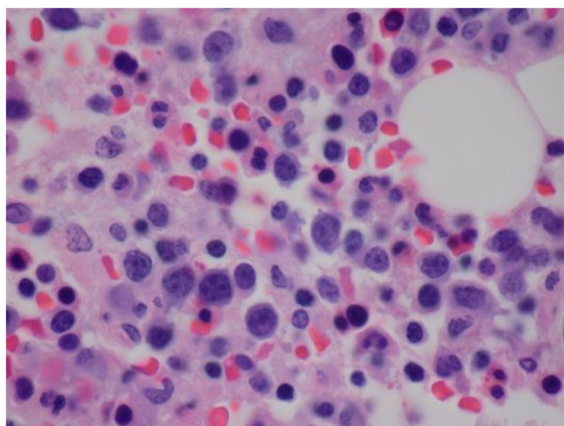


FIGURE 1

Histopathology of bone marrow demonstrating increased histiocytes and an abnormal population of large, atypical cells with round to irregular, occasionally convoluted nuclei, condensed chromatin, multiple distinct nucleoli and limited cytoplasm (Hematoxylin and Eosin (H&E) staining).

granzyme B and TIA1, and negative for perforin, BFI, TCR gamma, TCR delta, CD4, and CD8. A number of these large, atypical cells stained positively for Epstein-Barr encoded RNA (EBER) by *in situ* hybridization (Figure 3B). PCR for T-cell receptor beta showed a polyclonal pattern. Given the atypical cytotoxic NK cell population, marked atypia of the NK cells on CD56 immunostain, and less atypia among CD4+/CD8+ T cells, or CD20+ B cells, a diagnosis of aggressive NK cell leukemia (ANKL) was favored.

Genetic testing for germline variants through Blueprint Genetics Comprehensive Immune and Cytopenia Panel for HLH demonstrated microdeletion of 22q.11.2 of unclear significance. No germline pathogenic variants consistent with primary HLH were found. The specimens were sent for further review at the National Institutes of Health (NIH), which confirmed the diagnosis of ANKL.

Because his HLH parameters continued to worsen, the patient was started on an asparagine-based SMILE chemotherapy regimen (dexamethasone, ifosfamide, pegaspargase, etoposide) for ANKL

prior to a definitive diagnosis (17). He did not receive methotrexate, typically part of SMILE regimen, due to liver dysfunction. The goal of treatment was for disease control in order to allow for allogeneic stem cell transplant (18, 19).

His HLH parameters temporally improved, but then worsened at the 3-week mark of cycle 1 with marked hyperbilirubinemia, hypotension, and an increase in EBV viral load to 21 million copies/mL (Table 1). Accordingly, he was restarted on etoposide and ruxolitinib. On day 14 of cycle 2 of SMILE, his ferritin and EBV viral load continued to worsen with declining liver function. He was started on gemcitabine and oxaliplatin for disease control. Emopalumab, an interferon gamma inhibitor, was also given (20). The emopalumab was started quite late in his course as this medication requires special access in Canada and is not routinely available. He had transient clinical and biochemical responses to emopalumab with defervescence and improvement in ferritin (Figure 4).

His course was complicated by neutropenic infections with polymicrobial bacteremia (*Klebsiella pneumoniae*, *Escherichia coli*, *Roseomonas mucosa*, *Stenotrophomonas maltophilia*, and vancomycin-resistant *Enterococcus faecium*). Allo-HSCT was considered and a donor was identified, however it was not pursued given his worsening organ function, bacteremia, and inability to achieve partial or complete remission. He passed away due to severe lactic acidosis and multiorgan failure 78 days after his initial presentation.

Discussion

ANKL is a rare and aggressive hematologic malignancy which lacks a specific immunophenotypic or molecular diagnostic signature, making early diagnosis and treatment challenging. The presentation of ANKL varies, but typically presents as fulminant HLH with multi-organ failure, while the presence of B-symptoms (fevers, night sweats, unintentional weight loss) and lymphadenopathy often signifies leukemic disease (16). Several factors likely contribute to the immune dysregulation leading to HLH in ANKL and other NK cell disorders. First, the CD56^{bright} subset of neoplastic NK cells secretes high levels of

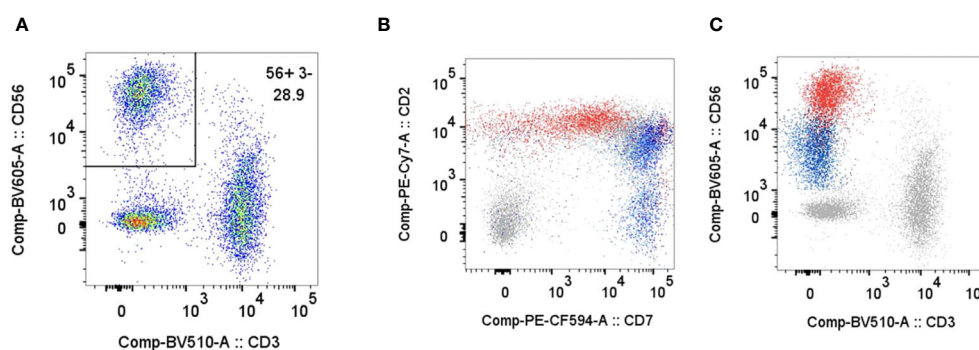


FIGURE 2

Flow cytometry on bone marrow aspirate specimen. Plots for CD3/CD56 (A), CD7/CD2 (B), CD3/CD56 (C) show a CD56+/CD3- NK cell population comprising 28.9% of the gated lymphocytes (CD45+, Low SSC, CD14-). The patient's NK cells demonstrate brighter CD56, slightly brighter CD2, and dimmer CD7 compared to normal NK cells. Plots in B and C show an overlay of the patient's gated lymphocytes in grey including abnormal NK cells in red, and NK cells from normal patients in blue.

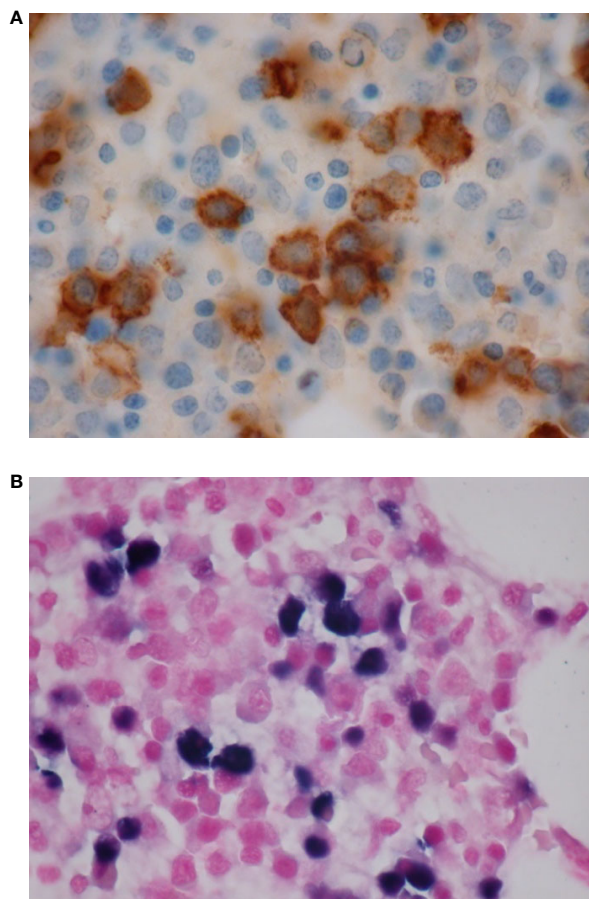


FIGURE 3

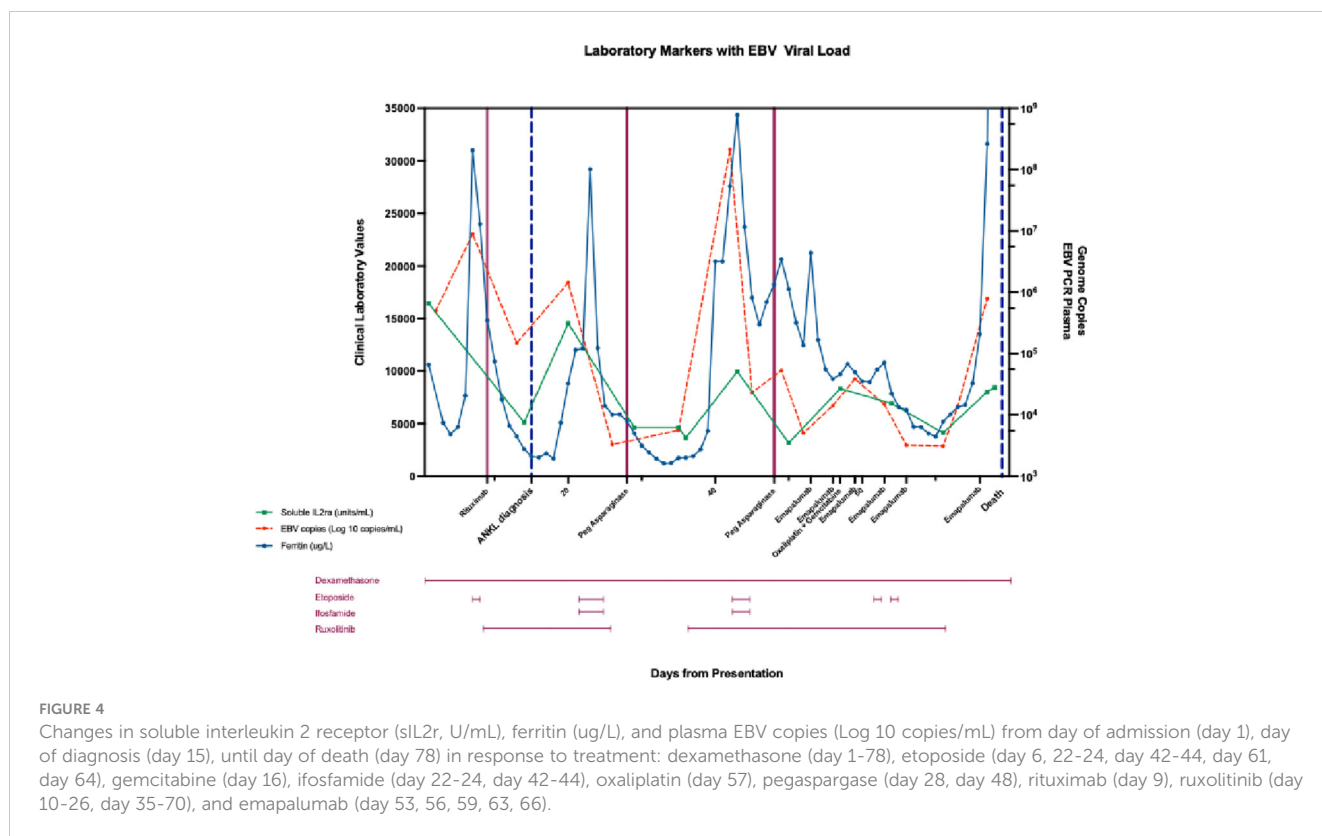
(A) Immunohistochemical staining of bone marrow with anti-CD56 monoclonal antibody. (B) *In situ* hybridization of bone marrow for Epstein-Barr virus encoded RNA (EBER), with numerous positive cells, of varying size.

interferon gamma (IFN- γ), resulting in activation of macrophages and histiocytes, leading to the pathologic immune activation seen in HLH (21). In a subset of patients with extranodal T/NK cell lymphoproliferative disorders, a recurrent somatic mutation, ECSIT-T419C activates the NF- κ B signaling pathway (22). This results in the downstream activation of tumor necrosis factor (TNF), IFN- γ , and interleukin-1 β (IL-1 β), suggesting that the aberrant production of these cytokines may be the driver of HLH seen in NK-cell malignancies, including ANKL.

The differential diagnosis of ANKL includes EBV extranodal NK/T cell lymphoma, which carries overlapping morphological and genetic features with ANKL, but has a less aggressive presentation. Even with early recognition, the prognosis is guarded, but the low burden of neoplastic cells in some cases, lack of standardized diagnostic criteria, and nonspecific morphological features can delay precise diagnosis and classification (2). This case highlights important lessons around the diagnosis of ANKL. First, morphology and immunophenotyping are important clues early in the disease presentation. The morphological finding of NK cells in blood and bone marrow should not be dismissed as simply reactive. Peripheral blood immunophenotyping is a widely available test for helping to rapidly distinguish between different subtypes of

EBV-HLH. Increased CD8 positive T cells are observed in 80% of non-neoplastic EBV-HLH, whereas patients with T/NK CAEBV will often have an increase population of aberrant CD4 positive T cells (7, 23). In this case, the increased NK cells on flow cytometry were highly suggestive of ANKL, and this rare diagnosis was confirmed with immunohistochemistry and consultation with a center of excellence in this disorder.

The differential diagnosis of EBV-HLH is broad and includes nonneoplastic EBV-HLH often driven by monogenic inborn errors of immunity, T or NK cell CAEBV, systemic EBV positive T-cell lymphoma of childhood, and T/NK cell lymphoproliferative disorders, including ENKL, and ANKL (7, 10, 11). However, there are significant clinicopathologic similarities between these diseases. The degree of marrow infiltrate in ANKL can vary from minimal to severe, which can be confused with non-neoplastic EBV-HLH in the bone marrow, and acute presentations of NK-CAEBV can overlap with ANKL (4). In differentiating ANKL from ENKL, nasal type, the former tends to be associated with gain of 1q and loss of 7p15.1-pp22.3 and 17p13.1 alterations compared to the latter, but this is not always the case (24). In our case, an initial diagnosis of non-neoplastic EBV-HLH was made on the absence of immunologic or pathologic evidence of malignancy, and lack of an



expanded atypical CD4+ T cell population or prior history of chronic, high-grade EBV viremia suggestive of NK cell CAEBV. While further immunologic studies revealed an atypical cytotoxic CD56+/CD7- NK cell population, the pathologic diagnosis of ANKL was challenging due to its rarity, and L-asparaginase based chemotherapy was initiated prior to confirmatory diagnosis given the aggressive course and high mortality with delayed treatment.

Outcomes are poor in ANKL, but once diagnosed, the goals of therapy are to control the EBV-HLH and proceed to allogeneic stem cell transplantation (allo-SCT) whenever possible. A Korean study of 21 patients demonstrated overall response rates of 33% to 40% with L-asparaginase-based regimens (25). Remissions are short-lived with median progression-free survival (PFS) 3.9 months. However, some patients can achieve durable remission with allo-SCT; in one study, for patients who achieved even-free survival at twelve months, the OS was 85.2% at 5 years (19). Novel therapies for better control of EBV-HLH leading into transplant are needed. JAK inhibition has shown promise in HLH, particularly in pediatric HLH, but provided only transient partial response in this case. Likewise, emapalumab provided transient improvement in fever and HLH parameters but was initiated relatively late in the course of disease, due in large part to access issues in Canada. Emapalumab was approved by the United States Food and Drug Administration (FDA) based on a single arm study in primary HLH where patients received anti-IFN γ early in the course of their disease (20). However, recent anecdotal data suggests that it may not be as effective when given late in patients with malignancy-associated HLH (26). The immune checkpoint inhibitor nivolumab has shown promise in relapsed and refractory EBV-HLH but has limited utility in patients wherein allo-SCT is the definitive therapy, as nivolumab causes severe graft-vs-host disease and a prolonged wash out period is required (27).

Conclusion

This case highlights the diagnostic challenges in ANKL. EBV+ T/NK cell malignancies must be considered in patients with EBV-HLH, and these patients require early disease control and allogeneic stem cell transplantation.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

Ethical approval was not required for the study involving human samples in accordance with the local legislation and institutional requirements. Written informed consent for participation in this study was provided by the participants' legal guardians/next of kin. Written informed consent was obtained from the individual(s), and minor(s)' legal guardian/next of kin, for the publication of any potentially identifiable images or data included in this article.

Author contributions

CS: Conceptualization, Investigation, Visualization, Writing – original draft, Writing – review & editing. JD: Software, Visualization,

Writing – review & editing. AW: Data curation, Writing – review & editing. RS: Writing – review & editing. AG: Writing – review & editing. SP: Writing – review & editing. JC: Data curation, Writing – review & editing. LC: Conceptualization, Investigation, Methodology, Supervision, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. LC's research is supported by a philanthropic gift from the Hsu & Taylor Family to the UBC & VGH Hospital Foundation. This work was supported by the Division of Intramural Research of the National Institute of Allergy and Infectious Diseases.

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